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## The role of dendritic cells in autoimmunity

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### Abstract

Dendritic cells (DCs) initiate and shape both the innate and adaptive immune responses. Accordingly, recent evidence from clinical studies and experimental models implicates DCs in the pathogenesis of most autoimmune diseases. However, fundamental questions remain unanswered concerning the actual roles of DCs in autoimmunity, both in general and, in particular, in specific diseases. In this Review, we discuss the proposed roles of DCs in immunological tolerance, the effect of the gain or loss of DCs on autoimmunity and DC-intrinsic molecular regulators that help to prevent the development of autoimmunity. We also review the emerging roles of DCs in several autoimmune diseases, including autoimmune myocarditis, multiple sclerosis, psoriasis, type 1 diabetes and systemic lupus erythematosus.

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In 2011, the late Ralph Steinman was awarded the Nobel Prize in Physiology or Medicine for his part in the discovery of dendritic cells (DCs) and the important role they have in initiating the adaptive immune response. DCs are specialized sentinel cells that bridge the innate and adaptive immune systems without directly engaging in effector activities such as pathogen killing. DCs recognize pathogens using pattern recognition receptors, including Toll-like receptors (TLRs), and then they migrate to T cell areas of lymphoid organs to present pathogen-derived antigens to antigen-specific T cells. Activated DCs upregulate co-stimulatory molecules and produce cytokines that drive T cell priming and effector differentiation, and they activate various types of immune cells. In the absence of activation, antigen presentation by steady-state DCs might lead to T cell unresponsiveness and might promote tolerance<sup>1</sup>.

DCs comprise two major classes: plasmacytoid DCs (pDCs) and conventional or classical DCs (cDCs). The pDCs rapidly produce type 1 interferon (IFN) following activation through nucleic acid-sensing TLRs, such as TLR7 and TLR9. cDCs are dedicated antigen-presenting

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cells (APCs) that have a characteristic dendritic morphology and express high levels of MHC class II molecules. Mouse cDCs can be broadly categorized into two distinct subsets: the CD8<sup>+</sup> DCs (which are CD103<sup>+</sup> in tissues) and the CD11b<sup>+</sup> DCs (TABLE 1). The CD8<sup>+</sup> DC subset is highly efficient at mediating antigen cross-presentation to cytotoxic CD8<sup>+</sup> T cells. The full functional range of CD11b<sup>+</sup> DCs remains to be determined, although they are believed to preferentially present MHC class II-restricted antigens to CD4<sup>+</sup> T cells. There is substantial genetic and functional heterogeneity within each DC subset, especially in those subsets that are localized to barrier tissues such as the intestine<sup>2,3</sup>. Within each subset, some DCs may express higher levels of TLRs, may preferentially sense pathogens and may secrete pro-inflammatory cytokines, whereas other DCs may be more efficient at migration, antigen presentation and T cell priming. Such functional division into ‘detector’ and ‘presenter’ DCs might be a general feature of DC subsets in lymphoid organs and tissues<sup>4</sup>. Major advances have been made in the characterization of human DCs; these include the discovery of genetic DC deficiencies<sup>5</sup> and the characterization of DCs in the lymph nodes, tissues and inflammatory fluids<sup>6,7,116,117</sup>. Collectively, these studies have shown that there is a strong evolutionary conservation of the major DC subsets between mice and humans (TABLE 1).

Three decades ago, pioneering studies in a rat model of induced autoimmune neural inflammation identified DCs that were in close contact with T cells in inflammatory brain lesions<sup>8</sup>. Strikingly, DCs transferred from mice in which autoimmune neural inflammation had been induced were able to induce the disease in naive recipients, showing their extraordinary potential as APCs<sup>9</sup>. Since then, the crucial involvement of DCs in virtually every aspect of autoimmunity has been documented in patients as well as in animal models. In particular, novel models of constitutive and inducible DC ablation, as well as DC-specific gene targeting, have facilitated direct analysis of the roles of DCs in autoimmune diseases. In this Review, we briefly discuss the current ideas concerning the role of DCs in immunological tolerance and the general aspects of their involvement in autoimmunity. In addition, we review the recent findings regarding the roles that DCs are thought to have in either promoting or inhibiting pathology in various different autoimmune diseases, both in patients and in the associated animal models.

## The role of DCs in immune tolerance

### DCs in central tolerance

Medullary thymic epithelial cells (mTECs) are primarily responsible for the negative selection of autoreactive T cells in the thymus. They express the transcription factor autoimmune regulator (AIRE), which drives the low-level expression of many tissue-specific self antigens for presentation to developing thymocytes. Thymic DCs have been shown to cross-present these self antigens that have been acquired from mTECs<sup>10,11</sup>. In addition, mTECs might recruit thymic DCs in an AIRE-dependent manner and might facilitate the generation of regulatory T (T<sub>Reg</sub>) cells<sup>12</sup>. It is probable that the DC-dependent presentation of self antigens occurs in parallel with the direct presentation of these antigens by mTECs<sup>13</sup> and that it might be secondary to their presentation by mTECs; this has been suggested because the negative selection of thymocytes that are specific for a model self

antigen was shown to be unimpaired by DC ablation<sup>14</sup>. Finally, peripheral DCs might migrate into the thymus and might present peripheral self antigens to induce clonal deletion of T cells or to induce T<sub>Reg</sub> cell generation<sup>15,16</sup>. However, it is unclear whether this mechanism of tolerance is relevant and whether it occurs in conditions other than artificial ones. Overall, the role of DCs in central tolerance seems to be fairly limited and might be restricted to promoting tolerance to a minor subset of self antigens.

### DCs in peripheral tolerance

Pioneering studies in which antigens were delivered to DCs *in vivo* using DC-specific antibodies showed that DC-targeted model antigens induced profound T cell tolerance in the steady state<sup>17,18</sup>. In subsequent studies, antigens were delivered to DCs using Cre recombinase-induced expression of model antigens in DCs *in vivo*<sup>19</sup>. This genetic approach showed that steady-state antigen presentation by DCs induces a profound and irreversible unresponsiveness in CD8<sup>+</sup> T cells; this includes the upregulation of the inhibitory molecules programmed cell death protein 1 (PD1) and cytotoxic T lymphocyte antigen 4 (CTLA4) on CD8<sup>+</sup> T cells<sup>20</sup>. The tolerogenic capacity of DCs in this context was enforced by MHC class II-dependent interactions with T<sub>Reg</sub> cells, and MHC class II-deficient DCs were proposed to induce CD8<sup>+</sup> T cell-mediated autoimmunity<sup>21</sup>. Thus, DCs can induce peripheral T cell tolerance to immunodominant antigens that are expressed at high levels. However, the applicability of this model to the full range of endogenous self antigens remains to be investigated.

DCs were proposed to contribute to peripheral tolerance by facilitating the homeostasis of peripheral T<sub>Reg</sub> cells. DCs can induce T<sub>Reg</sub> cells in the presence of strong stimuli, such as transforming growth factor- $\beta$  (TGF $\beta$ ) and retinoic acid<sup>22,23</sup>, and can support the homeostatic proliferation of T<sub>Reg</sub> cells after their depletion<sup>24</sup>. DCs seem to contribute to the maintenance of T<sub>Reg</sub> cell populations in the peripheral blood, which requires the expression of the co-stimulatory molecules CD80 and/or CD86 on DCs<sup>25</sup>. It was recently suggested that migratory cDCs, but not resident cDCs, in the lymph nodes induce the development of T<sub>Reg</sub> cells that are specific for a particular self antigen, even though that self antigen is expressed by both DC populations<sup>26</sup>. However, constitutive DC ablation causes only a small reduction in T<sub>Reg</sub> cell numbers without any obvious functional consequences<sup>14,25</sup>. Conversely, the administration of the DC growth factor FMS-related tyrosine kinase 3 ligand (FLT3L) leads to an expansion of both the DC and the T<sub>Reg</sub> cell populations<sup>27–29</sup>. This expansion of the T<sub>Reg</sub> cell population was associated with reduced severity of intestinal inflammation and of graft-versus-host disease, showing that a tolerogenic environment had been induced. However, FLT3L treatment skews DC subset distribution, it expands myeloid non-DC populations and it might change the function of individual DCs. Thus, the observed expansion of the T<sub>Reg</sub> cell population might not be a simple consequence of the increased numbers of DCs; this leaves the idea that there is a simple linear relationship between DCs and T<sub>Reg</sub> cells open to question. Taken together, DCs seem to facilitate the induction and/or the maintenance of peripheral T<sub>Reg</sub> cells; however, they are neither strictly required for T<sub>Reg</sub> cell homeostasis nor do they seem to preferentially regulate T<sub>Reg</sub> cells compared to effector T cells.

