

Using dynamical reaction network to infer drugs selectivity in pharmacology

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USING DYNAMICAL REACTION NETWORK TO INFER DRUGS SELECTIVITY IN PHARMACOLOGY

Romain Yvinec

BIOS, INRA Centre Val-de-Loire

Outline

What is Drugs Selectivity?

Some examples

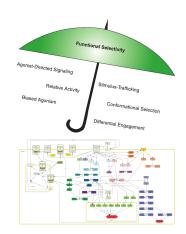
Bias quantification - standard method : operational mode

Biased quantification using dynamical mode

Functional selectivity, biased signaling

What is Drugs Selectivity?

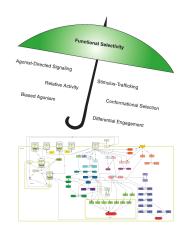
 Several reaction pathways are generally associated to a given receptor, and lead to various cell response.



Functional selectivity, biased signaling

What is Drugs Selectivity?

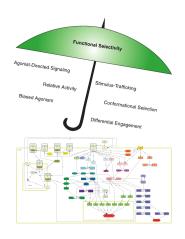
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- Differential activation of those reaction pathways, that differs between (natural or synthetic) ligand



Functional selectivity, biased signaling

What is Drugs Selectivity?

- Several reaction pathways are generally associated to a given receptor, and lead to various cell response.
- Differential activation of those reaction pathways, that differs between (natural or synthetic) ligand
- Drugs Selectivity =
 Ligand-dependent selectivity for certain signal transduction pathways at one given receptor



Key concept in pharmacology

- Drugs Selectivity (or Biased Signaling) is a key concept to be distinguish from
 - Partial or full agonist.
 - Antagonist, inverse agonist.
 - Affinity (K_d) , potency (EC_{50}) , efficacy (E_{max}) .

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- A bias might be context-dependent (cell type, physiological state, etc.)

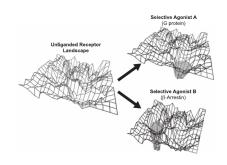
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 - Affinity (K_d) , potency (EC_{50}) , efficacy (E_{max}) .
- A bias might be context-dependent (cell type, physiological state, etc.)
- Biased agonism is becoming a major tool in drug discovery.
- \Rightarrow Candidate screening requires to accurately quantify bias.

Theoretical foundation

A receptor may adopt several spatial conformations, each of which has different activation pathway profiles.

Conformational selectivity = Ligand-specific modification of the energetic landscape, changing affinities and efficacies of signaling patways.





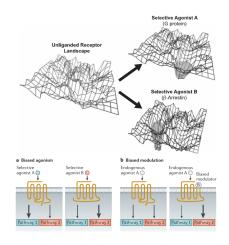
Kenakin, J Pharmacol Exp Ther (2011)

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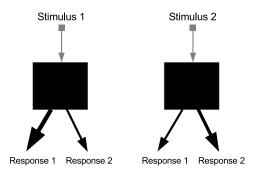
Similar concept : modulating bias





Minimal setting

To speak about signaling bias, one necessarily needs **two** ligands and **two** responses, in a **same** cellular context.



 \Rightarrow We always compare a ligand with respect to a reference one.

Outline

What is Drugs Selectivity?

Some examples

Bias quantification - standard method : operational model

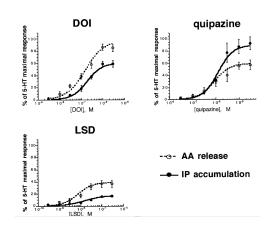
Biased quantification using dynamical mode

Serotonine receptor $5 - HT_{2C}$

- Quipazine is biaised towards PI accumulation with respect to AA production, compared to the reference agonist DOI.
- LSD is not biased.

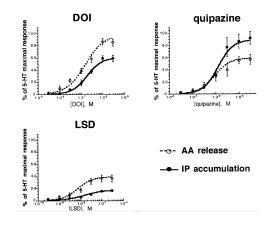


Berg et al., Mol. Pharmacol. (1998)



Serotonine receptor $5 - HT_{2C}$

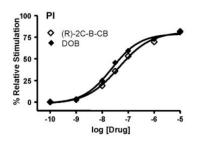
- Quipazine is biaised towards PI accumulation with respect to AA production, compared to the reference agonist DOI.
- LSD is not biased.
- \Rightarrow Bias due to an E_{max} difference.

