



Contribution of the Low-Density Lipoprotein Receptor Family to Breast Cancer Progression

Océane Champion, Tesnim Al Khalifa, Benoît Langlois, Jessica Thevenard-Devy, Stéphanie Salesse, Katia Savary, Christophe Schneider, Nicolas Etique, Stéphane Dedieu, Jérôme Devy

► To cite this version:

Océane Champion, Tesnim Al Khalifa, Benoît Langlois, Jessica Thevenard-Devy, Stéphanie Salesse, et al.. Contribution of the Low-Density Lipoprotein Receptor Family to Breast Cancer Progression. *Frontiers in Oncology*, 2020, 10, 10.3389/fonc.2020.00882 . hal-03053583

HAL Id: hal-03053583

<https://hal.science/hal-03053583>

Submitted on 11 Dec 2020

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

Contribution of the low-density lipoprotein receptor family to breast cancer progression

Océane Champion^{1,2,£}, Tesnim Al Khalifa^{1,2,£}, Benoit Langlois^{1,2}, Jessica Thevenard-Devy^{1,2},
Stéphanie Salesse^{1,2}, Katia Savary^{1,2}, Christophe Schneider^{1,2}, Nicolas Etique^{1,2}, Stéphane
Dedieu^{1,2*} & Jérôme Devy^{1,2*}

¹ Université de Reims Champagne-Ardenne (URCA), Reims, France.

² CNRS UMR 7369, Matrice Extracellulaire et Dynamique Cellulaire, MEDyC, Reims, France.

£ Co-first authors

* Co-senior authors

Email corresponding author: jerome.devy@univ-reims.fr

In review

Abstract

The low-density lipoprotein receptor (LDLR) family comprises fourteen single transmembrane receptors sharing structural homology and common repeats. These receptors specifically recognize and internalize various extracellular ligands either alone or complexed with membrane spanning co-receptors that are then sorted for lysosomal degradation or cell-surface recovery. As multifunctional endocytic receptors, some LDLR members from the core family were first considered as potential tumor suppressors due to their clearance activity against extracellular matrix degrading enzymes. LDLRs are also involved in pleiotropic functions including growth factors signaling, matricellular proteins and cell matrix adhesion turnover and chemoattraction, thereby affecting both tumor cells and their surrounding microenvironment. Therefore, their roles could appear controversial and dependent upon the malignancy state. In this review, recent advances highlighting the contribution of LDLR members to breast cancer progression are discussed with focus on (1) specific expression patterns of these receptors in primary cancers or distant metastasis and (2) emerging mechanisms and signaling pathways. In addition, potential diagnosis and therapeutic options are proposed.

The low-density lipoprotein receptor family and breast cancer: a state of art.

The low-density lipoprotein receptor (LDLR) gene family encodes single spanning transmembrane receptors usually referred to LDLR-related proteins (LRPs). The fourteen described members are LDLR, VLDLR, LRP1/CD91/A2MR, LRP1B, LRP2/megalin/GP330, LRP3, LRP4/MEGF7, LRP5, LRP6, LRP8/ApoER2, LRP10/LRP9, LRP11/SorLA, LRP12/ST7 and LRAD3 (see **Table 1**). Despite various homology levels, most members are clustered type I receptors sharing structural motifs (e.g., cysteine-rich complement-type repeats), involved in specific recognition of extracellular ligands, EGF-precursor homologous and β -propeller domains critical for protein folding and pH-dependent lysosomal release of ligands. The short intracellular tail encompasses motifs allowing the recruitment of scaffolds driving the endocytic machinery and intracellular signaling. The LDLR founding member was first identified as a frequently mutated etiological factor of familial hypercholesterolemia. LDLR functions were then extended to numerous physiopathological contexts such as vascular integrity, neurobiology, and cancer development due to their peculiar ability to control membrane compartmentalization of receptors and clearance of various classes of extracellular ligands. Some LRPs were thus implicated in the specific recognition of above 50 extracellular factors, among which several growth- or migration-regulatory molecules located in the tumor microenvironment (TME) of various cancers, including mammary cancers.

Breast cancer (BC) is one of the most diagnosed cancers among women worldwide and is the second leading cause of cancer death. Based on their histological features, breast tumors are divided into two subtypes, *in situ* breast carcinoma and invasive breast carcinoma. The first subtype is sub-classified as either ductal (DCIS) or lobular carcinoma *in situ* (LCIS). Invasive carcinomas are further categorized into several histological subtypes, such as infiltrating ductal, invasive lobular, ductal/lobular, mucinous (colloid), tubular, medullary and papillary carcinomas. Classification of BC based on molecular components is more useful for treatment planning and development of targeted therapies. In this classification, BC is mainly divided into hormone-receptor positive (ER^+/PR^+), human epidermal growth factor receptor-2 overexpressing ($HER2^+$) and triple negative (TNBC). Over the past decades, breakthroughs have been made in BC treatment including surgery, radiotherapy, chemotherapy, endocrine therapy, targeted therapy, and immunotherapy. Despite all these therapeutic options, TNBC remains associated with poor outcomes and a historical lack of targeted therapies. Regarding metastases from BC, the most common first site of distant spread is bone (41%), followed by lung (22%), brain (7%), and liver (7%). The remaining 20% of patients have multiple metastatic

sites [1]. In this review, our focus will be on the role played by the members of the LDLR family in BC by examining specifically their implications within the tumor microenvironment. The clinical relevance of targeting these receptors for developing new targeted therapies will be also discussed.

LRPs and breast cancer cells: a close and complex relationship

Obesity and hypercholesterolemia are risk factors for BC that negatively impact therapeutic efficacy [2, 3]. Higher levels of plasmatic cholesterol, LDL (low-density lipoprotein) and triglycerides and low circulating levels of HDL are frequently found in patients with BC [4]. Interestingly, LDL were reported to affect the sensitivity of tumor cells to radiotherapy in inflammatory BC [5] . LDL could affect the adhesion of BC cells involved in cell migration and proliferation and a difference in the quantity and type of lipid synthesis and storage has been demonstrated in basal-like ER⁻ compared to luminal ER⁺ BC cells [6]. Patients with BC usually exhibit elevated serum levels of oxidized LDL, reported to induce structural DNA alterations, a decrease in DNA repair and pro-oncogenic signaling pathways [7].

