



HAL
open science

Malacological survey in a bottle of water: a comparative study between manual sampling and environmental DNA metabarcoding approaches

Stephen Mulero, Eve Toulza, Anaïs Loisier, Meryl Zimmerman, Jean-François Allienne, Joséphine Foata, Yann Quilichini, Jean-Pierre Pointier, Olivier REY, Jérôme Boissier

► To cite this version:

Stephen Mulero, Eve Toulza, Anaïs Loisier, Meryl Zimmerman, Jean-François Allienne, et al.. Malacological survey in a bottle of water: a comparative study between manual sampling and environmental DNA metabarcoding approaches. *Global Ecology and Conservation*, 2021, 25, pp.e01428. 10.1016/j.gecco.2020.e01428 . hal-03106124

HAL Id: hal-03106124

<https://hal.science/hal-03106124v1>

Submitted on 11 Jan 2021

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

Running Head

Malacological monitoring by eDNA metabarcoding

Title

Malacological survey in a bottle of water: a comparative study between manual sampling and environmental DNA metabarcoding approaches

Author names and affiliations

Stephen Mulero^{1, *}, Eve Toulza¹, Anaïs Loisier¹, Meryl Zimmerman¹, Jean-François Allienne¹,
Joséphine Foata², Yann Quilichini², Jean-Pierre Pointier³, Olivier Rey^{1, □}, Jérôme Boissier^{1, □}

Contact Information

¹ University of Perpignan, IHPE UMR 5244, CNRS, IFREMER, Univ. Montpellier, F-66860 Perpignan, France.

² UMR SPE 6134, CNRS – Università di Corsica Pascal Paoli, 20250 Corte, Corsica, France.

³ PSL University, USR3278 CRIOBE EPHE-CNRS-UPVD, F-66860 Perpignan, France.

□ Both authors contributed equally to the study

Corresponding author

* Correspondence – stephen.mulero@outlook.com

Université de Perpignan Via Domitia, 52 Avenue Paul Alduy 66860 Perpignan cedex 9

E mail address

SM: stephen.mulero@outlook.com

ET: eve.toulza@univ-perp.fr

AL: anais.loisier@gmail.com

MZ: zimmermann.meryl@gmail.com

JFA: allienne@univ-perp.fr

JF: foata_j@univ-corse.fr

YQ: quilichini_y@univ-corse.fr

JPP: pointier@univ-perp.fr

OR: olivier.rey@univ-perp.fr

JB: boissier@univ-perp.fr

ABSTRACT

To assess the effect of anthropogenic activities on ecosystems, it is of prime importance to develop new tools enabling a rapid characterization of ecological communities. Freshwater ecosystems are particularly impacted and threatened by human activities and need thorough attention to preserve their biodiversity and the ecological services they provide. Studying such ecosystems is generally difficult because the associated organisms are hard to sample and to monitor. We present a ready-to-use environmental metabarcoding protocol to characterize and monitor the freshwater gastropods communities from water samples. The efficiency of this new tool was compared to a classical malacological survey at 19 sampled sites from 10 distinct rivers distributed over Corsica Island (France). From a single water sample, our eDNA monitoring tool provided a faithful characterization of the local malacofauna compared to the results obtained from the classical

malacological survey, with 97.1% of species detection confirmed by both methods. The present tool successfully detected the 11 freshwater snail species previously reported in Corsica by malacological survey but was limited at the genus level for some species. Moreover, our malacological survey allowed an update of the local distribution of a wide diversity of freshwater snails including invasive species (*i.e.* *Potamopyrgus antipodarum* and *Physa acuta*) as well as snail hosts of pathogens of medical and veterinary importance (*i.e.* *Bulinus truncatus* and *Galba truncatula*). These results strengthened a previous hypothesis of an eventual competitive interaction between *B. truncatus* and *P. antipodarum* that could limit the endemization of the urogenital bilharziasis in Corsica.

Keywords: biomonitoring, Corsica, environmental DNA, malacology, metabarcoding, snail communities

1. INTRODUCTION

Anthropogenic activities contribute to habitat fragmentation (Strayer and Dudgeon, 2010) or pollution (Blettler *et al.*, 2018) that have adverse effects on ecosystems (Parmesan and Yohe, 2003), including an important decrease of biodiversity (Waldron *et al.*, 2017). Freshwater ecosystems appear to be particularly threatened by human activities leading to local species extinction (Blettler *et al.*, 2018). According to the IUCN Red list, 46% of these ecosystems worldwide are endangered or vulnerable (Janssen *et al.*, 2016). These ecosystems host 10% of all known species despite representing only 0.8% of the Earth's surface (Balian *et al.*, 2008, Strayer

and Dudgeon, 2010). They also provide important ecological services for the development of human populations (Carpenter *et al.*, 2011).

Among freshwater species, gastropods have received particular attention for several reasons. First, they are suffering massive extinction notably due to habitat loss/degradation, the introduction of alien species and, in some cases, overexploitation (Bouchet *et al.*, 1999, Johnson *et al.*, 2013, Lydeard *et al.*, 2004, Strong *et al.*, 2007). Second, they play a key role in the functioning of most freshwater habitats. They are involved in the cycle of several biochemicals especially nitrogen (Hill and Griffiths, 2017) and are the prey of several fishes (Dillon, 2000b). Freshwater snails are also effective shredders and processors of vegetal matter in freshwater environment hence contributing to local matter and nutrient cycling (Dillon, 2000a). Finally, several freshwater gastropods are of medical or veterinary importance as they constitute intermediate hosts for several pathogens (*e.g. Trematoda*) (Lu *et al.*, 2018). Under the ongoing global changes, these diseases are currently (re-)emerging worldwide at an alarming rate (Jones *et al.*, 2008, Kincaid-Smith *et al.*, 2017). Hence, monitoring these snail species is crucial to address risk maps and thus prevent the transmission of these infectious diseases to nearby human and animal populations (Mulero *et al.*, 2020, Jones *et al.*, 2015).

Unfortunately, the monitoring of freshwater snails is challenging because: (i) there is no standardized monitoring protocol (Tallarico, 2016); (ii) their populations are spatially and temporally highly dynamic (Lamy *et al.*, 2012); and (iii) classical methods used to collect and to identify snail species are time-consuming, laborious and need malacological expertise (Vinarski and Kramarenko, 2015). In this context, the use of more sensitive molecular detection approaches such as environmental DNA (eDNA) are promising (Bohmann *et al.*, 2014).

According to Taberlet *et al.*, environmental DNA is a “complex mixture of genomic DNA from many different organisms found in an environmental sample” (Taberlet *et al.*, 2018, Taberlet *et al.*, 2012). This complex and heterogeneous community of DNA fragments originating from all surrounding organisms is either free in the environment or associated with organic or inorganic particles in water, soil or atmosphere (Pedersen *et al.*, 2015, Taberlet *et al.*, 2012). The use of eDNA approaches for monitoring targeted snail species already exist to detect invasive species such as *Potamopyrgus antipodarum* and *Crepidula fornicata* (Goldberg *et al.*, 2013, Miralles *et al.*, 2019, Clusa *et al.*, 2017); or gastropod species that transmit parasitic diseases such as *Galba truncatula*, *Austropelea tomentosa*, *Bulinus truncatus* and *Oncomelania hupensis* (Jones *et al.*, 2018, Mulero *et al.*, 2020, Fornillos *et al.*, 2019). In such a biodiversity assessment context, only one eDNA tool have very recently been developed to characterize communities of freshwater bivalves (Prié *et al.*, 2020). However, still is lacking a general eDNA-based tool to characterize the communities of freshwater gastropods.

To fill this gap, we here present the development of a ready-to-use eDNA-based diagnostic protocol for monitoring freshwater snail communities and its application in Corsica island. This study is constructed around four objectives (i) Developing a step-by-step environmental metabarcoding protocol. (ii) Comparing this protocol with a classical visual malacological survey approach in 19 sites from 10 rivers across Corsica Island (France). (iii) Providing an updated vision of the local malacofauna at the Island scale with focus on both invasive species and species with medical importance. (iv) Discussing the pros and cons of the emerging eDNA approaches compared to classical prospecting methods.

2. MATERIALS AND METHODS

2.1. Malacological survey and water sampling

The field work was conducted during August and September 2018 in the Corsica Island. This summer end period fulfil optimal conditions for the presence of freshwater snails populations that reach their highest density (Kincaid-Smith *et al.*, 2017). Overall, the sampling campaign consisting of both a water sampling for eDNA and a malacological survey were performed at 23 sites distributed over 13 different rivers (Figure 1). These sites are known for human recreational activities and are annually monitored by the French agency “Agence Régionale de Santé” de Corse (ARS, 2019).

2.1.1 Water sampling

Water samples for subsequent eDNA analyses were collected before the malacological survey and following a previously optimized protocol (Mulero *et al.*, 2020). At each sampling site, three liters of water were filtered in a single position from the river streambed just under the water surface. The water was collected downstream the prospected site to capture flowing eDNA from upstream transects, covering potentially the overall gastropod communities at a given site. Water filtrations were achieved using a sterile filtration unit with a 0.45 µm PES membrane of 90 mm diameter (VWR PES filter unit 1 L, model 514-0301) connected to a manual vacuum pump (Mityvac model MV8500). Afterwards, each filtration membrane was immediately stored in a 50 mL tube with a sterile Longmire buffer (Longmire *et al.*, 1997). Membranes were then stored in a dark cold room at 4°C until subsequent molecular analyses. During all the sampling process, precautions were taken to avoid any contamination (Taberlet *et al.*, 2012): operators used disposable sterile gloves and all reusable materials (*i.e.* scissors and forceps) were decontaminated

before and after each site by successively placing instruments in a 10% bleach bath, in a 90% ethanol bath and then in a DNA AWAY™ bath (Thermo Scientific) finalized by a flame sterilization (Taberlet *et al.*, 2012).

