



HAL
open science

Diallelic self-incompatibility is the main determinant of fertilization patterns in olive orchards

Roberto Mariotti, Saverio Pandolfi, Isabelle de Cauwer, Pierre Saumitou-Laprade, Philippe Vernet, Martina Rossi, Federica Baglivo, Luciana Baldoni, Soraya Mousavi

► To cite this version:

Roberto Mariotti, Saverio Pandolfi, Isabelle de Cauwer, Pierre Saumitou-Laprade, Philippe Vernet, et al.. Diallelic self-incompatibility is the main determinant of fertilization patterns in olive orchards. *Evolutionary Applications*, 2020, 10.1111/eva.13175 . hal-03078884

HAL Id: hal-03078884

<https://hal.science/hal-03078884>

Submitted on 17 Dec 2020

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

Diallelic self-incompatibility is the main determinant of fertilization patterns in olive orchards

Running title: Cross compatibility relationships in olive

Roberto Mariotti¹^, Saverio Pandolfi¹^, Isabelle De Cauwer²^, Pierre Saumitou-Laprade², Philippe Vernet², Martina Rossi¹, Federica Baglivo^{1,3}, Luciana Baldoni^{1*}, Soraya Mousavi¹

¹ CNR, Institute of Biosciences and Bioresources, 06028 Perugia, Italy

² Univ. Lille, CNRS, UMR 8198 - Evo-Eco-Paleo, F-59000 Lille, France

³ CNR, Institute for Mediterranean Agricultural and Forestry Systems, 06128 Perugia, Italy

^These authors have equally contributed to the present study

*Correspondence:

Luciana Baldoni

luciana.baldoni@ibbr.cnr.it

Abstract

Introduction - Self-incompatibility (SI) in flowering plants potentially represents a major obstacle for sexual reproduction, especially when the number of S-alleles is low. The situation is extreme in the commercially important olive tree, where *in vitro* pollination assays suggested the existence of a diallelic SI (DSI) system involving only two groups (G1 and G2). Varieties belonging to the same SI group cannot fertilize each other, such that successful fruit production is predicted to require pollination between varieties of different groups.

Methods and Results – To test this prediction, we explored the extent to which the DSI system determines fertilization patterns under field conditions. 117 olive cultivars were first genotyped using ten highly polymorphic dinucleotide Simple Sequence Repeat (SSR) markers to ascertain varietal identity. Cultivars were then phenotyped through controlled pollination tests to assign

This article has been accepted for publication and undergone full peer review but has not been through the copyediting, typesetting, pagination and proofreading process, which may lead to differences between this version and the [Version of Record](#). Please cite this article as [doi: 10.1111/EVA.13175](https://doi.org/10.1111/EVA.13175)

This article is protected by copyright. All rights reserved

each of them to one of the two SI groups. We then collected and genotyped 1,440 open pollinated embryos from five different orchards constituted of seven local cultivars with known group of incompatibility groups. Embryos genotype information were used: (i) to assign embryos to the most likely pollen donor genotype in the neighborhood using paternity analysis, and (ii) to compare the composition of the pollen cloud genetic among recipient trees in the five sites. The paternity analysis showed that the DSI system is the main determinant of fertilization success under field open pollination conditions: G1 cultivars sired seeds exclusively on G2 cultivars, and reciprocally. No self-fertilization events were observed.

Discussion - Our results demonstrate that DSI is a potent force determining pollination success among varieties within olive orchards used for production. They have the potential to improve management practices by guiding the selection of compatible varieties to avoid planting orchards containing sets of varieties with strongly unbalanced SI groups, as these would lead to suboptimal olive production.

Keywords

Incompatibility, paternity analysis, stigma test, olive cultivars, pollen donor, pollen cloud.

Acknowledgements

We are grateful to Vincent Castric for scientific discussions and careful reading of the MS. We appreciate also the Zagaria Olive Germplasm Conservation Field for their help and technical assistance. We thank Bouchaïb Khadari, Abdelmajid Moukhli, Laila Essalouh, Pierre Mournet and Ronan Rivallan for providing some plant material (INRA-CRRA Marrakech, Morocco). Special thanks to Giulia Cascini for the activities performed within the Training Program N. 0000318 A/16/CO. We also thank the owners of olive orchards exploited in the present manuscript: Luigi Brunelli, Giovanni Batta, Graziano Decimi, Antonio Pellerucci, Andrea and Marco Erolì. We are finally grateful to Riccardo Pandolfi for the picture gave us that I was submitted as potential cover picture.

Funding

The research was supported by the European Union's Horizon 2020 Research and Innovation Program Marie Skłodowska-Curie - Before Project (Grant Agreement No 645595); by the EU projects "OLIVE4CLIMATE – LIFE" (LIFE15 CCM/IT/000141); by the Rural Development Program of Umbria Region: years 2007-2013 – Measure 1.2.4 projects POLLINATOR – The best pollinators of olive varieties in Umbria; years 2014-2020 – Measure 16.2.1, projects INNO.V.O. - Development of alternative varieties to face the new challenges of olive growing, and OMEGA - Organizational models for the efficiency of Umbrian farms.

Conflict of Interest

The authors declare no conflicts of interest.

Data Availability Statement

The data that support the findings of this study, which are not already reported in the supplementary material files, are available from the corresponding author upon reasonable request.

MR. ROBERTO MARIOTTI (Orcid ID : 0000-0002-9111-5123)

DR. PIERRE SAUMITOU-LAPRADE (Orcid ID : 0000-0001-7413-3090)

Article type : Original Article

Diallelic self-incompatibility is the main determinant of fertilization patterns in olive orchards

Running title: Cross compatibility relationships in olive

Roberto Mariotti^{1^}, Saverio Pandolfi^{1^}, Isabelle De Cauwer^{2^}, Pierre Saumitou-Laprade², Philippe Vernet², Martina Rossi¹, Federica Baglivo¹, Luciana Baldoni^{1}, Soraya Mousavi¹*

¹ CNR, Institute of Biosciences and Bioresources, 06028 Perugia, Italy

² Univ. Lille, CNRS, UMR 8198 - Evo-Eco-Paleo, F-59000 Lille, France

[^]These authors have equally contributed to the present study

*Correspondence:

Luciana Baldoni

luciana.baldoni@ibbr.cnr.it

Abstract

Introduction - Self-incompatibility (SI) in flowering plants potentially represents a major obstacle for sexual reproduction, especially when the number of S-alleles is low. The

situation is extreme in the commercially important olive tree, where *in vitro* pollination assays suggested the existence of a diallelic SI (DSI) system involving only two groups (G1 and G2). Varieties belonging to the same SI group cannot fertilize each other, such that successful fruit production is predicted to require pollination between varieties of different groups.

Methods and Results – To test this prediction, we explored the extent to which the DSI system determines fertilization patterns under field conditions. 117 olive cultivars were first genotyped using ten highly polymorphic dinucleotide Simple Sequence Repeat (SSR) markers to ascertain varietal identity. Cultivars were then phenotyped through controlled pollination tests to assign each of them to one of the two SI groups. We then collected and genotyped 1,440 open pollinated embryos from five different orchards constituted of seven local cultivars with known group of incompatibility groups. Embryos genotype information were used: (i) to assign embryos to the most likely pollen donor genotype in the neighborhood using paternity analysis, and (ii) to compare the composition of the pollen cloud genetic among recipient trees in the five sites. The paternity analysis showed that the DSI system is the main determinant of fertilization success under field open pollination conditions: G1 cultivars sired seeds exclusively on G2 cultivars, and reciprocally. No self-fertilization events were observed.

Discussion - Our results demonstrate that DSI is a potent force determining pollination success among varieties within olive orchards used for production. They have the potential to improve management practices by guiding the selection of compatible varieties to avoid planting orchards containing sets of varieties with strongly unbalanced SI groups, as these would lead to suboptimal olive production.

Keywords

Incompatibility, paternity analysis, stigma test, olive cultivars, pollen donor, pollen cloud.

Abstract

Introduction - Self-incompatibility (SI) in flowering plants potentially represents a major obstacle for sexual reproduction, especially when the number of S-alleles is low. The situation is extreme in the commercially important olive tree, where *in vitro* pollination assays suggested the existence of a diallelic SI (DSI) system involving only two groups (G1 and G2). Varieties belonging to the same SI group cannot fertilize each other, such that successful fruit production is predicted to require pollination between varieties of different groups.

Methods and Results – To test this prediction, we explored the extent to which the DSI system determines fertilization patterns under field conditions. 117 olive cultivars were first genotyped using ten highly polymorphic dinucleotide Simple Sequence Repeat (SSR) markers to ascertain varietal identity. Cultivars were then phenotyped through controlled pollination tests to assign each of them to one of the two SI groups. We then collected and genotyped 1,440 open pollinated embryos from five different orchards constituted of seven local cultivars with known group of incompatibility groups. Embryos genotype information were used: (i) to assign embryos to the most likely pollen donor genotype in the neighborhood using paternity analysis, and (ii) to compare the composition of the pollen cloud genetic among recipient trees in the five sites. The paternity analysis showed that the DSI system is the main determinant of fertilization success under field open pollination conditions: G1 cultivars sired seeds exclusively on G2 cultivars, and reciprocally. No self-fertilization events were observed.

Discussion - Our results demonstrate that DSI is a potent force determining pollination success among varieties within olive orchards used for production. They have the potential to improve management practices by guiding the selection of compatible varieties to avoid planting orchards containing sets of varieties with strongly unbalanced SI groups, as these would lead to suboptimal olive production.

Keywords

Incompatibility, paternity analysis, stigma test, olive cultivars, pollen donor, pollen cloud.

