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A Fast, Simple, and Affordable Technique to Measure Oxygen Consumption in Living Zebrafish Embryos

Julie Somkhit,^{1,*} Roxane Loyant,^{2,*} Alexandre Brenet,¹ Rahma Hassan-Abdi,¹ Constantin Yanicostas,¹ Mathieu Porceddu,² Annie Borgne-Sanchez,² and Nadia Soussi-Yanicostas¹

Abstract

In all animal species, oxygen consumption is a key process that is partially impaired in a large number of pathological situations and thus provides informative details on the physiopathology of the disease. In this study, we describe a simple and affordable method to precisely measure oxygen consumption in living zebrafish larvae using a spectrofluorometer and the MitoXpress Xtra Oxygen Consumption Assay. In addition, we used zebrafish larvae treated with mitochondrial respiratory chain inhibitors, antimycin A or rotenone, to verify that our method enables precise and reliable measurements of oxygen consumption.

Keywords: respiration, zebrafish larvae, oxygen consumption

Introduction

RESPIRATION IS A KEY cellular function, which is altered in a great number of disorders and animal disease models, thus providing a meaningful window to interrogate the physiology of living cells and organisms.¹ Automated devices allowing precise measurement of both mitochondrial respiration (oxidative phosphorylation) and glycolysis have been developed in recent years, but their price and cost of usage restrain their use on a daily basis. To overcome this limitation, we describe here a simple and affordable method that allows precise measurement of oxygen consumption in living zebrafish larvae, using a simple spectrofluorometer and the MitoXpress Xtra Oxygen Consumption Assay (Agilent), a quantitative test relying on a fluorescent probe, which is quenched in a dose-dependent manner by O₂ molecules (Fig. 1A).

Results

To assess the ability of this method to detect fine changes in O₂ consumption in living zebrafish larvae, we measured O₂ consumption of pools of seven 5 days post-fertilization wild-type individuals treated with either vehicle (1% dimethyl

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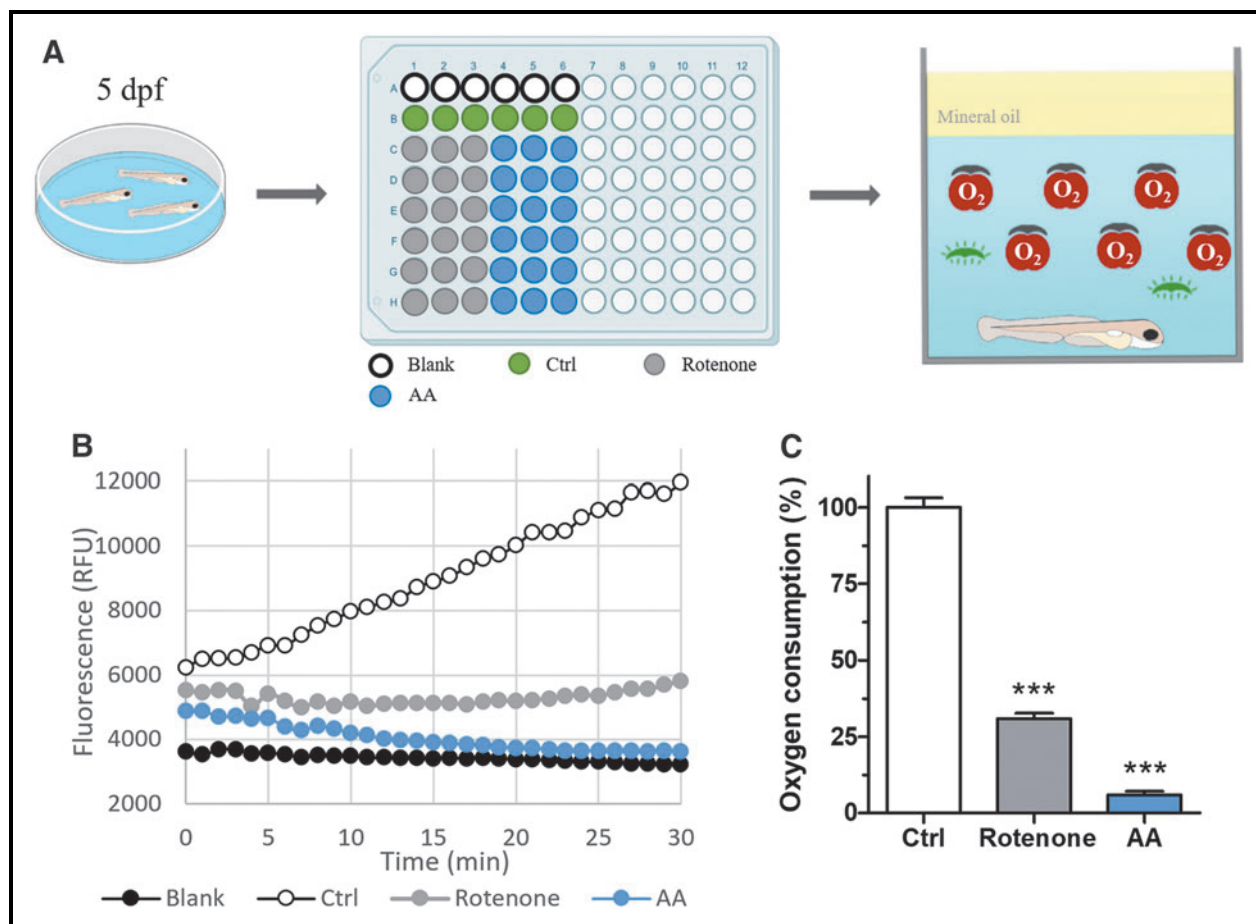


