



IRAP+ endosomes restrict TLR9 activation and signaling

Joël Babbdor, Delphyne Descamps, Aimé Cézaire Adiko, Mira Tohmé, Sophia Maschalidi, Irini Evnouchidou, Luiz Ricardo Vasconcellos, Mariacristina de Luca, Francois-Xavier Mauvais, Meriem Garfa-Traore, et al.

► To cite this version:

Joël Babbdor, Delphyne Descamps, Aimé Cézaire Adiko, Mira Tohmé, Sophia Maschalidi, et al.. IRAP+ endosomes restrict TLR9 activation and signaling. *Nature Immunology*, 2017, 18 (5), pp.509-518. 10.1038/ni.3711 . hal-03103680

HAL Id: hal-03103680

<https://hal.science/hal-03103680>

Submitted on 13 Sep 2021

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

IRAP endosomes restrict TLR9 activation and signaling

Joel Babbord^{1,2,3&}, Delphine Descamps^{1,2,3,4&}, Adiko Assi Aimé Cézairé^{9,10}, Mira Tohmé^{1,2,3,5},
Sophia Maschalidi^{1,2,3}, Irini Evnouchidou^{9,10}, Luiz Vasconcellos^{1,2,3}, Mariacristina De
Luca^{9,10}, Francois-Xavier Mauvais^{1,2,3}, Meriem Garfa-Traore^{1,2,3}, Melanie M. Brinkmann⁸,
Michel Chignard^{6,7}, Bénédicte Manoury^{1,2,3*}, Loredana Saveanu^{9,10*}

1. Institut National de la Santé et de la Recherche Médicale, Unité 1151, Paris, 75015, France
2. Centre National de la Recherche Scientifique, Unité 8253, Paris, 75015, France
3. Université Paris Descartes, Sorbonne Paris Cité, Faculté de médecine Paris Descartes, 75015 Paris, France
4. VIM, INRA, Université Paris-Saclay, 78350, Jouy-en-Josas, France.
5. Institut National de la Santé et de la Recherche Médicale, Unité 932, Institut Curie, 75005, Paris, France
6. Institut National de la Santé et de la Recherche Médicale, UMR S 938, CDR Saint-Antoine, Paris, France
7. Sorbonne Université, UPMC Univ Paris 06, UMR S 938, CDR Saint-Antoine, Paris, France
8. Helmholtz Centre for Infection Research, 38124 Braunschweig, Germany
9. Institut National de la Santé et de la Recherche Médicale, Unité UMR 1149, Centre de Recherche sur l'Inflammation, Paris, France
10. Université Paris Diderot, Faculté de Médecine Xavier Bichat, Paris, France

& These authors contributed equally to this work

* Co-senior authors

* Corresponding authors:

Loredana Saveanu, INSERM, Unité 1149, Faculté de Médecine Bichat, 16 rue Henri Huchard, 75018, Paris cedex 18, France
Bénédicte Manoury, INSERM, Unité 1151, Hôpital Necker-Enfants malades, 149 rue de Sèvres, 75743 Paris cedex 15, France
Emails: loredana.saveanu@inserm.fr, benedicte.manoury@inserm.fr

44 **Abstract**

45 Retention of intracellular Toll-Like Receptors (TLRs) in the endoplasmic reticulum prevents
46 their activation under basal conditions. TLR9, whose trafficking is still largely unknown, is
47 activated by sensing ligands in specific endosomal compartments. Here, we describe the
48 identification of IRAP (insulin responsive aminopeptidase) vesicles as major cellular
49 compartments for the early steps of TLR9 activation in dendritic cells (DCs). Both CpG and
50 TLR9 were found to be cargos of IRAP endosomes. In the absence of IRAP, CpG and TLR9
51 trafficking to lysosomes and TLR9 signaling were enhanced in DCs and in mice following
52 bacterial infection. IRAP stabilized CpG-containing endosomes by interacting with the actin
53 nucleation factor FHOD4, slowing down TLR9 activation in lysosomes. Thus, endosome
54 retention through IRAP interaction with the actin cytoskeleton is a mechanism that prevents
55 TLR9 hyper-activation in DCs.

56

57

58
 59 Innate and adaptive immune responses depend on the ability of toll-like receptors (TLRs) to
 60 discriminate between different classes of microbial products and initiate specific signaling
 61 cascades. While microbial products with no equivalent in mammalian cells, such as the
 62 components of the bacterial wall, are recognized by surface TLRs (1, 2, 4, 5 and 6), pathogen
 63 derived nucleic acids are sensed by intracellular TLRs (3, 7, 8 and 9). Recognition of nucleic
 64 acids by intracellular TLRs has an intrinsic potential to trigger autoimmune diseases through
 65 interaction with self nucleic acids ¹. To avoid inappropriate activation of endosomal TLRs,
 66 the trafficking of these receptors is tightly controlled. Thus, in basal conditions the receptors
 67 are located in the endoplasmic reticulum (ER) and translocate to endocytic vesicles only after
 68 cell stimulation by TLR ligands. Although all intracellular TLRs reside in the ER and require
 69 the chaperone Unc93b for their transfer to endosomes ^{2,3}, the trafficking pathways that move
 70 the receptors into the endocytic pathway show considerable variation among intracellular
 71 TLRs ⁴⁻⁶. For example, the TLR7/Unc93b complex traffics from Golgi stacks directly to
 72 endosomes using the clathrin adaptor AP4, while the TLR9/Unc93b complex is directed to
 73 the cell surface and reaches the endosomes via AP2-mediated clathrin-dependent endocytosis
 74 ⁶.
 75 In addition to the transfer into the endocytic pathway, a second step that controls the
 76 activation of endosomal TLRs is their partial proteolysis by an array of different proteases,
 77 specific for each TLR ^{5,7-12}.
 78 Although less often mentioned, the intracellular trafficking of its ligand also controls the
 79 activation of TLR9. TLR9 ligands (CpG) are internalized via clathrin-mediated endocytosis in
 80 early endosomes and translocate to late LAMP⁺ compartments ². TLR9 activation depends on
 81 CpG localization, since the abrogation of CpG translocation to LAMP⁺ vesicles by PIKfyve
 82 inhibitors decreased TLR9 signaling ^{13,14}. Thus, the intracellular trafficking of both, the ligand
 83 and the receptor are essential for the control of TLR9 activation.
 84 The complexity of TLR9 and CpG trafficking is rendered possible by the diversity and
 85 plasticity of the endocytic system. This system includes early endosomes that fuse to generate
 86 the sorting endosomes. From there, cargos are directed to different organelles, such as Rab4⁺
 87 fast recycling endosomes, Rab11⁺ slow recycling endosomes, the trans Golgi network (TGN)
 88 or lysosomes. Next to these universal routes of endosome trafficking, specialized cells, such
 89 as dendritic cells (DCs), display particular, albeit poorly characterized endosomal populations
 90 that affect TLR function, such as the VAMP3⁺ vesicles, which are involved in TLR9
 91 trafficking ¹⁵.

92 A particular and abundant endocytic population present in DCs, not yet investigated in the
 93 context of TLR signaling, is the slow recycling endosomes. They are characterized by the
 94 presence of the aminopeptidase IRAP (Insulin Responsive AminoPeptidase), a type II
 95 transmembrane protein composed of a catalytic site localized in the endosomal lumen and a
 96 cytosolic domain of 110 amino acids. We have previously demonstrated that in DCs, IRAP⁺
 97 vesicles are rapidly recruited to DC phagosomes, where the enzymatic activity of IRAP is
 98 involved in antigen processing during MHC-I cross presentation ^{16,17}.
 99 In addition to DCs, IRAP⁺ endosomes have been extensively studied in adipocytes, where they
 100 are called Glut4 storage vesicles (GSVs) and are rapidly transported to the cell surface under
 101 insulin stimulation. After the fusion of the endosomes with the cell membrane, Glut4 remains
 102 at the cell surface to facilitate glucose uptake, while IRAP is rapidly internalized ¹⁸. Thus,
 103 both in adipocytes and in DCs, IRAP displays a complex trafficking, sensitive to external
 104 regulation by insulin or phagocytic receptors activation. The regulated trafficking of IRAP
 105 depends on the cytosolic domain of the enzyme, which has been shown to interact with
 106 several proteins involved in vesicles formation or in cytoskeleton remodeling, such as the
 107 golgin p115 ¹⁹, vimentin ²⁰ and FHOS (formin homologue overexpressed in the spleen, also
 108 called FHOD1) ²¹. Whether these proteins and their interaction with the cytosolic domain of
 109 IRAP play a role in the complex trafficking of IRAP and IRAP⁺ endosomes is not known.
 110 Considering the dynamic and potentially regulated nature of IRAP⁺ endosomes, we wondered
 111 if IRAP plays a role in endosomal TLR trafficking and activation. We report here that the
 112 early step in TLR9 trafficking and CpG endocytosis requires IRAP. The absence of IRAP
 113 affects both CpG and TLR9 trafficking, leading to a dramatic increase in TLR9 signaling *in*
 114 *vitro* and *in vivo* following TLR9 stimulation. These results can be explained by the central
 115 role of IRAP in anchoring TLR9 endosomes to the actin cytoskeleton, which would limit
 116 TLR9-driven inflammatory responses. These findings provide a mechanistic explanation to
 117 the link between IRAP mutations and autoimmune disorders implicating TLR9 ²² and identify
 118 new factors and cellular pathways involved in TLR9 activation.

119

120 **RESULTS**

121

122 **IRAP deletion increases TLR9 response**

123 To address the role of IRAP in TLRs signaling, wild type (wt) and IRAP-deficient (ko) bone
 124 marrow derived dendritic cells (BM-DCs) were stimulated with ligands specific for
 125 intracellular and membrane TLRs: polyIC for TLR3, **Imiquimod for TLR7**, CpG-B for TLR9

126 and LPS for TLR4, and the production of the pro-inflammatory cytokines IL-6, IL-12p40,
 127 TNF $\square\square\square\square\square\square\square\square\square\square$. While IRAP deletion did not affect TLR3- and TLR4-dependent
 128 pro-inflammatory cytokine production, it enhanced pro-inflammatory cytokine production
 129 driven by TLR9 (**Fig. 1a**) and TLR7 activation (**Supplementary Fig. 1a**). These results
 130 suggest that IRAP affects the NF- κ B pathway downstream of TLR9 and probably TLR7.
 131 Since type I IFN production depends on TLR9 signaling through IRF-7, we wondered
 132 whether IRAP deletion affected this cytokine. We measured IFN- β production by wt and
 133 IRAP-deficient BM-DCs stimulated with TLR3, 4 and 9 ligands (**Fig. 1b**). IRAP deletion
 134 significantly increased only TLR9-driven IFN- β production but not IFN- β production by
 135 TLR3 and TLR4. Thus, IRAP disturbed the amplitude of both pro-inflammatory cytokine and
 136 type I IFN production in a TLR9-dependent manner in BM-DCs.
 137 To address whether the hypersensitivity of IRAP-deficient cells to TLR9 ligands was
 138 restricted to BM-DCs, which correspond to monocyte-derived inflammatory DCs, we purified
 139 conventional DCs (cDCs) and plasmacytoid DCs (pDCs) from the spleen and stimulated them
 140 with a TLR9 agonist. When incubated with CpG, IRAP-deficient spleen cDCs (**Fig. 1c**) and
 141 pDCs (**Fig. 1d**) produced significantly higher amounts of pro-inflammatory cytokines and
 142 INF- α than their wt counterparts. These results demonstrated that IRAP expression was
 143 required to prevent exacerbated inflammatory cytokine production in response to TLR9
 144 activation in all tested DC subsets.
 145 At least two mechanisms might account for the increased TLR9 response in IRAP-deficient
 146 DCs. IRAP⁺ vesicles, which are storage compartments sensitive to regulation by cell-specific
 147 stimulation in adipocytes, could store pro-inflammatory cytokines and control their trafficking
 148 and secretion. Alternatively, IRAP⁺ endosomes could directly influence TLR9 or CpG
 149 trafficking. We thus analyzed the intracellular localization of pro-inflammatory cytokines in
 150 wt and IRAP-deficient cells. While both IL-6 and IL-12 could not be detected in unstimulated
 151 cells, incubation with CpG resulted in staining for IL-6 and IL-12 in intracellular structures
 152 with a morphology indicative of Golgi stacks but devoid of IRAP staining (**Supplementary**
 153 **Fig. 1b**). The staining for the cis-Golgi matrix protein GM130, consistent with previous
 154 reports, confirmed IL-6 localization in Golgi stacks (**Supplementary Fig. 1c**)²³.
 155 Thus, IRAP⁺ endosomes are unlikely to be implicated in trafficking or secretion of IL-6 or IL-
 156 12(p40) and could be involved in a step upstream their synthesis. In support to this
 157 hypothesis, we found that DCs lacking IRAP expressed significantly higher levels of pro-
 158 inflammatory cytokine mRNAs than wt DCs upon CpG but not LPS stimulation (**Fig. 1e-f**).