Introduction

Despite a vast majority of species being hermaphroditic, self-incompatibility is pervasive among flowering plants (Barrett, 2010). During a given reproductive episode, this mechanism allows avoiding self-fertilization and inbreeding, but also reduces the number of effective male/female combinations. Understanding how self-incompatibility shapes the patterns of fertilization is crucial both in wild plant species, where a decrease in the number of compatible partners might threaten long term population persistence (Wagenius et al., 2007, Leducq et al., 2010), and in cultivated crops, where the number of compatible partners can directly impact fruit yield (Muñoz-Sanz et al., 2020). In self-incompatible fruit crops, compatible genotypes (often referred to as pollen donors) are frequently spread through fields/orchards, a management practice that is used for instance in apple, pear or apricot (Herrera et al., 2018, Kwon et al., 2015, Sanzol and Robbins, 2008). Despite the crucial impact of the mating system on fertilization patterns, the occurrence/functioning of self-incompatibility is not known for all crop species.

Cultivated olive (*Olea europaea* L., subsp. *europaea*, var. *europaea*), a wind pollinated species grown for fruit production, blooms profusely and produces pollen in great abundance, but only 2-3% of more than 500,000 flowers produced by a mature tree usually set fruits (Martin et al., 2005). While it is known that climatic conditions (abundant rain or high temperatures) during the pollination period can contribute to decrease the fruit set (Koubouris et al., 2009; Oteros et al., 2014), the availability of suitable pollen might also be a limiting factor in the field. Indeed, cultivated olive varieties are self-incompatible and require suitable pollen donors to ensure successful fertilization (Rugini and De Pace, 2016). Self-incompatibility and incompatibility between genotypes sharing SI specificities potentially represent important reproductive barriers in olive.

It is now well-established that olive possesses a sporophytic SI system (DSI), characterized by the unusual presence of only two SI specificities (Saumitou-Laprade et al., 2017a). Based on robust in-vitro stigma tests, each olive genotype can be reliably assigned to one of two SI groups: genotypes belonging to one group are incompatible with each other but compatible with all genotypes of the other group and *vice versa* (but see Breton et al., 2014, 2016; Farinelli et al., 2018). Up to now, this system has been exclusively observed in species within the Oleaceae family, associated with various mating systems: the hermaphrodite cultivated olive (Saumitou-Laprade et al., 2017a), the androdioecious *Phillyrea angustifolia* (Billiard et al., 2015; Saumitou-Laprade et al. 2010) and *Fraxinus ornus* (Vernet et al., 2016),

the polygamous *F. excelsior* (Saumitou-Laprade et al., 2018) and the hermaphrodite *Ligustrum vulgare* (De Cauwer et al., 2020).

A limitation of these data is that most of the empirical evidence obtained so far relies on stigma tests, i.e. observations of pollen tube trajectories in hand pollinated stigmas. A recent study using paternity analysis in a wild relative, the Saharan wild *Olea europaea* subsp. *laperrinei* (Besnard et al. 2020) confirmed the occurrence of only two SI groups in an experimental population, but the effect of DSI on fruit production remains to be demonstrated directly under field conditions (Farinelli et al. 2018). Obtaining such data is crucial, especially in the context of the current shift to super-intensive cultivation, which is increasingly based on mono-varietal olive orchards running the risk of economic failure by lack of consideration of the mating properties of the planted varieties.

In this study, we first characterize the SI phenotypes of 117 of the most commonly used olive varieties in the Mediterranean area, and we provide their multi-locus genotype at a set of genetic markers for varietal identification. Based on these data, we determined the extent to which the DSI system is shaping fertilization patterns in field conditions. This knowledge was obtained through paternity analysis of seeds produced in open pollination, after exhaustively genotyping their mother trees and as well as all potential pollen donors in a large 500m neighborhood using a set of highly polymorphic genetic markers. Overall, our results provide unequivocal support for the hypothesis that DSI is a potent force under field open pollination conditions and should be taken into account in the design of production orchards.

Materials and Methods

Plant material

Cultivars chosen for correspondence between genotype and SI group

A set of 117 olive cultivars, including elite varieties currently used in the most important olive oil producing countries, such as Arbequina, Koroneiki, Picual and Frantoio, has been phenotyped to determine the incompatibility group through *in vitro* stigma tests, following the procedure described in Saumitou-Laprade et al. (2017a). For 76 cultivars, information on incompatibility groups was extracted from the previous work of Saumitou-Laprade et al. (2017a and b) and 41 supplementary cultivars were *de novo* phenotyped (Supplementary Table S1). For *de novo* stigma test experiments, pollen, as well as fresh leaves for DNA extractions, were collected from the following olive varietal collections: the World Olive

Germplasm Collection of Marrakech (INRA, Morocco), the Zagaria Olive Germplasm Conservation Field (Enna, Italy), the Perugia Collection (Perugia, Italy) (Saumitou-Laprade et al., 2017a) and the Olive Collection established at CNR-IBBR (Perugia, Italy). The cross-compatible cultivars (cvs.) Leccino (belonging to the incompatibility group [G1]) and Dolce Agogia [G2] represented our recipient pair of tester genotypes: their stigmas were used as recipients to phenotype the pollen samples collected from the screened cultivars. Each cultivar was thus phenotyped for SI at the prezygotic stage and genotyped on a set of SSR markers (see below).

Experimental sites description and estimation of varietal composition

Olive orchards are generally constituted of thousands of trees possessing the same genotype and paternity analyses carried out in olive groves need to take into account the clonal origin of individuals, *i.e.* several replicas for each genotype, unlike what happens in natural systems in which each potential donor has a genotype different from all the others. Paternity analysis has rarely been used in olive groves due to the technical difficulties of genotyping all potential fathers surrounding the mother trees on which the progenies would be harvested (Mookerjee et al., 2005; Baruca Arbeiter et al., 2014). In the current study, we took advantage of previously gathered information about the identity of varieties in cultivated areas of an Italian olive-producing region to estimate paternity likelihoods. Given the abundance of olive trees in typical production regions, another difficulty is deciding the relevant scale at which the paternity analysis should be carried out, *i.e.* deciding which genotypes should be considered as potential pollen donors. The surface area that one should sample around mother trees to maximize the number of successful pollen donors in the data set will depend on pollen dispersal patterns. Since olive is wind pollinated, one might suspect that pollen can travel over long distances. However, even if pollen can travel over dozen, even hundreds of kilometers carried by the wind (Besnard et al., 2009; Hernández et al., 2011; Fernández-Rodríguez et al., 2014; Rojo et al., 2016), paternity analyses usually reveal that long distance effective pollen dispersal is not a ubiquitous characteristic of anemophilous species, including *Olea europaea* subsp. *cuspidata* (De Cauwer et al. 2012; Burczyk et al. 1997, 2002, 2004; Bacles and Ennos 2008; Kassa et al., 2018). In this subspecies, over 75% of the pollen dispersal occurred within less than 200 meters around mother trees (Kassa et al., 2018). Assuming pollen dispersal patterns are similar between the two related subspecies, we

chose to consider any tree within a radius of 500 meters around focal mother plants as a potential pollen donor.

In order to observe how the DSI system shapes gamete exchanges under field conditions, seven varieties growing in olive orchards of different areas of Umbria region (Italy) were selected for fruit harvesting. Five sites were chosen (Figure 1): Site 1 (Giano dell'Umbria, 42.839 N, 12.573 E), Site 2 (Narni, 42.491 N, 12.493 E), Site 3 (Amelia, 42.539 N, 12.467 E), Site 4 (Corciano, 43.150 N, 12.277 E) and Site 5 (Perugia, 43.097 N, 12.409 E). For each study site, all trees located in an area of 78.5 hectares were considered as potential contributors to the local pollen cloud (Table 1). About 30% of the total surveyed surface (112.06 hectares) corresponded to multiple olive groves and included 24,788 olive trees (Table 1B). The cultivars' occurrence at each site was ascertained by combining the results from previous genotyping work (Baldoni et al., 2003; Mariotti et al., 2009; Pandolfi et al., 2010), specific surveys based on morphological evaluations and interviews with olive producers for each area (Table 1A; Table 1B).

Criteria to select mother plants

In the Umbria region, where the study was conducted, there are about 7.5 million olive trees, corresponding to a few main cultivars. In the National Olive Cultivars Register, Umbria is represented by 11 genotypes, all included in our set of samples, either as mother plants or as possible pollen donors together with dozens of local or minor cultivars, in some cases represented by a few or single trees (Mousavi et al., 2019). Among them, cv. Moraiolo is the most widespread, representing about 35% of total olive trees, whereas cvs. Leccino and Dolce Agogia, have a medium diffusion, and cvs. Gentile Grande, Borgiona, San Felice and Raio show a local distribution restricted to a few hectares (Baldoni et al., 2003; Mariotti et al., 2009; Pandolfi et al., 2010).

In order to examine the effects of the site, the genotype and the SI group, seven cultivars grown in five different sites were selected (Table 1A). The site effect was tested by including cv. Moraiolo, a G1 cultivar widely used in Umbria and which was present in three different study sites: 1 (Giano), 2 (Narni), and 3 (Amelia). In order to study the SI group effect, we selected two cultivars belonging to G1 and G2 groups in three sites. In particular, in site 3 (Amelia), we selected the cultivars Moraiolo [G1] and Raio [G2], in site 4 (Corciano) Gentile Grande [G1] and Borgiona [G2] cultivars, and in site 5 (Perugia) the cvs. Leccino [G1] and Dolce Agogia [G2]. Finally, in order to assess the genotype effect within site, we selected

two [G1] cultivars, Moraiolo and San Felice in site 1 (Giano). In total, we collected fruits from nine different [Genotype x Site] combinations and on seven cultivars (four [G1] and three [G2]) in five different sites (Figure 1). In each site, 160 fruits were collected from four different trees from each cultivar (replicates A, B, C, and D), resulting in 36 sampled mothers.

