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# A Selective Sulfide Oxidation catalyzed by Heterogeneous Artificial Metalloenzymes Iron@NikA.

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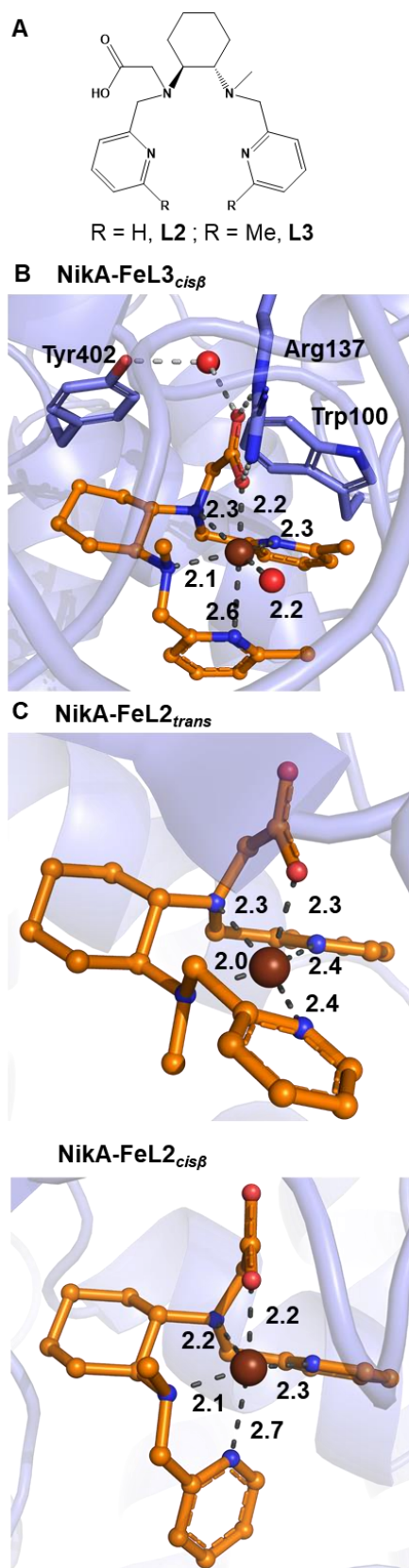
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**Abstract:** Performing a heterogeneous catalysis with proteins is still a challenge. Here, we demonstrate the importance of cross-linked crystals for sulfoxide oxidation by an artificial enzyme. The biohybrid consists of the insertion of an iron complex into a NikA protein crystal. The heterogeneous catalysts displays a better efficiency-with higher reaction kinetics, a better stability and expand the substrate scope compared to its solution counterpart. Designing crystalline artificial enzymes represents a good alternative to soluble or supported enzymes for the future of synthetic biology.

**Introduction.** Sustainable chemistry implies the discovery of new efficient but non-toxic solutions to synthesize molecules following eco-friendly strategies.<sup>[1]</sup> Biocatalysis, boosted by the development of synthetic biology, represents a great opportunity to fulfill the requirements of green chemistry but still suffers from comparisons with the metal-based chemical catalysis.<sup>[2]</sup> Biocatalysis and metal-based catalysis are two highly dynamic fields of research, ranging from enzyme and *in cellulo* processes to homogeneous or heterogeneous catalysis. Currently, each approach possesses advantages such as the mastering of chemo, regio and stereoselectivity and catalytic efficiency for enzymes while heterogeneous inorganic catalysts provide a better stability and recyclability. In order to optimize catalytic processes, enzymes can be engineered to gain stability and substrate promiscuity while MOFs, for example, are more and more sophisticated in order to allow the substrate control required to reach the expected selectivity. While natural enzymes have been engineered to gain higher activity<sup>[3]</sup> or been repurposed for new reactivities,<sup>[4]</sup> artificial enzymes issued of the seminal work of Whitesides *et al.*,<sup>[5]</sup> offer the opportunity to fill the gap between biocatalysis and chemical catalysis, providing new solutions for abiotic reactions with a special focus on stability.<sup>[6]</sup>