## Tolerogenic DCs: subset or state?

The role of DCs in self-tolerance might be primarily determined by their functional state or it might be mediated through a distinct ‘tolerogenic’ DC subset. Although DCs that have distinctly tolerogenic properties have been shown to arise in artificial systems<sup>30</sup>, genetic evidence for a tolerogenic DC subset in the steady state is lacking. For example, an indispensable role for intestinal CD103<sup>+</sup> DCs in T<sub>Reg</sub> cell induction has not been supported by loss-of-function analyses<sup>31,32</sup>. DCs that express perforin and that can kill cognate T cells might be an exception, although the identity of these DCs *in vivo* has not yet been established<sup>33</sup>. Another interesting exception might be peripheral DCs that express AIRE and that therefore present an unusually broad range of self antigens, including antigens that are normally restricted to other cell types. These DCs might correspond to very rare cells that have hallmarks of the DC lineage (such as MHC class II expression) but that have an unconventional surface phenotype, such as low levels of expression of the haematopoietic marker CD45 (M. Anderson, personal communication). Although the exact role of these DCs in peripheral tolerance remains to be established, the tolerogenic properties of cDCs seem to be primarily mediated by their functional state (for example, by the absence of activation).

## The role of DCs in autoimmunity: general aspects

### The numbers game

One important question about the role of DCs in autoimmunity has been whether the actual size of the overall DC population contributes to the development of autoimmunity. It was initially reported that the constitutive ablation of cDCs in inbred mice resulted in normal T cell development, the establishment of a normal peripheral T cell compartment without overt hyperactivation and normal T<sub>Reg</sub> cell numbers in most lymphoid organs<sup>14</sup>. However, DC-deficient mice were shown to develop an age-dependent myeloproliferative syndrome that was associated with increased serum concentrations of FLT3L. A subsequent study in mice of mixed backgrounds suggested that constitutive DC ablation does lead to autoimmune manifestations<sup>34</sup>. However, the nature and full course of the disease was not fully described nor was any evidence for T cell auto-reactivity provided in this study. Thus, the observed disease manifestations were apparently caused by myeloproliferation rather than by autoimmunity. Indeed, constitutive DC ablation in mice of an autoimmunity-prone background ameliorated rather than exacerbated the resulting disease<sup>35</sup>. Furthermore, the global constitutive ablation of specific DC subsets — including the CD8<sup>+</sup> cDCs<sup>31</sup> and pDCs<sup>36</sup> — did not cause any detectable autoimmunity. Thus, DCs seem to be dispensable for peripheral tolerance even though they might contribute to the process. In addition, their loss does not cause overt autoimmunity. This might reflect both the redundancy of tolerogenic DC function and the essential role of DCs in the activation of autoreactive T cells<sup>4</sup>.

Increased DC numbers (for example, after cytokine treatment) have been associated with T<sub>Reg</sub> cell induction and with the development of a tolerogenic environment<sup>27–29</sup>, as discussed above. Paradoxically, it was also suggested that impaired apoptosis of DCs might increase DC numbers and cause autoimmunity<sup>37–39</sup>. However, it remains unclear whether the *in vivo* blockade of apoptosis was limited to DCs in these studies; indeed, some of these

studies used a Cre-deleter strain that has a broad deletion range (BOX 1). Taken together, the association between increased DC numbers and autoimmunity has not been conclusively shown and its potential relevance remains debatable.

### DC-intrinsic regulation of autoimmunity

Although the mere gain or loss of DCs does not seem to initiate autoimmunity, changes in DC functionality might induce inflammation and/or autoimmune manifestations. The advent of DC-specific gene targeting (BOX 1) has facilitated the direct genetic manipulation of DC functionality and the analysis of its effect on autoimmunity. Several genes have been identified that induce spontaneous autoimmune and/or inflammatory manifestations when they are deleted in DCs<sup>40–46</sup> (TABLE 2). Most of these genes are negative regulators of pro-inflammatory signals (for example, A20 (also known as TNFAIP3) and SH2 domain-containing protein tyrosine phosphatase 1 (SHP1; also known as PTPN6)) or molecules that are upstream or downstream of anti-inflammatory signals (such as  $\alpha$ V $\beta$ 8 integrin and signal transducer and activator of transcription 3 (STAT3), respectively). Importantly, these are general negative regulators of immune activation and their deletion in other cell types also causes inflammation and/or autoimmunity. Thus, inappropriate DC activation is apparently sufficient to induce a broad range of autoimmune manifestations and it is actively opposed by multiple pathways and factors. However, cell-type specific negative regulators that prevent DC hyperactivation and autoimmunity remain mostly unknown. Given the powerful functionality and unique gene expression profile of DCs, we propose that such (unidentified) DC-specific regulators might exist and might have a crucial role in the maintenance of tolerance and in the prevention of autoimmunity.

### The role of DCs in autoimmune diseases

As discussed above, the involvement of DCs in tolerance and autoimmunity is complex and bidirectional (FIG. 1). Indeed, DCs might promote tolerance through multiple mechanisms, including through the generation and maintenance of T<sub>Reg</sub> cells, as well as through the induction of T cell unresponsiveness. Conversely, the powerful antigen presentation capacity of DCs might promote the priming and/or the effector differentiation of self-reactive T cells, either because of inappropriate activation signals or because of a cell-intrinsic breakdown of negative regulation. Below, we review the current evidence for these mechanisms from several major autoimmune diseases, which were primarily chosen because of the availability of data. We consider a somewhat special case of cardiac autoimmunity that is associated with molecular mimicry, as well as tissue-specific diseases of the central nervous system and skin. We also review two major diseases for which multigenic spontaneous mouse models are available: type 1 diabetes (T1D) and systemic lupus erythematosus (SLE).

### DCs in cardiac autoimmunity

Autoimmune myocarditis often occurs after viral infections (for example, coxsackievirus or adenovirus infections) and is associated with the production of autoantibodies against cardiac proteins, which suggests that there might be molecular mimicry between the pathogen-derived and the cardiac antigens. Similar cases of molecular mimicry have been

documented in rheumatic pancarditis, in which infection with group A *Streptococcus* (also known as *Streptococcus pyogenes*) initiates an immune response to host antigens in the perivascular connective tissue and in the myocardium. Adoptive transfer of DCs that are loaded with cardiac antigens can induce the infiltration of CD4<sup>+</sup> T cells into cardiac tissue and the development of experimental autoimmune myocarditis (EAM); this indicates that DCs are sufficient for inducing the disease<sup>47</sup>. The activation of DCs by the pro-inflammatory cytokines interleukin-1 (IL-1)<sup>48</sup> and granulocyte–macrophage colony-stimulating factor (GM-CSF)<sup>49</sup> was found to be crucial for efficient autoreactive T cell responses and EAM induction. Another study described the contribution of the RNA-sensing receptor TLR7 to EAM severity, suggesting that TLR7-expressing DCs (such as pDCs) have a role in driving the disease<sup>50</sup>. In a related model of pericardial inflammation caused by the injection of antigen-pulsed DCs, the cardiac proteoglycan biglycan was found to substitute for pathogen-associated stimuli and to induce cDC activation in the context of damaged cardiomyocytes<sup>51</sup>. Thus, in the inducible model of autoimmune myocarditis, the integration of cytokines and pathogen-derived or endogenous inflammatory stimuli by DCs is important for the development of the disease. However, it remains to be determined whether DCs are strictly necessary for the development of autoimmune myocarditis and whether they have a role in antigen mimicry-associated cardiac inflammation in patients.

