



HAL
open science

Membrane Fluidity as a New Means to Selectively Target Cancer Cells with Fusogenic Lipid Carriers

Julien Bompard, Annalisa Rosso, Leyre Brizuela, Saïda Mebarek, Loïc Blum, Ana-Maria Trunfio-Sfarghiu, Giovanna Lollo, Thierry Granjon, Agnès Girard-Egrot, Ofelia Maniti

► **To cite this version:**

Julien Bompard, Annalisa Rosso, Leyre Brizuela, Saïda Mebarek, Loïc Blum, et al.. Membrane Fluidity as a New Means to Selectively Target Cancer Cells with Fusogenic Lipid Carriers. *Langmuir*, 2020, 36 (19), pp.5134-5144. 10.1021/acs.langmuir.0c00262 . hal-03083631

HAL Id: hal-03083631

<https://hal.science/hal-03083631>

Submitted on 23 Dec 2020

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

SUPPORTING INFORMATION

Membrane Fluidity as a New Mean to Selectively Target Cancer Cells with Fusogenic Lipid Carriers

Julien Bompard¹, Annalisa Rosso², Leyre Brizuela¹, Saïda Mebarek¹, Loïc J. Blum¹, Ana-Maria Trunfio-Sfarghiu³, Giovanna Lollo², Thierry Granjon¹, Agnès Girard-Egrot¹, Ofelia Maniti^{1}*

¹: Institut de Chimie et Biochimie Moléculaires et Supramoléculaires, ICBMS UMR 5246, Univ Lyon, Université Lyon 1, CNRS, Lyon, France

²: Laboratoire d'Automatique, de Génie des Procédés et de Génie Pharmaceutique, LAGEPP UMR 5007, Univ Lyon, Université Lyon 1, CNRS, Lyon, France

³: Laboratoire de Mécanique des Contacts et Structures, LaMCoS UMR 5259, Univ Lyon, Université Lyon 1, CNRS, Lyon, France

