



Strong habitat and weak genetic effects shape the lifetime reproductive success in a wild clownfish population

Océane Salles, Glenn R. Almany, Michael L. Berumen, Geoffrey P. Jones, Pablo Saenz-Agudelo, Maya Srinivasan, Simon R Thorrold, Benoit Pujol, Serge Planes

► To cite this version:

Océane Salles, Glenn R. Almany, Michael L. Berumen, Geoffrey P. Jones, Pablo Saenz-Agudelo, et al.. Strong habitat and weak genetic effects shape the lifetime reproductive success in a wild clownfish population. *Ecology Letters*, 2020, 23 (2), pp.265-273. 10.24072/pci.evolbiol.100082 . hal-03020148

HAL Id: hal-03020148

<https://hal.science/hal-03020148>

Submitted on 23 Nov 2020

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

ECOLOGY LETTERS

Strong habitat and weak genetic effects shape the lifetime reproductive success in a wild clownfish population

Journal:	<i>Ecology Letters</i>
Manuscript ID	ELE-01130-2019.R1
Manuscript Type:	Letters
Date Submitted by the Author:	n/a
Complete List of Authors:	<p>Salles, Océane; PSL Université Paris: EPHE-UPVD-CNRS, USR 3278 CRILOBE; EPHE, Laboratoire d'Excellence 'CORAIL'</p> <p>Almany, Glenn; PSL Université Paris: EPHE-UPVD-CNRS, USR 3278 CRILOBE; EPHE</p> <p>Berumen, Michael; King Abdullah University of Science and Technology, Red Sea Research Center</p> <p>Jones, Geoffrey; James Cook University, ARC Centre of Excellence for Coral Reef Studies</p> <p>Saenz-Agudelo, Pablo; Universidad Austral de Chile, Instituto de Ciencias Ambientales y Evolutivas</p> <p>Srinivasan, Maya; James Cook University, ARC Centre of Excellence for Coral Reef Studies</p> <p>Thorrold, Simon; Woods Hole Oceanographic Institution, Biology</p> <p>Pujol, Benoit; PSL Université Paris: EPHE-UPVD-CNRS, USR 3278 CRILOBE; EPHE, Laboratoire d'Excellence 'CORAIL'</p> <p>PLANES, Serge; PSL Université Paris: EPHE-UPVD-CNRS, USR 3278 CRILOBE; EPHE, Laboratoire d'Excellence 'CORAIL'</p>

1
2
3
4
5
6
7
8
9
10
11
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

1 Strong habitat and weak genetic effects shape the lifetime
2 reproductive success in a wild clownfish population

3
4 1 Océane C. Salles^{1,2}, Glenn R. Almany^{1,2†}, Michael L. Berumen³, Geoffrey P. Jones⁴, Pablo
5 2 Saenz-Agudelo⁵, Maya Srinivasan⁴, Simon R. Thorrold⁶, Benoit Pujol^{1,2*}, and Serge Planes^{1,2*}
6 3

7 4 ¹ PSL Université Paris: EPHE-UPVD-CNRS, USR 3278 CRIOBE, Université de Perpignan, 52
8 5 Avenue Paul Alduy, 66860 Perpignan Cedex, France.

9 6 ² Laboratoire d'Excellence 'CORAIL', 58 avenue Paul Alduy, F-66360 Perpignan, France.

10 7 ³ Red Sea Research Center, Division of Biological and Environmental Sciences and Engineering,
11 8 King Abdullah University of Science and Technology, Thuwal, 23955, Saudi Arabia.

12 9 ⁴ ARC Centre of Excellence for Coral Reef Studies, and College of Science and Engineering, James
13 10 Cook University, Townsville, Queensland 4811, Australia.

14 11 ⁵ Instituto de Ciencias Ambientales y Evolutivas, Universidad Austral de Chile, 5090000 Valdivia,
15 12 Chile.

16 13 ⁶ Biology Department, Woods Hole Oceanographic Institution, Woods Hole, MA 02543, United
17 14 States of America.

18 15 †Deceased 24 March 2015.

19 16 *Both authors share senior authorship of this article
20 17

Authors' contributions: GPJ, SLRT, and SP designed the research program; OCS, BP, GRA, and MLB contributed new reagents/analytic tools; OCS and BP analyzed data; OCS, BP and SP wrote the manuscript and all authors contributed substantially to revise the paper.

Data accessibility statement: R programming protocols, script and codes, metadata and Rdata files to call upon any given parameter estimates from this study can be obtained on the Zenodo repository: <https://doi.org/10.5281/zenodo.3476529>

Short running title: Habitat drives clownfish local success

Keywords: multi-generational pedigree, additive genetic variation, maternal effects, environmental effects, adaptation, selection, heritability, evolvability

Type of article: Letters

Number of words in the abstract: 150, **in the main text (excluding abstract, acknowledgements, references, table and figure legends):** 5012, **in each text box:** N/A

Number of references: 54. **Number of figures:** 2. **Number of Tables:** 1. **Number of boxes:** 0

Person to whom correspondence should be sent: Benoit Pujol, USR3278 CRIOBE, Université de Perpignan, 52 Av. Paul Alduy, 66860 Perpignan cedex, France. Tel: +33 (0)4 30 19 23 24. Fax: +33 (0)4 68 50 36 86. Email: benoit.pujol@univ-perp.fr

1
2
3
4
5
6
7
8
9
10
11
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

Summary: The relative contributions of environmental, maternal and additive genetic factors to the Lifetime reproductive success (LRS) determine whether species can adapt to rapid environmental change. Yet to date, studies quantifying LRS across multiple generations in marine species in the wild are non-existent. Here we used 10-year pedigrees resolved for a wild orange clownfish population from Kimbe Island (PNG) and a quantitative genetic linear mixed model approach to quantify the additive genetic, maternal and environmental contributions to variation in LRS for the self-recruiting portion of the population. We found that the habitat of the breeder, including the anemone species and geographic location, made the greatest contribution to LRS. There were low to negligible contributions of genetic and maternal factors equating with low heritability and evolvability. Our findings imply that our population will be susceptible to short-term, small-scale changes in habitat structure and may have limited capacity to adapt to these changes.