### The role of DCs in neurological autoimmunity

In multiple sclerosis and its associated animal models — such as experimental autoimmune encephalomyelitis (EAE) — T cells that are specific for the myelin antigen are crucial effector cells that drive inflammation and tissue damage. Importantly, the rare autoreactive T cells that are present in the natural T cell repertoire need to encounter their cognate antigen on APCs at least twice for the development of EAE. During the initial priming phase, the population of T cells expands and the cells become polarized encephalitogenic T helper 1 (T<sub>H</sub>1) or T<sub>H</sub>17 effector cells. Subsequent interactions, which occur in close proximity to or within the brain, confer a crucial restimulation signal.

It was shown that cDCs are highly efficient in priming and polarizing encephalitogenic T cells in EAE caused by a myelin-associated peptide antigen, as well as in demyelinating disease caused by Theiler's murine encephalomyelitis virus<sup>52,53</sup>. Moreover, transgenic re-expression of MHC class II molecules specifically in the cDCs of MHC class II-deficient mice was sufficient to mediate peptide-induced EAE<sup>54</sup>. Notably, the subcutaneous delivery of the full-length protein antigen failed to achieve efficient T cell priming in this model, which suggests that in some situations DCs are insufficient as APCs. Furthermore, several models of DC lineage ablation showed that DCs are completely dispensable for encephalitogenic T cell priming in myelin protein-induced EAE<sup>55,56</sup>. Thus, DCs are involved in and might be sufficient for encephalitogenic T cell priming, but other types of APCs might substitute for DCs in inducible multiple sclerosis models.

DC-specific targeting approaches were recently used to identify several molecules that regulate the polarization capacity of DCs that prime T<sub>H</sub> cells; for instance, DC-specific ablation of mitogen-activated protein kinase p38 $\alpha$  (p38 $\alpha$  MAPK; also known as MAPK14) impaired the priming and the maintenance of T<sub>H</sub>17 responses and prevented disease in both

actively and passively induced models of EAE<sup>57</sup>. Given the evidence discussed above<sup>55,56</sup>, it is probable that p38 $\alpha$  MAPK-deficient DCs redirect T cells to a different effector cell fate rather than failing to prime them. In addition to such cell-intrinsic effects, DCs have a role in establishing the cytokine milieu that is required for T cell polarization. Thus, a DC-specific deficiency of  $\alpha$ V $\beta$ 8 integrin, which is required for the local activation of latent TGF $\beta$ , diminished the ability of cDCs to support T<sub>H</sub>17 cell responses and rendered mice resistant to peptide-induced — but not adoptively transferred — EAE<sup>58</sup>. Conversely, DC-intrinsic TGF $\beta$  signalling reduces the capacity of DCs to prime T cells and its specific blockade in DCs was shown to worsen EAE in immunization-induced and T cell receptor (TCR)-transgenic models<sup>59</sup>. Similarly, engagement of the surface-expressed receptor T cell immunoglobulin and mucin domain-containing protein 1 (TIM1; also known as HAVCR1) on DCs increases their capacity to drive effector T cell differentiation as opposed to T<sub>Reg</sub> cell induction. The antibody-mediated ligation of TIM1 exacerbated EAE, which suggests that aberrant DC hyperactivation might strongly contribute to the breach of tolerance in this model<sup>60</sup>.

IFN $\beta$  is used as an effective therapy for multiple sclerosis, thus the modulation of DC function by type I IFN is of particular interest in this disease. The genetic blockade of type I IFN induction or signalling exacerbates active EAE, possibly by facilitating T<sub>H</sub>17 cell effector differentiation or maintenance<sup>61,62</sup>. The suppression of T<sub>H</sub>17 cell responses by type I IFN requires the expression of the IFN $\alpha$ / $\beta$  receptor (IFNAR) by DCs but not by macrophages or microglia; this implicates DC-intrinsic IFNAR signalling in the control of T<sub>H</sub> cell differentiation<sup>62</sup>. In addition, administered IFN $\beta$  might directly inhibit DC functions, such as their migration, thereby reducing T cell priming<sup>63</sup>. Thus, DCs might represent important cellular targets of anti-inflammatory type I IFN signalling during the natural course of multiple sclerosis and of IFN $\beta$  therapy during the treatment of the disease.

Primed encephalitogenic effector T cells have to infiltrate the brain to induce autoimmune inflammation. However, the brain is a site of immune privilege that is protected by multiple mechanisms, including the blood–brain barrier. Notably, cDCs are present at all sites of immune cell entry to the brain, residing in the perivascular space of the blood–brain barrier and integrating into the physical barrier structures<sup>64,65</sup>. Thus, DCs might act as ‘gatekeepers’ that control T cell entry to the brain, presenting self antigen *in situ* to effector T cells but also inducing the T<sub>Reg</sub> cell activity that reduces inflammation and that maintains immune privilege at this site.

In the model of active relapsing EAE, CD11b<sup>+</sup> cDCs in the brain were proposed to drive and to maintain T<sub>H</sub>17 cell differentiation<sup>66</sup>. Indeed, extensive interactions between DCs and T cells were documented in the inflamed central nervous system (CNS) and were partially dependent on Cc-chemokine receptor 4 (CCR4) expression by DCs<sup>67</sup>. Both T<sub>H</sub>17 and T<sub>H</sub>1 cells in EAE secrete GM-CSF and this is essential for their effector functions<sup>68,69</sup>. GM-CSF activates the production of IL-23 by APCs, probably including DCs. Conversely, both constitutive and inducible ablation of DCs was shown to exacerbate EAE, which suggests that DCs might collectively ameliorate inflammation in the brain<sup>55</sup>. Furthermore, DC-specific expression of a myelin self antigen protected against the development of EAE and this was associated with T<sub>Reg</sub> cell induction via the expression of inhibitory PD1 ligand 1

(PDL1) on DCs. Similarly, pDC depletion was shown to exacerbate acute and relapsing EAE and to increase the production of inflammatory cytokines by T cells<sup>70</sup>. Moreover, the loss of MHC class II expression specifically on pDCs was shown to exacerbate EAE, apparently by reducing the induction of T<sub>Reg</sub> cells that were specific for myelin antigens after immunization<sup>71</sup>. Similarly, the constitutive ablation of pDCs increased EAE disease severity, which suggests that the pDCs have regulatory functions at all stages of EAE (S.H., unpublished observations).

Taken together, data from EAE models suggest that DCs are important — albeit not strictly required — for the T cell priming and the effector T cell polarization that sustains brain inflammation (FIG. 1). Conversely, strong genetic evidence suggests that DCs reduce T cell-mediated inflammation and promote T<sub>Reg</sub> cell differentiation at later stages of the disease. A major challenge now lies in extrapolating these data to multiple sclerosis in humans, in which the identity of the initiating event, the self antigens that are involved and the sites of T cell priming remain unresolved. Evidence for pro-inflammatory DC activity in multiple sclerosis has been reported<sup>72</sup>, but whether DCs also have tolerogenic activity remains to be established.

