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#### An individual based model for the eco-evolutionary emergence of 1 bipartite interaction networks 2 3 November 2, 2020 4 Odile Maliet<sup>a,\*</sup>, Nicolas Loeuille<sup>b</sup>, Hélène Morlon<sup>a</sup> 5 <sup>a</sup> Institut de biologie de l'Ecole normale supérieure (IBENS), Ecole normale supérieure, CNRS, INSERM, PSL 6 Research University, 75005 Paris, France 7 <sup>b</sup> Sorbonne Université, UPEC, CNRS, IRD, INRA, Institut d'Ecologie et des Sciences de l'Environnement, IEES, 8 F-75005 Paris, France q \* Corresponding author. email: odile.maliet@orange.fr 10

# <sup>11</sup> Authors contribution

OM, NL & HM conceived the study. OM wrote the simulation code and performed the analysis.
 OM, NL & HM wrote the manuscript.

## <sup>14</sup> 1 Data availability statement

Data sharing not applicable to this article as no new data were generated during the current study. The functions for running the simulations are available in the Rpackage RPANDA (Morlon et al., 2016).

## <sup>18</sup> 2 Short running title

<sup>19</sup> Emergence of bipartite interaction networks

## 20 3 Keywords

ecological networks ; mutualism ; antagonism ; nestedness ; coevolution

### 22 4 Count

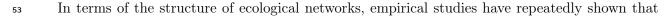
- 23 Main text : 5468 words
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### 28 Abstract

How ecological interaction networks emerge on evolutionary time scales remains unclear. Here 29 we build an individual-based eco-evolutionary model for the emergence of mutualistic, antagonis-30 tic and neutral bipartite interaction networks. Exploring networks evolved under these scenarios. 31 we find three main results. First, antagonistic interactions tend to foster species and trait diver-32 sity, while mutualistic interactions reduce diversity. Second, antagonistic interactors evolve higher 33 specialization, which results in networks that are often more modular than neutral ones; resource 34 species in these networks often display phylogenetic conservatism in interaction partners. Third, 35 mutualistic interactions lead to networks that are more nested than neutral ones, with low phylo-36 genetic conservatism in interaction partners. These results tend to match overall empirical trends. 37 demonstrating that structures of empirical networks that have most often been explained by eco-38 logical processes can result from an evolutionary emergence. Our model contributes to the ongoing 39 effort of better integrating ecological interactions and macroevolution. 40

# 41 Introduction

Species in ecological communities engage in a diverse set of antagonistic and mutualistic interac-42 tions such as predation, parasitism, pollination and seed dispersal. These interactions are thought 43 to have important consequences for species and trait diversity (Ehrlich and Raven, 1964; Van Valen, 44 1973; Hembry et al., 2014), as well as the structure of ecological networks (Bascompte and Jordano, 45 2007; Thébault and Fontaine, 2010; Suweis et al., 2013; Dormann et al., 2017). In terms of diversity, 46 empirical examples of very diverse communities seem more numerous in antagonistic (e.g. plant de-47 fense strategies, Futuyma and Agrawal, 2009) than mutualistic interactions (Hembry et al., 2014; 48 Chomicki et al., 2019). Eco-evolutionary models have indeed found that antagonistic interactions 49 foster, while mutualistic interactions impede, trait diversity (Yoder and Nuismer, 2010). If and how 50 antagonistic and mutualistic interactions affect the generation and maintenance of species diversity 51 is less clear (Weber et al., 2017). 52



this structure is highly non-random (Fontaine et al., 2011). Two main patterns have been reported: 54 modularity, with subsets of species interacting more strongly among each other than with the rest of 55 the community (May, 1972; Krause et al., 2003), and nestedness, with specialist species preferentially 56 interacting with generalists (Bascompte et al., 2003; Jordano et al., 2003; Lewinsohn et al., 2006; 57 Bascompte and Jordano, 2007; Thébault and Fontaine, 2010; Rohr et al., 2014). While several 58 factors play a role in explaining network structure, including intimacy (i.e. the degree of physical 59 proximity or integration of partner taxa, (Fontaine et al., 2011)) and phylogenetic scale (Beckett 60 and Williams, 2013), the nature of the interaction seems particularly important. In a comparison 61 of 95 networks, Fontaine et al. (2011) showed that non-intimate antagonistic networks are often 62 modular, while mutualistic networks are often nested, although there are clearly deviations from 63 this general pattern (Olesen et al., 2007; Pilosof et al., 2014). Many studies have sought to explain 64 the processes driving these non-random structures without reaching consensus. 65

One of the main hypotheses put forward for explaining non-random network structures is the 66 stability hypothesis. Species rich communities are not stable when species interact at random 67 (May, 1972; Krause et al., 2003; Jordano et al., 2003; Montova et al., 2006); this instability is 68 counteracted by nonrandom network structures that depend on the type of interaction (Fontaine 69 et al., 2011). In particular, stable species coexistence is reached in modular networks in the case of 70 antagonistic interactions, and in nested networks in the case of mutualistic interactions (Thébault 71 and Fontaine, 2010). Nestedness reduces effective interspecific competition (Bastolla et al., 2009) 72 and broadens the range of ecological conditions under which mutualistic species can coexist (Rohr 73 et al., 2014; Saavedra et al., 2016; Grilli et al., 2017), especially in the presence of adaptive foraging 74 (Valdovinos et al., 2016). Anti-modularity has a destabilizing effect in most ecological communities 75 (Grilli et al., 2016). Nestedness in mutualist networks and modularity in antagonist networks also 76 limit co-extinction cascades (May, 1972; Krause et al., 2003; Memmott et al., 2004; Stouffer and 77 Bascompte, 2011). 78

A second family of hypotheses to explain nonrandom, interaction-dependent structures, relies on trait-dependent interactions. For example, Santamaría and Rodríguez-Gironés (2007) and Rezende et al. (2007a) showed that interactions determined by either phenotypic difference or phenotypic

similarity can generate nested networks, in particular when several traits are involved and when 82 there is a phylogenetic signal in trait values. de Andreazzi et al. (2019) also recently showed 83 that a strong effect of species trait values on the probability for two species to interact helps in 84 explaining network structure, and in particular that trait matching fosters trait coevolution and 85 helps in explaining the structure of antagonistic networks. Related to the trait-based hypothesis is 86 the observation that the non-random structure of networks can emerge without being selected for 87 when interaction strength is inherited from the parent species (the "network spandrel" hypothesis 88 Maynard et al., 2018; Valverde et al., 2018). 89

Finally, differences in abundances across species can on their own generate non-random struc-90 tures. In a neutral world where individuals encounter and interact at random with one another, 91 individuals from rare specialist species are more likely to interact with individuals from abundant 92 generalists. As species abundance distributions are generally imbalanced, with many rare species 93 and a few abundant ones, this leads to nested networks (Vázquez, 2005; Vázquez et al., 2009; San-94 tamaría and Rodríguez-Gironés, 2007; Krishna et al., 2008; Staniczenko et al., 2013; Coelho and 95 Rangel, 2018). Hence, while the often-observed modularity of antagonistic networks cannot be ex-96 plained by neutral processes alone, the frequent nestedness of mutualistic networks can be a pure 97 result of neutral encounters. 98

