

Infestation of small seabirds by Ornithodoros maritimus ticks: Effects on chick body condition, reproduction and associated infectious agents

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1	Infestation of small seabirds by Ornithodoros maritimus ticks: effects on chick body
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25 Abstract:

Ticks can negatively affect their host by direct effects as blood feeding causing 26 anaemia or discomfort, or by pathogen transmission. Consequently, ticks can have an 27 important role in the population dynamics of their hosts. However, specific studies on 28 the demographic effects of tick infestation on seabirds are still scarce. Seabird ticks 29 30 have also the potential to be responsible for the circulation of little known tick-borne 31 agents, which could have implications for non-seabird species. Here, we report the 32 results of investigations on potential associations between soft tick Ornithodoros maritimus load and reproductive parameters of storm petrels Hydrobates pelagicus 33 breeding in a large colony in a cave of Espartar Island, in the Balearic archipelago. We 34 35 also investigated by molecular analyses the potential viral and bacterial pathogens 36 associated with O. maritimus ticks present at the colony. Lower nestling survival was recorded in the most infested area, deep in the cave, compared to the area near the 37 entrance. The parasite load was negatively associated with the body condition of the 38 39 nestlings. One pool of ticks tested positive for West Nile virus and 4 pools tested 40 positive for a Borrelia species which was determined by targeted nested PCR to have a 41 99% sequence identity with B. turicatae, a relapsing fever Borrelia. Overall, these 42 results show that further investigations are needed to better understand the ecology 43 and epidemiology of the interactions between ticks, pathogens and Procellariiform 44 species.

45

46 **Keywords:** *Borrelia*, mortality, storm petrels, soft ticks, tick-borne pathogens, WNV.

47

48 Introduction

Animals coexist with numerous parasite species with important ecological and evolutionary consequences (Hamilton and Zuk, 1982; Møller et al., 1993; Moore, 2002; Poulin, 2011). Hematophagous ectoparasites such as ticks, in particular, can damage their hosts directly by the fact of biting (i.e. dermatoid process) and blood-feeding (i.e. blood loss and associated anaemia) and/or indirectly through pathogen transmission (Jongejan and Uilenberg, 2004; Brites-Neto et al., 2015).

55 Ticks are divided into two groups: hard ticks (Ixodidae) and soft ticks (Argasidae). Both families can potentially transmit numerous pathogens of medical and 56 veterinary interest (Dietrich et al., 2011 and references therein). However, those 57 58 transmitted by soft ticks have been less studied due to the specialization of Argasidae 59 to hidden habitats (i.e. crevices) and the short time they spend for blood feeding on the host compared to hard ticks (Vial, 2009). Some of the most common worldwide 60 diseases caused by pathogens transmitted by soft ticks include: human tick-borne 61 62 relapsing fever (TBRF); viral encephalitis transmitted by ticks infesting seabirds, shore 63 birds and roosting birds; the African swine fever virus (ASFV); fowl spirochetosis 64 infections or anaplasmosis-like diseases and epizootic bovine abortion (Vial, 2009). Such pathogens can induce both lethal and sub-lethal effects on their hosts (Niebylski 65 66 et al., 1999; Gray and Bradley, 2006). Organisms present a wide range of responses to 67 tick-induced pathogens (Ramamoorthi et al., 2005). For instance, transmitted pathogens infecting the host might induce behavioral and/or physiological 68 69 modifications which interfere with immunity response, gene expression or energy 70 allocation, thus potentially affecting fitness. However, the effects of tick infestation 71 and tick-borne diseases on host populations are often poorly understood and

quantifications of sub-lethal effects are hard to assess (Nemeth et al., 2006; Dietrich etal., 2011).

Seabirds can be predictable hosts for ticks because they show colonial breeding 74 habits resulting in large and dense aggregations, high nest site fidelity and extended 75 76 breeding periods (Ramos et al., 2001; Schreiber and Burger, 2001). These breeding 77 characteristics of seabirds facilitate transmission and contribute to the support of large 78 and dense populations of ticks (Dietrich et al., 2011 and references therein). Severe 79 infestations of seabird colonies by ticks can have heavy negative effects (Duffy, 1983; Ramos et al., 2001; Dietrich et al., 2011). For example, Duffy (1983) documented a 80 large-scale nest desertion in Peruvian guano seabirds (cormorants, boobies and 81 82 pelicans) due to Ornithodoros amblus soft tick infestation at breeding sites. Similarly, 83 high densities of O. capensis s.s. seemed to be responsible for Sooty tern (Sterna *fuscata*) egg and newly hatched chick desertion at Seychelles in 1972 (Feare, 1976). 84 85 Colony abandonment or dispersal could be used as a strategy to avoid the negative 86 effects of parasites (Boulinier and Danchin, 1996). In fact, some cases of lethal effects 87 of ticks (i.e. *Ixodes uriae*) on seabirds have been documented, especially for young 88 nestlings (Boulinier and Danchin, 1996; Ramos et al., 2001) but very rarely in adults (Gauthier-Clerc et al., 1998). On the other hand, non-lethal but also negative effects 89 90 have been documented on seabird body condition and/or nestlings' growth rates (e.g. 91 Bosch and Figuerola, 1999; McCoy et al., 2002; Hipfner et al., 2019). However, 92 sometimes the effects of ticks are not evident until infestation levels are very high 93 (Gauthier-Clerc et al., 2003; Hipfner et al., 2019) or may depend on the environmental 94 conditions experienced by the hosts (McCoy et al., 2002). Although ticks can have an 95 important role in seabird population dynamics, specific studies on the demographic

