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Nanotechnologies: a key role in virus fight

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Abstract:

Zika, Ebola, Covid-19... Viruses from their nanoworld are at the origin of major health challenges and may cause pandemic threats. In the crusade for virus fight, being "small" may be an asset, for being active at the right scale. Nanosystems have recurrently been used to enhance the immune response of the body, by themselves or in combination with active agents (e.g. antigens) as well as for virus detection and isolation. This article is not an exhaustive review, but it envisions giving a general overview of the rationale behind the use of nanotechnology in virus fight, as illustrated by several significant examples including recent findings. Nanosystems, and in particular inorganic nanoparticles, offer a large variety of declinations that may be tailored in terms of physical and chemical characteristics to address the needs of virology. Among them, noble metals, metal oxides, mesoporous silica and calcium phosphates showed special promise. In particular, the bio-inspired character and modularity of calcium phosphates is appealing and allows envisioning further original developments in the fight against infections.

Keywords: virus, pandemic, nanotechnology, vaccine, adjuvant, calcium phosphate, apatite, nanocarrier

Main article:

It is a small world after all!

All the matter that surrounds us is based on associations of elementary particles forming atoms, themselves organized in molecules and crystals often of submillimetric dimensions. Microbial life in the form of bacteria, fungi and other microorganisms is ubiquitous on Earth and plays a fundamental role, as in plant nutrition in soils ¹. The human body itself counts at least as many bacterial cells as human cells ², and all of these cellular entities – measuring few microns at most – ensure the biological machinery of the whole body. The "small world" is not far from ruling our lives... and infectious pathogens also play the game.

Besides the vast majority of harmless bacteria necessary to the life of most superior animals including humans, a significantly more limited number of bacteria are, in contrast, pathogenic and may cause infectious diseases ³. Among illustrious bacteria-bound infections, on can cite for example

tuberculosis, cholera, or else bone infections involving *Staphylococcus aureus*. But other, smaller pathogens also come into play: viruses. They are an elaborate form of parasites, requiring penetration into live cells for viral replication. Viruses consist in a nucleic acid component expressing the virus genome (in the form of single- or double-stranded RNA or DNA) protected by a proteic envelope, the capsid. In some cases, the capsid itself is also surrounded by a phospholipid-based membrane ⁴. Among critically lethal viruses are Ebola, Rabies, HIV and others. Viruses such as the *Influenza* and more recently the Covid-19 (of the family of coronaviruses) exhibit lower fatality rates but can however cause major health issues especially when vaccines are now yet available ⁵. Additional difficulties in virus fight are the possible mutations operated by viruses as well as the eventuality of animal-to-human transfers... There is thus a need to develop virus fight strategies.

The challenges of virus fight

Especially when the human body cannot combat the viral infection by itself, if the symptoms are very invalidating, or else in the case of rapidly spreading viruses, a medical arsenal is needed to limit new infections in healthy people and help infected patients recover. Therapeutic tools rely on the search for antiviral compounds, in the development of vaccines, but also of non-vaccine immunotherapy strategies with the view to boost the immune response of the body ^{6, 7}. However, as in all therapeutic approach, there is a need to develop galenic pharmaceutical formulations that may allow a successful administration to the patients and an optimized *in vivo* efficacy of the active agents at the desirable dose. Taking into account the administration route (oral, nasal, by injection...), there is a need to develop galenic forms for transporting the drug/active agents while providing a stabilizing or protective effect, controlling the release, and possibly playing an enhancing effect. Here, nanotechnologies come into play...

The role of nanotechnologies in virus infection handling

The "nanoworld" is omnipresent in our modern societies. Whether in (opto)electronics and telecommunications, energy storage, sensors, transport, pollution control, biomedical sciences and other domains, nano-objects surround us. In link with the human body, it is not surprising that nanotechnologies should play a determining role in biomedicine, especially taking into account that most biologically-relevant molecules are smaller than 10 nm and thus belong to this nanoworld⁸. Also, another significant feature of nano-objects is their capacity to exhibit enhanced or new properties in comparison to their macroscopic counterparts, due to their high surface-to-volume ratios. In this context, nanotechnologies can prove helpful for the handling of virus infections, via several aspects (**Figure 1**): nanosystems can find applications for virus detection and the setup of diagnostic tests, for the separation/isolation of viral particles, for playing the role of adjuvants in vaccine formulations, and also as nanocarriers of active agents in non-vaccine approaches. Finally, some nano-objects can exhibit by themselves a direct antiviral activity. Illustrations of virus fight strategies involving nanotechnologies are given in the following.

Separation/isolation of viral particles

In the fight against the propagation of viruses, there is a need to be able to separate the viral particles from their evolving environment. This is in particular true to eliminate viruses from water, e.g. to limit drinking water contaminations. Several strategies have been investigated, including the use of nanocomposite membranes incorporating electrospun nanofibers, which was assessed relatively to the *Semliki* Forest virus in solution, pointing out a high virus removal efficiency (up to 98.9 %) ⁹. The selective adsorption of coronaviruses was also reported using biopolymer

nano/microspheres based on chitosan and genipin ¹⁰, and can be found useful for virus removal but also in virology research for isolating the viral particles.



Figure 1: Main uses of nanotechnology in virus fight

Detection and diagnostics

Another domain of interest for nanotechnologies in virology addresses the detection of viruses and the related setup of diagnostic tests. An example concerns the use of silver (Ag) NPs associated to DNA strands for the detection of the Dengue virus, via a colorimetric detection process, and high sensitivity was reported to detect different Dengue serotype RNAs ¹¹. Iron oxide nanoparticles of several compositions have also been used for virus detection purposes, among other nanosystems, as illustrated in the case of the Zika virus, exploiting a magnetic relaxation analysis ¹². In another context, it may be mentioned that nanotechnologies may also be exploited to detect/titrate antiviral drugs, as it was reported for instance using a zinc oxide (ZnO) NPs modified carbon electrode for the detection of Acyclovir via an electrochemical response ¹³.