Studies seeking to explain the non-random structure of species interaction networks have rarely 99 focused on their emergence over evolutionary time scales, despite empirical evidence that there are 100 evolutionary (phylogenetic) constraints to who interacts with whom (Rezende et al., 2007b; Elias 101 et al., 2013). They have instead often fixed the ecological context, for example by assuming an initial 102 network is subject to local extinctions (Thébault and Fontaine, 2010), or fixing the number of species 103 and either the species abundance distribution (Nuismer et al., 2013) or the species trait distribution 104 (Santamaría and Rodríguez-Gironés, 2007). There are notable exceptions though. Minoarivelo and 105 Hui (2016) used an adaptive dynamics framework to show that coevolution can lead to both nested 106 and modular networks for mutualistic interactions. In de Andreazzi et al. (2019), the authors 107 investigated the effect of trait coevolution on network structure, but with the number of both 108 species and interactions fixed according to empirical networks. In Poisot and Stouffer (2016), the 109

authors fitted to empirical networks a macroevolutionary model formalizing the evolution of species 110 interactions, and surprisingly did not detect major differences between antagonistic and mutualistic 111 networks. A recent study showed that neutral networks can evolve nested structures, probably as a 112 result of asymmetric abundances, but did not investigate the effect of mutualistic and antagonistic 113 interactions (Coelho and Rangel, 2018). Finally, Maynard et al. (2018) and Valverde et al. (2018) 114 showed that nestedness and modularity may emerge from speciation divergence dynamics within 115 antagonistic networks. The lack of a unified model mimicking the evolutionary emergence of species 116 interaction networks as species diversify clearly limits our understanding of the macroevolutionary 117 dynamics and consequences of inter-species interactions (Weber et al., 2017; Harmon et al., 2019). 118 Here we investigate the emergence of ecological networks as species coevolve, when individu-119 als engage into mutualistic, antagonistic or neutral interactions. We develop an individual-based, 120 stochastic eco-evolutionary model (BipartiteEvol) that allows us to assess the effect of interaction 121 type on species and trait diversity, network structure, and phylogenetic signal in interaction part-122 ners. We discuss results obtained when simulating the model in light of previous literature and 123 empirical observations. 124

### 125 Methods

# <sup>126</sup> An individual-based model for the eco-evolutionary emergence of bipartite in-<sup>127</sup> teraction networks

We develop an individual-based, stochastic model, with discrete time steps and fixed population 128 size, described in detail in our Supplementary Information. This model can be seen as an extension, 129 in two main directions, of the spatially explicit neutral model of biodiversity where dynamics are 130 modeled on a grid of N cells (Chave and Leigh Jr, 2002). As in the metacommunity version 131 of this model, our model best represents evolutionary outcomes in a closed community without 132 immigration. The first extension is that here each cell is occupied by two individuals, one from each 133 of two interaction guilds (guild A and B), and this co-occurrence determines interaction (Canard 134 et al., 2014). The second extension is that each individual is characterized by its (potentially 135

multidimensional) trait value, and has fitness determined by both its trait value and that of the 136 individual from the other guild occupying the cell. Offspring experience mutations that induce 137 genetic differentiation and small trait deviations from their parents (Loeuille and Leibold, 2014). 138 Genetic differentiation eventually leads to speciation (Manceau et al., 2015; Rosindell et al., 2015). 139 The nature of the interactions can affect evolutionary outcomes in an eco-evolutionary feedback 140 loop between who interacts with whom, the resulting adaptive pressures, and the gain or loss of 141 interactions linked to adaptations to these pressures. At each time step, we track the trait values 142 of each individual, their genealogical relationship, whether they experienced a mutation, and who 143 they interact with. At the end of the simulations, we construct the resulting species phylogenies 144 and species interaction networks. 145

*Eco-evolutionary dynamics* We start the simulations with a monomorphic population in each guild, all descending from a unique parent. At each time step we update the grid as follows:

- We select one individual from guild A at random and kill this individual. We record the trait value  $x_B$  of the individual of guild B present on this cell.
- We compute the fitness of all individuals of guild A present in the grid, should it interact with an individual of trait  $x_B$ .
- We select an individual from guild A to be the parent of the individual that replaces the killed
  one with probabilities proportional to each individual fitness. Hence, the filiation of the new
  individual depends on trait values and chance. We do not introduce dispersal limitation here,
  but this could be done in future work by selecting the parent with a probability proportional
  to its fitness times a dispersion kernel.
- The new individual from guild A has a probability  $\mu_A$  to experience a mutation, in which case its new trait is drawn independently in each dimension in a normal distribution centered on the parental trait with standard deviation  $\sigma$ . If no mutation occurs, the new individual inherits the trait value of its parent.
- We repeat the four previous steps for guild *B*.

6

• We record the genealogy obtained in each of the two guilds, as well as the mutations that occurred on this genealogy.

Modeling the effect of trait-specific interactions on fitness Each individual is characterized by 164 its trait value x, which can have any dimension d. The distance in trait space of two interacting 165 individuals determines the effect of the interaction on their fitness. Classical traits with fitness 166 effects include proboscis and floral tube length, color preferences, and organism sizes or phenology 167 in mutualist partners such as plants and their pollinators, or the presence of metabolic compounds 168 and the ability to metabolize these compounds in antagonist partners such as plants and their 169 herbivores. For example, a mismatch in phenology between a plant and its pollinator would induce 170 a cost in reproductive success for the plant and in the ability to feed for the pollinator. We use a 171 classical trait matching expression given by a Gaussian function, with maximal fitness effect when 172 the traits of two interacting individuals are similar (Fig. S1, Loeuille and Loreau (2005); Yoder 173 and Nuismer (2010)). The fitness function is parametrized by  $\alpha$ , which measures the specificity of 174 the trait matching: high  $\alpha$  values correspond to scenarios where fitness effects are preponderant in 175 interactors with very similar trait values (i.e. highly specialized interactions), while low  $\alpha$  values 176 correspond to more neutral scenarios. Thereafter for simplicity we refer to  $\frac{1}{\alpha}$  as "niche width". The 177 fitness function is further parametrized by a parameter r, the ratio between the maximum and 178 minimum fitness, that measures the overall effect of trait differences. 179

In mutualistic interactions, individuals from both guilds have higher fitness  $(W_A \text{ and } W_B)$  when they have similar trait values:

$$W_A(x_A, x_B) = \frac{1}{r_A - 1} + e^{-\|x_A - x_B\|^2 \times (\alpha_A^2/2)}$$

$$W_B(x_A, x_B) = \frac{1}{r_B - 1} + e^{-\|x_A - x_B\|^2 \times (\alpha_B^2/2)}$$
(1)

In antagonistic interactions, individuals from the consumer clade (B) have higher, and those from the resource clade (A) lower, fitness when they have similar trait values, so the fitness differs from the mutualistic scenarios only for clade A:

$$W_A(x_A, x_B) = \frac{1}{r_A - 1} + 1 - e^{-\|x_A - x_B\|^2 \times (\alpha_A^2/2)}$$

$$W_B(x_A, x_B) = \frac{1}{r_B - 1} + e^{-\|x_A - x_B\|^2 \times (\alpha_B^2/2)}$$
(2)

We obtained the neutral case by taking  $\alpha = 0$  in the expression for mutualistic fitness (taking  $\alpha = 0$  in the antagonistic version would yield identical simulations).