Within each site, the average distance between the four plant replicates (A, B, C, and D) of the same genotype was 20 meters (Figure 1). Regarding the four sites where two maternal varieties were selected (sites 1, 3, 4 and 5), the average distance between trees belonging to different varieties varied from 24 meters (Site 1) to 120 meters (Site 4). For each mother tree, healthy fruits were harvested at ripening stage in order to reach at least ten fully formed embryos from each cardinal direction of the canopy, resulting in a total of 1,440 embryos (Table 2 and Supplementary Figure S1). Pulp and pit were removed from each fruit, seeds were cut-up with a scalpel and embryos were pulled out and used for DNA extraction.

Varietal and embryo genotyping

DNA was extracted from leaf samples of *de novo* phenotyped trees (41 cultivars), from the 36 mother plants of the field test, as well as from their 1,440 embryos, by using the GeneElute Plant Genomic DNA Miniprep Kit (Sigma-Aldrich) and applying the standard manufacturer's instructions.

These samples were genotyped by using ten dinucleotide Simple Sequence Repeat (SSR) markers recognized as the most effective for cultivar characterization (Baldoni et al., 2009; El Bakkali et al., 2019; Trujillo et al., 2014; Mousavi et al., 2017a and b; Hosseini-Mazinani et al., 2014) and for parentage analyses (Mookerjee et al., 2005; Seifi et al., 2012; Beghe et al., 2017; Kassa et al., 2018). The analyzed loci included DCA3-5-9-16-18 (Sefc et al., 2000), EMO90 (de la Rosa et al., 2002), GAPU71B-101-103A (Carriero et al., 2002) and UDO-043 (Cipriani et al., 2002). PCR amplifications were performed separately in a final reaction volume of 25 μ l containing 25 ng of DNA, 1 \times PCR buffer, 200 μ M of each dNTP, 10 pmol of each forward and reverse primer, and 2 U of Q5 High-Fidelity DNA Polymerase (New England Biolabs), with an initial denaturation at 95°C for 5 min, followed by 40 cycles of 95°C for 30 s, annealing temperature as suggested by authors (50–60°C) for 30 s and 72°C for 25 s, followed by a final elongation at 72°C for 40 min.

PCR products were loaded on an ABI 3130 Genetic Analyzer (Applied Biosystems, Foster City, CA, USA) using the internal GeneScan 500 LIZ Size Standard (Thermo Fisher

Scientific). Fragment sizes were analyzed using GeneMapper 3.7 (Applied Biosystems). The profiles of mother plants, four replicates of each genotype from the same olive cultivar, were used as an additional internal controls to select the correct length for each allele during allele calling process.

Varietal identity of all genotyped individuals, including mother plants of the five study sites, was ascertained through the comparison of the SSR data obtained in our study with previously published genotypes (Trujillo et al., 2014; Mousavi et al., 2017a and b; El Bakkali et al., 2019). A consensus profile was built following the reference allele sizes provided by Baldoni et al. (2009) (Supplementary Table S1).

GenAlEx 6.501 software (Peakall and Smouse, 2006) was used to calculate the probability of identity (P_i), which provides an estimate of the average probability that two unrelated individuals drawn from the same population will have the same multilocus genotype. Moreover, the same software was utilized to build a genetic distance matrix by using the default parameters for codominant markers (Smouse and Peakall, 1999). The triangular matrix was exported into MEGA X software (Kumar et al., 2018) to construct a neighbor joining tree (Saitou and Nei et al., 1987). A dendrogram was built using the Figtree software 1.4.3 (Rambaut and Drummond, 2012) to allow the visualization of the clustering of individuals and incompatibility phenotypes in the data set.

Paternity analysis

Paternity was determined for the 1,440 embryos by using the maximum likelihood-based method described in Kalinowski et al. (2007) and implemented in CERVUS version 3.0.3 (Marshall et al., 1998). Because of the clonal nature of olive varieties, paternity could not be determined on the basis of individual putative pollen donor trees, but could still be assigned with very high power at the level of individual varieties. Accordingly, we considered all varieties in the 78.5 ha areas surrounding mother trees in a site as potential pollen donors. Using this approach, LOD scores were computed for every embryo – potential paternal variety combination and simulations were used to determine the critical Δ , *i.e.* the minimal difference in LOD scores between the two most likely paternal varieties required to assign paternity with 95% confidence. Allele frequencies used in the simulations for each study site were based on the genotypes of locally cultivated varieties weighted by their relative abundances based on orchard surface areas (Table 1A). Within each site, the number of potential paternal genotypes for each incompatibility group varied between six (Site 2) and

thirteen (Site 4). 10,000 offspring were simulated, allowing for selfing and using the following parameters: the number of candidate fathers was the number of cultivars occurring in each site; the proportion of sampled fathers was estimated based on the surface occupied by determined genotypes in each site (Table 1A). The proportion of ‘typed loci’, ‘mistyped’ and ‘minimum typed loci’ was always set at 0.99, 0.01 and 10, respectively with a relaxed value of 95% and the strict level at 99%. After simulation runs, paternity analyses were carried out within each site, determining for each embryo the two most likely fathers and deciding whether paternity could be assigned to the most likely father based on the critical Δ obtained from the simulations.

Male gamete heterogeneity among females

The multi-locus genotypes of the 36 maternal trees and the 1,440 embryos were then used to conduct a gametic analysis of molecular variance within and among mother plants using the TwoGener method, two generational analysis of pollen flow for codominant data (Smouse et al., 2001) implemented in the GenAlex 6.501 software (Peakall and Smouse, 2006). Mother plant/embryo comparisons were used to extract the haploid paternal contributions. A matrix of haploid genetic distances between all pairs of male gametes was built. An AMOVA was then conducted on this gametic distance matrix to partition the overall molecular variance among and within mothers. For all possible mother pairs (*i.e.* 630 comparisons), the proportion of the molecular variance was estimated using the ϕ_{FT} index. The significance of these pairwise ϕ_{FT} values was assessed using 1,000 random permutations of male gametes between mothers.

Results

Assignment of cultivars to the corresponding incompatibility group and genotype

Based on stigma tests, all 117 varieties could be clearly assigned to either of the two incompatibility groups, in line with Saumitou-Laprade et al (2017a). Specifically, for each variety used as pollen donor we consistently observed a compatibility reaction on one of the two cultivars used as stigma tester (either Leccino or Dolce Agogia). This compatibility reaction was characterized by several pollen tubes growing and converging through the stigmatic tissue towards the style and successive attrition, such that only one or two tubes reached the style. On the other stigma tester, we observed an incompatibility reaction with the absence of pollen tubes or with the presence of only short pollen tubes growing within the

stigma but never reaching the style (Supplementary Table S1, Supplementary Figure S2). With no exception, all varieties showed strictly asymmetrical reactions, i.e. none of them showed either compatibility or incompatibility on both tester lines. We observed that the two incompatibility groups are equally frequent among Italian (21 G1 and 27 G2; χ^2 test = 0.386, df = 1) and Spanish cultivars (10 G1 and 14 G2; χ^2 test = 0.234, df = 1) (Supplementary Table S1). The neighbor-joining tree highlighted the presence of synonymous cultivars, which clustered together (Supplementary Figure S3). In fact, from the 117 phenotyped cultivars a unique SSR profile was detected for 85 of them. The cases of shared profiles are reported in Supplementary Table S1. The probability of identity (P_i) was low in our data set ($P_i=7.5 \cdot 10^{-14}$), suggesting that the 10 SSR loci are polymorphic enough to distinguish unrelated individuals. Identical genotypes thus very likely correspond to clonal replicates, originally propagated from the same individual and named differently in various regions or countries. These identical genotypes were always assigned to the same SI group based on the stigma tests. The SSR profiles and SI group of all studied varieties are reported in Supplementary Table S1. The 36 olive trees selected as mother plants in the paternity analysis below all showed genetic profiles corresponding with their reference variety (7 different genotypes in total). Of these seven varieties, 4 were phenotyped as [G1], and 3 as [G2] (Table 2).

Paternity analysis

Paternity analysis in each of the five sampling sites allowed assigning paternity to a single variety in 82.91% of cases. The results highlighted the effect of the SI group on fertilization success (Table 2). Among the 1,440 progenies analyzed, no selfing events were detected, confirming strict self-incompatibility of the studied cultivars under field conditions. Over the whole data set, 18 different pollen donors were identified. Among the 1,188 embryos that were successfully assigned, not a single case of within-group fertilization was detected (Table 2, Supplementary Figure S4): the 960 embryos collected from the 24 [G1] mothers were sired by 12 paternal genotypes, all from the [G2] group; whereas the 480 embryos collected from twelve [G2] mothers were sired by six paternal genotypes, all from the [G1] group. Within sites, 20 out of 36 mothers were pollinated mainly by a single cultivar, siring more than 70% of embryos (Table 2),

Heterogeneity of the pollen cloud fertilizing individual trees

The gametic analysis of molecular variance within and among mother plants provides a powerful way to characterize the genetic structure of the pollen cloud. While most of the overall genetic variation in male gametes composition (83%) was found within mother plants, a statistically significant share of the total variation occurred among mothers ($\phi_{FT} = 0.169$, $P < 0.001$), indicating that individual trees are pollinated by distinct pollen clouds. Pairwise ϕ_{FT} values computed for all possible mother pairs in the data set are reported in Supplementary Table S2.