### DCs in psoriasis

Psoriasis is an inflammatory disease of a barrier tissue (the skin) and may not be primarily of autoimmune origin; however, it seems to have a strong autoimmune component. Keratinocytes in the psoriatic plaque are viewed as both ‘victims’ and ‘perpetrators’, as they drive the recruitment of immune cells by expressing antimicrobial peptides, chemokines and growth factors. Effector T cells that produce IL-17 and IL-22 — including CD4<sup>+</sup> T<sub>H</sub>17 cells and potentially  $\gamma\delta$  T cells — have a major role in psoriatic inflammation<sup>73,74</sup>, which raises the question of which cells and signals they are primed by.

One emerging model of psoriasis-like inflammation is induced by painting the skin with the TLR7 ligand imiquimod<sup>75</sup>, although TLR7-independent effects that are induced by the vehicle might also contribute to plaque formation<sup>76</sup>. It was recently shown that DC depletion in CD11c–diphtheria toxin receptor (DTR) mice protected against disease development in this model<sup>77</sup>. This study suggested that the production of IL-36 by skin cells activates DCs to produce IL-23 and to drive T<sub>H</sub>17 cell differentiation, which indicates that DCs are necessary for the disease. Conversely, DC-specific deletion of the A20-binding inhibitor of NF- $\kappa$ B inhibitor activation 1 (ABIN1; also known as TNIP1) caused TLR-induced cDC activation and hyperproduction of IL-23 and greatly exacerbated imiquimod-induced psoriasis (A. Ma, personal communication). Notably, polymorphisms in the *TNIP1* gene have been associated with psoriasis in patients. These observations suggest that the aberrant hyperactivation of cDCs might be sufficient for innate immune signals to induce psoriasis, and further investigation of cDC function in other models of the disease is warranted.

Type I IFN is sufficient to inadvertently induce flares of psoriasis in patients who have been treated for other conditions such as multiple sclerosis. Therefore, it has been postulated that type I IFN-producing pDCs have a major role in the initiation of psoriasis. Indeed, nascent psoriatic plaques show prominent infiltration of pDCs<sup>78</sup> and express the chemotactic factor chemerin, which recruits pDCs to the skin through chemokine-like receptor 1 (CMKLR1;

also known as CHEMR23)<sup>79</sup>. Furthermore, pDCs were shown to accumulate at the site of skin injury, to secrete type I IFN in a TLR-dependent manner and to promote either wound healing in normal mice or inflammation in mice from autoimmunity-prone backgrounds<sup>80,81</sup>. Using biochemical analysis of psoriatic plaques, Gilliet and colleagues<sup>82</sup> showed that activated keratinocytes produce the antimicrobial peptide LL37, which can bind to extracellular DNA and RNA that is generated in the skin during normal cell turnover or during injury<sup>82</sup>. The resulting nanoscale LL37–nucleic acid particles are protected from nuclease degradation, and they can induce type I IFN production by pDCs and activate cDCs<sup>82,83</sup> (FIG. 2). Unexpectedly, pDC ablation did not affect the severity of imiquimod-induced psoriatic plaque formation in mice<sup>118</sup>.

Thus, the pDC–type I IFN axis seems to have a prominent role in the initiation of inflammation and in the transition to a chronic disease, but it might be redundant in actual psoriatic lesion formation. By contrast, cDCs might be essential for IL-23 production, which drives T cell effector differentiation and plaque formation<sup>118</sup>. The imiquimod-induced model of psoriasis provides useful insights into the course of the disease, but a better understanding of the roles of pDCs and cDCs in psoriasis will be gained by studying these cells in spontaneous genetic models of the disease.

## DCs in T1D

Pancreatic  $\beta$ -islet cells are the source of the autoantigens that drive disease in T1D but they are unable to directly prime diabetogenic CD4<sup>+</sup> or CD8<sup>+</sup> T cell responses<sup>84</sup>, which suggests that cross-presenting APCs might be involved. Self antigens might be released after the death of  $\beta$ -islet cells, which occurs physiologically during pancreatic remodelling in non-obese diabetic (NOD) mice once they reach 2 weeks of age. Such  $\beta$ -islet cell-derived antigens can be captured by DCs and presented to autoreactive T cells after the migration of DCs to the pancreatic lymph nodes, thereby initiating T1D development<sup>85</sup>. Indeed, T1D development in NOD mice was shown to be prevented by inducible DC depletion and to be restored by the adoptive transfer of CD11b<sup>+</sup> DCs<sup>86</sup>. However, the CD11c–DTR model of DC ablation used in these studies is not entirely DC-specific and was shown to deplete other potentially relevant cell types. Despite this caveat, these data suggest that  $\beta$ -islet cell death might initiate diabetogenic T cell responses via DCs. DCs are constitutively sampling and presenting  $\beta$ -islet cell-derived antigens to T cells in the pancreatic lymph nodes<sup>87</sup>, but why they induce active immune responses rather than tolerance in response to self antigens is not yet clearly understood. It was proposed that dying pancreatic  $\beta$ -islet cells release the DNA-binding protein high mobility group protein B1 (HMGB1), which is a known danger signal that can activate DCs through TLR2 and TLR4. Antibody-mediated blockade of HMGB1 reduced the incidence and the onset of T1D in NOD mice, and this was associated with a decreased expression of co-stimulatory molecules on CD11b<sup>+</sup> DCs<sup>88</sup>. These results are consistent with a protective role of TLR2 deficiency in T1D pathogenesis in NOD mice<sup>89</sup>. In addition, inflammatory signals such as tumour necrosis factor (TNF)<sup>90</sup> and IL-21 (REF. 91) were shown to promote the maturation of  $\beta$ -islet cell-associated DCs and their migration to the pancreatic lymph nodes.

The expression of IFN $\alpha$  is detected in the pancreatic  $\beta$ -islet cells of patients with T1D and in experimental models of T1D, and the induction of a type I IFN response accelerates T1D development<sup>92,93</sup>. It was shown that pDCs infiltrate pancreatic islets and the pancreatic lymph nodes in NOD mice; moreover, antibody-mediated blockade of type I IFN-mediated signalling reduced T1D development in this model<sup>94</sup>. A recent study showed that pDCs in the  $\beta$ -islet cells produce type I IFN and that this is essential for the development of T1D<sup>95</sup>. According to the proposed model, dying  $\beta$ -islet cells release DNA that forms complexes with DNA-specific antibodies and with cathelin-related antimicrobial peptide (CAMP), which are produced by B1 cells and neutrophils, respectively. The resulting immune complexes activate type I IFN production by pDCs in a TLR9-dependent manner, possibly promoting cDC activation and autoreactive T cell priming<sup>95</sup>. In patients with T1D, pDCs were shown to present  $\beta$ -islet cell-derived antigens that were obtained by immune complex capture more efficiently than cDCs, thereby directly activating diabetogenic T cells<sup>96</sup>. Thus, aberrant pDC-mediated type I IFN production seems to be a common mechanism that leads to pathogenesis in several autoimmune diseases, including T1D (FIG. 2).

In line with the putative tolerogenic functions of DCs that are discussed above, these cells have also been proposed to protect NOD mice from developing T1D. The administration of FLT3L or G-CSF protected NOD mice from T1D development and was associated with an expansion of the DC population and an enhanced generation of T<sub>Reg</sub> cells<sup>97-99</sup>. However, these treatments might modulate the function of the expanded DC population and/or they might affect multiple non-DC cell types (as discussed above). The depletion of DCs using the CD11c-DTR system in TCR-transgenic NOD mice that have progressed to insulinitis was shown to exacerbate T1D and this was suggested to be due to the loss of pDCs<sup>86</sup>. However, the CD11c-DTR system is not known to appreciably deplete pDCs, which leaves this result open to alternative explanations. A recent study described the expression of the  $\beta$ -islet cell-derived antigen insulin in a distinct AIRE<sup>+</sup> DC subset, and DC-specific deletion of insulin caused insulinitis in wild-type mice<sup>100</sup>. The molecular and cellular identity of this DC subset, as well as its role in T1D development in NOD mice, remains to be established.