96 effects of tick infestation on seabirds are still scarce (Dietrich et al., 2011; Rodríguez et
97 al., 2019). Moreover, the ultimate mechanisms responsible for lethal and sub-lethal
98 effects of ticks (i.e. direct effect or indirect effect due to pathogen transmission) on
99 seabirds remain largely unknow (Yabsley et al., 2012).

100 Espartar Island, located on the Balearic archipelago, hosts the largest Spanish 101 colony of Mediterranean storm petrel (Hydrobates pelagicus melitensis), estimated at 102 750-1250 breeding pairs (Picorelli, unpublished data). Other seabirds (gulls and 103 shearwaters) also breed in the island, but storm petrels mainly concentrate in cliff 104 caves, sharing their breeding habitat with Balearic shearwaters (Puffinus 105 mauretanicus). Balearic shearwaters breed in lower densities and start reproduction 106 much earlier (i.e. February; Guilford et al., 2012) than storm petrels (i.e. May-June; 107 Ramírez et al., 2016). A systematic monitoring of the breeding parameters of storm 108 petrels breeding at the largest colony of the island, the cap de Migdia cave (~250 109 breeding pairs) started in 2014 (Mínguez et al., 2015). During the first years of 110 monitoring (2014-2016), we detected a high mortality of nestlings (see results) 111 compared to other storm petrel Mediterranean colonies in which nestling survival 112 typically varies between 90-95% (Sanz-Aguilar et al., 2009). Moreover, we experienced 113 recurrent tick bites during the daylight monitoring of the cave, a very unusual fact 114 when compared with our experience monitoring other storm petrel colonies. Ticks were identified as the soft tick O. maritimus (Vermeil and Marguet, 1967), included in 115 116 the complex O. (Carios) capensis that includes eight described species that parasitize 117 tropical and temperate colonial seabird species (Hoogstraal et al., 1979; Khouri, 2011; 118 Dupraz et al., 2016). Identification of *O. maritimus* was performed by morphology (i.e. 119 median dorsal line of mamillae disc short) and confirmed by an expert on the group

(Dupraz, M., personal communication). In addition, based on Dupraz et al., (2016) *O. maritimus* is mostly restricted to the West Paleartic Region where its presence
parasitizing seabirds have been extensively reported (Dietrich et al., 2011). The species *O. capensis* s.s. is also present in the same region, but almost exclusively associated
with pigeons and it can be morphologically separated from *O. maritimus* (Khoury et al.,
2011).

Both bites to researchers and mortality of storm petrel chicks were especially 126 127 evident in a particular area of the cave, the inner chambers. Consequently, in 2018 we evaluated the potential differences of O. maritimus load between different parts of the 128 colony as a proxy of tick abundance and evaluated their effects on nestlings' body 129 130 condition. We used this information to infer the potential effect of ticks on the 131 reproductive parameters of storm petrels. Moreover, we also used a high throughput real-time PCR system to test for the presence in O. maritimus ticks of the most 132 133 common tick-borne infectious agents including bacteria, parasites and viruses 134 potentially harboured by ticks.

135 Methods

136 Species, study site, field data and tick collection

137 The Mediterranean storm petrel is one of the smallest seabirds of the Order 138 Procellariiformes (average body mass, 28 g, Warham 1990). Storm petrels breed in 139 caves, burrows, under boulders or in crevices where they lay a single egg directly on 140 the floor (Fig. 1A). The egg is incubated by both partners for 40 days and chick rearing 141 lasts for about 63–70 days (Scott, 1970; Mínguez, 1994). The earliest clutches are laid 142 in the second half of April and the last eggs are laid in the first week of July (Ramírez et 143 al., 2016). Most fledglings leave colonies in August (Mínguez, 1994).

144 The study was conducted on Espartar Island, a 20.5 ha island located within the Es Vedrà, Es Vedranell i els illots de Ponent Natural Reserve (38° 57' 31" N, 1° 11' 44" E, 145 146 Fig. 2). Espartar Island has a characteristic steep relief with gentle slopes facing south and cliffs in the north. Storm petrels breed all around the island at low densities but 147 concentrate at high densities in natural caves. Our study site was located in one of 148 149 those caves, Cap des Migdia cave, that annually hosts around 250-300 storm petrel breeding pairs. The cave has two areas with different characteristics (Fig. 2): the 150 151 entrance and the inner chambers. The entrance of the cave is a wide and illuminated 152 area where nests are located in small cavities and rock debris. The inner chambers are narrow, dark and humid; in this area, nests are located in small cavities, rock debris 153 154 and dusty soil (Fig. 1A). The inner chamber of the cave also hosts 12 pairs of breeding 155 Balearic shearwaters.