Within-site/within-variety pairwise comparisons. Within each of the five selected sites, the four trees sampled for each studied variety allowed us to verify whether replicates of the same genotype located at close vicinity sampled male gametes from the same pollen cloud. Across the 54 within-site/within-genotype pairwise comparisons, the average ϕ_{FT} value was low (mean \pm SD = 0.002 ± 0.004) and with a single exception were not significantly different from zero (Supplementary Table S2). Overall, when replicates of the same genotype occurred in the same orchard, they thus tended to sample the same pollen cloud.

Between-sites/within-variety pairwise comparisons. Twelve Moraiolo mother plants were sampled in sites 1, 2 and 3 (four replicates per site). Geographic distances between these orchards were as follow: 39.08 km between orchard 1 and 2, 5.75 km between orchard 2 and 3 and 34.67 km between orchards 1 and 3 (Figure 1). All the 48 between-sites/within-variety pairwise comparisons were significant, indicating that replicates of the same genotype placed in orchards separated by a few to a few tens of kilometers sampled different pollen clouds ($\phi_{FT} = 0.229 \pm 0.066$), illustrating that local pollination environments can be different.

Between-group comparisons. Three study sites included both [G1] and [G2] mothers (site 3: Moraiolo [G1] and Raio [G2]; site 4: Gentile Grande [G1] and Borgiona [G2]; and site 5 Leccino [G1] and Dolce Agogia [G2]), allowing to jointly explore incompatibility group and site effects (Figure 2). Among all possible comparisons involving trees from these three sites, all pairwise ϕ_{FT} values were significant except for within-site/within-group comparisons (as described above). Two mother trees located in the same orchard but belonging to different groups sampled male gametes in pollen clouds that were as different as the pollen clouds sampled by two mother trees located in different orchards (Figure 2).

Finally, in one site, two varieties from [G1] were sampled (site 1, with four Moraiolo and four San Felice trees). Within this site, among the 16 comparisons involving the two varieties, 15 showed low but significant pollen cloud genetic differentiation (mean $\phi_{FT} \pm$ SD = 0.034 ± 0.014). This last result highlighted the fact that beyond the strong effects of self-incompatibility group and among site geographic distance documented above, different

genotypes belonging to the same group and located in the same orchard can sample slightly different pollen clouds.

Discussion

Theoretical predictions and DSI effect in orchards

The discovery of the diallelic self-incompatibility system in cultivated olive, with the presence of only two inter-compatible groups, implies that chances of fertile combinations among cultivars are limited to 50% of all possible crosses, if frequencies of each group are equal (Saumitou-Laprade et al., 2017a). In areas planted with olive varieties belonging to the same SI group or in mono-cultivar olive orchards, the limitation in compatible pollen may represent a very strong hindrance to fertilization and fruit production (Guerin et al., 2000).

Our study, conducted in five sites in Umbria, exemplify how strongly the DSI shapes the effective pollen flow in orchards. The results, we obtained in a situation of olive production, are in agreement with the conclusions of a recent study in the Saharan wild *Olea europaea* subsp. *laperrinei* (Besnard et al. 2020). Strict self-incompatibility appears to be the rule under field conditions for the seven olive cultivars used as mothers in our study and the 37 wild *Olea europaea* subsp. *laperrinei*. A phenomenon, generally thought to be a consequence of environmental factors interfering with the SI reaction or to result from the action of modifier genes (Busch & Schoen, 2008; Levin, 1996), is reported as “leaky” SI. As already discussed in (Saumitou-Laprade et al., 2017a), it is observed in some olive cultivars like Koroneiki (Marchese 2016) and Casaliva a homonym of Frantoio, (Moreno Sanz 2020). These cultivars belong to a SI group (Table S1) and present a low rate of selfing.

The main effect of the DSI remains in the distribution of the pollination success detected by the paternity analysis. Among the paternity assignments of 1,188 embryos resulting from open pollination of seven different cultivars by 22 different pollen donor varieties, not a single case of within-group fertilization was detected. This very strict within group incompatibility detected in cultivated olive is also in agreement with the complete absence of progenies produced by crosses within group reported in *Olea europaea* subsp. *laperrinei* (Besnard et al. 2020).

In the olive production areas across the Mediterranean Basin, flower fecundation is expected to be guaranteed by the presence of millions of olive trees therefore the availability of compatible pollen should be assured (Ferri et al., 2008; Pinillos and Cuevas, 2009;

Aguilera et al., 2013). Nevertheless, the presence of compatible pollen might seriously decrease in traditional areas of olive cultivation. Indeed, these areas are submitted to cultivar substitution, abandonment of ancient olive trees, plant death after severe winter frost or pathogenic attacks that reduce the number of genotypes present in a region.

At regional scale, within the two most important olive producing regions of Spain (Andalucía) and Italy (Apulia), the frequency of the two DSI groups were equilibrate in terms of number of cultivars. In Andalucía, Hojiblanca and Lechin de Sevilla [G1] traditionally grow close to Picual and Manzanilla Cacereña [G2] trees. In Apulia, popular varieties include Ogliarola Barese and Nociara [G1], as well as Coratina and Peranzana [G2]. Nevertheless, within the sites we studied in Umbria, while the number of [G1] and [G2] cultivars was similar, the G1 group was strongly overrepresented in terms of number of trees. This bias reaches 85% of [G1] for 5% [G2] and 10% unknown trees in site 2, which may locally result in [G2] pollen limitation.

The unavailability of compatible pollen could represent a serious problem mainly for the new cultivation areas, where olive growing has recently spread away from the traditional cultivation regions, towards other continents or at the opposite hemisphere, without the presence of any other olive or wild relative. The displacement of a few cultivars into new territories, as has occurred in the last 20 years, may result in risky situations because of the potential lack of compatible pollen. In these geographically isolated orchards with intensive mono-varietal olive plantations, pollination by compatible donors could be compromised (Wu et al., 2002; Mehri et al., 2003; Koubouris et al., 2010; Spinardi and Bassi, 2012). Based on our results, the fruiting failure reported in isolated mono-varietal orchards (Ayerza and Coates, 2004; Pinillos and Cuevas, 2009) may be related to limitation of compatible pollen, as the presence within orchards of both G1 and G2 cultivars represents a prerequisite for successful fertilization, and hence fruiting.

All these results and observations highlight how the knowledge of the SI group is of utmost importance to optimize pollination success in olive orchard. In order to contribute to this optimization, we provide a first list of 117 cultivars, including clonal cases with different varietal names, typed for SI group using stigma tests and fully genotyped with reliable SSR markers (Baltoni et al., 2009). This set of olive cultivars encompasses a large proportion of the entire variability of cultivated olive (Belaj et al., 2012 and 2018; Díez et al., 2012; Mousavi et al., 2017) and includes the most important olive cultivars for oil and table olive production.

Fertilization rates among compatible cultivars

Beyond the major effect of the mating system, we observed that recipient trees from different collection sites have been fertilized by genetically differentiated pollen clouds, and to a lesser extent this was also true among varieties sharing a given SI group. Pollen clouds pollinating the same genotype, *i.e.* Moraiolo, in three different olive orchards were significantly different. The genetic differentiation was strong and of the same order of magnitude as the differentiation observed between genotypes belonging to two different SI groups. These differences in pollen cloud among sites can be related to the varying contribution of different pollen donors among sites. For instance, on Moraiolo recipients, Pendolino sired 79.4% of embryos in site 1, 23.1% in site 2 and no embryos in site 3. The variable success of this variety was not clearly related to its local abundance within the three sites (15.4%, 40%, and 8% respectively). Explaining such patterns will require additional field experiments, involving more sites, considering the spatial positions of the trees, the main wind direction and the variation in flowering phenology among varieties and among sites.

Even within site, two varieties belonging to the same incompatibility group sampled slightly different pollen clouds, as exemplified in site 1 with the Moraiolo and San Felice recipients. While the level of genetic differentiation detected among mothers belonging to these two varieties was much lower than the levels of pollen cloud differentiation detected among sites or between groups, they were significant and higher than within site / within variety comparisons. Explaining why two different genotypes belonging to the same SI group do not sample the exact same pollen cloud will also require further studies.

In five out of nine combinations (mother/sites), a single cultivar sired more than 70% of the embryos even if other compatible pollen donors were available locally. Eighty-five percent of Moraiolo embryos were pollinated by cv. Raio; the coexistence of these cultivars was found in several ancient olive orchards of that area (Baldoni et al., 2003; Mariotti et al., 2009; Pannelli et al., 2010), highlighting a prominent affinity between these varieties already observed by farmers hundreds of years ago. Pendolino cultivar [G2], previously recognized as a good pollen donor (Solfanelli et al., 2006), was confirmed as a very important pollen donor for [G1] recipient cultivars. It contributed to fertilize four out of six [G1] mother genotypes and was the main pollen donor in one site for cvs. Moraiolo and San Felice, despite its low abundance in that area. The reasons why some compatible cultivars can succeed as pollen donors better than others are not yet established (Selak et al., 2014;

Montemurro et al., 2019). Precisely documenting the extent of this intriguing phenomenon and deciphering the factors involved will constitute an interesting next step.

Conclusions

The present work confirms the occurrence of only two SI groups within a wide range of genetically verified olive cultivars. It demonstrates how strongly DSI shapes the pollination patterns among cultivars in actual orchards and establishes that the assignment of genotypes to a SI group by *in vitro* stigma tests in the lab is fully predictive of the complete inability of a cultivar to pollinate another cultivar from the same SI group in the field. Now that the importance of the effect of the DSI on pollination patterns is validated under field conditions, the large-scale deployment of this approach at an agricultural level will require further analyses to obtain information on the incompatibility group of as much cultivars as possible. This may be facilitated by the recent discovery of STS markers linked to the SI phenotype (Mariotti et al., 2020).