Intrinsically-antiviral nanosystems

Several nanosystems were found to exhibit by themselves an intrinsic antiviral activity ¹⁴. This is for example the case of Ag NPs that were extensively studied ¹⁵ with regard to several viruses including HIV, Herpes simplex virus, Poliovirus type-1 and others. Various mechanisms were considered, in a virus-dependent manner, such as imposed conformational changes in the virus or membrane biochemical modifications (e.g. by interaction with sulfhydryl groups) upon contact with the NPs, or else a modification of the virus genome ¹⁴⁻¹⁷. In many cases, the Ag NPs were coated with an organic corona (e.g. polysaccharides, polyvinylpyrrolidone (PVP), sialic acid...) and a large variety of conditions have been tested, aiming various modes of antiviral action ¹⁵. An interest in using nanosized systems in virus fight is also the possibility to adjust their dimensions to the scale of viral particles, as was illustrated in the case of the Poliovirus (particle size range of 25-30 nm) ¹⁸. These authors suggested that "small" Ag NPs, typically around 7 nm, may easily interact with the slightly larger viral particles, therefore preventing their binding to cell receptors. Besides silver, gold (Au) is another noble metal that was tentatively studied, and antiviral outcomes of Au NPs were reported

for example versus HIV and Foot-and-mouse viruses. One proposed mechanism of action related to the blocking of receptors involved in virus entry into host cells ^{14, 19}.

Besides metal particles, several other systems are also investigated and used in virus fight. Carbonbased nanomaterials (based on fullerenes, carbon nanotubes, graphene-derived compounds...) have been considered, but to date their use in virology remains scarce ¹⁴. Glyco-dendro-fullerenes have for example been studied in relation with the Ebola virus and showed promising results ²⁰. Here, the accessibility of fullerene ligands to interact with viral receptors appears to be directly linked to their antiviral activity. The potential effects of selenium (Se), a non-metal element naturally present *in vivo* and supporting the activity of immunity cells, were also investigated: Se NPs were in particular reported to be effective in conjunction with the antiviral drug *zanamivir* for a synergistic treatment of the H1N1 *Influenza* virus ²¹.

Nanosized oxide particles have also been quite widely explored in view of direct antiviral activity. Iron oxides were for instance found to be active against Hepatitis C ^{14, 22}. In the treatment of the H1N1 Influenza virus, Fe₃O₄ NPs proved efficient in vitro (at the dose of 6.5 pg iron oxide/mL) in the inhibition of the virus when compared to the control ²³. Titanium dioxide (TiO₂) NPs were explored sporadically, e.g. in relation with the Influenza virus²⁴, although their potential toxicity is still being questioned nowadays ²⁵. Zirconia (ZrO₂) NPs (positively-charged, 200 nm wide) were shown recently to inhibit viral replication in the mouse with regards to the H5N1 Influenza virus and to protect the mice against the H5N1 infection ²⁶. This effect was related to an enhancement of the innate immunity in the early stage of infection, a reduction of the viral load and the limitation of proinflammatory cytokines influx in the lungs. Zinc oxide (ZnO) NPs were also documented in the fight against the H1N1 Influenza virus ²⁷. In that study, the coating of the surface of the NPs with polyethyleneglycol (PEG) macromolecules was found to increase very significantly the virus inhibition rate compared to uncoated NPs, from 52.2 to 94.6 %. This effect was related by the authors to the smaller particle size of the coated particles, which may penetrate passively in larger amount into cells. Copper oxides were also found to exhibit some antiviral properties. For instance, cuprous oxide (theoretically Cu₂O but no characterization was provided) NPs were reported to inhibit the effect of the Hepatitis C virus in vitro, which was related to an inhibition of cell entry for virus pseudoparticles ²⁸. Cupric oxide (CuO) NPs (40 nm in size) were, on the other hand, shown recently to be active against the Herpes simplex virus type 1, HSV-1, with a reduction of 83.3 % of viral load in Vero cells ²⁹. Although the exact mechanisms of action are not yet elucidated, the authors hypothesized an interference of the NPs with viral replication stages. The potentiality of released copper ions to result in the production of oxygenated radicals generating DNA denaturation and phospholipid membrane damages was also envisaged, as already pointed out in the fight against bacteria $^{29, 30}$. Tin oxide (SnO₂) nanowires were also tested against the HSV-1 virus, and shown to suppress virus entry and interfere with membrane fusion processes ³¹. Mesoporous silica (SiO₂) NPs have been investigated for their potential antiviral properties, as in the fight against HIV ³². Here, the hydrophilic/hydrophobic modularity of the particles, essentially via surface functionalization, is thought to play a direct role in preventing regular virus transduction by establishing strong interactions with the virus envelope. Apart from inorganic NPs, a wealth of polymeric nanosystems including dendrimers, liposomes,

chitosan-based NPs, etc. have also been considered, although in a limited manner for antiviral applications ¹⁴. However, it was essentially for their drug-carrier capacities rather than an intrinsic antiviral potential.

Nanocarriers of active agents

In the field of drug transport and release, nanotechnology is also often at the center of antiviral strategies. For example, the conjugation of noble metal NPs (Ag and Au) with the antiviral peptide *FluPep* was reported to enhance the inhibition of the *Influenza* virus compared to the peptide alone, which may be related to the high grafting density of the *FluPep* ligand on the surface of the NPs ³³. Cerium oxide (CeO₂) NPs were reported in link with Vesicular *stomatitis* virus ³⁴, and in association with interferon to allow its prolonged effect in the treatment of Herpes ³⁵. Other particulate systems were also investigated. For instance, lipid-coated mesoporous SiO₂ NPs were used to carry and

deliver the ML336 antiviral agent to inhibit encephalitic Alphavirus infection ³⁶: the lipid coating aimed at improving the colloidal stability of the particles, and the NPs efficacy was shown *in vitro* and *in vivo*, leading by a significant decrease in viral load. As mentioned previously, polymer-based NPs may be used to deliver active antiviral agents. For example, Rhodamine B-labeled poly(glycerol-adipate) NPs were prepared and contacted *in vitro* with a hepatic cell line transfected with *Hepatitis* C virus RNA, evidencing a greater NPs uptake in the cells compared to the non-transfected cells ³⁷. This observation directly illustrates one possible advantage of nanoparticle-based drug delivery as compared to the administration of the free drug, allowing for a potentially higher intracellular drug release (provided that the NPs may be sufficiently loaded with the drug); and this strategy may be expanded to other types of NPs thus opening new perspectives of research. Also, the association of drugs to nanocarriers may offer the possibility to target more precisely the affected organs and avoid accumulation of the drug at "off-target" sites ³⁷.