2.1.2 Malacological survey

At each site, the malacological survey was achieved along a total of 300 meters by the same operators. Three transects of 50 meters (downstream, at the referenced site and upstream) were prospected along the two shores of the river (3 x 2 transects). Within each transect, all freshwater snails were sampled manually or by scooping the grass on the water bench using a colander. Snails were then grouped according to their morphology and taxonomically identified at least at the genus, at best at the species level based on their morphological traits. The number of individuals of each species was semi-quantitatively estimated. If more than 100 snails were collected, we have attributed a “>100” score to the given species. At each site, GPS coordinates and several variables were collected including: the sampling day, the half day of sampling (*i.e.* A.M. or P.M.), water temperature & pH, water flow and altitude.

2.2. Molecular approaches

All pre-PCR molecular steps were performed under a sterile PCR hood decontaminated before and after each use as follow: the working surface was successively washed using 90% ethanol, 10% bleach and a DNA AWAY™ solution (Thermo Scientific™) and then exposed to UV light for 30 minutes. The reusable materials were decontaminated following the same protocol as for the field sampling and treated under UV exposure for 30 minutes.

2.2.1 DNA extractions

Total eDNA from each membrane was extracted using the DNeasy® Blood & Tissue kit (QIAGEN) following an adapted protocol (Mulero *et al.*, 2020). Briefly, membranes were subdivided into four equal parts to allow the use of 2 ml microcentrifuge tubes, these four parts were individually lysed at 65°C for one hour in a solution containing 567 µl of ATL buffers and 63 µl of proteinase K solution. Then a solution containing 630 µl of AL buffers and 630 µl of 100% ethanol was added in each tube. The remaining steps of the protocol were performed following the manufacturer instructions, except for that DNA was ultimately eluted in 100 µl (2 * 50 µl) of AE buffer pre-heated to 65°C. The resulting DNA extracts were then preserved at -20°C until subsequent steps.

In parallel, DNA was extracted from one individual of each of the 11 snail species collected during the malacological survey to produce a synthetic “mock community”. To this aim, we used the E.Z.N.A.® Tissue DNA kit (OMEGA bio-tek, Inc) following the “tissue protocol”. These snail DNA extractions were used as positive PCR and sequencing controls in the following steps.

To refine the molecular species assignments obtained from subsequent NGS analyses, we sequenced a 16S rRNA region for six snail species collected in Corsica. The 16Sbr-H primers were used following the PCR conditions described in (Saito *et al.*, 2018). The resulting amplicons contain the barcode region used for the metabarcoding. The PCR products were then sequenced on an ABI 3730xl sequencer at the GenoScreen platform (Lille, France). Sequences were obtained for *Ancylus fluviatilis* (MT361136), *Gyraulus* sp. (MT361134), *Physa acuta* (MT361133), *Pisidium casertanum* (MT361132), *Theodoxus fluviatilis* (MT361131) and *Galba truncatula* (MT361135).

2.2.2 PCR primers

The primer pair used in this study (Gast01) was previously developed and tested *in silico* (Taberlet *et al.*, 2018) but never tested and/or applied for empirical purposes. These primers target a 60-70 bp fragment of 16S mitochondrial rRNA using the Gast01F (5' CCGGTCTGAACTCAGATCA 3') and the Gast01R (5' TTTGTGACCTCGATGTTGGA 3') primer pair. Based on their *in silico* analyses, these primers theoretically amplify at least 1,280 species distributed among 456 genera, mainly related to *Gastropoda*. In our study, we first validated that these primers were able to produce a size-expected amplicon using qPCR for the 11 snail species collected during the malacological survey and eight additional snail species from previous field works (Table 1). These 19 DNA extracts were amplified by qPCR on a LightCycler®480 qPCR device following the same protocol as for the first PCR step of the library preparation, see 2.2.3 below.

2.2.3 Library preparation and MiSeq amplicon sequencing

Five negative controls and six positive controls were processed along with the environmental samples. Negative extraction controls consisted in ultrapure water samples processed following the same DNA extraction protocol as for the filtration membranes in each extraction runs (N = 2). PCR negative controls consisted in PCR reactions performed using water as template in each PCR run (N = 3). Positive controls consisted in two categories of mock communities. The first category consisted in an equimolar pool of 12 DNA extracts obtained from individuals of each snail species identified in Corsica and during previous field missions (Table 1). These mock communities are useful to detect PCR competition among different snail species. The second category of mock community consisted in pooling an equimolar quantity of PCR products individually obtained from 16 freshwater snail species (*i.e.* including the 11 identified snail species collected in Corsica

and five additional non-endemic species bred in our laboratory). These controls are useful to detect possible biases during the sequencing process.

Individual libraries were generated for each membrane section of the 23 filtration membranes and for each control, hence, representing a total of 103 libraries (23 x 4 = 92 eDNA samples + 11 controls). NGS libraries were prepared following the Illumina two-step PCR protocol, using mitochondrial 16S primers with Illumina adapters for the first locus-specific PCR. Both PCR were performed using the NEBNext[®] Ultra[™] II Q5[®] Master Mix M0544L (NEW ENGLAND BioLabs, USA). The first PCR reactions were performed in a final volume of 35 µl containing 3.5 µl of extracted DNA (concentrations ranging from 0.05 ng/µl to 8.52 ng/µl), 17.5 µl of 2X Q5 Master Mix and 14 µl of indexed primer mix (final concentration 0.4 µM) and ran on a Techne TC-PLUS thermal cycler PCR device (Techne, UK) using the following program: 98°C for 30 seconds followed by 40 cycles at 98°C for 10 seconds, 55°C for 20 seconds, 65°C for 8 seconds and a final elongation step of 2 minutes at 65°C. At this point, all PCR products were checked on a 2.5% agarose gel stained with ethidium bromide after electrophoresis.

The PCR products were individually indexed in the second PCR step consisting in eight cycles using the Nextera[™] XT Index (Illumina, San Diego, USA) following the manufacturer's instructions. Finally, the libraries were normalized using SequalPrep[™] plates (Thermo Fischer Scientific, USA) before pooling. The pooled libraries were then purified following the JetSeq[™] Clean protocol (Bioline, UK), checked on a Bioanalyzer High Sensitivity DNA kit (Agilent, USA) and quantified using a Qubit fluorometric quantification (Thermo Fisher Scientific, USA). Paired-end sequencing (2 x 250 cycles) was performed with a MiSeq Reagent Kit v2 on an Illumina MiSeq[™] instrument at the Bio-Environnement platform (University of Perpignan Via Domitia, France).

2.3. Data analyses

2.3.1 Bioinformatic pipeline

The resulting amplicon sequence dataset was processed using the Find Rapidly OTUs with Galaxy Solution (FROGS) pipeline implemented in Galaxy (Escudie *et al.*, 2018) at the Genotoul platform (Toulouse, France). (i) The amplicon dataset was first pre-processed, according to Gast01 primer specificities (*i.e.* amplicon size of 60 - 70 bp), we filtered out the sequences so as to keep amplicon sizes from 95 to 120 nucleotides (*i.e.* corresponding to the size of the targeted amplicon with Illumina adapters). (ii) The sequences kept were next clustered in operational taxonomic units (OTUs) using the swarm algorithm and using denoising and an aggregation distance of three nucleotides. (iii) The dataset was filtered out for chimeras using VSEARCH (Rognes *et al.*, 2016). Singletons and underrepresented clusters (*i.e.* <10 sequences) were removed as they correspond most likely to PCR error than rare sequences (Valentini *et al.*, 2016, Prié *et al.*, 2020). (iv) Each OTU was assigned to a species or taxon through a two-step MEGABLAST affiliation procedure. The first MEGABLAST analysis was restricted to *Mollusca* species. The second MEGABLAST analysis was performed without restricting parameters to check the robustness of the previous affiliations. The 10 best hits were kept for subsequent analysis. (v) The resulting OTUs were filtered following two criteria: first, only OTUs presenting a minimal blast coverage of 84% of amplicon length were kept; second, only OTUs presenting a pairwise identity above 89% with the affiliated sequence were kept, these thresholds are based on previously published protocols (Macher *et al.*, 2018) and on intraspecific sequences variations among snail strains. The remaining OTUs were considered as “unassigned”.

Lastly, to confirm that a given OTU was detected in a specific sample, we have adapted our validation criteria depending on OTUs abundances. An abundant OTU (*i.e.* ≥ 1000 copies considering all the samples) was considered present in a specific membrane when reaching at least 10 copies (overall the four membrane sections). Regarding poorly abundant OTU (*i.e.* < 1000 copies considering all the samples), we considered a sample positive if four or more copies of such OTU were distributed among two quarters of a same membrane.

3. RESULTS

3.1. Malacological survey

Overall, 11 distinct freshwater snail species were identified among the 23 sampled sites (Table 1-S1). These species were already registered in Corsica (INPN, 2020, IUCN, 2020) where 18 species of freshwater mollusk have been described so far, five of which inhabiting lentic ecosystems (Table S1) and (Mouthon, 1982, INPN, 2020). The species richness ranged from one (site 4) to six (site 23). The most common species was *P. antipodarum* (22/23 sites) and the rarest were *T. fluviatilis* (1/23 sites) and *Bithynia tentaculata* (3 individuals at site 13). Among the species identified, two are invasive (*i.e.* *P. antipodarum* and *P. acuta*) and two are intermediate hosts for Human and livestock parasites (*i.e.* *B. truncatus* and *G. truncatula*). Our semi-quantitative approach showed that *P. antipodarum* and *A. fluviatilis* were the most abundant species with 48.03 ± 46.38 and 26.68 ± 40.53 individuals, respectively (Table 1).

3.2. Molecular results

Over the 19 snail DNA extracts tested in qPCR with metabarcoding primers (Table 1), all but *P. casertanum* were positive. Regarding the equimolar pools of DNA, whatever pre- (Figure S2a) or post-PCR (Figure S2b) the sequencing of equimolar pools gave a number of reads for each species fitting to the expected ratio. This was higher evenness for post-PCR pools.