156 From 2014 to 2018, all accessible nests found in the study site (N = 312) were marked (Fig. 2A) and inspected every 2 weeks along the breeding season (May-157 158 September) to record reproductive parameters: hatching success (No.eggs hatched/No.eggs laid), fledgling success (No.chicks fledged/No.eggs hatched) and breeding success (No.chicks 159 160 fledged/No.eggs laid) (Mínguez et al., 2015). In addition, during the 2018 breeding season, 161 we inspected for parasites some adults and all the nestlings located alive and alone in 162 accessible nests at each monitoring visit. Given the extended breeding period of storm petrels and the mortality of some of the young nestlings inspected (N = 6), some 163 164 nestlings were inspected once and others multiple times. In total we performed 62 165 inspections of 23 nestlings reared at the entrance of the cave and 92 inspections of 37 166 nestlings reared at the inner chambers. 28 breeding adults nesting at the entrance and 167 24 nesting at the inner chambers were inspected once during the breeding season. We

168 did not capture the nestlings during the brooding period (i.e. when adults are covering 169 the nestling) to avoid potential negative manipulation effects during this critical stage 170 (Minguez and Oro, 2003). We visually inspected nestlings and adults pushing aside the down and feathers and counted the number of tick larvae attached to their skin (Fig. 171 1C). However, in order to avoid disturbance of breeding birds, inspections of adults 172 173 were faster and less exhaustive than inspections to nestlings. We measured their wing 174 length, tarsus and body mass using a ruler, a digital calliper and a digital balance. 175 Handling time was reduced to the minimum possible.

176 Assessing tick density in the cave is extremely challenging. O. maritimus ticks (Fig. 1B) are mainly nocturnal, feeding on the host rapidly at night in the nymphal and 177 178 adult life stages (Vial, 2009). During the day, they are buried in the ground and located 179 in small crevices and under stones (Vial, 2009). On the contrary, tick larvae can be 180 attached for a much longer time on their hosts, their blood meal lasting from hours to several days (Vial, 2009). Therefore, in order to evaluate the potential differences in 181 182 tick density on the different areas of the cave (i.e. entrance vs. inner chambers) we 183 used the data on the larvae tick load (i.e. the number of tick larvae attached to the skin) of storm petrel nestlings (Fig. 2C) as a proxy of the amount of ticks on the two 184 185 areas of the cave. At the end of the 2017 breeding season, ticks were searched on the 186 ground of the cave, under stones and in crevices, and collected for molecular analyses. 187 Ticks found together (i.e. under the same stone) were pooled. Until analysed, they 188 were stored at -70°C. Note that adult ticks were only found in the inner parts of the 189 cave (i.e. the chambers) although the entrance was also inspected.

190 Statistical analyses

191 First, we evaluated the effect of the area of the cave on the parasite load (i.e. 192 no. of attached tick larvae) experienced by the nestlings at each monitoring visit 193 conducted in 2018. Based on field observations, we also accounted for nestling age as a predictor of parasite load. Nestling age was determined at first capture using the 194 relationship between age and wing length (Age = (0.424*wing length) + 6.6102; R² = 195 196 0.95; Sanz-Aguilar, unpublished data) estimated at Benidorm Island (Spain), where an intense monitoring in 2015 allowed to establish the exact hatching date for 30 197 198 nestlings. This analysis was carried out using a generalized linear mixed model GLMM 199 with a Poisson distribution (log link function) and considering nestling identity as a 200 random effect.

201 Nestling and adult body condition was estimated individually using the 202 residuals from a least squares (OLS) linear regression of body mass against the 203 measure of wing (Owen and Cook, 1977). The residual indices were used as a proxy of 204 body condition: the individuals with positive residual values were considered in better 205 body condition than individuals with negative values (Jakob et al., 1996). We assessed 206 the association between the nestling body condition and the nesting area (entrance vs. 207 inner chambers) and the individual tick load (on a log scale). For tick load, we 208 considered either the actual tick load (No. of tick larvae) presented by each individual 209 at each monitoring or the maximum tick load, the maximum number of tick larvae 210 recorded for a given individual during any of the monitoring visits carried out during 211 the study period. As nestlings remain in the same nest during their development, this 212 last fixed individual covariate may be related with the tick load in the nest and 213 potentially experienced by the individual. This analysis was carried out using a GLMM 214 with a normal distribution (identity link function) and considering nestling identity as a

random effect. Adults were only measured once during the study period and no tick
larvae was found attached to them. Consequently, a GLM with a normal distribution
(identity link function) was used to analyse the potential differences in their body

218 condition between the entrance and inner chambers breeding areas.

