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ASAP: Assemble Species by Automatic Partitioning

Running title: Assemble Species by Automatic Partitioning

Nicolas Puillandre^{1,*}, Sophie Brouillet^{1,*}, Guillaume Achaz^{3,2,1}

¹ Institut Systématique Evolution Biodiversité (ISYEB), Muséum National d'Histoire Naturelle, CNRS, Sorbonne Université, EPHE, Université des Antilles. 57 rue Cuvier, CP 26, 75005 Paris, France.

² SMILE group, CIRB, UMR 7241, Collège de France, CNRS, INSERM, Paris, France.

³ Éco-anthropologie, Muséum National d'Histoire Naturelle, CNRS UMR 7206, Université de Paris, Paris, France

Corresponding author: N. Puillandre, puillandre@mnhn.fr.

*: equally contributed to the work.

23 **ABSTRACT**

24

25 We describe ASAP (Assemble Species by Automatic Partitioning), a new method to build
26 species partitions from single locus sequence alignments (*i.e.* barcode datasets). ASAP is
27 efficient enough to split datasets as large 10^4 sequences into putative species in several
28 minutes. Although grounded in evolutionary theory, ASAP is the implementation of a
29 hierarchical clustering algorithm that only uses pairwise genetic distances, avoiding the
30 computational burden of phylogenetic reconstruction. Importantly, ASAP proposes species
31 partitions ranked by a new scoring system that uses no biological prior insight of intra-
32 specific diversity. ASAP is a stand-alone program that can be used either through a graphical
33 web-interface or that can be downloaded and compiled for local usage. We have assessed its
34 power along with three others programs (ABGD, PTP and GMYC) on 10 real COI barcode
35 datasets representing various degrees of challenge (from small and easy cases to large and
36 complicated datasets). We also used Monte-Carlo simulations of a multi-species coalescent
37 framework to assess the strengths and weaknesses of ASAP and the other programs. Through
38 these analyses, we demonstrate that ASAP has the potential to become a major tool for
39 taxonomists as it proposes rapidly in a full graphical exploratory interface relevant species
40 hypothesis as a first step of the integrative taxonomy process.

41

42 **KEYWORDS**

43 ASAP, species delimitation, integrative taxonomy, ABGD, DNA barcoding.

44

45 INTRODUCTION

46

47 During the last 15 years, following the success of the DNA-barcoding projects and the
48 increase in sequencing capacities, many methods of species delimitation based on DNA
49 sequences have been developed. They can be roughly classified into two main categories. A
50 first one includes methods that compute the likelihood of competing partitions of species
51 hypotheses (“models”) in the so-called “multi-species coalescent” framework. In this
52 category, the most popular methods are SpedeSTEM (Ence & Carstens, 2011), BPP (Yang &
53 Rannala, 2014) and BFD (Leaché, Fujita, Minin, & Bouckaert, 2014), reviewed (with other
54 methods) in several articles (Camargo & Sites, 2013; Carstens, Pelletier, Reid, & Satler, 2013;
55 Fujita, Leaché, Burbrink, McGuire, & Moritz, 2012; Leavitt, Moreau, & Lumbsch, 2015;
56 Rannala, 2015). They were designed for multilocus data and are computationally (extremely)
57 demanding. As a consequence, they have been mainly applied to datasets with limited number
58 of sequences and species, and to well-studied groups, for which competing partitions of
59 species have been proposed in the literature; they generally correspond to species complexes,
60 typically in the grey zone (De Queiroz, 2005).

61 A second category of methods corresponds to exploratory ones, *i.e.* methods that propose *de*
62 *novo* species partitions, typically from a single-locus, DNA-barcoding-like, datasets.
63 Although sometimes criticized because a single gene tree poorly represents the species tree
64 (Degnan & Rosenberg, 2009; Nichols, 2001), these methods are widely used, as they are easy
65 to apply on DNA-barcoding datasets, even large, and precisely because they do not
66 necessitate pre-defined species hypotheses. The most popular ones are GMYC –General
67 Mixed Yule-Coalescent model– (Pons et al., 2006), PTP –Poisson Tree Process– (Zhang,
68 Kapli, Pavlidis, & Stamatakis, 2013), both first developed in a maximum likelihood

69 framework, and later extended to a Bayesian framework (Reid & Carstens, 2012), and ABGD
70 –Automatic Barcode Gap Discovery– (Puillandre, Lambert, Brouillet, & Achaz, 2012).
71 GMYC and PTP take as input a phylogenetic tree and estimate rates of branching events to
72 infer which part of the tree more likely follows a speciation model (the deepest part) and
73 which part follows a coalescent model (subtrees of the shallowest part). The species partition
74 is found by maximizing the likelihood of the transition between these two branching rates,
75 GMYC in absolute time (hence the need for an ultrametric tree), PTP in mutational time at
76 different nodes of the tree. GMYC and PTP first inferred a single transition event between the
77 two rates (speciation vs coalescent) and were later expanded to infer “multiple thresholds”,
78 allowing several transitions to occur in different subtrees (Kapli et al., 2017; Monaghan et al.,
79 2009).

80 Contrary to the two previous methods, ABGD uses only pairwise genetic distances (no tree is
81 inferred) and automatically identifies in their distribution the so-called “barcode gap”. This
82 gap marks the limit between the smaller intra-specific distances and the larger inter-specific
83 distances. From the gap, a distance threshold is estimated and used to partition the samples
84 into putative species. A coalescent model is used to identify the position of the most likely
85 barcode gap, based on a maximal genetic intraspecific divergence P defined *a priori* by the
86 user. Consequently, users must provide a range of P in which ABGD identifies one or several
87 barcode gaps and the method outputs the corresponding species partitions. For a single
88 dataset, ABGD thus eventually proposes several partitions that correspond to different prior
89 values P . In its recursive version, ABGD is applied on each group of the initial partition, and
90 eventually splits them when internal barcode gaps are detected.

91 The relative performances of these three exploratory methods, GMYC, PTP and ABGD,
92 sometimes together with less used methods (Flot, Coulloux, & Tillier, 2010; Ratnasingham &

93 Hebert, 2013) have been compared in various taxa: mammals (Derouiche, Vercammen,
94 Bouhadad, & Fernandes, 2017), amphibians (Vacher et al., 2017), squamates (Blair &
95 Bryson, 2017), fishes (Ramirez et al., 2017), echinoderms (Boissin, Hoareau, Paulay, &
96 Bruggemann, 2017), insects (Lin, Stur, & Ekrem, 2015), spiders (Ortiz & Francke, 2016),
97 crustaceans (Larson, Castelin, Williams, Olden, & Abbott, 2016), pycnogonids (Dömel,
98 Melzer, Harder, Mahon, & Leese, 2017), rotifers (Papakostas et al., 2016), annelids (Decaëns
99 et al., 2016), molluscs (Fourdrilis et al., 2016), flatworms (Scarpa et al., 2017), nemerts (Leasi
100 & Norenburg, 2014), cnidarians (Arrigoni et al., 2016), plants (Lithanatudom et al., 2017),
101 algae (Zou et al., 2016), lichens (Pino-Bodas, Burgaz, Teuvo, & Stenroos, 2018), fungi
102 (Alors, Lumbsch, Divakar, Leavitt, & Crespo, 2016) and foraminifera (André et al., 2014).

103 Although the results obtained with the various methods often vary depending on dataset
104 characteristics (e.g. Blair & Bryson, 2017), the main conclusions of these studies are:

- 105 1. all methods generally perform well (but see e.g. Dellicour & Flot, 2018) being mostly
106 congruent (*i.e.* providing similar species partitions) with each other and with the
107 species partitions inferred from independent data (e.g. other molecular markers,
108 morphological data, ecological data);
- 109 2. all of them perform poorly when the number of sampled individuals per species is too
110 low (Ahrens et al., 2016), or when the contrast of intra- vs. interspecific divergences is
111 mild. This contrast varies with species ages, mutation rates, population sizes, strengths
112 of the selection and degrees of within-species population structure (Pante et al., 2015;
113 Pentinsaari, Vos, & Mutanen, 2016; Ritchie, Lo, & Ho, 2016); mPTP was in particular
114 developed to overcome this issue (Kapli et al., 2017);
- 115 3. partitions proposed by the three methods sometimes differ, each of them being able to
116 infer the “correct” species when the two others fail. This led some authors to propose

117 that all three methods (among with eventually others) should be applied jointly and
118 compared (Ducasse, Ung, Lecointre, & Miralles, 2020);

119 4. Although there are several exceptions (e.g. Blair & Bryson, 2017), ABGD in
120 particular, and PTP to a lesser extent, tend to lump species more than GMYC
121 (Pentinsaari et al., 2016). Conversely, the multiple-threshold version of GMYC is
122 particularly prone to oversplitting (Fujisawa & Barraclough, 2013; Kekkonen &
123 Hebert, 2014).

124 In comparison with GMYC and PTP, ABGD has the advantage of being very fast, mainly
125 because it bypasses the phylogenetic reconstruction. Furthermore, because ABGD identifies a
126 species partition for each value of P defined a priori, several partitions may be proposed,
127 reflecting the uncertainty stemming from the data and encouraging the user to evaluate the
128 relevance of the ABGD partitions in the light of other data, as it is recommended in an
129 “integrative taxonomy” approach. However, ABGD does not provide a score for each
130 partition that would help the user to identify the “best” partition(s), and this probably
131 constitutes the main drawback of ABGD (judging from the numerous comments and
132 questions the authors of ABGD have received from the users).

133 In this article, we describe a new method of species delimitation, still based on pairwise
134 genetic distances, but which implementation provides a score for each defined partition and
135 overcomes the challenge of *a priori* defining P . Our new algorithm, ASAP (Assemble Species
136 by Automatic Partitioning), still provides several partitions, more or less fine-grained, but
137 ranked using a new scoring system. Importantly, we also develop a full graphical web-
138 interface to ease its usage. However, ASAP, like any other method, must not replace the
139 taxonomist work, as any partition of species must be subsequently tested against other

140 evidences in an integrative taxonomy framework. This is especially crucial as ASAP uses
141 single-locus data that are known to bear weaknesses.