Defining species and phylogenies Based on the resulting genealogies with mutations, we define 187 species following the model of Speciation by Genetic Differentiation (Manceau et al., 2015), except 188 that we allow s, the number of mutations needed to belong to different species, to vary (similarly 189 to what is done in Rosindell et al. (2015)). Species are thus the smallest monophyletic group of 190 individuals from the genealogy such that two individuals separated by less than s mutations belong 191 to the same species. This is a protracted mode of speciation, and the time needed for speciation 192 to complete increases with parameter s. This species definition allows for polymorphic species. 193 We compute the resulting species-level phylogenies from the genealogy with mutation positions 194 (Manceau et al., 2015). Speciation occurs as the result of accumulating mutations, whatever their 195 effect on the phenotype. It can thus merely result from drift, and we expect species diversity to 196 build up under neutral dynamics. However, if trait diversity is favored – as would be the case if the 197 presence of predators generates disruptive selection – we expect to see an increased species diversity 198 as more mutations are likely to get fixed. 199

From individuals' interaction to species interaction networks The interaction network is defined 200 at the scale of the entire grid and is based on individuals' co-occurrence: we consider that two 201 individuals interact if they co-occur in the same cell. Next, at the species level, we consider both 202 a quantitative and a binary network. For the quantitative network, we take the strength of the 203 interaction between two species to be the number of pairs of individuals of these species interacting 204 together. For the binary network, we consider that two species interact if at least one pair of 205 individuals of these species interact (that is if their interaction strength in the quantitative network 206 is nonzero). While interactions in our networks are constructed based on only co-occurrence, they 207 depend on trait values through the effect of traits on invasion probabilities. An interaction network 208

in our model thus results from the cumulative effects of smaller scale interaction events (as for example in Pillai et al., 2011).

We provide codes for running simulations of BipartiteEvol in RPANDA (Morlon et al., 2016, function sim.BipartiteEvol).

#### 213 Analyzing emerging patterns

In order to explore emerging patterns, we performed a series of simulations under neutral, 214 mutualistic and antagonistic scenarios. We followed the evolution of a (closed) community made of 215 N = 4000 individuals in each guild during 8e7 death events. This simulation duration was enough 216 for most of the simulations to have reached stationary state, at least for the summary statistics we 217 followed (Fig. S2-S3). We ran most of our simulations with a trait dimension d = 3, as ecological 218 networks are thought to be best described by traits with several vet few dimensions (Eklöf et al., 219 2013). To check the robustness of our result to trait dimensionality, we also ran simulations for 220 d = 1 and d = 10 for a selected parameter set (Fig S28-S34). We chose initial trait values 2 and 0 221 for guild A and B in all trait dimensions. We also chose a standard deviation  $\sigma = 1$ ; considering a 222 different  $\sigma$  would be equivalent to dividing  $\alpha_A$  and  $\alpha_B$  by  $\sigma$ . We held the mutation probabilities  $\mu_A$ 223 and  $\mu_B$  constant at 0.05. We fixed  $r_A$  at 10 and  $r_B$  at  $+\infty$ . In the case of mutualistic interactions, 224 this renders species from guild B obligate mutualists (their fitness is zero if they do not interact 225 with species from guild A, e.g. pollinators that entirely depend on a specific clade of plants) while 226 species from guild A are facultative mutualists (e.g. plants that can be pollinated by pollinators not 227 represented in B). In the case of antagonistic interactions,  $r_B = +\infty$  means that consumer species 228 entirely depend on interaction with their resources. Besides the neutral case ( $\alpha_A = \alpha_B = 0$ ), we 229 simulated all the combinations of  $\alpha_A$  and  $\alpha_B$  in 0.01, 0.02, 0.05, 0.1, 0.2, 0.5, 1, 10 for both mutualistic 230 and antagonistic interactions. We performed 10 simulations for each scenario and each parameter 231 set (20 in the neutral case). 232

At the end of each simulation, we built the resulting phylogenies and interaction networks using three different species definition thresholds s (s = 1, 10 and 50). We computed species richness, as well as trait diversity on each trait dimension as the variance in trait values across all individuals. In

addition, in order to investigate under which conditions trait coevolution between species on the two 236 sides of the network occurs, we measured the correlation in trait values of co-occurring individuals. 237 We computed nestedness and modularity using the R-package bipartite (Almeida-Neto and Ulrich. 238 2011; Dormann et al., 2008)). Nestedness was measured with the NODF metric using the func-239 tion nested, with method = "weighted NODF" for quantitative networks and method = "NODF2" 240 for binary networks. Modularity was computed for quantitative networks only, using the function 241 computeModules (the corresponding quanBiMo algorithm is not adapted to binary networks (Dor-242 mann and Strauss, 2014)). We compared the binary nestedness values to the distribution of values 243 obtained for two null models. In the first, thereafter called NM1, the network connectance is kept 244 constant (method "shuffle.web" in the function nullModel). In the second, thereafter called NM2, 245 row and column sums of the interaction matrix are kept constant (method "r2d"). This is equivalent 246 to randomly reassigning a position on the grid for all individuals regardless of their trait values, 247 which is also how we generate interactions in our neutral simulations and provides a way to cor-248 rect for species abundances. We also compared the quantitative nestedness and modularity values 249 to the distribution of values obtained for NM2 (NM1 is applicable to only binary networks). We 250 computed Z-scores, defined as  $\frac{x-\mu}{\sigma}$ , where for a given metric x is the value outputted by our simu-251 lation and  $\mu$  and  $\sigma$  are the mean and standard deviation of the values under the null model. While 252 Z-scores are not appropriate for comparing nestedness values between networks of different sizes 253 (Song et al., 2017), they are a good way of assessing the significance of nestdedness values against 254 a null model. Finally, we computed the phylogenetic signal of interaction partners using a Mantel 255 test that assesses the significance of the correlation between the phylogenetic distance of two species 256 and the dissimilarity of their interaction partners. In order to limit the effect of species definition 257 on our results, we used a phylogenetic metric to quantify this dissimilarity. We used the (weighted) 258 fraction of unshared phylogenetic branch length between the two sets of interaction partners (com-259 puted using quantitative uniFrac (Lozupone et al., 2007), "d 1" in the function GUniFrac from 260 the R package GUniFrac (Chen, 2012)). We also performed the analyses with a non-phylogenetic 261 metric, the Jaccard dissimilarity index – computed with the function cluster\_similarity from 262 the R-package clusteval – for comparison. 263

### 264 Results

Figure 1 shows two typical simulations, one mutualistic and the other antagonistic, that illustrate 265 general results concerning the evolution of trait values and species interaction networks. Other 266 figures in the main text show emergent properties of mutualistic, antagonistic and neutral networks 267 when the niche width of species from guild A (i.e. resources in antagonistic networks and facultative 268 mutualists in mutualistic networks) varies and that of guild B (i.e. consumers in antagonistic 269 networks and obligate mutualists in mutualistic networks) is fixed, and for three-dimensional traits. 270 Figures in the Supplementary Material report results for other parameter sets, including results 271 when the niche width of species from guild B varies and that of guild A is fixed, and for trait 272 dimension d = 1 and d = 10. Results for these trait dimensions were qualitatively similar (Fig 273 S28-S34). 274

### 275 Trait diversity

In mutualistic scenarios, trait values stay fairly constant through time (Fig. 1A) and trait 276 diversity is lower than in neutral scenarios (Fig. 2A & Fig. S4). Trait matching in mutualistic 277 networks results in stabilizing selection that constrains trait evolution on both sides of the network. 278 Trait diversity within a guild is generally constrained by niche width in this guild, but not by niche 279 width in the interacting guild, even if exceptions occur in extreme cases when niche width in one of 280 the two guilds is smaller than, or comparable to, the effect size of mutations (here fixed to  $\sigma = 1$ , 281 Fig. 2A & Fig. S4). The correlation between the traits of interacting individuals is slightly positive 282 but stays very low (Fig. 2B & Fig. S5). 283

Patterns are strikingly different in antagonistic scenarios, where clusters of traits progressively emerge from co-evolutionary dynamics (Fig. 1B). Disrupting selection acting on resource species typically increases trait diversity compared to neutral scenarios for both consumers and resources, with similar levels of diversity in the two guilds (Fig. 2A & Fig. S4). Trait diversity increases with the niche widths of both resource and consumer species, but collapses when consumers have a larger niche than resources (Fig. 2A & Fig. S4). The traits of interacting individuals are positively correlated, suggesting strong trait coevolution (Fig. 2B & Fig. S5).