In mammary tumor tissues, LDLR expression is higher and cholesteryl ester accumulation is associated with an increase of Ki67 expression and poor clinical outcome [8, 9]. BC cells express higher LRP1 and LDLR, allowing a better uptake of LDL-C from the blood [10]. Cholesterol may also generate 27-hydroxycholesterol, an oestrogen mimetic involved in epithelial-to-mesenchymal transition(EMT) in ER⁺ BC cells [11, 12]. In addition, LDLR and acyl-CoA:cholesterol acyltransferase-1 are increased in HER2 positive and triple-negative tumors compared to luminal A tumors [7].

Among LDLR, the giant receptors are represented by LRP1, LRP1B and LRP2, sharing strong structure homologies but showing discrepancies in term of endocytic kinetics and expression pattern [13]. LRP2/Megalin is required for the internalization of Vitamin D and its activation to 1,25-OH Vitamin D. A reduced expression was found in some BC leading to a decrease of its nuclear receptor VDR activation, which plays an important anti-proliferative role [14]. *LRP2* mRNA was also detected at fairly high levels in invasive BC but with extremely high variability [15].

LRP1B, a close homologue of LRP1, is among the top 10 significantly mutated genes in human cancer [16, 17]. LRP1B is mutated in circulating tumor cells from BC and may participate in human mammary gland carcinogenesis [18]. The nuclear localization of its intracellular domain is significantly related to poor prognosis in patients with invasive ductal breast carcinoma and

to a significant decrease of both disease-free and overall survival in patients with luminal A type breast carcinoma [19].

LRP1 was initially identified as a tumor suppressor controlling by endocytic clearance the extracellular matrix degrading enzymes in the microenvironment of various invasive tumors [20]. In BC models, α 2-macroglobulin/LRP1-dependent uptake of pepsin is involved in the control of the invasive potential of luminal and TNBC cells [21]. However, other studies support a more complex view of LRP1 functions in tumor cells. The serine protease inhibitor PN-1/SerpinE2, which is highly expressed in ER⁺ and high grade BC, stimulates lung metastasis of mammary tumor cells through LRP1-dependent secretion of MMP-9 [22]. By contrast, SerpinE2 and LRP1 were identified among the genes induced by ZEB-1, an EMT driver that limits the expression of LRP1-targeting miRNAs, thereby triggering tumor cell autocrine factors which predict poor survival in early stage of BC [23]. LRP1 can exert a dramatic control of tumor cell plasticity and migratory capacities. Its silencing in TNBC cells increased cellular rigidity, decreased cellular protrusions and finally impaired migration [24]. Converging data illustrate the important role of Hsp90 α binding to LRP1 during EMT-related events in BC [25-28]. Hypoxia leads to HIF1- α -dependent secretion of Hsp90 α by TNBC cells. Its specific binding to LRP1 stimulates tumor development and metastatic lung colonization [27]. This interaction and subsequent pro-metastatic signals are reinforced by clusterin in both luminal and TNBC models [28]. Interestingly, within extracellular space, Hsp90 α is absent from the normal microenvironment suggesting promising opportunities for targeted therapy [27]. These studies underline the versatility of LRP1 functions in breast TME and support ongoing research to identify the specific molecular interfaces mobilized by the receptor that could be targeted to control aggressive behavior of tumor cells. A less characterized member of LRPs, SorLA/LRP11, was recently involved in the endocytic trafficking of HER2 [29]. The depletion of SorLA was reported to affect lysosomal function and sensitize HER2-overexpressing cells resistant to targeted therapy. Its targeting could therefore affect compartmentalized pools of oncogenic receptors and restore efficacy of conventional treatments.

LRP5 and LRP6, as co-receptors of the Wnt/ β -catenin pathway, are directly involved in breast tumorigenesis. Wnt ligands such as frizzled homolog 7 and LRP6 are overexpressed in TNBC [30], whereas Wnt antagonists are frequently silenced by methylation in BC [31]. Blockade or silencing of LRP6 in SUM1315 basal BC cells results in a re-expression of epithelial markers and a decreased capacity to self-renew and metastasize [32]. Similarly, LRP6 downregulation in MDA-MB-231 decrease the pool of BC stem cells [33]. These effects are more pronounced

on TNBC cell migration and invasion [34]. The use of benzimidazole compounds on TNBC cells exerts anticancer activity by inhibiting the Wnt/ β -catenin pathway. Prodigiosin and other compounds decrease the phosphorylation of LRP6 (active form), and inhibit the activation of mTORC and Wnt/ β -catenin signaling [35-39]. The disruption of lipid rafts in TNBC cells is associated with a decrease of LRP6 and β -catenin expression, cell proliferation and migration [40]. Besides this Wnt/ β -catenin canonical pathway, LRP5 was also reported to be involved in the uptake of glucose in mammary epithelial cells, through Apolipoprotein E (ApoE) binding. The glucose uptake is essential for regulating the growth rate of these cells [41]. A soluble LRP6 ectodomain can prevent tumor progression, by inhibiting cell migration and metastasis, by limiting the Frz-mediated non-canonical pathway activation in breast tumor cells [42]. The function of LRP8/ApoER2, strongly expressed in ER negative breast tumors was recently described in breast tumor initiating cells, which constitute a clinical challenge of the pathology [43]. Interestingly, its depletion impairs TNBC cell proliferation and promotes apoptosis [44]. LRP8 depletion also leads to Wnt/ β -catenin signaling inhibition, decreases the pool of BC cells, limits their tumorigenic potential in murine xenografts, and finally restores TNBC cell sensitivity to chemotherapy [43]. An overview of the complex and multiple LRPs-mediated signaling pathways is shown in **Fig. 1**.

Functional interplay between LRPs and cells within the tumor microenvironment

The breast TME encompasses multiple cell types including fibroblasts, immune cells, adipocytes, and endothelial cells [45]. In human breast tumors, fibroblasts are the most abundant stromal cells and high levels of LRP1 expression was reported [46]. In fibroblasts, LRP1 binds to CTGF, PDGF and TGF β and interacts with their respective receptors, thereby modulating their mitogenic or contractile capacities [47-51]. Similarly, to LRP1, LRP6 interacts closely with PDGFR β and TGF- β RI in pericytes and is involved in their trans-differentiation into myofibroblasts in response to TGF β or CTGF. Therefore it stimulates the PDGF-BB-dependent proliferation of established myofibroblasts via β -catenin independent mechanisms [52]. Likewise, Wnt7a secreted by aggressive breast tumor cells promotes the activation of stromal fibroblasts through TGF β signaling [53]. In cancer-associated fibroblast from mammary tumors, the stabilization of LRP6 at cell-surface by DKK3 stimulates β -catenin and YAP/TAZ signals, promoting pro-tumorigenic functions such as ECM stiffening [54]. Interestingly, pro-cath-D hypersecreted by cancer cells in the breast TME stimulates fibroblast outgrowth by inhibiting the release of LRP1 β (intracellular domain), which is able to regulate gene transcription [55].

