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Myocardial perfusion simulation for coronary artery disease: a coupled patient-specific multiscale model

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Abstract

Patient-specific models of blood flow are being used clinically to diagnose and plan treatment for coronary artery disease. A remaining challenge is bridging scales from flow in arteries to the micro-circulation supplying the myocardium. Previously proposed models are descriptive rather than predictive and have not been applied to human data. The goal here is to develop a multiscale patient-specific model enabling blood flow simulation from large coronary arteries to myocardial tissue. Patient vasculatures are segmented from coronary computed tomography angiography data and extended from the image-based model down to the arteriole level using a space-filling forest of synthetic trees. Blood flow is modeled by coupling a 1D model of the coronary arteries to a single-compartment Darcy myocardium model. Simulated results on five patients with non-obstructive coronary artery disease compare overall well to $[^{15}\text{O}]\text{H}_2\text{O}$ PET exam data for both resting and hyperemic conditions. Results on a patient with severe obstructive disease link coronary artery narrowing with impaired myocardial blood flow, demonstrating the model's ability to predict myocardial regions with perfusion deficit. This is the first report of a computational model for simulating blood flow from the epicardial coronary arteries to the left ventricle myocardium applied to and validated on human data.

keywords: Heart, Hemodynamics, MBF (Myocardial Blood Flow), Coronary artery disease, PET perfusion map

1 Introduction

Coronary artery disease (CAD), affecting millions of people each year, is the leading cause of death world-wide. Numerous cardiac exams are in clinical use for assessment of CAD, often relying on medical imaging to quantify anatomical and physiological measures prognostic for patient risk. Non-invasive testing modalities most widely utilized have demonstrated only modest diagnostic performance resulting in unnecessary hospital procedures³⁷ costing billions of dollars annually. Some diagnostic modalities involve invasive protocols, putting patients at increased risk. With advances over the last few decades in medical imaging and functional modeling, patient-specific models (PSMs) have emerged as a non-invasive, cost-saving and integrative approach to assessing CAD⁴².

One of the remaining challenges in applying PSMs to quantify blood flow is to connect the disparate scales of cardiovascular physiology⁴⁴. The coronary arteries and the myocardium exhibit scale-specific properties and hence require different modeling approaches to sufficiently capture their behaviour at each scale.

Faced with the inherent challenges of modeling multiscale phenomena in the coronary circulation, many models focus either on the macro or micro vasculature. Taylor et al. describe a PSM to simulate blood flow inside large coronary arteries extracted from coronary Computed Tomography Angiography (cCTA) imaging data, to identify hemodynamically significant lesions via estimation of Fractional Flow Reserve (FFR)⁴². This method, named FFR_{CT} , relies on solving the 3D Navier-Stokes equations^{27,42}. FFR_{CT} is cleared for clinical use by the U.S. Food and Drug Administration (FDA), has been extensively validated in multiple prospective clinical studies^{28,32,35,15} and applied clinically in tens of thousands of patients to date³⁶. Other investigators have described alternate approaches for estimation of FFR with reduced order models of blood flow^{18,40} or hybrid methods⁹.

However in this and other PSMs to model coronary blood flow, functional data is limited to the larger epicardial vessels due to the image resolution: the behaviour of the downstream coronary circulation is approximated by terminal vessel boundary conditions. Hence, although the effect of individual lesions in the epicardial coronary arteries on vessel blood flow

can be modeled, the consequence of disease on blood flow to the myocardial tissue cannot be directly evaluated.

PSMs that seek to model the coronary microcirculation in more detail are still maturing. To handle the enormous number of small arteries, pre-arterioles, arterioles and capillaries in the modeling of whole heart coronary trees, most proposed methods leverage a porous-media model governed by Darcy's law⁷. Michler et al.³¹ have developed and tested a multi-compartment porous model in a porcine myocardium, using cryomicrotome data providing vessels down to the arteriole level. Limitations include the multicompartment computational cost and the difficulty in parameterizing the model with patient-specific data. Recently Alves et al.² proposed a porous model applied to human data, which is able to simulate contrast agent transport and quantify perfusion. This model correlated well with perfusion MRI data, but was only applied to 2D slices of the myocardium.

A few multiscale models have been developed bridging functional analysis in large vessels with tissue perfusion. A major limitation is the anatomical gap between macro and micro scales. To extend the functional analysis, Smith et al.⁴¹ extrapolated a canine coronary anatomy from epicardial coronaries to small vessels by synthetic networks for the arterial and venous systems. 1D flow models in the latter were connected by 0D microcirculatory components: results exhibit realistic vessel pressure distribution, and inside the myocardium the spatial flow heterogeneity follows a fractal pattern as previously described³. Hyde et al.²¹ used animal cryomicrotome data to connect epicardial arteries and microvessels, with a one-way coupling between a 1D coronary model and a multi-compartment porous model: inclusion of vascular data significantly improves the continuum perfusion results in comparison to a more simplistically parameterized model. Conversely, knowledge of perfusion¹⁶ and coronary flow repartition^{33,9} may improve coronary model boundary conditions. Finally, Lee et al.²⁹ proposed a two-way coupling between a 1D coronary model and a single compartment poromechanical model, simulated in a porcine geometry extracted from cryomicrotome. This model reproduces layer-dependent perfusion pattern during the whole heart cycle. This framework also includes contrast agent transport modeling and was used to simulate per-

fusion images, demonstrating a perfusion deficit in the neighbouring region of a simulated stenosis.

So far all models connecting coronary blood flow to myocardial perfusion have been developed on animal data and are descriptive rather than predictive. These models demonstrate the expected characteristics of spatial flow distribution inside the myocardium, but have not been compared with ground-truth data. Furthermore, when relying on destructive image acquisition such as cryomicrotome, these methods are not suited for clinical application.

The aim of this work is to develop a multiscale patient-specific model on human data, enabling blood flow simulation from the large coronary arteries to myocardial tissue. The model is adapted to the patient's coronary arterial network, which is extended with a synthetic vasculature²³. The computational model couples a 1D flow model in coronaries with a single compartment porous model for the myocardium. We applied this method to 6 patients with suspected CAD who underwent cCTA and [¹⁵O]H₂O Positron Emission Tomography (PET)¹³ prior to invasive coronary angiography, which demonstrated non-obstructive CAD in 5 patients and obstructive CAD in 1 patient. We analyzed hemodynamic results both along the coronary vasculature and inside the myocardium with respect to the literature, and compared simulated perfusion with [¹⁵O]H₂O PET perfusion data.

2 Materials and Methods

Regarding ethics, this post hoc substudy comprises the 6 patients mentioned above, from the PACIFIC trial (NCT01521468). The study complied with the Declaration of Helsinki, the study protocol was approved by the VUmc Medical Ethics Review Committee, and all patients provided written informed consent.

This section further consists of five parts: generation of the coronary arterial network from the aorta down to the arteriole level, the one-dimensional blood flow model, the myocardium perfusion model, the coupling of the one-dimensional blood flow model and the myocardium model, and the definition of post-processed quantities.

In modeling the coronary arterial circulation, the aorta, epicardial coronary arteries and left ventricle myocardium are first segmented from patient cCTA image data using custom methods (HeartFlow Inc., US). The coronary trees are then extended from the image-based model down to the arteriole level using a space-filling synthetic forest of arterial trees. The downstream coronary circulation is either taken into account by a terminal resistance, when the coronary model is used stand-alone, or by a myocardium model. The latter is thus described along with its coupling with the coronary model. In both models, blood is considered as an incompressible Newtonian fluid. In this paper we only compute steady-state hemodynamic quantities, seeking to model only mean flow and pressure under resting and hyperemic conditions to study myocardial perfusion. Finally, relevant quantities are derived from the simulations for comparison with the literature and $[^{15}\text{O}]\text{H}_2\text{O}$ PET perfusion data. The complete modeling pipeline is summarized in Fig. 1. Imaging protocols for cCTA and $[^{15}\text{O}]\text{H}_2\text{O}$ PET are provided in Electronic Supplementary Material (ESM) section 5.1.

2.1 Vascular network generation

The vascular networks are patient-specific hybrid vasculatures composed of (1) the aorta and epicardial coronary vessels segmented from cCTA and (2) synthetic trees of the downstream arteries. The synthetic coronary trees are generated as described in Jaquet et al.²³. Briefly, synthetic tree roots are defined at segmented coronary outlets and additionally along the sides of the segmented vessels according to the patient’s branching pattern. These additional roots are added to represent the smaller branches expected to be missed in the segmentation due to the limited cCTA spatial resolution. Only synthetic tree roots close to the left ventricular myocardium are identified for vascular tree generation. A target flow to each tree root is estimated based on the root diameter and the total baseline flow defined later in equation 2. The synthetic tree is generated by minimizing the total generated vascular volume, and constrained by patient-specific priors which are the segmented vessels and left ventricular myocardium. The competitive growth between multiple synthetic trees is driven by tree target flows. Tree generation terminates once a certain number of terminal segments is

reached.

The resulting hybrid vasculature is composed of two bifurcating trees, the left and right (RCA) coronary arteries branching of the aorta. The left side bifurcates into the left anterior descending (LAD) and left circumflex (LCX) arteries. Each vasculature is characterized by the number of terminal segments n_{term} . Each terminal segment outlet i , $i = 1 \dots n_{\text{term}}$, is identified in the manuscript with superscript T,i.

Due to initialization or geometrical constraints, some synthetic trees do not extend properly during the vascular growth computation. They do not reach their target flow and generate few terminal segments, typically with large diameters. To limit discrepancy between terminal segments and to improve scale consistency of the synthetic network, we remove those synthetic trees arising from the segmented vessels which reach less than 20% of their target flow.

2.2 Coronary model

2.2.1 Model description

In the vascular network defined above, blood flow is modeled with the one dimensional approximation of the Navier-Stokes equations¹⁷. Combined with the mass balance equation, the system of equations describing steady blood flow along the centerline axis of each vessel is:

$$\frac{\partial Q}{\partial z} = 0 \tag{1a}$$

$$\frac{\partial}{\partial z} \left(\alpha \frac{Q^2}{S} \right) + \frac{S}{\rho} \frac{\partial p}{\partial z} + 8\pi\nu \frac{Q}{S} = 0 \tag{1b}$$

with z the coordinate of the centerline axis, Q and p the flow rate and pressure respectively, S the cross-sectional area of the vessel, α a geometry-related parameter, ρ the blood density and ν the kinematic viscosity. Flow rate and pressure are respectively reported in mL min^{-1} and mmHg . Blood density is set to 1.06 g cm^{-3} and dynamic viscosity to $0.053 \text{ g cm}^{-1} \text{ s}^{-1}$. The constant $\alpha = 4/3$ is obtained for a parabolic velocity profile. For area-increasing regions,

α values are adjusted to prevent full pressure recovery downstream (Table 1). The values are empirically derived to minimize differences between 1D and 3D CFD solutions.

At each bifurcation of the vascular network, conservation of mass and continuity of pressure provide a relationship between vessel unknowns, completing the system of equations. In contrast to the segmented arteries which have variable vessel areas S , for synthetic vessels the system is practically reduced to Poiseuille law as S remains constant along each segment.

At the left and right coronary trees inlet, the average aortic pressure over a cardiac cycle is set to $P_{AO} = 93$ mmHg. The same aortic pressure is maintained at hyperemia, as FFR remains relatively constant with variations in aortic pressure values and heart rate^{14,24}.

At terminal segments of the network, flow rate boundary conditions are imposed. Given these boundary conditions, flow rate solutions are first obtained for each segment up to the vasculature root. Pressure solutions can then be computed from the root down to the terminal segments. When solving the 1D equations, the flow rate boundary conditions and the synthetic tree geometry are updated iteratively to reflect rest and hyperemic conditions as described in the next section.

Note that segmented coronary arteries are discretized based on their centerline: 1D meshes consist of nodes and elements (connections between consecutively connected nodes). The spacing between two nodes is 0.01 cm. For the synthetic part of the vasculature, as the model is reduced to 0D, no spatial discretization is needed. The spatial location of each synthetic segment is tracked using starting and ending points. This framework has been implemented in an in-house code.

2.2.2 Parameterization for rest and hyperemic conditions

Resting conditions The total myocardial baseline flow $Q_{\text{rest,LV}}^{\text{tot}}$ is estimated from the segmented left ventricular myocardial volume V_{LV} , following the relationship defined by Choi et al.⁸:

$$Q_{\text{rest,LV}}^{\text{tot}} = \zeta \times V_{\text{LV}}^{\gamma} \quad (2)$$

Table 1: Multiplication factor to the default α value of $4/3$ for area-increasing regions under resting and hyperemic conditions.