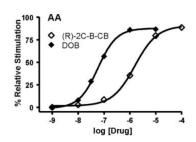




Berg et al., Mol. Pharmacol. (1998)

Serotonine receptor $5 - HT_{2A}$



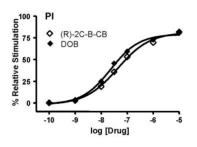


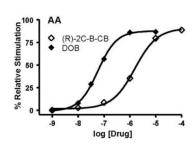
 (R) - 2C - B - CB is biaised towards PI accumulation with respect to AA production, compared to the reference agonist DOB.



Urban et al., J Pharmacol Exp Ther (2007)

Serotonine receptor $5 - HT_{2A}$





- (R) 2C B CB is biaised towards PI accumulation with respect to AA production, compared to the reference agonist DOB.
- \Rightarrow Bias due to an EC_{50} difference.



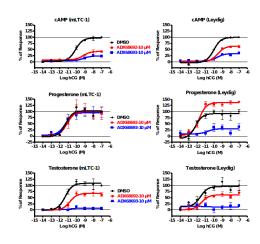
Urban et al., J Pharmacol Exp Ther (2007)

Steroidogenesis modulated by NAM

Some negative allosteric modulators (NAM) can biased Progesterone production with respect to Testosterone production, under stimulation of LH/CG receptor by hCG.



Ayoub et al., *Mol. Cell. Endocrinol* (2016)



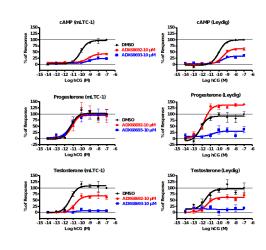
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⇒ Selective (biased) allosteric modulation



Ayoub et al., *Mol. Cell. Endocrinol* (2016)

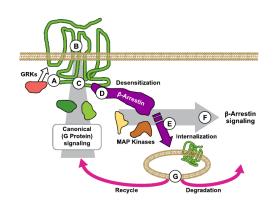


Many more examples on GPCR (principle drug target)

Many GPCR's are known to have biased ligands (G / β -arrestin)



Kenakin, *Chem Rev* (2017)



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Bias quantification - standard method : operational model

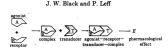
Biased quantification using dynamical mode

Operational model

Dose-response data are fitted with the function

$$y = E_{tot} \frac{\tau^n[L]^n}{([L] + Ka)^n + \tau^n[L]^n}.$$

- Response at equilibrium of a Michaelis-Menten type model.
- Ka = Dissociation constant of the couple Ligand/Receptor
- τ = Efficacy coefficient of the transduction pathway





Black and Leff, *Proc.* R. Soc. Lond. B (1983)

Operational model

Dose-response data are fitted with the function

$$y = E_{tot} \frac{\tau^n[L]^n}{([L] + Ka)^n + \tau^n[L]^n}.$$

For n = 1,

- $EC_{50} = \frac{Ka}{\tau + 1}$
- Efficacy $y_{\infty}/E_{tot} = \frac{\tau}{\tau+1}$

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Black and Leff, *Proc.* R. Soc. Lond. B (1983)

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- Efficacy $y_{\infty}/E_{tot} = \frac{\tau}{\tau+1}$

Then, we define

⇒ Transduction coefficient :

$$R := \log\left(\frac{\tau}{\mathit{Ka}}\right)$$

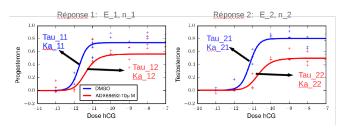
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Black and Leff, *Proc.* R. Soc. Lond. B (1983)

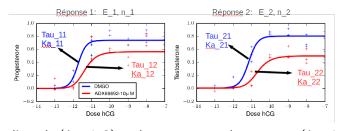
Bias quantification : with the operational model



Two ligands (j = 1, 2) and **two** measured responses (i = 1, 2): Each dose-response data is fitted with the operational model:

$$y_{ij} = E_i \frac{\tau_{ij}^{n_i}[L]^{n_i}}{([L] + Ka_{ij})^{n_i} + \tau_{ij}^{n_i}[L]^{n_i}}.$$

Bias quantification : with the operational model

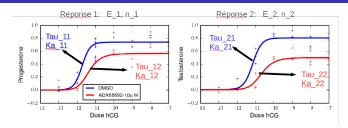


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For a given response i, we calculate $\Delta_i \log(\tau/Ka) = \log(\tau_{i2}/Ka_{i2}) - \log(\tau_{i1}/Ka_{i1})$.