Introduction

Lifetime reproductive success (LRS) – the number of successful offspring an individual contributes to the next generation – is a critical variable underpinning ecological and evolutionary responses to the environment (Clutton-Brock 1988). Several factors including different environmental, parental, and additive genetic effects can influence LRS (Hendry *et al.* 2018). If LRS is exclusively a phenotypic response to the conditions experienced by individuals, populations will be severely impacted by rapid environmental change and there is no prospect of adaptive microevolution. Maternally transmitted responses to environmental conditions can mediate the effect of those changes on the next generation (Mousseau & Fox 1998). However, it is the additive genetic variation in LRS that determines the rate of adaptation of a population to the environmental demand (Fisher 1930). To date, few studies have distinguished these relative contributions to LRS over multiple generations in wild populations (Hendry *et al.* 2018). This situation is changing as long-term, individual-based ecological studies with multi-generational pedigrees provide the necessary longitudinal information to quantify the different components of LRS (Pemberton 2008). Such studies are imperative as we seek to understand the ability of species to withstand or adapt to accelerating climate change (Munday *et al.* 2017).

Intergenerational responses to selection are a product of the interplay between evolutionary and ecological mechanisms that ultimately shape inherited variation in fitness-related traits. Environmentally driven mechanisms (*e.g.*, phenotypic plasticity, genetic assimilation) can facilitate (Ghalambor *et al.* 2007; Danchin *et al.* 2019) or constrain the microevolutionary response to selection (Pujol *et al.* 2018). However, in the absence of genetic variation for LRS, these mechanisms will likely have little effect on a negligible rate of adaptive evolution. In quantifying additive genetic variation, it is important to distinguish between *heritability* and *evolvability* (Wheelwright *et al.* 2014). Narrow sense heritability is widely used as a measurement of the population potential to respond to selection (Mousseau & Roff 1987). It is the additive genetic variance standardized by the

1
2
3 83 total phenotypic variance. Low heritability values can either reflect low additive genetic variance or
4
5 84 large environmental, or residual effects. Evolvability is the mean-standardized additive genetic
6
7
8 85 variance (Houle 1992). Environmental or maternal effects do not affect it, which makes it a more
9
10 86 appropriate metric in the comparison of evolutionary potential between traits, populations and
11
12 87 species. When measured for a surrogate of fitness like LRS, it estimates the expected proportional
13
14 88 change per generation in population mean fitness given a unit strength of selection. Heritability for
15
16 89 LRS reveals whether its additive genetic variance represents a non-trivial proportion of its total
17
18 90 variance in the actual environmental context of a given wild population. Together, evolvability and
19
20 91 heritability inform us about how much environmental change a wild population can withstand on the
21
22 92 basis of its evolutionary potential.
23
24
25
26 93
27
28 94 The few (n=15) long-term, individual-based studies that have quantified additive genetic variation,
29
30 95 heritability and evolvability of LRS in wild populations have all focused on terrestrial species (Postma
31
32 96 2014; Hendry *et al.* 2018; Table S1). They have largely confirmed low additive genetic variation and
33
34 97 evolvability (~0.08) for LRS, which nevertheless reflects some evolutionary potential (Hendry *et al.*
35
36 98 2018). Whether marine fish and terrestrial species reveal the same patterns remains unknown to date.
37
38 99 Until recently, quantifying LRS in marine organisms with a pelagic larval stage has been considered
39
40 100 impossible because of the difficulties in following the fate of offspring from one generation to the
41
42 101 next. However, there is increasing evidence of some degree of natal philopatry or self-recruitment in
43
44 102 local marine populations (Jones *et al.* 2009). Genetic parentage analysis is making it possible to assign
45
46 103 a significant proportion of offspring to their parents (Planes *et al.* 2009) and construct
47
48 104 multigenerational pedigrees for the offspring that return to their natal population (Salles *et al.* 2016).
49
50 105 As in most quantitative genetic studies of LRS, it is impossible to measure the recruitment of marine
51
52 106 fish dispersing juveniles at other locations. The regional component of LRS, which would inform us
53
54 107 on fitness variation beyond the local scale is impossible to obtain. Measuring the local component of
55
56
57
58
59
60

LRS in marine fish is nevertheless an opportunity as in any other species because it estimates the relative contribution of local fish to the population self-recruitment and replenishment.

For coral reef fishes, quantifying environmental and genetic components of LRS and assessing its evolvability in wild populations is of great contemporary importance. Between 30 to 50% of the world's coral reefs have been lost and those remaining are considered highly vulnerable (van Hooidonk *et al.* 2016). The rapid loss of suitable habitat is widely acknowledged to be contributing to a decline in reef fish populations and biodiversity (Jones *et al.* 2004). The potential for adaptation of reef fish is uncertain and near future environmental conditions predicted under climate change should have a dramatic effect on their reproductive success (Munday *et al.* 2013). To date, environmental, maternal and additive genetic contributions to LRS in wild coral reef fish populations have not been assessed. However, recent work establishing high levels of natal philopatry in some coral reef fishes (Jones *et al.* 2005) that allowed for resolving their pedigree across multiple generations (Salles *et al.* 2016) opened the way for quantifying the local component of LRS for the first time in coral reef fishes.

Here, we focus on the entire local population of the orange clownfish *Amphiprion percula* at Kimbe Island, Papua New Guinea where each year ~half the juveniles successfully recruiting are progeny of local breeding pairs (Salles *et al.* 2016). We use multi-generational pedigrees of up to 5 generations obtained from biennial DNA sampling over 10 years (Salles *et al.* 2016) and apply a quantitative genetic linear mixed model approach (Kruuk & Hill 2008) to quantify the additive genetic, maternal and environmental components of variation in LRS for the self-recruiting portion of the population. Habitat effects were quantified by examining this local LRS for individuals resident in two different anemone species and from different geographic locations around the island. By integrating habitat data with the pedigree information in a quantitative genetic generalized linear mixed model, we were able to assess the relative contribution of additive genetic, maternal and habitat effects to local LRS.