Despite all our precautions, four among the 23 eDNA water samples (from sites 10, 13, 18 and 19 – see Figure 1) were possibly contaminated and were thus discarded. In fact, amplicons corresponding of *A. fluviatilis*, *G. truncatula* and *B. truncatus* were found in the negative PCR controls associated with these samples. Hence, the subsequent metabarcoding data analysis was performed on the 19 remaining sites distributed over 10 rivers of Corsica.

Excluding the four contaminated sites, the whole sequencing (controls + field samples) generated more than 14 M of clusters with an average of 97,000 sequences per library. After analyses, 8.7 M of sequences were affiliated (Figure 2). These sequences were grouped among 488 OTUs with 43 OTUs corresponding to *Mollusca* species. After removing the sequences from control samples, on the 7,290,067 remaining sequences, 6% (445,638) were affiliated to *Mollusca*, including 424,580 sequences affiliated to *Gastropoda*; 10% (715,308) were unaffiliated and 84% (6,129,121) were affiliated to non-mollusk species (Figure 2). Whatever the taxonomic group, the OTU affiliations were generally limited to the genus and rarely reached the species level (*e.g.* only four out of 11 freshwater snail species found by metabarcoding were affiliated to a single species name). However, the 16S sequences obtained from snail DNA extracts collected in Corsica, allowed to recover the corresponding species name for major *Mollusca* OTUs. All the genus or species detected using eDNA corresponded to species already identified in Corsica and no new genus was detected compared to our malacological survey. Hence, considering all prospected sites,

we detected 61.1% (11/18) of historically known freshwater mollusks in Corsica and 84.6% (11/13) of species occurring in lotic systems (Table S1) and (INPN, 2020, Mouthon, 1982). The detected snail species belong to three subclasses of the *Gastropoda* (i.e. *Neritimorpha*, *Caenogastropoda*, and *Heterobranchia*) (Figure 3). The non-detected subclasses (e.g. *Vetigastropoda* and *Patellogastropoda*) are related to marine snail species. Beyond gastropods species, the present protocol allowed the detection of the bivalve *P. casertanum*.

3.3. Comparing the malacological survey to the eDNA monitoring

The malacological survey and the eDNA monitoring provided similar results (Figures 3 & 4). At the Island scale, all species identified during the malacological survey were also detected by eDNA monitoring. For some species such as the two *Gyraulus* species, the eDNA monitoring accuracy was limited to the genus level, one of these *Gyraulus* species has also not been affiliated by classical malacological survey either.

At the site level, the eDNA monitoring confirmed the detections obtained by malacological survey for 97.1% (67/69) of the detections. Only two occurrences of *A. fluviatilis* (site 17) and *Gyraulus* sp. (site 20) detected visually during the malacological survey, were not detected by eDNA monitoring (Figure 4). Conversely, the malacological survey confirmed the detections obtained by eDNA monitoring for 77% (67/87) of the detections (Figure 4). Moreover, eDNA monitoring detected on average 1.8 ± 2.84 more species per site than the malacological survey although this difference was not significant (Wilcoxon rank test; $W = 24$, $P = 0.11$). Based on the eDNA approach, *B. truncatus* was detected at 11 sites while only visually detected at 4 sites (Figures 4 & 5b). At the river scale, the results obtained from direct malacological survey and from the eDNA monitoring approach gave even more congruent results (Figure 3).

Among all the freshwater species detected in Corsica, two species are hosts for the transmission of trematodes of medical and veterinary importance (*i.e.* *B. truncatus*, Figure 5b and *G. truncatula*, Figure 5e) and two other are invasive species (*i.e.* *P. antipodarum*, Figure 5d and *P. acuta*, Figure 5f). The two invasive species display a wide distribution range (17 and 9 sites among the 19 monitored sites irrespective of the method) while the two species of medical/veterinary importance were mainly distributed in the Southern part of the Island. We were able to compare the current distributions of *B. truncatus* and *P. antipodarum* obtained within this study with those obtained in 1966, during previous malacological survey realized in Corsica for these two species (Doby *et al.*, 1966a). The distribution of *B. truncatus* has greatly diminished over the last 60 years and is currently limited to 4 rivers of the Porto-Vecchio region (*i.e.* Cavu, Solenzara, Osu and Tarcu; Berry *et al.*, 2014) for which we identified these species with the two approaches (Figure 5a & b). Contrarily, *P. antipodarum* has invaded most of the sites we have prospected and is now present on the east side of Corsica (Figure 5c & d).

4. DISCUSSION

One of the key steps for monitoring ecosystems and to quantify the effect of anthropogenic activities on biodiversity is to develop new tools enabling rapid and exhaustive characterization of the overall intrinsic species diversity. This is particularly true for freshwater ecosystems that undergo rapid changes and that are particularly threatened nowadays (Carpenter *et al.*, 2011). So far, the use of environmental DNA (eDNA) has been shown to be an efficient tool for monitoring fish (Hanfling *et al.*, 2016), arthropod (Krol *et al.*, 2019) and bivalve (Prié *et al.*, 2020) communities in freshwater environments. Here, we present the first application of an eDNA

monitoring method enabling the non-invasive characterization of entire gastropods communities from freshwater samples.

4.1. Benchmarking the eDNA monitoring

Our comparative approach between a classical (*i.e.* visual) prospection and an eDNA-based malacological survey highlights the ability of eDNA to characterize faithfully freshwater snail communities from water samples. Considering the 19 sites successfully diagnosed, 97.1% (67/69) of total occurrences reported by the malacological survey were confirmed using eDNA monitoring. A noticeable benefit of eDNA approaches compared to malacological survey is the less time-consuming sampling effort needed per site; only one sample of three liters realized by one operator (\approx 30 minutes from sampling to sample preservation) is needed (Mulero *et al.*, 2020). In comparison, the time needed to realize six measure units in classical malacological survey is much longer (>2 hours) depending on field practicability and experimenter skill level. However, whatever the visual taxonomical identification of specimen under binocular or the molecular lab experiment for metabarcoding are both time consuming activities. In the present study, we did not try to optimize the volume of water sampled. However, in the case of larger sampling campaigns it might be interesting to determine the lowest volume of water necessary to ensure the same diagnosis efficiency. We recently shown, using a targeted approach, that lower water volumes down to one liter allows detecting *B. truncatus* without losses in efficacy (Mulero *et al.*, 2020).

Using the eDNA monitoring approach, we tended to detect more species per site than with the malacological survey. Indeed, over the 87 detection events found using the eDNA monitoring, 77% of them (67) were corroborated by visual observations during the malacological survey. This apparent higher richness found by eDNA metabarcoding approach compared to classical survey was already reported in previous studies (Deiner *et al.*, 2017, Hanfling *et al.*, 2016, Valentini *et*

al., 2016, Prié *et al.*, 2020). Three non-exclusive hypotheses could explain the higher efficacy of eDNA compare to classical methods. First, DNA transportation through the river flow allows collecting eDNA from upstream the targeted site. In fact, in lotic systems, eDNA is known to disperse in a given way with waterflow (Hanfling *et al.*, 2016, Deiner *et al.*, 2016). Second, eDNA-based methods are expected to outcompete visual surveys for detecting cryptic or rare species (*e.g.* Dejean *et al.*, 2012). Cryptic freshwater mollusk species are particularly difficult to detect and previous studies estimated that about 20% of the species present at a single site are undetected after a single prospecting session during malacological surveys (Dubart *et al.*, 2019). Moreover, detecting biases are also common in malacology because populations of snails are highly dynamic in space and time (Lamy *et al.*, 2012). Interestingly, our eDNA approach has been particularly useful to detect *P. casertanum* compared to classical malacological survey, this species being known to be difficult to detect visually because of his preferred niche (*i.e.* under boulders) and its small size. Third, false positives constitute one major limitation of eDNA-based protocols (Cristescu and Hebert, 2018). These biases are generally related to either primer amplification biases or inappropriate field and/or laboratory protocols (Cristescu and Hebert, 2018). Regarding primer amplification biases, further applications of the current tool are needed to evaluate these errors considering that it is impossible to test the primers with the DNA of all known organisms *in vitro*. In our study, such biases are likely to be limited because our controlled mock communities were detected with no apparent amplification & sequencing biases between species. Concerning DNA contamination, the four sites that could have been contaminated were discarded. Moreover, the detections found using eDNA monitoring yet not confirmed by malacological survey, followed an ecologically realistic distribution. For *G. truncatula*, the two sites found positive only by eDNA monitoring were distributed on the Gravona River, downstream from a site where *G. truncatula*

was detected by the two approaches (*i.e.* site 5). The two other sites were distributed on the Rizzanese river in which *Lymnaeid* snails hosts of *F. hepatica* were previously identified (Gretillat, 1963). Noteworthy, this species prefers temporary water body, but could be found in river shore and associated trickle (Dreyfuss *et al.*, 2009), which might explain the observed relatively low densities.

Regarding *B. truncatus*, the detection differences are the most noticeable between the two approaches, the seven additional sites were mainly distributed in the Gravona River and nearby rivers. For each of these rivers, *B. truncatus* was previously identified (Doby *et al.*, 1966a). These non-randomly distributed detections in rivers where this species is historically known to be present support rather the non-detection of certain species by malacological survey, than false positive eDNA detection. However, a thorough field survey of these rivers is needed to validate our results.