#### <sup>291</sup> Species richness

Consistent with temporal dynamics of trait diversity, species diversification is lower in mutualistic than antagonistic networks (Fig. 1A versus B). Species richness is systematically larger in antagonistic than in mutualistic networks, regardless of niche width and species definition (Fig. 2C,D & Fig. S6-S9).

In mutualistic scenarios, species richness tends to be lower than in neutral scenarios (Fig. 2C,D & Fig. S6-S9). Species richness is comparable to what is obtained in the neutral case when species' niches are so large that individuals have approximately the same fitness regardless of who they interact with. But for narrower niches, trait matching is more influential and stabilizing selection inhibits speciation. Species richness within a guild thus decreases when niches in that guild become narrower (Fig. 2C,D & Fig. S6-S9).

In antagonistic scenarios, species richness is in general higher for resource than for consumer 302 species (Fig. 2C,D & S7-S9). The effect of niche width on species richness depends on the species 303 definition threshold s. For a low s, resource species richness is similar to what is obtained in neutral 304 simulations and unaffected by niche width of either resource or consumer species (Fig. 2C & S7). 305 Consumer species richness tends to be lower than in neutral simulations (Fig. 2C & S7). It is not 306 affected by the niche width of resource species (Fig. 2C) but decreases when the niches of consumer 307 species become narrower, as long as it remains larger than the effect size of mutations. For a higher 308 s (around 50, Fig. 2D & S9), species richness is affected by niche width in a similar way for resource 309 and consumer species, and can be either higher or lower than in neutral simulations. Cases when 310 species richness is higher than in neutral simulations correspond to scenarios with intermediate 311 resource niche width, and narrower niche widths for consumer than resource species (Fig. 2D, S9). 312

### 313 Network structure

Mutualistic interactions typically lead to the progressive emergence of nested networks (Figure 1A), while antagonistic interactions lead to modular networks (Figure 1B). When we look at network metrics without any comparison to a null model, most networks that have higher nestedness values than those in the neutral case were generated in mutualistic scenarios, while most of those that have higher modularity values than those in the neutral case were generated in antagonistic scenarios(Fig. 3A, Fig S10 & S15).

When compared to the null model that corrects for only connectance (NM1), all networks show up as significantly nested, including neutral ones (Fig. 3B, S12-S13 & S17-S18). Z-values are very high and mostly depend on the diversity of the community, with higher Z-values obtained for more diverse communities (Fig. S14).

When compared to the null model that corrects for abundance (NM2), neutral models are neither 324 nested nor anti-nested, and they are not modular either (Fig. 3C,D & Fig. S19-S20, S22-S23). 325 Mutualistic networks are similar to neutral ones. The only deviation occurs when the facultative 326 mutualists have an intermediate niche width that is larger than that of the obligate mutualists; 327 in this case, mutualistic networks are slightly anti-nested and modular (Fig. S19-S20, S22-S23). 328 Antagonistic networks deviate sharply from neutral ones; they most often exhibit a modular, anti-329 nested structure that peaks at intermediate resource niche width (Fig. 3C.D) and small consumer 330 niche width (Fig. S19, S22). The results stav qualitatively similar for s = 10 (Fig. S20, S23). 331

### 332 Phylogenetic signal

While modules in antagonistic networks seem strongly constrained by phylogenetic history (Fig. 333 1B), interactions in mutualistic networks seem rather independent from this history (Fig. 1A). 334 Indeed, the Mantel correlation between phylogenetic distance and similarity in interaction partners 335 is generally weak and non-significant on both sides of mutualistic networks, comparable to what is 336 found for neutral networks (Fig. 4A & B). The correlation is stronger in antagonistic networks, and 337 significant in many simulations (Fig. 4A,B). This phylogenetic signal is often stronger for resource 338 than for consumer species and higher for an intermediate consumer niche width and/or wider re-339 source niche width (Fig. 4B, Fig. S26). Results are qualitatively similar across species definition 340 thresholds and dissimilarity metrics, although fewer networks display significant correlation values 341 for s = 1 than for s = 10 and for the Jaccard than for the Unifrac metric (Fig. S24-S27). 342

### 343 Discussion

Our model allows a direct comparison of the effect of antagonistic, mutualistic, and neutral interactions on the dynamics of trait and species diversity as well as on the structure of the interaction network. We find that antagonistic interactions enhance both trait and species diversity, and result in modular interaction networks with phylogenetic signal in interaction partners. Mutualistic interactions instead limit trait and species diversity, and result in interaction networks that otherwise resemble neutral networks, with a tendency for nestedness rather than modularity, and low phylogenetic signal in interacting partners.

We discuss these results in light of empirical observations, keeping in mind that our model 351 best mimics eco-evolutionary dynamics in a closed metacommunity, where diversity is the result 352 of speciation-extinction dynamics, while most empirical networks are built at the community level, 353 where diversity can also be introduced through immigration, which can change qualitative patterns 354 (as in Maynard et al., 2018). Similar to what has been done in developments of the neutral bio-355 diversity model, our model could be developed to include dispersal limitation and to sample local 356 communities at different spatial scales, or as a continent-island model, where the local community 357 receives immigrants from the metacommunity. This would also help understand how local networks 358 assemble from 'meta-networks' (Gravel et al., 2011; Morlon et al., 2014). Other limitations of our 359 model include the 'zero-sum' assumption that total population size remains constant, not account-360 ing for sexual reproduction, and not varying the degree of intimacy, which can all affect network 361 structure. 362

Our result that antagonistic interactions tend to enhance, while mutualistic ones impede, trait 363 and species diversity, is generally consistent with previous empirical and theoretical studies (Ehrlich 364 and Raven, 1964; Yoder and Nuismer, 2010; Janz, 2011; Hembry et al., 2014). For example, previous 365 studies suggest that investment in defense traits results in higher diversification rates in North 366 American milkweeds (Agrawal et al., 2009), while specialized pollination in Yucca-moth interactions 367 does not increase Yucca diversification (Smith et al., 2008). Consistently, Armbruster and Muchhala 368 (2009) showed that in several groups of angiosperms, diversity promotes floral specialization – 369 through character displacement – rather than the reverse. Still, we do not exclude the possibility 370

that in some systems mutualism has a positive impact on diversification, as has been suggested by 371 other studies, such as in the case of fig-wasp interactions (Cruaud et al., 2012). Indeed our model 372 may output different results regarding the effect of mutualism if we allowed total population sizes 373 to vary and/or if we accounted for dispersal limitation and/or sexual reproduction. Mutualistic 374 interactions may indeed increase the density of individuals a community can sustain, either through 375 more efficient feeding and reproduction or by opening new adaptive zones. This could in turn allow 376 the maintenance of more diverse communities (Rosenzweig et al., 1995; Emerson and Kolm, 2005; 377 Joy, 2013). It has also been proposed that geographical isolation is necessary for mutualism to 378 promote speciation, and our non-spatial model cannot account for this potential effect (Thompson 379 and Cunningham, 2002; Kay and Sargent, 2009). In the case of pollination, mutualistic interactions 380 can also facilitate reproductive isolation, which cannot be modeled by our asexual model (van der 381 Niet and Johnson, 2012). Our results suggest that mutualism on its own, in the absence of such 382 mechanisms, is unlikely to promote diversity. 383