We also calculated the evolvability and heritability of local LRS to evaluate its evolutionary potential to respond to selection at the scale of the Kimbe island population.

Material and methods

Study population and data collection

A natural population of orange clownfish (*A. percula*) living in the reef surrounding Kimbe Island (Fig. 1a; 5°12'22.56" S, 150°22'35.58" E), West New Britain Province of Papua New Guinea, was surveyed every second year from 2003 to 2013. Here, *A. percula* lives in a mutualistic association with one of two host sea anemone species, *Heteractis magnifica* (Fig. 1b) and *Stichodactyla gigantea* (Fig. 1c). We geographically located and tagged a total of 310 anemones (176 *H. magnifica* and 134 *S. gigantea*) that were occupied by *A. percula* on the entire reef surrounding the island.

These two anemone species are remarkably different in terms of the micro-habitat they provide, including a wide range of shapes, sizes, depth distributions and surrounding substrata (Dunn 1981). Although we did not directly measure these variables (other than depth), the combination of host anemone species (*H. magnifica* or *S. gigantea*) with one of the three geographical areas covering the entire reef around the island (northern, western or eastern areas, Figure 1a) where it is located describes a 'habitat' variable for each fish that encompasses a suite of biotic and abiotic environmental conditions. These geographical areas correspond to the different coasts of the island that reflect possible environmental effects of the geographic location. In total, the combination of the two host anemone species with the three geographical areas allowed us to describe six different habitats.

Clownfish within one anemone live in group of typically three to five individuals in the Kimbe Island population. The size-based dominance hierarchy in *Amphiprion* allows us to determine the reproductive status of each individual (Fricke 1979). The female is the largest, the male is the second

largest, and the non-breeders rank progressively lower in the hierarchy as they decrease in size. If the single female adult of a group dies, then the male changes sex to female, and the largest non-breeder from the anemone becomes sexually mature as male. Reproduction occurs year round, with females laying several hundred eggs in a clutch near the pedal disk of the host anemone each lunar month. The eggs hatch after ~7 days of paternal care into larvae that spend ~10 days in the pelagic environment (Berumen *et al.* 2010) before settling to an anemone, either at their natal location (Kimbe Island) or elsewhere (Planes *et al.* 2009).

Divers captured fish by using hand nets. Individuals were measured *in situ* using calipers, fin-clipped (size > 35mm) or collected whole (size < 35mm) for genetic analysis and then released back on the same anemone. Small pieces of fin tissue were preserved in 95% ethanol in 2-mL vials. The biggest fish in each anemone was identified as the female, the second largest individual was assumed to be the male, and all other individuals were classified as non-breeders. We extracted DNA from all samples and genotyped them at 22 polymorphic microsatellite loci (Bonin *et al.* 2015). Then, we identified the individuals sampled multiple times over the years by using the Excel macro GenAlex v6.5 (Peakall & Smouse 2012) to compare multilocus genotypes from 2003, 2005, 2007, 2009, 2011 and 2013. Individuals were in average sampled 2.88 ± 0.04 times (mean \pm SE) over the six surveys (1% of individuals persisted over the 10-yr period, Salles *et al.* 2016). The 2-yr sampling scheme precluded calculating a precise measurement of the age of individuals, in particular for fish sampled in 2003 during the first sampling period, which age was unknown. The total duration of this long term survey did not allow us to obtain many replicated measurements within individuals before and after sex change (n=41 individuals). Estimating sex dependent additive genetic variance is precluded in this case because some effects cannot be disentangled as the clownfish only changes sex in one direction (from male to female). This change is always associated with a change of sexual partner and with an increase in female body size, which we expect to generate a confounding effect between a

1
2
3 185 female condition and its genetic quality. We therefore did not consider sex in our model as a result of
4
5 186 data and analytical limitations.
6
7
8 187
9
10 188 *Pedigree used for quantitative genetic analysis*
11
12 189 Pedigree reconstruction was conducted by assigning juvenile fish to parental pairs on the basis of
13
14 190 their multilocus genotypes (Salles *et al.* 2016). We used the software FaMoz (Gerber *et al.* 2003).
15
16
17 191 This approach is based on the calculation of LOD scores (Log of the odd-ratio comparison) for any
18
19 192 potential parentage relationship. It determined critical thresholds to accept or reject assignments by
20
21 193 simulating true and false parent-offspring pairs. Further details on parentage analyses and pedigree
22
23 194 reconstruction are given in Salles *et al.* (2016a). We kept assignments to known parental pairs, but
24
25 195 rejected assignments to single adults. In the context of overlapping generations, we used the year of
26
27 196 first sampling and the anemone of each parental couple as information to avoid possible false
28
29 197 assignments. As a result, sibship links could not be confused with parental links. The same individual
30
31 198 can be related to its offspring with either a paternal or maternal link because of sex changes. We
32
33 199 identified the mother and the father based on the size of the two parents on the year of first capture.
34
35 200 The original population pedigree includes 2927 clownfish over five generations including 121
36
37 201 families, 987 paternal, 987 maternal, 1809 full-sib, 412 maternal half-sibs, 248 paternal half-sib, 135
38
39 202 maternal grandmothers, 135 maternal grandfathers, 278 paternal grandmothers, 278 paternal
40
41 203 grandfathers and 218 cousins (Salles *et al.* 2016). For this study, we excluded from the original
42
43 204 pedigree the 1192 individuals that were removed from the habitat at the juvenile stage (size < 35mm,
44
45 205 10 to 458 days old). The final pedigree used for this study includes 1735 individuals from five
46
47 206 generations (Fig. S1). We used the R package ‘pedantics’ (Morrissey & Wilson 2010) to assess the
48
49 207 power of the resolved pedigree to detect significant quantitative genetic parameters (Fig. S1).
50
51
52 208
53
54 209 *LRS: the individual contribution to self-recruitment*
55
56
57
58
59
60