Segmented vessel node	α value
<i>Resting conditions</i>	
In bifurcating region	0.29
Outside bifurcating region	0.34
<i>Hyperemic conditions</i>	
In bifurcating region	0.72
Outside bifurcating region	0.36

with γ set to 0.75^8 . The coefficient ζ is assigned a value of $3.41 \text{ mL}^{0.25} \text{ min}^{-1}$. This value was determined empirically using a left ventricular myocardial mass distribution of patients enrolled in previously completed clinical studies^{34,28} by fitting to a normal distribution of myocardial blood flow with a median of $1 \text{ mL min}^{-1} \text{ g}^{-1}$.

The flow at each terminal segment that is perfusing the left ventricle is initialized based on its radius:

$$q_{\text{rest},i}^{\text{T},i} = \frac{(r^{\text{T},i})^{2.7}}{\sum_{i=1}^{n_{\text{term}}} (r^{\text{T},i})^{2.7}} Q_{\text{rest,LV}}^{\text{tot}} \quad (3)$$

where $r^{\text{T},i}$ is the terminal segment radius. An iterative process is used to maintain the values of these baseline flow boundary conditions as much as possible. If they create excessive pressure losses in the tree (and thus too low terminal resistance compared to an ideal minimum terminal resistance), then synthetic trees are dilated to replicate the physiological dilation of arterioles and small arteries. The iterative process for resting conditions is described in detail in ESM section 5.2.1.

Hyperemic conditions The hyperemic conditions approximate a stress state where the coronary vessels are maximally dilated as occurring when administering adenosine for invasive FFR measurement or perfusion imaging (ESM section 5.1). Wilson et al. showed that

under these conditions, the total coronary resistance falls to a fourth of the resting value⁴³. Thus, the total ideal hyperemic flow is defined as:

$$Q_{\text{stress,LV}}^{\text{tot}} = 4Q_{\text{rest,LV}}^{\text{tot}} \quad (4)$$

and the diameters of all synthetic segments are dilated to their maximal capacity (40%) from the initial, undilated values, consistent with a Poiseuille relationship between resistance and diameter. Thus, some of the dilation capacity can be used at rest to accommodate oxygen demand of the perfused myocardium at baseline and any remaining dilation capacity is used to simulate maximum hyperemia. Ideal terminal segment flows are calculated as:

$$q_{\text{stress}}^{\text{T,i}} = 4q_{\text{rest}}^{\text{T,i}} \quad (5)$$

The iterative process for hyperemic conditions is described in detail in ESM section 5.2.2.

Arteries not perfusing the left ventricle For the vessels not perfusing the left ventricle (mostly vessels in proximal RCA), terminal segment flow boundary conditions are:

$$q_{\text{rest}}^{\text{T,i}} = \frac{(r^{\text{T,i}})^{2.7}}{n_{\text{term,non-LV}} \sum_{i=1} (r^{\text{T,i}})^{2.7}} Q_{\text{rest,non-LV}}^{\text{tot}} \quad (6)$$

where $r^{\text{T,i}}$ denotes the terminal segment radius, $n_{\text{term,non-LV}}$ the number of outlets not perfusing the left ventricle, and $Q_{\text{rest,non-LV}}^{\text{tot}} = 0.2 \cdot Q_{\text{rest,LV}}^{\text{tot}}$ (see¹¹). Ideal hyperemic flow is defined as:

$$q_{\text{stress,non-LV}}^{\text{tot}} = 4q_{\text{rest,non-LV}}^{\text{tot}} \quad (7)$$

2.3 Myocardium model

For simplicity, hereafter the left ventricle myocardium and the septum are referred to as the myocardium.

2.3.1 Model description

Blood flow in the myocardium is modeled by a single compartment Darcy model^{7,10}:

$$\mathbf{w} + \mathbf{K}\nabla p = 0 \quad (8a)$$

$$\nabla \cdot \boldsymbol{\omega} = \beta_{\text{source}} (p_{\text{source}} - p) - \beta_{\text{sink}} (p - p_{\text{sink}}) \quad (8b)$$

with \mathbf{K} the permeability tensor, $\boldsymbol{\omega}$ the Darcy velocity, p the capillary bed pressure, p_{source} and p_{sink} the source and sink pressure terms respectively, β_{source} and β_{sink} parameters describing the conductance of flow entering and exiting the myocardium respectively.

In terms of physiological meaning, source and sink terms represent respectively the flow entering the myocardium through the coronary model outlets and the venous system drainage. Flow within the myocardium is driven by pressure differences. A no-flux boundary condition is applied to the myocardial wall, which is considered impermeable. Equations 8a and 8b are combined as a single Poisson equation which is solved for p using P1 elements, implemented in the FreeFEM framework²⁰. Myocardial meshes consist of $\approx 500,000$ tetrahedral elements with an average element volume of 2×10^{-4} mL, a sufficient resolution to obtain mesh-independent solutions with regards to myocardial blood flow.

2.3.2 Parameterization

We consider an isotropic permeability field with constant value $K = 2 \times 10^{-5} \text{ cm}^2 \text{ Pa}^{-1} \text{ s}^{-1}$ (see⁷). The coefficient β_{source} is assumed constant over the entire myocardial volume and estimated as:

$$\beta_{\text{source}} = \frac{Q^{\text{tot}}}{(\bar{p}_{\text{source}} - \bar{p})V_{\text{LV}}} \quad (9)$$

where V_{LV} denotes the myocardial volume, Q^{tot} the total myocardial blood flow, \bar{p}_{source} the average pressure of all source terms, and $\bar{p} = 15 \text{ mmHg}$ the targeted average capillary pressure⁷. Notice that the proportionality to Q^{tot} ensures that β_{source} is increased from rest to hyperemia. This increase reflects the vasodilatory response of the myocardium under hyperemia.

The sink terms are homogeneously distributed over the myocardial volume, thus the coefficient β_{sink} is assumed constant and estimated as:

$$\beta_{\text{sink}} = \frac{Q^{\text{tot}}}{(\bar{p} - p_{\text{sink}})V_{\text{LV}}} \quad (10)$$

Hence β_{sink} also increases under hyperemia reflecting increased micro-vessel recruitment and

venous elastic dilation. p_{sink} is regarded as the reference pressure and is thus equal to 0 mmHg.

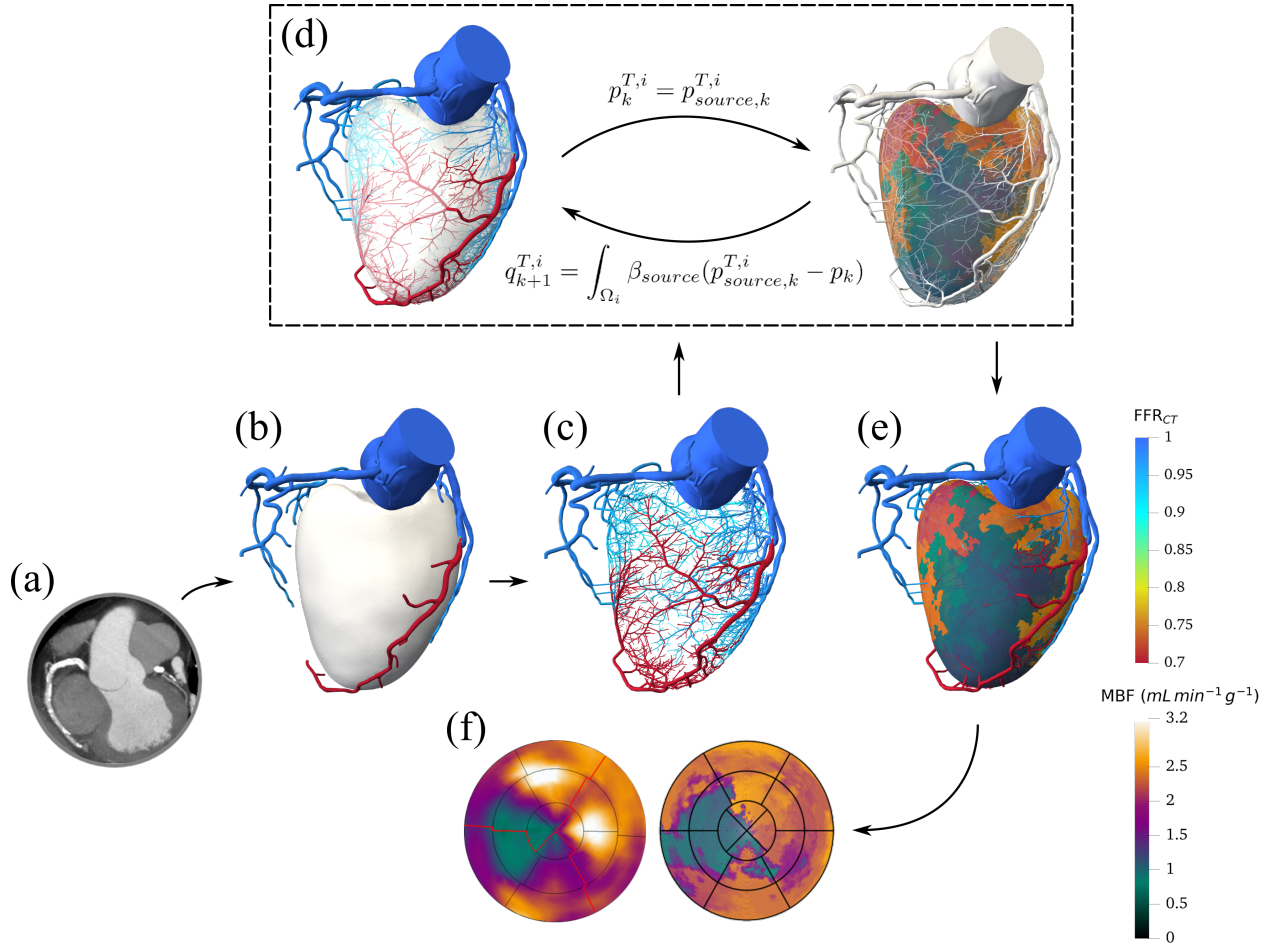


Figure 1: Illustration of the pipeline for a patient with disease in the left anterior descending coronary artery. (a) CT imaging data. (b) Segmented geometry and FFR_{CT} analysis. (c) FFR_{CT} results in segmented and synthetic vasculature. (d) Illustration of the coupling loop, demonstrating quantities exchanged between the coronary model (left) and the myocardium model (right) at coupling iteration k . (e) Hyperemic MBF for coupled model. (f) Comparison of simulated (right) and $[^{15}\text{O}]\text{H}_2\text{O}$ PET (left) hyperemic perfusion maps.

2.4 Coupling of the two models

The coronary and the myocardium model are strongly coupled. The interaction between the two models takes place at the terminal segment outlets which correspond to the source terms in the porous model description.

Each coronary outlet flow is associated to a respective perfusion territory in the myocardium. Those territories Ω_i are estimated from a discrete weighted Voronoi tessellation³⁹ using the terminal segment diameters as weights (equivalent to a Laguerre tessellation). Consequently, segments with larger diameters, carrying a higher amount of flow, are assigned larger perfusion territories. For blood flow, the coupling process involves an initialization loop, followed by iterations coupling the two models and is described in detail in ESM section 5.3. Exchange quantities at coupling iteration k are illustrated in Fig. 1d. The coupling framework has been developed in an in-house code.

2.5 Post-processing of results

Simulations provide pressure and flow rate information at different levels. FFR_{CT} is computed as the pressure at a given vessel location in the coronary tree divided by aortic inlet pressure. The clinical threshold between positive and negative results is 0.8. Note that for the segmented vasculature, FFR_{CT} results are visualized in 3D by projecting centerline values to the arterial wall.

Another quantity of interest is the Myocardial Blood Flow (MBF) for a given perfusion volume, defined as:

$$\text{MBF}_j = \frac{Q_j}{\Omega_j} \quad (11)$$

where Q_j is the flow rate associated to a perfusion volume Ω_j . When Ω_j corresponds to a Voronoi territory, Q_j is the associated terminal segment flow $q^{T,j}$. One can decompose the myocardial volume into smaller volumes and calculate the MBF value of each such volume. Depending on the volume decomposition chosen, characterization of MBF is possible at different spatial resolutions by computing statistics (mean, SD and range) over the MBF

values of the sub-volumes.