159 The increase in cytokines, both at the mRNA and protein level, in IRAP deficient cells
160 following TLR9 stimulation should be correlated with an enhanced TLR9 signaling. To
161 specifically investigate TLR9 signaling, we tested the association of the MyD88 adaptor with
162 the transcription factors NF- κ B and IRF7. The proximity ligation assay (Duolink), which
163 detects protein complexes *in situ* ²⁴, demonstrated a significantly increased association of
164 MyD88 with NF- κ B (**Fig. 2a**) and of MyD88 with IRF7 (**Fig. 2b**) in IRAP-deficient cells, as
165 compared with wt cells.

166 As a consequence of TLRs activation, the mitogen-activated protein kinase ERK is rapidly
167 phosphorylated. Indeed, ERK phosphorylation was increased in IRAP deficient cells after
168 CpG stimulation, as detected by immunoblot (**Fig. 2c**). In addition, the last step of NF- κ B
169 activation, the phosphorylation of I κ B- α , was also significantly increased in IRAP-deficient
170 cells after CpG but not LPS treatment (**Fig. 2d**), indicating an enhanced TLR9 signaling in the
171 absence of IRAP.

172

173 **IRAP-deficient mice display a hyper-inflammatory phenotype driven by TLR9** 174 **activation**

175 Our *in vitro* data demonstrated the regulation of TLR9 signaling by IRAP. We next addressed
176 the question whether IRAP-deficient mice display increased TLR9 activation. To test this, we
177 measured the level of IL-6 in the serum of wt and IRAP-deficient mice 2 h after intravenous
178 injection of PBS, CpG-B or LPS. While wt and IRAP-deficient animals responded identically
179 to PBS and LPS injection, TLR9 stimulation led to higher levels of IL-6 in the serum of mice
180 lacking IRAP than in wt animals (**Fig. 3a**).

181 We wondered if the observed exacerbated pro-inflammatory TLR9 signaling might affect the
182 innate immune response during a bacterial infection. *Pseudomonas aeruginosa* (*P.*
183 *aeruginosa*) is an opportunistic Gram-negative bacterium that activates several TLRs
184 including TLR9 on alveolar macrophages (AM) and epithelial cells ²⁵. TLR9-deficient mice
185 were recently shown to be resistant to *P. aeruginosa* infection, suggesting that TLR9
186 signaling can have deleterious effects in this model ²⁶. In order to test whether the TLR9-
187 dependent hyper-activation observed in IRAP-deficient mice could affect survival upon
188 bacterial pulmonary infection, we intranasally inoculated IRAP-deficient and wt mice with
189 10⁶ cfu of *P. aeruginosa* and monitored them for survival. At least 36% of wt mice survived
190 during the two weeks of observation, while all IRAP-deficient mice died within 72 h after
191 infection (**Fig. 3b**). We then investigated the correlation between mice survival and the

inflammatory response monitored in the lungs 24 h post-infection. In infected IRAP-deficient mice, broncho-alveolar lavage (BAL) fluid contained higher levels of KC, IL-6, TNF- α and IL-1 β than in wt mice (**Fig. 3c**). Since AMs are the first innate immune cells to encounter bacteria in the lungs, we isolated AM and tested their cytokine production upon TLR9 and TLR4 stimulation *in vitro*. While a response was barely detectable in AM isolated from wt mice, IRAP-deficient AMs secreted substantial higher amounts of all pro-inflammatory cytokines tested upon CpG stimulation (**Fig. 3d**) in comparison to wt mice. In contrast, IRAP deficiency did not alter IL-6 and TNF- α secretion upon LPS stimulation, indicating that the hyper-inflammatory phenotype produced was restricted to TLR9 (**Fig. 3d**).

To control for a potential difference between the two mouse strains in their ability to clear bacteria from the lungs, we measured the pulmonary bacterial load and found it to be identical in IRAP-deficient and wt mice (**Fig. 3e**). Consistent with an identical ability of both strains to clear bacteria, *P. aeruginosa* infection led to a similar accumulation of neutrophils and macrophages/monocytes in the airways of both groups (**Supplementary Fig. 2a**). In addition, myeloperoxidase activity that mirrors neutrophil degranulation was similar in BALs from both wt and IRAP-deficient mice (**Supplementary Fig. 2b**). Altogether, these experiments suggest that following *P. aeruginosa* infection, IRAP-deficient mice died earlier probably because of an excessive inflammatory response driven by TLR9 hyper-stimulation.

210

211 **IRAP enzymatic activity is not involved in TLR9 activation**

Our results showed that proper regulation of TLR9 signaling required IRAP. Since IRAP is an aminopeptidase, we wondered if the enzymatic activity of IRAP was involved in the control of TLR9 activation. To investigate this possibility, we tested the effect of an inactive form of IRAP on TLR9 activation. IRAP, similar to all M1 aminopeptidases, has a Zn²⁺ atom in the active site²⁷ and contains the canonical zinc-binding amino acid motif HELAH, which is essential for the enzymatic activity. A form of IRAP in which the HELAH sequence was changed into H \underline{A} LAH (E465A substitution) co-localized, like the wild-type protein, with syntaxin 6 (Stx6), a SNARE of IRAP⁺ vesicles^{16,17} (**Fig. 4a**). Both the wt and mutated form of IRAP were well expressed, as shown by immunoblotting with anti-IRAP antibodies (**Fig. 4b**) but the mutated form was enzymatically inactive (**Fig. 4c**). When we reconstituted IRAP-deficient BM-DCs with wild-type IRAP (**Fig. 4d**) or enzymatically inactive IRAP (**Fig. 4e**), pro-inflammatory cytokine production upon CpG stimulation was similar to wt cells. These results demonstrated that the enzymatic activity of IRAP is not involved in the control of TLR9 activation.

226

227 **CpG and TLR9 are cargos of IRAP vesicles**

228 Since IRAP enzymatic activity was not involved in TLR9 activation, we wondered if IRAP
229 could interfere directly with TLR9 or CpG trafficking. Analysis of CpG-FITC by confocal
230 microscopy demonstrated that CpG massively colocalized with IRAP. After 20 min of
231 endocytosis, half of the internalized CpG was found in IRAP vesicles, where it was retained
232 for at least 1 h (**Fig. 5a**). Concomitant with IRAP-CpG colocalization, we observed a
233 significant increase of co-localization between IRAP and TLR9-GFP (**Fig. 5b**). While only
234 20% of TLR9 was found in IRAP vesicles early after CpG stimulation, the TLR9 ligand was a
235 major and persistent cargo of IRAP endosomes.

236 Since both TLR9 and its ligand trafficked via IRAP vesicles, we wondered if IRAP vesicles
237 overlap with VAMP3, a marker of an endosomal population through which TLR9 traffics
238 towards lysosomes¹⁵. IRAP, as well as the small GTPase Rab14 and the Q-SNARE Stx6, two
239 others markers of IRAP⁺ vesicles in DCs^{16,17} coincided with VAMP3 endosomes
240 (**Supplementary Fig. 3**). These results suggest that IRAP vesicles are a new intermediate
241 compartment between early endosomes and the final destination of TLR9, which is the
242 LAMP⁺ lysosome. This conclusion is also supported by the absence of colocalization between
243 IRAP and LAMP that we have shown previously^{16,17}.

244

245 **IRAP absence increases the susceptibility of TLR9 to lysosomal processing**

246 Since CpG and TLR9 are cargos of IRAP vesicles, we wondered if IRAP deletion could
247 change the trafficking of CpG and TLR9. Analysis of CpG-FITC trafficking demonstrated
248 that the proportion of TLR9 ligand transported to LAMP⁺ vesicles was significantly higher in
249 IRAP-deficient cells than in wt cells, an effect that was obvious at early time points (**Fig. 6a**).
250 The accelerated transport to lysosomes of CpG, in IRAP-deficient DCs, was potentially
251 correlated with a change in the intracellular distribution of TLR9-GFP, which was found in
252 lysosomes even in the absence of CpG stimulation (**Fig. 6b**). The presence of TLR9 in the
253 LAMP⁺ compartment correlates with proteolytic generation of a highly active C-terminal
254 fragment of the receptor in DCs and macrophages^{7,8,11,12,28}. In agreement with the lysosomal
255 localization of TLR9 in unstimulated IRAP-deficient cells, we found that in IRAP-deficient,
256 but not in wt primary mouse embryonic fibroblasts transfected with TLR9-GFP and
257 UNC93B-Cherry, the majority of immunoprecipitated TLR9-GFP corresponded to its
258 processed form (**Fig. 6c**). As expected from the functional assays of TLR stimulation (**Fig.**
259 **1a-b**), TLR3 did not colocalize with IRAP and intracellular localization of TLR3 was not

260 affected by IRAP deletion (**Supplementary Fig. 4**). Thus, in the absence of IRAP, TLR9, but
261 not TLR3, was targeted to lysosomes without cell stimulation.
262 To ensure that the lysosomal expression of TLR9 in IRAP-deficient cells was not the
263 consequence of TLR9-GFP expression by nucleofection, we investigated the localization of
264 endogenous TLR9. Since the only antibody that is specific for TLR9 (**Fig. 6d**) was not
265 sensitive enough to detect TLR9 in whole cell lysate, we isolated early and late phagosomes
266 from wt and IRAP-deficient DCs. While in the absence of CpG, TLR9 was not recruited to
267 phagosomes in wt cells, we detected the active C-terminal form of TLR9 in IRAP-deficient
268 late phagosomes. Thus, similar to the TLR9-GFP fusion, endogenous TLR9 was recruited to
269 phago-lysosomes without CpG treatment in the absence of IRAP. Intriguingly, when the cells
270 were stimulated with CpG, TLR9 recruitment to early and late phagosomes was identical
271 between wt and IRAP-deficient cells (**Fig. 6e**). However, despite the similar recruitment to
272 phago-lysosomes of endogenous TLR9 in both wt and IRAP-deficient cells upon CpG
273 treatment, TLR9 signaling was exacerbated only in IRAP-deficient cells. This apparent
274 contradiction can be explained by an increased accessibility of TLR9 to CpG in IRAP-
275 deficient cells, suggested by the accelerated translocation of internalized CpG-FITC to
276 lysosomes (**Fig. 6a**).