Acknowledgements

We are grateful to Vincent Castric for scientific discussions and careful reading of the MS. We appreciate also the Zagaria Olive Germplasm Conservation Field for their help and technical assistance. We thank Bouchaïb Khadari, Abdelmajid Moukhli, Laila Essalouh, Pierre Mournet and Ronan Rivallan for providing some plant material (INRA-CRRA Marrakech, Morocco). Special thanks to Giulia Cascini for the activities performed within the Training Program N. 0000318 A/16/CO. We also thank the owners of olive orchards exploited in the present manuscript: Luigi Brunelli, Giovanni Batta, Graziano Decimi, Antonio Pellerucci, Andrea and Marco Erolì. We are finally grateful to Riccardo Pandolfi for the picture gave us that I was submitted as potential cover picture.

Data Availability Statement

The data that support the findings of this study, which are not already reported in the supplementary material files, are available from the corresponding author upon reasonable request.

Funding

The research was supported by the European Union's Horizon 2020 Research and Innovation Program Marie Skłodowska-Curie - Before Project (Grant Agreement No 645595); by the EU projects "OLIVE4CLIMATE – LIFE" (LIFE15 CCM/IT/000141); by the Rural Development Program of Umbria Region: years 2007-2013 – Measure 1.2.4 projects POLLINATOR – The best pollinators of olive varieties in Umbria; years 2014-2020 – Measure 16.2.1, projects INNO.V.O. - Development of alternative varieties to face the new challenges of olive growing, and OMEGA - Organizational models for the efficiency of Umbrian farms.

Conflict of Interest

The authors declare no conflicts of interest.

References

- Aguilera, F., Ruiz Valenzuela, L. (2013). Time trend in the viability of pollen grains in the ‘Picual’ olive (*Olea europaea* L.) cultivar. *Palynology* 37(1), 28-34. doi: 10.1080/01916122.2012.662920
- Ayerza, R., Coates, W. (2004). Supplemental pollination–increasing olive (*Olea europaea*) yields in hot, arid environments. *Exp. Agric.* 40, 481–491. doi: 10.1017/S0014479704002133
- Bacles, C. F., Ennos, R. A. (2008). Paternity analysis of pollen-mediated gene flow for *Fraxinus excelsior* L. in a chronically fragmented landscape. *Heredity* 101, 368-380. doi: 10.1038/hdy.2008.66
- Baldoni, L., Ricciolini, C., Munari, C., Pannelli, G., Arcioni, S. (2003). DNA characterization of olive varieties and ecotypes in Umbria. *Proc. Nat. Cong. Olive Germplasm and Oil Typicality*, pp. 312-315.
- Baldoni, L., Cultrera, N.G., Mariotti, R., Ricciolini, C., Arcioni, S., Vendramin, G.G., et al. (2009). A consensus list of microsatellite markers for olive genotyping. *Mol. Breed.* 24, 213–231. doi: 10.1007/s11032-009-9285-8
- Barrett SCH. (2010). Understanding plant reproductive diversity. *Philosophical Transactions of the Royal Society of London B* 365, 99–109.
- Baruca Arbeiter, A., Jakše, J., Bandelj, D. (2014). Paternity analysis of the olive variety “Istrska Belica” and identification of pollen donors by microsatellite markers. *Sci. World J.* ID 208590. doi: 10.1155/2014/208590
- Beghe, D., Piotti, A., Satovic, Z., de la Rosa, R., Belaj, A. (2017). Pollen-mediated gene flow and fine-scale spatial genetic structure in *Olea europaea* subsp. *europaea* var. *sylvestris*. *Ann. Bot.* 119, 671–679. doi: 10.1093/aob/mcw246
- Belaj, A., del Carmen Dominguez-García, M., Atienza, S.G., Urdíroz, N.M., de la Rosa, R., Satovic, Z., et al. (2012). Developing a core collection of olive (*Olea europaea* L.) based on molecular markers (DArTs, SSRs, SNPs) and agronomic traits. *Tree Genet. Genomes* 8, 365-378. doi: 10.1007/s11295-011-0447-6
- Belaj, A., De La Rosa, R., Lorite, I.J., Mariotti, R., Cultrera, N.G.M., Beuzón, C.R., et al. (2018). Usefulness of a new large set of high throughput EST-SNP markers as a tool for olive germplasm collection management. *Front. Plant Sci.* 9, 1320. doi: 10.3389/fpls.2018.01320

- Besnard, G., Baali-Cherif, D., Bettinelli-Riccardi, S., Parietti, D., Bouguedoura, N. (2009). Pollen-mediated gene flow in a highly fragmented landscape: consequences for defining a conservation strategy of the relict Laperrine's olive. *C. R. Biol.* 332, 662-672. Doi: 10.1016/j.crvi.2009.02.003
- Besnard, G., Cheptou, P. O., Debbaoui, M., Lafont, P., Huguény, B., Dupin, J., Baali-Cherif, D. (2020). Paternity tests support a diallelic self-incompatibility system in a wild olive (*Olea europaea* subsp. *laperrinei*, Oleaceae). *Ecol. Evol.* 10, 1876–1888. doi: 10.1002/ece3.5993
- Billiard, S., Husse, L., Lepercq, P., Godé, C., Bourceaux, A., Lepart, J., et al. (2015). Selfish male-determining element favors the transition from hermaphroditism to androdioecy. *Evolution* 69, 683-693. doi: 10.1111/evo.12613
- Breton, C.M., Bervillé, A. (2012). New hypothesis elucidates self-incompatibility in the olive tree regarding S-alleles dominance relationships as in the sporophytic model. *C. R. Biol.* 335, 563–572. doi: 10.1016/j.crvi.2012.07.006
- Breton, C. M., Farinelli, D., Shafiq, S., Heslop-Harrison, J. S., Sedgley, M., Bervillé, A. J. (2014). The self-incompatibility mating system of the olive (*Olea europaea* L.) functions with dominance between S-alleles. *Tree Genet. Genomes* 10, 1055-1067. doi: 10.1007/s11295-014-0742-0
- Breton, C. M., Farinelli, D., Koubouris, G., Bervillé, A. (2016). A model based on S-allele dominance relationships to explain pseudo self-fertility of varieties in the olive tree. *Euphytica* 210, 105-117. doi: 10.1007/s10681-016-1708-0
- Breton, C.M. Bervillé, A.J. (2018). Identification of olive pollen donor trees and pollinizers under controlled pollination-environment using STR markers. *Aust. J. Crop Sci.* 12, 1566. doi: 10.21475/ajcs.18.12.10.p1071
- Burczyk, J. Prat, D. (1997). Male reproductive success in *Pseudotsuga menziesii* (Mirb.) Franco: the effects of spatial structure and flowering characteristics. *Heredity*, 79, 638. doi: 10.1038/hdy.1997.210
- Burczyk, J., Adams, W.T., Moran, G.F., Griffin, A.R. (2002). Complex patterns of mating revealed in a *Eucalyptus regnans* seed orchard using allozyme markers and the neighbourhood model. *Mol. Ecol.* 11, 2379-2391. doi: 10.1046/j.1365-294X.2002.01603.x
- Burczyk, J., Lewandowski, A., Chalupka, W. (2004). Local pollen dispersal and distant gene flow in Norway spruce (*Picea abies* [L.] Karst.). *For. Ecol. Manag.* 197, 39-48. doi: 10.1016/j.foreco.2004.05.003

- Carriero, F., Fontanazza, G., Cellini, F., Giorio, G. (2002). Identification of simple sequence repeats (SSRs) in olive (*Olea europaea* L.). *Theor. Appl. Genet.* 104(2-3), 301-307. doi: 10.1007/s001220100691
- Cipriani, G., Marrazzo, M.T., Marconi, R., Cimato, A., Testolin, R. (2002). Microsatellite markers isolated in olive (*Olea europaea* L.) are suitable for individual fingerprinting and reveal polymorphism within ancient cultivars. *Theor. Appl. Genet.* 104(2-3), 223-228. doi: 10.1007/s001220100685
- Cruden, R.W. (2000). Pollen grains: why so many? *Plant Syst. Evol.* 222, 143–165. doi: 10.1007/bf00984100
- De Cauwer, I., Arnaud, J.F., Klein, E.K., Dufay, M. (2012). Disentangling the causes of heterogeneity in male fecundity in gynodioecious *Beta vulgaris* ssp. *maritima*. *New Phytol.* 195, 676–687. doi: 10.1111/j.1469-8137.2012.04191.x
- De Cauwer, I., Vernet, P., Billiard, S., Godé, C., Bourceaux, A., Ponitzki, C., Saumitou-Laprade, P. (2020). Widespread coexistence of self-compatible and self-incompatible phenotypes in a diallelic self-incompatibility system in *Ligustrum vulgare* (Oleaceae). bioRxiv 2020.03.26.009399; doi: 10.1101/2020.03.26.009399
- De la Rosa, R., James, C. M., & Tobutt, K. R. (2002). Isolation and characterization of polymorphic microsatellites in olive (*Olea europaea* L.) and their transferability to other genera in the Oleaceae. *Mol. Ecol. Notes* 2, 265-267. doi: 10.1046/j.1471-8286.2002.00217.x
- Díez, C. M., Imperato, A., Rallo, L., Barranco, D., Trujillo, I. (2012). Worldwide core collection of olive cultivars based on simple sequence repeat and morphological markers. *Crop Sci.* 52, 211-221. doi: 10.2135/cropsci2011.02.0110
- El Bakkali, A., Essalouh, L., Tollon, C., Rivallan, R., Mournet, P., Moukhli, A., et al. (2019). Characterization of Worldwide Olive Germplasm Banks of Marrakech (Morocco) and Córdoba (Spain): Towards management and use of olive germplasm in breeding programs. *PloS One* 14(10), e0223716. doi: 10.1371/journal.pone.0223716
- Fernández-Rodríguez, S., Skjøth, C.A., Tormo-Molina, R., Brandao, R., Caeiro, E., Silva-Palacios, I., et al. (2014). Identification of potential sources of airborne *Olea* pollen in the Southwest Iberian Peninsula. *Int. J. Biometeorol.* 58, 337-348. doi: 10.1007/s00484-012-0629-4
- Farinelli, D., Breton, C., Koubouris, G., Famiani, F., Villemur, P., & Bervillé, A. (2018). Reply to Saumitou-Laprade et al. (2017). Controlling for genetic identity of varieties,

pollen contamination and stigma receptivity is essential to characterize the self-incompatibility system of *Olea europaea* L. *Evol. Appl.* 11, 1465-1470. doi: 10.1111/eva.12633

