



HAL
open science

Electrical Sensing of sickled red blood cells subpopulations in microfluidic device

Tieying Xu, Maria A Lizarralde-Iragorri, Jean Roman, Emile Martincic,
Valentine Brousse, Olivier Français, Wassim El Nemer, Bruno Le Pioufle

► **To cite this version:**

Tieying Xu, Maria A Lizarralde-Iragorri, Jean Roman, Emile Martincic, Valentine Brousse, et al..
Electrical Sensing of sickled red blood cells subpopulations in microfluidic device. MicroTAS, Oct
2020, on-line, France. hal-03078878

HAL Id: hal-03078878

<https://hal.science/hal-03078878v1>

Submitted on 4 Jan 2021

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

ELECTRICAL SENSING OF SICKLED RED BLOOD CELLS SUBPOPULATIONS IN MICROFLUIDIC DEVICE

Tieying Xu¹, Maria A. Lizarralde-Iragorri², Jean Roman¹, Emile Martincic³, Valentine Brousse²,
Olivier Français⁴, Wassim El Nemer², and Bruno Le Pioufle^{5,*}

¹Université Paris-Saclay, ENS Paris-Saclay, CNRS, Institut d'Alembert, SATIE, Gif sur Yvette, France

²Université de Paris, UMR_S1134, BIGR, Inserm, Paris, France

³C2N, CNRS, Université Paris-Sud, Université Paris-Saclay, Palaiseau, France

⁴ESYCOM, Univ Gustave Eiffel, CNRS, ESIEE Paris, Marne-la-Vallée, France and

⁵Université Paris-Saclay, ENS Paris-Saclay, CNRS, Institut d'Alembert, LUMIN, Gif sur Yvette, France

ABSTRACT

This paper discusses the electrical sensing of red blood cell (RBC) rigidity due to Hemoglobin S (HbS) polymerization in sickle cell disease. For this purpose sickle RBCs subpopulations, differentiated by cell density, were flown through a microfluidic device equipped with integrated microelectrodes. Differences in transit time and electrical blockade were observed and analyzed.

KEYWORDS: Sickle Cell Disease, Electrical Sensing, Microfluidic

INTRODUCTION

Sickle cell disease (SCD) is a monogenic disorder caused by a single mutation in the β -globin gene (c20.A >T) [1]. This mutation induces the abnormal production of Hemoglobin S, that under hypoxia conditions polymerizes, inducing red blood cell (RBC) morphological and physiological changes as increased rigidity, decreased flexibility, and less deformability [2].

While circulating in microcapillaries and under different deoxygenation conditions, different subpopulations of sickle (SS) RBCs can be observed, their morphology might vary between elongated cells (irreversible sickle cells ISC) or biconcave (reticulocytes), and will depend on HbS polymerization and hydration [3].

By using a microfluidic device containing a microrestrictions network and equipped with electrical sensing electrodes, SS RBCs subpopulations can be discriminated due to their ability to deform and squeeze through the restriction zone. The restriction unit allows to reproduce the mechanical stress on RBCs in the blood microcirculation. The width of the restriction progressively decreases down to 5 μ m (See figure 1.A.), while its thickness is 2 μ m. A pair of electrodes is located under this restriction.

THEORY

The microfluidic restriction network is filled by the conductive solution of 12.5mS/cm. A voltage excitation (2V, 10 000Hz) is applied to the pair of gold electrodes (See figure 1.A.) bordering this restriction. The complex impedance of the restriction including (or not) the red blood cell, was monitored using the current response to the voltage excitation. A Gaussian distribution was obtained for the transit time, for which the mean transit time (μ) as well as the dispersion (variance σ^2) was estimated for each peak concerning a SS RBCs subpopulation.

EXPERIMENTAL

After centrifugation and several washes a RBC pellet was obtained from sickle and control samples blood. A percoll fractionation was performed in order to obtain the different RBC subpopulations:

Low Dense (LD) enriched in reticulocytes, Dense (D), and High Dense (HD) enriched in ISCs.

Under the depressurization of 250mbar, RBCs suspension of 2% hematocrit was loaded in the device and flown through the restriction network shaped in the microfluidic device.

RESULTS AND DISCUSSION

For the RBCs collected from one healthy donor, the mean of the transit time was 7.2ms, (See figure 1.B.). In general, SS RBCs showed longer transit time than control RBCs. Differences between the subpopulations were also observed, as rigidity increases a decreasing transit time was observed: LD 9.3ms, D 8.5ms and HD 7.5ms (See

figure 1.C,D,E.). Moreover considering the variance of each SS RBC subpopulation, HD sickled cells are less dispersed.