219 Finally, we evaluated the potential effect of the year and area of the cave on

220 reproductive parameters: hatching success, fledgling success and breeding success.

221 This analysis was carried out using a GLMM with a binomial distribution (logit link

function) and considering nest identity as a random effect.

223 Model selection was performed using the Akaike's information criterion (AIC; 224 Burnham and Anderson, 2002). We considered models to be equivalent when the 225 difference in AIC with the best model (ΔAIC) was < 2 (Burnham and Anderson, 2002).

226 RNA and genomic DNA extraction from the ticks

After collection and storage, ticks were identified and used for DNA/RNA 227 228 extractions. The extraction protocols used on the ticks followed those outlined in 229 Michelet et al. (2014) and Gondard et al. (2018). All ticks were washed for 5 min in an 230 ethanol bath, 10 min in two successive water baths and placed individually in sterile 231 tube and crushed in 300 μ l of Dulbecco's modified eagle medium (DMEM) with 10% 232 fetal calf serum using Precellys[®]24 Dual homogenizer (Bertin, France). The supernatant 233 was divided in 3 fractions: 100 μ l for the DNA extraction, 100 μ l for the RNA extraction and the rest was used as back-up and conserved at -80°C. Genomic tick DNA was then 234 235 extracted using the Wizard genomic DNA purification kit (Promega, France) according 236 to the manufacturer's instruction. Purified DNA and RNA were eluted into 50 μ l of 237 rehydration solution and 50 µl of RNase-free water, respectively, and conserved at -

238 80°C. Tick DNA and RNA quality was assessed via the amplification of the ITS2 region

and COI gene respectively (Michelet et al., 2014; Gondard et al., 2018).

240 High throughput real-time PCR system

Ticks were analysed for the most common tick-borne infectious agents using the BioMark real-time PCR amplification system. The DNA primer chip developed by Michelet et al. (2014) includes primers for detecting 28 bacterial species, 12 parasite species and the RNA primer chip developed by Gondard et al. (2018) includes 21 viruses. Twenty-two ticks pooled in eleven pools of a maximum of five adult ticks and were analysed by both systems in duplicate.

All RNAs were reverse transcribed into cDNAs using random primers and oligos 247 248 (dT). The remaining methods followed those of Michelet et al. (2014) and Gondard et 249 al. (2018). DNA and cDNA pre-amplifications were performed using the TaqMan 250 PreAmp Master Mix (Applied Biosystems, France) according to the manufacturer's 251 instructions. Primers for bacteria or viruses were pooled by combining equal volumes 252 of each primer to have 200 nM of each. The pre-amplification was performed in a final volume of 5 µL containing 2.5 µL TaqMan PreAmp Master Mix (2x), 1.2 µL pooled 253 254 primer mix (0.2) and 1.3 mL DNA. Thermal cycling conditions were as follows: one cycle 255 at 95°C for 10 min, 14 cycles at 95°C for 15 s and 4 min at 60°C (Michelet et al., 2014; 256 Gondard et al., 2018). The quantitative PCR reactions were then performed using 6-257 258 carboxyfluorescein (FAM) and black hole quencher (BHQ1)-labeled TaqMan probes

259 (Michelet et al., 2014; Gondard et al., 2018) with TaqMan Gene expression Master

260 Mix, in accordance to the manufacturer's instructions (Applied Biosystem, France). PCR

cycling comprised 5 min at 95°C, 45 cycles at 95°C for 10 s, 15 s at 60°C and 10 s at

40°C. Data were acquired on the BioMark real-time PCR system and analyzed using the
Fluigdim real-time PCR Analysis software. The assays were performed in duplicate
using two negative water controls per chip and *Escherichia coli* strain EDL933 was
added in each run to control for internal inhibition (Michelet et al., 2014).

A nested PCR using primers for the detected bacteria (targeting *Borrelia* gene *fla*; Loh et al., 2016) and a real-time PCR using primers for the detected virus (targeting the NS5 region from all flavivirus; (Weissenböck et al., 2002) were used to confirm the presence of the detected infectious agent in the samples. Amplicons were sequenced by Eurofins MWG Operon (Germany) and assembled using the BioEdit software (Ibis Biosciences, Carlsbad). An online BLAST (National Center for Biotechnology

272 Information) was used to identify the sequenced organism.

273 Results

274 Tick infestation

275 We did not find tick larvae attached to adult birds. In the case of nestlings, we 276 observed that tick larvae were mainly attached to the skin under the wings, the head and the neck, which are the areas with lower density of down and feathers (own 277 278 observation). 80% of the nestling inspected at the inner chambers (N = 37) and 43% of 279 the nestling inspected at the entrance (N = 23) presented at least one tick larvae 280 during the study period. The percentage of nestlings infested by at least one tick larvae showed temporal variations and decreased at the end of the study period, especially at 281 282 the entrance (Table 1).