Ferri, A., Bellini, E., Padula, G., Giordani, E. (2008). Viability and in vitro germinability of pollen grains of olive cultivars and advanced selections obtained in Italy. *Adv. Hortic. Sci.* 22(2), 1000-1007.

Frankel, R. Galun, E. (2012). *Pollination mechanisms, reproduction and plant breeding.* Springer Science & Business Media. doi: 10.1007/978-3-642-81059-6_2

Guerin, J., Collins, G., Sedgley, M. (2000). Selection and breeding of olive cultivars. *Outlook Agric.* 29, 269–274. doi: 10.5367/000000000101293338

Hernández-Ceballos, M.A., García-Mozo, H., Adame, J.A., Domínguez-Vilches, E., Benito, A., Bolívar, J.P., Galán, C. (2011). Synoptic and meteorological characterisation of olive pollen transport in Córdoba province (south-western Spain). *Int. J. Biometeorol.* 55, 17-34. doi: 10.1007/s00484-010-0306-4

Herrera, S., Rodrigo, J., Hormaza, J. I., Lora, J. (2018). Identification of self-incompatibility alleles by specific PCR analysis and S-RNase sequencing in apricot. *Int. J. Mol. Sci.* 19, 3612. doi: 10.3390/ijms19113612

Hosseini-Mazinani, M., Mariotti, R., Torkzaban, B., Sheikh-Hassani, M., Ataei, S., Cultrera, N. G., et al. (2014). High genetic diversity detected in olives beyond the boundaries of the Mediterranean Sea. *PLoS One* 9(4), e93146. doi: 10.1371/journal.pone.0093146

Iwano, M. Takayama, S. (2012). Self/non-self discrimination in angiosperm self-incompatibility. *Curr. Opin. Plant Biol.* 15, 78–83. doi: 10.1016/j.pbi.2011.09.003

Kalinowski, S.T., Taper, M.L., Marshall, T.C. (2007). Revising how the computer program Cervus accommodates genotyping error increases success in paternity assignment: Cervus Likelihood Model. *Mol. Ecol.* 16, 1099–1106. doi: 10.1111/j.1365-294X.2007.03089.x

Kassa, A., Konrad, H., Geburek, T. (2018). Mating pattern and pollen dispersal in the wild olive tree (*Olea europaea* subsp. *cuspidata*). *Tree Genet. Genomes* 14, 3. doi: 10.1007/s11295-017-1215-z

Koubouris, G.C., Metzidakis, I.T., Vasilakakis, M.D. (2009). Impact of temperature on olive (*Olea europaea* L.) pollen performance in relation to relative humidity and genotype. *Environ. Exp. Bot.* 67, 209–214. doi: 10.1016/j.envexpbot.2009.06.002

- Koubouris, G.C., Metzidakis, I.T., Vasilakakis, M.D. (2010). Influence of cross-pollination on the development of parthenocarpic olive (*Olea europaea*) fruits (shotberries). *Exp. Agric.* 46, 67–76. doi: 10.1017/s0014479709990500
- Kumar, S., Stecher, G., Li, M., Knyaz, C., Tamura, K. (2018). MEGA X: Molecular evolutionary genetics analysis across computing platforms. *Mol. Biol. Evol.* 35, 1547–1549. doi: 10.1093/molbev/msy096
- Kwon, S. I., Yoo, J., Lee, J., Moon, Y. S., Choi, C., Jung, H. Y., Lee, D. H., Kim, C. K., Kang, I. K. (2015). Evaluation of crab apples for apple production in high-density apple orchards. *J. Plant Biotechnol.* 42, 271-276. doi: 10.5010/JPB.2015.42.3.271
- Leducq, J. B., Gosset, C. C., Poiret, M., Hendoux, F., Vekemans, X., Billiard, S. (2010). An experimental study of the S-Allele effect in the self-incompatible plant *Biscutella neustriaca*. *Cons. Genet.* 11, 497–508. doi: 10.1007/s10592-010-0055-2
- Mariotti, R., Pannelli, G., Cultrera, N., Baldoni, L. (2009). Evolution of the olive varieties: the case of Umbria. *Eleiva, Proc. Cong. The origins of the Tuscan olive patrimony, San Quirico d’Orcia, Siena.*
- Mariotti R., Fornasiero A., Mousavi S., Cultrera N.G.M., Brizioli F., Pandolfi S., et al. (2020). Genetic mapping of the incompatibility locus in olive and development of a linked STS marker. *Front. Plant Sci.* 10, 1760. doi: 10.3389/fpls.2019.01760.
- Marshall, T.C., Slate, J.B.K.E., Kruuk, L.E.B., Pemberton, J.M. (1998). Statistical confidence for likelihood- based paternity inference in natural populations. *Mol. Ecol.* 7, 639-655. doi: 10.1046/j.1365-294x.1998.00374.x
- Martin, G.C., Ferguson, L., Sibbett, G.S. (2005). Flowering, pollination, fruiting, alternate bearing, and abscission, p. 49-54. In: *Olive Production Manual*. Sibbett, G.S., Ferguson, L., Coviello, J.L., Lindstrand, M. (Eds.). University of California, Agriculture and Natural Resources, Oakland, California.
- McCallum, M. (2005). *Tiberis Navigabilis: Commercial activity between Rome and the Middle Tiber Basin during the Roman period*. State University of New York at Buffalo.
- Mehri, H., Mehri-Kamoun, R., Msallem, M., Faïdi, A., Polts, V. (2003). Reproductive behavior of six olive cultivars as pollinizer of the self-incompatible olive cultivar Meski. *Adv. Hortic. Sci.* 42–46.

- Miller, A.J. Gross, B.L. (2011). From forest to field: perennial fruit crop domestication. *Am. J. Bot.* 98, 1389–1414. doi: 10.3732/ajb.1000522
- Montemurro, C., Dambruoso, G., Bottalico, G., Sabetta, W. (2019). Self-incompatibility assessment of some Italian olive genotypes (*Olea europaea* L.) and cross-derived seedling selection by SSR markers on seed endosperms. *Front. Plant Sci.* 10, 451. doi: 10.3389/fpls.2019.00451
- Mookerjee, S., Guerin, J., Collins, G., Ford, C., Sedgley, M. (2005). Paternity analysis using microsatellite markers to identify pollen donors in an olive grove. *Theor. Appl. Genet.* 111, 1174–1182. doi: 10.1007/s00122-005-0049-5
- Moreno-Sanz, P., L. Lombardo, S. Lorenzi, F. Michelotti and M. S. Grando (2020). "Genetic Resources of *Olea europaea* L. in the Garda Trentino Olive Groves Revealed by Ancient Trees Genotyping and Parentage Analysis of Drupe Embryos." *Genes* 11(10): 1171
- Mousavi, S., Mariotti, R., Bagnoli, F., Costantini, L., Cultrera, N. G., Arzani, K., et al. (2017a). The eastern part of the Fertile Crescent concealed an unexpected route of olive (*Olea europaea* L.) differentiation. *Ann. Bot.* 119, 1305–1318. doi: 10.1093/aob/mcx027
- Mousavi, S., Mariotti, R., Regni, L., Nasini, L., Bufacchi, M., Pandolfi, S., et al. (2017b). The first molecular identification of an olive collection applying standard simple sequence repeats and novel expressed sequence tag markers. *Front. Plant Sci.* 8, 1283. doi: 10.3389/fpls.2017.01283
- Mousavi, S., Stanzione, V., Mencuccini, M., Baldoni, L., Bufacchi, M., Mariotti, R. (2019). Biochemical and molecular profiling of unknown olive genotypes from central Italy: determination of major and minor components. *Eur. Food Res. Technol.* 245, 83–94. doi: 10.1007/s00217-018-3142-0
- Muñoz-Sanz, J. V., Zuriaga, E., Cruz-García, F., McClure, B., Romero, C. (2020). Self-(in)compatibility systems: target traits for crop-production, plant breeding, and biotechnology. *Front. Plant Sci.* 11, 195. doi: 10.3389/fpls.2020.00195
- Oteros, J., Orlandi, F., García-Mozo, H., Aguilera, F., Dhiab, A.B., Bonofiglio, T., et al. (2014). Better prediction of Mediterranean olive production using pollen-based models. *Agron. Sustain. Dev.* 34, 685-694. doi: 10.1007/s13593-013-0198-x
- Pandolfi, S., Baldoni, L., Mariotti, R., Alagna, F., Cultrera, N., Mattioli, T. (2010). Variability of olive and evolution of the landscape in the area of Trasimeno Lake. In:

Wonderland in the Landscape-Cultural Mosaic, Idea, Image, Illusion. Palmanova (UD).