These hybrid systems are made of two building blocks, a protein chosen for its capacity to host tightly an unnatural active site, usually an inorganic catalyst. Both parts of these artificial enzymes have their own functionality, which is the control of the reaction selectivity for the protein and the nature of the reaction for the inorganic complex. This exciting strategy is eager to reproduce enzyme properties, in particular the kinetic and the enantioselectivity of a reaction. Stereoselectivity is nowadays well documented and sulfoxidation reaction is often a benchmark of hybrid systems as chiral sulfoxides centers are of great interest in the pharmaceutical industry, with the worldwide famous drugs Omeprazole® or Modafinil®.<sup>[7]</sup> So far, only two systems have been engineered for API syntheses.<sup>[8]</sup> Accordingly, the design of artificial sulfide oxygenases has been developed, differentiated by the mode of synthesis of the hybrid systems. First, the *de novo* active site approach consists of stabilizing the exogenous molecules (mostly Schiff bases complexes), by supramolecular interactions with the amino acids present inside a protein cavity (such as the Ni importer NikA,<sup>[9]</sup> Xylanase 10 A,<sup>[10]</sup> serum albumins<sup>[11]</sup> or phytase).<sup>[12]</sup> The Trojan horse strategy uses the same mode of interaction but with a natural substrate of the protein playing the role of an anchor. The latter is modified by a coordination moiety, as successfully demonstrated with streptavidin,<sup>[13]</sup> BSA<sup>[14]</sup> or antibodies against steroids (testosterone) and the protein neocarzinostatin.<sup>[15]</sup> Second, dative interactions between vanadate anions, iron or Mn and an amino acid is also demonstrated in the case of myoglobin<sup>[16]</sup> or streptavidin.<sup>[17]</sup> To ensure the stabilization of the non-natural active site in the cavity, a third approach uses covalent bonding via cysteine residues from engineered myoglobin or  $\beta$ -lactoglobulin as protein target.<sup>[18]</sup> Finally, enzyme from scratch design is illustrated by the mimochrome series, in which the peptide environment is synthesized chemically and result of peptide self-assembly processes.<sup>[19]</sup> All these systems provided from good to excellent selectivity for sulfoxide formation (up to 100%) and stereoselectivity (up to 94% for serum albumins/Mn Schiff bases and streptavidin/ $\text{VO}_4^{2-}$ ) while yields oscillated between 50% and 100%. The major difference between all these systems lies on the number of TON, a criterion for stability. A TON over 1 000 was measured only in two cases, the mimochrome Mn VIa<sup>[19]</sup> and FeLiBu/BSA.<sup>[14]</sup> So far, industrial criteria for a use of artificial enzymes have not been reached and there is still a need for improvement. As observed in the field of chemical catalysis, the stability is increased when the molecular catalyst is grafted on surfaces and a similar approach can be used for enzymes. Several strategies exist from immobilization in/on MOFs or grafting on nanoparticles to encapsulation in a polymer matrix, micelle or liposome.<sup>[20]</sup> One attractive strategy, *i.e.* the use of cross-linked enzyme crystals (CLEC), is quite astonishing because of its various assets. On the one hand, protein crystals achieve the highest enzyme concentration while preserving its integrity. In addition, the high solvent content affords the advantages of microporous materials, allowing the easy diffusion of molecules through solvent channels. On the other hand, with the CLEC technology, the intermolecular lattice contacts within the crystals and chemical cross-links formed with bifunctional reagents such as glutaraldehyde stabilizes enzymes.



**Figure 1.** **A:** Structures of ligands **L2** and **L3**. **B:** Crystal structure of NikA-FeL3 binding site. **FeL3** under the only *cisβ* topology observed in this study. **C:** Crystal structure of NikA-FeL2 binding site. **FeL2** under either the *trans* or the *cisβ* topology, as observed in a previous study.<sup>[21]</sup> Fe is depicted as a brown sphere. Distances are in Å.