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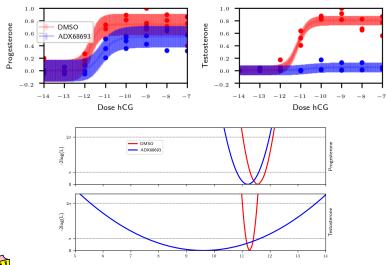
$$y_{ij} = E_i \frac{\tau_{ij}^{n_i}[L]^{n_i}}{([L] + Ka_{ij})^{n_i} + \tau_{ij}^{n_i}[L]^{n_i}}.$$

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The **Bias** is then defined by

$$\Delta\Delta \log(au/ extsf{Ka}) = \Delta_2 \log(au/ extsf{Ka}) - \Delta_1 \log(au/ extsf{Ka})$$

Statistical consideration : parameter confidence interval and (un-)identifiability





Data2Dynamics: Raue A., et al. Bioinformatics (2015)

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Bias quantification - standard method : operational mode

Biased quantification using dynamical model

Time-dependent bias?

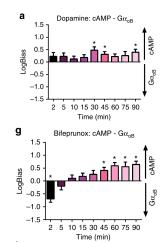
The role of kinetic context in apparent biased agonism at GPCRs

Carmen Klein Herenbrink¹, David A. Sykes², Prashant Donthamsetti^{3,4}, Meritxell Canals¹, Thomas Coudrat¹, Jeremy Shonberg⁵, Peter J. Scammells⁵, Ben Capuano⁵, Patrick M. Sexton¹, Steven J. Charlton², Jonathan A. Javitch^{3,4,6}, Arthur Christopoulos¹ & J Robert Lane¹

- Bias value may change according to the response time after stimulation.
- Kinetic explanation:
 Ligands with a slow binding kinetics may have changing bias value according to time.



Klein Herenbrink et al., *Nat. Commun* (2016)

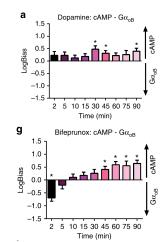


Time-dependent bias?

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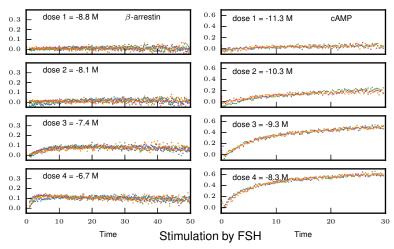
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- Bias value may change according to the response time after stimulation.
- Kinetic explanation:
 Ligands with a slow binding kinetics may have changing bias value according to time.
- ⇒ We need to take into account dynamic patterns in bias quantification



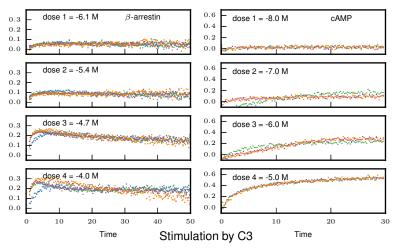
Dynamic data (on FHSR in HEK cells)

Instead of focusing on dose-response curves, we deal with **kinetic data** performed at several doses (here : induced BRET data)



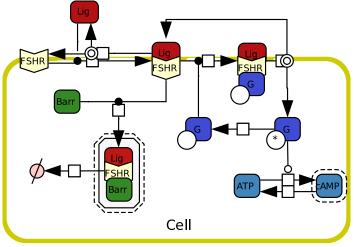
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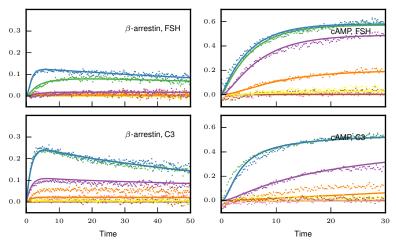
Principle of the methodology