Our model generates clear structural differences between antagonistic and mutualistic com-384 munities. We find that antagonist interactions generate modular networks, regardless of whether 385 modularity values are corrected for abundance or not. This modularity emerges as a response to 386 reciprocal specialization and coevolution between resources and consumers, as demonstrated by the 387 positive correlation between the traits of interacting partners. Our results regarding nestedness are 388 much more contingent on whether or not nestedness values are corrected for connectance and/or 389 abundance. Raw nestedness values are higher in mutualistic than neutral (and antagonistic) com-390 munities, as observed in empirical networks (Thébault and Fontaine, 2010; Fontaine et al., 2011). 391 When we do not correct for species abundances, all the networks, including neutral and antagonis-392 tic networks, are significantly nested. Finally, when we correct for species abundances, nestedness 393 values are non-significant for neutral communities, and either non-significant or significantly lower 394 than those obtained for the null model for antagonist (and a few mutualist) communities. These 395 results are consistent with the literature, as most empirical or theoretical studies that found signif-396 icant nestedness in bipartite networks did not use a null model correcting for species abundances 397 (Thébault and Fontaine, 2010; Lewinsohn et al., 2006; Bascompte et al., 2003), while those that 398

corrected for abundances did not find a clear nested pattern (Vázquez, 2005; Staniczenko et al., 399 2013; Canard et al., 2014). The nestedness signal observed in bipartite networks may thus well 400 be linked to uneven species abundance distributions, in agreement with the neutral hypothesis 401 (Vázquez, 2005; Vázquez et al., 2009; Santamaría and Rodríguez-Gironés, 2007; Krishna et al., 402 2008; Staniczenko et al., 2013; Coelho and Rangel, 2018). In this case, understanding differences in 403 raw nestedness values between mutualist, neutral, and antagonist communities boils down to under-404 standing why communities are increasingly asymmetrical in abundances as we move from antagonist 405 to neutral and finally mutualist communities. Under our model, the relatively low asymmetry of 406 species abundances in antagonist networks could be linked to the coexistence of several abundant 407 species facilitated by modular structures. The strong asymmetry of species abundances in mutualist 408 networks could be linked to the transient nature of many rare species; we indeed often observed a 409 single adaptive optimum in our simulated mutualist networks, suggesting that mutants could fre-410 quently be maladapted. In addition to this dominating effect of abundance, trait-based preferential 411 interactions reduce nestedness in antagonist communities, while this effect is generally insignificant 412 in mutualist communities in which trait values are quite constrained. The differences in network 413 structure between antagonistic and mutualistic communities generated by our model are consis-414 tent with the classical dichotomy between antagonistic and mutualistic structures (Thébault and 415 Fontaine, 2010; Fontaine et al., 2011). While many empirical examples show that this dichotomy is 416 not as general as once thought (Pilosof et al., 2014; Olesen et al., 2007), our model shows that it can 417 emerge from simple evolutionary rules at the individual level. Relaxing simplifying hypotheses of 418 our model could provide a more nuanced view. For example, sexual reproduction, spatial structure, 419 fluctuation in population sizes and intimacy could generate modular mutualistic communities, as is 420 sometimes observed in empirical data (Olesen et al., 2007), by allowing trait diversification and the 421 creation of clusters in trait space. 422

Our results regarding phylogenetic signal, which measures evolutionary conservatism in interaction partners, are also consistent with empirical observations for antagonistic communities, in which resource species generally show a stronger phylogenetic signal than consumer species, once again in accordance to empirical observations (Krasnov et al., 2012; Elias et al., 2013; Fontaine and

Thébault, 2015). In plant-herbivore systems, the stronger conservatism in interaction partners seen 427 in plants has sometimes been interpreted as the combination of factors acting on both sides of the 428 network. Chemical defenses in plants are difficult and therefore slow to evolve and thus display a 429 high phylogenetic conservatism. In herbivores on the contrary, resource shifts among close relatives 430 to avoid enemies and/or reproductive interference are frequent, reducing phylogenetic conservatism 431 (Fontaine and Thébault, 2015). However, in our simulations trait evolutionary rates, which are 432 controlled by the mutation probability and the effect size of mutations, were symmetrical between 433 resource and consumer species. We did not include enemies or reproductive interference either. We 434 instead suggest that the difference in phylogenetic signal between resource and consumer species 435 is linked to a difference in the nature of the selection pressure they experience. Resource species 436 experience a selection pressure to avoid consumption, and they can evolve in almost any direction to 437 escape consumers (only the few directions in trait space favorable to consumers must be avoided). 438 Consumers instead experience a selection pressure to evolve specific traits adapted to consuming 439 the existing resource species, and it is thus not uncommon to see resource shifts and convergence 440 in traits, even between rather phylogenetically distant species, which weakens phylogenetic signal. 441 In mutualistic communities, our model generally does not display a significant phylogenetic signal. 442 While a tendency towards lower phylogenetic signal in mutualist compared to antagonistic commu-443 nities has been observed in empirical communities (Fontaine and Thébault, 2015), it is significant 444 (Rezende et al., 2007b). This disconnect between our model and empirical networks is likely linked 445 to the restricted evolved trait range produced by our mutualistic model, and the result could change 446 if sexual reproduction, spatial structure and/or variable population size were accounted for. 447

The general qualitative patterns discussed above hold for a large range of parameter space with a noticeable effect of niche width – which in our model is closely related to selection pressures – on the values of the different metrics we measured. In antagonistic communities, we found the strongest patterns – for all observed metrics, including diversity, network structure and phylogenetic signal – when niche width is larger for resources than for consumers. This situation corresponds to the case when selection pressures promote the evolution of specialization to specific resources in consumers. As detailed above, this specialization leads to modular structures, and imposes a strong

disruptive selection pressure on resources that enhances diversity and strengthens phylogenetic signal 455 by preventing trait convergence. In mutualistic scenarios, niche width in one guild impacts mostly 456 trait and species diversity in that guild, but not in the interacting guild; small niche width indeed 457 results in stronger stabilizing selection that constrains diversity, but does not affect the strength 458 of stabilizing selection in the interacting guild. The only cases when niche width impacts network 459 structure in mutualist networks occur when there is a very high asymmetry in niche width between 460 the two guilds, in which case species from the small niche guild will evolve specialization in response 461 to the high trait variability in the large niche guild, leading to the emergence of modular networks 462 that resemble these seen in antagonistic scenarios, or some empirical mutualistic networks Olesen 463 et al. (2007). The patterns were robust to trait dimensionality, which we did not expect considering 464 previous literature (Gilman et al., 2012; Ispolatov et al., 2016). Future work could investigate this 465 aspect more thoroughly, along with adding limits to the trait space and/or a correlation between 466 the traits. Trait correlations could produce interesting emerging behaviors, especially if they are 467 asymmetrical between the two guilds. 468