Tick larvae load (i.e. No. of tick larvae on storm petrel nestlings) was higher in the inner chambers (mean 4.18 ± 0.69 SE) of the cave than at the entrance (mean 1.13 ± 0.85 SE), and decreased as nestlings became older (Table 2, Fig. 3). The age-related

decrease in tick load was more rapid at the entrance of the cave (Fig. 3). Six of the

nestlings inspected died (one at the entrance and 5 at the inner chambers). Nestlings

288 found dead presented high numbers of tick larvae on them at the last alive inspection

(mean = 20.8; range 2-40). We documented the mortality of 10% of the nestlings

detected as infested by ticks at the entrance (No._{dead}/No._{infested}= 1/10) and the

291 mortality of 17% of the nestlings infested by ticks in the inner chambers

292 (No._{dead}/No._{infested}= 5/30).

293 Body condition

Adults showed no differences in body condition at the entrance and the inner chambers (Table 3). For nestlings, differences in body condition were mainly related

with individual tick load (either the actual or the maximum tick load) and not with the

effect the zone *per se* (Table 2). The best model indicated that nestlings with the

298 highest tick larvae recorded during any of the monitoring visits (i.e. the maximum tick

load) experienced lower body condition (Table 2, Fig. 4).

300 *Reproductive parameters of storm petrels*

301 Individuals breeding in the entrance of the cave showed higher mean 302 performances in breeding parameters, especially regarding the fledging success (Table 303 4; $HS_{entrance} = 0.771 \pm 0.025 SE$; $FS_{entrance} = 0.903 \pm 0.021 SE$; $BS_{entrance} = 0.682 \pm 0.028 SE$) 304 than individuals breeding in the inner chambers (HS_{chambers} = 0.646 ± 0.018 SE; FS_{chambers} 305 = 0.516 ± 0.025 SE; BS_{chambers} = 0.320 ± 0.018 SE). Hatching success and fledgling 306 success varied over time and between zones, the additive models being preferred over 307 the models with interactions (Table 4). The differences between zones were higher for 308 fledgling success (i.e. nestling survival) than for hatching success (Fig. 5). Breeding

309 success showed the lowest temporal variation, the constant model being preferred

over the temporal models (Table 4, Fig. 5).

311 Molecular analyses

312 The West Nile virus (WNV) was detected in duplicate on one tick pool over the 11 that were analysed with the design targeting the genotype 3 of this virus (known to 313 314 be transmitted by ticks). Although the Pan-Flavivirus real-time PCR did not succeed to 315 confirm this result, nor the attempt of viral isolation into mammalian cell culture, this 316 may be expected if the concentration of the virus is low. This was the only virus 317 detected among the 21 that were searched. Four pools of ticks tested positive for a *Borrelia* sp. which was determined by targeted nested PCR to have a 99% sequence 318 319 identity with *B. turicatae*, a relapsing fever *Borrelia* (GenBank accession number: 320 MK732470).

321 Discussion

322 In this study, we show spatially explicit differences of infestation by the soft tick 323 *O. maritimus* of nestling storm petrels in a large cave on Espartar Island. Tick infestation was higher in the cave chambers and less so in the entrance. The within-324 325 colony differences in parasite density may be related to different and not exclusive 326 factors as the presence of Balearic shearwaters breeding in the inner parts of the cave 327 and the *per se* characteristics of the cave (Dietrich et al., 2011 and references therein). First, tick population dynamic parameters (notably survival) are known to depend on 328 329 local climatic conditions such as temperature and humidity (Sonenshine and Roe, 330 2013). The studied colony is located inside a cave where environmental conditions 331 (especially deep in the cave) are expected to be quite stable. Moreover, humidity is 332 higher at the inner chambers than at the entrance (own observation). These factors

333 can facilitate oviposition and tick survival, especially in the inner chambers (Vial, 2009. 334 Second, although parasites and infectious agents responsible for diseases are often 335 strongly host-specific, Ornithodoros spp. show indiscriminate host feeding and short 336 time for feeding completion compared to hard ticks (Vial, 2009). Balearic shearwaters 337 start to visit the colonies in November and the last chicks fledge in June (Guilford et al., 338 2012). Storm petrels start to visit the colonies in March and the last chicks fledge in late September (Ramírez et al., 2016). Moreover, storm petrels show asynchronous 339 340 breeding: the first eggs are laid at the end of April and the last at the beginning of July (Mínguez, 1994). Reproductive asynchrony of storm petrels may promote the 341 reproduction of ticks (Møller et al., 1993). Consequently, hosts can be available in the 342 343 cave during 11 months per year and seabird nestlings during 4-5 months, favouring the 344 production of several tick generations per year and high local population densities 345 (Vial, 2009).

