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31P MRS assessments of mitochondrial dysfunction in patients with peripheral arterial disease undergoing revascularization.

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Synopsis

Obliterative arterial disease of the lower limbs is a disease that obstructs lower extremities arteries, resulting in reduced lower limb perfusion and possibly mitochondrial dysfunction. Mitochondrial function of the calf assessed via 31P MRS at moderate and low exercise intensities before and after revascularization and phase contrast angiography of the posterior tibial artery enabled the assessment of vascular and mitochondrial contributions of the patients.

Introduction

Obliterative arterial disease of the lower limbs also known as peripheral arterial occlusive disease (PAOD) is defined as partial or total obstruction of one, or more, lower extremity arteries most often of atherosclerotic origin [1,2]. It is a common pathology whose five-year mortality of a patient with PAOD is about 30%, mainly of cardiovascular origin. In addition, most of the literature suggests that lower extremity mitochondrial function is severely reduced in PAOD. The main objective is to study skeletal muscle mitochondrial function by 31P spectroscopy (τ PCr) before and after revascularization in patients with PAOD using an exercise paradigm that approaches the ischemic stress stage and

using an alternative exercise paradigm that is well below ischemic stress[3,4].

This approach allow insight into the relative contribution of the vascular and mitochondrial components to the degradation of muscle function in PAOD. The overall hypothesis was that slowed τ PCr in PAOD is the result of reduced skeletal muscle perfusion. Successful revascularization should be accompanied by an immediate improvement of the τ PCr in the absence of mitochondriopathy. On the other hand, in the case of mitochondrial disease, the τ PCr should remain altered in early post-revascularization and despite the lifting of the vascular factor.

Methods

Experimental protocol

Four patients (male, age=60±6 yrs old, mean±SD) were measured before and after revascularization. Each patient was supine on the MR bed of the scanner, with a surface coil 31P/1H Tx/Rx under the calf muscle.

For dynamic measurements, an MR compatible ergometer was used during isometric plantar flexion. MR scans were performed using a non-localized, MR-FID sequence with saturation bands done with adiabatic pulses[5].

Maximum Voluntary Contraction (MVC) force were performed for 1-2 s. Prior localization was done with a dixon sequence (FA 9deg , TR 4.1ms, TE 1.4ms, matrix size 118×192 , FOV 169×206). Thus, resting 31P acquisitions were done with a 90deg FA, TR 30s,10 averages, bandwidth 2500Hz, and saturation bands placed on anterior muscles. Then, 2 dynamic protocols were performed with TR 4s, but with different dynamic resolution times (time between two exercises), see figure 1. Thus, a moderate intensity exercise protocol, expected to approach the ischemic stage as well as a low intensity gated exercise protocol that would be well above the ischemic limit were performed by each subject before and after revascularization. A total of 30 contractions were performed in the moderate protocol at a rate of 0.25 Hz and a total of 15 contractions were performed in the gated protocol at a rate of 0.05 Hz. In addition, resting flow in the posterior tibial artery was assess with ECG gated phased contrast MR angiography.

Data Processing

Data was processed using CSIAPO software[7] for phasing, MATLAB(The MathWorks) and the method QUEST(QUantitation based on QUantum ESTimation)[8]. For the gated protocol, the last 10 contractions cycles were averaged together, resulting in 10 points during the contraction/recovery cycle. In addition, 10 spectra were averaged over the initial rest. The following formula was used to estimate τ PCr gated:

$$\tau = -\Delta t / \ln[D / (D + Q)],$$

where D represents the PCr drop to the steady state (rest PCr–max PCr after steady state is reached), Q is the PCr change in the steady state (max PCr in the steady state–minimum PCr), and t=time between contractions[6]. pH was determined using the frequency difference between PCr and Pi as:

$$\text{pH} = \text{pKa1} + \log((\text{FreqPi} - \text{FreqPCr}) - \text{pKaM}) / \text{pKaM} - (\text{FreqPi} - \text{FreqPCr}),$$

with pKa1=6.75, pKaM=3.27, pKaM=5.69 and FreqPCr and FreqPi the resonant frequencies of PCr

and Pi[9].

For blood velocity and flow analyses, velocity was extracted from each time point across the cardiac cycle. Flow was the product of the vessel CSA and velocity averaged across the vessel. Data were compared using paired t-tests to examine differences between exercise protocol and between measurement times with significance at $p < 0.05$.

Results

Figure 2 shows a sample stack plot of spectra from the moderate exercise protocol and the low intensity gated protocol; Table 1 reports the MRS results. In general, the PCr recovery time constant was substantially longer for the moderate compared to the gated exercise protocol before surgery ($\tau = 72.2s$ vs. $34.5s$, respectively, $p = 0.012$). Following surgery, there was a 33% improvement of the τ_{PCr} for the moderate protocol ($p = 0.039$). However, there was no significant improvement in τ_{PCr} for the gated protocol. Blood flow parameters are reported in Table 1. The arterial waveforms are shown in Figure 3. The waveforms in 3 cases are characterized as monophasic. Following revascularization, there was an apparent increase in peak blood velocity that approached significance. This was particularly the case for Subj01 and Subj03 who adopted a biphasic arterial waveform following revascularization.

Discussion

The results primarily indicate that blood flow is a large determinant of the measured PCr time constant under standard exercise conditions. This is demonstrated both when comparing the moderate to the low intensity protocol and when examining the moderate exercise PCr time constant post-surgery. These results encourage the quantification of blood flow when assessing mitochondrial function in conditions of compromised flow. Alternatively, a “gated” low intensity protocol may be utilized to independently assess mitochondrial function in conditions of compromised blood flow, oxygen delivery, or oxygen consumption.

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