Remarkably, CLEC, as an immobilization technique, represents a great option to give access to biocatalysis in environments that are otherwise mostly incompatible with enzyme function, including prolonged exposure to high temperatures, extreme pH, near-anhydrous organic solvents and aqueous-organic solvent mixtures.<sup>[22]</sup> Moreover, they can generally be stored indefinitely at room temperature.<sup>[23]</sup> Up to now, the CLEC technology has been benchmarked as industrial biocatalysts for reductase or hydrolase activities but its application to redox metalloenzymes remains to be explored further. Notably, only Ueno *et al.* and our group have independently

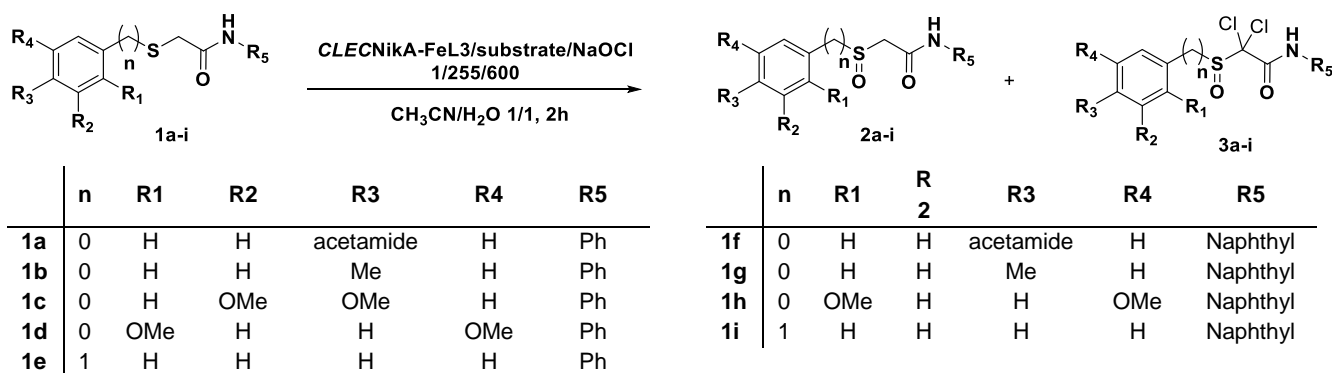
designed artificial metalloenzymes *in crystallo* for alkene transformations and O<sub>2</sub> activation, showing the high impact of a crystalline hybrid catalyst on stability and activity.<sup>[24]</sup>

In order to benchmark this approach on thioether sulfoxidation, we designed cross-linked artificial sulfide oxidases *in crystallo*, based on our previous results obtained in solution, by the incorporation of iron complexes with nitrogen-based ligands into NikA. In this study, the results revealed the better catalytic efficiencies of heterogeneous *CLECNika-FeLn* compared to their analogs in solution (*soNika-FeLn*) on thioether oxidation, and particularly the stability of the *CLECNika-FeL3* system.