**corresponding author : ofelia.maniti@univ-lyon1.fr*

Number of pages: 7

Number of figures: 6

Number of tables: 0

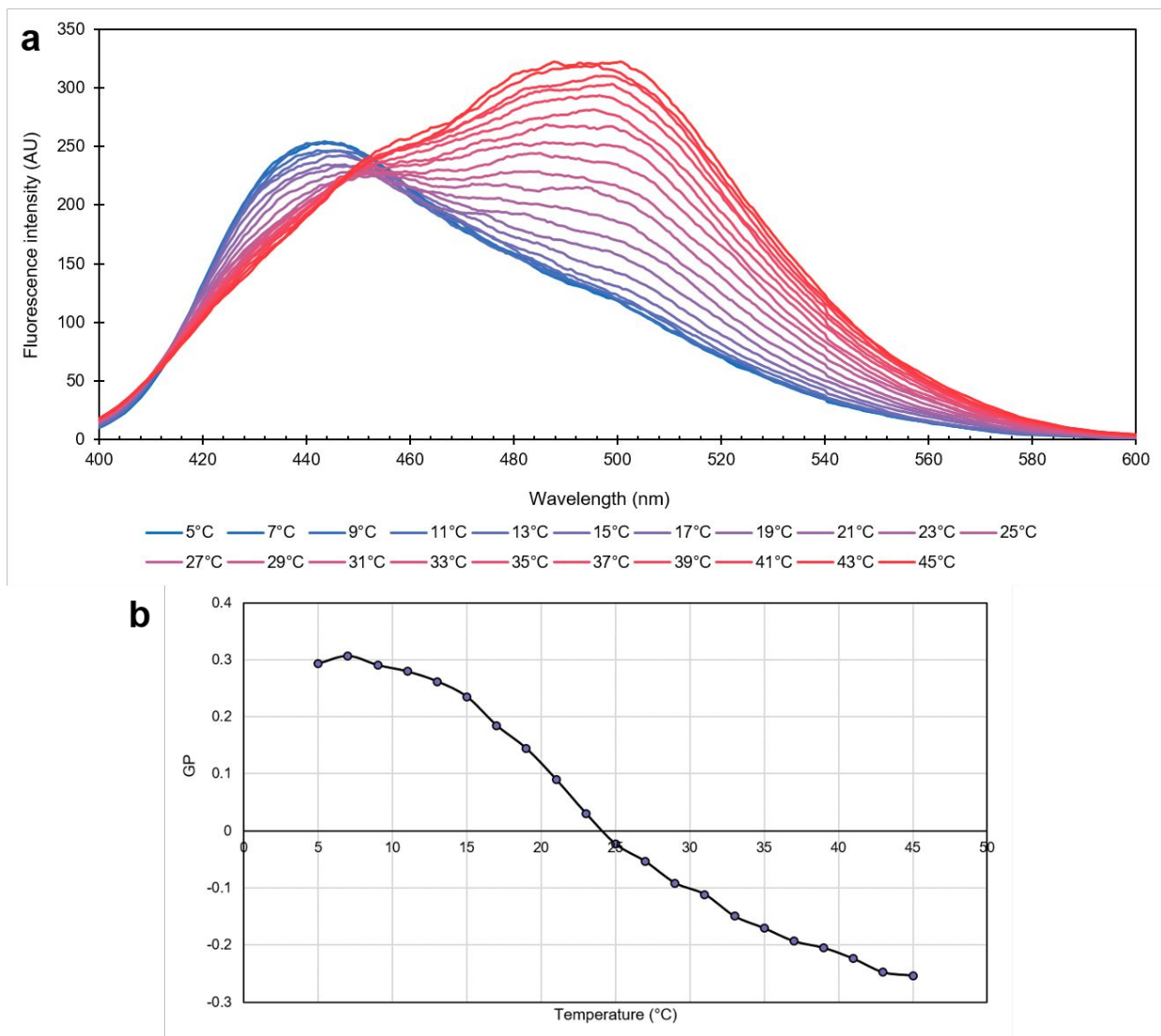


Figure S1. DiIol accurately reports the membrane fluidity state of liposome membranes.

a) Fluorescence spectra of DMPC liposomes incubated with DiIol for 15 min, from 5°C to 45°C.

b) GP measurements, showing an inflexion at 24°C, the usual phase-transition temperature of DMPC.