346 Nestling body condition was negatively correlated with tick load, which may 347 explain the higher nestling mortality in the inner parts of the colony. Our results agree with other studies on seabirds that have detected high nest desertion and/or nestling 348 349 mortalities of individuals infested by ticks or in areas with high tick densities (e.g. 350 Feare, 1976; Duffy, 1983; Ramos et al., 2001; Reeves et al., 2006; Descamps 2013). In 351 addition, nestling mortality in the inner parts of the cave, where tick prevalence was 352 higher and ticks were more abundant, was very high (48%). We cannot directly analyse 353 the effect of tick infestation on nestling survival due to our reduced sample size of 354 nestlings inspected before dying (i.e. for which we had counted the number of tick 355 larvae attached to them when alive). This is because the majority of the mortalities 356 occurred during the first two weeks after hatching and many nestlings were born and

357 died between our monitoring visits (own observation). However, the low survival of 358 storm petrel nestlings located in the inner parts of the Espartar's cave matches with 359 the high mortality estimates provided for other seabird species nestlings infested by 360 ticks (Feare, 1996; Ramos et al., 2001). For example, Ramos et al. (2001) documented a 361 mortality of 63% of roseate terns (Sterna dougallii) infested by hard ticks Amblyomma 362 loculosum versus a nestling mortality of 17% of non-infested nestlings during a harsh breeding season with food shortage. On the contrary, during a breeding season with 363 364 benign environmental conditions, only 24% of the tick-infested nestlings died (Ramos 365 et al., 2001).

In fact, environmental conditions may influence the effect of parasites on hosts 366 367 (Ramos et al., 2001; McCoy et al., 2002). At Espartar's colony, during our 5 years of 368 monitoring the fledgling success showed annual variations that may be related with 369 differential tick densities or environmental conditions affecting storm petrels and or 370 interacting with their parasites (McCoy et al., 2002), factors that should be studied in 371 the future. However, mortality in the inner chambers of the cave was always very high. On the contrary, at the entrance of the cave, nestling mortality showed the usual low 372 373 levels (~10%) estimated at other storm petrel colonies (Sanz-Aguilar et al., 2009) 374 where tick prevalence is lower (Merino et al., 1999). For example, in Benidorm Island 375 only one of the 34 storm petrel nestlings inspected by Merino et al. (1999) in 1996 376 presented a single Ornithodoros spp. tick larvae. Additionally, a tick larvae was detected only on 4 out of 27 storm petrel nestlings inspected at Benidorm Island on 377 378 25th June 2019 (tick larvae prevalence of 15%; own unpublished data). Monitoring of 379 tick infestation could be conducted to explore potential increase over time, such as 380 that reported as a function of warmer winter temperature in an arctic population of

381 Brünnich's guillemots (Uria lomvia) (Descamps, 2013). Prevalence of ticks infesting 382 seabirds has been shown to be very variable among seabird species, breeding colonies 383 and/or years ranging 0% and > 70% (Feare, 1974; Merino et al., 1999; Ramos et al., 2001; Descamps, 2013; Hipfner et al., 2019). However, the effects of ticks on hosts 384 385 depend also on the tick load, the host species, the age of the host (e.g. the effects can 386 be different for nestlings and adults), and/or the environmental conditions, as explained above (Feare, 1974; Merino et al., 1999; Ramos et al., 2001; McCoy et al., 387 388 2002; Descamps, 2013; Hipfner et al., 2019). For example, Feare (1976) documented a 67% of tick prevalence on Sooty terns and high mortality effects, while Hipfner et al. 389 390 (2019) documented tick prevalence > 60% for two species of auklet nestlings without 391 relevant survival effects.

Although we cannot determine the ultimate cause of nestlings' death, there are 392 393 several non-exclusive possibilities (Dietrich et al., 2011; Yabsley et al., 2012). First, 394 anaemia caused by blood loss could be lethal for young nestlings infested by high 395 numbers of ticks (Wanless et al., 1997). In fact, several studies have evidenced 396 negative effects of ticks on nestlings' body condition, at least during harsh 397 environmental conditions or high parasite loads (Morbey, 1996; Bosch and Figuerola, 398 1999; McCoy et al., 2002; Hipfner et al., 2019). Given their very small body size (< 10 g 399 when hatching, Davis, 1957), storm petrel nestlings could be more susceptible to 400 ectoparasites as ticks than larger seabird species (Merino et al., 1999). In fact, no 401 mortality was recorded for nestlings of Balearic shearwaters (adult weight ~500 g) at 402 the study site during the study period. In this vein, Hipfner et al. (2019) did not find 403 negative effects of tick infestation on survival or nestlings growth of two medium size 404 seabird species (adult weight 150-500 g), Cerorhinca monocerata and Ptychoramphus

405 *aleuticus,* at normal tick *I. uriae* loads; but they found negative effects at very high tick loads, in accordance with other evidences in other seabird species (Descamps, 2013). 406 407 In agreement with our results, they also documented a reduction in tick load with 408 nestling age, being essentially zero at fledging (Hipfner et al., 2019). However, contrary 409 to hard Ixodes ticks, soft Ornithodoros nymph and adult ticks are only attached to their 410 hosts for very short blood meals, at night (Vial, 2009), making it difficult to establish the actual tick load experienced by the hosts. We could only count the larvae ticks 411 412 feeding on nestlings during the monitoring visits, while adults and nymphs were likely 413 also feeding on both adult and chick storm petrels. However, adults breeding in both 414 areas showed similar body condition, which suggest that ticks may have higher 415 negative effects on nestlings than on adults.