**Results and discussion.** Previously, we designed an artificial system that was capable of sulfoxidation in HEPES buffer solution. Thanks to a screening approach, sulfide molecules able to bind to the cavity of the nickel-binding protein NikA were selected. Among them, a series of substrates with the R1-S-CH<sub>2</sub>-CONH-R2 motif (S- and N-substituted thioglycolamides, Scheme 1) have been transformed into their sulfoxides with high chemoselectivity, a TON of 199 and a TOF of 0.7 min<sup>-1</sup> for the substrate **1b** in buffered solution.<sup>[9]</sup> The reaction was catalyzed by the artificial enzyme *soNika-FeL2*, synthesized by the incorporation of an iron N2Py2 complex **FeL2** into the NikA scaffold. **L2** is a variation of the BPMCN ligand<sup>[25]</sup> obtained by the replacement of one N-methyl moiety by a glycine moiety (Figure 1A, R = H). This modification ensures the stabilization of the complex **FeL2** (with a K<sub>d</sub> in the μM range) inside the NikA cavity *via* a salt bridge between the Arg137 and the carboxylate moiety.<sup>[21]</sup> The X-ray structure of the hybrid revealed that the complex adopted different topologies, oscillating between a *trans* and a *cisβ* topology (differing by the pyridine ring positions to each other, Figure 1C). In the *trans* topology (Figure 1C, top), the two pyridines are *cis* to each other in the equatorial plane of the complex while in the *cisβ* (Figure 1C, bottom), the two pyridine rings coordinate *cis* to each other but in perpendicular planes. This last topology has been observed in [6-Me<sub>2</sub>-BPMCN](OTf)<sub>2</sub> (where OTf is trifluoromethanesulfonate anion).<sup>[25]</sup> This mixture of topologies is deleterious to envisage an efficient metal based enantioselective control. This urged us to synthesize the dimethyl 6-Me<sub>2</sub>-BPMCN<sup>[25]</sup> variant, **FeL3** (Figure 1A, R = Me).

The ligand **L3** and the complex syntheses are described in the supplementary material. It has to be noted that the ferrous complex appeared more appropriate to ensure its stabilization in the NikA cavity. Actually, compared to the ferric state, the ferrous state, with a charge of +1 should facilitate the insertion of the complex into the positively charged NikA binding site. Accordingly, the soaking was performed under an inert atmosphere. The complex **FeL3** displayed a UV-visible spectrum characteristic of a ferrous ion featuring a unique charge transfer band in the visible region at 365 nm, attributed to a transition from the Fe(II) “t2g” orbitals to the π\* pyridine orbitals (Figure S1).<sup>[26]</sup> The small absorption coefficient (ε = 750 L.mol<sup>-1</sup>.cm<sup>-1</sup>) indicates an iron high-spin state, S=2. The complex was also identified by ESI-MS with a single positive fragment at m/z 451 (100%) corresponding to a [**L3**-H+Fe(II)]<sup>+</sup> ion (Figure S2).

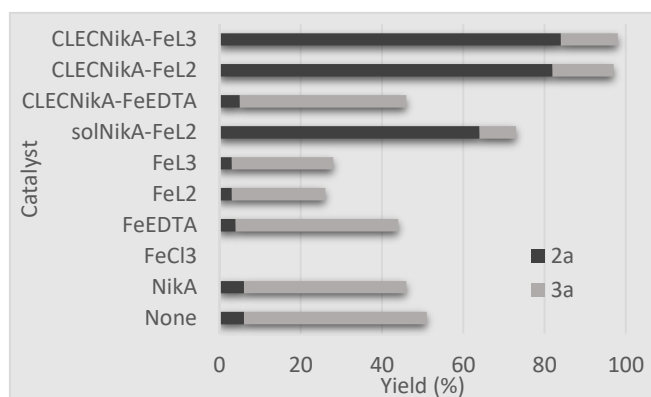
The X-ray structure of NikA-**FeL3** was solved at 1.9 Å resolution. Crystallographic data statistics are summarized in table S1. Two molecules are present in the asymmetric unit (called A and B). In both molecules, the ligand **L3** is fully occupied while Fe(II) is only partly occupied (with an occupancy of about 0.7). For clarity, only molecule A will be discussed for the description of **FeL3** binding mode in NikA cavity. In contrast to NikA-**FeL2** (Figure 1C), the X-ray structure revealed that only one ligand conformation, the *cisβ* one, was observed (Figure 1B). The bulkiness of the methyl group on the pyridine rings has then led to the destabilization of the *trans* topology observed previously for NikA-**FeL2**. Depending on the structures, the Fe(II) ion is either pentacoordinated or hexacoordinated by two nitrogen atoms from the amino groups (Fe-N = 2.1 and 2.3 Å), one oxygen atom from the carboxylate group (Fe-O = 2.2 Å) and one nitrogen atoms from one pyridine ring (Fe-N = 2.3 Å). When Fe(II) is hexacoordinated, the nitrogen atom from the pyridine ring *trans* to the carboxylate group is bound to Fe(II) with a Fe-N distance of 2.6 Å. This pyridine group is flexible (with a B-factor of about 60) rendering the electron density map difficult to model and highlighting the lability of this coordination bond (Figure S3). Whatever the coordination number of the metal is, a water molecule binds to the iron (Fe-OH<sub>2</sub> = 2.2 Å). As previously observed in all structures of NikA/metal complexes hybrids described so far, **FeL3** is mainly stabilized in the protein pocket by a salt bridge between the carboxylate group of **L3** and Arg137. In addition, an interaction with a structural water molecule made one donating H-bond with the oxygen atom of the carboxylate group involved in the salt bridge with Arg137, one accepting bond with the oxygen OH of Tyr402 and one donating H-bond with the main chain O of Arg137. As in the case of NikA-**FeL2**<sub>*cisβ*</sub> structure, one π-stacking interaction was also observed between one pyridyl group and the Trp100 side chain. Interestingly, in both structure, the ligand topology led to an elongation of the Fe-N<sub>pyridine</sub> distance *trans* to the carboxylate, underlining a suspected *trans* effect of the anion. This effect is also combined with its position in an open space, excluding any interactions with the protein structure.