Adipocytes are mainly engaged during BC progression through a metabolic crosstalk with adjacent tumor cells and adopt a modified phenotype called cancer-associated adipocytes [56]. Resulting dysfunctional adipocytes overexpressed fatty acid, cholesterol, triglycerides, hormones, but also adipokines, inflammatory cytokines and proteases that are linked to cancer progression [56]. LRP-1 is highly expressed in preadipocytes and is involved in adipocyte differentiation, especially through its regulation of peroxisome proliferator-activated receptor γ [57]. LRP1 has also been demonstrated to be upregulated in obese mouse adipocytes and obese human adipose tissues [57] and to regulate insulin receptor and GLUT4 trafficking and activation [58, 59]. Through modulation of Wnt5a signaling, LRP1 controls cholesterol storage and fatty acid synthesis during adipocyte differentiation [60]. An attenuated endocytosis of apoA5 by adipocytes was demonstrated in both adipose tissue from obese patients and insulin resistant adipocytes. The mechanism underlying this phenomenon might be related to a reduced endocytic activity of LRP1 and/or an attenuated insulin-dependent movement of LRP1 from intracellular structures to the cell surface [61]. These mechanisms may lead to excessive augmentation of triglycerides storage and abnormal metabolism of adipocytes, hence promoting the development of obesity and obesity-associated disorders such as BC.

LRP1 is also abundantly expressed by various immune cells and its function in HSP-mediated antigen presentation and subsequent innate immune response is well described in macrophages and dendritic cells [62]. LRP1 also inhibits macrophage-driven inflammation by decreasing cell-surface abundance of the TNF receptor-1 and $\text{I}\kappa\text{B}$ kinase/NF- κB intracellular activation [63]. By contrast, production of sLRP1 (shed or soluble LRP1) by macrophages induces pro-inflammatory factors synthesis such as IL-10, TNF- α , and MCP-1 [64]. Macrophage inflammatory protein-1a/CCL3, known to amplify inflammation, is overproduced in the absence of LRP1 in myeloid cells, leading to enhanced CCR5-expressing monocyte recruitment to tumors and cancer angiogenesis [65].

Recent studies have demonstrated the crucial angio-modulatory actions of LRP family members in various solid tumors, including BCs [23, 38, 66-69]. LDL and VLDL (very low density lipoprotein) are involved in the secretion of diffusible angiogenic factors by BC cells, such as amphiregulin [69]. Moreover, the binding of Wnt3a to LRP6 stimulates VEGF production by TNBC cells [38]. The stoichiometry of Wnt ligands and their secreted regulators such as Dickkopf (DKK) seems instrumental to fine-tune LRP5/6 functions in the TME. DKK1 was indeed described as anti-angiogenic, whereas DKK2 binding to LRP6 triggers potent induction of endothelial cell sprouting [70]. LRP1 is widely expressed in various endothelial cells and its

specific binding to tPA alone or complexed with uPA/PAI-1 induces vascular permeability in the blood-brain-barrier (BBB) or in lung microvasculature, two major sites of BC cells metastatic homing [71, 72]. LRP1 controls multiple aspects of endothelial cell metabolism [73] and participates to the control of intercellular junctionality, morphogenesis and proliferation [71, 74, 75]. Interestingly and as stated above, during epithelial-to-mesenchymal transition in breast tumors, LRP1 expression was derepressed through ZEB-1-dependent inhibition of LRP1-targeting miRNAs, thereby contributing to vascular mimicry of breast tumor cells [23]. The induction of this endocytic receptor could thus reinforce endothelial interface of breast tumor cells and facilitate their metastatic dissemination.

Clinical significance of LRPs as biomarker in the context of breast tumors

A large scale study conducted on solid tumors indicates that mostly LDLR mRNA are overexpressed in breast invasive carcinomas with *LRP2* mRNA being the most expressed, but no correlation with patient survival was observed [15]. Only, a few studies are focused on LRP1 expression in BC patient samples. LRP1 was first immunohistochemically studied in fresh frozen tissue from primary invasive breast carcinomas, ductal *in situ* carcinomas and axillary lymph-node metastases in 1996 [46]. LRP1 staining appeared intense in all stromal fibroblasts both outside and within the tumor tissue and scattered in macrophages and mast cells. Interestingly, epithelial cells, endothelial cells and lymphocytes appeared negative for LRP1. A more recent immunohistochemistry study of LRP1 expression, performed on infiltrating ductal breast carcinomas, brought different results as cytoplasmic LRP1 overexpression was identified in tumor cells in addition to non-neoplastic stromal cells, whereas normal ductal cells were always negative [76]. Concerning *LRP1* polymorphism, although C766T mutation was firstly reported as significantly higher in patients with BC [77], the increased risk of BC development associated to LRP1 polymorphism is not definitely established. Of note, neoadjuvant chemotherapy in BC did not impair LRP1 expression [9].

A better understanding of LRPs functionalities may lead to efficient therapeutic strategies

Therapeutic approaches involving LRPs developed in oncology, particularly in BC, aim to address the endocytic properties of these receptors as vectorization tools. One of the remaining therapeutic concerns for BC patients is related to metastases. Brain metastases occur in about 15 to 30% of women with stage IV BC. The targeting of the BBB, formed by endothelial cells, astrocytes and pericytes embedded in the capillary basement membrane, remains critical for treating brain metastases. As LRP1 transports ligands such as β -Amyloid or tPA across the

BBB and is expressed at high levels in this tissue [78], it appears as a promising candidate for targeted therapy against metastatic BC cells. In this context, the main therapeutic approaches use the Angiopep-2, an LRP1 binding peptide first identified by Demeule and colleague [79]. Combined with three paclitaxel residues, this molecule (namely GRN1005, formerly known as ANG1005) binds to LRP1, crosses the BBB and allows a better drug delivery in the brain compartment [68, 79]. Phase I/II clinical trials with ANG1005/GRN1005 show that treatment is safe and brings clinical benefit for both peripheral metastatic BC and brain metastasis, even if the tumor had previously developed resistance to conventional taxanes. Interestingly, an open-label Phase III study will start in 2020 to investigate whether ANG1005 can prolong patient survival in HER2-negative BC patients with newly diagnosed leptomeningeal disease and previously treated brain metastases (NCT03613181). Angiopep-2 can also be useful to target BC cells overexpressing LRP1. For instance, Angiopep-2 was used to decorate nanoparticles combined with doxycycline (Angio-DOX-DGL-GNP) in TNBC to facilitate the drug penetration and accumulation in BC cells [80].