Here we consider 4 distinct levels of spatial resolution encountered in clinical practice, detailed in Table 2. The 3D simulation results depict MBF values on a scale with characteristic volume of 10^{-4} mL ("voxels" resolution level in cCTA imaging). The whole myocardial volume is decomposed into 17 standardized segments proposed by the American Heart Association (AHA)⁶ ("AHA segments" resolution level). Based on the AHA segmentation, 3D MBF results are visualized with a 2D perfusion map representing the myocardial volume. Perfusion maps depict MBF values derived from the resolution of the $[^{15}\text{O}]\text{H}_2\text{O}$ PET image, which has a characteristic volume of 6.4×10^{-2} mL, while indicating the approximate boundaries of each AHA segment. The apex segment is not included, since its volume is negligible.

The local flow heterogeneity is assessed with the fractal analysis described in Bassingthwaite et al.³. The Relative Dispersion ($\text{RD} = \text{SD}/\text{mean}$) values of the flow distribution at each scale are plotted in logarithmic scale against myocardial territory volumes of increasing size. Obtaining a line expresses self-similarity, its slope relating to the Fractal Dimension (FD) as: $\text{slope} = 1 - \text{FD}$. A FD value equal to 1 indicates that the heterogeneity is uniform at all scales. The higher the FD, the more heterogeneous the flow becomes when probed at finer scales.

3 Results

The generated synthetic vasculatures consist of varying number of terminal segments n_{term} depending on the study. The synthetic trees that did not extend properly during vascular growth due to constraints were trimmed but they carried less than 2% of the total number of terminal segments.

Table 2: Different levels of spatial resolution considered for MBF calculation.

Resolution level	Notation	Total number of volumes	Order of magnitude	Description
Whole myocardium	MBF_{myo}	1	100 mL	Organ scale. MBF_{myo} is calculated according to eq. 11, where the entire myocardial volume is considered as the perfusion volume.
Main coronary arteries	MBF_{trees}	3	35 mL	Each perfusion volume corresponds to the perfusion territory of each main coronary artery (RCA, LAD, LCX).
AHA segments	MBF_{AHA}	17	6 mL	Perfusion volumes correspond to AHA segments detailed in ⁶ .
Voxels	MBF_{voxels}	$\approx 1,000,000$	10^{-4} mL	High resolution from cCTA, for characterization of MBF at the local level.

3.1 Baseline behaviour: Coronary model versus coronary-myocardium coupled model in a reference case

We explore the main behavior of the coronary and coupled model by comparing simulation results in both resting and hyperemic conditions for a single patient with non-obstructive CAD (Patient 1). A synthetic vasculature with around 3000 terminal segments defines the reference case. We investigate the main differences between the two models regarding blood flow behaviour in the coronaries and inside the myocardium.

While the mean coronary outlet flow remains the same between the two models, the standard deviation of outlet flows is increased by around 25% for rest and hyperemia in the coupled model compared to the coronary model (Table 3).

Pressure results along the vasculature are evaluated based on the diameter-defined Strahler order vessel numbering system²⁶. A global view of the pressure drop in hyperemic conditions for the whole vasculature is provided in Fig. 2a, where pressure values for each vessel are grouped according to the vessel's Strahler order. While such a description can be useful in identifying median pressure and range at different scales, it can often be misleading: pressure may increase with decreasing order (see median pressure in orders 9-8). For this reason one should not interpret the median, minimum or maximum pressure per order as representative pressure drop along individual paths.

To emphasize this point, pressure drop results for paths spanning from segmented vessels (orders 9-11) to terminal segments (mainly orders 5-7) are plotted individually (Fig. 2b). For visualization purposes, only one randomly selected path per main coronary tree is displayed for both models in resting and hyperemic conditions. As expected, results show a larger pressure drop in hyperemia compared to rest, with minor differences between the two models. Pressure is always decreasing when going down a Strahler order. Similar results were found for all paths.

Next, we assess the blood flow results inside the myocardium. For a given model, mean MBF remains practically constant across the different levels of resolution detailed in Table 2.

Table 3: Blood flow results along vasculature and inside the myocardium for vascular networks with varying number of terminal segments for Patient 1.

Number of terminal segments	Coronary model			Coupled model		
	3,000	6,000	12,000	3,000	6,000	12,000
Terminal segment flows ($[\text{mL min}^{-1}] \times 10^{-2}$)						
Resting conditions	$2.89 \pm$	$1.44 \pm$	$0.73 \pm$	$2.84 \pm$	$1.42 \pm$	$0.71 \pm$
	2.25	1.1	0.56	3.62	2.14	1.05
Hyperemic conditions	$9.73 \pm$	$4.86 \pm$	$2.44 \pm$	$9.49 \pm$	$4.73 \pm$	$2.36 \pm$
	7.72	3.79	1.92	12.33	7.31	3.58
MBF _{AHA} ($\text{mL min}^{-1} \text{g}^{-1}$)						
Resting conditions	$1.24 \pm$	$1.22 \pm$	$1.23 \pm$	$1.14 \pm$	$1.14 \pm$	$1.14 \pm$
	0.68	0.52	0.54	0.05	0.05	0.05
	(0.39–	(0.48–	(0.43–	(1.04–	(1.04–	(1.03–
	3.24)	2.42)	2.39)	1.21)	1.2)	1.21)
Hyperemic conditions	$4.17 \pm$	$4.11 \pm$	$4.12 \pm$	$3.84 \pm$	$3.83 \pm$	$3.79 \pm$
	2.27	1.75	1.82	0.31	0.29	0.33
	(1.28–	(1.58–	(1.41–	(3.27–	(3.33–	(3.18–
	10.32)	7.75)	7.77)	4.32)	4.28)	4.3)

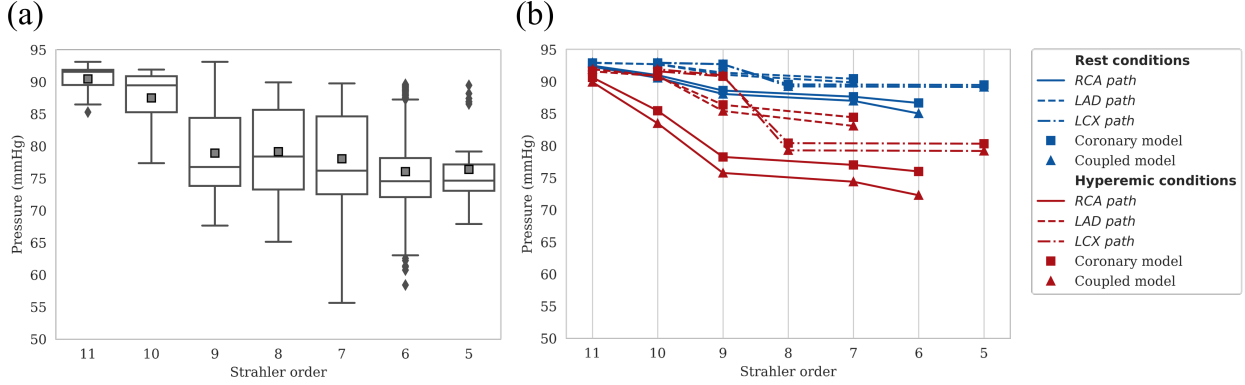


Figure 2: Pressure results along vasculature for reference case. (a) Pressure for vessels in each Strahler order for hyperemic conditions (coupled model). Median and mean pressure values for each order are depicted with horizontal lines and grey squares, respectively. (b) Pressure along 1 random path in each of the three main coronary trees (RCA, LAD and LCX) for resting and hyperemic conditions.

However, increasing the resolution leads to a significant increase in the standard deviation of MBF values, as locally heterogeneous regions which were previously averaged out are revealed (Fig. 3).

Comparing the two models, no significant difference is observed in predicted mean MBF. However, when taking into account the variance and range of MBF values, the coronary model displays extremely heterogeneous results. On the other hand, the coupled model exhibits a more homogeneous distribution of MBF values which seems physiologically more realistic. In hyperemic conditions, computed mean MBF increases by a factor of approximately 3.4 across all scales, reflecting the increase of total flow in the system. The behaviour of the two models remains the same compared to resting conditions.

Finally, the impact of the synthetic vasculature geometry on the results is assessed by generating different vasculatures for Patient 1. In particular, we investigate (1) the effect of the number of terminal segments (ESM section 5.4.1), and (2) the effect of the inherent randomness in the tree generation method (ESM section 5.4.2). The coupled model demonstrated robustness in the geometrical variability of the synthetic vasculature for both studies.

For the rest of this paper, we generate vasculatures with 12000 terminal segments in order to explicitly model as many vessels as possible, while maintaining a manageable computational cost.

Thereafter we consider the AHA segment resolution as the default scale for MBF calculation, unless otherwise stated. For simplicity, the AHA subscript is dropped and MBF_{AHA} will be denoted by MBF.

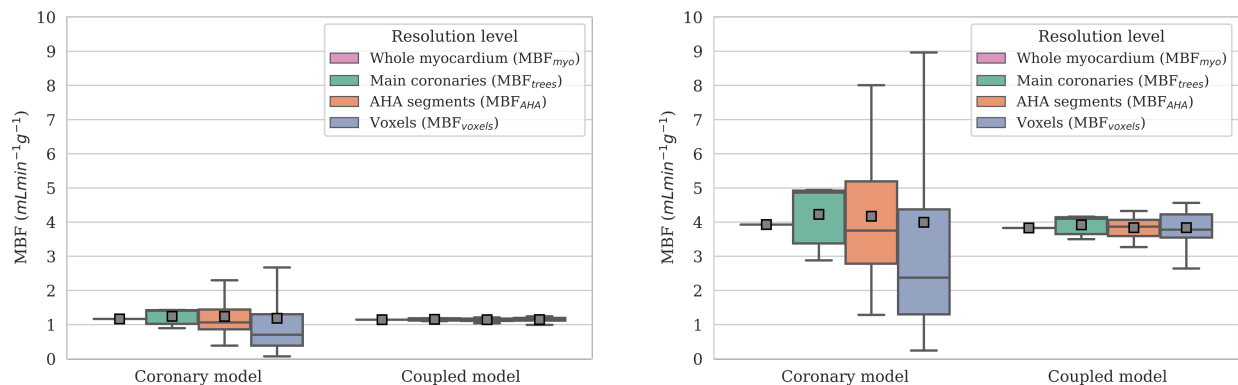


Figure 3: MBF results for the reference case at different levels of spatial resolution detailed in Table 2, for resting (left) and hyperemic (right) conditions. Mean MBF values for each scale are depicted with grey squares. Coronary model outliers for the "AHA segments" and "voxels" resolutions are not depicted for legibility purposes; other scales and the coupled model do not present outliers.

3.2 Patients with non-obstructive CAD

We extend the study to 5 patients (Patient 1 included) with non-obstructive CAD (see ESM section 5.5 for patient clinical data). Generated vasculatures achieved similar levels of vascular growth for 4 out of 5 patients, extending down to Strahler order 4 (though segments of that order represent $< 0.5\%$ of synthetic segments). Most synthetic segments belong to orders 5 and 6, and constitute 20% - 30% and 45% - 55% of the total number of segments, respectively. The vascular network for Patient 5 was the only one that reached a Strahler

order of 3 ($< 1\%$ of segments), with 7%, 33% and 41% of synthetic segments belonging to orders 4, 5 and 6, respectively. Diameter distribution within each Strahler order is provided for each patient in ESM section 5.6.

Blood flow in the vascular network Blood flow results along the vasculature are initially compared to Kassab et al.²⁵, where vascular trees were generated based on porcine statistical data. For this purpose, elements (series of segments belonging to the same Strahler order) are grouped by their Strahler order and mean flow for each order is calculated. Coronary model results for resting conditions follow a similar logarithmic law as in²⁵ (Fig. 4). The law is maintained in hyperemic conditions with similar slope but higher intercept, as the total flow is increased. Similar results are obtained for the coupled model with the exception of flows at order 4, which deviate from the logarithmic law. This is due to the low amount of segments belonging to that order (with the exception of Patient 5), which does not allow for calculation of reliable statistics.

Findings for the reference case regarding terminal segment flows are extended for the rest of the patients: standard deviation of outlet flows is increased in the coupled model compared to the coronary model, while mean flow remains the same regardless of the model. From resting to hyperemic conditions, the mean outlet flow increase ranges from a factor of 2.7 to 3.35 across patients. The initial target factor of 4 is not attained due to the network resistance. Both conditions have the same relative flow heterogeneity.