Vaccine adjuvants

The development of vaccines is one of the keys to virus fight. In continuation with the idea of transporting active agents as in the above section, adjuvants are often included in the formulation of vaccines. Adjuvants may indeed play several potential roles: to optimize or prolong the delivery of the active components, to boost the antiviral activity, to limit the antigen dose, etc. The adjuvancy strategy was discovered in the 1920s-1930s by Glenny *et al.* ³⁸ who stated that "*if the [antigen] is too rapidly absorbed or eliminated, only slight stimulation may occur, however large the dose injected*". This work was relating in particular to the use of potassium alum as additive to "*precipitate the toxoid*" for treating Diphtheria. Although this pathology is linked to a bacterium rather than a virus, the concept of adjuvants was born. Nanosized adjuvants are particularly well suited for this task, taking into account the submicron scale in question here. Several strategies may be found. Nanoemulsion is one of them, as illustrated for example in the treatment of the Foot-and-mouth disease virus, where a water-in-oil nano-emulsion adjuvant was prepared and tested, leading to a higher cell-mediated immune response than the commercially available ISA-206 adjuvant ³⁹.

Organic, polymer-based nanoparticles were also found pertinent in the role of adjuvant, as illustrated by chitosan-based NPs loaded with inactivated Poliovirus and coated with alginate, where electrostatic interactions explained the retention of charged antigenic particles ⁴⁰. In another context, poly- γ -glutamic acid NPs were also tested in the mouse versus the *Influenza* virus, in the form of a mucosal adjuvant, and proved to induce cell-mediated immune responses and a neutralization of antibody production ⁴¹.

Inorganic NPs were often considered as vaccine adjuvants (as pioneered by Glenny et al. with alum ³⁸), taking into account their stability, large surface area and tailorable physical-chemical characteristics. Whether already approved by healthcare Regulatory agencies or still in development, inorganic adjuvants have been reviewed on several occasions taking into account their significance in this field (e.g. ⁴²⁻⁴⁴). Additionally, the role of several physical-chemical features such as chemical composition, nanoparticle size, morphology, shape, surface charge and hydrophobicity has been reviewed lately ⁴⁴. Universal conclusions cannot be made straightforwardly as biological outcomes are rather dependent on the type of nanosystem considered. Aluminum-based adjuvants, represented by the generic term "alum", have been widely used in vaccines since the 1930s. However, some potential drawbacks may be identified; in particular aluminum was reported to be neurotoxic and potentially related to Alzheimer's disease ⁴⁵ and other pathologies, although some aspects are still being debated on this matter. Also, the activity of alum relies essentially on humoral immune response (Th2 helper T lymphocytes) and is "relatively ineffective for vaccines requiring a Th1 response" as for example for treating the HIV virus ⁴⁴. Finally, new adjuvants may prove to be ultimately more efficient than alum, since nanotechnologies become increasingly more elaborate. The search for alternative compounds to alum opened the way to several exploratory works. ZnO NPs were tested in mice and showed a boosting effect on the immune system (on lymphocytes Th2 but not Th1, as for alum) ⁴⁶. It may be noted that a more balanced Th1 to Th2 ratio was obtained with cobalt oxide (Co₃O₄) NPs ⁴⁷. Noble metal NPs (Ag, Au) can also be suited as vaccine adjuvants. An example is the use of Au NPs in the development of DNA vaccines for the HIV virus ⁴⁸. The role of the shape (spherical, rod, cubic) and size of Au NPs coated with West Nile virus (WNV) envelope protein has been scrutinized *in vitro* and *in vivo* ⁴⁹: different cytokine pathways were activated depending on these characteristics. Other alternatives to alum have been explored, including in a comparative study in rats, carried out in the 1990s and involving iron oxides, alumina (Al₂O₃), SnO₂, ZrO₂, SiO₂, ground glass and talcum ⁵⁰. The authors reported a correlation between "*the magnitude of adjuvancy*" and the degree of inflammatory response and surface area while the hydrophilic/hydrophobic character of the particles did not seem directly correlated. In that work, alumina and iron oxides appeared as the most promising among the compounds tested. In a recent study, dendrimer-like mesoporous SiO₂ NPs were developed to address the Foot-and-mouth disease virus, and were found to be well adapted as nanocarriers of large proteins/antigens ⁵¹. Layered double hydroxides (LDHs) were tested as potential vaccine adjuvants since years 2010s ^{43, 52, 53}. A study was, in particular, undergone with various types of LDHs and the *in vitro* response of the immune system was found to be greatly dependent on the LDH chemical composition ^{43, 52}. These systems may represent a relevant family of vaccine adjuvants to further develop in the future.

Besides alum, calcium phosphates (CaP) represent another major family of inorganic compounds that have been explored for years in the domain of vaccine adjuvants. This may be rationalized on the basis of their similarity to the mineral part of bones and teeth (apatitic CaP have been present naturally in all Vertebrates for millions of years) and may thus be considered as "intrinsically" biocompatible ⁵⁴. Furthermore, CaP compounds – and more particularly nanocrystalline apatites – exhibit a high surface reactivity and are greatly tailorable in terms of chemical composition and surface features, opening the way to a wealth of biomedical applications as reviewed recently ⁵⁵ (**Figure 2**). Plus, they are biodegradable *in vivo* and lead to natural metabolites (calcium and phosphate ions already present in body fluids) that the host body can eliminate easily.