**Scheme 1.** Details of the sulfoxidation reaction and the substrates used in this study.

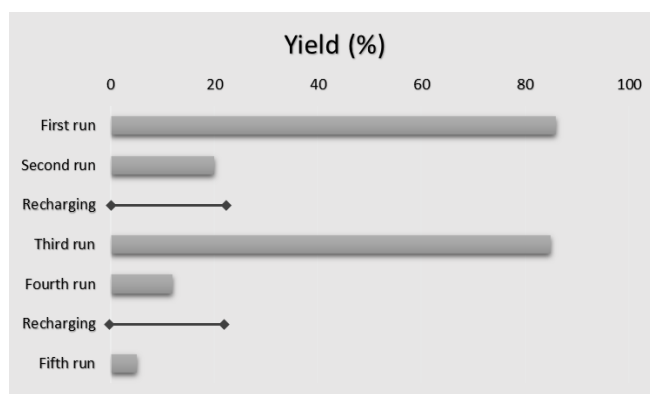
Confident that the complex was stabilized in NikA crystals, the CLEC synthesis was then undertaken in two steps. First, NikA-FeEDTA crystals were soaked with either **FeL3** or **FeL2** for four days in a glove box. The crystals were then soaked with a solution of glutaraldehyde for 5 hours to yield *CLECNiKA-FeL2* and *CLECNiKA-FeL3*, which were subsequently washed and stored in a 1:1 solution of  $\text{CH}_3\text{CN}:\text{H}_2\text{O}$ .

To ascertain the role of the crystalline material on the catalysis, we have first undertaken to use the substrates of our earlier study in 2013<sup>[9]</sup> under the similar reaction conditions but the solvent, here a mixture of aerated water and acetonitrile (Scheme 1). This medium will then help to dissolve water-insoluble substrates, in order to evaluate the substrate scope of the biohybrid catalysts. Actually, *CLECNiKA-FeL2* and *CLECNiKA-FeL3* (100 CLEC, 31  $\mu\text{M}$ ) catalyzed the oxidation by NaOCl of a series of nine thioglycolamides **1a-i**. *CLECNiKA-FeLn* were compared to the complexes alone and *soNikA-FeL2* for **1a** oxidation.<sup>[9]</sup> For each biohybrid, protein concentration was measured either by Size Exclusion Chromatography coupled to Multi-Angle-Laser-Light-Scattering (SEC-MALLS) or by using the Rose Bengal protocol.<sup>[27]</sup> The metal content was measured by ICP-AES and the metal/protein ratio was closed to one for all biohybrids. The determination of X-ray structures of the different NikA/complexes hybrids demonstrated the full occupancy of **FeLn** inside NikA molecules (see supplementary material).