Pannelli, G., Pandolfi, S., Baldoni, L., Bongi, G. (2010). Selection and use of ancient olive trees in Umbria. Proc. IV Nat. Cong. Mediterranean Plants, Sarli G., Alvino A., Cervelli C., 93-104.

Peakall, R.O.D., Smouse, P.E. (2006). GENALEX 6: genetic analysis in Excel. Population genetic software for teaching and research. Mol. Ecol. Notes 6, 288–295. doi: 10.1111/j.1471-8286.2005.01155.x

Pinillos, V., Cuevas, J. (2009). Open-pollination provides sufficient levels of cross-pollen in Spanish monovarietal olive orchards. HortSci. 44, 499–502. doi: 10.21273/hortsci.44.2.499

Rambaut, A., Drummond, A.J. (2012). FigTree version 1.4.0. Available: <http://tree.bio.ed.ac.uk/software/figtree/>.

Rojo, J., Orlandi, F., Pérez-Badia, R., Aguilera, F., Dhiab, A.B., Bouziane, H., et al. (2016). Modeling olive pollen intensity in the Mediterranean region through analysis of emission sources. Sci. Total Environ. 551-552, 73–82. doi: 10.1016/j.scitotenv.2016.01.193

Rugini E., De Pace C. (2016) Olive Breeding with Classical and Modern Approaches. In: The Olive Tree Genome. Compendium of Plant Genomes. Springer, Cham. doi: 10.1007/978-3-319-48887-5_10

Saitou, N., Nei, M. (1987). The Neighbor-joining method: a new method for reconstructing phylogenetic trees. Mol. Biol. Evol. 4(4), 406-425.

Sanzol, J., Robbins, T. P. (2008). Combined analysis of S-alleles in European pear by pollinations and PCR-based S-genotyping; correlation between S-phenotypes and S-RNase genotypes. J. Am. Soc. Hortic. Sci. 133, 213-224. doi: 10.21273/JASHS.133.2.213

Saumitou-Laprade, P., Vernet, P., Vassiliadis, C., Hoareau, Y., de Magny, G., Dommée, B., Lepart, J. (2010). A self-incompatibility system explains high male frequencies in an androdioecious plant. Science 327, 1648–1650. doi: 10.1126/science.1186687

Saumitou-Laprade, P., Vernet, P., Vekemans, X., Billiard, S., Gallina, S., Essalouh, L., et al. (2017a). Elucidation of the genetic architecture of self-incompatibility in olive: Evolutionary consequences and perspectives for orchard management. Evol. Appl. 10, 867–880. doi: 10.1111/eva.12457

- Saumitou-Laprade, P., Vernet, P., Vekemans, X., Castric, V., Barcaccia, G., Khadari, B., et al. (2017b). Controlling for genetic identity of varieties, pollen contamination and stigma receptivity is essential to characterize the self-incompatibility system of *Olea europaea* L. *Evol. Appl.* 10, 860–866. doi:10.1111/eva.12498.
- Saumitou-Laprade, P., Vernet, P., Dowkiw, A., Bertrand, S., Billiard, S., Albert, B., et al. (2018). Polygamy or subdioecy? The impact of diallelic self-incompatibility on the sexual system in *Fraxinus excelsior* (Oleaceae). *Proc. R. Soc. B Biol. Sci.* 285, 20180004. doi: 10.1098/rspb.2018.0004
- Sefc, K.M., Lopes, M.S., Mendonça, D., Santos, M.R.D., Machado, M.L.D.C., Machado, A.D.C. (2000). Identification of microsatellite loci in olive (*Olea europaea*) and their characterization in Italian and Iberian olive trees. *Mol. Ecol.* 9(8), 1171-1173. doi: 10.1046/j.1365-294x.2000.00954.x
- Seifi, E., Guerin, J., Kaiser, B., Sedgley, M. (2012). Sexual compatibility of the olive cultivar ‘Kalamata’ assessed by paternity analysis. *Span. J. Agric. Res.* 10, 731–740. doi: 10.5424/sjar/2012103-501-11
- Selak, G.V., Cuevas, J., Ban, S.G., Perica, S. (2014). Pollen tube performance in assessment of compatibility in olive (*Olea europaea* L.) cultivars. *Sci. Hortic.* 165, 36–43. doi: 10.1016/j.scienta.2013.10.041
- Smouse, P.E. Peakall, R.O.D. (1999). Spatial autocorrelation analysis of individual multiallele and multilocus genetic structure. *Heredity* 82, 561-573. doi: 10.1046/j.1365-2540.1999.00518.x
- Smouse, P.E., Dyer, R.J., Westfall, R.D., Sork, V.L. (2001). Two-generation analysis of pollen flow across a landscape. I. Male gamete heterogeneity among females. *Evolution* 55, 260–271. doi: 10.1111/j.0014-3820.2001.tb01291.x
- Solfanelli, C., Bartolini, S., Vitagliano, C., Lorenzi, R. (2006). Immunolocalization and quantification of IAA after self-and free-pollination in *Olea europaea* L. *Sci. Hortic.* 110(4), 345-351. doi: 10.1016/j.scienta.2006.06.026
- Spinardi, A. Bassi, D. (2012). Olive fertility as affected by cross-pollination and boron. *Sci. World J.* 2012, ID 375631, 8. doi: 10.1100/2012/375631
- Torimaru, T., Wennström, U., Lindgren, D., Wang, X.R. (2012). Effects of male fecundity, interindividual distance and anisotropic pollen dispersal on mating success in a Scots pine (*Pinus sylvestris*) seed orchard. *Heredity* 108(3), 312-321. doi: 10.1038/hdy.2011.76

- Accepted Article
- Trujillo, I., Ojeda, M.A., Urdiruz, N.M., Potter, D., Barranco, D., Rallo, L., Diez, C.M. (2014). Identification of the Worldwide Olive Germplasm Bank of Córdoba (Spain) using SSR and morphological markers. *Tree Genet. Genomes* 10, 141–155. doi: 10.1007/s11295-013-0671-3
- Vernet, P., Lepercq, P., Billiard, S., Bourceaux, A., Lepart, J., Dommée, B., Saumitou-Laprade, P. (2016). Evidence for the long-term maintenance of a rare self-incompatibility system in Oleaceae. *New Phytol.* 210, 1408–1417. doi: 10.1111/nph.13872
- Wagenius, S., Lonsdorf, E., Neuhauser, C. (2007). Patch aging and the S-Allee effect: breeding system effects on the demographic response of plants to habitat fragmentation. *Am. Nat.* 169, 383-397. doi: 10.1086/511313
- Wu, S.B., Collins, G., Sedgley, M. (2002). Sexual compatibility within and between olive cultivars. *J. Hortic. Sci. Biotechnol.* 77, 665-673. doi: 10.1080/14620316.2002.115115544

Figure legends

Figure 1. Map of the five study sites in the Umbria region of Italy: Site 1 (Giano dell'Umbria), Site 2 (Narni), Site 3 (Amelia), Site 4 (Corciano) and Site 5 (Perugia). Each circle represents an area of 78.5 hectares (*i.e.* 500 meters around mother trees). The yellow color highlights the orchard where mother plants were located, while the red color indicates other olive orchards nearby. Pink [G1] and blue [G2] arrows show the position of each mother tree from which fruits were collected.

Figure 2. Mean ϕ_{FT} values observed for different types of pairwise comparisons in sites 3, 4 and 5, where mothers from both groups were sampled. The graph summarizes the genetic differences found among and within the collecting sites in relation with incompatibility group.

Table 1. Distribution of the 22 varieties present in the five study sites in Umbria, Italy (Figure 1). **A:** For each observed cultivar, in column "% within site" is given the percentage of trees from that cultivar within site over the total number of trees present in this site (G1+G2+undetermined); in column "% within group" is given the percentage of trees from that cultivar within site over the total number of trees from the same SI group present in this site (G1 or G2). The cultivar of mother plants at each site is indicated as "*". **B:** For each site are presented: the total surface of olive orchards within site (hectares), the total number of olive trees within site, the number of G1 and G2 trees, as well as their proportion in % over the total number of trees (G1+G2+undetermined).