Figure 2: Nanocrystalline calcium phosphate apatites: a platform for nanomedicine (modified from ref. 55)

Calcium phosphates: relevant bioinspired compounds

The first use of CaP as vaccine adjuvant was reported for the delivery of Diphtheria anatoxin by Relyveld *et al*. in the 1960s ⁵⁶, as an alternative to pre-existing aluminium-based formulations, in the case of allergic patients. In this case, brushite (CaHPO₄) particles was used. The authors reported several advantages of CaP adjuvants: no signs of allergy were detected, large scale production is easy, antigen adsorption is complete and immediate at neutral pH, and it does not have the inconvenient of aluminum phosphate which requires a maturation stage at acidic pH for adsorption to be complete. Also, the formulations obtained were stable over several years ⁵⁶. CaP have then been used in vaccinal fight against several viral infections as Poliovirus and *Hepatitis* B⁴². However, often, no precise characterization information is given about the used CaP particles. Jiang et al. have reported a physical-chemical analysis of a commercial CaP adjuvant and showed that it was composed of a nonstoichiometric hydroxyapatite phase, $Ca_{10-x}(PO_4)_{6-x}(HPO_4)_x(OH)_{2-x}$ ⁵⁷. Synthesis parameters of the CaP particles proved to condition the final association with the active component of the vaccine ⁵⁸, and the degree of nonstoichiometry appeared to be one relevant criterion. Indeed, temperature, pH and maturation time are key parameters that may control the final characteristics of nanocrystalline apatites⁵⁹. Recently, CaP adjuvants have been prepared with more controlled features than traditional ones ⁶⁰, although detailed structural and compositional analyses were not reported. The absence of side effects and the potency of the CaP adjuvant were shown for example in a study on Herpes simplex virus 2 (HSV-2) ⁶⁰, where the authors reported that "[these data] strongly suggest that CaP nanoparticles significantly enhance the antibody response to HSV-2 antigen". Here, the release of the antigen from the CaP surface was considered to happen progressively over an extended period of time. In another study, other authors claimed the effect of CaP nanoparticles for the setup of a vaccinal formulation (final NPs concentration: 0.3 % wt./vol., typical NPs size: 450-500 nm) in relation to the H1N1 Influenza virus in infected mice ⁶¹. Despite the absence of structural and compositional information on the particles used, the formulation used showed promise in its efficiency and for antigen dose-sparing. A Phase I human clinical study is in progress in the US where CaP NPs were injected intramuscularly ⁶². Related US patent literature indicates that "there is [a] need for calcium phosphate core particles that can be effectively used as supports and matrices for sustained release of polynucleotide material (DNA or RNA) encoding immunogenic polypeptides" ⁶³. Studies in other scientific domains have indeed pointed out the possible adsorption of (poly)nucleotides onto apatite nanocrystals ⁶⁴⁻⁶⁶. Also, transfection was reported to be feasible using apatite NPs as carriers of nucleic acids ⁶⁷. The use of CaP NPs (50-100 nm) to develop a DNA vaccine against the Foot-and-mouth disease virus was also reported, and showed similar in vitro transfection efficacy as commercial lipofectamine as well as significant induced humoral immune response in vivo on rodents ⁶⁸. Another desirable feature for vaccine adjuvants is the possibility to envisage a variety of administration routes. Beyond injections, nasal/mucosal administration is for example also appealing and may be considered by patients as less invasive. The possible administration of CaP NPs (size < 50 nm) by inhalation was reported, although in another context (cardiovascular disorders)⁶⁹, and access to the pulmonary system of rodents was shown to be successful. In the field of vaccines, mucosal immunity was reported using CaP NPs as adjuvant in mice relatively to the Herpes Simplex Virus Type 2⁷⁰. One other appealing feature of CaP compounds, and especially of apatites, is their capacity to accommodate a large number of ionic substituents, therefore allowing for a modulation of physical-chemical but also biological properties ⁵⁵. In view of cancer immunotherapy, Mg- and Zn-doped CaP particles (β tricalcium phosphate (β -TCP) then apatite) were considered ⁷¹, and some compositions exhibited enhanced anti-tumor activity. Still in cancer immunotherapy, a pilot study on the development of a therapeutic antitumor vaccine employing CaP particles and self-antigens was reported ⁷², e.g. to face breast carcinoma or melanoma. The formulation (although made of microparticles) was well tolerated by patients and led to clinical improvements in some patients (activation of T-cell response). Although this does not address viral infections, these examples give information of how CaP-bearing vaccine formulations behave in humans. The potential role of CaP particles shape and composition on adjuvant efficacy was also mentioned, but without a systematic study 44, 73. In addition, the

association of CaP NPs with active molecules can be made either during CaP precipitation or in a subsequent step via adsorption on pre-existing particles. The adsorption capabilities of nanocrystalline biomimetic apatites are indeed well-known, as was demonstrated for a wealth of drugs/biomolecules in several contexts (e.g. antibacterial enzymes ⁷⁴, anti-tumor and anti-resorptive drugs ⁷⁵⁻⁷⁷, antibodies ⁷⁸, antibiotics ⁷⁹, etc.). With all these appealing features, bio-inspired calcium phosphates appear very promising as tailorable vaccine adjuvants to face several scenarios in virus fight and beyond.

Concluding remarks

Nanotechnology is part of our societies and plays a key role in the fight against infections, including for addressing a large variety of viruses. Whether as drug nanocarriers, vaccine adjuvants or for intrinsic antiviral properties, the inclusion of nanosystems in therapeutic galenic formulations has permitted significant improvements in terms of active agent stabilization, antigen dose sparing, controlled/prolonged release, adequate exposure of antigens, improved cell uptake, etc. Besides, nanosystems proved helpful for the separation and isolation of viral particles as well as in the development of detection and diagnostics tools.