Taken together, DCs seem to have an essential role in the pathogenesis of T1D through their ability to take up and present  $\beta$ -cell antigens to pathogenic T cells in the pancreatic lymph nodes. This process might happen early in life — that is, during the remodelling of pancreatic tissue — and it might be promoted by genetic predisposition factors such as the impaired clearance of dead cells. Although DCs might induce immunological tolerance against  $\beta$ -cell antigens in the absence of such a predisposition or they might suppress autoreactive T cell responses, these DC functions remain to be proved in defined genetic systems.

## DCs in SLE

SLE is often considered to be an antibody-driven autoimmune disease, unlike T1D or multiple sclerosis, which are thought to be primarily T cell-mediated. However, SLE shows a strong association with MHC haplotypes, which suggests that T cell responses are essential for the development of this disease. Indeed, even an SLE-like disease that is driven by TLR7 overexpression in mice, and that presumably involves an innate initiating stimulus,

is completely MHC haplotype-dependent<sup>101</sup>. This circumstantial evidence suggests that DCs are possibly involved in the presentation of chromatin and RNA-associated proteins to self-reactive T cells. However, the role of DCs in SLE has only been definitively studied in a single model of SLE — one that is driven by a mutation in the death receptor CD95 (also known as FAS). The loss of CD95 in DCs was found to be sufficient to induce disease manifestations in C57BL/6 mice<sup>38</sup>, although the specificity of the CD95 deletion remains to be confirmed (BOX 1). Furthermore, the constitutive deletion of DCs in SLE-prone MRL-*Fas<sup>Lpr</sup>* mice reduced T cell population expansion, plasmablast differentiation and target organ damage, which ameliorated the disease<sup>35</sup>. T cell priming was not substantially affected in the absence of DCs in this model, probably because lymphoproliferation seems to be a key initiating event in this model.

Conversely, DC-specific deletion of several negative regulators is sufficient to induce an SLE-like disease *in vivo* (TABLE 2). The SLE manifestations that are caused by DC-specific ablation of B lymphocyte-induced maturation protein 1 (BLIMP1; also known as PRDM1), a transcriptional repressor that controls the differentiation of B and T cells, are of particular interest<sup>42</sup>. Polymorphisms in the *PRDM1* gene have been associated with SLE in humans, which shows that it is a highly relevant candidate regulator. BLIMP1-deficient DCs were shown to induce T follicular helper cell differentiation via increased IL-6 production and, thus, they enhanced germinal centre reactions and humoral autoreactivity. Both the monoallelic and biallelic loss of *PRDM1* in DCs induced this phenotype, which highlights the sensitivity of DCs to the gene dosage of *PRDM1*. Strikingly, the resulting disease occurred predominantly in female animals, which recapitulates the strong bias towards female patients with SLE. This study shows that the aberrant activation of DCs might be sufficient to drive the entire spectrum of cellular and humoral autoreactivity in SLE and that it might contribute to the characteristic sex bias that is associated with the disease.

A prominent role for type I IFNs in SLE pathogenesis has been known since the 1970s, when high serum levels of type I IFN were noted to be associated with the exacerbation of SLE disease parameters. Subsequent studies reported a gene expression signature of type I IFN signalling in leukocytes from SLE patients that correlated with disease severity<sup>102</sup>. Type I IFNs might have pleiotropic effects in promoting this disease, for example, by inducing the maturation of monocytes and the stimulation of autoantibody production by B cells. Indeed, genetic ablation of type I IFN signalling ameliorates SLE development in animal models, such as in NZB/NZW-derived lupus-prone mouse strains<sup>103,104</sup>. Given the powerful type I IFN-producing capacity of pDCs, these cells have been proposed to be an important source of aberrant type I IFN secretion and major drivers of SLE progression<sup>105</sup>.

In recent years, multiple studies have provided strong — albeit indirect — evidence that pDCs have a role in SLE propagation. Patients with SLE have reduced numbers of pDCs in the blood and an increased accumulation of pDCs in tissue lesions<sup>106</sup>. Furthermore, pDCs can be activated by self-nucleic acids that are in a complex with antibodies<sup>107,108</sup>, or with DNA- or RNA-binding proteins such as HMGB1 (REF. 109). It was recently shown that complexes of DNA and LL37 are released from activated neutrophils as components of neutrophil extracellular traps (NETs) in a process termed ‘NETosis’. These complexes induce pDCs to secrete type I IFN, which in turn activates monocytes and neutrophils and

drives the vicious circle of immune activation that is seen in patients with SLE<sup>110,111</sup> (FIG. 2). Moreover, activated pDCs become resistant to glucocorticoids, which is a possible reason for the limited efficacy of these drugs in the treatment of SLE<sup>112,113</sup>. Thus, pDCs might represent an important component of SLE pathogenesis and they might provide an attractive therapeutic target. Indeed, pDC-specific therapy would not affect type I IFN production by other cell types and therefore would cause less immunosuppression than a general type I IFN blockade. However, causal evidence for the role of pDCs in SLE is lacking and awaits further genetic studies in experimental models of pDC lineage ablation.

## Conclusions and future directions

Although the experimental evidence reviewed above is obviously incomplete, some preliminary conclusions can be made. In each disease reviewed, substantial support exists for the pathogenic role of DCs, which drive the activation and effector differentiation of the relevant T cell populations (FIG. 1). In addition, some DC-based mechanisms, such as those involving type I IFN production by pDCs, might represent common mechanisms that lead to pathogenesis in autoimmune diseases as distinct as psoriasis, SLE and T1D (FIG. 2). Conversely, the evidence for the tolerogenic roles of DCs is less robust in most diseases other than EAE, which is an inducible model in which T cells are primed in the periphery but function in a unique, privileged tissue. Thus, the role of DCs in preventing autoimmunity might be confined to the earliest stages of spontaneous disease, at a time when this cannot be prospectively shown. One major challenge for the field is to extend the studies in inducible models, such as EAM, EAE and imiquimod-induced psoriasis, to spontaneous monogenic or (better yet) multigenic models (BOX 2). An even bigger challenge is to translate the findings in these models to the infinitely more complex human diseases. Although the possibility of DC-based therapies for autoimmune diseases does not appear to be close at hand, a better understanding of DC functions in autoimmunity would bring it closer to practice.