142

143 **MATERIAL AND METHODS**

144

145 **Overview of the ASAP software**

146 ASAP is a C self-contained program. Users can use ASAP either through a full graphical
147 web-interface (<https://bioinfo.mnhn.fr/abi/public/asap>), or download and compile the sources
148 for local usage (same url).

149 Our algorithm is an ascending hierarchical clustering, merging sequences into *groups* that are
150 successively further merged until all sequences form a single group. At each merging step, the
151 assignment of all sequences into groups is named a *partition*. The first partition contains as
152 many groups as sequences (no grouping was yet done) whereas the last partition is a single
153 group with all sequences inside. Larger groups are created by merging groups of the previous
154 partition together. We characterize all newly created partition in two complementary ways.
155 First, we assign to it a probability that quantifies the chances that each of its new groups is a
156 single species. Second, we compute the width of the barcode gap between the previous and
157 this new partition. Both metrics (probability and barcode gap width) are combined into a
158 single *asap-score* that is used to rank the partitions.

159

160 **ASAP in details**

161 i) Ranked distances

162 We first start by computing, when not provided, all pairwise distances between the n
163 sequences of the alignment. Distances are then ranked by increasing values. The efficiency of

164 the algorithm stems from the fact that each distance is only considered once in increasing
165 order for clustering purposes.

166 ii) Hierarchical clustering

167 The clustering process starts with a first partition where each sequence belongs to a different
168 group. ASAP then treats each of the ranked distances one by one in increasing order (equal
169 distances are treated together) as a threshold value for delimiting groups: sequences separated
170 by a distance equal to the current value d_C are clustered into the same group. Consequently,
171 when sequences that were in different groups are clustered together, the previous groups are
172 merged into a new larger group, and is associated to the current clustering distance, d_C .
173 Importantly, a new partition can have a single new group or several new ones when several
174 sequences from different groups are merged independently into different groups for the same
175 distance d_C . When a new partition is built, the clustering process pauses. ASAP then scores
176 all new groups with a probability of panmixia. It also scores the new partition using an ad-hoc
177 score computed from both the barcode gap width and probabilities of panmixia. After the
178 group(s) and partition scoring, ASAP then continues the clustering by looking after the next
179 distances until another partition is built. The algorithm stops when all sequences are merged
180 into a single final group.

181 iii) Computing p-values

182 **a. For each group:** we aim at computing a p-value for a newly created group that is a
183 merge of two or more subgroups. We compute Π_{intra} the average pairwise distance between
184 sequences within the subgroups and Π_{inter} the average pairwise distance among sequences of
185 different subgroups (Figure 1). We then compare Π_{intra} to its theoretical distribution,
186 computed by Monte-Carlo simulations of a neutral coalescent model assuming a single
187 panmictic species with a sample size m and a coalescent mutation rate $\theta = \Pi_{\text{inter}} / [2 \times (1 - 1/m)]$.

188 The value of θ is set so that in the simulations the distance between sequences connected by
189 the Most Recent Common Ancestor (MRCA) of the group (π_{inter}) is equal, on average, to the
190 observed one: $E[\pi_{\text{inter}}]=\Pi_{\text{inter}}$. This relates to the average time to the MRCA that is $2 \times (1-1/m)$,
191 expressed in coalescent time (Wakeley, 2009). We compute the p-value as the fraction of
192 replicates where the simulated π_{intra} is equal or lower than the observed Π_{intra} . The number of
193 replicates is updated on the fly to have correct estimations of low p-values. Put differently, it
194 quantifies under H_0 (one single species) the probability of observing a diversity Π_{intra} or less
195 within the subgroups given that the divergence between the subgroups is on average Π_{inter} .

196 **b. For partitions:** we compute the probability to observe π_{intra} or less diversity within
197 all subgroups of the *current* partition (that are groups of *previous* partition before the merge)
198 assuming that all new groups of the *current* partition are independent coalescent realizations
199 with θ estimated for each group independently.

200 iv) Recursive splits

201 Once a new partition is built, ASAP tests for each of the groups of the partition whether its p-
202 value is lower than a given risk (by default 1%) and consequently should be split. When a
203 group is split, ASAP recursively descends to all its subgroups and assesses whether they
204 should be split as well.

205 v) Relative barcode gap width

206 ASAP also computes a relative barcode gap width associated to the current partition
207 (Supplementary Material 1). The partition is associated to a threshold distance d_T that is the
208 mid-point between the current distance, d_C (with rank r_C), that triggered the merging and the
209 previous distance in the list d_{C-1} (with rank r_C-1). A barcode gap corresponds to a “jump” in
210 the distance values in only few ranks. While increasing only few ranks in the list, the distance
211 will “jump” from a value that is (much) less than d_T to a value that is (much) higher than d_T .

212 To quantify the barcode gap width, ASAP scans downward the distance list from d_{C-1} until it
213 finds the first distance smaller than $0.9d_{C-1}$: this is d_L which rank is r_L in the list. It then scans
214 from d_C the distance list upward until it finds the first distance above $1.1d_C$: this is d_H which
215 rank is r_H . The relative gap width W is defined as:

$$216 \quad W = [(d_H - d_L) / (d_H + d_L + 1)] / (r_H - r_L).$$

217 We normalized the difference of distance ($d_H - d_L$) by ($d_H + d_L + 1$) to compute the “relative”
218 width of the gap; the “+1” only prevents the ratio to be very high when distance values are
219 very small. The higher the W , the larger the barcode gap.

220 vi) Outputs

221 At the end of the clustering, ASAP scores and sorts all the different partitions using two
222 criteria: their p-value sorted (see iii.b) by increasing order (the smallest p-value has rank 1)
223 and their rank of relative barcode gap width (see v) sorted by decreasing order (the largest gap
224 has rank 1). The *asap-score* is the average of both ranks: the smaller, the better. Furthermore,
225 ASAP produces a graphical output where each node of the hierarchical clustering is color-
226 coded depending on its probability of being a panmictic species (see iii.a). Thus, the color
227 guides the user finding which nodes may be split into smaller groups. Several other graphical
228 options are provided to help the user navigate among partitions and choose the “most
229 relevant” partition, beyond a simple naive use of the *asap-score* (Supplementary Material 2).

230

231 **Tests on empirical data**

232 To compare the results obtained by four methods (ASAP, (m)PTP, (m)GMYC and ABGD),
233 we selected 10 empirical COI datasets covering various taxa (birds, mammals, amphibians,
234 insects, crustaceans and molluscs) and including 44 to 2,574 specimens that belong to 5 to
235 643 species (Table 1) (Borisenko, Lim, Ivanova, Hanner, & Hebert, 2008; Elias-Gutierrez,

236 Jeronimo, Ivanova, Valdez-Moreno, & Hebert, 2008; Hajibabaei, Janzen, Burns, Hallwachs,
237 & Hebert, 2006; Kerr et al., 2007; Puillandre, Cruaud, & Kantor, 2010; Puillandre, Baylac,
238 Boisselier-Dubayle, Cruaud, & Samadi, 2009; Puillandre, Fedosov, Zaharias, Aznar-
239 Cormano, & Kantor, 2017; Puillandre et al., 2011, 2012; Smith, Poyarkov Jr., & Hebert,
240 2008). Among them, five correspond to datasets published by one of the authors to facilitate
241 the interpretations of the results. An eleventh dataset, including 9,396 sequences of moths
242 (publicly available from BOLD), was used to estimate and compare the computation times of
243 ABGD and ASAP. A dataset of this size could not be analyzed by (m)GMYC or (m)PTP as
244 the phylogenetic reconstruction is too costly.

245 For all empirical datasets, we used the web version of ABGD, with default parameters. Only
246 the initial partitions were considered, and only the more stable partition(s) (i.e. the partition(s)
247 found with several P in the vicinity of the barcode gap) was (were) reported. For ASAP, we
248 used a recursive split probability of 0.01 (see iv), and report a) the partition with the best
249 *asap-score* as well as b) the partition that is closest to the “correct” one among the two best
250 partitions, according to their *asap-scores*. For GMYC and mGMYC, ultrametric trees were
251 reconstructed using BEAST 2 (Bouckaert et al., 2014), with an independent GTR substitution
252 model for each codon position. Relative divergence times were estimated using a relaxed log-
253 normal clock with a coalescent prior and a constant population size, following the
254 recommendations of Monaghan et al. (2009). The number of MCMC steps were 20M
255 (*Gemmuloborsonia*, *Benthomangelia*, *Lophiotoma* and *Eumunida* datasets), 100M
256 (Amphibians, Cladocera, Mammals, Sphingidae and Turridae datasets) and 200M (Birds
257 dataset), sampled every 2,000, 10,000 and 20,000 steps respectively. Convergence of the runs
258 was assessed using TRACER 1.6 (Rambaut & Drummond, 2014) to check that all effective

259 sample size values exceeded 200. Consensus trees were calculated after discarding the first
260 25% of the trees as burn-in, with the option “Common Ancestry” for node height.
261 For PTP and mPTP, the web server at <https://mptp.h-its.org/#/tree> was used, with default
262 parameters. The input tree was obtained with RAxML (Stamatakis, 2006), with an
263 independent GTR substitution model for each codon position. All phylogenetic analyses were
264 performed on the Cipres Science Gateway (<http://www.phylo.org/portal2>), using the BEAST2
265 on XSEDE (2.1 - 2.4.8) and RAxML-HPC2 on XSEDE (8.2.10) tools.

266

267 **Simulations**

268 We measured the power of ABGD, GMYC, (m)PTP and ASAP to retrieve the correct species
269 partition in various scenarios using Monte Carlo simulations. We used a “multispecies
270 coalescent” framework (Rannala & Yang, 2003) with different options and parameters using
271 Monte-Carlo simulations, as described previously (Puillandre et al., 2012). Note that
272 contrarily to the standard multispecies coalescent, the species tree is here drawn from a
273 probability distribution. The home-made C simulator is available upon request.