Among appealing nanosystems, inorganic nanoparticles have in particular raised great interest thanks to wide possibilities offered in terms of size, shape, porosity, resorption rate, surface features, etc. Noble metals, metal oxides, mesoporous silica and calcium phosphates have been especially investigated, besides aluminum-based compounds whose potential neural toxicity has been debated. The incorporation of nanosystems in therapeutic formulations requires approval from Regulatory agencies, but also more and more public acceptance; therefore privileging highly biocompatible compounds whenever possible is a safe prerequisite. Bio-inspired calcium phosphates, proved to be increasingly appealing for use in nanomedicine as in virus fight, and should allow even more original uses in the future. However, care should be taken in the characterization of the particles produced, and the use of complementary techniques is recommended to assess structural and compositional features among the CaP family of compounds. Independently of the type of nanosystem considered, exploration of the mechanism(s) underlying the enhancement of antiviral activity is necessary in order to better understand the key parameters.

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References:

1. Yadvinder, S.; Bijay, S.; Timsina, J., Crop Residue Management for Nutrient Cycling and Improving Soil Productivity in Rice-Based Cropping Systems in the Tropics. In *Advances in Agronomy*, Academic Press: **2005**; Vol. 85, pp 269-407.

2. Sender, R.; Fuchs, S.; Milo, R., Revised Estimates for the Number of Human and Bacteria Cells in the Body. *PLoS biology* **2016**, *14* (8), e1002533-e1002533.

3. McFall-Ngai, M., Care for the community. *Nature* **2007**, *445* (7124), 153-153.

4. Prasad, B. V. V.; Schmid, M. F., Principles of virus structural organization. *Advances in experimental medicine and biology* **2012**, *726*, 17-47.

5. Online resources : <u>https://www.livescience.com/56598-deadliest-viruses-on-earth.html;</u> <u>https://www.statista.com/statistics/1095129/worldwide-fatality-rate-of-major-virus-outbreaks-in-the-last-50-years/</u>

6. Dalgleish, A. G., Vaccines versus immunotherapy: overview of approaches in deciding between options. *Human vaccines & immunotherapeutics* **2014**, *10* (11), 3369-3374.

7. Dibo, M.; Battocchio, E. C.; Dos Santos Souza, L. M.; da Silva, M. D. V.; Banin-Hirata, B. K.; Sapla, M. M. M.; Marinello, P.; Rocha, S. P. D.; Faccin-Galhardi, L. C., Antibody Therapy for the Control of Viral Diseases: An Update. *Curr Pharm Biotechnol* **2019**, *20* (13), 1108-1121.

8. Milioni, D.; Tsortos, A.; Velez, M.; Gizeli, E., Extracting the Shape and Size of Biomolecules Attached to a Surface as Suspended Discrete Nanoparticles. *Analytical Chemistry* **2017**, *89* (7), 4198-4203.

9. Al-Attabi, R.; Rodriguez-Andres, J.; Schütz, J. A.; Bechelany, M.; des Ligneris, E.; Chen, X.; Kong, L.; Morsi, Y. S.; Dumée, L. F., Catalytic electrospun nano-composite membranes for virus capture and remediation. *Separation and Purification Technology* **2019**, *229*, 115806.

10. Ciejka, J.; Wolski, K.; Nowakowska, M.; Pyrc, K.; Szczubiałka, K., Biopolymeric nano/microspheres for selective and reversible adsorption of coronaviruses. *Materials Science and Engineering: C* **2017**, *76*, 735-742.

11. Vinayagam, S.; Rajaiah, P.; Mukherjee, A.; Natarajan, C., DNA-triangular silver nanoparticles nanoprobe for the detection of dengue virus distinguishing serotype. *Spectrochim Acta A Mol Biomol Spectrosc* **2018**, *202*, 346-351.

12. Shelby, T.; Banerjee, T.; Zegar, I.; Santra, S., Highly Sensitive, Engineered Magnetic Nanosensors to Investigate the Ambiguous Activity of Zika Virus and Binding Receptors. *Scientific Reports* **2017**, *7* (1), 7377.

13. Pavamana, M.; Shetti, N. P.; Malode, S. J.; Bukkitgar, S. D., Nano level detection and analysis of an antiviral drug at ZnO nanoparticles modified sensor. *Materials Today: Proceedings* **2019**, *18*, 1568-1573.

14. Kerry, R. G.; Malik, S.; Redda, Y. T.; Sahoo, S.; Patra, J. K.; Majhi, S., Nano-based approach to combat emerging viral (NIPAH virus) infection. *Nanomedicine* **2019**, *18*, 196-220.

15. Galdiero, S.; Falanga, A.; Vitiello, M.; Cantisani, M.; Marra, V.; Galdiero, M., Silver nanoparticles as potential antiviral agents. *Molecules* **2011**, *16* (10), 8894-918.

16. Lara, H. H.; Ayala-Nuñez, N. V.; Ixtepan-Turrent, L.; Rodriguez-Padilla, C., Mode of antiviral action of silver nanoparticles against HIV-1. *Journal of Nanobiotechnology* **2010**, *8* (1), 1.

17. Rónavári, A.; Kovács, D.; Igaz, N.; Vágvölgyi, C.; Boros, I. M.; Kónya, Z.; Pfeiffer, I.; Kiricsi, M., Biological activity of green-synthesized silver nanoparticles depends on the applied natural extracts: a comprehensive study. *International journal of nanomedicine* **2017**, *12*, 871-883.

18. Huy, T. Q.; Hien Thanh, N. T.; Thuy, N. T.; Chung, P. V.; Hung, P. N.; Le, A. T.; Hong Hanh, N. T., Cytotoxicity and antiviral activity of electrochemical - synthesized silver nanoparticles against poliovirus. *J Virol Methods* **2017**, *241*, 52-57.

19. Vijayakumar, S.; Ganesan, S., Gold nanoparticles as an HIV entry inhibitor. *Curr HIV Res* **2012**, *10* (8), 643-6.

20. Luczkowiak, J.; Muñoz, A.; Sánchez-Navarro, M.; Ribeiro-Viana, R.; Ginieis, A.; Illescas, B. M.; Martín, N.; Delgado, R.; Rojo, J., Glycofullerenes Inhibit Viral Infection. *Biomacromolecules* **2013**, *14* (2), 431-437.