The endocytic properties of LRP2 have also been used to improve the effectiveness of anticancer drugs in resistant BC cells [81]. In this context, lipid-polyethylenimine hybrid nanocarriers decorated with apolipoprotein E (Ap-LPN) were developed for improving siRNA delivery against clusterin in MCF7 BC cells, leading to increased cell chemosensitization toward paclitaxel.

Another strategy of tumor targeting was used with the NT4 peptide, a tetrabranch peptide from the human neurotensin, capable of binding LRP1 and LRP6 by mimicking ApoE and midkine heparin binding site [82]. Depau and collaborators showed that methotrexate conjugated with NT4 can overcome drug resistance in methotrexate-resistant human BC cells [83]. NT4 conjugated with other drugs (NT4-paclitaxel, NT4-5FdU) were tested in various animal models of human cancer, including an orthotopic mouse model of human BC, leading to improved drug activity as compared to unconjugated counterpart [84-86].

More recently, some LRPs have been identified as direct molecular targets for BC. LRP6 is probably the most promising target in the TNBC with its overexpression leading to Wnt signaling pathway activation together with tumorigenesis promotion [87]. Several drugs such as salinomycin, prodigiosin and niclosamide indeed induce LRP6 phosphorylation and degradation leading to decreased tumor growth [36, 37, 88]. Ren and collaborators have

suggested that soluble LRP6 ectodomain could also be used as an innovative anti-metastatic drug [42].

Conclusion and subjective points of view

Receptors from the LDLR family are increasingly emerging as key relevant biomarkers in oncology and potential therapeutic targets. Their multiple implications within the TME, variable expression related to tumor stages, together with molecular versatility, constitute the main challenges to better understand their functionalities. In Breast cancer, scientific evidence is fragmented, sometimes contradictory, and only a few clinical data are available. Potential prognostic value of these receptors is still unclear, thus preventing from demonstrating clinical benefits. Additional studies will be necessary to establish a link between LRPs and some events promoting obesity or metabolic diseases, particularly to improve the treatment of BC in post-menopausal patients. LRP1 is likely to be the most promising receptor because it constitutes an efficient drug-carrier within tumor cells. Very promising trials are ongoing in HER2-negative BC patients with metastasis. In addition, LRP1 could also be considered as an attractive therapeutic target in TNBC. However, its high molecular weight, intricate regulation, and sub-cellular localization together with its ability to bind multiple extracellular ligands within the same clusters, make current research extremely complex and can lead to contradictory conclusions. The use of more advanced *in vitro* multi-cellular and 3D tumor-based systems (tumoroids), with patient-derived cells will be key to deeper understand the functionality of this receptor. In the coming years and in order to consider LRP1 as an innovative vectorization tool, the approach should be focused on the endocytic properties of overexpressed LRP1 rather than on the modulation (eg, inhibition or reduction) of LRP1 expression.

Author Contributions

All authors listed have made a substantial, direct and intellectual contribution to the work, and approved it for publication.

Conflict of Interest

The authors declare that this work was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Abbreviations

BBB: blood-brain-barrier, BC: breast cancer, DCIS: Ductal carcinoma *in situ*, ECM: extracellular matrix, EMT: epithelial-to-mesenchymal transition, ER: estrogen receptor, HDL: High density lipoprotein, HER2: human epidermal growth factor receptor-2, LCIS : Lobular carcinoma *in situ*, LDL: low-density lipoprotein, LDLR: low-density lipoprotein receptor, , LRP: LDL receptor-related protein, PR: progesterone receptor, TME: tumor microenvironment, TNBC: triple negative breast cancer, VDR: vitamin D receptor, VLDL: very low density lipoprotein.

Acknowledgements

We acknowledge Arlette Thomachot and Dr Laetitia Devy for editorial assistance.

Legend

Figure 1. LRP-mediated signaling pathways and trafficking in breast tumor cells.

The most important members of the LDLR family exhibiting effects on breast cancer cells are represented with their associated extracellular ligands. Outside-in and inside-out pathways are represented by black and yellow arrows or lines, respectively. The yellow strikethrough line indicates an inhibition. A2M: alpha-2 macroglobulin, CT: cholesterol, ER: estrogen receptor, LXR: liver X receptor, VitD: vitamin D.

Table 1. The fourteen members of the low-density lipoprotein receptor family and their involvement in breast cancer.

The family core members are underlined in green, the structurally distant members in blue and the most distant members in orange. ND: not determined.