274 Briefly, for each replicate, we generate a species tree using either a Yule model (all lineages
275 have the same birth rate) or a radiation model (all species arose at the same time). Radiation
276 (hard polytomy) models cases where all speciation events follow each other quickly and
277 where no mutations have occurred between the first (the root) and the last speciation event.
278 We used a backward coalescent version of these models that we have previously used for
279 ABGD evaluation (Puillandre et al., 2012). For the radiation model a unique speciation event,
280 exponentially distributed with rate r , is drawn. For the Yule model $(n_{sp}-1)$ speciation events
281 are drawn with identical rate (Lambert & Stadler, 2013).

282 Once the species tree is obtained, we assign sequences to species uniformly, with at least 1
283 sequence per species. All species (current and ancestral) are assumed to be of equal effective
284 size (*i.e.* N individuals). The genealogy of the sequences is then simulated in backward time
285 using a standard Kingman coalescent process but forbidding coalescent events between
286 lineages from different species. Once the genealogy is obtained, a Poisson random number of
287 mutations – with mean $L\theta/2$, where L is the total tree length and θ the population mutation
288 rate – are distributed uniformly on the tree and the resulting polymorphic sites are generated.
289 The whole simulation process is tuned by 4 parameters:

- 290 - a total number of sequences n ,
- 291 - a number of species n_{sp} with one or more sequences,
- 292 - a speciation rate r , expressed in coalescent time (*i.e.* in N generations),
- 293 - a mutation rate θ , expressed in coalescent scale ($\theta = 2 N \mu$), set to $\theta=10$ for 600bp
294 of simulated sequence. Mutations are only substitutions following a Jukes-Cantor
295 model.

296 ABGD and ASAP use the pairwise distance matrix as input. For ABGD, we used a prior
297 value of 0.083 ($5 \times 10 / 600$) that is an excellent prior representing a situation where the user has
298 near perfect knowledge on maximal diversity within species. For GMYC and (m)PTP, we
299 used as input the ‘true’ gene genealogy (the one simulated for the replicates) not only to
300 fasten the simulation (*i.e.* skipping the phylogenetic reconstruction) but also to assess their
301 power when the phylogeny is perfectly reconstructed. We would like to emphasize that only
302 ASAP used unprocessed data (polymorphic sites) without any biological insights (no prior, no
303 phylogeny reconstruction nor calibration).

304

305 RESULTS

306

307 **Empirical datasets**

308 We first assessed the ability of ASAP through a proxy that is its ability to retrieve the
309 “correct” number of species in 10 empirical datasets (Table 1). The datasets were selected to
310 represent test cases of different sizes (from 44 sequences/5 species to 2,574 sequences/643
311 species). We first report the number of species predicted in the partition with the best *asap-*
312 *score* (ASAP 1st): we found that in 4/10 of the datasets, the partition with the best *asap-score*
313 is *very close* to the reference one (less than 5% difference in terms of species numbers) and
314 that 8/10 is *close* (less than 10% difference). If we also consider the partition with the second
315 best *asap-score* (ASAP 1st and 2nd), the degree of accuracy increases to 6/10 for the *very close*
316 ones and 9/10 for the *close* ones. This is a good indication that ASAP users should consider
317 not only the partition with the best *asap-score* but also few subsequent ones. It is important to
318 report that here no extra biological knowledge was considered for ASAP predictions. One
319 could for example use threshold distances (e.g. d_T or d_C) to prefer one partition over another
320 despite a poorer *asap-score* (e.g. in most clades intra-specific diversity is typically on the
321 order of 1%, not on the order of 10%). Obviously, other criteria and characters should also be
322 used to choose a final species partition, in an integrative taxonomy context.

323 One of the ASAP main qualities is that it is extremely fast compared to any method that relies
324 on tree reconstruction. The online version takes 45 seconds for the largest dataset of Table 1
325 (2,574 aligned sequences; 643 species) for all steps of the complete method: mainly creating
326 the distance matrix, performing the clustering and computing probabilities by Monte Carlo at
327 each node. We observed that the CPU time increases linearly with the number of species in
328 the datasets (Figure 2) and only to a lesser extent with the number of sequences (data not
329 shown). We estimate the CPU cost at 0.07 sec per species for the current web version. This

330 suggests that most of the CPU time is taken by probability estimations of significant nodes
331 (see method, section iii) (non-significant ones are not as costly in our implementation as we
332 increase the number of replicates only for nodes with low probabilities). The number of
333 significant nodes likely increases approximately linearly with the number of species. The time
334 for distance matrix computation and clustering both increase quadratically with the number of
335 sequences and are independent from the number of species.

336 On a curated unpublished moth dataset, it took 6 min 35 on the website to delimit 2,466
337 species (best *asap-score*) or 2,067 (second best *asap-score*) from 9,396 sequences.
338 Subsequent partitions with lower *asap-scores* are close to one or the other of these two first
339 partitions. Because of its rapidity, ASAP web server accepts up to 10^4 sequences (unlike the
340 ABGD server).

341 We also took the opportunity of analyzing the 10 datasets to assess the performance of other
342 methods: ABGD which is solely based on pairwise distances, PTP and mPTP that were run on
343 an ML trees (*i.e.* RaxML) and GMYC and mGMYC on an ultrametric trees estimated by a
344 Bayesian MCMC method (*i.e.* BEAST). Results (Table 1) show that ABGD performance is
345 similar to ASAP 1st-2nd, that PTP and mPTP tend to not perform very well, that GMYC
346 performs very well provided that the number of species is not too large and that, as previously
347 reported in the literature, mGMYC generally oversplits (Fujisawa & Barraclough, 2013;
348 Kekkonen & Hebert, 2014). Note that ABGD performances are somehow overestimated as
349 we report the partition that is the closest to the reference one over the whole range of P . We
350 could not use GMYC for the largest dataset as the Bayesian tree reconstruction did not
351 converge after several weeks of computation.

352

353 **Simulated datasets**

354 We then assess the theoretical performance of ASAP using Monte-Carlo simulations of a
355 multispecies coalescent framework. In brief, a random species tree is generated using either a
356 Radiation model, where all species arose in single event, or a Yule model, where the
357 speciation events occur at constant rate independently in all branches. In both model, we tune
358 the separation of time scales (speciation versus intra-specific coalescent events) using a
359 speciation rate that is expressed in coalescent time (*i.e.* N generations per unit of time). The
360 lower the speciation rate, the better the separation of time scales. For example, when the
361 speciation rate is 0.1, speciation events are 10 times slower than pairwise coalescent events
362 within species.

363

364 **The impact of speciation rate on ASAP**

365 We first examine the ability of ASAP to correctly retrieve four species in both speciation
366 models as a function of the speciation rate (from 0.001 to 1). We report in Figure 3 the
367 fraction of runs where ASAP was able to correctly retrieve the four species (top panel) and
368 the average number of predicted species, regardless of their composition (bottom panel). We
369 assess the quality of the partition with the best *asap-score* (ASAP 1st) as well as the quality of
370 the partition that is the closest to the truth among the two best partitions (ASAP 1st-2nd).

371 We observe that for low rates of speciation, the best partition proposed by ASAP correspond
372 exactly to the four species. This is an “easy” case where the two time scales are well
373 separated. As the speciation rate increases, both time scales overlap and it becomes harder to
374 delineate species using pairwise genetic differences at a single locus. When the speciation rate
375 is larger than 1, speciation events are more recent than intra-specific divergence so that
376 individuals within species are no more different than individuals between species.

377 ASAP performs usually better with the Radiation than with the Yule model. This is especially
378 striking for moderate speciation rate (e.g. 0.03). For radiations, most of the errors correspond
379 to oversplit, as illustrated by the average number of predicted species that is larger than four.
380 Under the Yule model with four species, there are three independent speciation events and
381 consequently there is a higher chance to generate at least one very recent speciation event that
382 would be invisible in regard of sequence divergence. Indeed, the most recent event is
383 exponentially distributed with rate $3r$. As a consequence, contrarily to the radiation model,
384 ASAP failures correspond for this rate to cases where it lumps the two closest species into a
385 single one.

386

387 **The impact of the number of species on ASAP**

388 Second, we explore the impact of the number of species for a fixed sample size of 200
389 sequences, with $r=0.01$, a moderately challenging speciation rate. We report the average
390 number of predicted species regardless of their composition for both the radiation and the
391 Yule models. Results (Figure 4) show a) that ASAP very well predicts the species under a
392 radiation model, regardless of the number of species and b) that it only finds a fraction of
393 them for the Yule model. Under the Yule model, the problem of finding a threshold between
394 intra- and inter-specific distance becomes harder as the most recent speciation event is
395 exponentially distributed with rate $r.(n_{sp}-1)$; the more species, the more recent the last
396 speciation event. Furthermore, the higher the number of species the higher the chance to have
397 a very old coalescent MRCA (Most Recent Common Ancestor) within one of the species.
398 This old MRCA translates into a high divergence among individuals of this species, which
399 would also obscure the threshold between intra- and inter-specific genetic divergences.

400

401 **The impact of the number of species on ABGD, PTP and GMYC**

402 We apply the same analysis to ABGD, (m)PTP and GMYC. We would like to emphasize
403 again that we assessed their power under optimal conditions: a single “excellent” prior for
404 ABGD representing a perfect knowledge of intraspecific diversity and the “true” simulated
405 tree for (m)PTP and GMYC, bypassing their main limitations, that is having a correctly
406 reconstructed phylogenetic tree. As a consequence, we here overestimate their power for
407 realistic biological situations where only a set of sequences is available (neither the true tree
408 nor prior knowledge of intraspecific diversity is known). ASAP, on the contrary, directly uses
409 the sequences and needs no prior biological insight or phylogenetic reconstruction.