Second, the low survival of nestlings in the inner part of the cave may be a
consequence of a massive adult abandonment of young nestlings as a result of high
parasite load, as shown in other species (Feare, 1976; King et al., 1977; Duffy, 1983;
Reeves et al., 2006). However, we think that this possibility is highly improbable. Some
field observations using camera trapping indicates that the adults came back to feed
the chicks when they were already dead. Moreover, neighbours of already dead chicks
continued incubating or rearing their surviving chicks (Fig. 1A).

Finally, ticks may be hosting infectious agents pathogenic for their seabird hosts, at least for young nestlings. Our study shows that two tick-borne agents known for their pathogenic potential for vertebrates were detected among the 11 pools of ticks that were analysed. We detected RNA from WNV genotype 3, which suggests that the virus is circulating in the storm petrel population, possibly relatively independently from other cycles of that virus in the area. West Nile virus can cause high mortality in

429 birds, particularly in naïve populations (George et al., 2015), and neurological disorders 430 in horses and humans. Moreover, the virus could also cause sub-lethal unknown 431 negative effects (Nemeth et al., 2006). The major vector species of WNV are mosquitoes, in particular ornithophilic species such as Culex pipiens (Calistri et al., 432 433 2010; Brugman et al., 2018). To date, little information is available on the mosquito 434 species associated to nesting areas of storm petrels, as well as the possible role of soft 435 ticks in the virus circulation. WNV genotype 3 have been previously isolated from O. 436 maritimus (Hoogstraal et al., 1976) and proved to be transmitted in laboratory by other soft tick species, such as O. moubata and O. capensis (Lawrie et al., 2004; 437 Hutcheson et al., 2005). Consequently, the O. capensis complex may act as a reservoir 438 439 of this genotype of WNV (Lawrie et al., 2004) and deserve to be further investigated. 440 Moreover, DNA from a Borrelia sp. very similar to relapsing fever B. turicatae was 441 detected. It may appear surprising to detect DNA of such as bacteria in ticks from 442 storm petrel because B. turicatae has been mostly reported in the southern USA and 443 Latin America (Schwan et al., 2005). Nevertheless, relapsing fever Borrelia has also 444 been detected in ticks from a storm petrel and shearwater colony in Japan (Takano et 445 al., 2009), in African penguins (Spheniscus demersus) (Yabsley et al., 2012), in brown 446 pelicans (Pelecanus occidentalis) (Reeves et al., 2006) and recently in ticks from Yellow-447 legged gull (Larus michahellis) nests in Algeria (Lafri et al., 2017). The detection of those Borrelia with a very similar sequence to B. turicatae thus suggests that further 448 449 investigations are required to understand how widespread are relapsing fever Borrelia 450 in seabird ticks (Kim et al., 2016). In addition, considering the lack of host specificity of 451 Ornithodoros spp. and the pathogenic potential of relapsing fever Borrelia for other 452 vertebrates, including humans, the question of the extent of the circulation of such

bacteria arises. Given the growing interest in relapsing fever *Borrelia* in America and
the biomedical tools that may become available to detect the past exposure of hosts
by tracking antibody levels (Lopez et al., 2013), investigations could rely on the
detection of the *Borrelia* in ticks, but also sometime more efficiently by detecting
antibodies in the vertebrate host (Armstrong et al., 2018).

458 **Conclusions**

In conclusion, our study provides evidence that O. maritimus may directly or 459 460 indirectly be the responsible of the extremely high mortality of storm petrel nestlings observed at the study site. Our results open exciting questions that need to be 461 addressed for a better understanding of the ecology and epidemiology of the 462 463 interactions between soft ticks and breeding Procellariiform species. Is the case of 464 Espartar unique among storm petrel colonies? What is the ultimate cause of nestling 465 mortality? Will the colony persist under such high levels of parasite infestation and 466 nestling mortality? Is it possible to detect any sub-lethal effects of ticks or pathogens 467 on adults? How do tick population dynamics work? Are the tick-borne virus and 468 bacteria circulating in the storm petrel population relatively independently from other 469 vertebrate host populations? Understanding how these host-parasite systems 470 function in space and time will require further challenging but potentially rewarding 471 multidisciplinary collaborations (taxonomy, ecology, and epidemiological studies). 472

473 **Conflict of interest**

474 The authors declare no conflicts of interest.

475

476 Ethical statement

477	This study was authorized by the Reserves des Vedrà es Vedranell i els illots de Ponent,
478	Balearic Government. All aspects of the study were performed according to guidelines
479	established for the ethical treatment of animals and complied with current Spanish
480	regulations. The collection of ticks was authorized by the Balearic Government
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496	Data accessibility
497	Data will be available via the Spanish National Research Council data repository. Sanz-
498	Aguilar, A., Payo-Payo, A., Igual, J. M., Rotger, A., Viñas Torres, M., Picorelli, V., 2019.
499	Storm petrel data sets. http://dx.doi.org/10.20350/digitalCSIC/

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- 681

Table 1. Prevalence of *O. maritimus* larvae on storm petrel nestlings during the

683 monitoring period at the entrance and the inner part of the colony (sample size in

684 parentheses).