21. Lin, Z.; Li, Y.; Guo, M.; Xiao, M.; Wang, C.; Zhao, M.; Xu, T.; Xia, Y.; Zhu, B., Inhibition of H1N1 influenza virus by selenium nanoparticles loaded with zanamivir through p38 and JNK signaling pathways. *RSC Advances* **2017**, *7* (56), 35290-35296.

22. Delaviz, N.; Gill, P.; Ajami, A.; Aarabi, M., Aptamer-conjugated magnetic nanoparticles for the efficient removal of HCV particles from human plasma samples. *RSC Advances* **2015**, *5* (97), 79433-79439.

23. Kumar, R.; Sahoo, G. C.; Chawla-Sarkar, M.; Nayak, M. K.; Trivedi, K.; Rana, S.; Pandey, K.; Das, V.; Topno, R.; Das, P., Antiviral effect of Glycine coated Iron oxide nanoparticles iron against H1N1 influenza A virus. *International Journal of Infectious Diseases* **2016**, *45*, 281-282.

24. Mazurkova, N. A.; Spitsyna, Y. E.; Shikina, N. V.; Ismagilov, Z. R.; Zagrebel'nyi, S. N.; Ryabchikova, E. I., Interaction of titanium dioxide nanoparticles with influenza virus. *Nanotechnologies in Russia* **2010**, *5* (5), 417-420.

25. Winkler, H. C.; Notter, T.; Meyer, U.; Naegeli, H., Critical review of the safety assessment of titanium dioxide additives in food. *J Nanobiotechnology* **2018**, *16* (1), 51.

26. Huo, C.; Xiao, J.; Xiao, K.; Zou, S.; Wang, M.; Qi, P.; Liu, T.; Hu, Y., Pre-Treatment with Zirconia Nanoparticles Reduces Inflammation Induced by the Pathogenic H5N1 Influenza Virus. *International journal of nanomedicine* **2020**, *15*, 661-674.

27. Ghaffari, H.; Tavakoli, A.; Moradi, A.; Tabarraei, A.; Bokharaei-Salim, F.; Zahmatkeshan, M.; Farahmand, M.; Javanmard, D.; Kiani, S. J.; Esghaei, M.; Pirhajati-Mahabadi, V.; Monavari, S. H.; Ataei-Pirkooh, A., Inhibition of H1N1 influenza virus infection by zinc oxide nanoparticles: another emerging application of nanomedicine. *Journal of Biomedical Science* **2019**, *26* (1), 70.

28. Hang, X.; Peng, H.; Song, H.; Qi, Z.; Miao, X.; Xu, W., Antiviral activity of cuprous oxide nanoparticles against Hepatitis C Virus in vitro. *J Virol Methods* **2015**, *222*, 150-7.

29. Tavakoli, A.; Hashemzadeh, M. S., Inhibition of herpes simplex virus type 1 by copper oxide nanoparticles. *J Virol Methods* **2020**, *275*, 113688.

30. Warnes, S. L.; Summersgill, E. N.; Keevil, C. W., Inactivation of murine norovirus on a range of copper alloy surfaces is accompanied by loss of capsid integrity. *Appl Environ Microbiol* **2015**, *81* (3), 1085-91.

31. Trigilio, J.; Antoine, T. E.; Paulowicz, I.; Mishra, Y. K.; Adelung, R.; Shukla, D., Tin Oxide Nanowires Suppress Herpes Simplex Virus-1 Entry and Cell-to-Cell Membrane Fusion. *PLoS ONE* **2012**, 7(10): e48147.

32. de Souza, E. S. J. M.; Hanchuk, T. D.; Santos, M. I.; Kobarg, J.; Bajgelman, M. C.; Cardoso, M. B., Viral Inhibition Mechanism Mediated by Surface-Modified Silica Nanoparticles. *ACS Appl Mater Interfaces* **2016**, *8* (26), 16564-72.

33. Alghrair, Z. K.; Fernig, D. G.; Ebrahimi, B., Enhanced inhibition of influenza virus infection by peptide-noble metal nanoparticle conjugates. *Beilstein J. Nanotechnol.* **2019**, 10, 1038–47.

34. Zholobak, N. M.; Olevinskaia, Z. M.; Spivak, N.; Shcherbakov, A. B.; Ivanov, V. K.; Usatenko, A. V., Antiviral effect of cerium dioxide nanoparticles stabilized by low-molecular polyacrylic acid. *Mikrobiol Z* **2010**, *72* (3), 42-7.

35. Shydlovska, O.; Kharchenko, E.; Osenniy, I.; Spivak, M.; Shcherbakov, A.; Zholobak, N., Nanoparticles of cerium dioxide – an effective antiviral agent and adjuvant of biologically active molecules. *ScienceRise:Biological Science* **2018**, 1 (10), 26-30.

36. LaBauve, A. E.; Rinker, T. E.; Noureddine, A.; Serda, R. E.; Howe, J. Y.; Sherman, M. B.; Rasley, A.; Brinker, C. J.; Sasaki, D. Y.; Negrete, O. A., Lipid-Coated Mesoporous Silica Nanoparticles for the Delivery of the ML336 Antiviral to Inhibit Encephalitic Alphavirus Infection. *Scientific Reports* **2018**, *8* (1), 13990.

37. Abo-zeid, Y.; Urbanowicz, R. A.; Thomson, B. J.; Irving, W. L.; Tarr, A. W.; Garnett, M. C., Enhanced nanoparticle uptake into virus infected cells: Could nanoparticles be useful in antiviral therapy ? *International Journal of Pharmaceutics* **2018**, *547* (1), 572-581.

38. Glenny, A. T.; Buttle, G. A. H.; Stevens, M. F., Rate of disappearance of diphtheria toxoid injected into rabbits and guinea - pigs: Toxoid precipitated with alum. *The Journal of Pathology and Bacteriology* **1931**, *34* (2), 267-275.

39. Chen, Z.; Zhang, S.; Li, Z.; Ma, G.; Su, Z., Construction of a stable w/o nano-emulsion as a potential adjuvant for foot and mouth disease virus vaccine. *Artificial cells, nanomedicine, and biotechnology* **2017**, *45* (5), 897-906.