410 The power assessments of the methods (Figure 4) show that ABGD retrieves well the correct
411 partition when speciation occur as a single radiation but has a limited power when speciations
412 follow a Yule model. On the contrary, we found that GMYC performs very well for the Yule
413 model but is less efficient for a radiation model. Interestingly mPTP consistently split a
414 constant small number of species. It thus performs poorly when the number of species is low
415 but quite well when the number of species is 50 or more.

416

417 **DISCUSSION**

418

419 We introduced a new species delimitation program, ASAP, fully exploratory, in the sense that
420 it does not require any *a priori* knowledge, neither on the number of species, the species
421 composition, or any biological information, such as a phylogenetic tree or *a priori*-defined
422 intraspecific genetic distances. Only pairwise genetic distances are used to build a list of
423 partitions ranked by a score. This composite score is computed using the probabilities of
424 groups to be panmictic species and the barcode gap widths. ASAP overcomes the two mains

425 limitations of ABGD, namely (i) the need for an *a priori* defined P and (ii) the lack of a
426 scoring system.

427 However, and contrary to some other methods, ASAP still outputs several partitions, ranked
428 by their *asap-scores*. A list of the “best” partitions (10 by default) is provided in the output
429 together with their gap-width score, their p-value, their threshold distance d_T and the number
430 of species they correspond to.

431 The graphical output of ASAP has four main components (Supplementary Material 2):

432 (1) a list of partitions ranked by their *asap-score* that putatively correspond to species
433 hypothesis,

434 (2) a plot of the *asap-score* as a function of d_C . We report the *asap-score* of all partitions
435 (not only the best ones) as a function of the clustering distance d_C to appreciate
436 whether all good partitions have similar d_C or whether “potentially good” partitions
437 can drastically differ in size.

438 (3) an ultrametric clustering tree of all sequences, where the distance to the leaves lengths
439 correspond to the distance d_C at which these sequences were clustered in the same
440 group. All nodes of this tree are color-coded depending on their p-value (the darker
441 the more it differs from a panmictic species).

442 (4) a “boxed-species” graph, where species hypotheses in the different partitions are
443 represented as vertical boxes in front of the ultrametric tree.

444 When a partition is selected by a click in any of the three panels, it is automatically
445 highlighted in the two other components.

446 We also propose a complementary representation, where we display the hierarchical tree with,
447 at its leaves, the 10 best ASAP partitions where their groups are depicted as boxes (that are
448 similar to the boxes of Figure 1).

449 We have evaluated ASAP strengths and weaknesses using both real and simulated data. Our
450 benchmark shows that ASAP performs well delivering partitions in a matter of minutes even
451 for datasets as large as 10^4 sequences. ASAP is thus meant to be applied on large single-locus
452 datasets when no species hypothesis is available, as typically produced in DNA-barcoding
453 projects. Although the web version limits the input to 10^4 sequences, more sequences can be
454 analyzed using a local command-line version of ASAP (sources are available on the
455 webservice).

456 The comparison with the other programs shows that ASAP and ABGD both perform well for
457 a Radiation model, because there are no “recent” invisible speciation events. Indeed, both
458 methods use a phenetic approach where similar sequences are simply clustered in the same
459 group/species. On the contrary, (m)GMYC and (m)PTP that are explicitly based on a
460 phylogenetic approach behave differently, performing quite well under a Yule model. More
461 generally, (m)GMYC and (m)PTP are both relying on a different property to propose species
462 hypotheses, compared to ABGD and ASAP: specimens belonging to the same species, *i.e.* to
463 the same diverging lineage, share a common evolutionary history, *i.e.* they form a clade.
464 Indeed, phenetic differences are calculated by simply counting the differences among
465 sequences, whereas the phylogenetic criterion requires the reconstruction of a proper
466 phylogenetic tree. This additional step in the (m)GMYC and (m)PTP methods potentially
467 introduces a bias, because a) phylogenetic trees reconstructed on a single locus may differ
468 drastically from the species tree, and b) the limited number of sites in a single marker may
469 lead to incorrectly reconstructed trees. Consequently, (m)GMYC and (m)PTP have been
470 shown to be sensitive to the reconstruction method (Tang, Humphreys, Fontaneto, &
471 Barraclough, 2014). On the contrary, it could be argued that relying only on genetic distances,
472 *i.e.* without testing if these differences actually correspond to distinct evolutionary histories,

473 and not to homoplasy, must be used with caution. Indeed, the efficiency of each method in
474 delimiting species probably depends on various characteristics of the species and datasets
475 (number of samples, number of species, population sizes...), and applying several methods to
476 a given dataset is a strategy commonly applied to maximize the probability to detect species
477 complexes, identified as groups of species whose limits vary depending on the method.

478 Importantly, several other methods can also be used to delimit species, such as BINs
479 (Ratnasingham & Hebert, 2013), Jmotu (Jones, Ghoorah, & Blaxter, 2011) or VSEARCH
480 (Rognes, Flouri, Nichols, Quince, & Mahé, 2016), among others (e.g. Rannala & Yang,
481 2020). We are also aware that the number of predicted species is only a proxy to assess the
482 performance of the different methods. Indeed, other metrics such as the F-measure (Larsen &
483 Aone, 1999) or the number of splits or merges (Ratnasingham & Hebert, 2013) give also
484 insightful information. Some of them are even implemented in meta-analysis software such as
485 LIMES (Ducasse, Ung, Lecointre, & Miralles, 2020), which could be used to perform a more
486 extensive benchmark of all existing methods using a wider spectrum of metrics.

487 More generally, and as advocated by the proponents of the integrative approach in taxonomy,
488 the use of a single marker with a single method of species delimitation should be avoided,
489 precisely because each method has its own limitations. Some methods are based on a phenetic
490 criteria (e.g. ASAP and ABGD) while others on phylogenetic criteria (e.g. (m)GMYC and
491 (m)PTP). Furthermore a single locus may not follow the species history, because of
492 introgression and incomplete lineage sorting. This is particularly true for species in the grey
493 zone, in which the gene tree may differ from the species tree, and the coalescent events may
494 be older than the speciation events (De Queiroz, 2005). For this reason, we recommend that
495 single-locus methods are to be used as a first step of the species delimitation process that is to
496 propose primary species *hypotheses*. This is for example useful in groups for which there is

497 no pre-existing hypotheses to test, or for which unknown/incorrectly delimited species
498 represent the majority of the diversity (e.g. microbial communities or hyperdiverse groups of
499 eukaryotes, such as insects, spiders, nematodes, mollusks...). Furthermore, DNA barcodes are
500 now routinely produced using NGS approaches, providing large numbers of sequences often
501 not assignable to known and sequenced species (Kennedy et al., 2020), and for which
502 methods such as ASAP are welcome to e.g. compare species diversity among sites.

503 In a second step it is then the responsibility of the taxonomist to evaluate with other methods
504 (in particular, methods that will evaluate alternative partitions of species) and/or lines of
505 evidence (such as other genetic markers, morphology or ecology) whether the proposed
506 hypotheses are robust, or not. In this context, methods such as ASAP, ABGD, (m)PTP and
507 (m)GMYC should thus be seen as a formalized and reproducible way to propose species
508 hypotheses in groups where no such hypotheses exist, or, if they do exist, that are better to be
509 ignored.

510

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516

517 **REFERENCES**

518 Ahrens, D., Fujisawa, T., Krammer, H.-J., Eberle, J., Fabrizi, S., & Vogler, A. P. (2016). Rarity and
519 incomplete sampling in DNA-based species delimitation. *Systematic Biology*, 65(3), 478–494.

520 Alors, D., Lumbsch, H. T., Divakar, P. K., Leavitt, S. D., & Crespo, A. (2016). An integrative approach for
521 understanding diversity in the *Punctelia rudecta* species complex (Parmeliaceae,
522 Ascomycota). *PLoS One*, *11*(2), e0146537.

523 André, A., Quillévéré, F., Morard, R., Ujiie, Y., Escarguel, G., De Vargas, C., ... Douady, C. J. (2014). SSU
524 rDNA divergence in planktonic foraminifera: molecular taxonomy and biogeographic
525 implications. *PLoS One*, *9*(8), e104641.

526 Arrigoni, R., Berumen, M. L., Chen, C. A., Terraneo, T. I., Baird, A. H., Payri, C., & Benzoni, F. (2016).
527 Species delimitation in the reef coral genera *Echinophyllia* and *Oxypora* (Scleractinia,
528 Lobophylliidae) with a description of two new species. *Molecular Phylogenetics and*
529 *Evolution*, *105*, 146–159.

530 Blair, C., & Bryson, R. W. (2017). Cryptic diversity and discordance in single-locus species delimitation
531 methods within horned lizards (Phrynosomatidae: *Phrynosoma*). *Molecular Ecology*
532 *Resources*, *17*(6), 1168–1182.

533 Boissin, E., Hoareau, T. B., Paulay, G., & Bruggemann, J. H. (2017). DNA barcoding of reef brittle stars
534 (Ophiuroidea, Echinodermata) from the southwestern Indian Ocean evolutionary hot spot of
535 biodiversity. *Ecology and Evolution*, *7*(24), 11197–11203.

536 Borisenko, A. V., Lim, B. K., Ivanova, N. V., Hanner, R. H., & Hebert, P. D. N. (2008). DNA barcoding in
537 surveys of small mammal communities: a field study in Suriname. *Molecular Ecology*
538 *Resources*, *8*, 471–479.

539 Bouckaert, R., Heled, J., Kühnert, D., Vaughan, T., Wu, C.-H., Xie, D., ... Drummond, A. J. (2014). BEAST
540 2: a software platform for Bayesian evolutionary analysis. *PLoS Computational Biology*, *10*(4),
541 e1003537.

542 Camargo, A., & Sites, J. Jr. (2013). Species delimitation: a decade after the renaissance. In I. ISBN:
543 978-953-51-0957-0 DOI: 10. 5772/52664 (Ed.), *The Species Problem - Ongoing Issues, book*

544 edited by Igor Ya. Pavlinov, ISBN 978-953-51-0957-0, Published: February 6, 2013 under CC BY
545 3.0 license. Pavlinov, I. Y.