277

278 **IRAP deletion reduces CpG and TLR9 retention in early endosomes**

279 Since the consequences of IRAP on TLR9 hyper-activation could come from properties of
280 IRAP on early endosome trafficking, we wondered if the early steps of CpG and TLR9
281 trafficking were modified by IRAP depletion. To analyze this, we used the early endosomal
282 antigen, EEA1, a tethering factor known to be involved in homotypic and heterotypic fusion
283 events of early endosomes. EEA1 recruitment to endosome is a mandatory step in endosome
284 maturation to lysosome since inhibition of EEA1 activity blocks phagosome maturation ²⁹.
285 Nevertheless, a fraction of EEA1⁺ vesicles display a slow maturation rate and do not fuse
286 rapidly to late endosomes ³⁰. To visualize TLR9 and CpG trafficking in EEA1⁺ vesicles, we
287 used BM-DCs from TLR9-GFP transgenic mice pulsed with CpG-biotin (**Fig. 7a**) in which
288 IRAP was depleted by lentiviral shRNA (shIRAP) delivery (**Fig. 7d**). Colocalization analysis
289 between TLR9-GFP and EEA1 showed that in IRAP-depleted cells, and not in control cells
290 (shNT), TLR9 displayed a vesicular staining in subdomains of EEA1 endosomes in the
291 absence of CpG (**Fig. 7a**, upper panels). Following CpG incubation, the trafficking of CpG
292 was also affected by IRAP depletion. While in control cells, 25% of internalized CpG
293 remained in EEA1 endosomes for 2 h, in IRAP-depleted cells, CpG was rapidly transferred

from EEA1⁺ vesicles to TLR9⁺ vesicles, as illustrated by the white arrow in the **Fig. 7a**, middle panel. As a consequence of the rapid transfer to TLR9⁺ vesicles in IRAP-depleted cells, the colocalization between CpG and TLR9 was 3 times higher in comparison to control cells (**Fig. 7b**). This enhanced localization of CpG in TLR9 positive vesicles in cells lacking IRAP was not the consequence of an increased up-take of CpG by IRAP-depleted cells (**Fig. 7c**).

Thus, IRAP depletion facilitated not only TLR9 processing but also TLR9 access to its ligand.

IRAP interaction with the FHOD4 formin provides a molecular mechanism for the control of TLR9 activation

Altogether, these results highlighted a role for IRAP in TLR9 and CpG retention in EEA1⁺ endosomes that was independent on its enzymatic activity. We reasoned that the effect of IRAP on TLR9 activation could be mediated by interactions with proteins that play roles in vesicular trafficking. Two cytoskeleton factors have been previously identified to interact with the cytosolic domain of IRAP: vimentin ²⁰ and FHOD1 (formin homology domain-containing proteins; synonym: FHOS-formin homologue overexpressed in the spleen) ²¹. Vimentin forms intermediate filaments, cytoskeleton components that are important for anchoring intracellular organelles ³¹. FHOD formins are proteins essential for actin polymerization and are involved in anchoring vesicles to the actin cytoskeleton ³². Thus, vimentin and formins could play a role in the control of IRAP-mediated trafficking of TLR9. Duolink and co-immunoprecipitation experiments in fibroblasts and in DCs failed to show a robust interaction between IRAP and vimentin (data not shown), which implies that the IRAP-vimentin interaction that has been demonstrated in adipocytes ²⁰ might be specific to that cell type.

To investigate the FHOD1-IRAP interaction in DCs, we first investigated the expression of FHOD1 in different cell types as reported in the gene expression database of the ImmGen ³³ consortium (**Fig. 8a**). The mRNA expression data recovered from ImmGen (<https://www.immgen.org/Databrowserpage.swf>) showed that while FHOD1 expression is restricted to a subset of macrophages, FHOD4, a formin from the same family, has a wider distribution and higher expression levels in monocytes and DCs. As a consequence, we tested the interaction between IRAP and FHOD4. Endogenous FHOD4, as well as a FHOD4-GFP fusion protein interacted with IRAP as demonstrated by reciprocal co-immunoprecipitations (**Fig. 8b** and **Supplementary Fig. 5a**) and by Duolink, an alternative method used to

investigate protein interaction *in situ*²⁴ (**Fig. 8c** and **Supplementary Fig. 5b**). Confocal microscopy showed that FHOD4 could be recruited to IRAP⁺ vesicles, together with a vesicular actin coat labeled by phalloidin (**Fig. 8d**). These results suggested that FHOD4 could anchor IRAP⁺ vesicles to the actin cytoskeleton.

To investigate if FHOD4-IRAP interaction was involved in TLR9 activation in DCs, we knocked-down FHOD4 expression using lentiviral transduction of shRNA in BM-DCs from TLR9-GFP transgenic mice. FHOD4 was reproducibly reduced by 95% (\pm 10%) in the cells transduced with **two shRNA (17 and 20)** targeting FHOD4 (**Fig. 8e**). In the absence of FHOD4 and CpG stimulation, 40% (\pm 5%) of the endogenous TLR9-GFP was found in the cleaved form (**Fig. 8f**). In agreement with the increased basal processing of TLR9-GFP, the GFP-fused TLR9 was found in lysosomes in FHOD4 depleted cells (**Fig. 8g**). Thus, TLR9 trafficking and processing were affected by FHOD4 depletion, like in the case of IRAP deletion.

Considering the impact of FHOD4 depletion on TLR9 localization, we expected to have an increased TLR9-driven inflammatory response in FHOD4 depleted BM-DCs. When incubated with CpG, FHOD4 depleted cells (wt-shFHOD4) secreted significantly more pro-inflammatory cytokines than the cells transduced with a non-targeting shRNA (wt-shNT) (**Fig. 8h** and **Supplementary Fig. 6a**). **Depletion of FHOD1 did not affected TLR9-GFP localization or activation (Supplementary Fig. 6c, d), consistent with its very low levels of expression in BMDCs (Supplementary Fig. 6b).** These results suggest that IRAP anchors endosomal vesicles to the actin cytoskeleton through its specific interaction with FHOD4, slowing their transport to lysosomes. A major effect of these molecular interactions is ensuring a limited interaction between TLR9 and its ligand, which prevents hyper-inflammation.

DISCUSSION

The capacity of intracellular TLRs to recognize host nucleic acids is a risk for auto-immunity. For example, inappropriate activation of endosomal TLRs by self DNA has a major role in inflammation that occurs in systemic lupus erythematosus, arthritis and psoriasis³⁴. To avoid hyper-activation of TLRs, their encounter with the ligands and ability to signal must be tightly regulated. First, exposure to ligands is restricted through the retention of TLRs in the ER in basal conditions³⁵ and second, downstream signaling depends on the partial proteolysis of TLRs that occurs in the endo-lysosomal compartment^{5,7,8,12,36}. Although these two steps apply to all intracellular TLRs, recent in-depth studies show that the trafficking routes used by

363 distinct endosomal TLRs to reach the endocytic pathway are different ^{5,6}. TLR9, like other
 364 TLRs, exits ER by interacting with Unc93b ³⁵. Unlike other TLRs, the TLR9-Unc93b
 365 complex reaches the cell surface and is later internalized into a poorly characterized
 366 endosomal compartment through AP-2 mediated endocytosis ⁶.
 367 In this study, we identified IRAP as a regulator of CpG and TLR9 intracellular trafficking and
 368 *in vivo* activation of TLR9. IRAP deficiency led to rapid transport of internalized CpG to
 369 lysosomes and to TLR9 localization in lysosomes, where TLR9 is cleaved into its active C-ter
 370 form in the absence of CpG ligand. We observed this aberrant TLR9 trafficking and
 371 processing not only for TLR9-GFP, but also for endogenous TLR9 detected with anti-TLR9
 372 antibodies. Lysosomal localization of TLR9 in IRAP-deficient cells might be a consequence
 373 of accelerated trafficking of its ligand, which normally is retained in IRAP⁺ vesicles for a long
 374 time. The aberrant trafficking of both the ligand and the receptor led to an uncontrolled
 375 inflammatory response to TLR9 ligands, which culminated with animal death following an
 376 infection with *P. aeruginosa*, a bacterium sensed by TLR9 ²⁶. Altogether, our results show
 377 that IRAP is required to avoid excessive TLR9-driven inflammatory responses. In view of
 378 these results, it is conceivable that IRAP plays a role in human autoimmune pathologies
 379 through its effects on TLR9 signaling. The recent identification of a genetic association
 380 between psoriasis, one of the autoimmune disorders implicating TLR9 activation, and a
 381 nonsense mutation in the LNPEP gene encoding IRAP is consistent with this hypothesis ²².
 382 The new role of IRAP and our co-localization experiments define an endosomal compartment
 383 that is described by the presence of Rab14 and Stx6 (**Supplementary Fig. 7**) and partially
 384 overlaps with VAMP3⁺ and EEA1⁺ vesicles. VAMP3 and TLR9 have been shown to co-
 385 localize in an intermediate step of the route that TLR9 follows towards lysosomes and which
 386 depends on the AP-3 adaptor ¹⁵. Our data suggest that IRAP vesicles delay the trafficking of
 387 CpG and TLR9 from EEA1⁺ endosomes to lysosomes, with important functional
 388 consequences. To understand how IRAP could mechanistically affect the dynamics of the
 389 early endosomal compartment in which CpG and TLR9 are retained, we screen for
 390 cytoskeletal proteins that might interact with IRAP and could interfere with endosomal
 391 motility. Indeed, considering that the cytosolic tail of IRAP was shown to interact with two
 392 cytoskeleton components, an actin nucleation factor, the formin FHOD1 ²¹ and the
 393 intermediate filament vimentin ²⁰, we hypothesized that these interactions ensure the
 394 anchoring of IRAP and the associated TLR9 endosomes to cytoskeleton. Whereas the
 395 interaction of IRAP with vimentin was not detectable in DCs (experiments not shown), we
 396 found that IRAP binds to FHOD4 formin.

Formins are major actin nucleation factors that drive the assembly of actin monomers into filamentous structures and remain associated with the barbed end during filament elongation³⁷. A knock-down of FHOD4 had effects similar to IRAP deletion on TLR9 trafficking and the cellular response to CpG. These results suggest that by promoting actin assembly on endosomes, FHOD4 prevents the transfer of endosomes to microtubules, delaying their retrograde transport towards lysosomes, as reported for the formin mDial³⁸. Actin polymerization around the endosomal vesicles containing TLR9 ligands has been shown to be also driven by the other key actin nucleation factor, Arp2/3, and to be essential in limiting TLR9 signaling³⁹. Interestingly, both, FHOD4 and Arp2/3 are activated by the same small GTPase of Rho family, Cdc42³⁷, suggesting that these two actin-remodeling factors might cooperate in the regulation of TLR9 signaling, like they cooperate in phagocytic cup formation⁴⁰.

The intervention of FHOD4 in interaction with IRAP for modulating TLR9 function has major implications for potential links between extracellular stimuli, such as cytokines and the ability of TLR9 to respond to its ligands. It has been previously reported that TLR responses can be inhibited by extracellular stimuli such as cytokines⁴¹ or integrin ligation^{42,43}. Since Cdc42 activation occurs downstream integrin, receptor tyrosine kinase or G-protein-coupled receptors signaling⁴⁴, it could affect actin polymerization and the anchoring of CpG⁺/TLR9⁺ vesicles by IRAP. Thus, the anchoring of CpG⁺/TLR9⁺ vesicles to actin cytoskeleton could be essential in regulating TLR9 dependent cellular responses to the environment.

ACKNOWLEDGEMENTS

We are grateful to N. Goudin and S. Benadda for advice on analysis and quantification of confocal microscopy images, to S. Keller for anti-IRAP rabbit antibodies and mice, to D. Billadeau for anti-FHODs antibodies and to F. Benvenuto, F. Perez and S. Blystone for plasmid vectors. This work received financial support from the French “Agence Nationale de Recherche”, ANR IRAP-DC, ANR CytoEndoStor, ANR 2010 MIDI 008 01 and ANR post-doctoral support to D. D. We thank the Foundation “ARC pour la Recherche sur le cancer” for the acquisition of Leica SP8 confocal microscope and for the financial support to J.B.