Mean pressure drop for the coupled model from the aortic root to tree outlets ranges from 6.5 mmHg to 10.6 mmHg at rest, and from 16.5 mmHg to 24.2 mmHg at hyperemia for patients 1 to 4, with similar values for the coronary model. Mean pressure drop is higher for Patient 5, as the vasculature extends down to an additional Strahler order, with 18.7 mmHg at rest and 36.4 mmHg at hyperemia. These results are consistent with values obtained in²⁵: pressure drop from root to vessels with Strahler order 6, 5 and 4 was reported at 5 ± 1.3 mmHg, 8.8 ± 1.7 mmHg and 24.5 ± 2.8 mmHg, respectively. To compare, note that for patients 1 to 4 the majority of terminal segments belong to orders 6 and 5, while for Patient

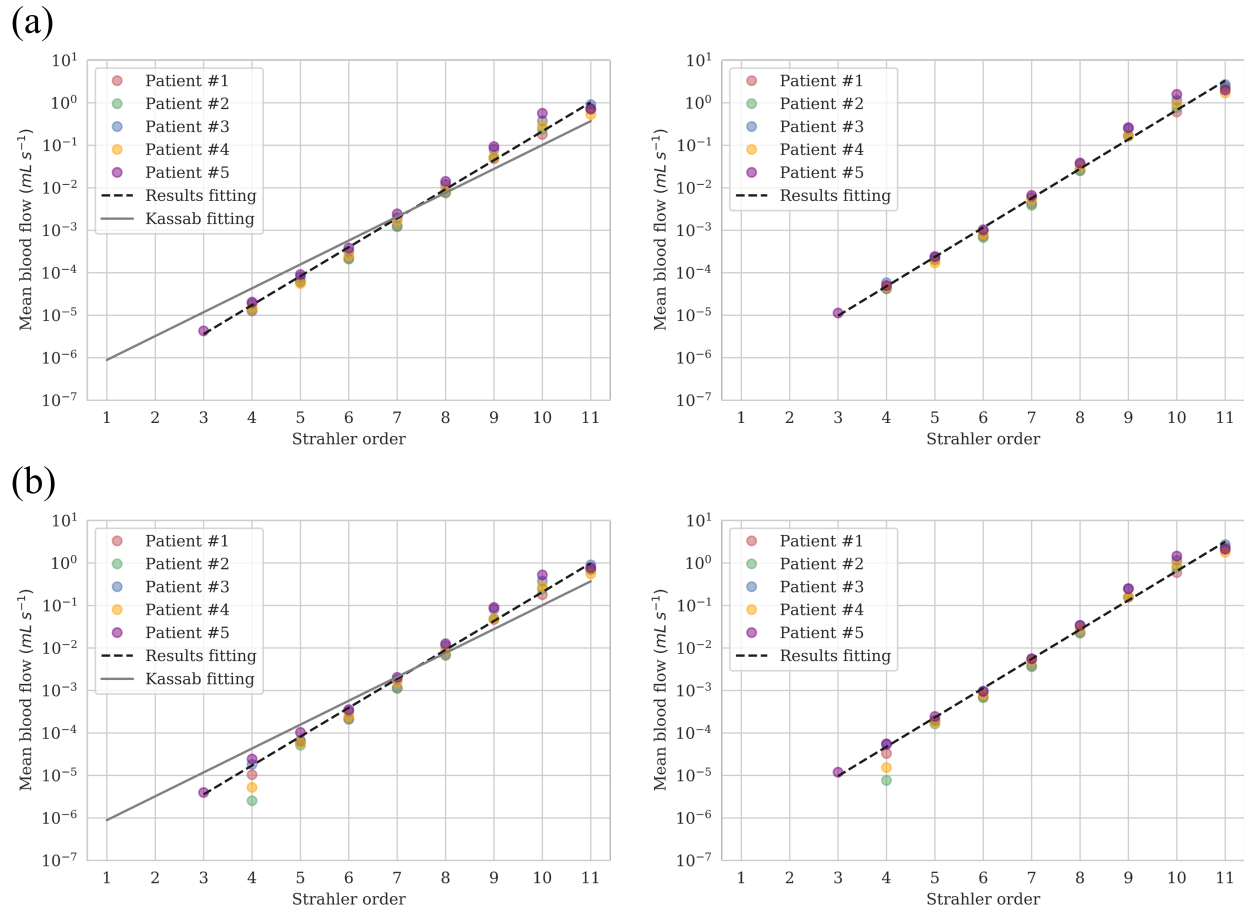


Figure 4: Mean blood flow of elements in the vascular network per Strahler order for (a) coronary model and (b) coupled model. For each model: resting conditions (left), hyperemic conditions (right). The slope of the fitted line for results is equal to 1.57 and 1.59 in resting and hyperemic conditions, respectively, for both models.

5 to orders 5 and 4. The spatial distribution of pressure highlights the patient-specificity of FFR_{CT} analysis (ESM Fig. 5.4).

From resting to hyperemic conditions, mean pressure drop increase ranges from a factor of 1.95 to 2.7 across patients. Note the pressure drop increase is thus less than the increase of flow from rest to hyperemia: this highlights the decrease of tree resistance due to synthetic network dilation, which partially compensates for the resistance increase in the segmented network due to increased flow.

Under resting conditions, the resistance of the overall coronary tree (segmented plus synthetic), defined as $[P_{\text{AO}} - \text{mean}_i(p^{\text{T},i})] / \sum_i q^{\text{T},i}$, accounts for 7% to 11.3% of the total resistance of the system (macro- and micro-vasculature) for patients 1 to 4 and for 20% for Patient 5. Coronary tree resistance plays a more significant role in hyperemic conditions, making up 17.7% to 26% of total resistance for patients 1 to 4 and 39.1% for Patient 5. The coronary tree resistance is thus non-negligible compared to the resistance of the downstream micro-vasculature. The dilation of the vessels in the synthetic network reduces the network resistance but not enough to compensate for the added inertial resistance of the increased flow from rest to hyperemia.

Blood flow inside the myocardium Following a fractal analysis (described in section 2.5), the local flow heterogeneity is assessed under resting conditions for the 5 patients and compared to literature (Fig. 5). Coronary model results are comparable to similar models relying on synthetic networks. In particular, fractal dimension values show good agreement with results obtained with the extended canine synthetic vasculature of Smith et al.⁴¹. The relative dispersion of flow is slightly lower in the coronary model at all scales, indicating a more homogeneous flow distribution. Compared to the synthetic network of Beard et al.⁴ based on porcine data, the coronary model exhibits a higher level of flow heterogeneity and lower fractal dimension, but within the same order of magnitude.

The fractal analysis on animal populations by Bassingthwaite et al.³ provides reference physiological values. Compared to these data, the coronary model results have consistently

higher RD at all scales, while on the contrary, the coupled model consistently lower. This suggests that blood flow predicted by the coronary model tends to be overly heterogeneous, flow predicted by the coupled model overly homogeneous, with physiological values in-between. Furthermore, the very low fractal dimension of the coupled model (close to 1) indicates that there exists only a small difference in heterogeneity across different levels of spatial resolution. The fractal dimension of the coronary model is closer to physiological data.

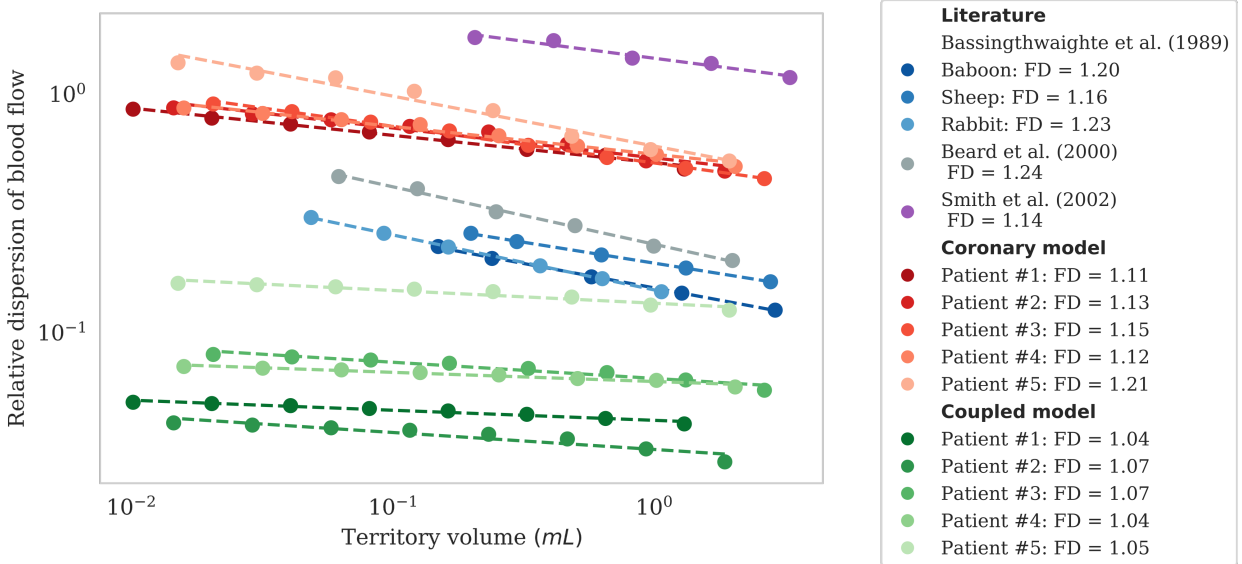


Figure 5: Fractal analysis at resting conditions for 5 patients with non-obstructive CAD and comparison with literature.

Next, MBF results for both models are compared with $[^{15}\text{O}]\text{H}_2\text{O}$ PET exam patient data (Fig. 6a). For resting conditions, mean MBF matched measured data regardless of the model used, with prediction error less than the mean MBF inter-patient variability. When taking into account the variance of the AHA segment MBF, the coronary model exhibits extremely heterogeneous flow distribution which does not correspond with patient data. This behaviour is consistent with the fractal analysis results. However, the coupled model overcomes this limitation demonstrating MBF values within measured range and better agreement overall. Despite the fact that fractal analysis revealed overly homogeneous flow at smaller scales, the considered spatial resolution is low enough that the model produces physiologically valid

results with predictive value. Results are similar for both resting and hyperemic conditions for Patients 1-4.

Simulated perfusion maps for hyperemic conditions are directly compared with $[^{15}\text{O}]\text{H}_2\text{O}$ PET perfusion maps. Coronary model perfusion maps do not correlate well with PET data due to their overly heterogeneous distribution and therefore are not displayed. Coupled model maps (Fig. 6b), while not capturing the exact spatial flow distribution, allow in general for a correct estimation of CAD’s impact on myocardial perfusion. Hyperemic results for Patient 5 did not match measured data, predicting a significant perfusion deficit which was not present in PET data. Lower simulated pressure at the terminal segment outlets led to lower hyperemic MBF. This patient had unique characteristics compared to other patients including higher hyperemic flow and coronary flow reserve (CFR) in the PET exam (Fig. 6a) and a synthetic vasculature upstream of the impaired region that was extended to lower Strahler orders.

3.3 Patient with obstructive CAD

Here we present preliminary results for an extension of the model to a patient with a lesion in the left anterior descending coronary artery (close to 80% reduction in diameter) - apparent from the FFR_{CT} analysis (Fig. 1c). The $[^{15}\text{O}]\text{H}_2\text{O}$ PET exam perfusion map for hyperemic conditions revealed a significant perfusion deficit in the myocardial region corresponding to the LAD tree. For this patient, estimating total coronary flow using the power-law relation between flow and myocardial mass⁸, led to significantly increased flow compared to PET data (24% and 56% excess flow in resting and hyperemic conditions respectively). For this reason, blood flow in the system was instead estimated with the MBF_{myo} value provided by the PET exam in order to determine if the relative effect of the stenosis could be modeled. Previous limitations of the coronary model in terms of flow heterogeneity still apply, thus the analysis is focused on the coupled model.