4.2. Environmental DNA for characterizing communities

A recent study focused on freshwater bivalves has proposed an eDNA metabarcoding protocol for the characterization of these communities (Prié *et al.*, 2020). The main differences between our study and the latter one are the volume of water sampled (30 L vs 3 L) and the use of several primer combinations in the study of Prié *et al.* It is interesting to notice that both approaches tend to the same conclusions: eDNA detects more species than a classical malacological survey and the sensitivity of both protocols is similar. Indeed, in a single sampling, at their spatial scale Prié *et al.*, (2020) have detected 60% of the known bivalve community and we have found 61% of known gastropods in Corsica. Despite different protocols and different targeted groups this shows replicability but also the limitation of the eDNA approach. The present eDNA metabarcoding protocol allowed to qualitatively characterize the Corsican gastropod fauna at global scale. However, as shown in previous eDNA-based metabarcoding studies, the accuracy of such

characterization highly depends on the reliability of currently available databases together with sequence size that sometimes results in a limitation for the taxonomic affiliation to the genus taxonomic level especially for closely related species (Krol *et al.*, 2019, Deiner *et al.*, 2017). Here, we couldn't distinguish the two distinct species belonging to the *Gyraulus* genus identified visually. These species could also be difficult to differentiate morphologically, that could explain confusions with *Planorbis planorbis* on public NCBI databases. At the eDNA era, such limitations clearly call for the improvement and reinforcement of public databases aiming at characterizing biodiversity at the taxonomic and molecular scale (Henry *et al.*, 2008, Balint *et al.*, 2018). To this end, we submitted 6 new 16S rRNA sequences of freshwater snail species identified in Corsica. However, to overcome these limitations in the immediate future, Krol *et al.*, (2019) recently suggested to use a combination of barcode to optimize taxonomic assignation. Recent studies also called for the need of reinforcing the collection of metadata in eDNA studies that could influence the detection of eDNA (Nicholson *et al.*, 2020). In this way, we collected environmental variables at all sampling sites including some that are generally neglected (*e.g.* pH, waterflow, water temperature; Nicholson *et al.*, 2020). However, the sampling of a wider diversity of freshwater environment is still needed to accurately assess the effect of these variables on our ability to detect species.

4.3. Environmental DNA for the risk assessment of parasitic diseases

Beyond the technological development, our study provides an updated picture of the distribution of snails communities in Corsica. Among the 11 species detected, two snail species are of medical/veterinary importance. An updated distribution map for these two species is of prime interest for determining the risk of the associated waterborne diseases. In particular, in 2013 an outbreak of Schistosomiasis emerged in Corsica, this disease being caused by a trematode parasite

using *B. truncatus* as host (Boissier *et al.*, 2016, Ramalli *et al.*, 2018). *Bulinus truncatus* was widely distributed in Corsica in the middle of the 20th century and at this time some authors warned authorities about the potential for such a local emergence of Schistosomiasis (Doby *et al.*, 1966b, Doby *et al.*, 1966a). Today, this species distribution is limited in four rivers in Southern Corsica (Cavu, Solenzara, Tarcu and Osu; Berry *et al.*, 2014). Using our eDNA monitoring tool, we detected the presence of *B. truncatus* in the Gravona River, although possibly at low densities and/or upstream the sampled sites. This result call for an urging field validation of the local presence of this species to prevent potential transmission to Humans in this highly frequented river near Ajaccio, the main agglomeration of Corsica.

Fasciola hepatica is another trematode species already present in Corsica that uses *G. truncatula* as intermediate host (Oviedo *et al.*, 1996). This parasite is responsible for the zoonotic Fasciolosis disease generally infecting livestock and Humans (Valero *et al.*, 2002). The present study provides an up-to-date geographic distribution of *G. truncatula* in Corsica, hence, delineating pastures presenting a potential risk of transmission to livestock and Humans. Interestingly, the eDNA monitoring tool also detected the presence of *G. truncatula* in the Rizzanese River, a region in which lymnaeids snails hosts of *F. hepatica* were previously identified (Gretillat, 1963) although not locally detected by visual inspection in this study. In a sanitary context, these results also call for thorough monitoring of this species in this particular river.

4.4. Environmental DNA for studying bioinvasions

Invasive snail species needs attention as they can drastically disturb the receiving ecosystems and impact the ecology of indigenous species by altering biotic composition of invaded ecosystems (Cowie, 2001, Strayer, 2010). In this context, we have detected two invasive snail species: *Physa acuta* and *Potamopyrgus antipodarum*. *Physa acuta* originating from North America (Vinarski,

2017) has invaded Europe likely through the activity of aquarium keepers and maritime trade (Vinarski, 2017). *Physa acuta* was first reported in Corsica in 1977 (Holoyak, 2019). Today, we observe that this species has a wider distribution in Corsica than reported in public databases (Holoyak, 2019, GBIF, 2020).

Potamopyrgus antipodarum originates from New Zealand (Goldberg *et al.*, 2013) and has been introduced in Europe in 1859 probably *via* human activities such as fish farming (Alexandre da Silva *et al.*, 2019). This parthenogenetic snail is widely distributed in Corsica and needs attention considering the important density that these populations can reach (*e.g.* >400,000/m²; Hall *et al.*, 2003). This snail species is known to negatively impact nutrient cycling or can competitively exclude native species (Goldberg *et al.*, 2013). *Potamopyrgus antipodarum* was first identified in Corsica in 1961 and have rapidly invaded rivers of the south-west side of the island (Doby *et al.*, 1966a). Today, we observe that this species is widely distributed and abundant all around the island (Figure 5d). Interestingly, the researchers that carried out the previous malacological survey hypothesized that this species could competitively exclude other freshwater snails' such as snail intermediate hosts for parasites (Doby *et al.*, 1966a). In the same study, the authors observed that *B. truncatus* hosting either *Schistosoma bovis* (an animal parasite historically present in Corsica) or *S. haematobium* (a human parasite currently present in Corsica) was present all around the island. The considerable restriction of *B. truncatus* distribution in parallel with the increase distribution of *P. antipodarum* seems to agree with Doby's *et al.* predictions.

However, environmental metabarcoding approaches are currently limited for quantifying these interactions *in natura*, and more generally when they are applied for conservation, and parasitological risk assessment purposes because these study areas require not only presence/absence information but also abundance data (Yates *et al.*, 2020). The relationship

between eDNA and species abundance is subjected to unexplained variations (Yates *et al.*, 2020) mainly because eDNA is heterogeneously released in the environment (Jo *et al.*, 2019) and DNA persistence depend on several environmental biotic and/or abiotic factors (Barnes *et al.*, 2014, Taberlet *et al.*, 2018).

5. Conclusion

In view of these results, the eDNA monitoring using metabarcoding approaches appears particularly useful for biomonitoring studies. Here we presented the first application of eDNA metabarcoding protocol for the characterization of entire gastropods communities from water samples. This eDNA-based tool provides very similar results when compared to the classical malacological survey and is useful for the detection of habitat hosting whether invasive snail species or species of medical and veterinary importance, while requiring fewer sampling efforts than the visual prospecting. Beyond these qualitative data, our results have given more support to previous hypotheses about a potential competitive interaction between *B. truncatus* and *P. antipodarum*. Indeed, this interaction needs to be experimentally validated but could play an important role in the endemization process of the urogenital bilharziasis in Corsica. Hence, the environmental metabarcoding is particularly suited for a first analysis of environments for which data on malacofauna are scarce. In a regular use context, the temporality of records could enable an accurate characterization of local snail communities in space and time leading to the early detection of phenomena such as biological invasions, biodiversity losses, and hence, paving the way for complementary studies in other fields.

Authors Contributions

JB., OR. and SM. conceived the study and the sampling design. AL., MZ., JF., YQ and SM. carried out the malacological survey and the field sampling. SM., JFA. and ET. processed the samples and performed molecular experiments and amplicon sequencing. SM., JB., OR. and ET., analyzed and interpreted the data. JPP. provided the malacological expertise. SM., JB. and OR. drafted the manuscript. JB., OR., SM., ET., AL., MZ., JFA., JF., YQ. and JPP. revised the manuscript. All authors read and approved the final manuscript.

Acknowledgements

We thank the French agency “Agence Régionale de Santé de Corse” (ARS) for the data relative to the prospected sites. This study is set within the framework of the “Laboratoire d’Excellence (LABEX)” TULIP (ANR-10-LABX-41).

Funding

This study was funded by the Région Occitanie, the European “Fonds Européen de Développement Régional” (FEDER); the French Agency for Food, Environmental and Occupational Health & Safety (ANSES) [PNREST 2019/1/059 Molrisk]; and the French “Agence Nationale de la Recherche” (ANR) [Program HySWARM ANR-18-CE35-0001].

REFERENCES

- Alexandre Da Silva, M. V., Nunes Souza, J. V., Souza, J. R. B. & Vieira, L. M. 2019. Modelling species distributions to predict areas at risk of invasion by the exotic aquatic New Zealand mudsnail *Potamopyrgus antipodarum* (Gray 1843). *Freshwater Biology*, 64, 1504-1518.
- ARS. 2019. *Qualité des eaux de baignade* [Online]. French minister for Solidarity and Health: French minister for Solidarity and Health. Available: <http://baignades.sante.gouv.fr/baignades/navigMap.do?idCarte=fra#a> [Accessed November 28, 2019 2019].
- Balian, E. V., Segers, H., Lévêque, C. & Martens, K. 2008. The Freshwater Animal Diversity Assessment: an overview of the results. *Hydrobiologia*, 595, 627-637.
- Balint, M., Nowak, C., Marton, O., Pauls, S. U., Wittwer, C., Aramayo, J. L., Schulze, A., Chambert, T., Cocchiararo, B. & Jansen, M. 2018. Accuracy, limitations and cost efficiency of eDNA-based community survey in tropical frogs. *Mol Ecol Resour*, 18, 1415-1426.
- Barnes, M. A., Turner, C. R., Jerde, C. L., Renshaw, M. A., Chadderton, W. L. & Lodge, D. M. 2014. Environmental conditions influence eDNA persistence in aquatic systems. *Environ Sci Technol*, 48, 1819-27.
- Berry, A., Mone, H., Iriart, X., Mouahid, G., Aboo, O., Boissier, J., Fillaux, J., Cassaing, S., Debuisson, C., Valentin, A., Mitta, G., Theron, A. & Magnaval, J. F. 2014. *Schistosomiasis haematobium*, Corsica, France. *Emerg Infect Dis*, 20, 1595-7.