428 **Figure Legends**

429 **Figure 1. IRAP deletion increases TLR9 response**

430 (a) Wt and IRAP-deficient (ko) BM-DCs were stimulated with different TLR ligands for 16 h
431 and the secretion of IL-6, IL-12(p40) and TNF- α in supernatants was measured by ELISA
432 (n=10 experiments, mean \pm SEM, * $p<0.05$, ** $p<0.01$). (b) Wt and ko BM-DCs were
433 incubated for 16 h with TLR ligands and IFN- β was measured by ELISA (n=3 experiments,
434 ** $p<0.01$). (c-d) Splenic cDCs (c) or pDCs (d) from wt and ko mice were isolated by cell
435 sorting (c) or anti-PDCA-1 magnetic beads (d), incubated overnight with CpG-B or CpG-A
436 and cytokine secretion was measured by ELISA (n=2 experiments, ** $p<0.001$). (e) Wt and ko
437 BM-DCs were incubated for 3 h with CpG-B or LPS and mRNA for TNF- α , IL-6 and IL-12
438 was quantified by RT-PCR using as reporters GAPDH and HPRT1. NS= non-stimulated
439 cells. (n= 4 experiments, data are represented as means \pm SEM, * $p<0.05$, ** $p<0.01$). (f)
440 mRNAs for IL-6 and IL-12 from wt and ko splenic pDCs, stimulated or not (NS) with CpG-
441 B, were measured by RT-PCR using the same reporter genes as in e (n= 3 experiments, means
442 \pm SEM, * $p<0.05$). See also **Supplementary Fig. 1**.

443

444 **Figure 2. Increased TLR9 signaling in IRAP-deficient DCs**

445 PLA for detection of MyD88/NF- κ B (a) or MyD88/IRF7 (b) proximity was performed with
446 specific antibodies against MyD88, NF- κ B and IRF7. The graphs show the quantification of
447 MyD88 interaction with NF- κ B (a) or IRF7 (b) by PLA. A minimum of 30 cells was
448 analyzed in each condition. (n=3 experiments, means \pm SD, ** $p<0.01$, *** $p<0.001$).
449 Statistical analysis was performed with Student t test. (c-d) Wt or ko BMDCs were incubated
450 with TLR ligands (CpG-B: 10 μ g/ml, LPS: 100 ng/ml) for 10, 20, 30, 40, 60 and 120 minutes.
451 Cells lysates were assessed for ERK (c) and I-KB- α (d) phosphorylation by Western Blot and
452 ELISA. Total ERK (t-ERK) and I-KB- α (t-I-KB- α) were also measured. Phosphorylated
453 ERK (p-ERK) proteins were quantified and normalized over total ERK (t-ERK) using
454 IMAGE J software. (c, d, n= 3 experiments, means \pm SEM, * $p<0.05$, ** $p<0.01$, ***
455 $p<0.001$, **** $p<0.0001$).

456

457 **Figure 3. IRAP-deficient mice display a hyper-inflammatory phenotype driven by TLR9** 458 **activation**

459 (a) IL-6 production was measured in serum of wt or IRAP-deficient (ko) mice 2 h after i.v.
460 injection of CpG-B (left panel) or LPS (right panel) (n=9 animals, mean \pm SEM, ** $p<0.01$).

(b) Wt and ko mice (n=9-11 in each group), were inoculated intranasally with *P. Aeruginosa* at 10^6 cfu/mouse. Animal survival was determined up to 7 days post-infection (n=3 independent experiments, * p<0.05). (c) Broncho-alveolar lavage (BAL) fluid levels of KC, IL-6, TNF- α and IL-1 β in wt and ko mice 24 h after intranasal inoculation of *P. Aeruginosa* (10^6 cfu/mice) (n=9 animals, 3 independent experiments, mean \pm SEM * p<0.05, ** p<0.01, *** p<0.001). (d) TNF- α , KC and IL-6 secretion in supernatants of non-stimulated (NS), CpG-B- or LPS-stimulated wt or ko alveolar macrophages (2 independent experiments, mean \pm SEM, * p<0.05, *** p<0.001). (e) Twenty-four hours post-infection, bacterial load was determined in lungs from wt and ko mice. (n=9 animals; graphs show mean \pm SEM of 2 independent experiments). See also **Supplementary Fig. 2**.

471

472 **Figure 4. IRAP enzymatic activity is not involved in TLR9 activation**

(a) IRAP-deficient (ko) BM-DCs reconstituted with active or inactive IRAP-HA by nucleofection were seeded on fibronectin-coated slides and stained with anti-Stx6 and anti-HA specific antibodies. **The graph shows the colocalization between IRAP and Stx6 (n=10 cells, two experiments).** (b) IRAP-deficient fibroblasts were transfected by electroporation with active or inactive IRAP and 36 h later, IRAP expression was analyzed by immunoblotting with anti-IRAP antibodies. (c) IRAP was immunoprecipitated with anti-IRAP antibodies from the fibroblasts transfected as in (b) and the aminopeptidase activity was tested by incubation of the beads with the colorimetric substrate Leu-AMC (2 independent experiments). (d-e) IRAP-deficient BM-DCs reconstituted with active (d) or inactive (e) IRAP were stimulated with TLR ligands for 16 h and IL-6 secretion was measured by ELISA (three (d) or two (e) independent experiments, mean \pm SEM, **p<0.01, ***p<0.001).

484

485 **Figure 5. CpG and TLR9 are cargos of IRAP endosomes**

(a) Wt BM-DCs were pulsed for 20 minutes with CpG-FITC, chased at 37°C for the indicated time-points, washed in PBS, fixed and stained with anti-IRAP and anti-FITC specific antibodies. (b) Wt BM-DCs were transfected with TLR9-GFP by nucleofection. Two days later, the cells were stimulated or not with CpG-B for 20 min, fixed and stained with anti-IRAP specific antibodies. See also **Supplementary Fig. 3**.

491

492 **Figure 6. IRAP absence increases the susceptibility of TLR9 to lysosomal processing**

(a) Wt and IRAP-deficient BM-DCs were pulsed for 20 minutes with CpG-FITC, chased at

37°C for the indicated time-points, washed in PBS, fixed and stained with anti-LAMP1 and anti-FITC specific antibodies (2 experiments, n=6 cells, *p<0.05, ***p<0.001). (b) IRAP-deficient and wt BM-DCs were transfected with TLR9-GFP by nucleofection and 48 h later stimulated or not with CpG-B for 20 or 120 min. The cells were fixed and stained with specific antibodies for LAMP1 (2 experiments, n=10 cells, *p<0.05, **p<0.01). (c) Wt and IRAP-deficient fibroblasts expressing TLR9-GFP and Unc93b-Cherry were lysed in 1% NP-40 and TLR9-GFP was immunoprecipitated with anti-GFP antibodies and analyzed by anti-GFP immunoblot. One experiment out of three is shown. The graph represents the quantification of GFP immunoblots from the three independent experiments. Phagosomes from wt, ko (e) and *tlr9*^{-/-} (TLR9-ko, d) BMDCs unstimulated or stimulated with CpG-B (10 µg/ml) were magnetically purified after 20 min or 120 min. Proteins expressed in phagosomes (10 µg) were resolved by SDS-PAGE and endogenous TLR9 and LAMP1 proteins were visualized by immunoblot. Data are representative of two experiments.

507

508 **Figure 7. IRAP deletion reduces CpG and TLR9 retention in early endosomes**

509 BM-DCs from TLR9-GFP transgenic mice were transduced with a lentivirus coding for a shRNA against IRAP (shIRAP) or a non-targeting shRNA (shNT). (a) The cells were pulsed or not with Biotinylated-CpG, washed and chased for the indicated time points. After fixation, the cells were stained with anti-EEA1 specific antibodies. (b) The graphs represent the percentage of colocalization between the proteins visualized in (a) (2 experiments, n=6 cells, *p<0.05, **p<0.01, ***p<0.001 ****p<0.0001). (c) CpG-FITC up-take by wt and IRAP-deficient BM-DCs was measured by flow cytometry. (d) The efficiency of IRAP knockdown was analyzed by immunoblotting, using anti-IRAP antibodies and anti α-Tubulin antibodies for the loading control.

518

519 **Figure 8. IRAP interaction with the FHOD family of formins controls TLR9 activation**

520 (a) IRAP (encoded by *Lnpep* gene), *FHOD1*, *FHOD4* and *TLR9* mRNA expression data were recovered from immgen (<https://www.immgen.org/Databrowserpage.swf>) for pDCs (pDCs_8+_sp sub-type), Monocytes (Mo_C6+_II+_Bl type) and Macrophages (MF_RP_Sp type). (b) Endogenous IRAP and FHOD4 were immunoprecipitated with antibodies anti-IRAP and anti-FHOD4 respectively and the precipitates were split in two and analyzed by immunoblot as indicated. (c) In situ IRAP/FHOD4 interaction was detected by Duolink assay in wt and IRAP-deficient (ko) BM-DCs using antibodies against IRAP and FHOD4 (2

independent experiments, 10 cells acquired). **(d)** Wt fibroblasts expressing FHOD4-GFP were fixed and stained with phalloidin and anti-IRAP antibodies. **40% (+/-5) of IRAP colocalized with FHOD4 (n=5 cells, 2 independent experiments).** **(e-g)** BM-DCs from TLR9-GFP transgenic mice were transduced with two lentiviruses coding for shRNAs against *FHOD4* (shFHOD4 17 and shFHOD4 20) or a non-targeting shRNA (shNT). **(e)** The efficiency of FHOD4 knock-down was analyzed by immunoblotting using anti FHOD4 antibodies. **(f)** Endogenous TLR9-GFP processing in cells transduced with shNT or shFHOD4 (20) was analyzed by immunoblot with anti-GFP antibodies. Control corresponds to untransduced wt BMDCs. **(g)** Endogenous TLR9-GFP localization in steady state conditions was analyzed by confocal microscopy using an anti-LAMP1 antibody. The graph represents the quantification of TLR9-GFP/LAMP1 colocalization (2 experiments, n=10 cells, ***p<0.001). **(h)** Wt and ko BMDCs were transduced with shNT (non-targeting) and shFHOD4 (20) lentiviruses and stimulated with different TLR ligands for 6 h. The secretion of IL-12(p40) in supernatants was measured by ELISA (n=3 experiments, mean \pm SEM, **p<0.01, *p<0.05). See also **Supplementary Fig. 5-6.**

References

- 1 Lee, B. L. & Barton, G. M. Trafficking of endosomal Toll-like receptors. *Trends in cell biology* 24, 360-369, doi:10.1016/j.tcb.2013.12.002 (2014).
- 2 Latz, E. *et al.* TLR9 signals after translocating from the ER to CpG DNA in the lysosome. *Nat Immunol* 5, 190-198, doi:10.1038/ni1028 (2004).
- 3 Leifer, C. A. *et al.* TLR9 is localized in the endoplasmic reticulum prior to stimulation. *J Immunol* 173, 1179-1183 (2004).
- 4 Fukui, R. *et al.* Unc93B1 restricts systemic lethal inflammation by orchestrating Toll-like receptor 7 and 9 trafficking. *Immunity* 35, 69-81, doi:10.1016/j.immuni.2011.05.010 (2011).
- 5 Garcia-Cattaneo, A. *et al.* Cleavage of Toll-like receptor 3 by cathepsins B and H is essential for signaling. *Proc Natl Acad Sci U S A* 109, 9053-9058, doi:10.1073/pnas.1115091109 (2012).
- 6 Lee, B. L. *et al.* UNC93B1 mediates differential trafficking of endosomal TLRs. *Elife* 2, e00291, doi:10.7554/eLife.00291 (2013).
- 7 Asagiri, M. *et al.* Cathepsin K-dependent toll-like receptor 9 signaling revealed in experimental arthritis. *Science* 319, 624-627, doi:10.1126/science.1150110 (2008).
- 8 Ewald, S. E. *et al.* Nucleic acid recognition by Toll-like receptors is coupled to stepwise processing by cathepsins and asparagine endopeptidase. *J Exp Med* 208, 643-651, doi:10.1084/jem.20100682 (2011).
- 9 Hipp, M. M. *et al.* Processing of human toll-like receptor 7 by furin-like proprotein convertases is required for its accumulation and activity in endosomes. *Immunity* 39, 711-721, doi:10.1016/j.immuni.2013.09.004 (2013).