In review

References

1. Berman, A.T., et al., *Incidence and patterns of distant metastases for patients with early-stage breast cancer after breast conservation treatment*. Clin Breast Cancer, 2013. **13**(2): p. 88-94.
2. Liu, J., et al., *Cholesterol-induced mammary tumorigenesis is enhanced by adiponectin deficiency: role of LDL receptor upregulation*. Oncotarget, 2013. **4**(10): p. 1804-18.
3. Lee, K., et al., *The Impact of Obesity on Breast Cancer Diagnosis and Treatment*. Curr Oncol Rep, 2019. **21**(5): p. 41.
4. Li, X., et al., *Status of lipid and lipoprotein in female breast cancer patients at initial diagnosis and during chemotherapy*. Lipids Health Dis, 2018. **17**(1): p. 91.
5. Wolfe, A.R., et al., *High-density and very-low-density lipoprotein have opposing roles in regulating tumor-initiating cells and sensitivity to radiation in inflammatory breast cancer*. International Journal of Radiation Oncology* Biology* Physics, 2015. **91**(5): p. 1072-1080.
6. Guan, X., et al., *Emerging roles of low-density lipoprotein in the development and treatment of breast cancer*. Lipids Health Dis, 2019. **18**(1): p. 137.
7. Cedo, L., et al., *HDL and LDL: Potential New Players in Breast Cancer Development*. J Clin Med, 2019. **8**(6).
8. de Gonzalo-Calvo, D., et al., *Intratumor cholesteryl ester accumulation is associated with human breast cancer proliferation and aggressive potential: a molecular and clinicopathological study*. BMC Cancer, 2015. **15**: p. 460.
9. Pires, L.A., et al., *Effect of neoadjuvant chemotherapy on low-density lipoprotein (LDL) receptor and LDL receptor-related protein 1 (LRP-1) receptor in locally advanced breast cancer*. Braz J Med Biol Res, 2012. **45**(6): p. 557-64.
10. dos Santos, C.R., et al., *LDL-cholesterol signaling induces breast cancer proliferation and invasion*. Lipids Health Dis, 2014. **13**: p. 16.
11. Torres, C.G., et al., *27-hydroxycholesterol induces the transition of MCF7 cells into a mesenchymal phenotype*. Oncol Rep, 2011. **26**(2): p. 389-97.
12. Cruz, P., et al., *Proliferation of human mammary cancer cells exposed to 27-hydroxycholesterol*. Exp Ther Med, 2010. **1**(3): p. 531-536.
13. Liu, C.X., et al., *The putative tumor suppressor LRP1B, a novel member of the low density lipoprotein (LDL) receptor family, exhibits both overlapping and distinct properties with the LDL receptor-related protein*. J Biol Chem, 2001. **276**(31): p. 28889-96.
14. Cabezas, F., P. Farfan, and M.P. Marzolo, *Participation of the SMAD2/3 signalling pathway in the down regulation of megalin/LRP2 by transforming growth factor beta (TGF-ss1)*. PLoS One, 2019. **14**(5): p. e0213127.
15. Gonias, S.L., et al., *Expression of LDL receptor-related proteins (LRPs) in common solid malignancies correlates with patient survival*. PLoS One, 2017. **12**(10): p. e0186649.
16. Knisely, J.M., et al., *Slow endocytosis of the LDL receptor-related protein 1B: implications for a novel cytoplasmic tail conformation*. Exp Cell Res, 2007. **313**(15): p. 3298-307.
17. Chen, H., et al., *Association of LRP1B Mutation With Tumor Mutation Burden and Outcomes in Melanoma and Non-small Cell Lung Cancer Patients Treated With Immune Check-Point Blockades*. Front Immunol, 2019. **10**: p. 1113.
18. Wang, Y., et al., *Single nucleotide variant profiles of viable single circulating tumour cells reveal CTC behaviours in breast cancer*. Oncol Rep, 2018. **39**(5): p. 2147-2159.
19. Asano, Y., et al., *Nuclear localization of LDL receptor-related protein 1B in mammary gland carcinogenesis*. J Mol Med (Berl), 2019. **97**(2): p. 257-268.
20. Dedieu, S. and B. Langlois, *LRP-1: a new modulator of cytoskeleton dynamics and adhesive complex turnover in cancer cells*. Cell Adh Migr, 2008. **2**(2): p. 77-80.
21. Kang, H.S., et al., *LRP1-dependent pepsin clearance induced by 2'-hydroxycinnamaldehyde attenuates breast cancer cell invasion*. Int J Biochem Cell Biol, 2014. **53**: p. 15-23.

22. Fayard, B., et al., *The serine protease inhibitor protease nexin-1 controls mammary cancer metastasis through LRP-1-mediated MMP-9 expression*. *Cancer Res*, 2009. **69**(14): p. 5690-8.
23. Langer, E.M., et al., *ZEB1-repressed microRNAs inhibit autocrine signaling that promotes vascular mimicry of breast cancer cells*. *Oncogene*, 2018. **37**(8): p. 1005-1019.
24. Berquand, A., et al., *A gentle approach to investigate the influence of LRP-1 silencing on the migratory behavior of breast cancer cells by atomic force microscopy and dynamic cell studies*. *Nanomedicine*, 2019. **18**: p. 359-370.
25. Boel, N.M., M.C. Hunter, and A.L. Edkins, *LRP1 is required for novobiocin-mediated fibronectin turnover*. *Sci Rep*, 2018. **8**(1): p. 11438.
26. Dong, H., et al., *Breast Cancer MDA-MB-231 Cells Use Secreted Heat Shock Protein-90alpha (Hsp90alpha) to Survive a Hostile Hypoxic Environment*. *Sci Rep*, 2016. **6**: p. 20605.
27. Sahu, D., et al., *A potentially common peptide target in secreted heat shock protein-90alpha for hypoxia-inducible factor-1alpha-positive tumors*. *Mol Biol Cell*, 2012. **23**(4): p. 602-13.
28. Tian, Y., et al., *Extracellular Hsp90alpha and clusterin synergistically promote breast cancer epithelial-to-mesenchymal transition and metastasis via LRP1*. *J Cell Sci*, 2019. **132**(15).
29. Pietila, M., et al., *SORLA regulates endosomal trafficking and oncogenic fitness of HER2*. *Nat Commun*, 2019. **10**(1): p. 2340.
30. Yang, L., et al., *FZD7 has a critical role in cell proliferation in triple negative breast cancer*. *Oncogene*, 2011. **30**(43): p. 4437-46.
31. Suzuki, H., et al., *Frequent epigenetic inactivation of Wnt antagonist genes in breast cancer*. *Br J Cancer*, 2008. **98**(6): p. 1147-56.
32. DiMeo, T.A., et al., *A novel lung metastasis signature links Wnt signaling with cancer cell self-renewal and epithelial-mesenchymal transition in basal-like breast cancer*. *Cancer Res*, 2009. **69**(13): p. 5364-73.
33. Ibrahim, S.A., et al., *Syndecan-1 (CD138) modulates triple-negative breast cancer stem cell properties via regulation of LRP-6 and IL-6-mediated STAT3 signaling*. *PLoS One*, 2013. **8**(12): p. e85737.
34. Ma, J., et al., *Role of Wnt Co-Receptor LRP6 in Triple Negative Breast Cancer Cell Migration and Invasion*. *J Cell Biochem*, 2017. **118**(9): p. 2968-2976.
35. Gangrade, A., et al., *Preferential Inhibition of Wnt/beta-Catenin Signaling by Novel Benzimidazole Compounds in Triple-Negative Breast Cancer*. *Int J Mol Sci*, 2018. **19**(5).
36. Wang, Z., et al., *Prodigosin inhibits Wnt/beta-catenin signaling and exerts anticancer activity in breast cancer cells*. *Proc Natl Acad Sci U S A*, 2016. **113**(46): p. 13150-13155.
37. Lu, W. and Y. Li, *Salinomycin suppresses LRP6 expression and inhibits both Wnt/beta-catenin and mTORC1 signaling in breast and prostate cancer cells*. *J Cell Biochem*, 2014. **115**(10): p. 1799-807.
38. Zhang, J., et al., *Kallistatin antagonizes Wnt/beta-catenin signaling and cancer cell motility via binding to low-density lipoprotein receptor-related protein 6*. *Mol Cell Biochem*, 2013. **379**(1-2): p. 295-301.
39. Lu, W., C. Lin, and Y. Li, *Rottlerin induces Wnt co-receptor LRP6 degradation and suppresses both Wnt/beta-catenin and mTORC1 signaling in prostate and breast cancer cells*. *Cell Signal*, 2014. **26**(6): p. 1303-9.
40. Badana, A.K., et al., *Lipid rafts disruption induces apoptosis by attenuating expression of LRP6 and survivin in triple negative breast cancer*. *Biomed Pharmacother*, 2018. **97**: p. 359-368.
41. Chin, E.N., et al., *Lrp5 Has a Wnt-Independent Role in Glucose Uptake and Growth for Mammary Epithelial Cells*. *Mol Cell Biol*, 2015. **36**(6): p. 871-85.
42. Ren, D.N., et al., *LRP5/6 directly bind to Frizzled and prevent Frizzled-regulated tumour metastasis*. *Nat Commun*, 2015. **6**: p. 6906.
43. Lin, C.C., et al., *Targeting LRP8 inhibits breast cancer stem cells in triple-negative breast cancer*. *Cancer Lett*, 2018. **438**: p. 165-173.
44. Maire, V., et al., *LRP8 is overexpressed in estrogen-negative breast cancers and a potential target for these tumors*. *Cancer Med*, 2019. **8**(1): p. 325-336.