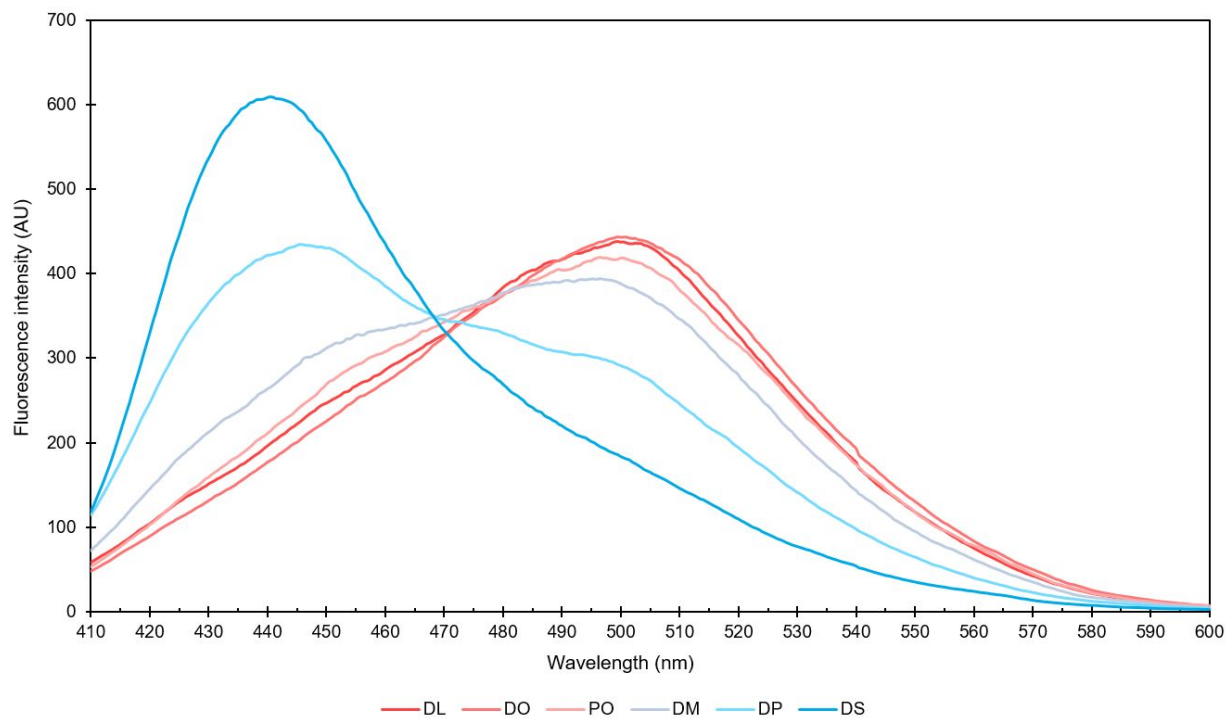


Figure S2. DiIol fluorescence spectra in liposomes with varying phase-transition temperatures reports differences in membrane fluidity states of liposomes made of 80% PC and 20% DOPE.

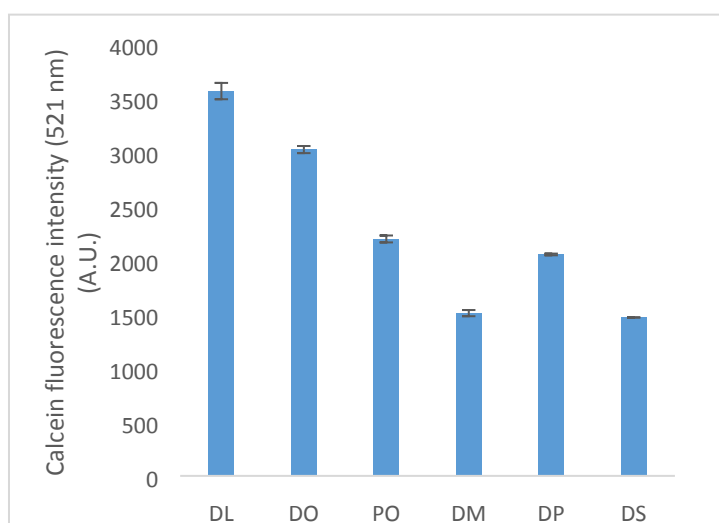


Figure S3. Calcein load in liposomes. Calcein fluorescence intensity at 521 nm (488 nm excitation) as a function of liposome preparation for a liposome concentration of 100 $\mu\text{g/ml}$. Calcein was loaded in the liposomes at a concentration of 5 μM . Free calcein was removed with size exclusion chromatography.