567 10 Maschalidi, S. *et al.* Asparagine endopeptidase controls anti-influenza virus immune
 568 responses through TLR7 activation. *PLoS Pathog* 8, e1002841,
 569 doi:10.1371/journal.ppat.1002841 (2012).
 570 11 Park, B. *et al.* Proteolytic cleavage in an endolysosomal compartment is required for
 571 activation of Toll-like receptor 9. *Nat Immunol* 9, 1407-1414, doi:10.1038/ni.1669
 572 (2008).
 573 12 Sepulveda, F. E. *et al.* Critical role for asparagine endopeptidase in endocytic Toll-like
 574 receptor signaling in dendritic cells. *Immunity* 31, 737-748,
 575 doi:10.1016/j.immuni.2009.09.013 (2009).
 576 13 Hayashi, K., Sasai, M. & Iwasaki, A. Toll-like receptor 9 trafficking and signaling for
 577 type I interferons requires PIKfyve activity. *Int Immunol* 27, 435-445,
 578 doi:10.1093/intimm/dxv021 (2015).
 579 14 Hazeki, K., Uehara, M., Nigorikawa, K. & Hazeki, O. PIKfyve regulates the
 580 endosomal localization of CpG oligodeoxynucleotides to elicit TLR9-dependent
 581 cellular responses. *PLoS One* 8, e73894, doi:10.1371/journal.pone.0073894 (2013).
 582 15 Sasai, M., Linehan, M. M. & Iwasaki, A. Bifurcation of Toll-like receptor 9 signaling
 583 by adaptor protein 3. *Science* 329, 1530-1534, doi:10.1126/science.1187029 (2010).
 584 16 Saveanu, L. *et al.* IRAP identifies an endosomal compartment required for MHC class
 585 I cross-presentation. *Science* 325, 213-217, doi:10.1126/science.1172845 (2009).
 586 17 Weimershaus, M. *et al.* Conventional dendritic cells require IRAP-Rab14 endosomes
 587 for efficient cross-presentation. *J Immunol* 188, 1840-1846,
 588 doi:10.4049/jimmunol.1101504 (2012).
 589 18 Keller, S. R., Scott, H. M., Mastick, C. C., Aebersold, R. & Lienhard, G. E. Cloning
 590 and characterization of a novel insulin-regulated membrane aminopeptidase from
 591 Glut4 vesicles. *J Biol Chem* 270, 23612-23618 (1995).
 592 19 Hosaka, T. *et al.* p115 Interacts with the GLUT4 vesicle protein, IRAP, and plays a
 593 critical role in insulin-stimulated GLUT4 translocation. *Mol Biol Cell* 16, 2882-2890,
 594 doi:10.1091/mbc.E05-01-0072 (2005).
 595 20 Hirata, Y. *et al.* Vimentin binds IRAP and is involved in GLUT4 vesicle trafficking.
 596 *Biochem Biophys Res Commun* 405, 96-101, doi:10.1016/j.bbrc.2010.12.134 (2011).
 597 21 Tojo, H. *et al.* The Formin family protein, formin homolog overexpressed in spleen,
 598 interacts with the insulin-responsive aminopeptidase and profilin IIa. *Mol Endocrinol*
 599 17, 1216-1229, doi:10.1210/me.2003-0056 (2003).
 600 22 Cheng, H. *et al.* Identification of a missense variant in LNPEP that confers psoriasis
 601 risk. *J Invest Dermatol* 134, 359-365, doi:10.1038/jid.2013.317 (2014).
 602 23 Stow, J. L., Low, P. C., Offenhauser, C. & Sangermani, D. Cytokine secretion in
 603 macrophages and other cells: pathways and mediators. *Immunobiology* 214, 601-612,
 604 doi:10.1016/j.imbio.2008.11.005 (2009).
 605 24 Leuchowius, K. J., Weibrecht, I. & Soderberg, O. In situ proximity ligation assay for
 606 microscopy and flow cytometry. *Curr Protoc Cytom* Chapter 9, Unit 9 36,
 607 doi:10.1002/0471142956.cy0936s56 (2011).
 608 25 Greene, C. M. *et al.* TLR-induced inflammation in cystic fibrosis and non-cystic
 609 fibrosis airway epithelial cells. *J Immunol* 174, 1638-1646 (2005).
 610 26 Benmohamed, F. *et al.* Toll-Like Receptor 9 Deficiency Protects Mice against
 611 *Pseudomonas aeruginosa* Lung Infection. *PLoS One* 9, e90466,
 612 doi:10.1371/journal.pone.0090466 (2014).
 613 27 Saveanu, L. & van Endert, P. The role of insulin-regulated aminopeptidase in MHC
 614 class I antigen presentation. *Front Immunol* 3, 57, doi:10.3389/fimmu.2012.00057
 615 (2012).

616 28 Onji, M. *et al.* An essential role for the N-terminal fragment of Toll-like receptor 9 in
617 DNA sensing. *Nat Commun* 4, 1949, doi:10.1038/ncomms2949 (2013).

618 29 Fratti, R. A., Backer, J. M., Gruenberg, J., Corvera, S. & Deretic, V. Role of
619 phosphatidylinositol 3-kinase and Rab5 effectors in phagosomal biogenesis and
620 mycobacterial phagosome maturation arrest. *J Cell Biol* 154, 631-644,
621 doi:10.1083/jcb.200106049 (2001).

622 30 Lakadamyali, M., Rust, M. J. & Zhuang, X. Ligands for clathrin-mediated endocytosis
623 are differentially sorted into distinct populations of early endosomes. *Cell* 124, 997-
624 1009, doi:10.1016/j.cell.2005.12.038 (2006).

625 31 Kim, S. & Coulombe, P. A. Intermediate filament scaffolds fulfill mechanical,
626 organizational, and signaling functions in the cytoplasm. *Genes Dev* 21, 1581-1597,
627 doi:10.1101/gad.1552107 (2007).

628 32 Goode, B. L. & Eck, M. J. Mechanism and function of formins in the control of actin
629 assembly. *Annu Rev Biochem* 76, 593-627,
630 doi:10.1146/annurev.biochem.75.103004.142647 (2007).

631 33 Heng, T. S. & Painter, M. W. The Immunological Genome Project: networks of gene
632 expression in immune cells. *Nat Immunol* 9, 1091-1094, doi:10.1038/ni1008-1091
633 (2008).

634 34 Rifkin, I. R., Leadbetter, E. A., Busconi, L., Viglianti, G. & Marshak-Rothstein, A.
635 Toll-like receptors, endogenous ligands, and systemic autoimmune disease.
636 *Immunological reviews* 204, 27-42, doi:10.1111/j.0105-2896.2005.00239.x (2005).

637 35 Brinkmann, M. M. *et al.* The interaction between the ER membrane protein UNC93B
638 and TLR3, 7, and 9 is crucial for TLR signaling. *J Cell Biol* 177, 265-275,
639 doi:10.1083/jcb.200612056 (2007).

640 36 Descamps, D. *et al.* Toll-like receptor 5 (TLR5), IL-1 β secretion, and asparagine
641 endopeptidase are critical factors for alveolar macrophage phagocytosis and bacterial
642 killing. *Proc Natl Acad Sci U S A* 109, 1619-1624, doi:10.1073/pnas.1108464109
643 (2012).

644 37 Kuhn, S. & Geyer, M. Formins as effector proteins of Rho GTPases. *Small GTPases*
645 5, e29513, doi:10.4161/sgtp.29513 (2014).

646 38 Fernandez-Borja, M., Janssen, L., Verwoerd, D., Hordijk, P. & Neefjes, J. RhoB
647 regulates endosome transport by promoting actin assembly on endosomal membranes
648 through Dia1. *Journal of cell science* 118, 2661-2670, doi:10.1242/jcs.02384 (2005).

649 39 Prete, F. *et al.* Wiskott-Aldrich syndrome protein-mediated actin dynamics control
650 type-I interferon production in plasmacytoid dendritic cells. *J Exp Med* 210, 355-374,
651 doi:10.1084/jem.20120363 (2013).

652 40 Seth, A., Otomo, C. & Rosen, M. K. Autoinhibition regulates cellular localization and
653 actin assembly activity of the diaphanous-related formins FRL α and mDia1. *J Cell*
654 *Biol* 174, 701-713, doi:10.1083/jcb.200605006 (2006).

655 41 Wimmer, N. *et al.* Lymphotoxin β receptor activation on macrophages induces
656 cross-tolerance to TLR4 and TLR9 ligands. *J Immunol* 188, 3426-3433,
657 doi:10.4049/jimmunol.1103324 (2012).

658 42 Acharya, M. *et al.* α Integrins combine with LC3 and atg5 to regulate Toll-like
659 receptor signalling in B cells. *Nat Commun* 7, 10917, doi:10.1038/ncomms10917
660 (2016).

661 43 Han, C. *et al.* Integrin CD11b negatively regulates TLR-triggered inflammatory
662 responses by activating Syk and promoting degradation of MyD88 and TRIF via Cbl-
663 b. *Nat Immunol* 11, 734-742, doi:10.1038/ni.1908 (2010).

664 44 Etienne-Manneville, S. Cdc42--the centre of polarity. *Journal of cell science* 117,
665 1291-1300, doi:10.1242/jcs.01115 (2004).