546 Carstens, B. C., Pelletier, T. A., Reid, N. M., & Satler, J. D. (2013). How to fail at species delimitation.
547 *Molecular Ecology*, in press.

548 De Queiroz, K. (2005). A unified concept of species and its consequences for the future of taxonomy.
549 *Proceedings of the California Academy of Sciences*, 56, 196–215.

550 Decaëns, T., Porco, D., James, S. W., Brown, G. G., Chassany, V., Dubs, F., ... Rossi, J.-P. (2016). DNA
551 barcoding reveals diversity patterns of earthworm communities in remote tropical forests of
552 French Guiana. *Soil Biology and Biochemistry*, 92, 171–183.

553 Degnan, J. H., & Rosenberg, N. A. (2009). Gene tree discordance, phylogenetic inference and the
554 multispecies coalescent. *Trends in Ecology and Evolution*, 24(6), 332–340.

555 Dellicour, S., & Flot, J. (2018). The hitchhiker’s guide to single-locus species delimitation. *Molecular*
556 *Ecology Resources*.

557 Derouiche, L., Vercammen, P., Bouhadad, R., & Fernandes, C. (2017). Genetic evidence supporting
558 the taxonomic separation of the Arabian and Northwest African subspecies of the desert
559 hedgehog (*Paraechinus aethiopicus*). *Gene*, 620, 54–65. doi: 10.1016/j.gene.2017.04.009

560 Dömel, J. S., Melzer, R. R., Harder, A. M., Mahon, A. R., & Leese, F. (2017). Nuclear and Mitochondrial
561 Gene Data Support Recent Radiation within the Sea Spider Species Complex *Pallenopsis*
562 *patagonica*. *Frontiers in Ecology and Evolution*, 4, 139.

563 Ducasse, J., Ung, V., Lecointre, G., & Miralles, A. (2020). LIMES: a tool for comparing species partition.
564 *Bioinformatics*, 36(7), 2282–2283.

565 Elias-Gutierrez, M., Jeronimo, F. M., Ivanova, N. V., Valdez-Moreno, M., & Hebert, P. D. N. (2008).
566 DNA barcodes for Cladocera and Copepoda from Mexico and Guatemala, highlights and new
567 discoveries. *Zootaxa*, 1839, 1–42.

568 Ence, D. D., & Carstens, B. C. (2011). SpedeSTEM: a rapid and accurate method for species
569 delimitation. *Molecular Ecology Resources*, *11*, 473–480.

570 Flot, J.-F., Couloux, A., & Tillier, S. (2010). Haplowebs as a graphical tool for delimiting species: a
571 revival of Doyle's "field for recombination" approach and its application to the coral genus
572 *Pocillopora* in Clipperton. *BMC Evolutionary Biology*, *10*(1), 1.

573 Fourdrilis, S., Mardulyn, P., Hardy, O. J., Jordaens, K., de Frias Martins, A. M., & Backeljau, T. (2016).
574 Mitochondrial DNA hyperdiversity and its potential causes in the marine periwinkle
575 *Melarhaphe neritoides* (Mollusca: Gastropoda). *PeerJ*, *4*, e2549.

576 Fujisawa, T., & Barraclough, T. G. (2013). Delimiting Species Using Single-locus Data and the
577 Generalized Mixed Yule Coalescent (GMYC) Approach: A Revised Method and Evaluation on
578 Simulated Datasets. *Systematic Biology*, *62*, 707–724.

579 Fujita, M. K., Leaché, A. D., Burbrink, F. T., McGuire, J. A., & Moritz, C. (2012). Coalescent-based
580 species delimitation in an integrative taxonomy. *Trends in Ecology and Evolution*, *27*, 480–
581 488.

582 Hajibabaei, M., Janzen, D. H., Burns, J. M., Hallwachs, W., & Hebert, P. D. N. (2006). DNA barcodes
583 distinguish species of tropical Lepidoptera. *Proceedings of the National Academy of Sciences*,
584 *103*(4), 968–971.

585 Jones, M., Ghoorah, A., & Blaxter, M. (2011). jMOTU and Taxonator: turning DNA barcode
586 sequences into annotated operational taxonomic units. *PLoS ONE*, *6*, e19259.

587 Kapli, P., Lutteropp, S., Zhang, J., Kobert, K., Pavlidis, P., Stamatakis, A., & Flouri, T. (2017). Multi-rate
588 Poisson tree processes for single-locus species delimitation under maximum likelihood and
589 Markov chain Monte Carlo. *Bioinformatics*, *33*(11), 1630–1638.

590 Kekkonen, M., & Hebert, P. D. N. (2014). DNA barcode-based delineation of putative species: efficient
591 start for taxonomic workflows. *Molecular Ecology Resources*, *14*(4), 706–715. doi:
592 10.1111/1755-0998.12233

593 Kennedy, S. R., Prost, S., Overcast, I., Rominger, A. J., Gillespie, R. G., & Krehenwinkel, H. (2020). High-
594 throughput sequencing for community analysis: the promise of DNA barcoding to uncover
595 diversity, relatedness, abundances and interactions in spider communities. *Development*
596 *Genes and Evolution*, 1–17.

597 Kerr, K. C. R., Stoeckle, M. Y., Dove, C. J., Weigt, L. A., Francis, C. M., & Hebert, P. D. N. (2007).
598 Comprehensive DNA barcode coverage of North American birds. *Molecular Ecology Notes*, 7,
599 535–543.

600 Lambert, A., & Stadler, T. (2013). Birth–death models and coalescent point processes: The shape and
601 probability of reconstructed phylogenies. *Theoretical Population Biology*, 90, 113–128.

602 Larsen, B., & Aone, C. (1999). *Fast and effective text mining using linear-time document clustering*.
603 16–22.

604 Larson, E. R., Castelin, M., Williams, B. W., Olden, J. D., & Abbott, C. L. (2016). Phylogenetic species
605 delimitation for crayfishes of the genus *Pacifastacus*. *PeerJ*, 4, e1915.

606 Leaché, A. D., Fujita, M. K., Minin, V. N., & Bouckaert, R. R. (2014). Species delimitation using
607 genome-wide SNP data. *Systematic Biology*, 63(4), 534–542.

608 Leasi, F., & Norenburg, J. L. (2014). The necessity of DNA taxonomy to reveal cryptic diversity and
609 spatial distribution of meiofauna, with a focus on Nemertea. *PLoS One*, 9(8), e104385.

610 Leavitt, S. D., Moreau, C. S., & Lumbsch, H. T. (2015). The dynamic discipline of species delimitation:
611 progress toward effectively recognizing species boundaries in natural populations. In *Recent*
612 *Advances in Lichenology* (pp. 11–44). Springer.

613 Lin, X., Stur, E., & Ekrem, T. (2015). Exploring genetic divergence in a species-rich insect genus using
614 2790 DNA barcodes. *PloS One*, 10(9), e0138993.

615 Lithanatudom, S. K., Chaowasku, T., Nantarat, N., Jaroenkit, T., Smith, D. R., & Lithanatudom, P.
616 (2017). A First Phylogeny of the Genus *Dimocarpus* and Suggestions for Revision of Some
617 Taxa Based on Molecular and Morphological Evidence. *Scientific Reports*, 7(1), 6716.

618 Monaghan, M. T., Wild, R., Elliot, ., Fujisawa, T., Balke, M., Inward, D. J. G., ... Vogler, A. P. (2009).
619 Accelerated species inventory on Madagascar using coalescent-based models of species
620 delineation. *Systematic Biology*, *58*, 298–311.

621 Nichols, R. (2001). Gene trees and species trees are not the same. *Trends in Ecology and Evolution*,
622 *16*(7), 358–364.

623 Ortiz, D., & Francke, O. F. (2016). Two DNA barcodes and morphology for multi-method species
624 delimitation in Bonnetina tarantulas (Araneae: Theraphosidae). *Molecular Phylogenetics and*
625 *Evolution*, *101*, 176–193.

626 Pante, E., Puillandre, N., Viricel, A., Arnaud-Haond, S., Aurelle, D., Castelin, M., ... Samadi, S. (2015).
627 Species are hypotheses: avoid connectivity assessments based on pillars of sand. *Molecular*
628 *Ecology*, *24*(3), 525–544. doi: 10.1111/mec.13048

629 Papakostas, S., Michaloudi, E., Proios, K., Brehm, M., Verhage, L., Rota, J., ... Fontaneto, D. (2016).
630 Integrative taxonomy recognizes evolutionary units despite widespread mitonuclear
631 discordance: evidence from a rotifer cryptic species complex. *Systematic Biology*, *65*(3), 508–
632 524.

633 Pentinsaari, M., Vos, R., & Mutanen, M. (2016). Algorithmic single-locus species delimitation: effects
634 of sampling effort, variation and nonmonophyly in four methods and 1870 species of beetles.
635 *Molecular Ecology Resources*.

636 Pino-Bodas, R., Burgaz, A. R., Teuvo, A., & Stenroos, S. (2018). Taxonomy of *Cladonia angustiloba* and
637 related species. *The Lichenologist*, *50*(3), 267–282.

638 Pons, J., Barraclough, T. G., Gomez-Zurita, J., Cardoso, A., Duran, D. P., Hazell, S., ... Vogler, A. P.
639 (2006). Sequence-based species delimitation for the DNA taxonomy of undescribed insects.
640 *Systematic Biology*, *55*, 595–609.

641 Puillandre, N., Cruaud, C., & Kantor, Y. I. (2010). Cryptic species in *Gemmuloborsonia* (Gastropoda:
642 Conoidea). *Journal of Molluscan Studies*, *73*, 11–23.

643 Puillandre, N., Lambert, A., Brouillet, S., & Achaz, G. (2012). ABGD, Automatic Barcode Gap Discovery
644 for primary species delimitation. *Molecular Ecology*, *21*, 1864–1877.