- Blettler, M. C. M., Abrial, E., Khan, F. R., Sivri, N. & Espinola, L. A. 2018. Freshwater plastic pollution: Recognizing research biases and identifying knowledge gaps. *Water Res*, 143, 416-424.
- Bohmann, K., Evans, A., Gilbert, M. T., Carvalho, G. R., Creer, S., Knapp, M., Yu, D. W. & De Bruyn, M. 2014. Environmental DNA for wildlife biology and biodiversity monitoring. *Trends Ecol Evol*, 29, 358-67.
- Boissier, J., Grech-Angelini, S., Webster, B. L., Allienne, J. F., Huyse, T., Mas-Coma, S., Toulza, E., Barre-Cardi, H., Rollinson, D., Kincaid-Smith, J., Oleaga, A., Galinier, R., Foata, J., Rognon, A., Berry, A., Mouahid, G., Henneron, R., Mone, H., Noel, H. & Mitta, G. 2016. Outbreak of urogenital schistosomiasis in Corsica (France): an epidemiological case study. *Lancet Infect Dis*, 16, 971-9.
- Bouchet, P., Falkner, G. & Seddon, M. B. 1999. Lists of protected land and freshwater molluscs in the Bern Convention and European Habitats Directive: are they relevant to conservation? *Biological Conservation*, 90, 21-31.
- Carpenter, S. R., Stanley, E. H. & Vander Zanden, M. J. 2011. State of the World's Freshwater Ecosystems: Physical, Chemical, and Biological Changes. *Annual Review of Environment and Resources*, 36, 75-99.
- Clusa, L., Miralles, L., Basanta, A., Escot, C. & Garcia-Vazquez, E. 2017. eDNA for detection of five highly invasive molluscs. A case study in urban rivers from the Iberian Peninsula. *PLoS One*, 12, e0188126.
- Cowie, R. 2001. Invertebrate invasions on Pacific Islands and the replacement of unique native faunas: a synthesis of the land and freshwater snails. *Biological Invasions*, 3, 119-136.

- Cristescu, M. E. & Hebert, P. D. N. 2018. Uses and Misuses of Environmental DNA in Biodiversity Science and Conservation. *Annual Review of Ecology, Evolution, and Systematics*, 49, 209-230.
- Deiner, K., Bik, H. M., Machler, E., Seymour, M., Lacoursiere-Roussel, A., Altermatt, F., Creer, S., Bista, I., Lodge, D. M., De Vere, N., Pfrender, M. E. & Bernatchez, L. 2017. Environmental DNA metabarcoding: Transforming how we survey animal and plant communities. *Mol Ecol*, 26, 5872-5895.
- Deiner, K., Fronhofer, E. A., Machler, E., Walser, J. C. & Altermatt, F. 2016. Environmental DNA reveals that rivers are conveyor belts of biodiversity information. *Nat Commun*, 7, 12544.
- Dejean, T., Valentini, A., Miquel, C., Taberlet, P., Bellemain, E. & Miaud, C. 2012. Improved detection of an alien invasive species through environmental DNA barcoding: the example of the American bullfrog *Lithobates catesbeianus*. *Journal of Applied Ecology*, 49, 953-959.
- Dillon, R. 2000a. Gastropod autecology. In: Dillon, R. T. (ed.) *The Ecology of Freshwater Molluscs*. Cambridge: Cambridge University Press.
- Dillon, R. T. 2000b. Predation. In: Dillon, R. T. (ed.) *The Ecology of Freshwater Molluscs*. Cambridge: Cambridge University Press.
- Doby, J. M., Rault, B., Deblock, S. & Chabaud, A. 1966a. Bullins et bilharzioses en Corse. *Annales de Parasitologie Humaine et Comparée*, 41, 337-349.
- Doby, J. M., Rault, B., Deblock, S. & Chabaud, A. 1966b. Snails and bilharziasis in Corsica. Distribution, frequency and biology of "*Bulinus truncatus*". *Ann Parasitol Hum Comp*, 41, 337-349.

- Dreyfuss, G., Vareille-Morel, V. & Rondelaud, D. 2009. Les habitats de *Lymnaea truncatula* Müller (Mollusque) le long de deux rivières. *Annales de Limnologie - International Journal of Limnology*, 33, 67-72.
- Dubart, M., Pantel, J. H., Pointier, J. P., Jarne, P. & David, P. 2019. Modeling competition, niche, and coexistence between an invasive and a native species in a two-species metapopulation. *Ecology*, 100, e02700.
- Escudie, F., Auer, L., Bernard, M., Mariadassou, M., Cauquil, L., Vidal, K., Maman, S., Hernandez-Raquet, G., Combes, S. & Pascal, G. 2018. FROGS: Find, Rapidly, OTUs with Galaxy Solution. *Bioinformatics*, 34, 1287-1294.
- Fornillos, R. J. C., Sato, M. O., Tabios, I. K. B., Sato, M., Leonardo, L. R., Chigusa, Y., Minamoto, T., Kikuchi, M., Legaspi, E. R. & Fontanilla, I. K. C. 2019. Detection of *Schistosoma japonicum* and *Oncomelania hupensis* quadrasi environmental DNA and its potential utility to schistosomiasis japonica surveillance in the Philippines. *PLoS One*, 14, e0224617.
- GBIF. 2020. *The Global Biodiversity Information Facility* [Online]. Available: <https://www.gbif.org/what-is-gbif> [Accessed November 26 2020].
- Goldberg, C. S., Sepulveda, A., Ray, A., Baumgardt, J. & Waits, L. P. 2013. Environmental DNA as a new method for early detection of New Zealand mudsnails (*Potamopyrgus antipodarum*). *Freshwater Science*, 32, 792-800.
- Gretillat, S. 1963. [Epidemiology of Certain Trematode Diseases of Domestic Animals in Corsica (Bovine Bilharziosis and Bovine and Ovine Distomiasis). Observations Conducted during a Mission Accomplished during the Autumn of 1962]. *Ann Parasitol Hum Comp*, 38, 471-81.

- Hall, R., Tank, J. and Dybdahl, M. 2003. Exotic snails dominate nitrogen and carbon cycling in a highly productive stream. *Frontiers in Ecology and the Environment*, 1, 407-411.
- Hanfing, B., Lawson Handley, L., Read, D. S., Hahn, C., Li, J., Nichols, P., Blackman, R. C., Oliver, A. & Winfield, I. J. 2016. Environmental DNA metabarcoding of lake fish communities reflects long-term data from established survey methods. *Mol Ecol*, 25, 3101-19.
- Henry, P.-Y., Lengyel, S., Nowicki, P., Julliard, R., Clobert, J., Čelik, T., Gruber, B., Schmeller, D. S., Babij, V. & Henle, K. 2008. Integrating ongoing biodiversity monitoring: potential benefits and methods. *Biodiversity and Conservation*, 17, 3357-3382.
- Hill, W. R. & Griffiths, N. A. 2017. Nitrogen processing by grazers in a headwater stream: riparian connections. *Freshwater Biology*, 62, 17-29.
- Holoyak, D. T. 2019. Inventaire National du Patrimoine Naturel (2019). Données de malacologie continentale transmises par D. T. Holoyak & M. B. Seddon. Version 1.1. UMS PatriNat (OFB-CNRS-MNHN), Paris. Occurrence dataset <https://doi.org/10.15468/wwdhh0> [Accessed via GBIF.org on November 26 2020].
- INPN. 2020. *National Inventory of Natural Heritage* [Online]. Muséum national d'Histoire naturelle. Available: <https://inpn.mnhn.fr> [Accessed February 11 2020].
- IUCN. 2020. *International Union for Conservation of Nature* [Online]. International Union for Conservation of Nature. Available: www.iucn.org [Accessed February 10 2020].
- Janssen, J. A. M., Rodwell, J. S., Garcia-Criado, S., Gubbay, S., Haynes, T., Nieto, A., Sanders, N., Landucci, F., Loidi, J., Ssymank, A. & Valachovič, M. 2016. European Red List of Habitats Part 2. Terrestrial and freshwater habitats. *European Publications Office*, 2.

- Jo, T., Murakami, H., Yamamoto, S., Masuda, R. & Minamoto, T. 2019. Effect of water temperature and fish biomass on environmental DNA shedding, degradation, and size distribution. *Ecol Evol*, 9, 1135-1146.
- Johnson, P. D., Bogan, A. E., Brown, K. M., Burkhead, N. M., Cordeiro, J. R., Garner, J. T., Hartfield, P. D., Lepitzki, D. A. W., Mackie, G. L., Pip, E., Tarpley, T. A., Tiemann, J. S., Whelan, N. V. & Strong, E. E. 2013. Conservation status of freshwater gastropods of Canada and the United States. *Fisheries*, 38, 247-282.
- Jones, K. E., Patel, N. G., Levy, M. A., Storeygard, A., Balk, D., Gittleman, J. L. & Daszak, P. 2008. Global trends in emerging infectious diseases. *Nature*, 451, 990-3.
- Jones, R. A., Brophy, P. M., Davis, C. N., Davies, T. E., Emberson, H., Rees Stevens, P. & Williams, H. W. 2018. Detection of *Galba truncatula*, *Fasciola hepatica* and *Calicophoron daubneyi* environmental DNA within water sources on pasture land, a future tool for fluke control? *Parasit Vectors*, 11, 342.
- Jones, R. A., Williams, H. W., Dalesman, S. & Brophy, P. M. 2015. Confirmation of *Galba truncatula* as an intermediate host snail for *Calicophoron daubneyi* in Great Britain, with evidence of alternative snail species hosting *Fasciola hepatica*. *Parasit Vectors*, 8, 656.
- Kincaid-Smith, J., Rey, O., Toulza, E., Berry, A. & Boissier, J. 2017. Emerging Schistosomiasis in Europe: A Need to Quantify the Risks. *Trends Parasitol*, 33, 600-609.
- Krol, L., Van Der Hoorn, B., Gorsich, E. E., Trimbos, K., Bodegom, P. M. V. & Schrama, M. 2019. How Does eDNA Compare to Traditional Trapping? Detecting Mosquito Communities in South-African Freshwater Ponds. *Frontiers in Ecology and Evolution*, 7, 260.