As MBF is significantly impaired, this patient provides a suitable framework for evaluation of the model’s capacity for vascular dilation. As demonstrated in Fig. 7b, synthetic

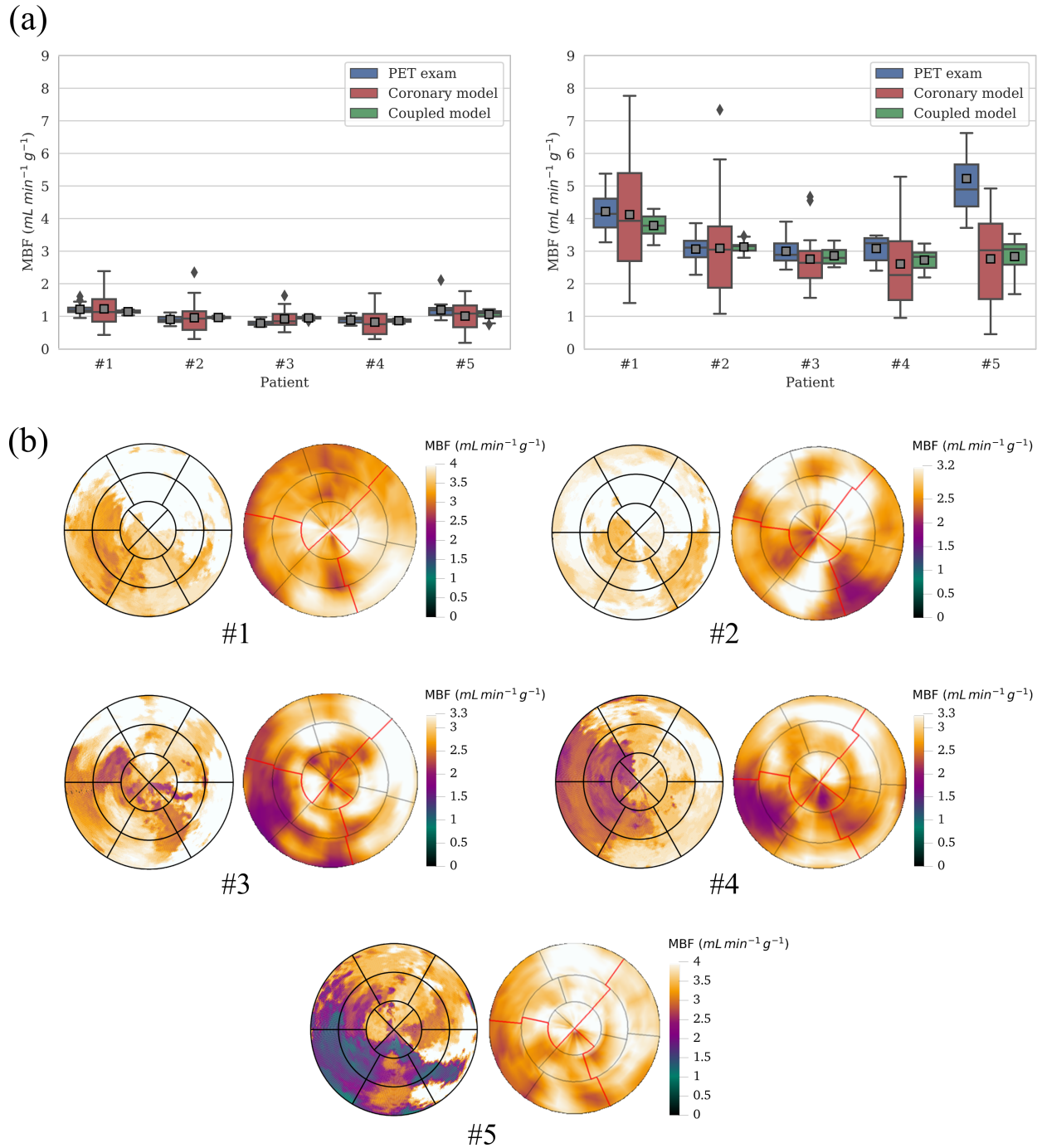


Figure 6: MBF analysis for 5 patients with non-obstructive CAD. (a) Comparison of MBF_{AHA} results with $[^{15}O]H_2O$ PET exam data for resting (left) and hyperemic (right) conditions. Mean MBF_{AHA} values are depicted with grey squares. (b) Comparison of simulated perfusion maps (left) to $[^{15}O]H_2O$ PET exam perfusion maps (right) for the coupled model under hyperemic conditions. Note the upper limit of color map range is truncated to match PET exam range.

segments arising from the LAD were dilated up to 15% under resting conditions, in contrast to healthy trees arising from RCA and LCX where dilation was minimal (up to 3%). These results highlight the model’s ability to account for flow deficits downstream of diseased vessels and adapt diameters accordingly, reflecting the underlying physiological response to ensure adequate myocardial perfusion at rest in patients with epicardial disease.

Pressure drop along three random paths in each main coronary artery is shown in Fig. 7a. LAD paths display higher pressure drop in both resting and hyperemic conditions compared to healthy coronary trees, which takes place almost exclusively in large vessels (between orders 11 and 10). This significant drop early in the vasculature strongly indicates the existence of a hemodynamically significant stenosis in an epicardial vessel of the LAD tree. In addition, outlet pressure decreases almost twice as much from rest to hyperemia in LAD paths (≈ 20 mmHg) compared to RCA and LCX paths (≈ 10 mmHg), reflecting a larger difference between resting and hyperemic flow in regions perfused by LAD trees.

While the total blood flow in the system was explicitly tuned to match PET exam MBF_{myo} , distribution of flow throughout the coronary tree follows the same procedure as previously described. As illustrated in the perfusion map simulated under hyperemia (Fig. 1f), the exact region of the MBF perfusion deficit in the myocardium (in green) is predicted by the coupled model. Under resting conditions, even though synthetic segments corresponding to the impaired region were dilated (Fig. 7b), a lower perfusion is still visible: the simulated map exhibits the same level of lower perfusion as in the PET exam, but the spatial spread is different. In fact, under hyperemia MBF in the perfusion deficit zone is $0.92 \text{ mL min}^{-1} \text{ g}^{-1}$ for a myocardium average MBF of $1.9 \text{ mL min}^{-1} \text{ g}^{-1}$, which is more drastic than at rest where the lower perfusion is of $0.73 \text{ mL min}^{-1} \text{ g}^{-1}$ for an average MBF of $0.81 \text{ mL min}^{-1} \text{ g}^{-1}$.

4 Discussion

In this paper, a multiscale method for simulation of coronary and myocardial blood flow is presented and applied to human data. Patient-specific three-dimensional coronary artery

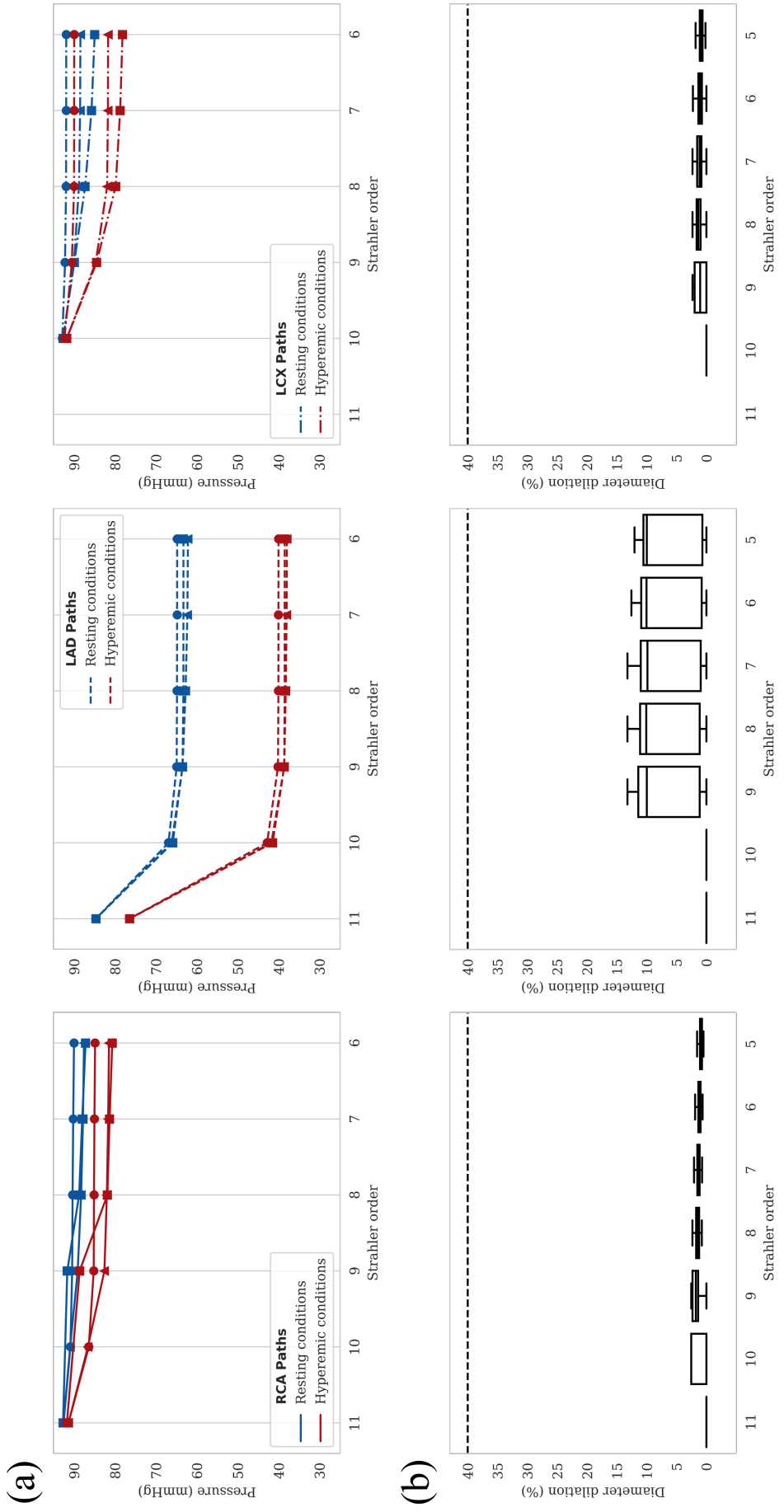


Figure 7: Pressure and dilation analysis along vasculature for a patient with LAD lesion. (a) Pressure along 3 random paths of each main coronary tree for resting and hyperemic conditions. Each path is differentiated with a circle, triangle or square. (b) Diameter dilation per Strahler order in resting conditions for each coronary tree: RCA (left), LAD (center), LCX (right). The horizontal dotted line represents maximum dilation capacity.

models were segmented using cCTA image data and synthetic trees were generated down to the arteriole level. A stand-alone coronary model and a coronary-myocardium coupled model were investigated with the aim of simulating myocardial perfusion in health and disease. The coupled model demonstrated robustness to variations of the generated vasculature (different initialization, number of terminal segments) and better agreement with $[^{15}\text{O}]\text{H}_2\text{O}$ PET exam data overall. In this section, we discuss some of our modeling assumptions, we propose alternative parameterization choices and provide directions for future work. Limitations and clinical potential of the model are also discussed.

4.1 Model assumptions and parameterization

Only steady-state hemodynamic quantities were computed in this paper, which is sufficient for simulated MBF to be comparable to $[^{15}\text{O}]\text{H}_2\text{O}$ PET data, keeping in mind that the acquisition is performed over multiple cardiac cycles. Pulsatile flow simulations would necessitate additional assumptions, more parameters to estimate³⁰, or other patient-specific data not easily measurable, overall introducing additional complexity which is outside of the scope of the current study. In future work, incorporation of flow pulsatility in the model would potentially allow for assessing the risk of plaque rupture in the coronaries.

Moreover, blood was modeled as a Newtonian fluid. For larger epicardial vessels segmented from cCTA, shear rate was actually above 10s^{-1} for all patients in this study. Recent 3D work¹ has demonstrated for mild to severe coronary stenoses that the Newtonian model did not lead to a significantly different centerline velocity compared to three non-Newtonian models, validated by clinical measurements. Moreover, while viscosity changes can affect velocity profiles, it has a minor effect on flow and pressure losses³⁸. The effect of viscosity uncertainty on perfusion could be assessed in future work, but it would require additional patient-specific measurements (hematocrit, plasma viscosity). For smaller synthetic vessels (Strahler orders 4-5), future work could also consider the Fahraeus-Lindqvist effect and plasma skimming.

A single Darcy compartment was used in the myocardium model with homogeneous flow

conductance parameters (β_{source} and β_{sink}). Previous studies have considered more complex porous model descriptions with multiple compartments modeling different scales of the micro-circulation^{31,21}, but this approach makes the flow conductance parameters difficult to estimate and relies on complex anatomical information. The simpler myocardium model proposed herein enables parameter estimation using easier-to-derive patient-specific variables, such as the total coronary flow and the myocardial volume, for which estimation laws already exist^{8,42}. This simplified approach is sufficient for MBF characterization at the AHA segment level as demonstrated by the good agreement with PET exam data. However, finer details of the flow are not captured which can be seen in the almost uniform flow at lower spatial resolutions (Fig. 5). This could be tackled with a homogenization approach on parameter estimation²².