45. Soysal, S.D., A. Tzankov, and S.E. Muenst, *Role of the Tumor Microenvironment in Breast Cancer*. Pathobiology, 2015. **82**(3-4): p. 142-52.
46. Christensen, L., et al., *Immunohistochemical localization of urokinase-type plasminogen activator, type-1 plasminogen-activator inhibitor, urokinase receptor and alpha(2)-macroglobulin receptor in human breast carcinomas*. Int J Cancer, 1996. **66**(4): p. 441-52.
47. Newton, C.S., et al., *Platelet-derived growth factor receptor-beta (PDGFR-beta) activation promotes its association with the low density lipoprotein receptor-related protein (LRP). Evidence for co-receptor function*. J Biol Chem, 2005. **280**(30): p. 27872-8.
48. Yang, M., et al., *Tyrosine phosphorylation of the LDL receptor-related protein (LRP) and activation of the ERK pathway are required for connective tissue growth factor to potentiate myofibroblast differentiation*. FASEB J, 2004. **18**(15): p. 1920-1.
49. Muratoglu, S.C., et al., *Low density lipoprotein receptor-related protein 1 (LRP1) forms a signaling complex with platelet-derived growth factor receptor-beta in endosomes and regulates activation of the MAPK pathway*. J Biol Chem, 2010. **285**(19): p. 14308-17.
50. Schnieder, J., et al., *Loss of LRP1 promotes acquisition of contractile-myofibroblast phenotype and release of active TGF-beta1 from ECM stores*. Matrix Biol, 2019.
51. Hu, K., et al., *Tissue-type plasminogen activator promotes murine myofibroblast activation through LDL receptor-related protein 1-mediated integrin signaling*. J Clin Invest, 2007. **117**(12): p. 3821-32.
52. Ren, S., et al., *LRP-6 is a coreceptor for multiple fibrogenic signaling pathways in pericytes and myofibroblasts that are inhibited by DKK-1*. Proc Natl Acad Sci U S A, 2013. **110**(4): p. 1440-5.
53. Avgustinova, A., et al., *Tumour cell-derived Wnt7a recruits and activates fibroblasts to promote tumour aggressiveness*. Nat Commun, 2016. **7**: p. 10305.
54. Ferrari, N., et al., *Dickkopf-3 links HSF1 and YAP/TAZ signalling to control aggressive behaviours in cancer-associated fibroblasts*. Nat Commun, 2019. **10**(1): p. 130.
55. Derocq, D., et al., *Cathepsin D is partly endocytosed by the LRP1 receptor and inhibits LRP1-regulated intramembrane proteolysis*. Oncogene, 2012. **31**(26): p. 3202-12.
56. Choi, J., Y.J. Cha, and J.S. Koo, *Adipocyte biology in breast cancer: From silent bystander to active facilitator*. Prog Lipid Res, 2018. **69**: p. 11-20.
57. Masson, O., et al., *LRP1 receptor controls adipogenesis and is up-regulated in human and mouse obese adipose tissue*. PLoS One, 2009. **4**(10): p. e7422.
58. Jedrychowski, M.P., et al., *Proteomic analysis of GLUT4 storage vesicles reveals LRP1 to be an important vesicle component and target of insulin signaling*. J Biol Chem, 2010. **285**(1): p. 104-14.
59. Actis Dato, V. and G.A. Chiabrando, *The Role of Low-Density Lipoprotein Receptor-Related Protein 1 in Lipid Metabolism, Glucose Homeostasis and Inflammation*. Int J Mol Sci, 2018. **19**(6).
60. El Asmar, A. and Z. El Rassi, *Hepatoblastoma in childhood, long term survival achieved: 2 case reports and literature review*. Int J Surg Case Rep, 2016. **21**: p. 55-8.
61. Zheng, X.Y., et al., *Apolipoprotein A5 regulates intracellular triglyceride metabolism in adipocytes*. Mol Med Rep, 2017. **16**(5): p. 6771-6779.
62. Pawaria, S. and R.J. Binder, *CD91-dependent programming of T-helper cell responses following heat shock protein immunization*. Nat Commun, 2011. **2**: p. 521.
63. Mantuano, E., et al., *LDL receptor-related protein-1 regulates NFkappaB and microRNA-155 in macrophages to control the inflammatory response*. Proc Natl Acad Sci U S A, 2016. **113**(5): p. 1369-74.
64. Gorovoy, M., et al., *Inflammatory mediators promote production of shed LRP1/CD91, which regulates cell signaling and cytokine expression by macrophages*. J Leukoc Biol, 2010. **88**(4): p. 769-78.
65. Staudt, N.D., et al., *Myeloid cell receptor LRP1/CD91 regulates monocyte recruitment and angiogenesis in tumors*. Cancer Res, 2013. **73**(13): p. 3902-12.