LRS, which when measured at the scale of the local population is also the contribution of an individual to self-recruitment, corresponds to the total number of offspring produced during its lifetime and recruiting into Kimbe Island (*e.g.*, the local breeder population). We used biennial measurements of their reproductive success (using field-data from 2003, 2005, 2007, 2009, 2011 and 2013) to compare LRS between individuals because some fish were still alive at the end of sampling and others might have already reproduced before the first sampling year. The LRS corresponds here to the total number of descendants produced on a biennial basis that successfully recruited into Kimbe Island population, which provided us with repeated measures over the period of the survey from 2003 to 2013. In the Supplementary information we present results from an alternative approach based on the De-lifing method (DL). The calculation of DL takes into account the temporal variation of the population growth and estimates the contribution of every clownfish to biennial changes in population size through both reproduction and survival (Coulson *et al.* 2016). Statistical problems with the use of DL, potentially leading to precision issues, have been pointed out (Dupont *et al.* 2017). DL was used in two of the 15 studies where the genetic variation of fitness was quantified in wild populations (Table S1), which limits our ability to discuss its properties. We therefore also provided DL results in this study in the supplementary section.

Quantitative genetic generalized linear mixed model approach

Similarities between relatives living in contrasted micro-habitats allowed us to evaluate simultaneously the genetic and habitat components of LRS. Repeated ‘records’ on individuals made it possible to estimate permanent environmental effects, and therefore account for intra-individual and unmeasured environmental trait variation across time. Permanent environmental effects also account for a part of non-additive genetic effects (Wilson *et al.* 2010). LRS variance was partitioned into six random effects: Additive genetic (V_A), Maternal (V_M), Natal Habitat (V_{NH}), Resident Habitat (V_{RH}), Permanent Environment (V_{PE}) and Residual (V_R) variances by using the ‘animal model’ quantitative genetic approach (Kruuk 2004). This Linear Mixed Model (LMM) approach uses

pedigree information to extract the additive genetic component. This approach is more powerful than traditional analyses (*e.g.*, parent–offspring regressions) because it takes into account every relationship link in a pedigree. Maternal variance was modeled using the mother’s identity as a random effect, allowing maternal effects to include both genetic and environmental maternal effects. Permanent environmental effects were modeled by including the identity of individuals as a random effect. The LRS variance is the sum of six variance components:

$$V_{LRS} = V_A + V_M + V_{NH} + V_{RH} + V_{PE} + V_R \quad (1)$$

Quantitative genetic models were computed as univariate GLMMs using the ‘MCMCglmm’ package (Hadfield 2010) in R version 3.5.1 (R.Core.Team 2018), with LRS as a Poisson response variable. Using this Bayesian framework facilitates parameter estimation for non-Gaussian traits. We used parameter expanded priors for all analyses ($V=1$, $nu=0.02$), which are often referred to as ‘non informative’ priors although such denomination can be debated, as we wanted posterior estimates to be determined from the data and not from the priors (Morrissey *et al.* 2014). We ran model MCMC chains over 1,000,000 iterations with initial burning of 10,000 iterations and a thinning of 1,000 iterations. Historically, the Deviance Information Criterion (DIC) was often used to compare models and assess the significance of the random variance components in this type of approach. However, it is becoming less commonly used since it was recognized as an inappropriate tool for model comparisons of the same type than quantitative genetic GLMM analyses (Spiegelhalter *et al.* 2014). Effects of variance components were considered statistically supported if their posterior distributions did not overlap zero (Wilson *et al.* 2010).

Variance Components

Variance components were estimated as the mode of the posterior distributions established on the MCMC sample and we reported lower and upper limits of the 95% credible intervals. For the six variance components, we calculated their relative contribution to the sum of all variance components, thereby expressing their effects as percentages of the total phenotypic variance (V_{LRS}). As a result,

we obtained standard narrow sense heritability estimates for LRS (h^2) by applying the basic formula ($h^2 = V_A / V_P$), and similarly maternal effects by estimating the proportion of total phenotypic variance explained by the maternal variance ($m^2 = V_M / V_P$). Evolvability (I_A) of LRS is the additive genetic variance divided by the squared mean of the LRS (Houle 1992). Analyses assumed a Poisson distribution and provided parameter estimates on a statistically convenient latent scale for non-Gaussian traits. We therefore back-transformed these estimates onto the observed data scale to improve our inferences by using the ‘QGImm’ package (de Villemereuil *et al.* 2016). We used the function ‘QGparams’ to estimate additive components such as V_A and h^2 , and ‘QGicc’ to estimate broader sense components such as V_M and m^2 , V_{NH} , V_{RH} , V_{PE} and V_R . Although parameter estimates transformed back on the data-scale are expected to be upward biased, their ratio is reliable, and hence the estimators derived from their relative proportions such as h^2 . It is necessary to point out two specific aspects of this back transformation on the observed data scale. First, V_R is estimated on the basis of the additive over-dispersion term in the nonlinear model and its value cannot be interpreted similarly to the usual residual variance term estimated by classical quantitative genetic linear mixed models. Second, the sum of the variance components estimated on the data scale are not additive and therefore not expected to sum up to the value of the phenotypic variance calculated directly on the raw data. For the sake of clarity and comparison, we present the results on the latent scale and the observed data scale. We calculated the 95% credibility intervals from the posterior distributions of observed parameters for all the variance components and other estimates expressed on their basis by using the ‘HDIInterval’ package (Meredith & Kruschke 2016).