**Figure 2.** **1a** oxidation by various catalysts under the following conditions: Catalyst:NaOCl:**1a** : 1:255:600; [cat] = 37  $\mu\text{M}$  in HEPES 10 mM pH 7; 2 hours except for CLEC [cat] = 31  $\mu\text{M}$  in  $\text{CH}_3\text{CN}:\text{H}_2\text{O}$  1/1 v:v ; 2 hours

First, the uncatalyzed reaction led to a small amount of sulfoxide (less than 5%) and an achiral formation of the dichlorinated product **3a** (up to 5%) (Scheme 1). A similar lack of reactivity was observed with any iron salts or the **FeLn** complexes alone. It has to be noted that the protein NikA itself is inactive, while other biomolecules such as phytase or serum albumin produced sulfoxides.<sup>[11b, 28]</sup> Conversely, the *soNikA-FeL2* hybrid, in buffered solution, produced sulfoxide **2a** with a moderate yield and high selectivity (64% (163 TON) and 90%, respectively) (Table S2). The protein host helps to greatly annihilating the production of **3a**, causes a drastic gain in selectivity and a twenty-fold increase in yield. Second, oxidation of **1b** was performed with *soNikA-FeL2* and *soNikA-FeL3*. **2b** was formed with a slight difference in yields, 63% versus 50%, respectively. Third, oxidation of **1a** was performed with *CLECNiKA-FeL2* and *CLECNiKA-FeL3*. Using comparable catalyst concentrations, the yield in sulfoxide **2a** was increased by nearly 30% (82% (209 TON) and 84% (214 TON), for *CLECNiKA-FeL2* and *CLECNiKA-FeL3*, respectively) and the selectivity was kept (85 vs 90%). Higher efficiencies were then obtained when switching to CLEC with the reference substrate. It is also proven that the complex **FeLn** controls the reactivity, as the *CLECNiKA-FeEDTA* was fully inactive. The similar results obtained for both *CLECNiKA-FeL2* and *CLECNiKA-FeL3* indicate that the methyl substitution on the pyridine ring were not deleterious for the reactivity. However, the presence of only one ligand topology for *CLECNiKA-FeL3* has no effect at all on the enantioselectivity of the sulfoxidation since an enantioselective excess of only 10% was observed whatever the complex is, as in solution.



**Figure 3.** Yields in **2a** during recharging experiment with *CLECNika-FeL3* as catalyst and **1a** as substrate in standard experimental conditions (100 CLEC).