40. Tafaghodi, M.; Kersten, G.; Jiskoot, W. I. M., Nano-adjuvanted polio vaccine : Preparation and characterization of chitosan and trimethylchitosan (TMC) nanoparticles loaded with inactivated polio virus and coated with sodium alginate. *Nanomedicine Journal* **2014**, *1* (4), 220-228.

41. Okamoto, S.; Matsuura, M.; Akagi, T.; Akashi, M.; Tanimoto, T.; Ishikawa, T.; Takahashi, M.; Yamanishi, K.; Mori, Y., Poly(gamma-glutamic acid) nano-particles combined with mucosal influenza

virus hemagglutinin vaccine protects against influenza virus infection in mice. *Vaccine* **2009**, *27* (42), 5896-905.

42. Paneque-Quevedo, A. A., Inorganic compounds as vaccine adjuvants. *Biotecnología Aplicada* **2013**, *30*, 250-256.

43. Maughan, C. N.; Preston, S. G.; Williams, G. R., Particulate inorganic adjuvants: recent developments and future outlook. *J Pharm Pharmacol* **2015**, *67* (3), 426-49.

44. Li, X.; Wang, X.; Ito, A., Tailoring inorganic nanoadjuvants towards next-generation vaccines. *Chemical Society Reviews* **2018**, *47* (13), 4954-4980.

45. Tomljenovic, L., Aluminum and Alzheimer's disease: after a century of controversy, is there a plausible link? *J Alzheimers Dis* **2011**, *23* (4), 567-98.

46. Matsumura, M.; Nagata, M.; Nakamura, K.; Kawai, M.; Baba, T.; Yamaki, K.; Yoshino, S., Adjuvant effect of zinc oxide on Th2 but not Th1 immune responses in mice. *Immunopharmacol Immunotoxicol* **2010**, *32* (1), 56-62.

47. Cho, W. S.; Dart, K.; Nowakowska, D. J.; Zheng, X.; Donaldson, K.; Howie, S. E., Adjuvanticity and toxicity of cobalt oxide nanoparticles as an alternative vaccine adjuvant. *Nanomedicine (Lond)* **2012**, *7* (10), 1495-505.

48. Xu, L.; Liu, Y.; Chen, Z.; Li, W.; Wang, L.; Wu, X.; Ji, Y.; Zhao, Y.; Ma, L.; Shao, Y.; Chen, C., Surface-engineered gold nanorods: promising DNA vaccine adjuvant for HIV-1 treatment. *Nano Lett* **2012**, *12* (4), 2003-12.

49. Niikura, K.; Matsunaga, T.; Suzuki, T.; Kobayashi, S.; Yamaguchi, H.; Orba, Y.; Kawaguchi, A.; Hasegawa, H.; Kajino, K.; Ninomiya, T.; Ijiro, K.; Sawa, H., Gold Nanoparticles as a Vaccine Platform: Influence of Size and Shape on Immunological Responses in Vitro and in Vivo. *ACS Nano* **2013**, *7* (5), 3926-3938.

50. Naim, J. O.; van Oss, C. J.; Wu, W.; Giese, R. F.; Nickerson, P. A., Mechanisms of adjuvancy: I--Metal oxides as adjuvants. *Vaccine* **1997**, *15* (11), 1183-93.

51. Liu, Z.; Ru, J.; Sun, S.; Teng, Z.; Dong, H.; Song, P.; Yang, Y.; Guo, H., Uniform dendrimer-like mesoporous silica nanoparticles as a nano-adjuvant for foot-and-mouth disease virus-like particle vaccine. *Journal of Materials Chemistry B* **2019**, *7* (21), 3446-3454.

52. Williams, G. R.; Fierens, K.; Preston, S. G.; Lunn, D.; Rysnik, O.; De Prijck, S.; Kool, M.; Buckley, H. C.; Lambrecht, B. N.; O'Hare, D.; Austyn, J. M., Immunity induced by a broad class of inorganic crystalline materials is directly controlled by their chemistry. *Journal of Experimental Medicine* **2014**, *211* (6), 1019-1025.

53. Li, A.; Qin, L.; Zhu, D.; Zhu, R.; Sun, J.; Wang, S., Signalling pathways involved in the activation of dendritic cells by layered double hydroxide nanoparticles. *Biomaterials* **2010**, *31* (4), 748-756.

54. Gomez-Morales, J.; Iafisco, M.; Delgado-Lopez, J. M.; Sarda, S.; Drouet, C., Progress on the preparation of nanocrystalline apatites and surface characterization: Overview of fundamental and applied aspects. *Progress in Crystal Growth and Characterization of Materials* **2013**, *59* (1), 1-46.

55. Drouet, C.; Choimet, M.; Simon, M.; Devès, G.; Barberet, P.; Seznec, H.; Rassu, G.; Marsan, O.; Tourrette, A., Colloidal Apatite Particles : A Multifunctional Platform in (Nano)Medicine. *Juniper Online Journal of Materials Science* **2020**, 6(1): 555676.

56. Relyveld, E. H.; Henocq, E.; Raynaud, M., Study of the antidiphtheria vaccination of allergic subjects with a pure anatoxin absorbed on calcium phosphate. *Bulletin of the World Health Organization* **1964**, *30* (3), 321-325.

57. Jiang, D.; Premachandra, G. S.; Johnston, C.; Hem, S. L., Structure and adsorption properties of commercial calcium phosphate adjuvant. *Vaccine* **2004**, *23* (5), 693-8.

58. Relyveld, E. H., Preparation and use of calcium phosphate adsorbed vaccines. *Dev Biol Stand* **1986**, *65*, 131-6.

59. Vandecandelaere, N.; Rey, C.; Drouet, C., Biomimetic apatite-based biomaterials : on the critical impact of synthesis and post-synthesis parameters. *J. Mater. Sci: Mater. Med* **2012**, *23*, 2593-2606.

60. He, Q.; Mitchell, A. R.; Johnson, S. L.; Wagner-Bartak, C.; Morcol, T.; Bell, S. J., Calcium phosphate nanoparticle adjuvant. *Clinical and diagnostic laboratory immunology* **2000**, *7* (6), 899-903.