I)We start with a sufficiently detailed chemical reaction network



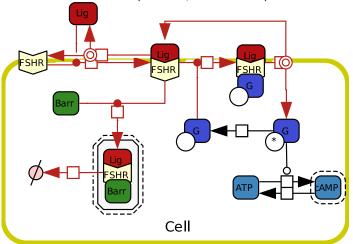
Principle of the methodology

I)We start with a sufficiently detailed chemical reaction network to accurately fit the data (one **separate** model for each Ligand)

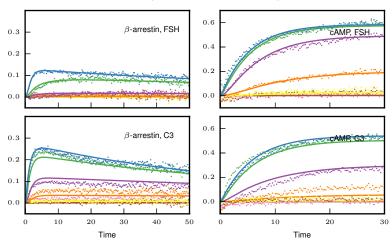


Principle of the methodology

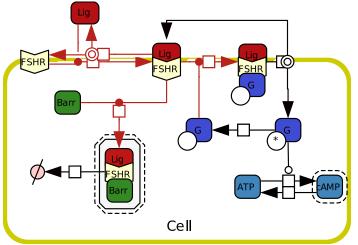
II) We fit **all data at once**, using some **common** parameters (initial concentration of molecules, measurement parameters...) and some **different** ones (kinetic parameters...)



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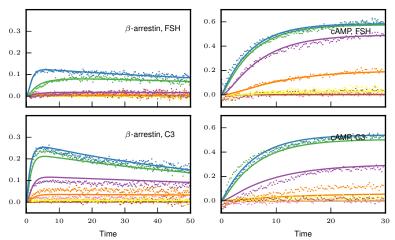
III) We use L^1 -penalization to find ligand specific parameters





Data2Dyanmics : Steiert, Timmer and Kreutz, *Bioinformatics* (2016)

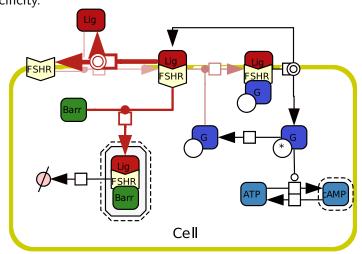
III) We use L^1 -penalization to find **ligand specific parameters**, keeping the fit 'as good as before'



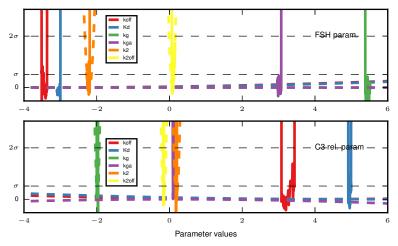


Steiert. Timmer and Kreutz. Bioinformatics (2016)

IV) After re-optimization, the set of distinct (ligand-specific) kinetic parameters gives us an accurate description of ligand specificity.

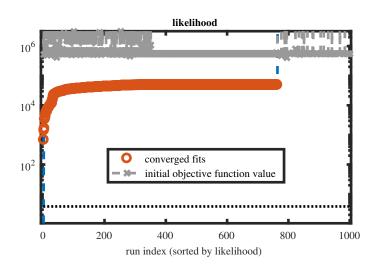


V) Significant differences between parameters is assessed by PLE

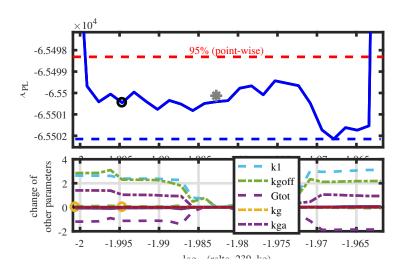


 \rightarrow here : C3 is biased towards β -arr, compared to cAMP, in comparison to FSH.

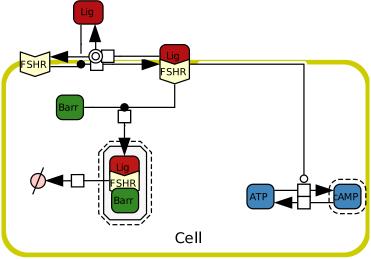
Practical problems...



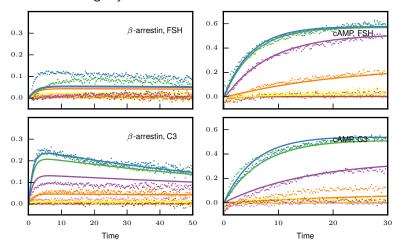
Practical problems...