Our eco-evolutionary model allowed us to study the effect of different types of interactions 469 on the emergence of ecological networks as species and traits diversify. Despite the simplicity of 470 the processes involved, the model generated clear differences between mutualistic and antagonistic 471 communities that are broadly consistent with empirical observations. There is a lot of room for 472 future developments, such as accounting for geographical processes (Thompson, 2005), dispersal 473 from meta- to local communities, and the simultaneous effects of different interaction types (Fontaine 474 et al., 2011; Montesinos-Navarro et al., 2017). Developing an inference tool associated to this model 475 would also be an important step to quantify, for example, the actual effect of trait matching or trait 476 differences on selection pressures and trait coevolution from empirical data (Manceau et al., 2017) 477 . Such efforts are required if we want to better understand the macroevolutionary consequences of 478 species interactions (Weber et al., 2017; Harmon et al., 2019). 479

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### 487 References

- 488 Agrawal, A. A., M. Fishbein, R. Halitschke, A. P. Hastings, D. L. Rabosky, and S. Rasmann. 2009.
- Evidence for adaptive radiation from a phylogenetic study of plant defenses. Proceedings of the
  National Academy of Sciences 106:18067–18072.
- <sup>491</sup> Almeida-Neto, M. and W. Ulrich. 2011. A straightforward computational approach for measuring
  <sup>492</sup> nestedness using quantitative matrices. Environmental Modelling & Software 26:173–178.
- Armbruster, W. S. and N. Muchhala. 2009. Associations between floral specialization and species
  diversity: cause, effect, or correlation? Evolutionary Ecology 23:159.
- Bascompte, J. and P. Jordano. 2007. Plant-animal mutualistic networks: the architecture of biodiversity. Annu. Rev. Ecol. Evol. Syst. 38:567–593.
- <sup>497</sup> Bascompte, J., P. Jordano, C. J. Melián, and J. M. Olesen. 2003. The nested assembly of plant–
  <sup>498</sup> animal mutualistic networks. Proceedings of the National Academy of Sciences 100:9383–9387.
- Bastolla, U., M. A. Fortuna, A. Pascual-García, A. Ferrera, B. Luque, and J. Bascompte. 2009. The
  architecture of mutualistic networks minimizes competition and increases biodiversity. Nature
  458:1018–1020.
- Beckett, S. J. and H. T. Williams. 2013. Coevolutionary diversification creates nested-modular
   structure in phage-bacteria interaction networks. Interface focus 3:20130033.

- <sup>504</sup> Canard, E., N. Mouquet, D. Mouillot, M. Stanko, D. Miklisova, and D. Gravel. 2014. Empirical
  <sup>505</sup> evaluation of neutral interactions in host-parasite networks. The American Naturalist 183:468–
  <sup>506</sup> 479.
- <sup>507</sup> Chave, J. and E. G. Leigh Jr. 2002. A spatially explicit neutral model of  $\beta$ -diversity in tropical <sup>508</sup> forests. Theoretical population biology 62:153–168.
- <sup>509</sup> Chen, J. 2012. Gunifrac: generalized unifrac distances. R package version 1:2012.
- <sup>510</sup> Chomicki, G., M. Weber, A. Antonelli, J. Bascompte, and E. T. Kiers. 2019. The impact of mutu<sup>511</sup> alisms on species richness. Trends in ecology & evolution .
- <sup>512</sup> Coelho, M. T. P. and T. F. Rangel. 2018. Neutral community dynamics and the evolution of species
  <sup>513</sup> interactions. The American Naturalist 191:421–434.
- 514 Cruaud, A., N. Rønsted, B. Chantarasuwan, L. S. Chou, W. L. Clement, A. Couloux, B. Cousins,
- G. Genson, R. D. Harrison, P. E. Hanson, et al. 2012. An extreme case of plant-insect codiversification: figs and fig-pollinating wasps. Systematic Biology 61:1029–1047.
- de Andreazzi, C. S., J. Astegiano, and P. R. Guimarães Jr. 2019. Coevolution by different functional
  mechanisms modulates the structure and dynamics of antagonistic and mutualistic networks.
  Oikos .
- Dormann, C. F., J. Fründ, and H. M. Schaefer. 2017. Identifying causes of patterns in ecological
  networks: opportunities and limitations. Annual Review of Ecology, Evolution, and Systematics
  48:559–584.
- <sup>523</sup> Dormann, C. F., B. Gruber, and J. Fründ. 2008. Introducing the bipartite package: analysing <sup>524</sup> ecological networks. interaction 1:0–2413793.
- Dormann, C. F. and R. Strauss. 2014. A method for detecting modules in quantitative bipartite
   networks. Methods in Ecology and Evolution 5:90–98.
- Ehrlich, P. R. and P. H. Raven. 1964. Butterflies and plants: a study in coevolution. Evolution
  18:586–608.

- Eklöf, A., U. Jacob, J. Kopp, J. Bosch, R. Castro-Urgal, N. P. Chacoff, B. Dalsgaard, C. Sassi,
  M. Galetti, P. R. Guimaraes, et al. 2013. The dimensionality of ecological networks. Ecology
  letters 16:577–583.
- Elias, M., C. Fontaine, and F. F. van Veen. 2013. Evolutionary history and ecological processes
  shape a local multilevel antagonistic network. Current Biology 23:1355–1359.
- <sup>534</sup> Emerson, B. C. and N. Kolm. 2005. Species diversity can drive speciation. Nature 434:1015.
- Fontaine, C., P. R. Guimarães, S. Kéfi, N. Loeuille, J. Memmott, W. H. van Der Putten, F. J. van
  Veen, and E. Thébault. 2011. The ecological and evolutionary implications of merging different
  types of networks. Ecology letters 14:1170–1181.
- Fontaine, C. and E. Thébault. 2015. Comparing the conservatism of ecological interactions in plant–
  pollinator and plant–herbivore networks. Population Ecology 57:29–36.
- Futuyma, D. J. and A. A. Agrawal. 2009. Macroevolution and the biological diversity of plants and
  herbivores. Proceedings of the National Academy of Sciences 106:18054–18061.
- Gilman, R. T., S. L. Nuismer, and D.-C. Jhwueng. 2012. Coevolution in multidimensional trait
  space favours escape from parasites and pathogens. Nature 483:328.
- Gravel, D., F. Massol, E. Canard, D. Mouillot, and N. Mouquet. 2011. Trophic theory of island
  biogeography. Ecology letters 14:1010–1016.
- Grilli, J., M. Adorisio, S. Suweis, G. Barabás, J. R. Banavar, S. Allesina, and A. Maritan. 2017.
  Feasibility and coexistence of large ecological communities. Nature communications 8:0.
- Grilli, J., T. Rogers, and S. Allesina. 2016. Modularity and stability in ecological communities.
  Nature communications 7:1–10.
- Harmon, L. J., C. S. Andreazzi, F. Débarre, J. Drury, E. E. Goldberg, A. B. Martins, C. J. Melián,
  A. Narwani, S. L. Nuismer, M. W. Pennell, et al. 2019. Detecting the macroevolutionary signal
  of species interactions. Journal of evolutionary biology 32:769–782.