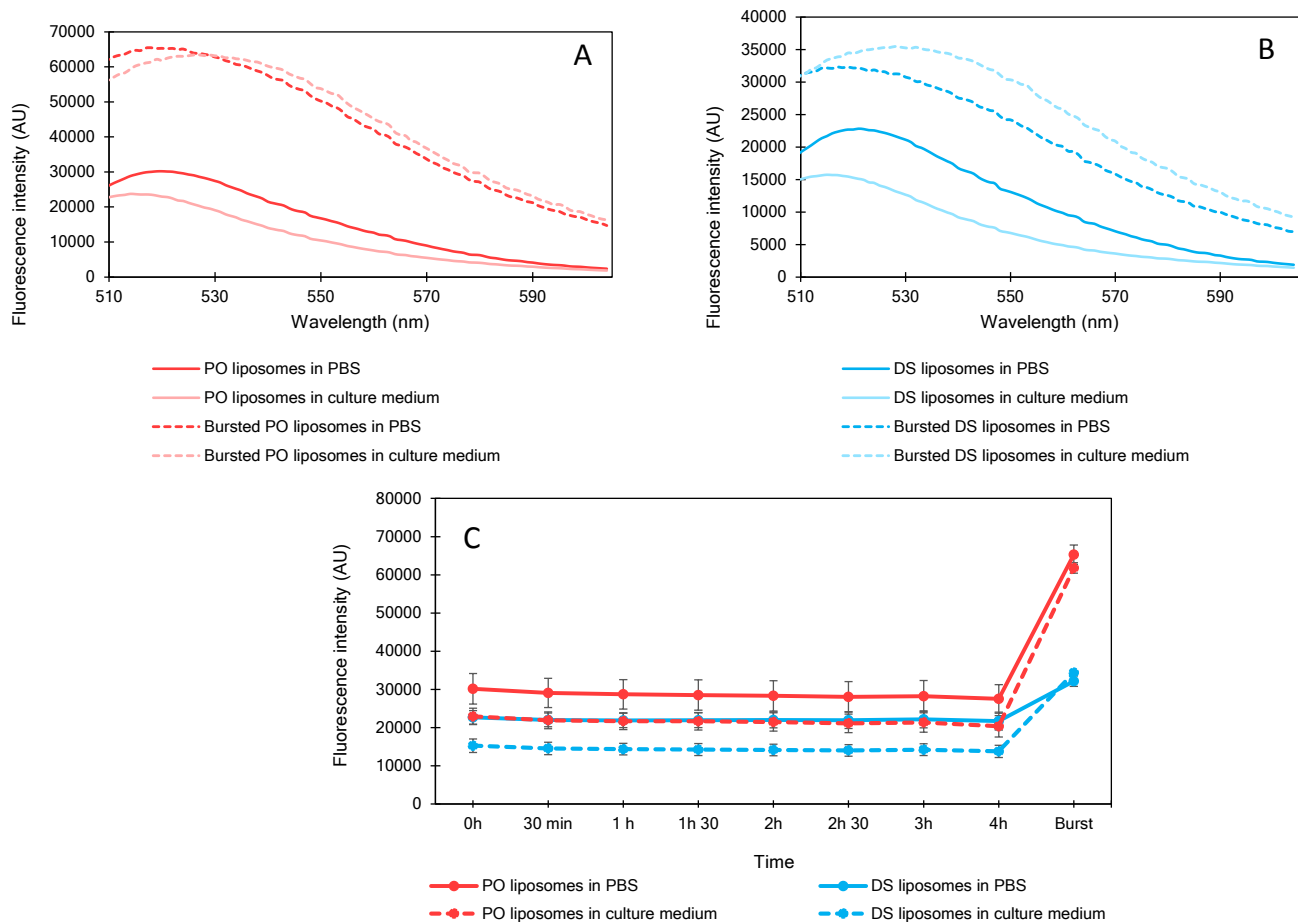


Figure S4: Liposome stability in cell culture medium. PO (A) and DS (B) liposomes were loaded with 60 mM calcein and diluted either in PBS or in FBS-containing DMEM medium. Fluorescence spectra were recorded after excitation at 488 nm before (full lines) and after (dotted lines) liposome burst with 10 μ l HCl 1M. The fluorescence intensity increased after liposome burst indicated release from quenching. C. Evolution of calcein fluorescence (measured at maximum emission wavelength, 521 nm) with time in PBS (full lines) or in FBS 10 % cell culture medium. Fluorescence was low and constant for several hours, both in PBS and in cell culture medium, but drastically increased after liposome burst indicating that FBS-containing media did not affect liposome integrity.