666
667

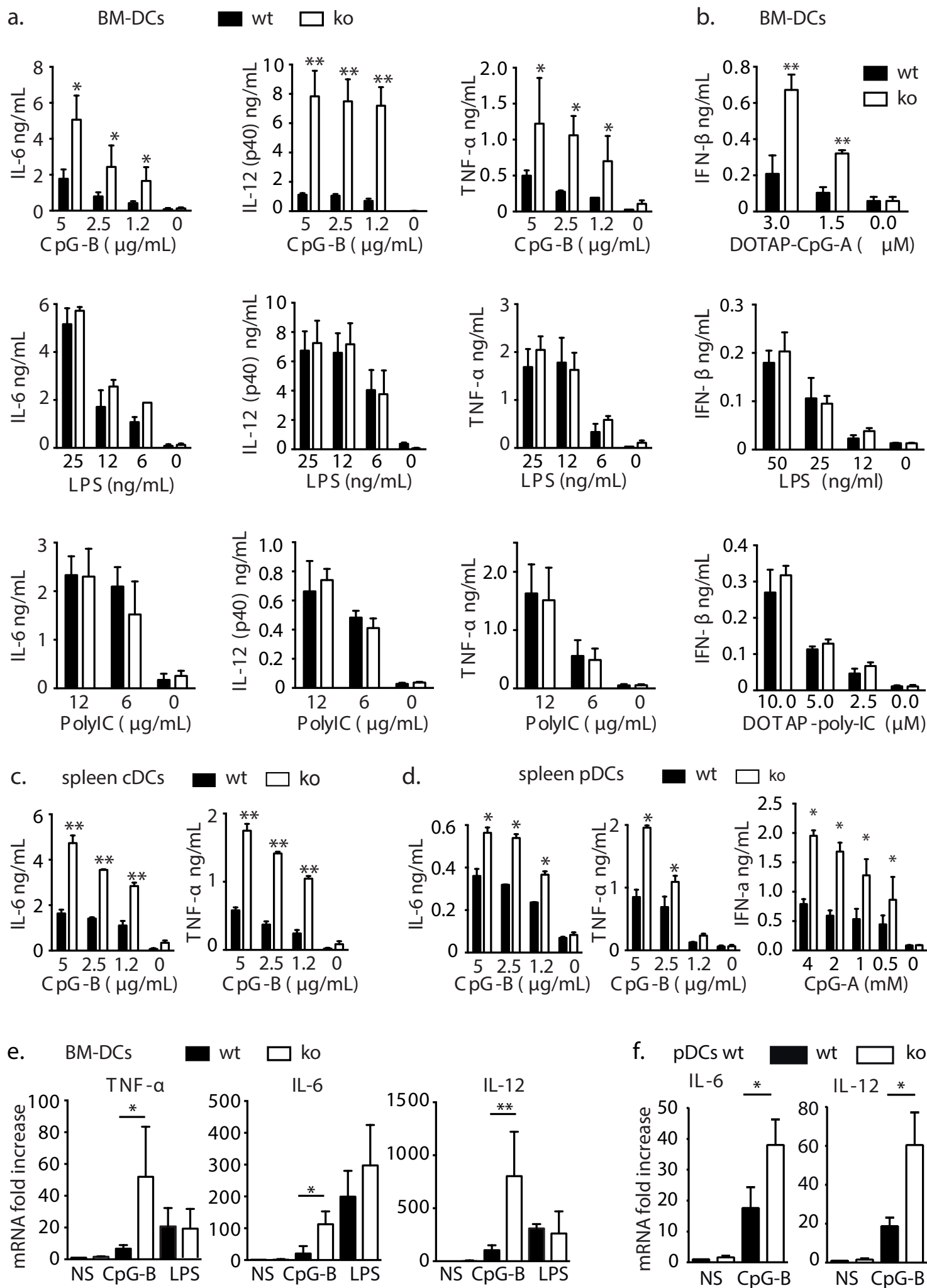


Figure 1 Babdor et al.

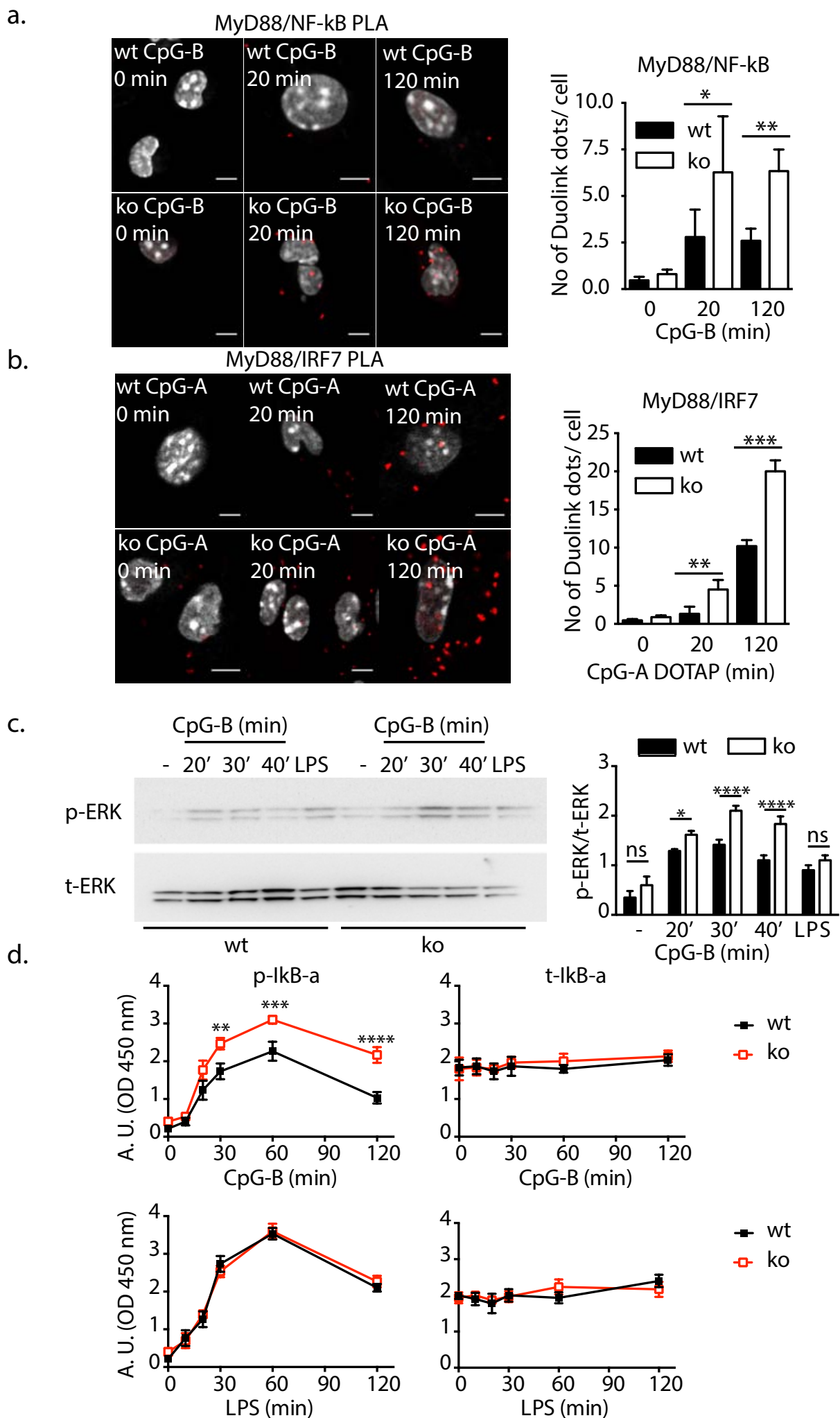


Figure 2. Babdor et al.

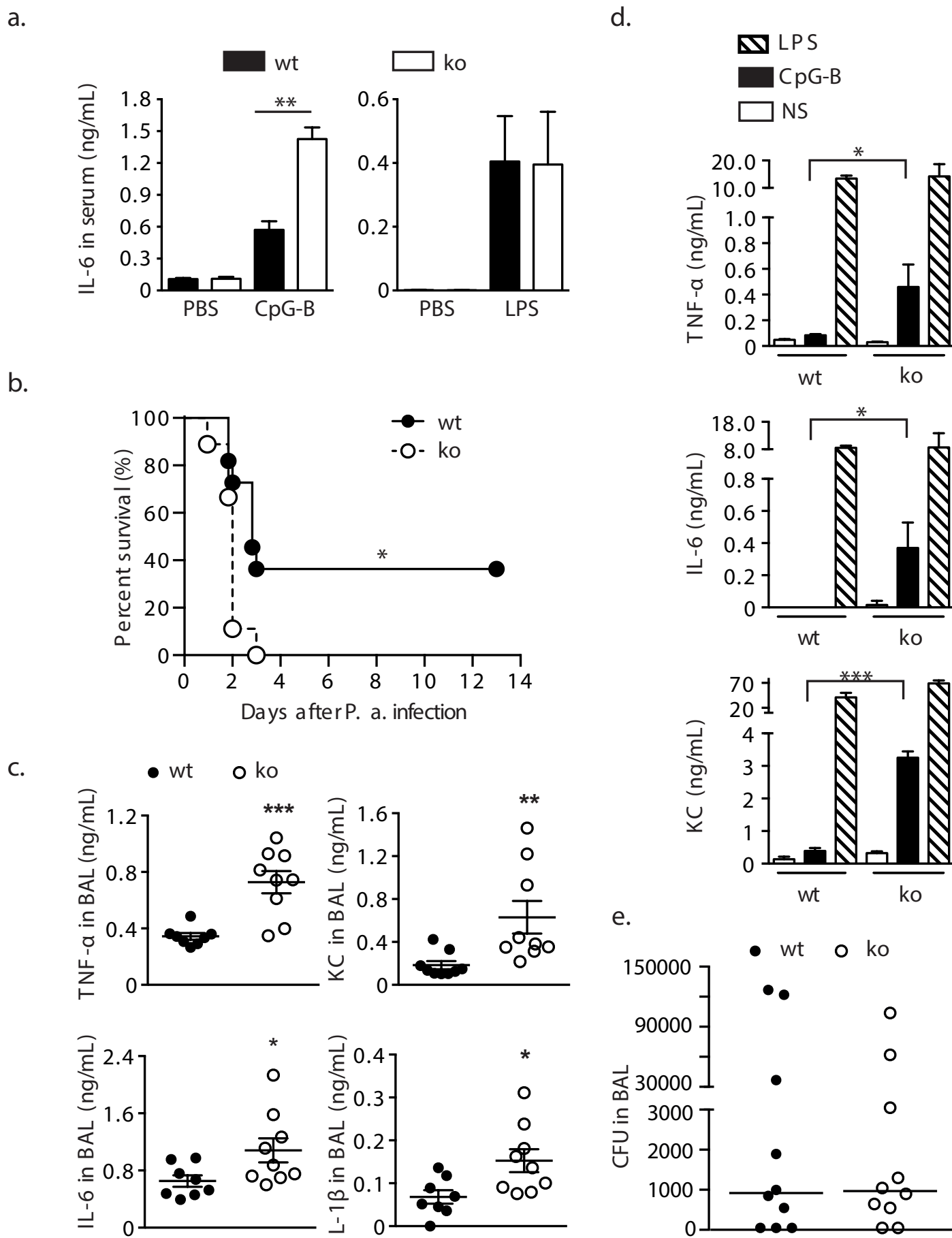
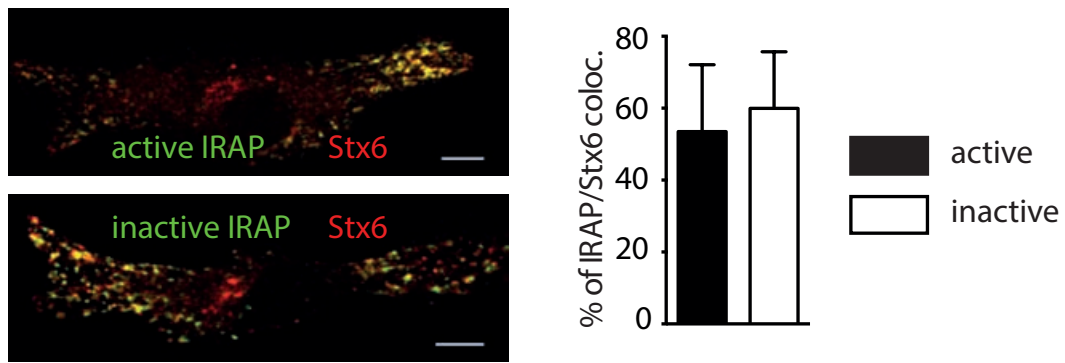
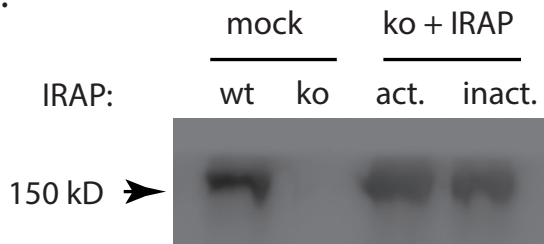


Figure 3. Babdor et al.