FIG. 1. Real-time measurement of oxygen consumption in 5 dpf zebrafish larvae. (A) Scheme of the experimental setup used to measure oxygen consumption *in vivo* in zebrafish larvae, treated or not with MRC inhibitors, with an example of a filled 96-well microplate. Pools of seven 5 dpf larvae treated with MRC inhibitors or untreated (control) were transferred to a 96-well microplate (Greiner Bio-One International) and then incubated in the presence of the MitoXpress Xtra probe, (Supplementary Data) a fluorescent molecule whose fluorescence is quenched in a dose-dependent manner by O₂ through direct molecular interactions, thus making fluorescence intensity inversely proportional to the amount of free O₂ dissolved in the medium. (B) Real-time measurement of the oxygen consumption in larvae exposed to MRC inhibitors or untreated, and blank (E3 medium without larva). (C) Quantification of the oxygen consumption of larvae exposed to either 2 μ M rotenone or 10 μ M AA. Whiskers represent the standard error of the mean. Rotenone ($n=36$), AA ($n=35$), control ($n=18$), and blank ($n=24$), one-way ANOVA with Tukey's Multiple Comparison Test: *** $p < 0.0001$. AA, antimycin A; ANOVA, analysis of variance; dpf, days post-fertilization; MRC, mitochondrial respiratory chain.

sulfoxide [DMSO]), 2 μ M rotenone, a specific inhibitor of the mitochondrial respiratory chain (MRC) complex I,² or 10 μ M antimycin A (AA), a specific inhibitor of the MRC complex III (Fig. 1A).^{3,4} As previously reported, we found that 1% DMSO concentration had no effect on larval development.⁵

Results showed that oxygen consumption of larvae exposed to 2 μ M rotenone and 10 μ M AA was 31% and 6%, respectively, of that observed in non-treated controls (Fig. 1B, C, Supplementary Data), indicating that the method described allows reliable measurements of oxygen consumption. Thus, this simple and direct method can be used to measure respiration *in vivo* in zebrafish intoxication and disease models.

Disclosure Statement

No competing financial interests exist.

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Supplementary Material

Supplementary Data

References

1. Frazier AE, Thorburn DR, Compton AG. Mitochondrial energy generation disorders: genes, mechanisms, and clues to pathology. *J Biol Chem* 2019;294:5386–5395.
2. Lindahl PE, Öberg KE. The effect of rotenone on respiration and its point of attack. *Exp Cell Res* 1961;23:228–237.
3. Huang LS, Cobessi D, Tung EY, Berry EA. Binding of the respiratory chain inhibitor antimycin to the mitochondrial Bc1 complex: a new crystal structure reveals an altered intramolecular hydrogen-bonding pattern. *J Mol Biol* 2005;351:573–597.
4. Rieske JS. Inhibitors of respiration at energy-coupling site 2 of the respiratory chain. *Pharmacol Ther* 1980;11:415–450.
5. Alavi Naini SM, Yanicostas C, Hassan-Abdi R, Blondeel S, Bennis M, Weiss RJ, *et al.* Surfen and oxalyl surfen decrease tau hyperphosphorylation and mitigate neuron deficits in vivo in a vertebrate model of tauopathy. *Transl Neurodegener* 2018;7:6.

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