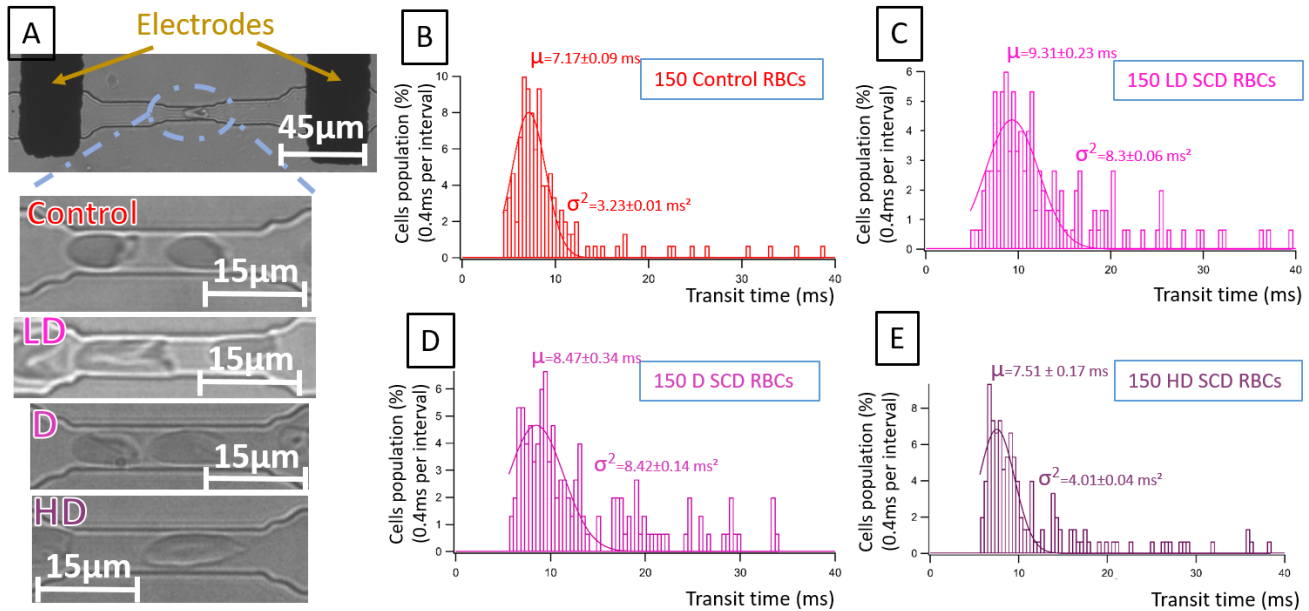


Figure 1: A. The microrestriction engaged by a pair of gold electrodes and different kind of RBCs flowing within the microchannel; B. The Gaussian distribution of the transit time for control RBCs ($n=150$), C. for Low density (LD) sickled cells ($n=150$), D. for Dense (D) sickled cells ($n=150$) and E. for High density (HD) sickled cells ($n=150$).

Due to the elongated shape acquired by the HD dense cells once the HbS polymerizes a small surface contact between HD RBC and the wall of the restriction unit is reflected in the short transit time (See in figure 1.E.). By the contrary, the biconcave shape and hydrated conditions of reticulocytes present in the LD RBC fraction force the LD RBCs to squeeze and deform through the smaller restriction unit increasing its transit time (See figure 1.C.).

CONCLUSION

Through an electrical monitoring of RBC transit time in the microfluidic channel, a distinction between SS RBCs subpopulations was done. This approach, consisting to electrical sense the RBCs in a microfluidic device mimicking the in vivo microcirculation, is very promising for the detection of RBCs abnormalities in different hematopathologies, as well as a promising tool to evaluate treatments related with RBC morphology and physiology.

ACKNOWLEDGEMENTS

The authors acknowledge the Labex LaSIPS (ANR-10-LABX-0040-Lasips), the “Ecole Doctorale EOBÉ” of Université Paris-Saclay, the “Institut d’Alember” and the “CNRS” for project fundings.

REFERENCES

- [1] L. Pauling, H. A. Itano, S. J. Singer, and I. C. Wells, “Sickle Cell Anemia, a Molecular Disease,” *Science* (80-), vol. 110, no. 2865, pp. 543–548, Nov. 1949.
- [2] M. J. Stuart and R. L. Nagel, “Sickle-cell disease,” *Lancet*, vol. 364, no. 9442, pp. 1343–1360, Oct. 2004.
- [3] M. A. Lizarralde Iragorri *et al.*, “A microfluidic approach to study the effect of mechanical stress on erythrocytes in sickle cell disease,” *Lab Chip*, vol. 18, no. 19, pp. 2975–2984, 2018.

CONTACT

* Bruno Le Pioufle : bruno.le-pioufle@ens-paris-saclay.fr