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## Glossary

|                           |  |
|---------------------------|--|
| <b>Type I interferon</b>  | (Type I IFN). A family of cytokines that comprises IFN $\beta$ and multiple subtypes of IFN $\alpha$ , which all signal through a common IFN $\alpha/\beta$ receptor (IFNAR). Type I IFNs are typically induced by viral infection and can confer an antiviral state growth arrest and/or apoptosis in host cells, as well as being able to recruit and to activate multiple immune cell types |
| <b>Cross-presentation</b> | The ability of certain antigen-presenting cells to load peptides that are derived from exogenous antigens on MHC class I molecules. This property is uncommon, as most cells exclusively present   |

peptides from their endogenous proteins on MHC class I molecules. Cross-presentation is essential for the initiation of immune responses to viruses that do not infect antigen-presenting cells

**DC ablation**

The process by which dendritic cells (DCs) are depleted; for example using diphtheria toxin expression or administration. Diphtheria toxin or the diphtheria toxin receptor can be specifically expressed in DCs, which facilitates constitutive or diphtheria toxin-inducible DC ablation, respectively. Limitations to this method include the potential depletion of other cell types, the nonspecific toxicity of the administered diphtheria toxin and secondary effects owing to substantial cell death

**FMS-related tyrosine kinase 3 ligand**

(FLT3L). An important growth factor in dendritic cell (DC) development. Its receptor FLT3 is expressed on haematopoietic progenitor cells as well as on all DCs, and FLT3L administration causes uneven expansion of most DC subsets

**Type 1 diabetes**

(T1D also known as insulin-dependent or juvenile diabetes). A disease that arises in children and young adults and that is caused by the destruction of pancreatic insulin-producing  $\beta$ -islet cells. It is thought that  $\beta$ -islet cell destruction is mediated by  $CD4^+$  and  $CD8^+$  T cells that are specific to  $\beta$ -islet cell autoantigens, such as insulin, zinc transporter 8 (ZNT8), islet-specific glucose-6-phosphatase catalytic subunit-related protein (IGRP) and chromogranin A. Inbred non-obese diabetic (NOD) mice represent an excellent spontaneous model of T1D, a disease which is controlled by multiple genetic loci

**Systemic lupus erythematosus**

(SLE). A disease characterized by the production of autoantibodies against self-DNA, chromatin, and RNA-associated proteins. The resulting immune complexes are deposited and induce inflammation in multiple tissues, particularly in kidney glomeruli (glomerulonephritis). Mouse models of spontaneous SLE include: the NZB/NZW strain and its derivative strains; *MRL-Fas<sup>Lpr</sup>* mice with a mutation in the death receptor CD95; and models with overexpression of Toll-like receptor 7 (TLR7; for example, *Yaa* locus-containing strains and *Tlr7*-transgenic mice)

**Autoimmune myocarditis**

A form of heart inflammation that is commonly associated with dilated cardiomyopathy, which is the most common cause of heart failure in young adults. It can be modelled in mice with experimental autoimmune myocarditis (EAM), which is induced by immunization with cardiac proteins such as the  $\alpha$ -myosin heavy chain

|  |   |
|--|---|
| <b>Multiple sclerosis</b>                        | A disease that is most frequent in young female adults as a relapsing-remitting disease. It involves inflammation and focal neurodegeneration of the white matter of the central nervous system, which results from an autoimmune response to the components of the myelin sheath   |
| <b>Experimental autoimmune encephalomyelitis</b> | (EAE). A classical induced model of multiple sclerosis. EAE can be either actively induced by immunization against proteins of the myelin sheath, or passively induced by the adoptive transfer of encephalitogenic T helper cells. Transgenic models of EAE expressing myelin-specific T cell receptors are also widely studied  |
| <b>Psoriasis</b>                                 | A chronic inflammation of the skin that is characterized by vascular hyperplasia and keratinocyte hyperproliferation, and is accompanied by a local pro-inflammatory milieu and immune cell infiltration. Mouse models typically recapitulate only some aspects of the disease and include genetic manipulation or chemical activation of immune responses (for example, with the Toll-like receptor 7 agonist imiquimod) |

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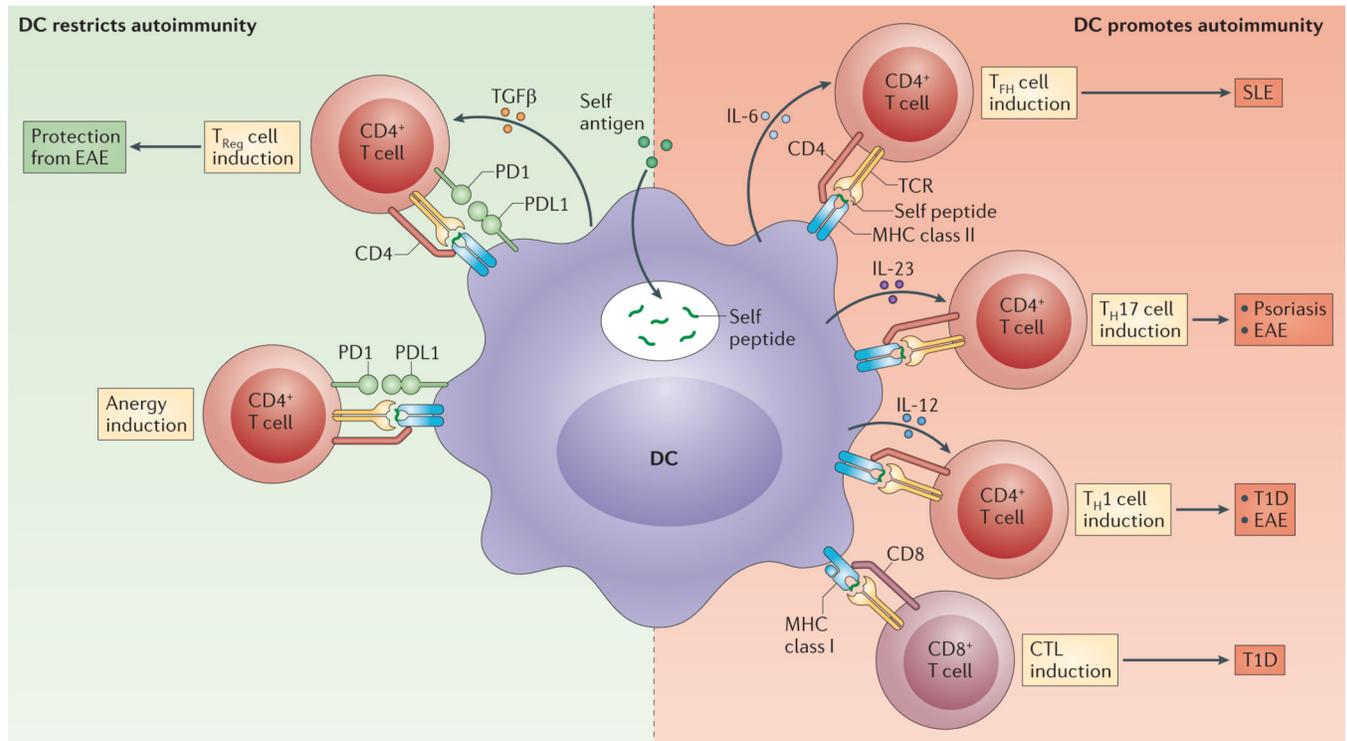
**Box 1****Advantages and limitations of DC-specific gene targeting**

Dendritic cell (DC)-specific gene targeting in mice uses conditional LoxP-flanked (floxed) alleles combined with ‘deleter’ strains that express Cre recombinase specifically in DCs. The available deleter strains have Cre transgenes controlled by the *Itgax* gene, which encodes the DC-specific marker CD11c. However, CD11c is also expressed at lower levels by several immune cell types, including natural killer (NK) cells and activated T cells. The mouse strain expressing Cre recombinase under the control of the 5 kilobase *Itgax* promoter<sup>38</sup> was recently shown to mediate Cre recombination in a broad range of immune cells<sup>114</sup>. The strain expressing Cre recombinase under the control of the 160 kilobase genomic fragment of *Itgax*<sup>115</sup> mediates Cre recombination in the majority of classical DCs and plasmacytoid DCs, but only in approximately 10% of non-DCs (including T cells and NK cells). In combination with Cre-inducible diphtheria toxin or diphtheria toxin receptor (DTR), this strain enables the efficient and specific ablation of the classical DC lineage. However, the DC-intrinsic nature of the observed phenotypes should be ascertained in every case, for example, through comparison with other deleter strains.