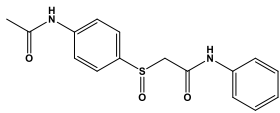
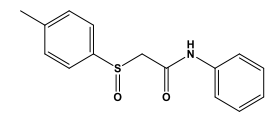
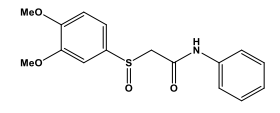
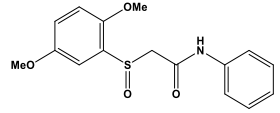
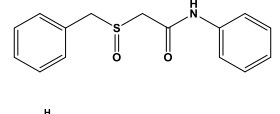
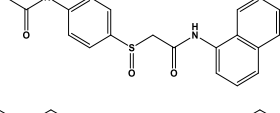
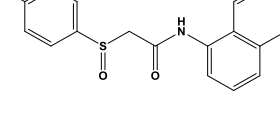
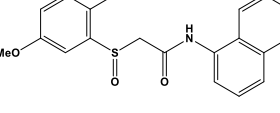
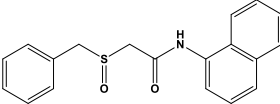
The *CLECNika-FeL3* stability was also evaluated (Figure 3). The **1a** transformation was tested with a batch of 100 CLEC. At the end of the first run, substrate and product were extracted and the CLEC were washed twice with a solution of CH<sub>3</sub>CN:H<sub>2</sub>O. After the removal of the supernatant, a fresh solution of substrate and oxidant was added and a second catalytic run was performed. A drop from 86% to 20% of the sulfoxide yield was observed, attesting that the catalytic system is rather unstable under harsh oxidative conditions. To discriminate between the catalyst and the protein degradation, after the second run, a reloading step was done by soaking *in situ* the *CLECNika-FeL3* with a fresh **FeL3** solution. A third catalytic run was then performed leading to a yield of 84% in **2a**. The full restored activity cannot be attributed to the complex alone in solution (see Figure 2, **FeL3** complex activity is only 3%), suggesting that active CLEC were recovered and that the iron complex is degraded at first while the NikA protein scaffold seems less impacted. Nevertheless, after a new consecutive run, as expected, the activity dropped again but could not be restored after CLEC soaking with **FeL3**. Therefore, the NaOCl is too harsh for the complex and the protein, leading to a moderate stability. The use of other oxidants such as hydrogen peroxide (even in the presence of acetic acid)<sup>[29]</sup> or *tert*-butyl hydroperoxide, were not conclusive with both lower yields and slower kinetics. The results nevertheless displayed an activity higher than most of the other hybrid systems in solution reported in the literature, with a total of 470 TON obtained after the fourth run. It has to be mentioned that in similar conditions but in solution, a second run did not yield any product.

In addition, the CLEC strategy allowed us to expand the substrate scope to water-insoluble substrates (Table 1), using a 15 μM of *CLECNika-FeL3* (50 CLEC) as catalyst for screening. It has to be noted that in these experimental conditions, **1a** conversion after 2h was less efficient (15% instead of 84%) as the catalyst concentration was half of the amount used above. This nearly six-fold reduced yield was used as reference for comparison. We observed that the *CLECNika-FeL3* is efficient for **1a-d** oxidation with a high selectivity for sulfoxide formation, reproducing the trends observed in solution with the hybrid *soNika-FeL2* (**1b** still more reactive than **1a**, **1c** or **1d**),<sup>[9]</sup> suggesting an electrophilic character of the oxidizing species. This is also confirmed by the use of a 4-nitrophenyl-*N*-phenylthioglycolamide derivative, which was reluctant to any transformation under oxidative conditions. The lower transformation of **1c** and **1d**, compared to *soNika-FeL2* in solution,<sup>[9]</sup> strongly suggests that steric effects related to the methoxy groups are more important in the solid state since both substrates were totally soluble.

Thanks to the stability of the CLEC toward organic solvents, more lipophilic substrates **1e-1i** were then targeted. This new catalytic system allows now to oxidize less enriched thioethers (and aliphatic) like **1e**. Interestingly, the use of a CH<sub>3</sub>CN : H<sub>2</sub>O allowed the solubilization of naphthyl moiety containing substrates **1f-i**. Comparing **1f** to **1a**, the presence of a naphthyl versus a phenyl moiety strongly decreases the reactivity and steric effects can be evoked. This is also confirmed by comparing **1b** and **1g** with a yield divided by 2 after 2 hours. Nevertheless, **2g** and **2i** were obtained with an efficiency at the level of **2a**. The best yield is observed for **2e**, related again to steric effects. The best yields obtained for **2b** and **2e** then pushed us to test the catalytic system under a 31 μM concentration for a direct comparison with the standard reaction conditions (Figure 2, Table S2). The results after two hours gave a 100% selectivity and a yield of 80% (204 TON) and 89% (227 TON) for **2b** and **2e**, respectively, in the line with the results obtained for **2a**. The test the limits of the catalyst, **1b** was oxidized with only 0.09% of catalyst, giving a slightly increased yield (50%) of **2b** but a 8 fold increased TON (570). To conclude, by switching from homogeneous to heterogeneous conditions, we have then afforded a larger promiscuity for substrates.