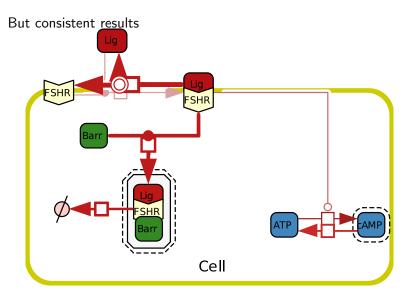


Kinetic model without G-protein

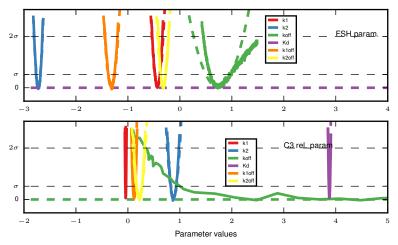


We obtain a slightly worse fit



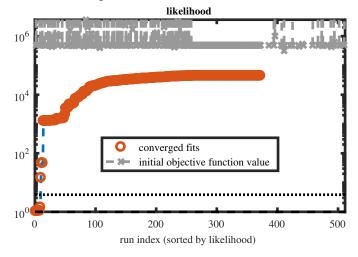


And "better" parameter identifiability

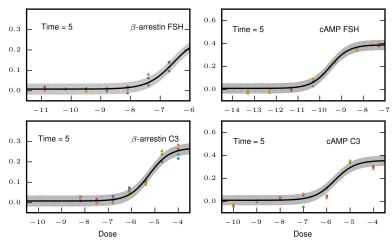


C3 is biased towards β -arr, compared to cAMP, in comparison to FSH.

And "better" convergence curves

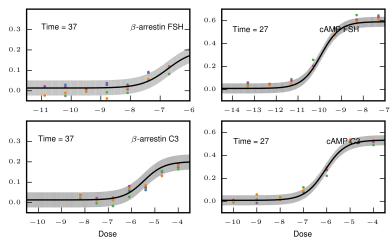


We systematically calculate bias value using standard method (operational model on dose-response curves :)



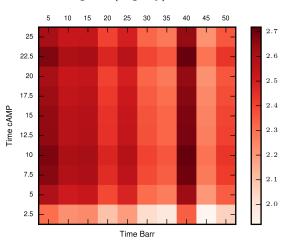
Bias=2.3 : C1 is biased towards β -arr, compared to cAMP, in comparison to FSH

We systematically calculate bias value using standard method (operational model on dose-response curves :)



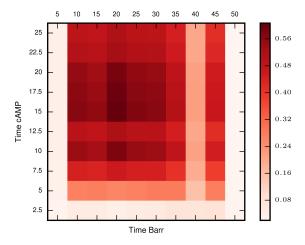
Bias=2.64 : C1 is biased towards β -arr, compared to cAMP, in comparison to FSH

We systematically calculate bias value using standard method Different times gives (slightly) different bias values



C1 is biased towards β -arr, compared to cAMP, in comparison to FSH

We systematically calculate bias value using standard method Uncertainty can be large according to the time of measurement



Summary

- Notion of signaling bias to quantify differential activation of several pathways by a Ligand at a given receptor.
- Standard quantification has several drawbacks (no time, limited to sigmoid scenario, et).
- We gave a kinetic interpretation of Ligand biased, which rely on dynamic (ODE) modeling and parameter estimation with L¹ penalization.

Summary

- Notion of signaling bias to quantify differential activation of several pathways by a Ligand at a given receptor.
- Standard quantification has several drawbacks (no time, limited to sigmoid scenario, et).
- We gave a kinetic interpretation of Ligand biased, which rely on dynamic (ODE) modeling and parameter estimation with L^1 penalization.
- ⇒ How to deal with "fuzzy/noisy" PLE / Densely sampled time data?
- ⇒ How to deal with non uniqueness of the penalized solution?
- ⇒ How to perform a model reduction that would lead to both a satisfactory fit and identifiable parameters?

Thanks for your attention!