645 Puillandre, N., Baylac, M., Boisselier-Dubayle, M.-C., Cruaud, C., & Samadi, S. (2009). An integrative
646 approach to species delimitation in *Benthomangelia* (Mollusca: Conoidea). *Biological Journal
647 of the Linnean Society*, *96*(3), 696–708.

648 Puillandre, N., Fedosov, A. E., Zaharias, P., Aznar-Cormano, L., & Kantor, Y. I. (2017). A quest for the
649 lost types of *Lophiotoma* (Gastropoda: Conoidea: Turridae): integrative taxonomy in a
650 nomenclatural mess. *Zoological Journal of the Linnean Society*, *181*(2), 243–271.

651 Puillandre, N., Macpherson, E., Lambourdière, J., Cruaud, C., Boisselier-Dubayle, M.-C., & Samadi, S.
652 (2011). Barcoding type specimens helps to identify synonyms and an unnamed new species
653 in *Eumunida* Smith, 1883 (Decapoda: Eumunididae). *Invertebrate Systematics*, *25*(4), 322–
654 333. doi: 10.1071/IS11022

655 Puillandre, N., Modica, M.-V., Zhan, Y., Sirovich, L., Boisselier, M.-C., Cruaud, C., ... Samadi, S. (2012).
656 Large-scale species delimitation method for hyperdiverse groups. *Molecular Ecology*, *21*(11),
657 2671–2691. doi: 10.1111/j.1365-294X.2012.05559.x

658 Rambaut, A., & Drummond, A. J. (2014). *Tracer v1.6*. Available from <http://beast.bio.ed.ac.uk/Tracer>.

659 Ramirez, J. L., Birindelli, J. L., Carvalho, D. C., Affonso, P. R., Venere, P. C., Ortega, H., ... Galetti Jr, P.
660 M. (2017). Revealing Hidden Diversity of the Underestimated Neotropical Ichthyofauna: DNA
661 Barcoding in the Recently Described Genus *Megaleporinus* (Characiformes: Anostomidae).
662 *Frontiers in Genetics*, *8*, 149.

663 Rannala, B., & Yang, Z. (2020). Species delimitation. In *Phylogenetics in the Genomic Era*
664 (Scornavacca, C., Delsuc, F., Galtier, N.).

665 Rannala, B. (2015). The art and science of species delimitation. *Current Zoology*, *61*(5), 846–853.

666 Rannala, Bruce, & Yang, Z. (2003). Bayes estimation of species divergence times and ancestral
667 population sizes using DNA sequences from multiple loci. *Genetics*, *164*(4), 1645–1656.

668 Ratnasingham, S., & Hebert, P. D. (2013). A DNA-based registry for all animal species: the Barcode
669 Index Number (BIN) system. *PLoS One*, 8(7), e66213.

670 Reid, N. M., & Carstens, B. C. (2012). Phylogenetic estimation error can decrease the accuracy of
671 species delimitation: a Bayesian implementation of the general mixed Yule-coalescent
672 model. *BMC Evolutionary Biology*, 12(1), 196.

673 Ritchie, A. M., Lo, N., & Ho, S. Y. (2016). Examining the sensitivity of molecular species delimitations
674 to the choice of mitochondrial marker. *Organisms Diversity & Evolution*, 1–14.

675 Rognes, T., Flouri, T., Nichols, B., Quince, C., & Mahé, F. (2016). VSEARCH: a versatile open source
676 tool for metagenomics. *PeerJ*, 4, e2584.

677 Scarpa, F., Sanna, D., Cossu, P., Lai, T., Curini-Galletti, M., & Casu, M. (2017). A molecular approach to
678 the reconstruction of the pre-Lessepsian fauna of the Isthmus of Suez: the case of the
679 interstitial flatworm *Monocelis lineata* sensu lato (Platyhelminthes: Proseriata). *Journal of*
680 *Experimental Marine Biology and Ecology*.

681 Smith, M. A., Poyarkov Jr., N. A., & Hebert, P. D. N. (2008). CO1 DNA barcoding amphibians: take the
682 chance, meet the challenge. *Molecular Ecology Resources*, 8, 235–246.

683 Stamatakis, A. (2006). RAxML-VI-HPC: maximum likelihood-based phylogenetic analyses with
684 thousands of taxa and mixed models. *Bioinformatics*, 22, 2688–2690.

685 Tang, C. Q., Humphreys, A. M., Fontaneto, D., & Barraclough, T. G. (2014). Effects of phylogenetic
686 reconstruction method on the robustness of species delimitation using single-locus data.
687 *Methods in Ecology and Evolution*, 5(10), 1086–1094.

688 Vacher, J.-P., Kok, P. J., Rodrigues, M. T., Lima, J. D., Lorenzini, A., Martinez, Q., ... Gaucher, P. (2017).
689 Cryptic diversity in Amazonian frogs: Integrative taxonomy of the genus *Anomaloglossus*
690 (Amphibia: Anura: Aromobatidae) reveals a unique case of diversification within the Guiana
691 Shield. *Molecular Phylogenetics and Evolution*, 112, 158–173.

692 Wakeley, J. (2009). *Coalescent theory: an introduction*.

693 Yang, Z., & Rannala, B. (2014). Unguided species delimitation using DNA sequence data from multiple
694 loci. *Molecular Biology and Evolution*, msu279.

695 Zhang, J., Kapli, R., Pavlidis, P., & Stamatakis, A. (2013). A general species delimitation method with
696 applications to phylogenetic placements. *Bioinformatics, Advance Access*.

697 Zou, S., Fei, C., Song, J., Bao, Y., He, M., & Wang, C. (2016). Combining and comparing coalescent,
698 distance and character-based approaches for barcoding microalgae: A Test with *Chlorella*-
699 like species (Chlorophyta). *PloS One*, 11(4), e0153833.

700

701 **DATA ACCESSIBILITY STATEMENT**

702

703 ASAP is available at <https://bioinfo.mnhn.fr/abi/public/asap>. Data sharing is not applicable to
704 this article as no new data were created or analyzed in this study.

705 The software used to simulate multispecies coalescent with random speciation time was
706 written in C and is available upon request, as well as the simulated datasets. All real datasets
707 are directly accessible from the ASAP website.

708

709 **AUTHOR CONTRIBUTIONS**

710

711 SB, GA and NP designed the method; GA developed the algorithm and tested it on simulated
712 datasets; SB wrote the program and created the web-interface; NP performed the tests on real
713 datasets; GA and NP wrote the manuscript.

714

715 **TABLES**

716

717 **Table 1.** Results of the analyses of the empirical datasets.

718

Dataset	Reference	#seq	#spec	ASAP 1 st	ASAP 1 st -2 nd	ABGD	PTP	mPTP	GMYC	mGM YC
<i>Benthomangelia</i>	Puillandre et al. 2009	44	5	2/4/5	5	5	6	5	5	11
<i>Gemmuloborsonia</i>	Puillandre et al. 2010	80	5	5	5	5	5	5	5	8
<i>Lophiotoma</i>	Puillandre et al. 2017	276	10	9	10	9	17	13	10	12
<i>Eumunida</i>	Puillandre et al. 2011	127	16	16	16	16	18	16	16	24
Amphibians	Smith et al. 2008	339	39	20	37	38	44	33	38	49
Cladocera	Elias-Gutierrez et al. 2008	355	58	54	54	53	60	54	67	89
Mammals	Borisenko et al. 2008	521	73	66	66	76	73	55	80	95
Turridae	Puillandre et al. 2012	1,000	87	81	88	87	103	69	95	115
Sphingidae	Hajibabaei et al. 2006	989	107	107	107	98	135	105	140	159
Birds	Kerr et al. 2007	2,574	643	527	529	601	634	475	n.a.	n.a.

719

720 Each line represents a dataset which numbers of sequences (#seq) and species (#spec) are
721 reported in the provided reference. We compare the “true” number of species to the
722 predictions made by the partition ranked first by ASAP (ASAP 1st), by the “best” partition
723 among the two first predicted by ASAP (ASAP 1st-2nd), the “best” partition by ABGD and the
724 unique partition predicted by PTP, mPTP, GMYC and mGMYC. There is no partition for
725 Birds by GMYC and mGMYC as we were not able to obtain a Bayesian tree given the large
726 number of sequences. Cells were colored in dark grey when predictions were very accurate (at
727 most 5% different from the referenced number of species) and with light grey when accurate
728 (between 5% and 10%).

729 **FIGURE CAPTION**

730

731 **Figure 1. An illustration of the clustering algorithm on a small dataset of nine sequences.**

732 On the lower part, we report how ASAP proceeds (downward in the figure) through the list of
733 ranked distances (on the left), merging successively sequences into groups (highlighted in
734 colored blocks). For each new group, ASAP computes a p-value that this new group is a
735 panmictic species (values reported on the right part) based on pairwise differences within
736 (intra) and between (inter) subgroups. Furthermore, each time a new group is created, a new
737 partition is built (a sequence of blocks in the central part) that is associated to the current
738 distance d_C . The distances d_C at which the partitions are instantiated are represented in a
739 phenetic tree (top part). Each node is a group, each horizontal dashed line is a partition. For
740 each newly created partition, ASAP also computes a probability of panmixia (p-val) and a
741 relative gap width metrics (W). Then using their respective ranks (given in parenthesis),
742 ASAP computes an ad-hoc ASAP-score: the lower the score, the better the partition.

743

744 **Figure 2. The computation time of ASAP as a function of the number of species.**

745 Illustrating the linear relationship, we estimate that on the current webserver, computation
746 time is seconds 0.07 seconds per species.

747

748 **Figure 3. Performance of ASAP as a function of the speciation rate.** For two alternative

749 models of speciation (Radiation and Yule), we report the fraction of replicates where ASAP
750 find the four correct species (top panels). We considered either only the partition with the best
751 *asap-score* (ASAP-1) or the partitions ranked first and second (ASAP-1/2). Obviously, the
752 later has better performance. We also report the average number of predicted species,
753 regardless they are correct or not (bottom panels). Each point is evaluated on 500 replicates.