Several refinements in the parameterization of the coupled model are possible. While the mechanical properties of the myocardial tissue are non-uniform and anisotropic⁷, the myocardial permeability tensor \mathbf{K} is considered herein as constant and isotropic. Note that simulations with an increased K value, to account for the additional vessels recruited under hyperemia¹⁹, did not result in any noticeable difference in terms of perfusion. Furthermore, the aortic pressure is based on a population average. Patient-specific values could help fine-tune the range and spatial distribution of MBF for each patient but are variable at rest and difficult to predict under hyperemic conditions.

As shown in the simulated perfusion maps, even though the main MBF characteristics are adequately captured, the spatial distribution of flow does not exactly match PET exam maps. Note that for any given MBF distribution, a corresponding source-sink field can be determined and vice versa. An interesting direction for future work to improve results such as for Patient 5, would be to estimate spatially-heterogeneous flow conductance parameters via a machine learning model, utilizing PET exam data and other patient-specific features.

4.2 Limitations

In the current model, total flow in hyperemic conditions cannot exceed four times the resting total flow. As a result, the model is unable to predict MBF of patients with CFR higher than 4, like Patient 5. Interestingly for this patient, in a separate simulation with increased hyperemic total flow based on PET data, a perfusion deficit with similar magnitude is still present. This indicates that the upstream vasculature is unable to accommodate additional flow, which could be a potential drawback of the synthetic network generation method.

Challenges with the estimation of total flow are apparent from the example of the patient with a severe LAD lesion as the myocardial-mass-based total flow over-estimated the baseline flow. While there is significant variability in resting MBF¹³, our current estimation method is able to capture the reported median level of resting MBF for patients with non-obstructive CAD, as seen in Fig. 6a (rest). For the obstructive CAD patient, our method overestimated total flow by 24%, which is nevertheless within the range reported by Danad et al. for obstructive CAD patients. Studies in additional patients with obstructive CAD may be needed to determine if parameters in addition to the myocardial mass are needed to predict absolute resting blood flow in such patients. Note, in separate simulations, the coupled model was still able to identify the perfusion deficit region, even when the flow estimation was based on myocardial mass, albeit with over-estimated absolute MBF values.

Results on a patient with LAD stenosis demonstrated the model’s ability to mimic the physiological response of arterial dilation. While vasculature downstream of a diseased epicardial vessel was partially dilated at rest to accommodate more flow, the spatial results show that, as for some of CAD non-obstructive patients, the model may need to include more heterogeneous parameterization.

Future work will include a study on how the coupled model affects the prediction of FFR_{CT} , especially for cases near the cut-off value. Finally, more patients are needed for further validation of the model.

4.3 Conclusion and perspectives on clinical relevance

A full pipeline from cCTA to simulation of MBF with potential for clinical application is described. Absolute hyperemic MBF has been shown to have better prognostic⁵ and diagnostic¹³ performances compared to other clinical metrics, like CFR. By utilizing readily available information from a CT scan, this approach enables quantitative assessment of CAD's impact on myocardial perfusion. With the exception of one non-obstructive CAD case for which the model needs to be further refined, simulated MBF matched [¹⁵O]H₂O PET exam data at the AHA segment scale, a widely used resolution in clinical practice. In case of severe CAD, a direct link between coronary artery narrowing and impaired myocardial blood flow is achieved. This framework also makes possible the calculation of ischemic burden, an important clinical metric for evaluation of CAD severity. At the same time, it provides a testbed to explore different combinations of positive/negative FFR and presence/absence of perfusion deficit and better elucidate their different contributions for treatment choice.

As FFR_{CT} analysis is already utilized in clinical practice, incorporation of this method could enhance its current capabilities by extending the analysis to the myocardium: it would constitute a diagnostic tool capable of multiscale simulation of blood flow from the epicardial coronary arteries to the myocardial tissue and enable identification of those regions of the myocardium with diminished blood flow or at risk of infarction in the event of atherosclerotic plaque rupture.

Conflict of interest statement

Authors affiliated with HeartFlow report 'other from HeartFlow Inc.', during the conduct of the study. Authors from Esiee and Inria and Dr. Knaapen report 'grant from HeartFlow Inc.'. In addition, Dr. Taylor, Schaap and Jaquet have a patent U.S. 10,262,101. L. Grady, M. Schaap, C.A. Taylor, C. Jaquet, Systems and Methods for Predicting Perfusion Deficits from Physiological, Anatomical, and Patient Characteristics issued, and a patent U.S. 10,405,925. C. Jaquet, M. Schaap, Y. Bai, L. Grady, C.A. Taylor, Systems and Meth-

ods for Determining and Visualizing a Functional Relationship Between a Vascular Network and Perfused Tissue issued.

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5 Electronic Supplementary Material (ESM)

5.1 Imaging protocols

The imaging data for coronary artery and myocardium segmentation from cCTA and for perfusion map ground truth from $[^{15}\text{O}]\text{H}_2\text{O}$ PET was acquired as described by Danad et. al.¹².

5.1.1 Coronary Computed Tomography Angiography (cCTA)

Patients underwent coronary angiography on a 256-slice CT scanner (Philips Brilliance iCT, Philips Healthcare, Best, the Netherlands) with a collimation 128 x 0.625 mm and a tube rotation time of 270 ms. To visualize the coronary artery lumen a bolus of 100 mL iobitidol

(Xenetix 350) was injected intravenously 5.7 mL s^{-1} , with immediately after a 50 mL saline chaser. The scan was triggered with an automatic bolus tracking technique. The region of interest was placed in the descending thoracic aorta with a threshold of 150 HU. Metoprolol 50 to 150 mg was administered orally if patients had a prescan $\text{HR} \geq 65$ beats per minute (bpm) one hour before the start of the CT protocol. If necessary, 5 to 25 mg metoprolol was given intravenously during the scan to achieve a heart rate < 65 bpm. All patients received 800 μg of sublingual nitroglycerine immediately before cCTA.

5.1.2 $[^{15}\text{O}]\text{H}_2\text{O}$ Positron Emission Tomography (PET)

Patients had to refrain from taking products containing caffeine or xanthine 24 hours before imaging. Patients fasted for at least 4 hours before the scan protocol. All patients were imaged on a hybrid PET/CT device (Philips Gemini TF 64, Philips Healthcare, Best, The Netherlands). During resting conditions as well as vasodilator stress induced by intravenous infusion of adenosine ($140 \mu\text{g kg}^{-1} \text{ min}^{-1}$), a 370 MBq of $[^{15}\text{O}]\text{H}_2\text{O}$ was used as a perfusion tracer. For more information on the cardiac $[^{15}\text{O}]\text{H}_2\text{O}$ PET protocol, image acquisition and quantification of MBF, see Danad et al.¹³.

5.2 Parameterization of coronary model in resting and hyperemic conditions: Detailed description

5.2.1 Resting conditions

Using the initialized flows in eq. 3, we calculate ideal baseline resistance at each terminal segment:

$$R_{\text{base}}^{\text{T},i} = \frac{P_{\text{AO}}}{q_{\text{rest}}^{\text{T},i}} \quad (12)$$

For each terminal segment, an expected minimum resistance is estimated as follows:

$$R_{\text{min}}^{\text{T},i} = \frac{1}{4} R_{\text{base}}^{\text{T},i} \quad (13)$$

Here, we assume a uniform factor but this can vary for each patient. Anatomically, the factor represents a maximal radius dilation capacity of 40% based on uniform dilation of a tree to

achieve a 4-fold reduction in resistance⁴³. Note that minimum resistances are calculated once and fixed for the whole parameterization process.

Using the terminal segment flows as boundary conditions for the first iteration, flow and pressure are computed for the entire tree by solving the 1D equations. The computed terminal segment pressure $p^{T,i}$ and the assigned terminal segment flow $q_{\text{rest}}^{T,i}$ determine the simulated terminal segment resistance $R_{\text{sim}}^{T,i}$:

$$R_{\text{sim}}^{T,i} = \frac{p^{T,i}}{q_{\text{rest}}^{T,i}} \quad (14)$$

If the simulated terminal segment resistance is lower than the minimum resistance value $R_{\text{min}}^{T,i}$, we update both the geometry and the flow. We dilate the radius of the terminal segment and its upstream synthetic segments by multiplying them with a uniform factor of $\left(\frac{R_{\text{base}}^{T,i}}{R_{\text{min}}^{T,i}}\right)^{0.25} = \sqrt{2}$. We reduce the terminal segment flow by δq by solving the following equation

$$(q_{\text{sim,rest}}^{T,i} + \delta q)R_{\text{min}}^{T,i} = P_{AO} - R_p^{T,i}(q_{\text{sim,rest}}^{T,i} + \delta q) \quad (15)$$

where $R_p^{T,i} = \frac{P_{AO} - P_{\text{sim,rest}}^{T,i}}{q_{\text{sim,rest}}^{T,i}}$.

If the simulated terminal segment resistance is greater or equal to $R_{\text{min}}^{T,i}$, we update the geometry by multiplying by a factor of $\left(\frac{R_{\text{base}}^{T,i}}{R_{\text{sim}}^{T,i}}\right)^{0.25}$ but maintain the same flow $q_{\text{rest}}^{T,i}$.

Note that each terminal segment can have a different radius dilation factor as the resistance of the terminal segment will depend on the pressure loss accumulated along the path. For the upstream segments branching to child segments with different dilation factors, bigger dilation factors are chosen to dilate them. Several iterations are necessary to obtain convergence as both the geometry and boundary conditions are updated in each iteration. The convergence criteria used:

$$\max_i \left| \frac{R_{\text{sim},n+1}^{T,i} - R_{\text{sim},n}^{T,i}}{R_{\text{sim},n+1}^{T,i}} \right| < 1\% \quad (16)$$

with n the iteration counter. Note that for synthetic trees arising from the segmented vessel outlets, the dilation propagation stops before reaching the root segment since this was directly observed in the CT data. For synthetic trees arising along a segmented vessel, the

dilation propagation is applied up to and including the root segment. This ensures that only arteriole and small artery vessels are dilated since they contribute most to coronary resistance modulation and are dilated most under hyperemic conditions. Further, the CT data is obtained with the use of sublingual nitrates so the segmented tree is assumed to be fully dilated at rest.

5.2.2 Hyperemic conditions

The minimum resistance at each terminal segment is computed using the ideal terminal segment flow:

$$R_{\min,\text{stress}}^{T,i} = \frac{P_{\text{AO}}}{q_{\text{stress}}^{T,i}} \quad (17)$$

Since the whole system is already dilated to the maximum radius dilation capacity, only terminal flows can be changed. The system is first solved using $q_{\text{stress}}^{T,i}$ as a boundary condition at each terminal segment outlet. With the resulting simulated pressures $p_{\text{stress}}^{T,i}$ and set flows, we calculate the simulated resistance:

$$R_{\text{sim},\text{stress}}^{T,i} = \frac{p_{\text{stress}}^{T,i}}{q_{\text{stress}}^{T,i}} \quad (18)$$

In each iteration, the terminal segment flow is incremented by δq by solving the following equation:

$$(q_{\text{sim},\text{stress}}^{T,i} + \delta q) R_{\min,\text{stress}}^{T,i} = P_{\text{AO}} - R_p^{T,i} (q_{\text{sim},\text{stress}}^{T,i} + \delta q) \quad (19)$$

where $R_p^{T,i} = \frac{P_{\text{AO}} - p_{\text{sim},\text{stress}}^{T,i}}{q_{\text{sim},\text{stress}}^{T,i}}$.

Consequently when the simulated resistance $R_{\text{sim},\text{stress}}^{T,i}$ is less than the minimum resistance $R_{\min,\text{stress}}^{T,i}$, the flow is decreased. Otherwise flow is increased.

Simulation is repeated with the updated terminal flow values. Upon iteration, the computed terminal resistance values converge to the minimum resistance values. The convergence criteria is:

$$\max_i \left| \frac{R_{\text{sim},\text{stress},n}^{T,i} - R_{\min,\text{stress}}^{T,i}}{R_{\min,\text{stress}}^{T,i}} \right| < 1\% \quad (20)$$

5.3 Coupling method

For blood flow, the coupling process involves an initialization loop, followed by iterations coupling the two models. The variable k represents the coupling iteration counter.

