

Importance of chloroplastic/mitochondrial interactions during elicitor- induced cell death in tobacco leaves: acceleration by light and low oxygen

Rosine de Paepe, Marie Garmier, Pierrick Priault, Guillaume Vidal, Simon Driscoll, Reda Djebbar, Martine Boccara, Chantal Mathieu, Christine Foyer

▶ To cite this version:

Rosine de Paepe, Marie Garmier, Pierrick Priault, Guillaume Vidal, Simon Driscoll, et al.. Importance of chloroplastic/mitochondrial interactions during elicitor- induced cell death in tobacco leaves: acceleration by light and low oxygen. 14th International Congress of Photosynthesis, Jul 2007, Glasgow, United Kingdom. hal-03078887

HAL Id: hal-03078887

https://hal.science/hal-03078887

Submitted on 16 Dec 2020

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers. L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

Importance of chloroplastic/mitochondrial interactions during elicitor- induced cell death in tobacco leaves: acceleration by light and low oxygen.

De Paepe R¹ Garmier M¹, Priault P², Vidal G¹, Driscoll S³, Djebbar R⁴, Boccara M⁵, Mathieu C¹, and Foyer CH³

- 1 Institut de Biotechnologie des Plantes, UMR-CNRS 8618, Université Paris-Sud Sud, 91405 Orsay France. ² Ecologie, Systématique et Evolution, UMR-CNRS 8079, Université Paris-Sud Sud, 91405 Orsay, France. ³ Crop Performance and Improvement Division, Rothamsted Research, Harpenden, U.K. ⁴Laboratoire de Physiologie Végétale, Université des Sciences et de la Technologie Houari Bournedienne,, Alger, Algérie ⁵ Atelier de bioinformatique, Université Pierre et Marie Curie Paris 6 75005 Paris, France

INTRODUCTION

Light and oxygen are considered important in the execution of plant programmed cell death (PCD), principally because they are involved in the generation of reactive oxygen species (ROS)

We previously showed that the bacterial elicitor harpin N_{ea}from Erwinia amylovora induces an hypersensitive response (HR)-like PCD process in N. sylvestris leaves

In the light, harpin induces a respiratory burst involving alternative oxidase (AOX) engagement as determined by oxygen isotope discrimination (Vidal et al., 2007), changes in chloroplastic (Boccara et al. 2007), stomatal opening (Boccara et al. 20001) and changes in the expression of antioxidant genes (Garmier et al., 2002).

Here, we compare the HR-like responses of leaves elicited the

- in the light and in darkness
- in air and under very low oxygen (VL ox: < 0,1%)

SUMMARY

The present work shows that, in contrast with other plant PCD systems, neither light nor oxygen are necessary to harpin N-induced cell death. In all conditions, the HR-lire response is an active processs, involving contrasting responses of defence and antioxidant genes. Moreover, examination of response signatures in mitochondria (respiration) and in chloroplasts (photosynthesis), together with the intracellular localisation of ROS accumulation,, provides compelling evidence for strong interactions between respiration and photosynthetic metabolism in the modulation of the cell death. While extra-mitochondrial ROS are not key signalsin the execution of the regulated cell suicide programme, mitochondrial signals could be of major importance.

- REFERENCES

 Boccara et al (2001) IR thermography revealed a role for mitochondria in presymptomatic cooling during harpin-induced hypersensitive response. Plant J. 28, 663ara et al. (2007) Early chloroplastic alterations analysed by optical coherence tomography (OCT) during harpin-induced hypersensitive response. Plant J., in
- lochem. 40, 561-566

 Vidal et al. (2007) Lack of respiratory chain Complex I impairs AOX engagement and modulates redox signaling during elicitor-induced cell death in tobacco. Plant

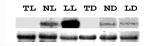
RESULTS

I.1 Symptoms induction

Harpin N-induced cell death was induced in the light and in darkness, in air and under anoxia

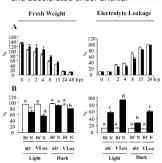


I.2 Accumulation of transcripts of the tobacco Tnt1 transposon 4 hours post harpin N, indicates an active cell death process in all conditions



TL: untreated illuminated leaf; NL: harpin 4h pi, in the ight and atm ox; LL: harpin 4h pi, in the light and atm ox; LL: harpin 4h pi, in the light and under VL ox; TD: untreated leaf in the dark; ND: harpin 4h pi, in the dark and under atm ox; LD: harpin 4h pi, in the dark and under VL ox.

However. fresh weight and electrolyte measurements indicate that cell death was delayed in the dark and accelerated under anoxia.



A A comparison of harpin elicitation in the light (350 μmol m² s¹) and in the dark under atm CO₂ and O₂ concentrations; open columns: light, closed columns: dark; h pi: hours post harpin elicitation.

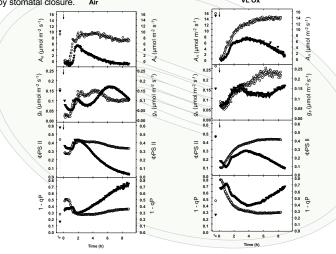
B. A comparison of harpin (N) and buffer (Bf) infiltration under atm ox s and VL ox (< 0.1%), at the 5 h time point; open columns; buffer, closed columns: harpin N.

II Photosynthetic and Respiratory signatures of harpin-elicited leaves

II.1 Photosynthetic and fluorescence parameters of light-elicited leaves

Collapse of photosynthesis from two hours post elicitation is caused by inactivation of PSII and not by stomatal closure.

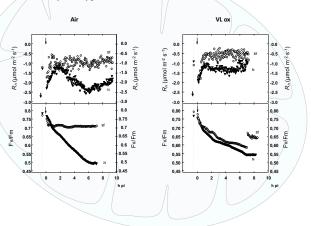
VL 0x by stomatal closure.



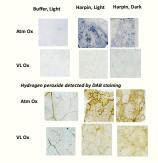
II.2 CO2 release in leaves infiltrated in the dark

The respiratory burst induced in air was nearly abolished under VLox, probably due to the inhibition of AOX which has lower

affinity for oxygen than COX.

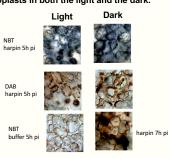


III.1 In situ detection of ROS 5 h post harpin elicitation.



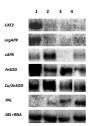
ROS accumulation peaked 4-6 h pi in both the light and the dark. In contrast it was strongly attenuated

III. 2 Microscopic examination indicate accumulation of reactive oxygen species inside chloroplasts in both the light and the dark.



A: harpin 3h pi, light; B: harpin 5h pi, light; C: harpin 9 pi, dark; D: harpin 3h pi, light; E: harpin 5h pi, light; F: harpin 5h pi, dark; G: harpin 7h pi, light; H: harpin 7h pi, dark; I: buffer 5h pi, light. Red arrows: chloroplasts; black arrows: collapsed cells.Bar: $1 \text{ cm} = 100 \ \mu\text{m}$

III.3 Expression of antioxidant and defence genes 4 h post harpin elicitation (Northern).



1: untreated leaf in the light under atm ox; 2: harpin-infiltrated leaf in the light under atm ox; 3: harpin-infiltrated leaf in the dark under atm ox; 4: harpin-infiltrated leaf in the light

eased transcript levels of organellar antioxidant, CAT2, orgAPX and chlFeSOD, were observed in all

On the other hand, cytosolic Cu/Zn SOD and cAPX accumulated only in the light under atm conditions.

The PAL defence gene accumulated in all cases.