Cultivars	A																							
	Sites																							
	1				2				3				4				5							
SI group	% within site	% within group	Mother plants	% within site	% within group	Mother plants	% within site	% within group	Mother plants	% within site	% within group	Mother plants	% within site	% within group	Mother plants	% within site	% within group	Mother plants	% within site	% within group	Mother plants			
Moraiolo	G1	25	29.4	*	65	76.5	*	50	71.4	*	13	18.6	10	15.9										
Gentile Grande	G1										5	7.1			*									
Leccino	G1	15	17.6		10	11.8		10	14.3		20	28.6	20	31.7	*									
Frantoio	G1	15	17.6		10	11.8		10	14.3		20	28.6	30	47.6										
San Felice	G1	30	35.3	*																				
Villastrada	G1												1	1.6										
Leccio del Corno	G1												2	3.2										
Orbetana	G1										5	7.1												
Pocciolo	G1										5	7.1												
Gentile di Montone	G1										2	2.9												
Pendolino	G2	2	15.4		2	40.0		2	8.0		1	3.4												
Raio	G2							20	80.0	*														
Borgiona	G2										5	17.2	*	2	7.4									

Dolce Agogia	G2	2	15.4					14	48.3	23	85.2	*
Canino	G2			1	20.0		1	4.0				
Bosana	G2	3	23.1					5	17.2			
Maurino	G2	2	15.4	2	40.0		2	8.0				
Nostrale di Rigali	G2	2	15.4					2	6.9			
Coratina	G2									2	7.4	
Itrana	G2							2	6.9		0.0	
Piantone di Mogliano	G2	1	7.7									
Piangente	G2	1	7.7									
Undetermined		2		10		5		1		10		

B	Site 1	Site 2	Site 3	Site 4	Site 5	All Sites
Total surface of olive orchards within site (ha)	41.15	18.87	14.97	11.89	25.18	112.06
Total number of olive trees within site	9,115	3,275	3,319	3,431	5,648	24,788
Number of [G1] trees	7,748	2,784	2,323	2,402	3,558	18,815
Total [G1] %	85	85	70	70	63	75.90
Number of G2 trees	1,185	164	830	995	1,525	4,698
Total [G2] %	13	5	25	29	27	19.00
% of undetermined trees	2	10	5	1	10	5.10

Table 2. Percentage of embryos assigned with relaxed value of 95% confidence to different pollen donors obtained by likelihood based paternity tests in the nine cultivar/site combinations (four replicates for each maternal genotype per site, see Supplementary Table S2). Pollen donors (assigned fathers) and pollen recipients (mother plants) are cultivars of olive trees present in Umbria (see Table 1A). SI Group: Self-Incompatibility group for each cultivar, G1 or G2. In grey: within SI group crosses. +/-: presence/absence of trees from the cultivar in the site. In *italics*, the pollen donors never detected among the analyzed embryos.

Sampling sites		Site 1	Site 2	Site 3	Site 1	Site 4	Site 5	Site 3	Site 4	Site 5				
Pollen recipients		Moraiolo	Moraiolo	Moraiolo	San Felice	Gentile Grande	Leccino	Raio	Borgiona	Dolce Agogia				
Pollen donors	SI Group	G1	G1	G1	G1	G1	G1	G2	G2	G2				
Moraiolo	G1	+	+	+	+	+	+	68.8	+	8.1	+	9.4	+	
Gentile Grande	G1	-	-	-	-	-	+	-	75.0	+	-	-	-	
Leccino	G1	+	+	+	+	+	+	11.9	+	8.8	+	11.9	+	
Frantoio	G1	+	+	+	+	+	+	8.1	+	8.1	+	8.8	+	
<i>San Felice</i>	<i>G1</i>	+	-	-	+	-	-	-	-	-	-	-	-	
Villastrada	G1	-	-	-	-	-	-	-	-	-	10.6	+	+	
Leccio del Corno	G1	-	-	-	-	-	-	-	-	-	10.6	+	+	
<i>Orbetana</i>	<i>G1</i>	-	-	-	-	-	+	-	+	-	-	-	-	
<i>Pocciolo</i>	<i>G1</i>	-	-	-	-	-	+	-	+	-	-	-	-	
<i>Gentile di Montone</i>	<i>G1</i>	-	-	-	-	-	+	-	+	-	-	-	-	
Pendolino	G2	78.1	+	23	+	+	48.8	+	10.0	+	-	+	+	-
Raio	G2	-	-	85.0	+	-	-	-	-	-	+	-	-	-
Borgiona	G2	-	-	-	-	-	10.0	+	70.0	+	-	+	+	+
Dolce Agogia	G2	+	-	-	8.1	+	40	+	10.0	+	-	+	+	+

Canino	G2	-	35	+	8.1	+	-	-	-	+	-	-
Bosana	G2	+	-	-	-	-	+	18.8	+	-	+	-
Maurino	G2	+	+	6.3	+	10.0	+	-	-	+	-	-
Nostrale di Rigali	G2	+	-	-	-	13.1	+	+	-	-	+	-
Coratina	G2	-	-	-	-	-	-	-	8.1	+	-	+
Itrana	G2	-	-	-	-	-	-	8.1	+	+	+	+
Piantone di Mogliano	G2	0.6	+	-	-	8.1	+	-	-	-	-	-
Piangente	G2	6.3	+	-	-	-	+	-	-	-	-	-
<i>Undetermined</i>		15	42	0.6	11.9	11.9	11.9	11.9	11.9	11.3	0.0	48.7
Number of embryos*		160	160	160	160	160	160	160	160	160	160	160

*For each combination (mother-cultivar/site) paternity assignments are obtained from 160 embryos collected on four trees ($40 \times 4 = 160$) of a single cultivar in a single orchard.

Supporting Information

Supplementary Table S1. List of 85 olive genotypes corresponding to 117 known cultivars analyzed by stigma test, including those with different names and identical SSR profile. Data on corresponding incompatibility group ([G1] and [G2]), main country of cultivation, source of plant material (for SSR genotyping and/or stigma test) and consensus allele lengths (base pair) at 10 SSR loci are provided. . The two incompatibility groups are equally frequent among Italian (21 G1 and 27 G2; Chi² test = 0.386, df = 1) and Spanish cultivars (10 G1 and 14 G2; Chi² test = 0.234, df = 1).

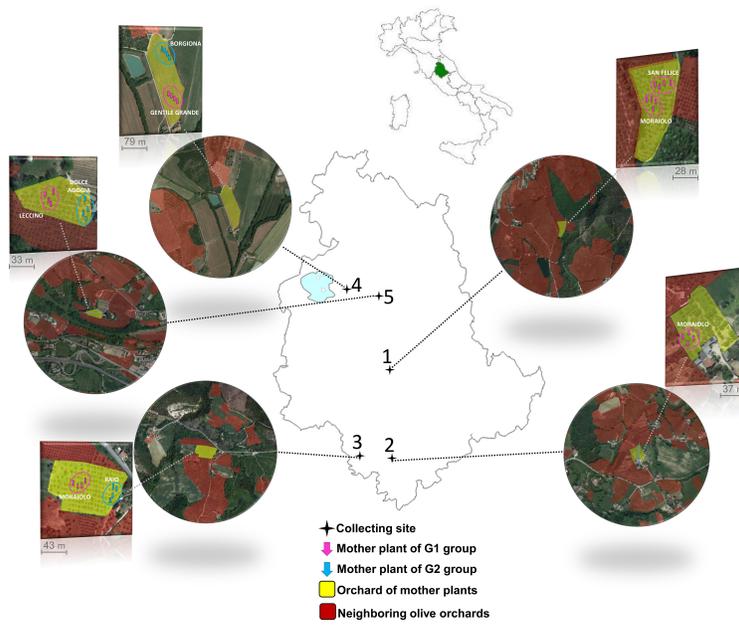
Supplementary Table S2. Percentages and absolute frequencies of embryos assigned to different pollen donors by paternity analysis. The percentage is calculated on the total number of embryos harvested on the four replicates of a given mother plant genotype assigned to given pollen donor. Plant A, B, C and D indicate the four replicates of 40 embryos assigned for paternity for a given genotype. Pairwise population of ϕ FT values calculated starting on the TwoGener genetic distance matrix: 1,440 seedlings and 36 mother plants were considered using 999 permutations. In bold are reported the cultivars belonging to the DSI group 1.

Supplementary Figure S1. Graphical representation of the experimental design.

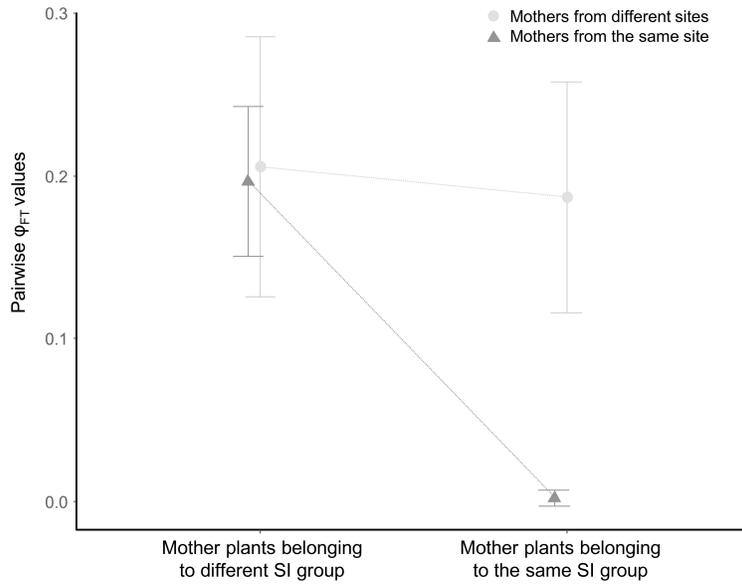
Supplementary Figure S2. Photographs of the different steps of the stigma tests. A) Freshly opened flowers; B) Pistil with pollinated stigma; C, D) Cases of compatible reaction between pollen and stigma (G1 x G2), where pollen grains have germinated and pollen tubes are growing towards the style; E, F) Incompatible reaction (G1 x G1 or G2 x G2): pollen grains produce short skewed tubes (red circles in the picture E) or do not germinate at all.

Supplementary Figure S3. Neighbor Joining tree showing the clustering of all analyzed cultivars. Line colors refer to the group of incompatibility: pink indicates G1 varieties while blue refers to G2 varieties.

Supplementary Figure S4. Distribution of observed LOD scores for the most likely father of each embryo in the five study sites. The distribution is bimodal in each site, with unassigned embryos (light grey) always displaying strongly negative values. This suggests that unassigned embryos were likely sired by unknown pollen donors, either because the actual pollen donor was outside the 500 m radius (thus not included as a potential father in the analysis) or because the pollen donor was missed in our survey.



eva_13175_f1.jpg



eva_13175_f2.jpg