754

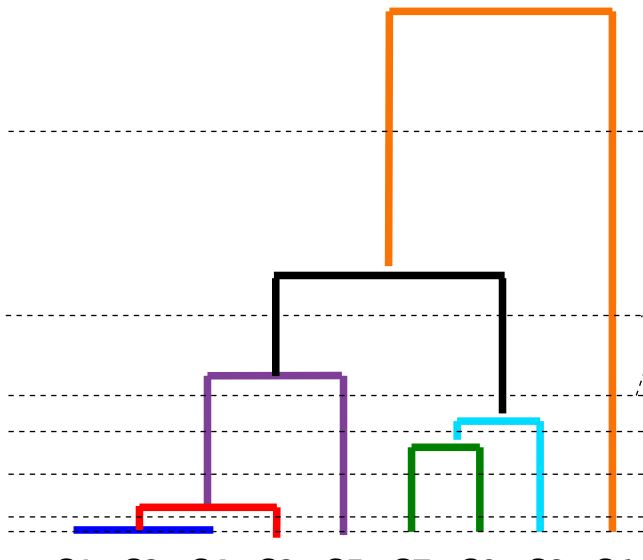
755 **Figure 4. Power of ASAP, ABGD, PTP and GMYC to predict the correct number of**

756 **species among 200 sequences.** We vary the number of true species from 4 to 60 in the
757 Radiation and in the Yule model. Each point is an average of 500 replicates and vertical error
758 bars mark the standard deviation.

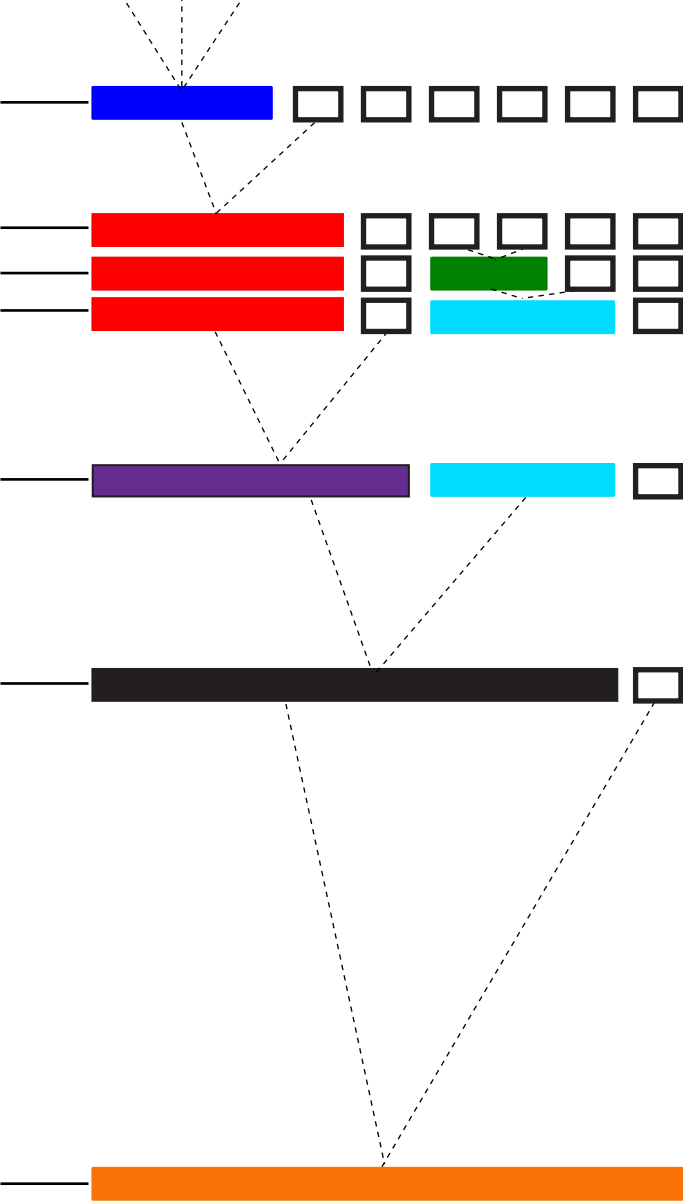
759

- 760 Supplementary Material 1: Computation of the relative barcode gap width.
761
762
763 Supplementary Material 2: Graphical output of ASAP.

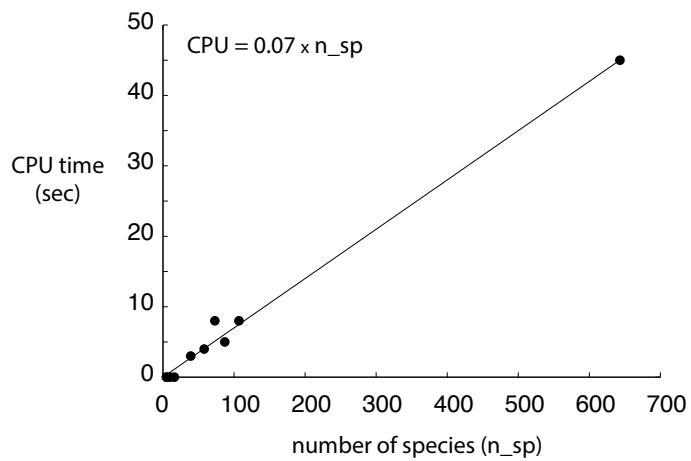
	ranked	distances
1	S1,S3	0.0029
2	S1,S4	0.0029
3	S3,S4	0.0029
4	S1,S2	0.0058
5	S2,S3	0.0058
6	S2,S4	0.0058
7	S7,S9	0.0231
8	S7,S8	0.0289
9	S1,S5	0.0405
10	S3,S5	0.0405
11	S4,S5	0.0405
12	S8,S9	0.0405
13	S2,S5	0.0434
14	S1,S7	0.0665
15	S3,S7	0.0665
16	S4,S7	0.0665
17	S5,S7	0.0665
18	S2,S7	0.0694
19	S1,S9	0.0723
20	S3,S9	0.0723
21	S4,S9	0.0723
22	S5,S9	0.0723
23	S2,S9	0.0751
24	S5,S8	0.0751
25	S1,S8	0.0809
26	S3,S8	0.0809
27	S4,S8	0.0809
28	S2,S8	0.0838
29	S6,S7	0.1301
30	S6,S9	0.1329
31	S1,S6	0.1416
32	S3,S6	0.1416
33	S4,S6	0.1416
34	S2,S6	0.1445
35	S5,S6	0.1445
36	S6,S8	0.1474



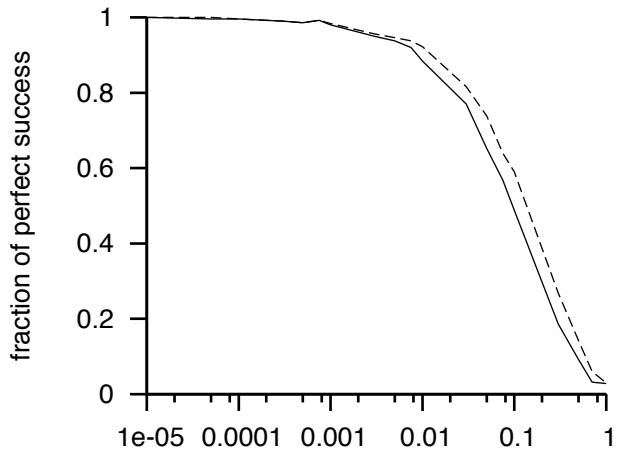
d_T	p-val	Wx100	asap score
0.1301	0.76 (2)	0.57 (2)	2
0.0665	0.76 (3)	0.28 (7)	5
0.0405	0.22 (1)	0.57 (3)	2
0.0289	0.90 (5)	1.11 (1)	3
0.0231	1.00 (6)	0.50 (4)	5
0.0058	0.81 (4)	0.32 (5)	4.5
0.0029	1.00 (6)	0.09 (6)	6



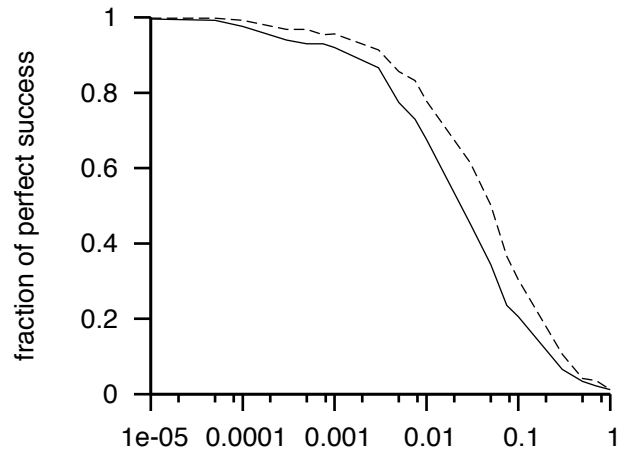
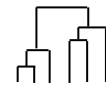
For the nodes		
π_{intra}	π_{inter}	p-val
-	0.0029	n/a
0.0029	0.0058	0.81
-	0.0231	n/a
0.0231	0.0347	0.90
0.0044	0.0412	0.22
0.0218	0.0734	0.76
0.0495	0.1409	0.76