During the initialization loop ($k = 0$), 1D equations are solved iteratively to generate possibly dilated geometry with converged flow and pressure fields as described in the parameterization section, either for rest or hyperemic conditions. Initial porous model parameters are determined using the converged flow and pressure values. The converged outlet pressures $p_{k=0}^{\text{T},i}$ are used to estimate the β_{source} and β_{sink} parameters of the myocardium model. The initialized parameters are maintained constant along all following coupling iterations unless otherwise stated.

In the coupling iterations, the resulting pressure of the coronary model at each terminal segment end $p_k^{\text{T},i}$ is used as input to the porous model, defining the source pressure in each tessellation territory Ω_i . The porous model is then solved for flow values for each perfusion territory, that are used as new inputs for the coronary model, $q_{k+1}^{\text{T},i}$. For iterations with $k > 0$, the coronary model solves for pressure with given flows while keeping the same geometry and boundary conditions. The coupling loop is illustrated in Fig. 1d. The coupling convergence is established considering the terminal segment flow values between iteration k and iteration $k + 1$:

$$\left| \frac{q_k^{\text{T},i} - q_{k+1}^{\text{T},i}}{q_k^{\text{T},i}} \right| < 1\% \text{ for all } i \quad (21)$$

In practice for a few cases, in particular the patient with significant obstructive disease, a relaxation scheme was necessary.

Note that in principle, each perfusion territory is assigned to one and only one segment. However in some cases, the scale of the segment outlets is much smaller than the resolution of the myocardial mesh. As a result, during the Voronoi tessellation computation, two segments can be assigned to the same perfusion territory, which can cause problems to the coupling algorithm. In order to avoid this, we locally refine the myocardial mesh and rerun the tessellation. If the issue persists as the segments are too close to each other and an

excessively refined mesh resolution is required, we modify the vasculature by trimming the smallest terminal segment and recompute the tessellation.

5.4 Sensitivity to the synthetic vascular network

In this section, we assess the impact of the vascular network on the results by generating different synthetic vasculatures for the same patient as in section 3.1 (Patient 1).

5.4.1 Varying number of terminal segments

We generated three distinct synthetic vasculatures with 3000, 6000 and 12000 terminal segments, respectively.

Vasculature analysis We evaluate the vascular depth reached in each vasculature using the Strahler order system²⁶. Overall, increasing the number of terminal segments leads to a greater percentage of synthetic segments with lower Strahler orders (5-6) and a lower percentage of segments with higher Strahler orders (7-9) (Fig. 5.1a). This implies that vasculatures with higher number of terminals achieved greater depth. In particular, close to 20% of synthetic segments in the 12000-terminals vasculature were extended down to Strahler order 5, while $< 5\%$ and $< 1\%$ of segments reached that order in the 3000 and 6000-terminal vasculatures, respectively.

Further investigating the diameter distribution of terminal segments reveals that both smaller diameters and greater terminal diameter uniformity is achieved with an increase in number of terminal vessels (Fig. 5.1b). Mean and standard deviation of terminal diameters are $195 \pm 48 \mu\text{m}$, $155 \pm 38 \mu\text{m}$ and $123 \pm 30 \mu\text{m}$ for the 3000, 6000 and 12000 terminal vascular trees, respectively.

Hemodynamic results Blood flow in the three generated vasculatures is summarized in Table 3. In terms of blood flow in the coronary arteries, increasing the number of terminals leads to a more homogeneous flow distribution at the outlet level for both coupled and

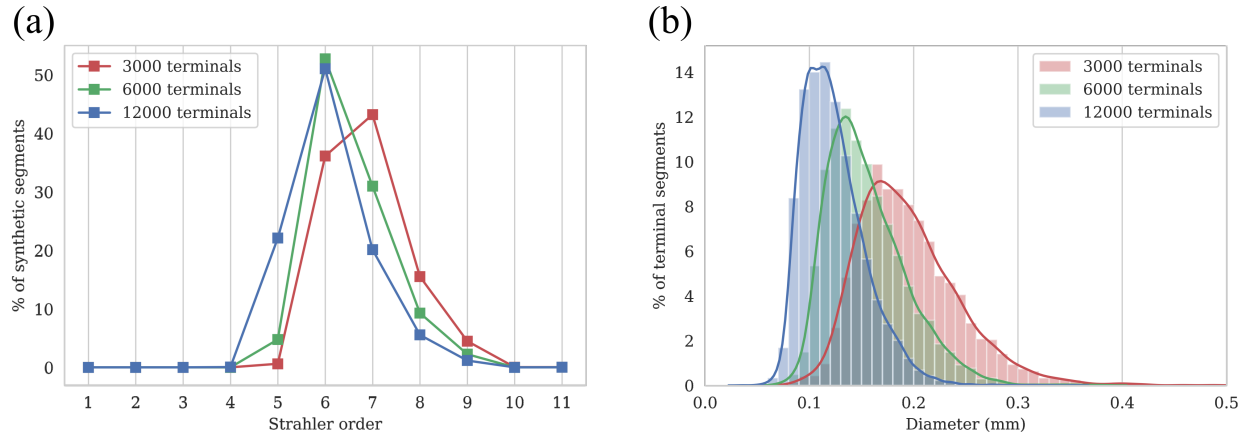


Figure 5.1: Geometrical analysis of vascular networks comprising 3000, 6000 and 12000 terminal segments. (a) Percentage of synthetic segments in each Strahler order. (b) Distribution of terminal segment diameters.

coronary models, as suggested by the lower standard deviation of terminal flows (Table 3). This behaviour reflects the more homogeneous diameter distribution. Naturally, the mean terminal segment flow is also lower, as the same amount of total flow is distributed in more segments.

In terms of blood flow in the myocardium, mean MBF remains almost identical regardless of the number of terminals and the model used. While the coronary model MBF values remain extremely heterogeneous, a slight reduction is observed when increasing the terminal segments (see reduced SD, Table 3). In contrast, the coupled model exhibits the same level of homogeneity regardless of vasculature, showing robustness to the variation in number of terminal segments.

The level of heterogeneity for both models is also illustrated in their respective perfusion maps: coronary model maps consist of significantly over- and under-perfused regions, while the coupled model ones are quite homogeneous. In terms of spatial distribution, although perfusion maps are not identical, the main perfusion features are present across vasculatures (Fig. 5.2).

For all the above considered characteristics, varying the total number of terminal segments has a similar impact on the results in both resting and hyperemic conditions.

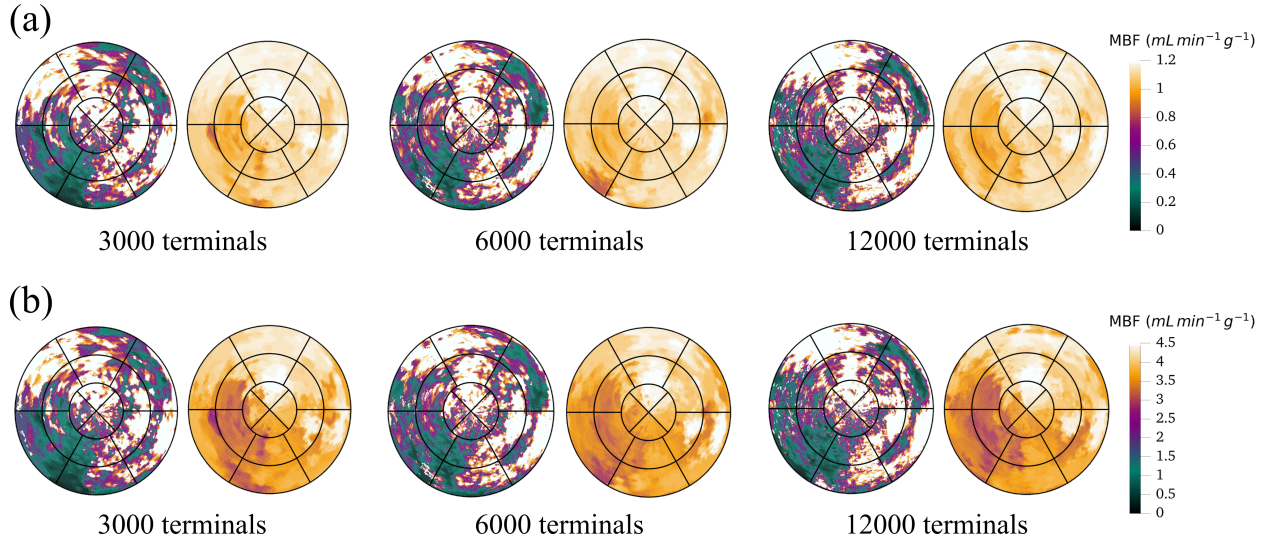


Figure 5.2: Simulated perfusion maps for vasculatures with varying number of terminal segments for (a) resting and (b) hyperemic conditions. For each vasculature: coronary model (left), coupled model (right). Note the upper limit of the color map matches the maximum MBF value of the coupled model across vasculatures. Coronary model MBF values in perfusion maps reach up to $77 \text{ mL min}^{-1} \text{ g}^{-1}$ and $243 \text{ mL min}^{-1} \text{ g}^{-1}$ in resting and hyperemic conditions, respectively, across vasculatures.

To conclude, the 12000-terminals vasculature demonstrates preferable morphological features (greater vascular depth, more uniform outlet diameter size) leading to a minor, albeit noticeable, improvement in the coronary model results. In order to explicitly model as many vessels as possible while maintaining a manageable additional computational cost, we consider 12000 terminal segments as the default choice for this paper.

5.4.2 Varying randomness of the vasculature

Here we generate 5 vasculatures comprising 12000 terminal segments. All synthetic trees start from the same roots on the segmented vessels, but grow differently based on the inherent randomness in the tree generation as described in²³. Thus these 5 "seeds" lead to different but similar-looking synthetic networks.

MBF results for both models across different seeds are presented in Fig. 5.3. The

choice of seed is only significant in the global flow heterogeneity of the coronary model (Fig. 5.3a, 5.3b). In fact, choosing a particular seed has a larger impact on the standard deviation of simulated MBF for the coronary model, compared to varying the number of terminal segments. The coupled model provides consistent results regardless of the seed, demonstrating robustness in the geometrical variability of the vasculature.

The associated perfusion maps display some regional differences, but their spatial distribution of flow has similar main characteristics (Fig. 5.3c). For example, regions from 7 to 9 o'clock consistently have lower MBF compared to the rest of the map. Results are similar for resting and hyperemic conditions.