**Box 2****Key questions and future directions for the field**

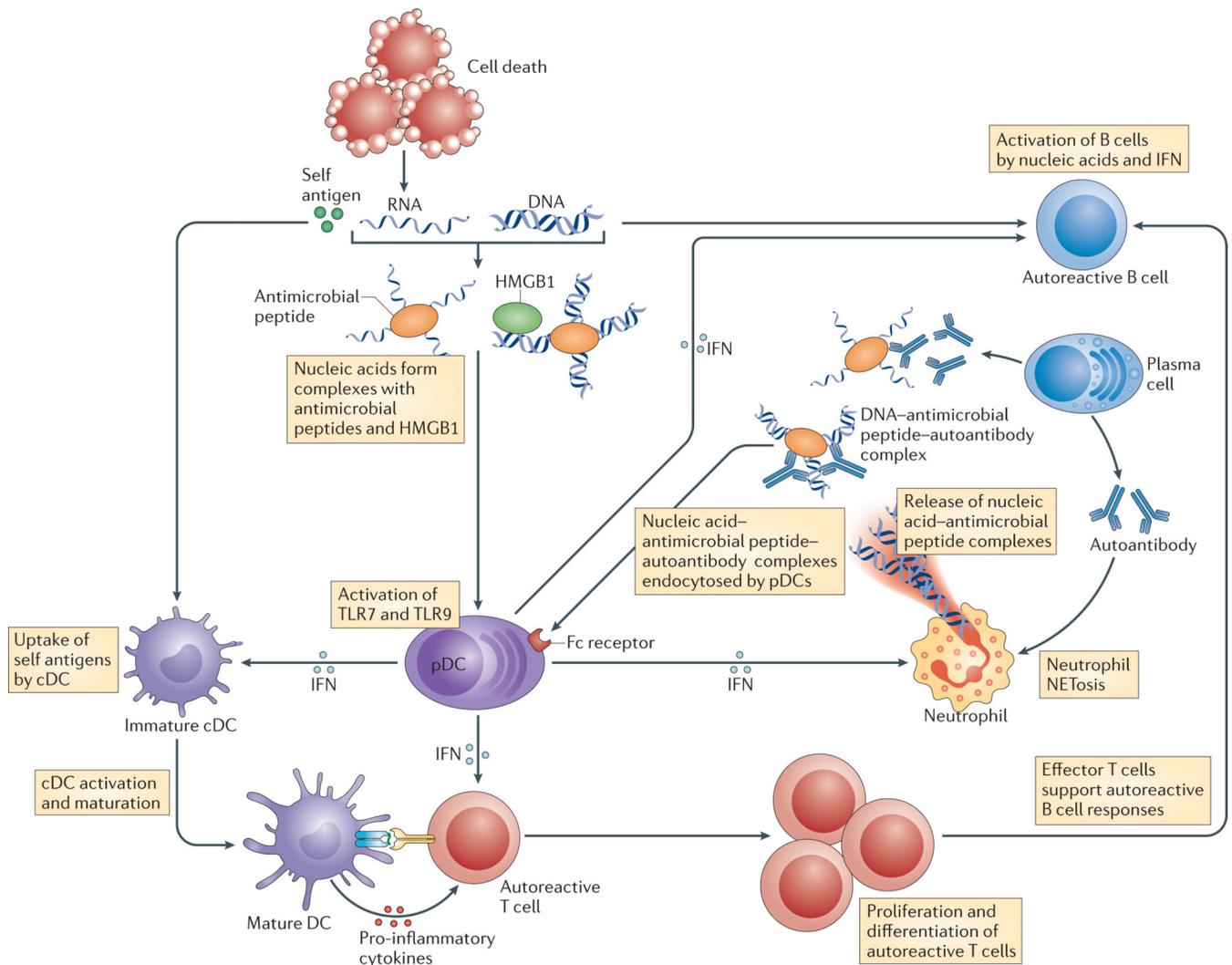
- What is the specific role of individual dendritic cell (DC) subsets (as opposed to the general population of DCs) in human autoimmune disease and in spontaneous multigenic animal models of autoimmunity?
- Are ‘tolerogenic DCs’ a developmentally and genetically distinct subset, a functional state or an artificial (albeit useful) cell type?
- Is there a ‘special relationship’ between DCs and regulatory T ( $T_{Reg}$ ) cells that differs from the relationship of DCs with effector T cells? What is the role of DCs in  $T_{Reg}$  cell induction and  $T_{Reg}$  cell-mediated suppression of T effector cell function?
- What are the environmental factors and cell-extrinsic signals that lead to abnormal DC activation and to the ensuing breach of tolerance?
- Are there DC-specific cell-intrinsic regulators that prevent DC activation and autoimmunity?
- Does the self-nucleic-acid-induced type I interferon production by plasmacytoid DCs have a causative role in autoimmune diseases? Can it work in reverse to oppose autoimmunity — for example, in multiple sclerosis?

The answers to these questions would provide not only essential insights into the causes and mechanisms of autoimmunity, but also potential targets for rational therapeutic approaches.



**Figure 1. Potential roles of DCs in autoreactive T cell responses**

Depending on the inflammatory context and the expression of cell-intrinsic regulators, dendritic cell (DC)-mediated presentation of self antigens might promote or inhibit autoimmune responses; for example, the presentation of self antigens to T cells in the context of the programmed cell death protein 1 (PD1)—PD1 ligand 1 (PDL1) interaction and/or transforming growth factor- $\beta$  (TGF $\beta$ ) signalling can lead to anergy in self-reactive T cells or it can promote their development into regulatory T ( $T_{Reg}$ ) cells (left panel). By contrast, if DCs take up and present self antigens to T cells in the context of pro-inflammatory mediators (such as interleukin-6 (IL-6), IL-12 and IL-23), they can promote the development of self-reactive effector  $CD4^+$  T cells and cytotoxic T lymphocytes (CTLs). These self-reactive T cells might contribute to pathological autoimmune responses, such as experimental autoimmune encephalomyelitis (EAE) in mice, or systemic lupus erythematosus (SLE), psoriasis and type 1 diabetes (T1D) in patients. TCR, T cell receptor;  $T_{FH}$ , T follicular helper;  $T_H$ , T helper.



**Figure 2. The production of type I IFN by pDCs as a common mechanism of pathogenesis**  
 The production of type I interferon (IFN) by plasmacytoid dendritic cells (pDCs) is a common mechanism of pathogenesis in type 1 diabetes, systemic lupus erythematosus and psoriasis. Dying tissue cells release nucleic acids; these form large complexes with antimicrobial peptides, such as LL37 in humans and cathelin-related antimicrobial peptide (CAMP) in mice, and with endogenous DNA-binding proteins, such as high mobility group protein B1 (HMGB1). These DNA and RNA complexes activate pDCs via Toll-like receptor 9 (TLR9) and TLR7, respectively, and they induce the production of type I IFN. In turn, type I IFN promotes T cell activation, autoantibody production by B cells and the release of neutrophil extracellular traps (NETs) that consist of immune complexes that are preferentially endocytosed by pDCs via Fc receptors. Self-nucleic acids also activate classical DCs (cDCs) and they promote the release of inflammatory cytokines and the priming of T cells that are specific for self antigens in a process that is also facilitated by type I IFN<sup>82,83,95,110,111</sup>.