61. Morçöl, T.; Hurst, B. L.; Tarbet, E. B., Calcium phosphate nanoparticle (CaPNP) for dosesparing of inactivated whole virus pandemic influenza A (H1N1) 2009 vaccine in mice. *Vaccine* **2017**, *35* (35 Pt B), 4569-4577.

62. Morcol, T.; Nagappan, P.; Nerenbaum, L.; Mitchell, A.; Bell, S., Particulate Drug Delivery Systems for Protein Drugs: An overview of Noninvasive Insulin Delivery Using Calcium Phosphate Particle Technology. **2008**; pp 223-241.

63. Bell, S. J. D.; Morcol, T.; He, Q. Therapeutic calcium phosphate particles and methods of manufacture and use. US Patent US8431221B2.

64. Grunenwald, A.; Keyser, C.; Sautereau, A. M.; Crubézy, E.; Ludes, B.; Drouet, C., Adsorption of DNA on biomimetic apatites: Toward the understanding of the role of bone and tooth mineral on the preservation of ancient DNA. *Applied Surface Science* **2014**, *292*, 867-875.

65. Choimet, M.; Tourrette, A.; Drouet, C., Adsorption of nucleotides on biomimetic apatite: The case of cytidine 5' monophosphate (CMP). *Journal of Colloid and Interface Science* **2015**, 456, 132-137.

66. Hammami, K.; Feki, H. E.; Marsan, O.; Drouet, C., Adsorption of nucleotides on biomimetic apatite: The case of adenosine 5' monophosphate (AMP). *Applied Surface Science* **2015**, *353*, 165-172.

67. Welzel, T.; Radtke, I.; Meyer-Zaika, W.; Heumann, R.; Epple, M., Transfection of cells with custom-made calcium phosphate nanoparticles coated with DNA. *Journal of Materials Chemistry* **2004**, *14* (14), 2213-2217.

68. Joyappa, D. H.; Ashok Kumar, C.; Banumathi, N.; Reddy, G. R.; Suryanarayana, V. V. S., Calcium phosphate nanoparticle prepared with foot and mouth disease virus P1-3CD gene construct protects mice and guinea pigs against the challenge virus. *Veterinary Microbiology* **2009**, *139* (1), 58-66.

69. Miragoli, M.; Ceriotti, P.; Iafisco, M.; Vacchiano, M.; Salvarani, N.; Alogna, A.; Carullo, P.; Ramirez-Rodriguez, G. B.; Patricio, T.; Esposti, L. D.; Rossi, F.; Ravanetti, F.; Pinelli, S.; Alinovi, R.; Erreni, M.; Rossi, S.; Condorelli, G.; Post, H.; Tampieri, A.; Catalucci, D., Inhalation of peptide-loaded nanoparticles improves heart failure. *Sci Transl Med* **2018**, *10* (424).

70. He, Q.; Mitchell, A.; Morcol, T.; Bell, S. J. D., Calcium phosphate nanoparticles induce mucosal immunity and protection against herpes simplex virus type 2. *Clinical and diagnostic laboratory immunology* **2002**, *9* (5), 1021-1024.

71. Wang, X.; Li, X.; Onuma, K.; Sogo, Y.; Ohno, T.; Ito, A., Zn- and Mg- containing tricalcium phosphates-based adjuvants for cancer immunotherapy. *Sci Rep* **2013**, *3*, 2203.

72. Ciocca, D. R.; Frayssinet, P.; Cuello-Carrión, F. D., A pilot study with a therapeutic vaccine based on hydroxyapatite ceramic particles and self-antigens in cancer patients. *Cell stress & chaperones* **2007**, *12* (1), 33-43.

73. Lee, D. D.; Aiolova, M. Calcium phosphate delivery vehicle and adjuvant. US Patent Application 20030082232, A1.

74. Weber, C. G.; Mueller, M.; Vandecandelaere, N.; Trick, I.; Burger-Kentischer, A.; Maucher, T.; Drouet, C., Enzyme-functionalized biomimetic apatites: concept and perspectives in view of innovative medical approaches. *Journal of Materials Science-Materials in Medicine* **2014**, *25*, 595-606.

75. Al-Kattan, A.; Errassifi, F.; Sautereau, A.-M.; Sarda, S.; Dufour, P.; Barroug, A.; Santos, I. D.; Combes, C.; Grossin, D.; Rey, C.; Drouet, C., Medical potentialities of biomimetic apatites through adsorption, ionic substitution, and mineral/organic associations: Three illustrative examples. *Advanced Engineering Materials* **2010**, *12* (7), B224-B233.

76. Sarda, S.; Iafisco, M.; Pascaud-Mathieu, P.; Adamiano, A.; Montesi, M.; Panseri, S.; Marsan, O.; Thouron, C.; Dupret-Bories, A.; Tampieri, A.; Drouet, C., Interaction of Folic Acid with Nanocrystalline Apatites and Extension to Methotrexate (Antifolate) in View of Anticancer Applications. *Langmuir* **2018**, *34* (40), 12036-12048.

77. lafisco, M.; Drouet, C.; Adamiano, A.; Pascaud, P.; Montesi, M.; Panseri, S.; Sarda, S.; Tampieri, A., Superparamagnetic iron-doped nanocrystalline apatite as a delivery system for doxorubicin. *Journal of Materials Chemistry B* **2016**, *4* (1), 57-70.

78. lafisco, M.; Varoni, E.; Di Foggia, M.; Pietronave, S.; Fini, M.; Roveri, N.; Rimondini, L.; Prat, M., Conjugation of hydroxyapatite nanocrystals with human immunoglobulin G for nanomedical applications. *Colloids and Surfaces B: Biointerfaces* **2012**, *90* (1), 1-7.

79. Cazalbou, S. ; Bertrand, G. ; Drouet, C. Tetracycline-loaded biomimetic apatite: an adsorption study. *J Phys Chem B*. **2015**, 119(7),3014-24.