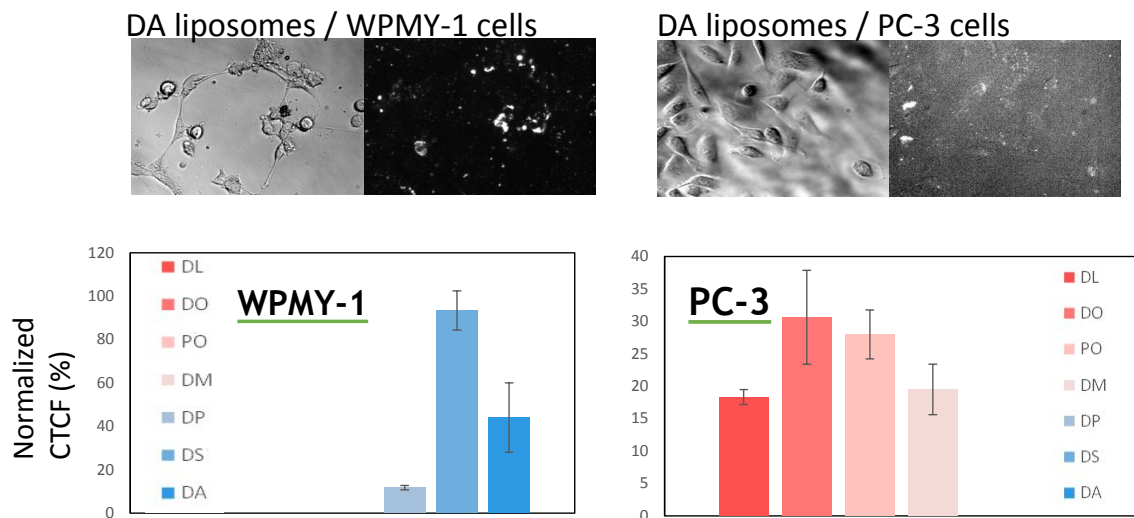


Figure S5: DA liposomes (composed of 78 % of DAPC (C20:0), 20 % DOPE and 2 % NBD-PE) interact with WPMY-1 but not PC-3 cells. Top: fluorescence microscopy images of WPMY-1 (left) or PC-3 cells incubated with DA liposomes. Bottom: Fluorescence intensity (CTCF) quantified on fluorescence images and normalized over the maximum fluorescence intensity value. Left: WPMY-1, right: PC-3 cells incubated with DL, DO, PO, DM, DP, DS and DA liposomes.

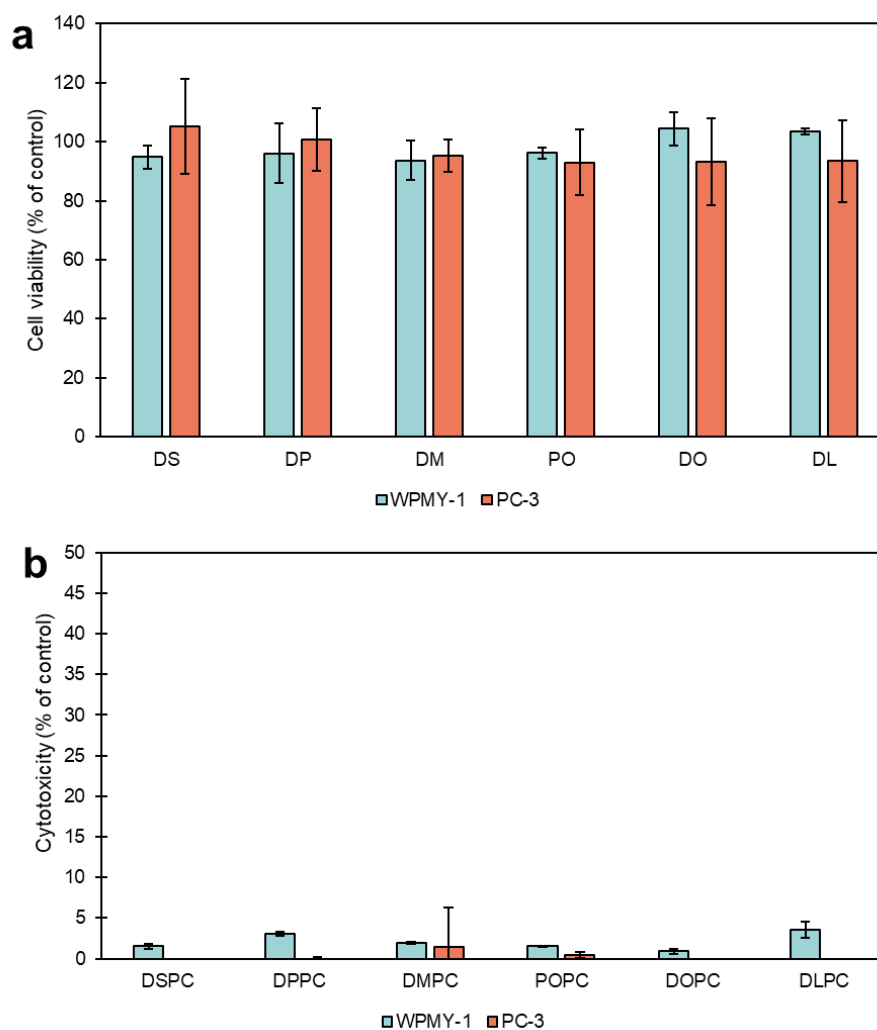


Figure S6. Liposomes do not affect cell viability and show no cytotoxic effect. a) Cell viability quantification, assessed on plated cells with the MTT assay 2h30 after liposome treatment. Results expressed as a percentage of cell viability relative to an untreated control corresponding to 100% viability. b) Cytotoxicity quantification, assessed on the same cells than (a) with the LDH assay. Results expressed as a percentage of cell death relative to an untreated control corresponding to 0% toxicity and a control treated with 10% Triton X-100 corresponding to 100% toxicity.