- Lamy, T., Pointier, J. P., Jarne, P. & David, P. 2012. Testing metapopulation dynamics using genetic, demographic and ecological data. *Mol Ecol*, 21, 1394-410.
- Longmire, J. L., Maltbie, M. & Baker, R. J. 1997. Use of "lysis buffer" in DNA isolation and its implication for museum collections. *Museum of Texas Tech University*. doi: <https://doi.org/10.5962/bhl.title.143318>.
- Lu, X. T., Gu, Q. Y., Limpanont, Y., Song, L. G., Wu, Z. D., Okanurak, K. & Lv, Z. Y. 2018. Snail-borne parasitic diseases: an update on global epidemiological distribution, transmission interruption and control methods. *Infect Dis Poverty*, 7, 28.
- Lydeard, C., Cowie, R. H., Ponder, W. F., Bogan, A. E., Bouchet, P., Clark, S. A., Cummings, K. S., Frest, T. J., Gargominy, O., Herbert, D. G., Hershler, R., Perez, K. E., Roth, B., Seddon, M., Strong, E. E. & Thompson, F. G. 2004. The global decline of nonmarine mollusks. *BioScience*, 54, 321-330.
- Macher, J. N., Vivancos, A., Piggott, J. J., Centeno, F. C., Matthaei, C. D. & Leese, F. 2018. Comparison of environmental DNA and bulk-sample metabarcoding using highly degenerate cytochrome c oxidase I primers. *Mol Ecol Resour*, 18, 1456-1468.
- Miralles, L., Parrondo, M., Hernandez De Rojas, A., Garcia-Vazquez, E. & Borrell, Y. J. 2019. Development and validation of eDNA markers for the detection of *Crepidula fornicata* in environmental samples. *Mar Pollut Bull*, 146, 827-830.
- Mouthon, J. 1982. Les mollusques dulcicoles - Données biologiques et écologiques - Clés de détermination des principaux genres de bivalves et de gastéropodes de France. *Bulletin Français de Pisciculture*, 1-27.

- Mulero, S., Boissier, J., Allienne, J., Quilichini, Y., Foata, J., Pointier, J. & Rey, O. 2020. Environmental DNA for detecting *Bulinus truncatus*: A new environmental surveillance tool for schistosomiasis emergence risk assessment. *Environmental DNA*, 2, 161-174.
- Nicholson, A., Mcisaac, D., Macdonald, C., Gec, P., Mason, B. E., Rein, W., ... Hanner, R. H. 2020. An analysis of metadata reporting in freshwater environmental DNA research calls for the development of best practice guidelines. *Environmental DNA*. doi: 10.1002/edn3.81.
- Oviedo, J. A., Bargues, M. D. & Mas-Coma, S. 1996. The intermediate snail host of *Fasciola hepatica* on the mediterranean island of Corsica. *A.P.E.*, 56, 217-220.
- Parnesan, C. & Yohe, G. 2003. A globally coherent fingerprint of climate change impacts across natural systems. *Nature*, 421, 37-42.
- Pedersen, M. W., Overballe-Petersen, S., Ermini, L., Sarkissian, C. D., Haile, J., Hellstrom, M., Spens, J., Thomsen, P. F., Bohmann, K., Cappellini, E., Schnell, I. B., Wales, N. A., Caroe, C., Campos, P. F., Schmidt, A. M., Gilbert, M. T., Hansen, A. J., Orlando, L. & Willerslev, E. 2015. Ancient and modern environmental DNA. *Philos Trans R Soc Lond B Biol Sci*, 370, 20130383.
- Prié, V., Valentini, A., Lopes-Lima, M., Froufe, E., Rocle, M., Poulet, N., Taberlet, P. & Dejean, T. 2020. Environmental DNA metabarcoding for freshwater bivalves biodiversity assessment: methods and results for the Western Palearctic (European sub-region). *Hydrobiologia*. doi: <https://doi.org/10.1007/s10750-020-04260-8>.
- Ramalli, L., Mulero, S., Noel, H., Chiappini, J. D., Vincent, J., Barre-Cardi, H., Malfait, P., Normand, G., Busato, F., Gendrin, V., Allienne, J. F., Fillaux, J., Boissier, J. & Berry, A.

2018. Persistence of schistosomal transmission linked to the Cavu river in southern Corsica since 2013. *Euro Surveill*, 23, 2-5.
- Rognes, T., Flouri, T., Nichols, B., Quince, C. & Mahe, F. 2016. VSEARCH: a versatile open source tool for metagenomics. *PeerJ*, 4, e2584.
- Saito, T., Hirano, T., Prozorova, L., Tu Do, V., Sulikowska-Drozd, A., Sitnikova, T., Surenkhorloo, P., Yamazaki, D., Morii, Y., Kameda, Y., Fukuda, H. & Chiba, S. 2018. Phylogeography of freshwater planorbid snails reveals diversification patterns in Eurasian continental islands. *BMC Evol Biol*, 18, 164.
- Strayer, D. L. 2010. Alien species in fresh waters: ecological effects, interactions with other stressors, and prospects for the future. *Freshwater Biology*, 55(suppl. 1), 152-174.
- Strayer, D. L. & Dudgeon, D. 2010. Freshwater biodiversity conservation: recent progress and future challenges. *Journal of the North American Benthological Society*, 29, 344-358.
- Strong, E. E., Gargominy, O., Ponder, W. F. & Bouchet, P. 2007. Global diversity of gastropods (*Gastropoda; Mollusca*) in freshwater. *Hydrobiologia*, 595, 149-166.
- Taberlet, P., Bonin, A., Zinger, L. & Coissac, E. 2018. *Environmental DNA: For biodiversity research and monitoring*, Oxford, UK, Oxford University Press.
- Taberlet, P., Coissac, E., Hajibabaei, M. & Rieseberg, L. H. 2012. Environmental DNA. *Mol Ecol*, 21, 1789-93.
- Tallarico, L. D. F. 2016. Freshwater Gastropods as a Tool for Ecotoxicology Assessments in Latin America*. *American Malacological Bulletin*, 33, 330-336.
- Valentini, A., Taberlet, P., Miaud, C., Civade, R., Herder, J., Thomsen, P. F., Bellemain, E., Besnard, A., Coissac, E., Boyer, F., Gaboriaud, C., Jean, P., Poulet, N., Roset, N., Copp,

- G. H., Geniez, P., Pont, D., Argillier, C., Baudoin, J. M., Peroux, T., Crivelli, A. J., Olivier, A., Acqueberge, M., Le Brun, M., Moller, P. R., Willerslev, E. & Dejean, T. 2016. Next-generation monitoring of aquatic biodiversity using environmental DNA metabarcoding. *Mol Ecol*, 25, 929-942.
- Valero, M. A., Panova, M., Comes, A. M., Fons, R. & Mas-Coma, S. 2002. Patterns in size and shedding of *Fasciola hepatica* eggs by naturally and experimentally infected murid rodents. *J Parasitol*, 88, 308-13.
- Vinarski, M. V. 2017. The history of an invasion: phases of the explosive spread of the physid snail *Physella acuta* through Europe, Transcaucasia and Central Asia. *Biological Invasions*, 19, 1299-1314.
- Vinarski, M. V. & Kramarenko, S. S. 2015. How does the discrepancies among taxonomists affect macroecological patterns? A case study of freshwater snails of Western Siberia. *Biodiversity and Conservation*, 24, 2079-2091.
- Waldron, A., Miller, D. C., Redding, D., Mooers, A., Kuhn, T. S., Nibbelink, N., Roberts, J. T., Tobias, J. A. & Gittleman, J. L. 2017. Reductions in global biodiversity loss predicted from conservation spending. *Nature*, 551, 364-367.
- Yates, M. C., Glaser, D. M., Post, J. R., Cristescu, M. E., Fraser, D. J. & Derry, A. M. 2020. The relationship between eDNA particle concentration and organism abundance in nature is strengthened by allometric scaling. *Mol Ecol*, 2020;00, 1-15.

Table 1. Snail species used for assessing the detection range of the Gast01 PCR primers *in vitro*.

Snail species	Origin	Date	Provided by	Average headcount per site
<i>Bulinus forskalii</i>	South-Africa	2017	Huyse T., KU Leuven, Belgium	–
<i>Bulinus globosus</i>	Senegal	2012	Huyse T., KU Leuven, Belgium	–
<i>Bulinus africanus</i>	South-Africa	2017	Huyse T., KU Leuven, Belgium	–
<i>Radix natalensis</i> ^{a,b}	South-Africa	2017	Huyse T., KU Leuven, Belgium	–
<i>Galba schirazensis</i> ^{a,b}	Cuba	2013	Hurtrez-Boussès S., IRD Montpellier, France	–
<i>Galba cubensis</i> ^{a,b}	Cuba	2013	Hurtrez-Boussès S., IRD Montpellier, France	–
<i>Pseudosuccinea columella</i> ^{a,b}	France	2011	Hurtrez-Boussès S., IRD Montpellier, France	–
<i>Biomphalaria glabrata</i> ^{a,b}	Brazil	2018		–
<i>Bulinus truncatus</i> ^{a,b}	Corsica, France	2018		2.28 (\pm 13.95) from 0 to >100
<i>Bithynia tentaculata</i> ^b	Corsica, France	2018		0.02 (\pm 0.28) from 0 to 3
<i>Radix balthica</i> ^b	Corsica, France	2018		0.11 (\pm 1.21) from 0 to 13
<i>Ancylus fluviatilis</i> ^{a,b}	Corsica, France	2018		26.68 (\pm 40.53) from 0 to >100
<i>Galba truncatula</i> ^b	Corsica, France	2018		0.06 (\pm 0.38) from 0 to 3
<i>Gyraulus laevis</i> ^{a,b}	Corsica, France	2018		0.06 (\pm 0.34) from 0 to 3
<i>Gyraulus</i> sp. ^{a,b}	Corsica, France	2018		2.99 (\pm 8.66) from 0 to 50
<i>Physa acuta</i> ^{a,b}	Corsica, France	2018		17.38 (\pm 34.29) from 0 to >100
<i>Potamopyrgus antipodarum</i> ^{a,b}	Corsica, France	2018		48.03 (\pm 46.38) from 0 to >100
<i>Pisidium casertanum</i> ^{a,b}	Corsica, France	2018		1.45 (\pm 7.10) from 0 to 50
<i>Theodoxus fluviatilis</i> ^b	Corsica, France	2018		0.87 (\pm 5.26) from 0 to 50

^aDNA extracts used for the preparation of equimolar pools of DNA (N = 12). ^bDNA extracts used for the preparation of equimolar pools of PCR products (N = 16). The displayed median headcount for each species reported by the malacological survey consider the six measure units conducted on each sites as separate values.