- Hembry, D. H., J. B. Yoder, and K. R. Goodman. 2014. Coevolution and the diversification of life.
  The American Naturalist 184:425–438.
- Ispolatov, I., V. Madhok, and M. Doebeli. 2016. Individual-based models for adaptive diversification
  in high-dimensional phenotype spaces. Journal of theoretical biology 390:97–105.
- Janz, N. 2011. Ehrlich and raven revisited: mechanisms underlying codiversification of plants and enemies. Annual review of ecology, evolution, and systematics 42.
- Jordano, P., J. Bascompte, and J. M. Olesen. 2003. Invariant properties in coevolutionary networks of plant-animal interactions. Ecology letters 6:69–81.
- Joy, J. B. 2013. Symbiosis catalyses niche expansion and diversification. Proceedings of the Royal
   Society of London B: Biological Sciences 280:20122820.
- Kay, K. M. and R. D. Sargent. 2009. The role of animal pollination in plant speciation: integrating
  ecology, geography, and genetics. Annual Review of Ecology, Evolution, and Systematics 40:637–
  656.
- Krasnov, B. R., M. A. Fortuna, D. Mouillot, I. S. Khokhlova, G. I. Shenbrot, and R. Poulin.
  2012. Phylogenetic signal in module composition and species connectivity in compartmentalized
  host-parasite networks. The American Naturalist 179:501–511.
- Krause, A. E., K. A. Frank, D. M. Mason, R. E. Ulanowicz, and W. W. Taylor. 2003. Compartments
  revealed in food-web structure. Nature 426:282.
- Krishna, A., P. R. Guimaraes Jr, P. Jordano, and J. Bascompte. 2008. A neutral-niche theory of
  nestedness in mutualistic networks. Oikos 117:1609–1618.
- Lewinsohn, T. M., P. Inácio Prado, P. Jordano, J. Bascompte, and J. M Olesen. 2006. Structure in
  plant–animal interaction assemblages. Oikos 113:174–184.
- Loeuille, N. and M. A. Leibold. 2014. Effects of local negative feedbacks on the evolution of species
  within metacommunities. Ecology letters 17:563–573.

- Loeuille, N. and M. Loreau. 2005. Evolutionary emergence of size-structured food webs. Proceedings
  of the National Academy of Sciences 102:5761–5766.
- <sup>579</sup> Lozupone, C. A., M. Hamady, S. T. Kelley, and R. Knight. 2007. Quantitative and qualitative  $\beta$ <sup>580</sup> diversity measures lead to different insights into factors that structure microbial communities. <sup>581</sup> Applied and environmental microbiology 73:1576–1585.
- Manceau, M., A. Lambert, and H. Morlon. 2015. Phylogenies support out-of-equilibrium models of
  biodiversity. Ecology letters 18:347–356.
- Manceau, M., A. Lambert, and H. Morlon. 2017. A unifying comparative phylogenetic framework
   including traits coevolving across interacting lineages. Systematic biology 66:551–568.
- 586 May, R. M. 1972. Will a large complex system be stable? Nature 238:413.
- Maynard, D. S., C. A. Serván, and S. Allesina. 2018. Network spandrels reflect ecological assembly.
  Ecology letters 21:324–334.
- Memmott, J., N. M. Waser, and M. V. Price. 2004. Tolerance of pollination networks to species
  extinctions. Proceedings of the Royal Society of London. Series B: Biological Sciences 271:2605–
  2611.
- <sup>592</sup> Minoarivelo, H. and C. Hui. 2016. Trait-mediated interaction leads to structural emergence in mu-<sup>593</sup> tualistic networks. Evolutionary ecology 30:105–121.
- Montesinos-Navarro, A., F. Hiraldo, J. L. Tella, and G. Blanco. 2017. Network structure embracing
   mutualism-antagonism continuums increases community robustness. Nature ecology & evolution
   1:1661–1669.
- Montoya, J. M., S. L. Pimm, and R. V. Solé. 2006. Ecological networks and their fragility. Nature
  442:259.
- Morlon, H., S. Kefi, and N. D. Martinez. 2014. Effects of trophic similarity on community composition. Ecology letters 17:1495–1506.

- Morlon, H., E. Lewitus, F. L. Condamine, M. Manceau, J. Clavel, and J. Drury. 2016. Rpanda: an r
  package for macroevolutionary analyses on phylogenetic trees. Methods in Ecology and Evolution
  7:589–597.
- Nuismer, S. L., P. Jordano, and J. Bascompte. 2013. Coevolution and the architecture of mutualistic
  networks. Evolution 67:338–354.
- Olesen, J. M., J. Bascompte, Y. L. Dupont, and P. Jordano. 2007. The modularity of pollination
   networks. Proceedings of the National Academy of Sciences 104:19891–19896.
- Pillai, P., A. Gonzalez, and M. Loreau. 2011. Metacommunity theory explains the emergence of
  food web complexity. Proceedings of the National Academy of Sciences 108:19293–19298.
- Pilosof, S., M. A. Fortuna, J.-F. Cosson, M. Galan, C. Kittipong, A. Ribas, E. Segal, B. R. Krasnov, S. Morand, and J. Bascompte. 2014. Host-parasite network structure is associated with
  community-level immunogenetic diversity. Nature communications 5:1–9.
- Poisot, T. and D. Stouffer. 2016. How ecological networks evolve. bioRxiv Page 071993.
- Rezende, E. L., P. Jordano, and J. Bascompte. 2007a. Effects of phenotypic complementarity and
  phylogeny on the nested structure of mutualistic networks. Oikos 116:1919–1929.
- Rezende, E. L., J. E. Lavabre, P. R. Guimarães, P. Jordano, and J. Bascompte. 2007b. Non-random
  coextinctions in phylogenetically structured mutualistic networks. Nature 448:925–928.
- Rohr, R. P., S. Saavedra, and J. Bascompte. 2014. On the structural stability of mutualistic systems.
  Science 345:1253497.
- Rosenzweig, M. L. et al. 1995. Species diversity in space and time. Cambridge University Press.
- Rosindell, J., L. J. Harmon, and R. S. Etienne. 2015. Unifying ecology and macroevolution with
  individual-based theory. Ecology letters 18:472–482.
- Saavedra, S., R. P. Rohr, J. M. Olesen, and J. Bascompte. 2016. Nested species interactions promote
  feasibility over stability during the assembly of a pollinator community. Ecology and evolution
  6:997–1007.

- Santamaría, L. and M. A. Rodríguez-Gironés. 2007. Linkage rules for plant-pollinator networks:
  trait complementarity or exploitation barriers? PLoS biology 5:e31.
- Smith, C. I., O. Pellmyr, D. M. Althoff, M. Balcazar-Lara, J. Leebens-Mack, and K. A. Segraves.
  2008. Pattern and timing of diversification in yucca (agavaceae): specialized pollination does not
  escalate rates of diversification. Proceedings of the Royal Society of London B: Biological Sciences
  275:249–258.
- Song, C., R. P. Rohr, and S. Saavedra. 2017. Why are some plant-pollinator networks more nested
  than others? Journal of Animal Ecology 86:1417–1424.
- Staniczenko, P. P., J. C. Kopp, and S. Allesina. 2013. The ghost of nestedness in ecological networks.
  Nature communications 4:1391.
- Stouffer, D. B. and J. Bascompte. 2011. Compartmentalization increases food-web persistence. Pro ceedings of the National Academy of Sciences 108:3648–3652.
- Suweis, S., F. Simini, J. R. Banavar, and A. Maritan. 2013. Emergence of structural and dynamical
   properties of ecological mutualistic networks. Nature 500:449.
- Thébault, E. and C. Fontaine. 2010. Stability of ecological communities and the architecture of
  mutualistic and trophic networks. Science 329:853–856.
- <sup>642</sup> Thompson, J. N. 2005. The geographic mosaic of coevolution. University of Chicago Press.
- Thompson, J. N. and B. M. Cunningham. 2002. Geographic structure and dynamics of coevolutionary selection. Nature 417:735.
- Valdovinos, F. S., B. J. Brosi, H. M. Briggs, P. Moisset de Espanés, R. Ramos-Jiliberto, and N. D.
- Martinez. 2016. Niche partitioning due to adaptive foraging reverses effects of nestedness and
  connectance on pollination network stability. Ecology letters 19:1277–1286.
- Valverde, S., J. Piñero, B. Corominas-Murtra, J. Montoya, L. Joppa, and R. Solé. 2018. The architecture of mutualistic networks as an evolutionary spandrel. Nature ecology & evolution 2:94–99.