Results

Habitats dominantly shape the Lifetime Reproductive Success in the clownfish population

Biennial estimates of the Lifetime Reproductive Success (LRS) measured inside the area of the Kimbe island population ranged from 0 to 13, with a phenotypic variance $V_{LRS}=1.31$ and an average value of 0.54 ± 0.05 (mean \pm SE) offspring per individual for a two-year period. Because clownfish live

1
2
3 288 in strong association with their anemone, we were able to identify and geo-locate the precise position
4
5 289 and habitat where breeders contributed more to the local replenishment of the population (Fig. 1a).
6
7 290 Breeders that produced more self-recruiting offspring lived in Kimbe Island's eastern area and mostly
8
9 291 in *S. gigantea* anemones. Our analysis also revealed fish that did not contribute at all to the local
10
11 292 replenishment of the population over the 10-year monitoring period. These fish represented 25% of
12
13 293 the pairs of local breeders and were located in 48 *H. magnifica* and 30 *S. gigantea* of the 310 anemones
14
15 294 monitored in both deep and shallow waters (Fig. 1a).
16
17
18 295
19
20
21 296 Quantitative genetic generalized linear mixed models on the latent and the observed data scale gave
22
23 297 very close results. Our results on the scale of observed data showed that Natal Habitat and Resident
24
25 298 Habitat explained respectively 19.1% and 5.7% of the variance in local LRS, furthermore, residual
26
27 299 and permanent environment explained respectively 41.0% and 31.0%, whereas additive genetic
28
29 300 effects and maternal effects were very weak and explained 1.3% and 1.9% respectively (Fig. 2a,
30
31 301 variances on observed data-scale). Similar results were obtained for DL (see Supplementary
32
33 302 Information for more details).
34
35 303
36
37
38
39 304 *Low evolvability and low heritability for LRS*
40
41
42 305 The modes of the posterior distributions estimating the additive genetic variance for LRS, expressed
43
44 306 on the latent and observed data-scales, were extremely small (Table 1). On the observed data-scale,
45
46 307 we found $V_A=0.030$ ($CI_{95\%}$ 4.94×10^{-4} to 0.060). This could be linked to the statistical power of our
47
48 308 pedigree (Fig. S1). Our model nevertheless placed fairly restricted bands on the 95% credible
49
50 309 intervals (Table 1). Credible intervals did not overlap the zero but were close. The extent to which
51
52 310 these very low values of genetic estimates are not null must therefore be considered with caution.
53
54 311 LRS evolvability estimated on the observed data-scale, which evaluated the micro-evolutionary
55
56 312 change of the number of self-recruiting offspring that can be reached by the population, was equal to
57
58 313 0.103 ($CI_{95\%}$ 1.661×10^{-3} to 0.511). In other words, 0.103 additional juveniles were added to the
59
60

average number of juveniles originating and recruiting in the population per generation. The heritability estimate expressed on the observed data-scale was $h^2=0.013$ (CI_{95%} 4.951×10^{-5} to 1.227×10^{-2}) for LRS (Table 1). We can therefore estimate the maximum response (R) to selection (S), expected in theory, in the presence of strong selection pressures acting on the Kimbe Island orange clownfish population by using the Breeder's equation $R=h^2\times S$ (Lush 2008). The low to negligible value of the LRS heritability means that the maximum genetic change of the population average LRS would never exceed ~ 0.020 offspring per generation. Similar results were obtained for DL (see Supplementary Information for more details).

Weak maternal effects for LRS

Our analysis detected maternal variance for the LRS but it was also extremely small to the extent that it might be considered as null (Table 1): $V_M=0.019$ (CI_{95%} 2.966×10^{-5} to 2.044×10^{-2}). It made very little contribution to the total variance in LRS ($m^2=1.9\%$, expressed on observed data-scale, Fig. 2). The habitat occupied by the mother (Natal Habitat) had a stronger effect on LRS than the mother herself. The relative contribution of individuals to the population replenishment was indeed influenced by the Natal Habitat to an extent of 19.1% for LRS. Similar results were obtained for DL (see Supplementary Information for more details).

Discussion

Strong habitat and weak genetic effects on LRS

Our study revealed that biennial LRS in the Kimbe Island orange clownfish population quantified over five generations was largely explained by ecological factors. Host anemone species, depth and geographical location explained $\sim 25\%$, and permanent environmental effects $\sim 30\%$ of its variation, with only weak maternal (1.9%) and additive genetic effects (1.3%). Intrinsic biological characteristics of anemone species (*e.g.*, size, shape and toxicity) affecting the life-history traits of their resident clownfish might explain strong habitat effects (Chausson *et al.* 2018). In addition, the

higher toxicity of *S. gigantea* (Nedosyko *et al.* 2014) might provide better protection against predators of eggs attached to the host anemone, but this hypothesis remains to be tested. The geographical location also appears to be important. Most successful individuals were close to the land in shallow water on *S. gigantea* and in deeper lagoons for *H. magnifica*, which might promote greater local retention of larvae. Mechanisms responsible for geographical differences in LRS around Kimbe Island remain unknown (Berumen *et al.* 2010). Some breeders likely have a different reproductive success beyond the sampled population, through dispersers rather than self-recruiters. Inside the Kimbe island clownfish population, weak genetic effects on LRS indicate a low to negligible rate of adaptive evolution in progress and raise concern about the ability of this population to adapt to rapid climate change.

350

351 *Susceptibility to habitat change*

Habitat largely drove LRS variation. Individuals that happen to settle on particular anemones and places do well. The dependence on habitat quality of LRS inside the Kimbe island population suggests this species will be extremely susceptible to habitat degradation over ecological time scales. Detrimental direct and indirect anthropogenic impacts on reef anemones habitats are already affecting numerous clownfish species (Saenz-Agudelo *et al.* 2011). *S. gigantea* anemones located in shallow waters are likely to be disproportionately more impacted by increasing water temperatures and irradiance (Hobbs *et al.* 2013). If these habitats are differentially impacted, this will affect clownfish contributions to the local replenishment of the population and compromise population persistence.