Finally, in addition to the role of CLEC on substrate promiscuity, the most important output of this study is the kinetics of the S1 oxidation. We estimated a 5-fold increase of the sulfoxide turnover frequency to 3.7 min<sup>-1</sup> against 0.7 min<sup>-1</sup> in solution under standard conditions. In addition, we measured the kinetics when H<sub>2</sub>O<sub>2</sub> was used as the oxidant, taking advantage of the slower reactivity. We then measured a 3 fold increase (0.34 min<sup>-1</sup> vs 0.12 min<sup>-1</sup> Figure S4). This rate increase emphasizes the positive impact of the combination of a high catalyst density within the crystals and a better catalyst stability on the catalytic efficiency.

**Table 1.** Substrate **1a-1i** oxidation by *CLECNikA-FeL3*.

	Product 2a-2i	<i>CLECNikA-FeL3</i>	Yield 2a-2i (%)	TON
2a		-	3	-
		+	15	38
		+*	84	214
2b		-	0	-
		+	28	71
		+*	80	202
2c		-	0	-
		+	1	2
		+**	50	570
2d		-	3	-
		+	17	43
2e		-	2	-
		+	30	77
2f		-	4	-
		+	89	227
2g		-	5	-
		+	15	40
2h		-	0	-
		+	0	-
2i		-	3	-
		+	15	38

Experimental conditions: 1:255:600; [cat] = 15  $\mu$ M in in  $\text{CH}_3\text{CN}:\text{H}_2\text{O}$  1/1 v:v for 2 hours. \*With 31  $\mu$ M CLEC. \*\* With 1:1150:1200.

**Conclusion.** This study shows that the transition from homogeneous to heterogeneous conditions does not affect the selectivity of the *soNikA-FeLn* artificial enzymes, confirming that the switch of conditions consists of a real transposition of the catalysis. It also confirms that the reaction mechanism of the *soNikA-FeLn* is conserved. The use of *CLECNikA-FeLn* has increased the reactivity by nearly  $1/3^{\text{rd}}$  in one standard run of catalysis, 3.5-fold when lowering the catalyst concentration to 0.09% and 3-fold when reloading the catalyst. The moderate stability of the catalysts closely relates to the kinetic of the reaction, as the faster the oxidant reacts with the substrate, the less it degrades the catalyst, related to the nucleophilicity of the sulfide substrate. No positive results were conclusive for enantioselectivity implying that NikA engineering approaches have to be considered in the future. For industrial approaches, the CLEC technology should be developed for Modafinil® or Esomeprazole® transformation. Finally, this work illustrates the mode of action of artificial CLEC on substrate promiscuity and kinetics enhancement.

## Acknowledgements

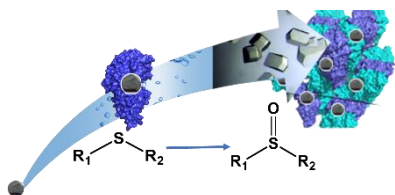
We thank the Agence Nationale pour la Recherche for grant (ANR-14-CE06-0005-01), and the CEA, CNRS and Univ. Grenoble-Alpes for their institutional support. This work has been partially supported by Labex ARCANE (ANR-11-LABEX-0003-1) and CBH-EUR-GS (ANR-17-EURE-0003). We thank the staffs from the BM-30A beamline of the European Synchrotron Facility in Grenoble

**Keywords:** Heterogeneous Catalysis • Oxidation • Artificial Metalloenzymes • Sulfoxidation k • Bioinspired catalyst • Crystal Catalysis

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## Entry for the Table of Contents



**Crystals are not fragile.** Using protein crystals as supports for inorganic catalyst, we have designed a methodology of heterogeneous catalysis by artificial enzymes. The hybrid solids, made from the association of the NikA protein and an iron complex with a N2Py2 ligand, provide access to a larger substrate scope, a greater stability and greater kinetics for sulfide transformations than the homogeneous counterpart.