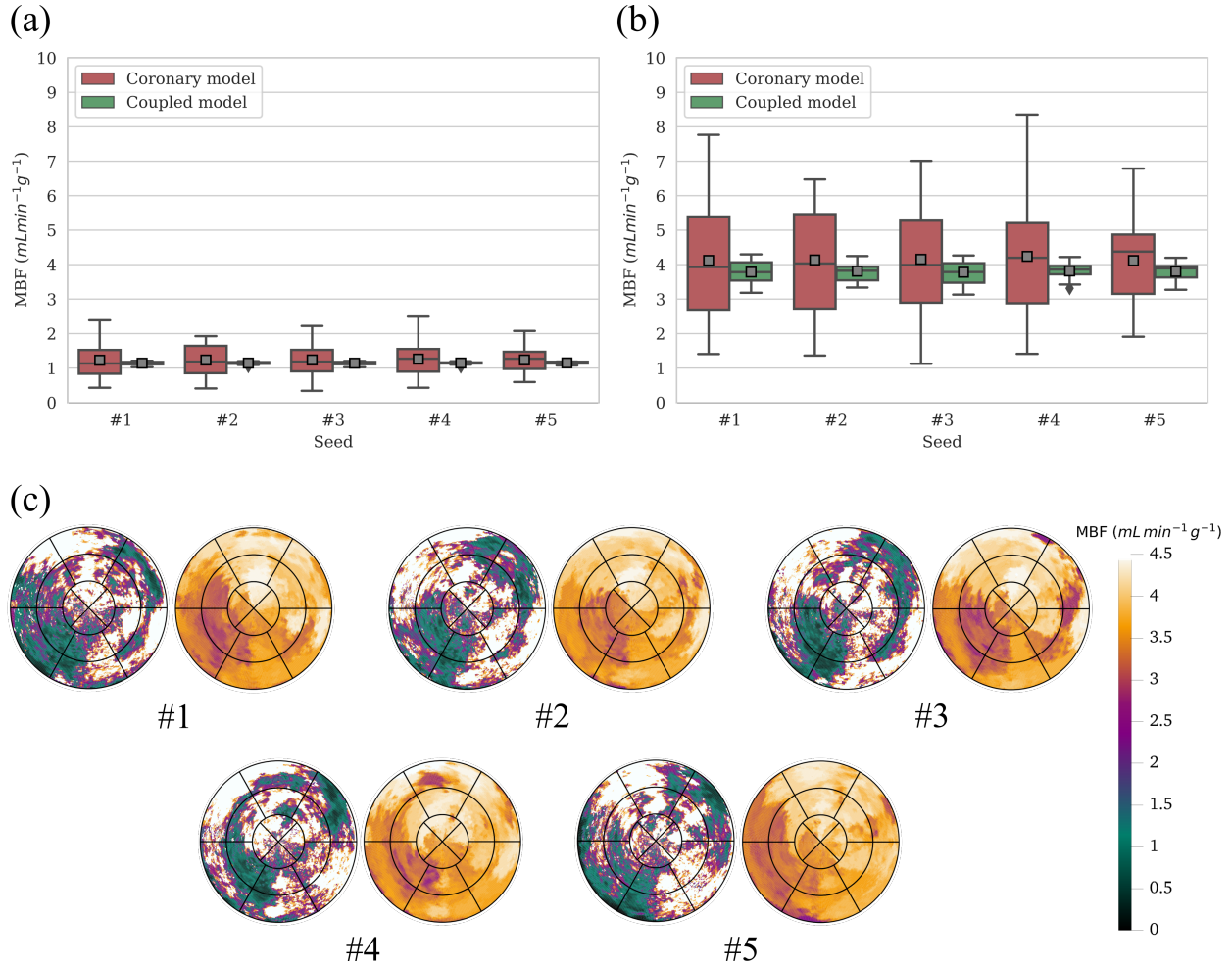


Figure 5.3: MBF_{AHA} results for 5 random seeds for (a) resting and (b) hyperemic conditions. Mean MBF_{AHA} values are depicted with grey squares. (c) Simulated perfusion maps for hyperemic conditions. For each seed: coronary model map (left), coupled model map (right). Note the upper limit of the color map matches the maximum MBF value of the coupled model across seeds. Coronary model MBF values in perfusion maps reach up to $225 \text{ mL min}^{-1} \text{ g}^{-1}$ across seeds.

5.5 Patient selection and characteristics

For this study, a total of 6 patients were selected to have high cCTA image quality and contain a wide distribution of heart dominance (3 right, 2 codominant, 1 left). They are grouped based on non-obstructive CAD (5 patients) or obstructive CAD (1 patient), with obstructive CAD defined as $> 30\%$ reduction in diameter in at least one location on the larger segmented vessels.

Table 4: Patient characteristics of 6 patients with suspected CAD who underwent cCTA and [^{15}O]H $_2$ O Positron Emission Tomography (PET)¹³ prior to invasive coronary angiography, which demonstrated non-obstructive CAD in 5 patients and obstructive CAD in 1 patient. ACE = angiotensin-converting enzyme; ARB = angiotensin receptor blocker.

	Patient 1	Patient 2	Patient 3	Patient 4	Patient 5	Patient with obstructive CAD
Age	54	59	59	56	46	53
Sex	female	female	male	male	female	male
Body mass index (kg m ⁻²)	22.95	28.08	27.90	20.38	29.32	25.93
Heart dominance	right	right	codominant	codominant	left	right
<i>Comorbidities</i>						
Current smoker	No	No	No	Yes	No	No
Smoking history	No	No	No	Yes	No	Yes
Diabetes (type I)	No	No	No	No	No	No
Diabetes (type II)	No	No	No	No	No	No
Hypertension	Yes	No	No	No	No	No
Hypercholesterolemia	No	Yes	No	No	No	Yes
Family history of CAD	Yes	No	No	Yes	Yes	No
<i>Medications</i>						
Acetylic acid	Yes	Yes	Yes	Yes	No	Yes
Statin	Yes	Yes	Yes	Yes	No	Yes
ACE inhibitors	No	No	Yes	No	No	Yes
ARBs	Yes	No	No	No	No	No
Long acting nitrates	Yes	No	No	No	No	No
Beta blockers	Yes	No	No	Yes	No	Yes
Calcium channel blockers	Yes	No	No	No	No	Yes
Symptoms	atypical angina	aspecific chest pain	atypical angina	typical angina	aspecific chest pain	typical angina

5.6 Additional illustrations

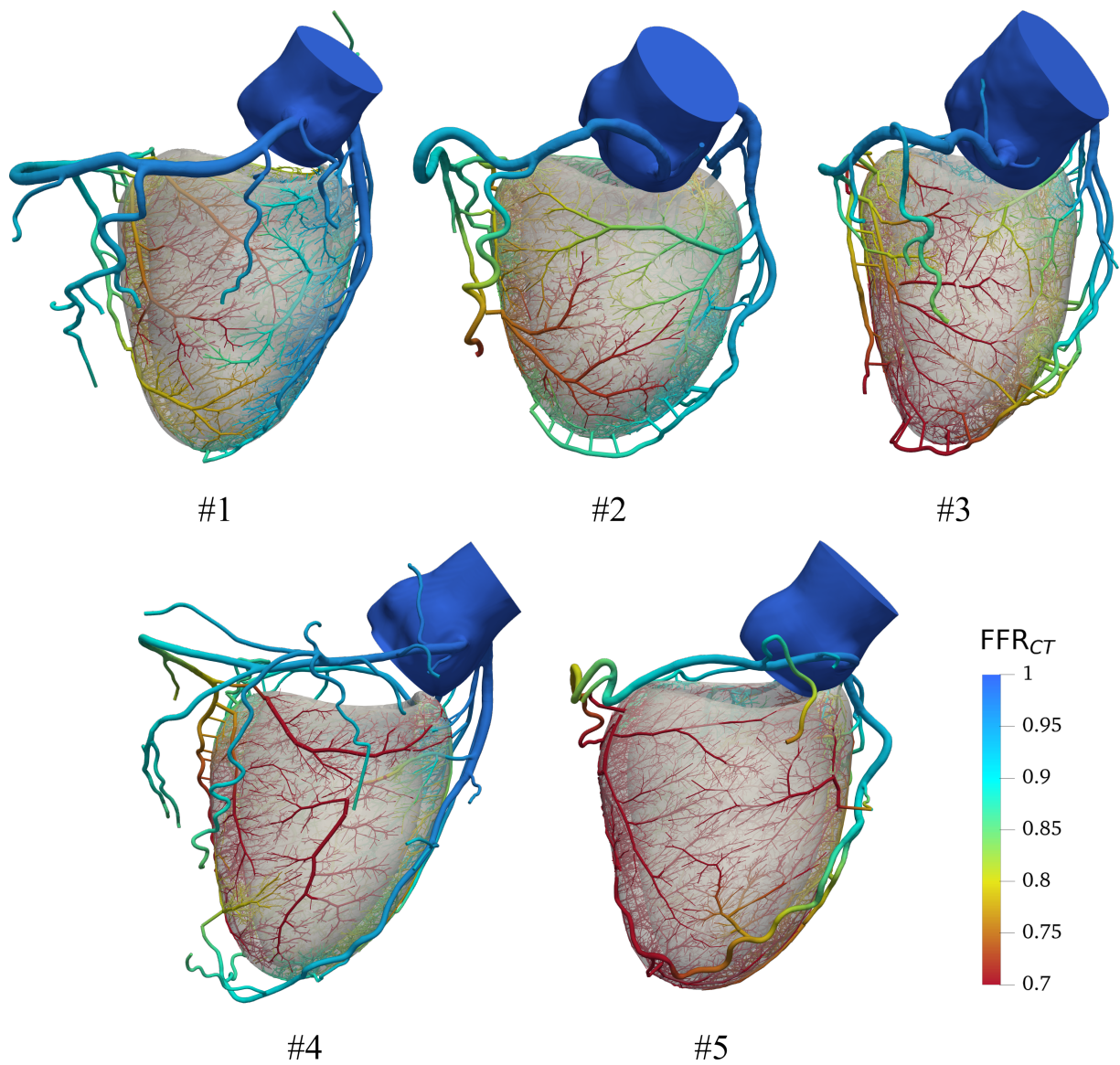


Figure 5.4: FFR_{CT} analysis along the segmented and synthetic vasculature for 5 patients with non-obstructive CAD (hyperemic conditions).

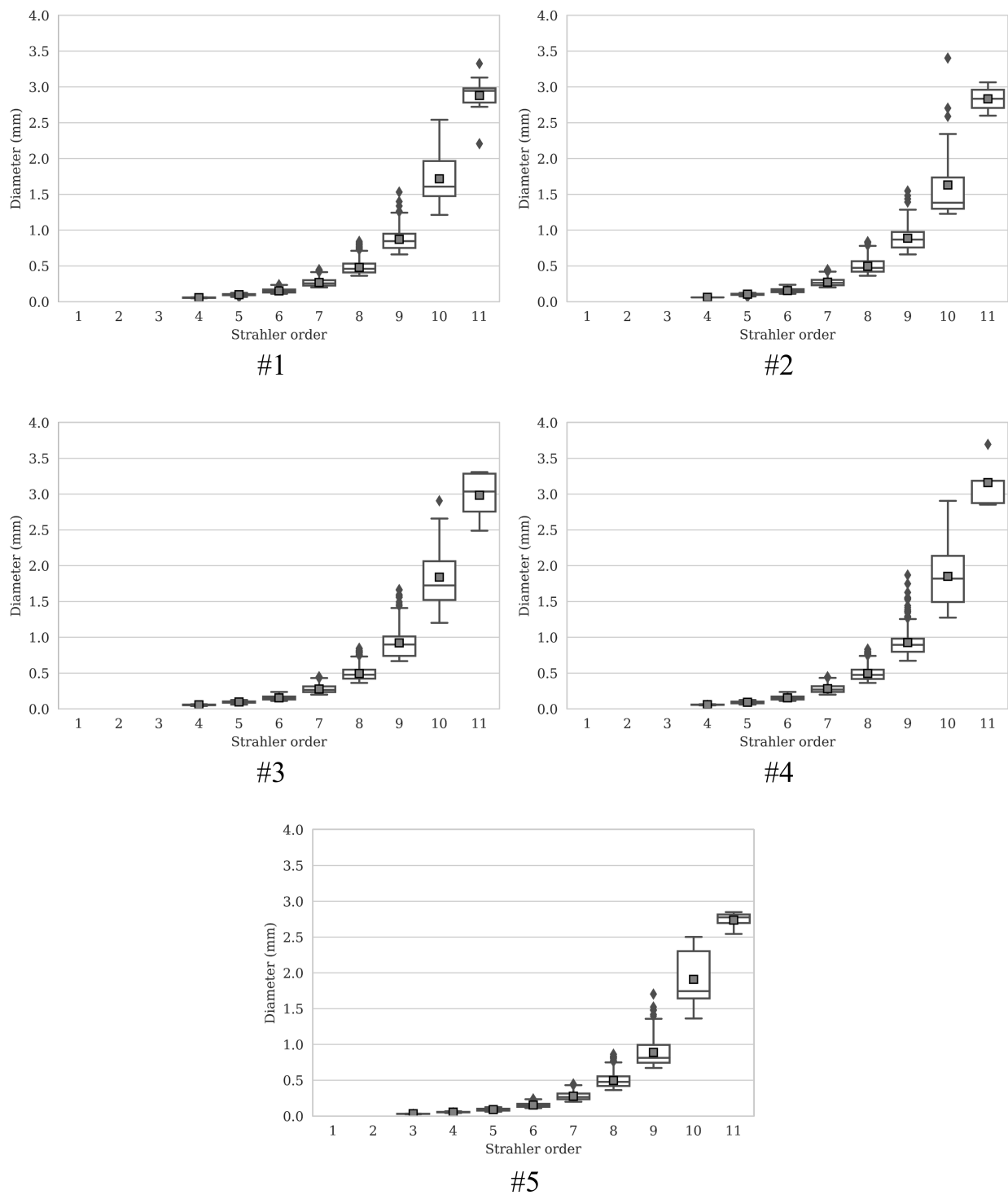


Figure 5.5: Diameter distribution of segmented vessels (orders 9-11) and undilated synthetic segments (orders 3-9) within each Strahler order for five patients with non-obstructive CAD. Vasculatures comprise 12000 terminal segments.