66. Njah, K., et al., *A Role of Agrin in Maintaining the Stability of Vascular Endothelial Growth Factor Receptor-2 during Tumor Angiogenesis*. Cell Rep, 2019. **28**(4): p. 949-965 e7.
67. Pencheva, N., et al., *Convergent multi-miRNA targeting of ApoE drives LRP1/LRP8-dependent melanoma metastasis and angiogenesis*. Cell, 2012. **151**(5): p. 1068-82.
68. Li, F. and S.C. Tang, *Targeting metastatic breast cancer with ANG1005, a novel peptide-paclitaxel conjugate that crosses the blood-brain-barrier (BBB)*. Genes Dis, 2017. **4**(1): p. 1-3.
69. Lu, C.W., et al., *VLDL and LDL, but not HDL, promote breast cancer cell proliferation, metastasis and angiogenesis*. Cancer Lett, 2017. **388**: p. 130-138.
70. Min, J.K., et al., *The WNT antagonist Dickkopf2 promotes angiogenesis in rodent and human endothelial cells*. J Clin Invest, 2011. **121**(5): p. 1882-93.
71. Makarova, A.M., et al., *Urokinase-type plasminogen activator (uPA) induces pulmonary microvascular endothelial permeability through low density lipoprotein receptor-related protein (LRP)-dependent activation of endothelial nitric-oxide synthase*. J Biol Chem, 2011. **286**(26): p. 23044-53.
72. Wang, X., et al., *Lipoprotein receptor-mediated induction of matrix metalloproteinase by tissue plasminogen activator*. Nat Med, 2003. **9**(10): p. 1313-7.
73. Mao, H., L. Xie, and X. Pi, *Low-Density Lipoprotein Receptor-Related Protein-1 Signaling in Angiogenesis*. Front Cardiovasc Med, 2017. **4**: p. 34.
74. Mao, H., et al., *LRP1 Regulates Retinal Angiogenesis by Inhibiting PARP-1 Activity and Endothelial Cell Proliferation*. Arterioscler Thromb Vasc Biol, 2016. **36**(2): p. 350-60.
75. Vezina, A., et al., *Cerebrovascular Angiogenic Reprogramming upon LRP1 Repression: Impact on Sphingosine-1-Phosphate-Mediated Signaling in Brain Endothelial Cell Chemotactism*. Mol Neurobiol, 2018. **55**(4): p. 3551-3563.
76. Catasus, L., et al., *Low-density lipoprotein receptor-related protein 1 is associated with proliferation and invasiveness in Her-2/neu and triple-negative breast carcinomas*. Hum Pathol, 2011. **42**(11): p. 1581-8.
77. Benes, P., et al., *C766T low-density lipoprotein receptor-related protein 1 (LRP1) gene polymorphism and susceptibility to breast cancer*. Breast Cancer Res, 2003. **5**(3): p. R77-81.
78. Zhao, Y., et al., *The role of the low-density lipoprotein receptor-related protein 1 (LRP-1) in regulating blood-brain barrier integrity*. Rev Neurosci, 2016. **27**(6): p. 623-34.
79. Thomas, F.C., et al., *Uptake of ANG1005, a novel paclitaxel derivative, through the blood-brain barrier into brain and experimental brain metastases of breast cancer*. Pharm Res, 2009. **26**(11): p. 2486-94.
80. Hu, G., et al., *Peptide mediated active targeting and intelligent particle size reduction-mediated enhanced penetrating of fabricated nanoparticles for triple-negative breast cancer treatment*. Oncotarget, 2015. **6**(38): p. 41258-74.
81. Xue, H.Y. and H.L. Wong, *Targeting megalin to enhance delivery of anti-clusterin small-interfering RNA nanomedicine to chemo-treated breast cancer*. Eur J Pharm Biopharm, 2012. **81**(1): p. 24-32.
82. Falciani, C., et al., *Cancer selectivity of tetrabrached neurotensin peptides is generated by simultaneous binding to sulfated glycosaminoglycans and protein receptors*. J Med Chem, 2013. **56**(12): p. 5009-18.
83. Depau, L., et al., *Coupling to a cancer-selective heparan-sulfate-targeted branched peptide can by-pass breast cancer cell resistance to methotrexate*. Oncotarget, 2017. **8**(44): p. 76141-76152.
84. Falciani, C., et al., *Synthesis and biological activity of stable branched neurotensin peptides for tumor targeting*. Mol Cancer Ther, 2007. **6**(9): p. 2441-8.
85. Falciani, C., et al., *Design and in vitro evaluation of branched peptide conjugates: turning nonspecific cytotoxic drugs into tumor-selective agents*. ChemMedChem, 2010. **5**(4): p. 567-74.
86. Brunetti, J., et al., *Tumor-selective peptide-carrier delivery of Paclitaxel increases in vivo activity of the drug*. Sci Rep, 2015. **5**: p. 17736.

87. Roslan, Z., et al., *The Roles of Low-Density Lipoprotein Receptor-Related Proteins 5, 6, and 8 in Cancer: A Review*. J Oncol, 2019. **2019**: p. 4536302.
88. Lu, W., et al., *Niclosamide suppresses cancer cell growth by inducing Wnt co-receptor LRP6 degradation and inhibiting the Wnt/beta-catenin pathway*. PLoS One, 2011. **6**(12): p. e29290.
89. Gallagher, E.J., et al., *Elevated tumor LDLR expression accelerates LDL cholesterol-mediated breast cancer growth in mouse models of hyperlipidemia*. Oncogene, 2017. **36**(46): p. 6462-6471.
90. He, L., et al., *Up-regulated expression of type II very low density lipoprotein receptor correlates with cancer metastasis and has a potential link to β -catenin in different cancers*. BMC cancer, 2010. **10**(1): p. 601.
91. He, L., Y. Lu, and J. Guo, *Type II VLDLR promotes cell migration by up-regulation of VEGF, MMP2 and MMP7 in breast cancer cells*. The Chinese-German Journal of Clinical Oncology, 2013. **12**(8): p. 374-378.
92. Arun, B., C. Shiang, and Y. Qi, *High throughput siRNA screen identifies LRP8 as druggable metabolic regulator in triple-negative breast cancer*. SM Journal of Oncology and Hematology, 2016. **1**(1): p. 1-8.
93. Chlon, T.M., et al., *Retinoids modulate expression of the endocytic partners megalin, cubilin, and disabled-2 and uptake of vitamin D-binding protein in human mammary cells*. The Journal of nutrition, 2008. **138**(7): p. 1323-1328.
94. Lindvall, C., et al., *The Wnt co-receptor Lrp6 is required for normal mouse mammary gland development*. PloS one, 2009. **4**(6).
95. Maubant, S., et al., *LRP5 regulates the expression of STK40, a new potential target in triple-negative breast cancers*. Oncotarget, 2018. **9**(32): p. 22586.