Bios Team, PRC, INRA (Tours, Fr)

- * Eric Reiter
- ⋆ Pascale Crépieux
- * Anne Poupon
- * Francesco De Pascali

United Arab Emirates University

⋆ Mohammed Ayoub

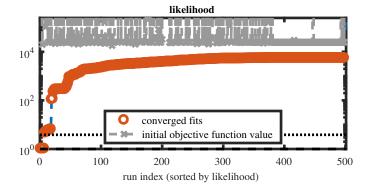


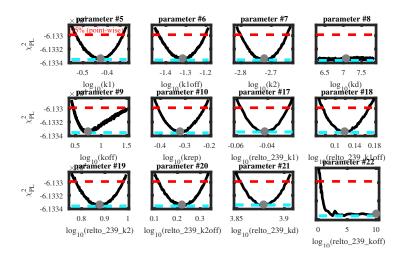
M. Ayoub et al., Molecular and Cellular Endocrinology 436 (2016)

L. Riccetti et al., Scientific Reports 7:940 (2017)

R.Y. et al., Methods in Molecular Biology, in press (2018)

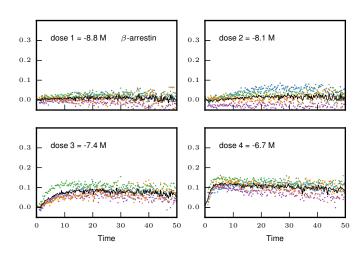






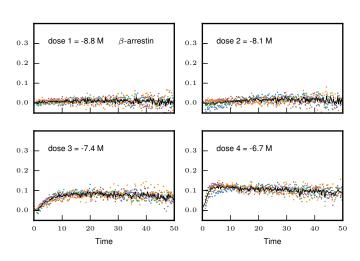
("trick" to minimize variance...)

Original "raw" data



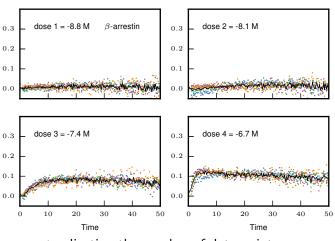
("trick" to minimize variance...)

"Adjusted" data



("trick" to minimize variance...)

"Adjusted" data



 $\boldsymbol{+}$ adjusting the number of data points \dots

Other extensions

Dose-dependent bias



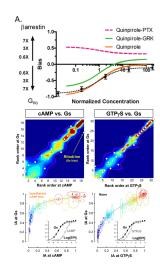
Barak and Peterson et al., Biochem. (2012) Extension of the operational model

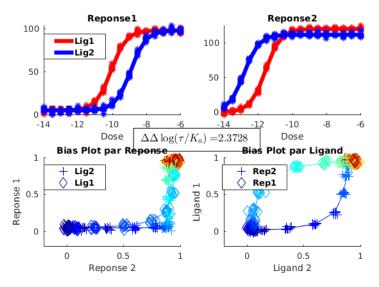


Kenakin, *Chem. Rev.* (2017) Method based on Intrinsic activities and rank ordering

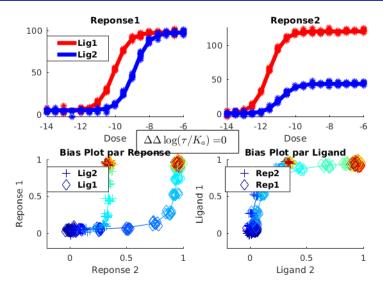


Onaran et al., *Sci. Rep.* (2017)

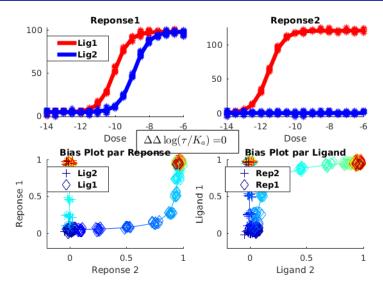




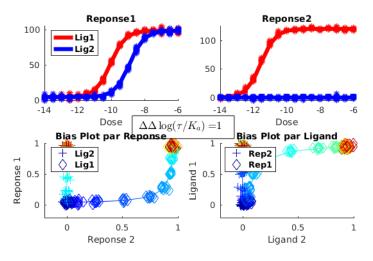
A strong bias is usually 'apparent' on dose-response curves or bias plot



But there may be counter-intuitive situation...



But there may be counter-intuitive situation...



But there may be counter-intuitive situation...

... and those situations occur in real life!