- van der Niet, T. and S. D. Johnson. 2012. Phylogenetic evidence for pollinator-driven diversification
  of angiosperms. Trends in Ecology & Evolution 27:353–361.
- <sup>652</sup> Van Valen, L. 1973. A new evolutionary law. Evol Theory 1:1–30.
- Vázquez, D. P. 2005. Degree distribution in plant–animal mutualistic networks: forbidden links or
  random interactions? Oikos 108:421–426.
- Vázquez, D. P., N. P. Chacoff, and L. Cagnolo. 2009. Evaluating multiple determinants of the
  structure of plant-animal mutualistic networks. Ecology 90:2039–2046.
- Weber, M. G., C. E. Wagner, R. J. Best, L. J. Harmon, and B. Matthews. 2017. Evolution in a
  community context: on integrating ecological interactions and macroevolution. Trends in Ecology
  & Evolution 32:291–304.
- Yoder, J. B. and S. L. Nuismer. 2010. When does coevolution promote diversification? The American
   Naturalist 176:802–817.

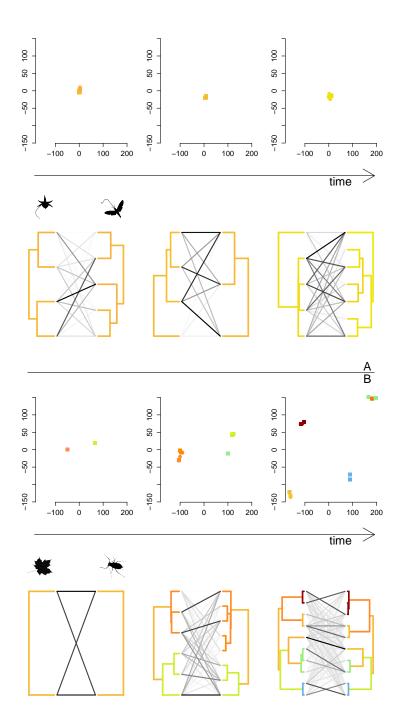


Figure 1: Example outputs of the eco-evolutionary model for mutualistic (panel A) and antagonistic (panel B) interactions. In each panel the upper row shows the evolution of trait values (in a three-dimensional trait space, point color representing the third trait dimension). The lower row shows the evolution of interaction networks and associated phylogenies. Darker links in the network correspond to interactions of higher strength; branches in the phylogenies are colored according to the value of the trait's third dimension so that it matches the colors of the corresponding dots in the upper row. Round dots correspond to species from guild A, square ones to species from guild B.

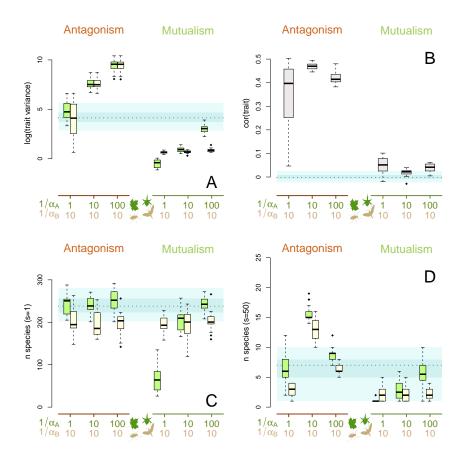


Figure 2: Effect of interaction type on the emergence of trait and species diversity. A : Logarithm of the variance of the first dimension of the species trait as a function of niche width for guild A (green boxplots) and B (white boxplots). The blue area shows values obtained for neutral simulations (range of values, quartiles and median). Antagonistic interactions commonly lead to higher, and mutualistic scenarios lower, trait diversity than in neutral simulations. Results for the other two trait dimensions are similar (results not shown). B : Correlation of the first dimension of the traits of the interacting individuals. Traits are consistently positively correlated in antagonistic scenarios, and correlation values stay pretty low in mutualistic ones. Results were similar for the other two trait dimensions (results not shown). C : Species richness in each guild for a species definition threshold s = 1. Diversity is higher in antagonistic scenarios than in mutualistic ones but always stays bellow that obtained for neutral scenarios. D : Species number in each guild for a species definition threshold s = 50. Diversity is higher in antagonistic scenarios than in mutualistic ones, and a few antagonistic scenarios display a diversity that is higher than in neutral simulations.

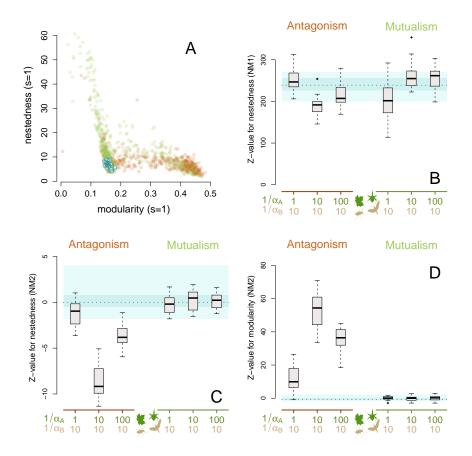


Figure 3: Effect of interaction type on nestedness and modularity A : Each point shows quantitative modularity and nestedness values for a single simulated network. The networks that have high nestedness values are mostly mutualistic ones (in green), while those with high modularity values are mostly antagonistic ones (in red). Blue points show the results for neutral simulations. B : Z-values for the binary NODF metric corrected by null model NM1 as a function of niche width. The blue area shows values obtained for neutral simulations (range of values, quartiles and median). All networks show up as significantly nested. C : Z-values for the quantitative NODF corrected by null model NM2. Mutualistic and neutral networks have similar levels of nestedness and are neither significantly nested nor anti-nested. A large proportion of antagonistic networks are significantly anti-nested. D : Z-values for the quantitative modularity metric corrected by null model NM2. Mutualistic and neutral networks have similar levels of modularity metric corrected by null model NM2. Mutualistic networks have similar levels of modularity metric corrected by null model NM2. Mutualistic and neutral networks have similar levels of modularity metric corrected by null model NM2. Mutualistic and neutral networks have similar levels of modularity and are not significantly modular. Most of the antagonistic networks are significantly modular. Results are shown for a species definition threshold s = 1.

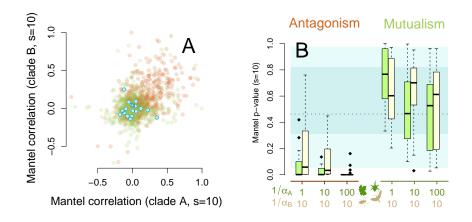


Figure 4: Effect of interaction type on phylogenetic signal in interaction partners. A : Each point shows, for a single simulation, the Mantel correlation between phylogenetic distance and dissimilarity in interaction partners on both side of the network. The correlation is higher for antagonistic (in red) than for mutualistic (in green) or neutral (in blue) networks. B : Mantel p-value as a function of niche width. The blue area shows values obtained for neutral simulations (range of values, quartiles and median). White boxplots show results for guild A and green boxplots for guild B. Results are shown for the Unifrac dissimilarity metric and a species definition threshold s = 1.