Figure 1. Map referencing the 23 sites prospected across Corsica Island for malacological survey and for water eDNA sampling. All these sites are lotic and distributed among 13 different rivers (darker lines). These sites are annually monitored by the French agency ARS, coordinates and environmental variables available upon request. The sites in black were sampled but removed from the analysis due to potential DNA contaminations (see text for details).

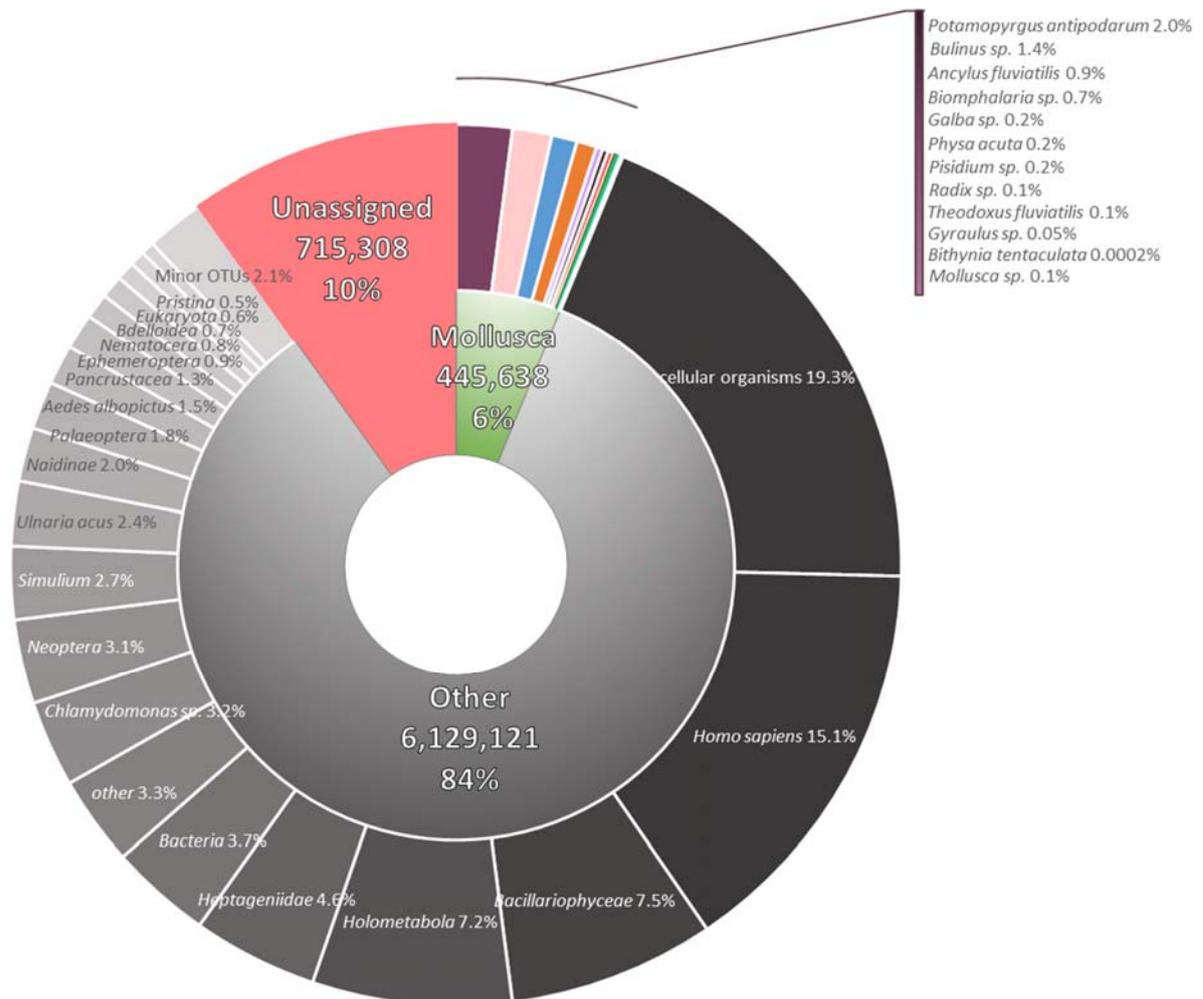


Figure 2. Major OTUs ranking based on sequence abundances. These sequences were obtained from eDNA samples collected in the field and do not contain those from experimental controls. Each taxon is based on merged OTUs (*i.e.* all OTUs affiliated to a same species are merged in one category).

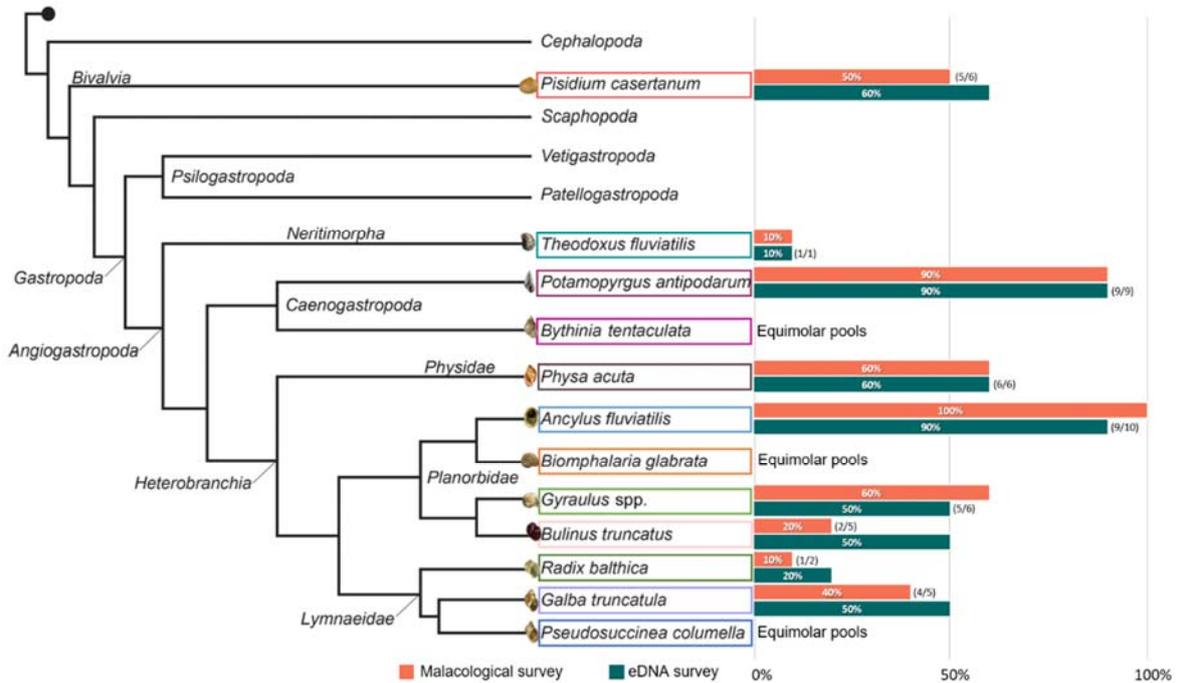


Figure 3. Simplified phylogeny of molluscs from Cunha & Giribet, (2019) highlighting all species reported in Corsica Island during the present study and species used to test primers (mock communities). Horizontal barplots represent species occurrences recorded at the river scale (N = 10) based on the malacological survey (in orange) and on the eDNA metabarcoding results (in blue). Numbers into brackets show the number of rivers positives for both methods on the total of positive rivers for the method showing the highest number of positive for a given species.