360

361 *Low to negligible evolutionary potential*

Our findings provide the first empirical support for a wild marine population to Fisher's fundamental theorem of selection that additive genetic variance in fitness is depleted under selection and tends towards zero in a population reaching evolutionary equilibrium (Gustafsson 1986). Although normal and expected, low heritability and evolvability in LRS is concerning given the increasing rate of

environmental change. The low to negligible scope for adaptive evolution (estimated by evolvability) and the low to negligible genetic potential for responding to selection (estimated by heritability) may not be a problem for gradual environmental change. At this rate, it would take around at least 10 generations for the population average LRS to increase by one juvenile, which highlights the stability of the demographic rate of self-recruitment in this population. Our results therefore support the hypothesis that the population is at evolutionary equilibrium (no genetic changes) in a context of environmental stability over the timescale of the survey.

Connectivity as a plausible cause

Our findings were at first surprising because immigration accounts for on average 44% of the juvenile recruitment (Salles *et al.* 2016). Average dispersal distance in Kimbe Bay is between 10 and 20km, providing substantial connectivity among adjacent reefs and potential dispersal of up to 100km (Pinsky *et al.* 2017). The associated gene flow would be expected to bring new genetic variants and thereby increase genetic variation (Facon *et al.* 2008). Under such scenario, selection for self-recruitment, and thereby against migrants, has to be strong to keep the population at evolutionary equilibrium. An alternative scenario is that homogenization by gene flow results in most immigrants sharing a similar genetic background. As a result, no new genetic variants are brought in the population by gene flow and low genetic variation is maintained (Pujol *et al.* 2010). Low genetic variation for LRS implies that evolution by selection at the local scale is extremely limited in its current state. However, this does not imply a dead end for adaptive evolution in this clownfish population because several mechanisms can provide adaptive evolutionary potential over the long term (Pujol *et al.* 2018).

Slight but probably negligible maternal genetic effect

Additional adaptive evolutionary potential can in theory be provided to a population by maternal effects (Räsänen & Kruuk 2007). In the Kimbe Island orange clownfish, we found that maternal

effects explained 1.9% of the LRS variance, which is quite small, even if it was more than additive genetic effects. Maternal effects therefore increased slightly the low to negligible rate of LRS change by adaptive evolution. One must note that this increase is nearly negligible. It is likely that this small value reflects the genetic component of the maternal effect. The habitat of birth, which is also the maternal habitat, probably encompassed a non-negligible part of the maternal environmental effects (Germain & Gilbert 2014). There is growing awareness that maternal environmental effects can contribute to adaptation in natural populations, especially when maternal and offspring environments are positively correlated (Shama 2015). It might even buy some time for adaptive evolution through slow genetic change to occur (Levis & Pfenning 2016).

Towards a wider sample of contemporary rates of adaptive evolution in the wild

In our study, LRS estimated local fish contribution to the local population replenishment. As with most wild population pedigrees, this excludes the dispersal fitness because the amount of offspring that successfully dispersed somewhere else is unknown (Kruuk *et al.* 2000). Its genetic variation evaluates the rate of adaptive evolution inside the Kimbe island population. While there are no comparable data from marine systems, 15 studies conducted on terrestrial vertebrates have also estimated the additive genetic variation and the heritability of LRS (Table S1). The number of estimates of maternal effects on LRS variation are extremely rare (McFarlane *et al.* 2014 and references therein). A majority of these studies similarly found low to negligible contributions of additive genetic effects in the wild. Marine fish, mammals and birds revealed the same patterns. There is little evidence of additive genetic variance for LRS in studies conducted in the wild. This situation is less clear for maternal effects, partly because studies remain scarce. Very low but significant additive and maternal genetic variations for fitness indicate that there was some genetic change over the course of the survey but very limited potential for short-term adaptive evolution.

A wider use of LRS does not preclude caution

LRS studies in wild populations with a known pedigree have multiplied during the last thirty years. Although useful insights were gained, one must note that the use of LRS as a surrogate for fitness is not well suited for iteroparous species (Grafen 1988). This is particularly true when population growth varies between reproductive seasons (Coulson *et al.*, 2006). Furthermore, although quantitative genetic approaches decomposing the genetic and environmental components of LRS are theoretically correct, they might provide imprecise parameter estimates in the wild because temporal and spatial environmental effects (e.g., condition dependent LRS, cohort effects) might not have been adequately corrected for. Caution must therefore be taken when interpreting the additive genetic variance of LRS as adaptive evolutionary rate or evolutionary potential in wild populations where it might in fact reflect other ecological parameters. Our quantitative genetic model explained a large amount of environmental variation in LRS in the Kimbe island clownfish population, which supports the reliability of our genetic estimates.

Conclusion

The major outcome of this study is that the heterogeneity of the habitat of the Kimbe Island orange clownfish had a profound influence on the individual contribution to the local population replenishment over five generations. This finding implies that habitat ecology is crucial for this clownfish population. In terms of future persistence, expected changes in habitat quality and configuration over relatively short time scales might affect the ability of fish to self-recruit. This ability harbored low to negligible additive genetic and maternal genetic variation. As a consequence, this population potential for rapid evolutionary change of LRS by selection, and therefore its rate of adaptive evolution, can be considered negligible in the current state of the population. This finding, which is in line with other studies on the topic, stresses the importance of environmental mechanisms (e.g., plasticity) that have the potential to enable rapid adaptive responses. From the perspective of management, our results caution against hoping for local adaptive evolutionary responses and lend support to focusing conservation efforts on maintaining habitat quality.

1
2
3
4
5
6
7
8
9
10
11
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

Acknowledgements

This research was supported by Laboratoire d’Excellence CORAIL, Expenditure Review Committee, Coral Reef Initiatives for the Pacific, the Global Environment Facility Coral Reef Targeted Research Connectivity Working Group, National Science Foundation, the Australian Research Council Centre of Excellence Coral Reef Studies, The Nature Conservancy, Total Foundation, James Cook University, King Abdullah University of Science and Technology, and Woods Hole Oceanographic Institution. Research visas were approved by the Papua New Guinea (PNG) government and research protocols were endorsed by the Board of Mahonia Na Dari Research and Conservation Centre, Kimbe, PNG.