Radiation model

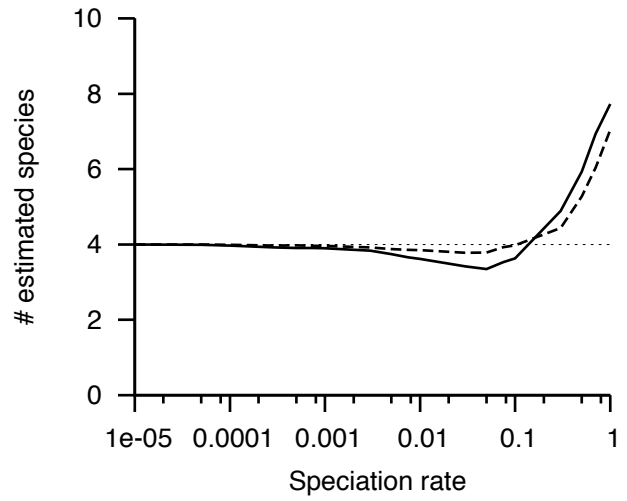
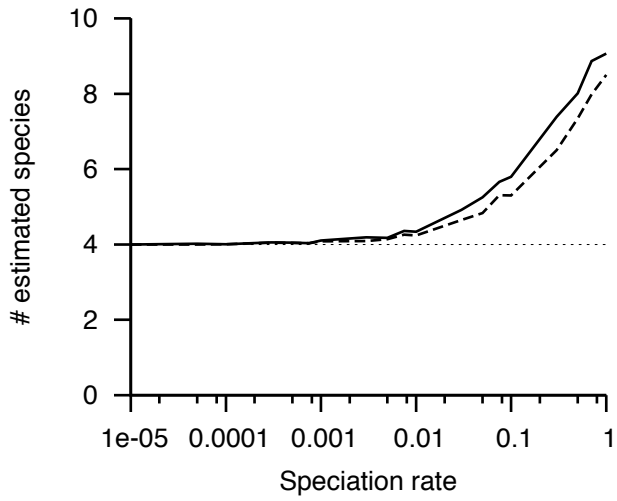


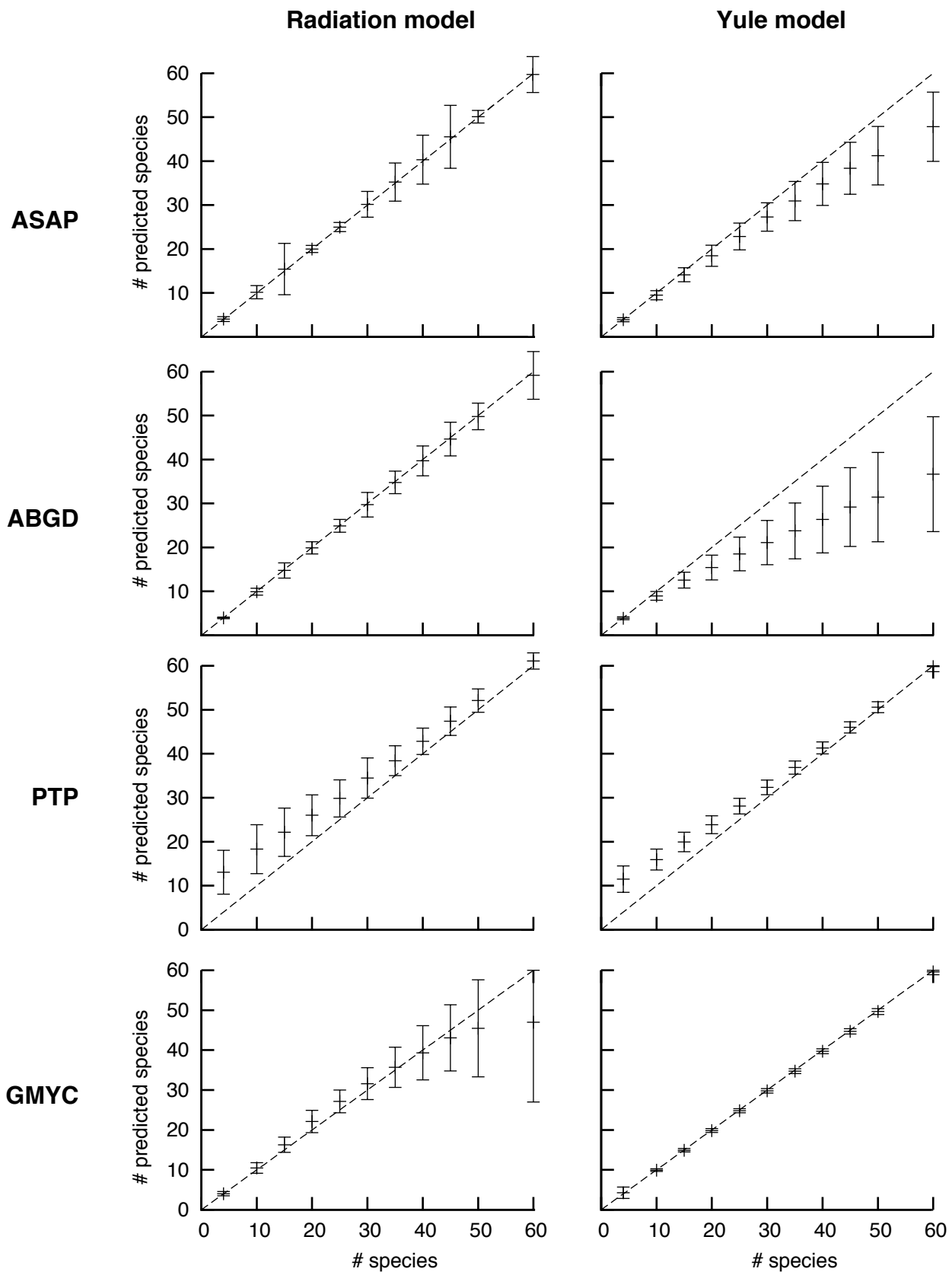
Yule model



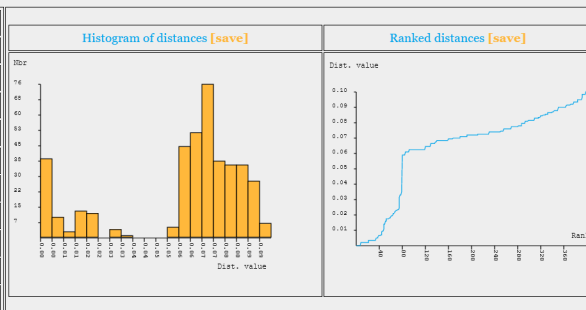
ASAP-1 ———

ASAP-1/2 - - - -

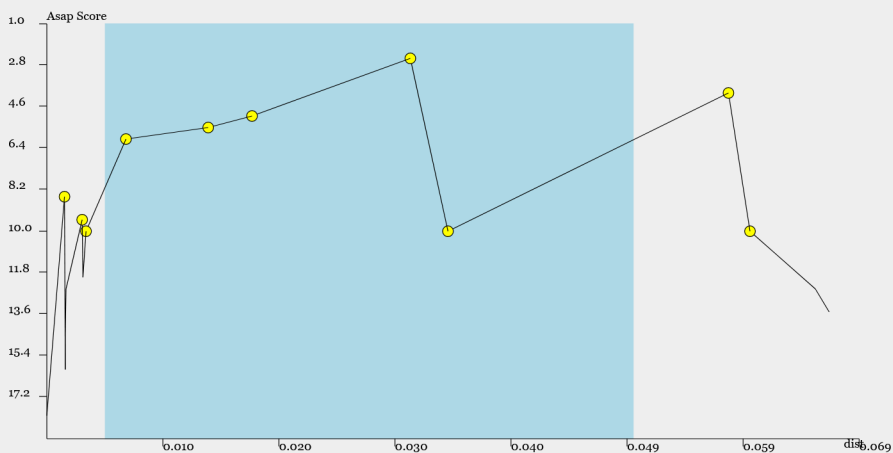




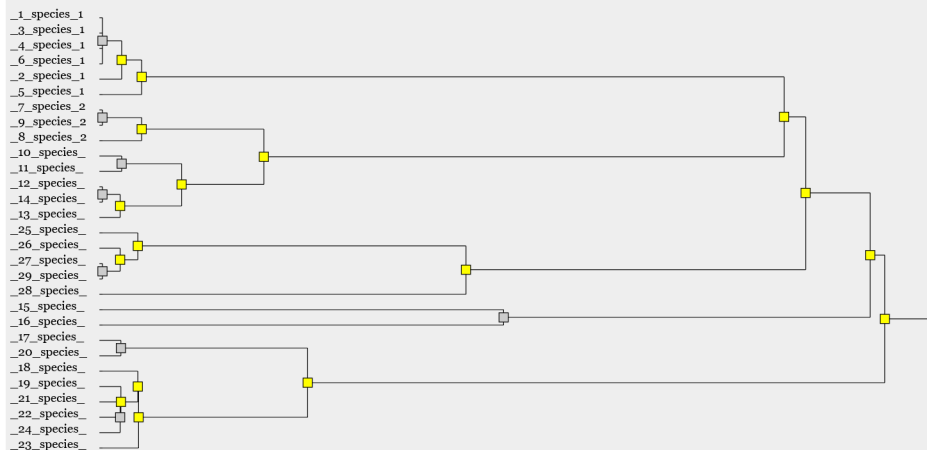
Nb of groups	ASAP score	Proba/ranking	Slope/ranking	Max Dist. for grouping	Text
* 7	1.50	1.16e-01 (1)	2.001266e-01 (2)	0.024263	list esp
* 5	3.00	1.60e-01 (2)	1.048301e-01 (4)	0.046165	list esp
* 8	4.00	3.41e-01 (3)	3.865965e-02 (5)	0.015651	list esp
* 9	4.50	6.87e-01 (8)	2.010442e-01 (1)	0.010307	list esp
* 10	5.00	3.93e-01 (4)	2.325606e-02 (6)	0.005104	list esp
23	7.50	4.81e-01 (6)	1.762736e-02 (9)	0.000761	list esp
15	8.50	7.50e-01 (9)	2.133008e-02 (8)	0.002371	list esp
12	9.00	4.77e-01 (5)	5.378038e-03 (13)	0.003255	list esp
4	9.00	9.74e-01 (11)	2.259335e-02 (7)	0.059945	list esp
* 6	9.00	1.00e+00 (15)	1.787349e-01 (3)	0.032616	list esp



[View/Save Boxed species graph here](#)



Legend: ● <0.001 ● <0.05 ● <0.1 ● >0.1 ● N/A —



[View/save curves and dendrogram here](#)