	03/07/19	12/07/19	19/07/19	02/08/19	23/08/19
Entrance	100% (3)	57% (7)	28% (18)	5% (21)	0% (13)
Inner chambers	100% (6)	78% (9)	89% (19)	52% (33)	8% (25)

Table 2. Model selection of the effects of zone and age on tick *O. maritimus* load of
Storm petrel nestlings in Cap des Migdia Cave. df, degrees of freedom; Dev, Deviance;
ΔAIC, Akaike's information criterion difference with the best model; "+", additive
effect.

Effect	df	Dev	ΔΑΙC
Chick age*Zone	5	613.204	0
Chick age+Zone	4	634.002	18.797
Chick age	3	648.081	30.877
Zone	3	927.631	310.426
Null model	2	939.427	320.222

Table 3. Model selection of the effects of the zone, the individual tick *O. maritimus*larvae load and the maximum tick larvae load recorded for each individual during the
monitoring period on body condition index of storm petrel nestlings and adults. Note
that for adults, only the zone is tested as no tick larvae were found attached to them.
Notation as in Table 1.

Group	Effect	df	Dev	ΔAIC
Nestlings	Log(Maximum No. ticks +1)	3	-34.232	0
Nestlings	Log(Maximum No. ticks +1)+Zone	4	-34.246	1.986
Nestlings	Log(No.ticks +1)*Zone	5	-35.808	2.424
Nestlings	Log(No.ticks +1)	3	-31.538	2.694
Nestlings	Log(Maximum No. ticks +1)*Zone	5	-34.268	3.964
Nestlings	Null model	2	-28.061	4.172
Nestlings	Log(No.ticks +1)+Zone	4	-31.779	4.454
Nestlings	Zone	3	-29.136	5.096
Adults	Null model	1	0.446	0
Adults	Zone	2	0.440	1.627

- **Table 4.** Model selection of the effects of year and zone on the reproductive
- parameters: Hatching success (HS), Fledgling success (FS) and Breeding Success (BS) of
- storm petrels breeding in Cap des Migdia Cave. Notation as in Table 1.
- 705

Parameter	Effect	df	Dev	ΔΑΙϹ
HS	Year+Zone	7	1180.305	0
HS	Year*Zone	11	1178.445	6.141
HS	Zone	3	1197.818	9.513
HS	Year	6	1194.097	11.793
HS	Null model	2	1210.125	19.820
FS	Year+Zone	7	661.359	0
FS	Year*Zone	11	654.552	1.193
FS	Zone	3	679.188	9.829
FS	Year	6	734.838	71.480
FS	Null model	2	756.804	85.446
BS	Zone	3	1130.221	0
BS	Year+Zone	7	1127.815	5.595
BS	Year*Zone	11	1120.786	6.565
BS	No effect (constant)	2	1203.073	70.852
BS	Year	6	1200.250	76.029

- 707 Figure 1. A) Inner chambers of the storm petrel colony at Espartar Island. In the
- photograph can be observed one incubating adult, three alive nestlings and two dead
- nestlings; B) adults soft tick (O. maritimus) and C) larvae of soft tick (O. maritimus) on a
- 710 storm petrel nestling.



- **Figure 2**. A) Map of the study area and the studied species; B) Infographic of Cap des
- 713 Migdia Cave study areas: Entrance and inner chambers. Representation is informative
- 714 but not topologically accurate.



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Figure 3. A) Relationship between tick *O. maritimus* larvae load and storm petrel
nestlings age. Dotted lines are the individuals' predictions and solid lines the mean
prediction, Table 2, with grey and black lines representing the entrance and the inner
chambers of the colony respectively. B) Boxplot showing the distribution of the
number of tick *O. maritimus* larvae per storm petrel nestlings at the entrance and the
inner chambers of the colony.



Figure 4. A) Relationship between body condition index of storm petrel nestlings and
the maximum tick *O. maritimus* load (log scale) detected for each individual (Table 3),
the black line represent the estimate of the model and the grey area the SE. Points
indicate the individual predictions.



Figure 5. A) Annual mean (and 95% CI) of hatching success (HS), B) fledgling success
(FS) and C) breeding success (BS) of storm petrels breeding in the entrance (grey) and
the inner chambers (black) of Cap des Migdia Cave between 2014 and 2018. Estimates
from the model including the additive effects of zone and year (Table 4).