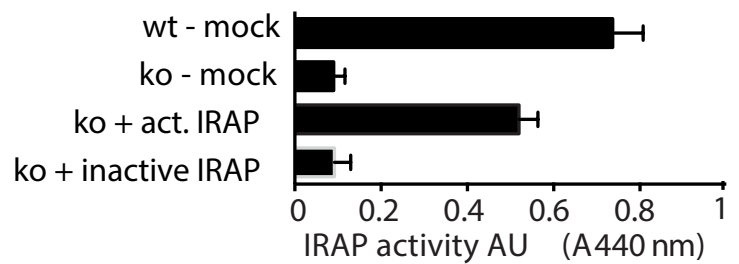
a.



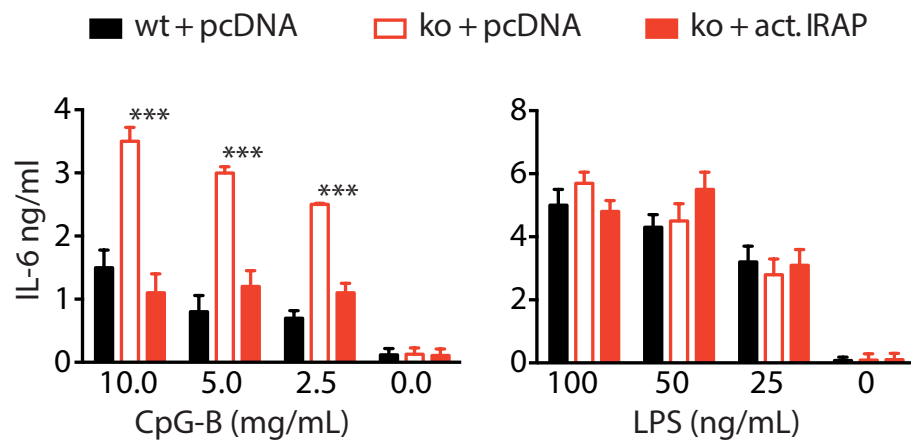
b.



c.



d.



e.

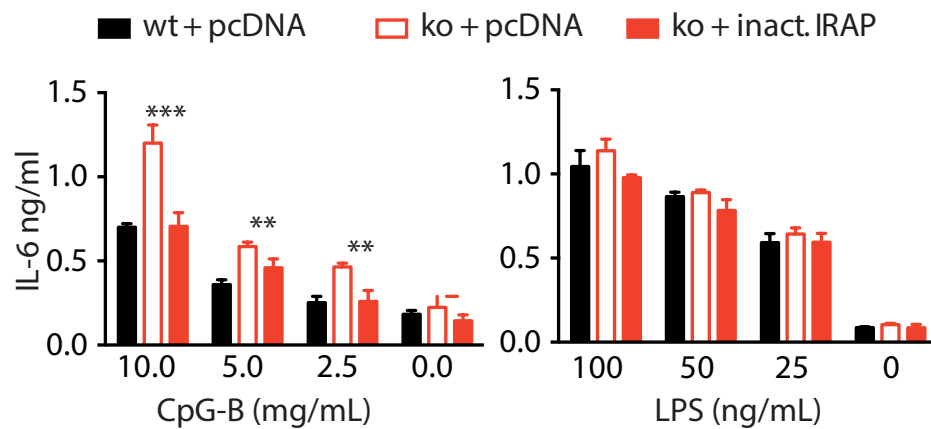
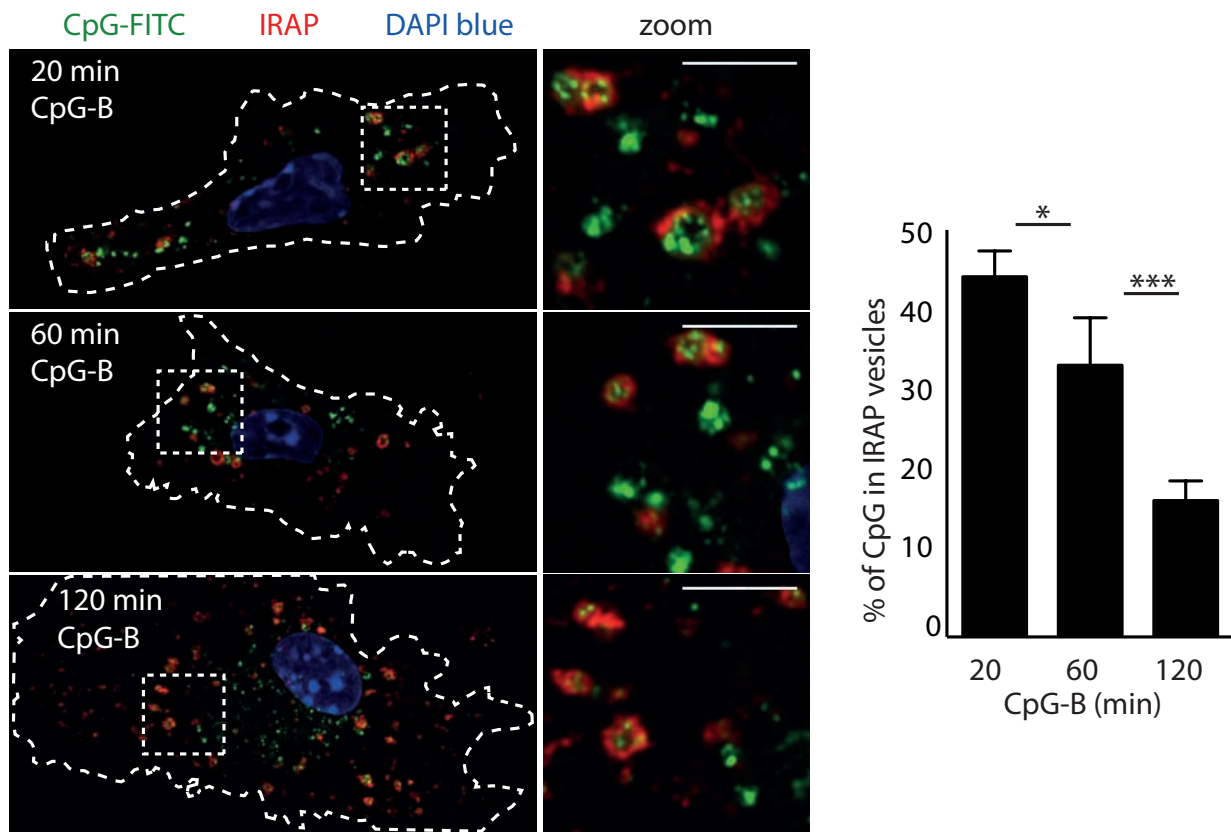


Figure 4. Babdor et al

a.



b.

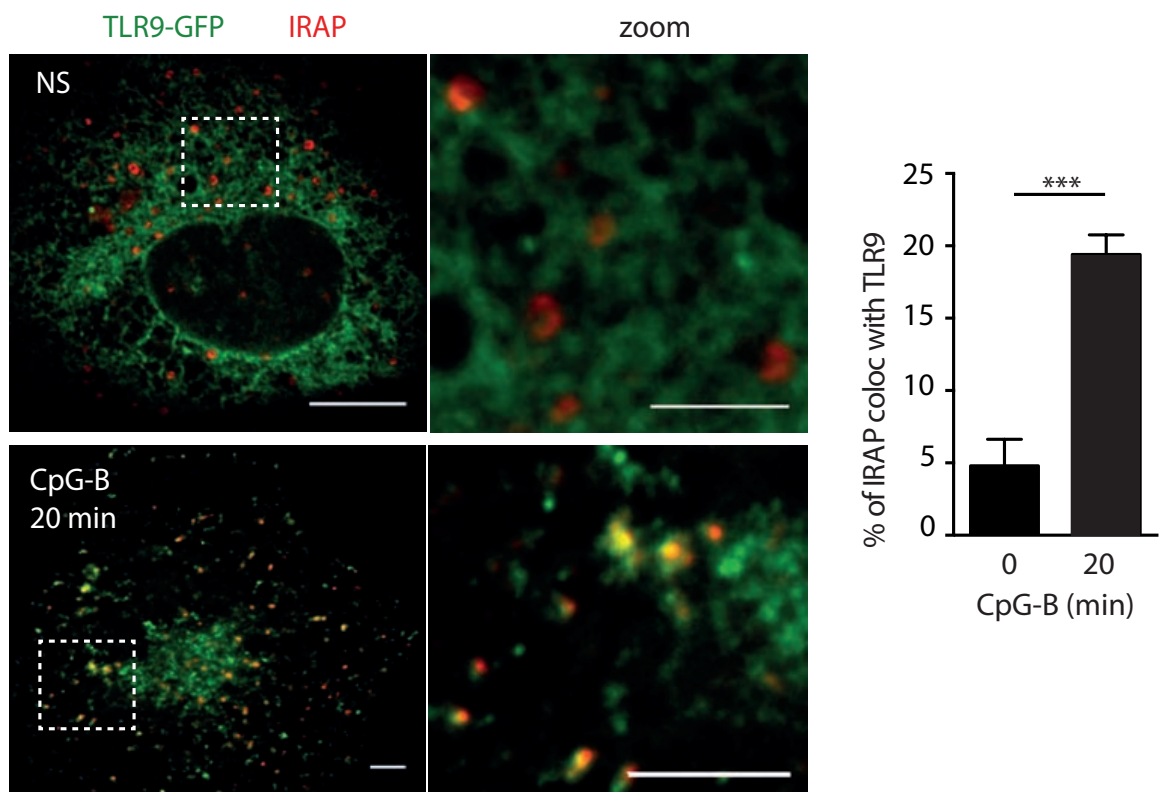


Figure 5 Babdor et al.