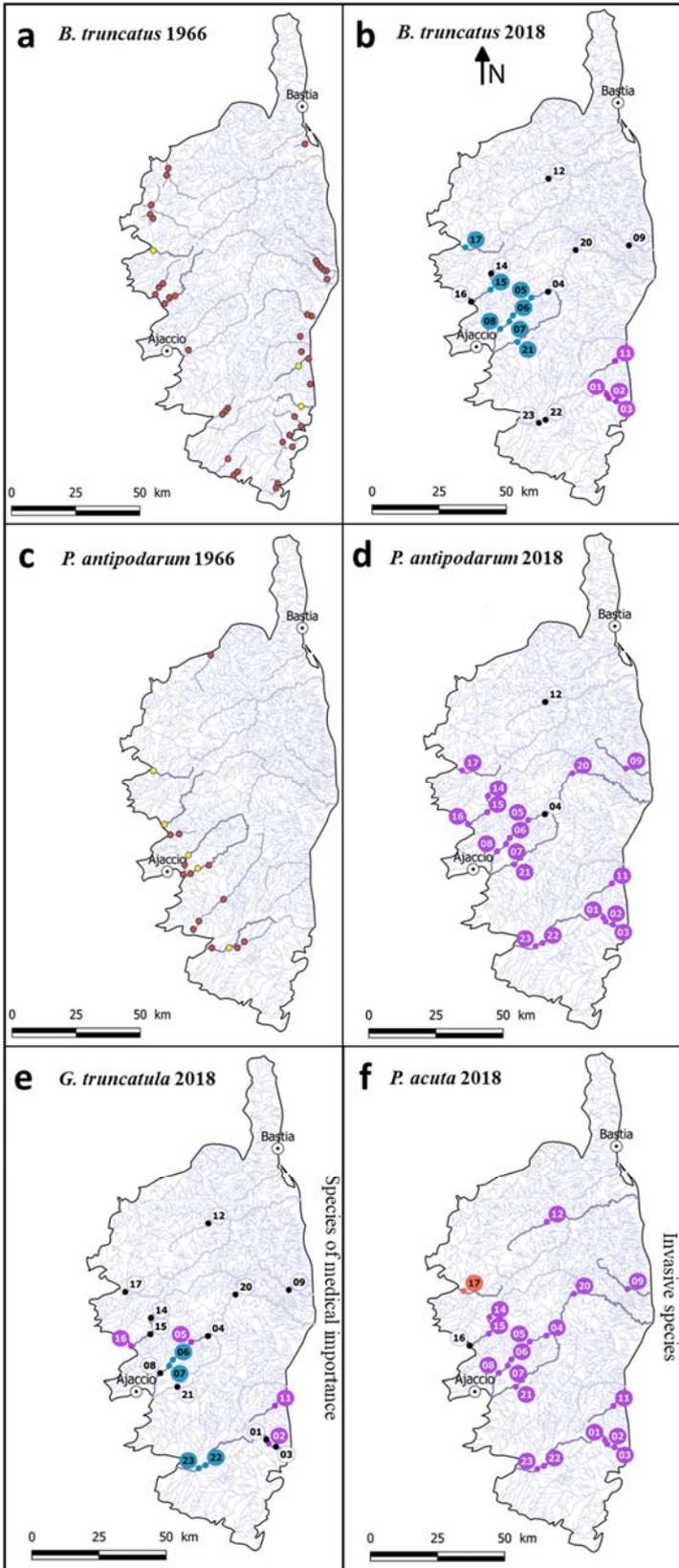


Figure 5. Maps displaying the current and past distributions of freshwater snail species of medical/veterinary and ecological interest, **a.** *Bulinus truncatus* in 1966, an intermediate host of the urogenital schistosomiasis infecting Humans; **b.** *Bulinus truncatus* in 2018; **c.** *Potamopyrgus antipodarum* in 1966, an invasive snail species from New Zealand; **d.** *Potamopyrgus antipodarum* in 2018; **e.** *Galba truncatula*, an intermediate host of the liver fluke infecting livestock and Humans and **f.** *Physa acuta*, an invasive snail species from North America. Black dots are sites where the given species is absent for the two monitoring methods; Purple dots are sites where the given species was detected with the two monitoring methods; Blue dots are sites where the given species was detected using the eDNA monitoring only; Orange dots are sites where the given species was detected using the malacological survey only; Yellow dots are sites where the given species was identified in our study in 2018 and in the study of Doby *et al.* in 1966.

Disclosures

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

Availability of data and materials

The dataset supporting the conclusions of this article is included within the article.

Table S1. Freshwater mollusks that were historically and presently detected in Corsica.

Species name	Habitat	Identified in sites	Average headcount per site
<i>Galba truncatula</i>	Lotic, Lentic	2, 5, 11, 16	0.06 (\pm 0.38) from 0 to 3
<i>Bulinus truncatus</i>	Lotic, Lentic	1, 2, 3, 11	2.28 (\pm 13.95) from 0 to >100
<i>Bithynia tentaculata</i>	Lotic, Lentic	13	0.02 (\pm 0.28) from 0 to 3
<i>Ancylus fluviatilis</i>	Lotic	All except 16	26.68 (\pm 40.53) from 0 to >100
<i>Potamopyrgus antipodarum</i>	Lotic, Lentic	All except 4 and 12	48.03 (\pm 46.38) from 0 to >100
<i>Physa acuta</i>	Lentic, Lotic	3, 6, 7, 8, 9, 15, 16, 21, 23	17.38 (\pm 34.29) from 0 to >100
<i>Pisidium casertanum</i>	Lotic, ditches, ponds	8, 11, 15, 21, 22, 23	1.45 (\pm 7.10) from 0 to 50
<i>Gyraulus laevis</i>	Lentic, lotic	20, 23	0.06 (\pm 0.34) from 0 to 3
<i>Gyraulus</i> sp.	Lentic, lotic	6, 7, 12, 14, 15, 17, 20, 23	2.99 (\pm 8.66) from 0 to 50
<i>Radix balthica</i>	Lentic, Lotic	1, 2	0.11 (\pm 1.21) from 0 to 13
<i>Theodoxus fluviatilis</i>	Lotic	23	0.87 (\pm 5.26) from 0 to 50
<i>Stagnicola palustris</i>	Ponds	–	
<i>Planorbis planorbis</i>	Ponds, lentic	–	
<i>Moitessieria corsica</i>	Groundwater	–	
<i>Sphaerium lacustre</i>	Lentic	–	
<i>Unio mancus</i>	Lotic	–	
<i>Acroloxus lacustris</i>	Lentic	–	
<i>Hippeutis complanatus</i>	Ponds, ditches	–	

These data are recovered from the work of Mouthon, 1982 and confirmed with the INPN 2020 database. Lentic = still water, Lotic = flowing water. The displayed median headcount for each species reported by the malacological survey consider the six measure units conducted on each sites as separate values.

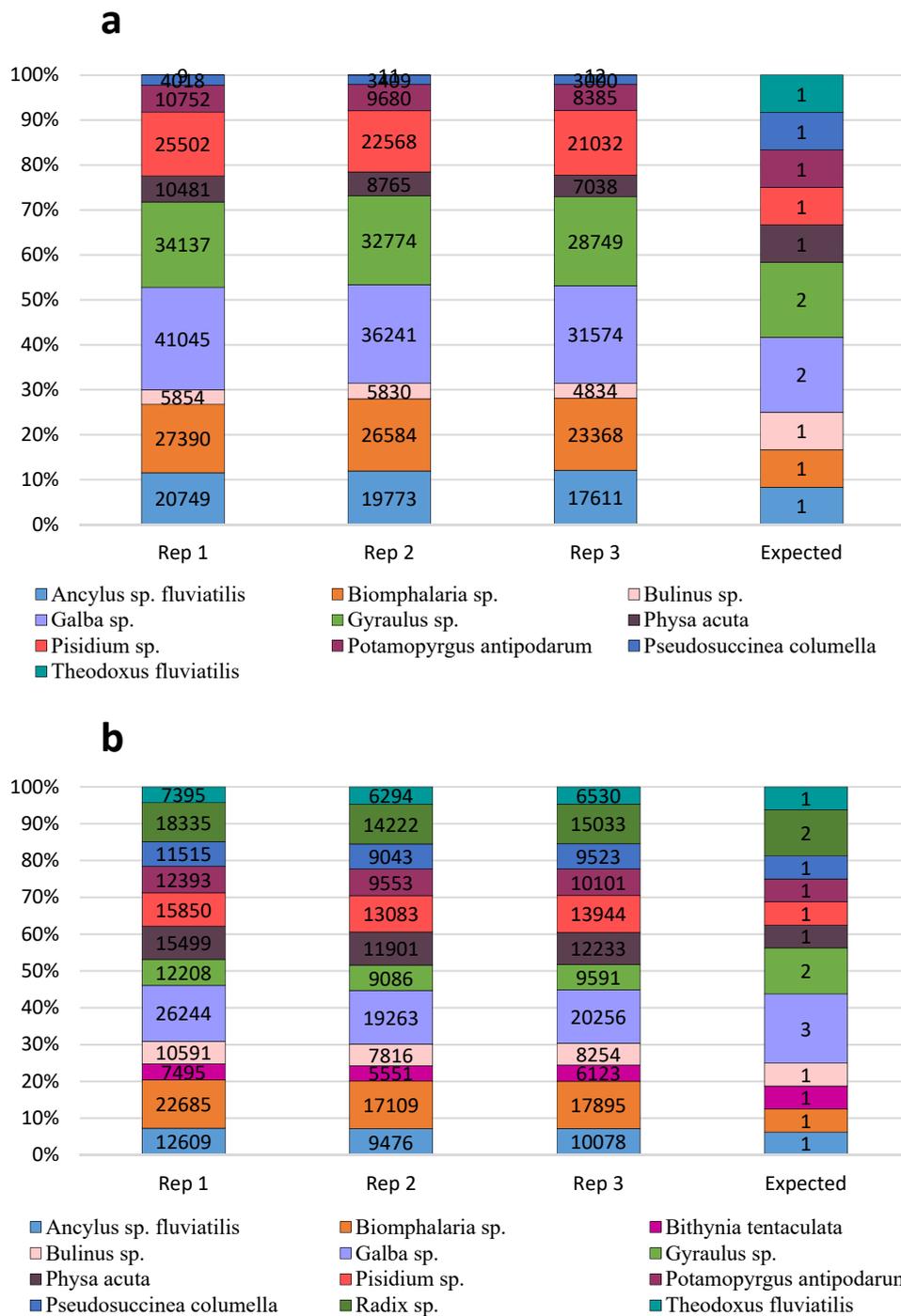


Figure S2. Number of sequences generated per species in equimolar controls, **a.** results for pooled DNA (N = 12) and **b.** results for pooled PCR products (N = 16). The first three columns are associated to the three technical replicate realized for each pool category, the fourth

column give the expected composition of the sample in number of distinct DNA ordered per genus.