We thank the large number of volunteers who assisted in the field and collected tissue samples. Mahonia and FeBrina provided essential logistic support. We are grateful to the traditional owners of the reefs near Kimbe Island for allowing us access to their reefs. We also thank Pierre de Villemereuil, Jarrod Hadfield, Michael Morrissey, Caroline Thomson and Isabel Winney for useful discussions, comments, and help with the method.

We thank Ecology Letters’ editor Tim Coulson for precious comments that improved this manuscript. We also thank PCI Evol. Biol for the recommendation of a previous version of this article (<https://doi.org/10.24072/pci.evolbiol.100082>), and in particular Philip Munday for recommending our study and two non anonymous reviewers: Loeske Kruuk and Juan Diego Gaitan-Espitia for precious comments

Conflict of interest: Authors declare no conflict of interests

Supplementary material

Additional Supporting Information may be found in the online version 3 of this preprint on the Zenodo repository: <https://doi.org/10.5281/zenodo.3476529>.

The Additional Supporting Information file contains:

Table S1. Previous estimates of fitness heritability and maternal effects on fitness in 15 wild populations.

Figure S1. Pedigree data and power analysis for the Kimbe Island orange clownfish population.

Supplementary methods, results and discussion on De-living measures.

References

- Berumen, M.L, Walsh, H.L., Raventos, N., Jones, G.P., Starczak, V. *et al.* (2010). Otolith geochemistry does not reflect dispersal history of clownfish larvae. *Coral Reefs*, 29, 883-891.
- Bonin, M.C., Saenz-Agudelo, P., Harrison, H.B., Nanninga, G.B., van der Meer, M.H. *et al.* (2015). Characterisation and cross-amplification of microsatellite markers in four species of anemonefish (*Pomacentridae*, *Amphiprion* spp.). *Mar. Biodivers.*, 46, 135-150.
- Chausson J, Srinivasan M & Jones GP (2018). Host anemone size as a determinant of social group size and structure in the orange clownfish (*Amphiprion percula*). *Peer J* 6:e5841.
- Clutton-Brock, T.H. (1988). Reproductive success. In Reproductive Success (Clutton-Brock, T.H. ed.), 472-486, University of Chicago Press.
- Coulson, T., Benton, T.G., Lundberg, P., Dall, S.R.X., Kendall, B.E. & Gaillard, J.M. (2006). Estimating individual contributions to population growth: evolutionary fitness in ecological time. *Proc. R. Soc. Lond. B. Biol. Sci.*, 273, 547-555.
- Danchin, E., Pocheville, A., Rey, O., Pujol, B. & Blanchet, S. (2019). Epigenetically facilitated mutational assimilation: epigenetics as a hub within the Inclusive Evolutionary Synthesis. *Biol. rev.*, 94, 259-282.

- de Villemereuil, P., Schielzeth, H., Nakagawa, S. & Morrissey, M. (2016). General methods for evolutionary quantitative genetic inference from generalized mixed models. *Genetics*, 204, 1281-1294.
- Dunn, D.F. (1981). The clownfish sea-anemones - *Stichodactylidae* (coelenterata, actiniaria) and other sea-anemones symbiotic with pomacentrid fishes. *T. Am. Philo. Soc.*, 71, 3-113.
- Dupont, P., Cohas, A., Allainé, D. & Pradel, R. (2017). Testing determinants of the annual individual fitness: An overall mean mixture model for de-lifing data. *Methods Ecol. Evol.*, doi: 10.1111/2041-210X.12908.
- Facon, B., Pointier, J.-P., Jarne, P. Sarda, V. & David, P. (2008). High genetic variance in life-history strategies within invasive populations by way of multiple introductions. *Curr. Biol.*, 18, 363-367.
- Fisher, R.A. (1930). The genetical theory of natural selection. Clarendon Press, Oxford, U.K.
- Fricke, H.W.Z. (1979), Mating system, resource defense and sex change in the anemonefish *Amphiprion akallopisos*. *Tierpsychol.*, 50, 313-326.
- Gerber, S., Chabrier, P., Kremer, A. (2003). FaMoz: A software for parentage analysis using dominant, codominant and uniparentally inherited markers. *Mol. Ecol. Res.*, 3(3), 479-481.
- Germain, R.M. & Gilbert, B. (2014). Hidden responses to environmental variation: maternal effects reveal species niche dimensions. *Ecol. Lett.*, 17, 662-669.
- Ghalambor, C.K., MacKay J.K., Carroll, S.P., *et al.* (2007). Adaptive versus non-adaptive phenotypic plasticity and the potential for contemporary adaptation in new environments. *Funct. Ecol.*, 21, 394-407.
- Grafen, A. (1988). On the uses of lifetime reproductive success. In *Reproductive success: studies of individual variation in contrasting breeding systems*. Clutton-Brock T.H., University of Chicago Press, Chicago, Illinois, pp. 454-471.
- Gustafsson, L. (1986). Lifetime reproductive success and heritability: Empirical support for Fisher's fundamental theorem. *Am. Nat.*, 128: 761-764.