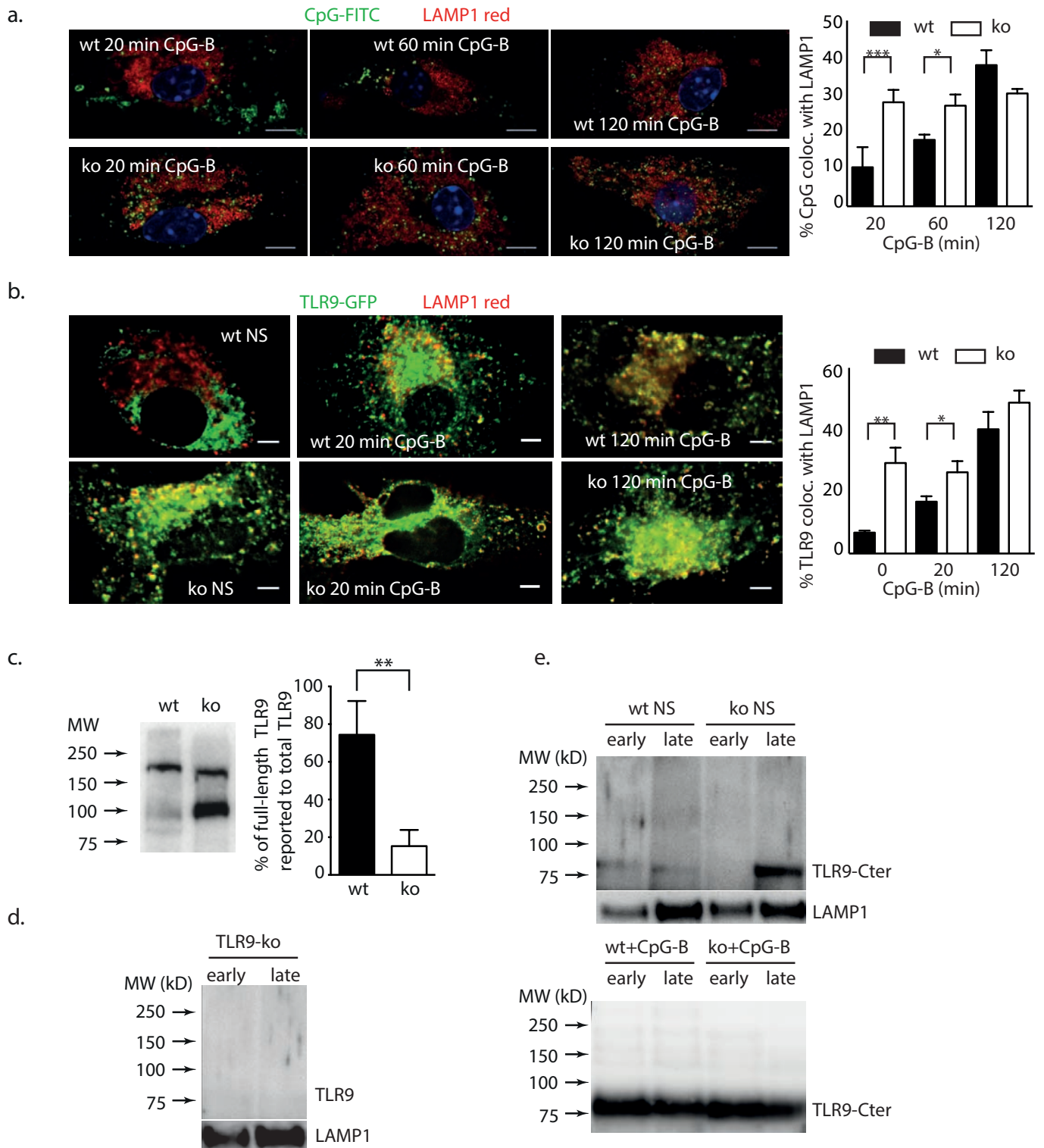
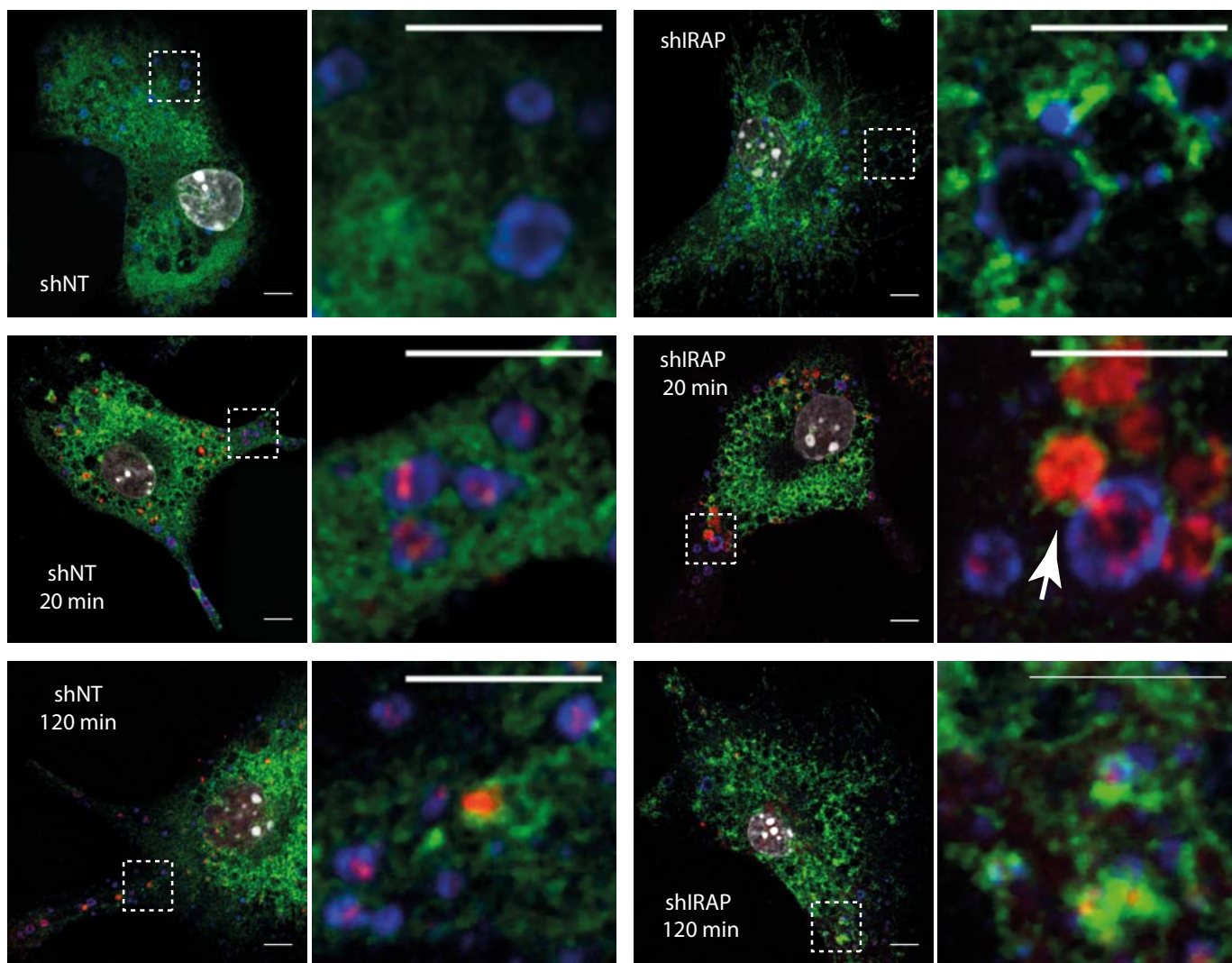


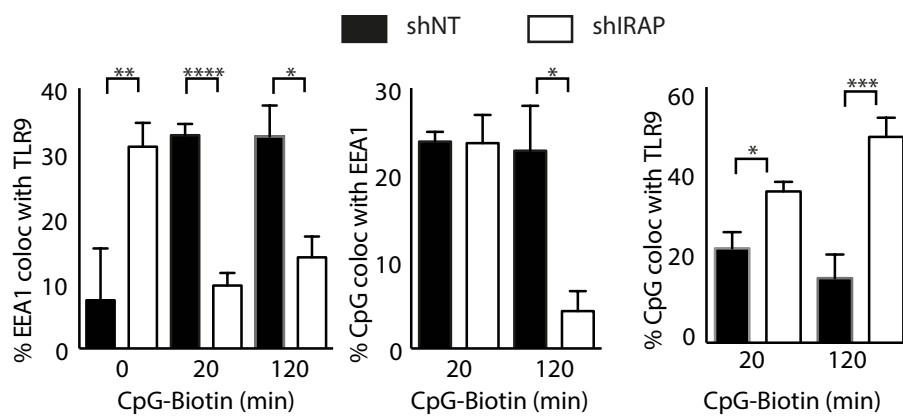
Figure 6_ Babbior et al.

a.

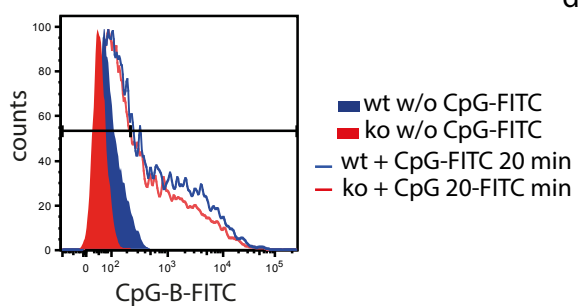
TLR9-GFP tg CpG-Biotin red EEA1 blue



b.



c.



d.

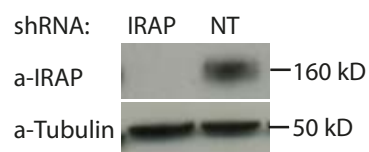


Figure 7 Babbior et al.

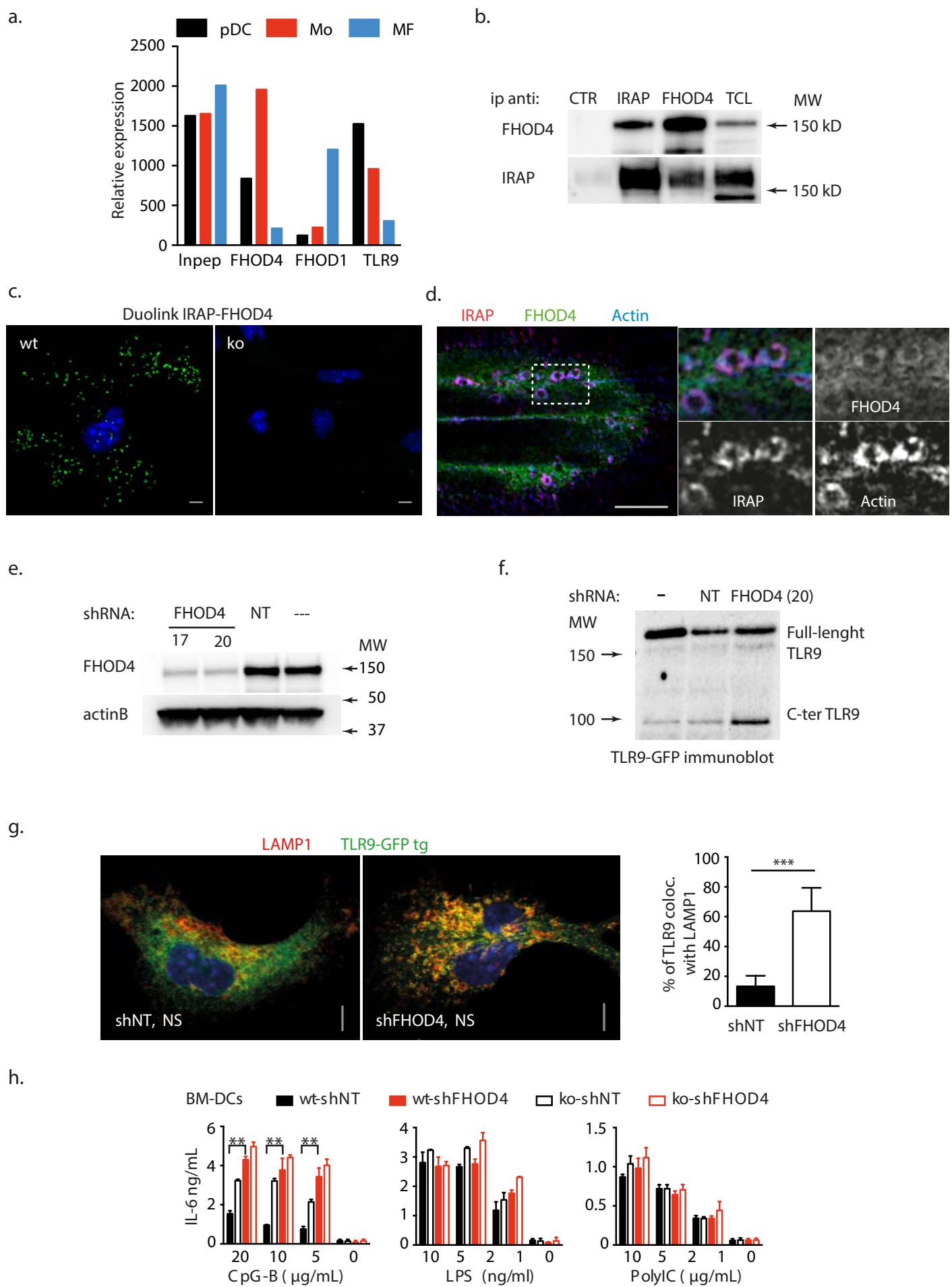


Figure 8 Babdor et al.