Figure 1.TIF

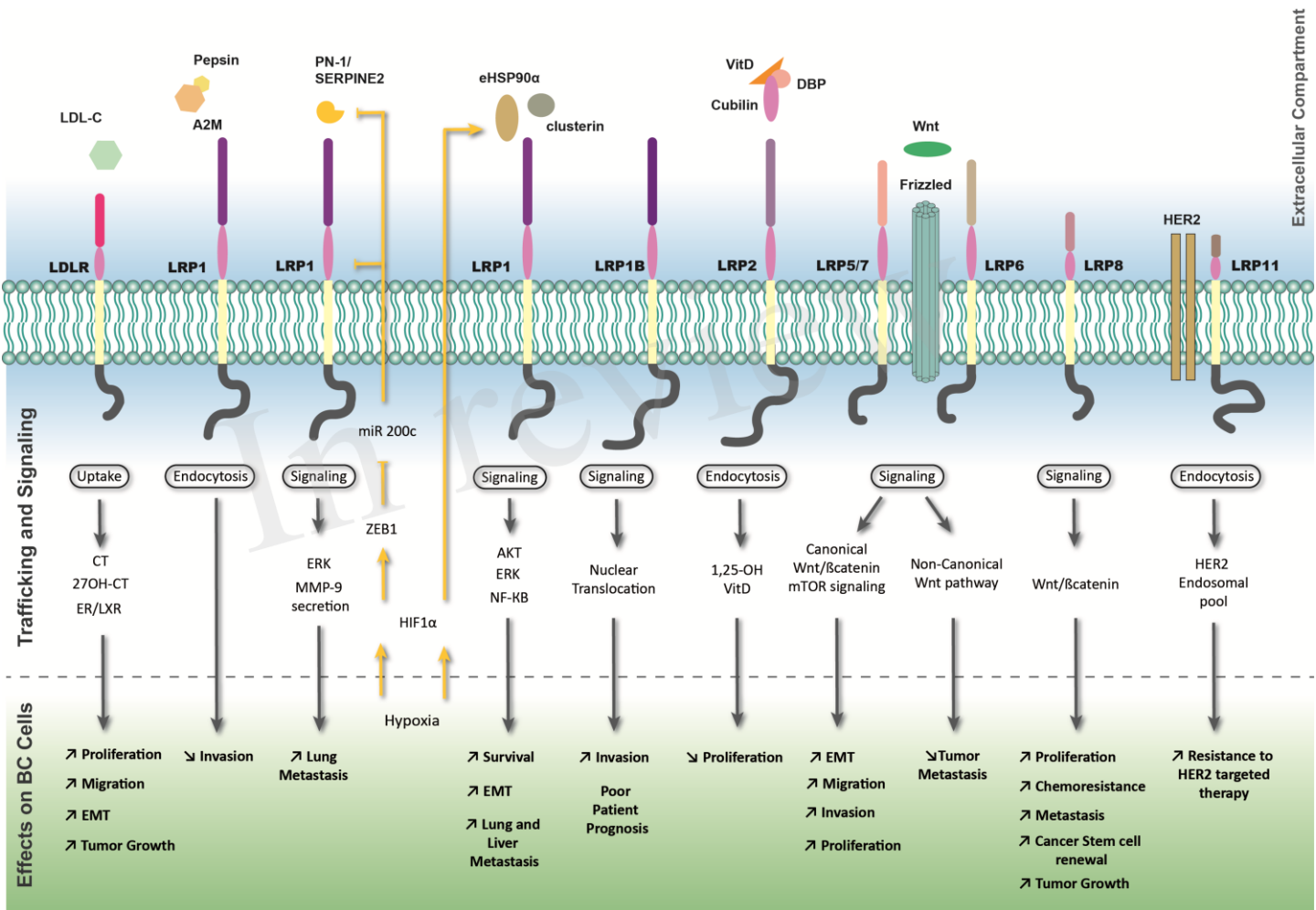


TABLE 1 | The 14 members of the low-density lipoprotein receptor family and their involvement in breast cancer.

LRP isotype	Alternative name	MW (kDa)	Tissue distribution	How involved in breast cancer
LDLR		120-160	Ubiquitous	• Overexpressed in HER2⁺ and TNBC (MDA-MB-231)(1) • Overexpression accelerates LDL cholesterol-mediated BC growth in mouse models of hyperlipidemia (2)
VLDL-R (type II)		96	Abundant in heart, skeletal muscle, ovary and kidney	• Up-regulated expression correlates with BC metastasis (3) • Promotes BC cell migration by up-regulating VEGF, MMP2 and MMP7 (4) • Survival factor in TNBC (5, 6)
LRAD3	Ldlrad3	50	Neurons	None
LRP1	α 2MR APOER CD91	600	Ubiquitous (liver, brain, adipose tissue, fibroblasts and tissue stroma)	• Overexpressed in aggressive HER2⁺ and TNBC and associated with increased invasion (7) • Stimulates TNBC migration (8) • C766T polymorphism is suspected to increase risk of BC development (9)
LRP1B	LRP-DIT	515	Especially in brain, thyroid, skeletal muscles, testis, ovary, colon	• Considered as tumor suppressor in several cancer types but not in BC • Intracellular nuclear localization correlates with poor prognosis in invasive ductal BC (10)
LRP2	Megalin GP330	517/600	Placenta, kidney, mammary epithelial cells	• High mRNA levels in invasive BC (11) • Mutated in circulating tumor cells from BC (12) • Upregulated in T-47D (13)
LRP3	hLRP105	105	Widely expressed (ovary, heart, brain, liver, pancreas, prostate and small intestine, skeletal muscle)	• ND
LRP4	MEGF7	212	Bone, cartilage, muscle, brain	• ND
LRP5	LR3 LRP7	216	Widely expressed (including in mammary epithelium) with high expression in liver	• Overexpressed in TNBC and basal-like BC (14, 15) • Stimulates STK40 expression and cell viability in TNBC (15) • Regulates glucose uptake in mammary epithelial cells (16)
LRP6		180	Co-expressed with LRP5 during embryogenesis and in adult tissues	• Overexpressed in TNBC and basal-like BC (14) • Role in TNBC cell migration and invasion (MDA-MB-231 and BT549) (15) • Increases the pool of stem cells in TNBC (17)
LRP8	APOER2	106	Brain, placenta, platelets	• Overexpressed in TNBC and ER⁻ BC (18, 19) • Positive regulator of BC stem cells in TNBC, supports chemoresistance and metastasis (18) • Suggested as novel therapeutic target in TNBC (6)
LRP10	LRP9 in mouse	76	Ubiquitous (Leukocyte, lung, placenta, small intestine, liver, kidney, spleen, thymus, colon, skeletal muscle, heart)	• ND
LRP11	sorLA LR11	53	Substantial levels in kidney, testis, ovary, lymph nodes, vascular smooth muscle cells and nervous system	• Drives resistance to anti-HER2 therapy (20)
LRP12	ST7	94	Heart, skeletal muscle, brain, lung, placenta and pancreas	ND

The family core members are underlined in green, the structurally distant members are in blue, and the most distant members are in orange. ND: not determined.