Table 1

Major subsets of DCs in humans and mice\*

| DC subset   | Phenotype   |   | Specific transcription factors | Specific mediators produced upon activation | Specific antigen-presentation capacities                                    | Location                         | Tissue condition |
|---|---|---|--------------------------------|---|---|----------------------------------|------------------|
|   | Mice  | Humans  |                                |   |   |                                  |                  |
| Plasmacytoid DC   | CD11 <sup>c</sup> <sup>int</sup> , MHC class II <sup>int</sup> , B220, BST2 and SIGLEC-H  | CD11 <sup>c</sup> MHC class II <sup>int</sup> , CD123, BDCA2 and BDCA4                                | TCF4 (also known as E2-2)      | Type 1 IFN                                  | Present and Cross-present peptides only after activation                    | Lymphoid organs                  | Steady state     |
| CD11b <sup>+</sup> ESAM <sup>hi</sup> cDC               | CD11 <sup>c</sup> <sup>hi</sup> , MHC class II <sup>hi</sup> , CD11b, CX <sub>3</sub> CR1 <sup>low</sup> and ESAM <sup>hi</sup> | CD11 <sup>c</sup> <sup>hi</sup> MHC class II <sup>hi</sup> , CD11b and BDCA1                          | Notch 2                        | ND  | Present peptides on MHC class II molecules to CD4 <sup>+</sup> T cells      | Lymphoid organs                  | Steady state     |
| CD11b <sup>+</sup> ESAM <sup>low</sup> cDC <sup>‡</sup> | CD11 <sup>c</sup> <sup>hi</sup> , MHC class II <sup>hi</sup> , CD11b, CX <sub>3</sub> CR1 <sup>hi</sup> and ESAM <sup>low</sup> | CD11 <sup>c</sup> <sup>hi</sup> , MHC class II <sup>hi</sup> , CD11b, CD16 and CD14                   | ND                             | TNF and IL-12                               | ND  | Lymphoid organs                  | Steady state     |
| CD8α <sup>+</sup> cDC                                   | CD11 <sup>c</sup> <sup>hi</sup> , MHC class II <sup>hi</sup> , XCR1, CLEC9A and CD8α  | CD11 <sup>c</sup> <sup>int</sup> MHC class II <sup>hi</sup> , XCR1, CLEC9A and BDCA3                  | BATF3                          | IL-12                                       | Cross-present peptides on MHC class I molecules to CD8 <sup>+</sup> T cells | Lymphoid organs                  | Steady state     |
| CD103 <sup>+</sup> cDC                                  | CD11 <sup>c</sup> <sup>hi</sup> , MHC class II <sup>hi</sup> , CD103, XCR1 and CLEC9A   | CD11 <sup>c</sup> <sup>hi</sup> MHC class II <sup>hi</sup> , XCR1, CLEC9A and BDCA3                   | BATF3                          | ND  | Cross-present peptides on MHC class I molecules to CD8 <sup>+</sup> T cells | Peripheral tissues               | Steady state     |
| CD11b <sup>+</sup> cDC                                  | CD11 <sup>c</sup> <sup>hi</sup> MHC class II <sup>hi</sup> , CD103 <sup>‡</sup> , CD11b and CD24                                | CD11 <sup>c</sup> <sup>hi</sup> , MHC class II <sup>hi</sup> , CD11b, SIRPα <sup>‡</sup> and BDCA1 // | IRF4 and Notch 2 <sup>‡</sup>  | IL-6 and IL-23                              | Present peptides on MHC class II molecules to CD4 <sup>+</sup> T cells      | Lungs and gut                    | Steady state     |
| Interstitial cDC  | CD11 <sup>c</sup> <sup>hi</sup> , MHC class II <sup>hi</sup> , X <sub>3</sub> CR1 and CD11b                                     | CD11 <sup>c</sup> <sup>hi</sup> , MHC class II <sup>hi</sup> , CD11b, CD16 and CD14                   | ND                             | TNF and IL-12                               | ND  | Peripheral tissues               | Steady state     |
| Langerhans cell   | CD11 <sup>c</sup> <sup>hi</sup> MHC class II <sup>hi</sup> , Langerin and CD205   | CD11 <sup>c</sup> <sup>hi</sup> MHC class II <sup>hi</sup> , CD1a, Langerin and CD205                 | ND                             | ND  | ND  | Epidermis                        | Steady state     |
| Monocyte-derived DC                                     | CD11 <sup>c</sup> <sup>hi</sup> , MHC class II <sup>hi</sup> , CD11b, CX <sub>3</sub> CR1 and CD209                             | CD11 <sup>c</sup> <sup>hi</sup> MHC class II <sup>hi</sup> , CD11b and CD11b                          | ND                             | TNF and iNOS                                | Present peptides on MHC class II molecules to CD4 <sup>+</sup> T cells      | Inflamed lymph nodes and tissues | Inflamed         |

BATF3, basic leucine zipper transcription factor ATF-like 3; BDCA, blood dendritic cell antigen; BST2, bone marrow stromal antigen 2; cDC, classical dendritic cell; CLEC9A, C-type lectin domain family 9 member A; CX<sub>3</sub>CR1, CX<sub>3</sub>C-chemokine receptor 1; ESAM, endothelial cell-selective adhesion molecule; IFN, interferon; IL-12, interleukin-12; iNOS, inducible nitric oxide synthase; IRF4, interferon-regulatory factor 4; ND, not described; SIGLEC, sialic acid-binding immunoglobulin-like lectin; SIRPα, signal-regulatory protein-α; TCF4, transcription factor 4; TNF, tumour necrosis factor; XCR1, XC-chemokine receptor 1.

\* The table (which is by no means all-inclusive) shows the main cell surface markers, subset-specific transcription factors and predominant functional features of the putative orthologous human and mouse dendritic cell subsets<sup>3-5,116,117</sup>.

‡ It is unclear whether the human and mouse subsets of these DCs are orthologous.

§ Expression has been reported in the gut.

// Expression has been reported in the lungs.

¶ The expression of Notch 2 is ubiquitous; however, its function is only required in this DC subset in the gut and not in the lungs.

**Table 2**

## Cell-intrinsic negative regulators of DC function in autoimmunity

| Protein deleted in DCs (gene targeted)*        | Disease phenotype  | Mechanism of action   | Refs  |
|--|--|---|-------|
| $\alpha$ V $\beta$ 8 integrin ( <i>Itgb8</i> ) | Polyclonal immune activation, production of autoantibodies and development of colitis  | DCs cannot activate latent TGF $\beta$ to induce regulatory T cell conversion   | 42    |
| STAT3 ( <i>Stat3</i> )                         | Cervical lymphadenopathy and ileocolitis   | DCs are unresponsive to inhibitory signals delivered by IL-10 and they show enhanced induction of T cell priming and T <sub>H</sub> 1 cell differentiation              | 43    |
| BLIMP1 ( <i>Prdm1</i> )                        | Increased germinal centre reaction, production of DNA-specific antibodies and development of SLE-like disease (females only) | Spontaneous production of IL-6 and the induction of T follicular helper cell differentiation by DCs   | 44    |
| A20 ( <i>Tnfrsf3</i> )                         | Polyclonal immune activation, ankylosing arthritis and colitis   | Spontaneous DC maturation, increased inflammatory cytokine production and enhanced T cell activation by DCs   | 45    |
|  | Polyclonal immune activation and development of SLE-like disease   | Reduced apoptosis and spontaneous maturation of DCs, increased inflammatory cytokine production by DCs, and increased uptake and presentation of apoptotic cells by DCs | 46    |
| SHP1 ( <i>Ptpn6</i> )                          | Polyclonal immune activation and development of SLE-like disease   | Spontaneous DC maturation, and increased inflammatory cytokine production and induction of T <sub>H</sub> 1 cell differentiation by DCs                                 | 47,48 |

BLIMP1, B lymphocyte-induced maturation protein 1; DC, dendritic cell; IL, interleukin; SHP1, SH2 domain-containing protein tyrosine phosphatase 1; SLE, systemic lupus erythematosus; STAT3, signal transducer and activator of transcription 3; TGF $\beta$ ; transforming growth factor- $\beta$ ; T<sub>H</sub>1, T helper 1.

\* Genes are shown that, when deleted in DCs using a DC-specific Cre-deleter mouse strain<sup>115</sup>, caused spontaneous inflammatory and/or autoimmune manifestations.