- 521 Hadfield, J.D. (2010). MCMC methods for multi-response generalized linear mixed models: The
 522 MCMCglmm R Package. *J. Stat. Softw.*, 33, 1-22.
- 523 Hendry, A.P., Schoen, D.J., Wolak, M.E. & Reid, J. (2018). The contemporary evolution of fitness.
 524 *Ann. Rev. Ecol. Evol. Syst.*, 49, 457-476.
- 525 Hobbs, J.P.A., Frisch, A.J., Ford, B.M., Thums, M., Saenz-Agudelo, P., Furby, K.A. *et al.* (2013).
 526 Taxonomic, spatial and temporal patterns of bleaching in anemones inhabited by
 527 anemonefishes *Plos One*, 8, e70966.
- 528 Houle, D. (1992). Comparing evolvability and variability of quantitative traits. *Genetics*, 130, 195-
 529 204.
- 530 Jones, G.P., Almany, G.R., Russ, G.R., Sale, P.F., Steneck, R.S., van Oppen, M.J.H. *et al.* (2009)
 531 Larval retention and connectivity among populations of corals and reef fishes: history, advances
 532 and challenges. *Coral Reefs* 28, 307-325.
- 533 Jones, G.P., McCormick, M.I., Srinivasan & M., Eagle, J.V. (2004) Coral decline threatens fish
 534 biodiversity in marine reserves. *Proc. Natl. Acad. Sci. USA*, 101, 8251-8253.
- 535 Jones, G.P., Planes, S., Thorrold, S.R., (2005) Coral reef fish larvae settle close to home. *Curr. Biol.*
 536 15, 1314-1318.
- 537 Kruuk, L.E.B., Clutton-Brock, T.H., Slate, J., Pemberton, J.M., Brotherstone, S. & Guinness, F.E.
 538 (2000). Heritability of fitness in a wild mammal population. *Proc. Natl. Acad. Sci. USA*, 97,
 539 698-703.
- 540 Kruuk, L.E.B. (2004). Estimating genetic parameters in natural populations using the 'animal model'.
 541 *Philos. Trans. R. Soc. Lond. B. Biol. Sci.*, 359, 873-890.
- 542 Kruuk, L.E.B. & Hill, W.G. (2008). Introduction. Evolutionary dynamics of wild populations: the use
 543 of long-term pedigree data. *Proc. R. Soc. Lond. B. Biol. Sci.*, 275, 593-596.
- 544 Levis, N.A. & Pfenning, D.W. (2016). Evaluating 'Plasticity-First' Evolution in Nature: Key Criteria
 545 and Empirical Approaches. *Trends Ecol. Evol.*, 31, 563-574.
- 546 Lush, J.L. (2008). *Animal Breeding Plans*. Iowa State University Press, Ames, Iowa, 444 pp.

- McFarlane, S.E., Gorrell, J.C., Coltman, D.W., Humphries, M.M., Boutin, S. & McAdam, A.G. (2014). Very low levels of direct additive genetic variance in fitness and fitness components in a red squirrel population. *Ecol. and Evol.*, 4, 1729-1738.
- Meredith, M. & Kruschke, J. (2016) HDInterval: *highest (posterior) density intervals*. R package version 0.1.3. Available at: <https://CRAN.R-project.org/package=HDInterval>.
- Morrissey, M.B., de Villemereuil, P., Doligez, B. & Gimenez, O. (2014). Bayesian approaches to the quantitative genetic analysis of natural populations. In: *Quantitative genetics in the wild*, {eds. Charmantier, A., Garant, D., Kruuk, L.E.B.} Oxford University Press, Oxford, pp. 228-253.
- Morrissey, M.B. & Wilson, A.J. (2010). Pedantics: an r package for pedigree-based genetic simulation and pedigree manipulation, characterization and viewing. *Mol. Ecol. R.*, 10, 711-719.
- Mousseau, T.A. & Roff, D.A. (1987). Natural selection and the heritability of fitness components. *Heredity*, 59: 181-197.
- Mousseau, T.A. & Fox, C.W. (1998). The adaptative significance of maternal effects. *Trends Ecol. Evol.*, 13, 403-407.
- Munday, P.L., Donelson, J.M. & Domingos, J.A. (2017). Potential for adaptation to climate change in a coral reef fish. *Glob. Change Biol.*, 23, 307-317.
- Munday, P.L., Warner, R.R., Monroe, K., Pandolfi, J.M. & Marshall, D.J. (2013) Predicting evolutionary responses to climate change in the sea. *Ecol. Lett.*, 16, 1488–1500.
- Nedosyko, A.M., Young, J.E., Edwards, J.W. & da Silva, K.B. (2014) Searching for a toxic key to unlock the mystery of anemonefish and anemone symbiosis. *Plos One* 9, 970–976.
- Peakall, R. & P.E. Smouse (2012). GenAlex 6.5: Genetic analysis in Excel. Population genetic software for teaching and research-an update. *Bioinformatics*, 28(19), 2537-2539.
- Pemberton, J.M. (2008). Wild pedigrees: the way forward. *Proc. Roy. Soc. B.* 175, 613-621.
- Pinsky, M.L., Saenz-Agudelo, P., Salles, O., Almany, G.R., Bode, M. *et al.* (2017). Marine dispersal scales are congruent over evolutionary and ecological time. *Curr. Biol.*, 27, 149-154.

- Planes, S., Thorrold, S.R. & Jones, G.P. (2009) Larval dispersal connects fish populations in a network of marine protected areas. *Proc. Nat. Acad. Sci. USA*, 106, 5693–5697.
- Postma, E. (2014). Four decades of estimating heritabilities in wild vertebrate populations: Improved methods, more data, better estimates? In *Quantitative Genetics in the Wild*. {eds. Charmantier, A., Garant, D., Kruuk, L.E.B.}. Oxford University Press, Oxford, pp. 16-33.
- Pujol, B., Blanchet, S., Charmantier, A., Danchin, E., Facon, B., Marrot, P. *et al.* (2018). The missing response to selection in the wild. *Trends Ecol. Evol.*, 33, 337-346.
- Pujol, B., Obbard, D.J. & Pannell, J.R. (2010). Symptoms of population range expansion: lessons from phenotypic and genetic differentiation in hexaploid *Mercurialis annua*. *Plant Ecol. Div.*, 3, 103-108.
- Räsänen, K. & Kruuk, L.E.B. (2007). Maternal effects and evolution at ecological time-scales. *Funct. Ecol.*, 21, 408-421.
- R.Core.Team (2018). *R: A language and environment for statistical computing*. R Foundation for Statistical Computing, Vienna, Austria. Available at: <http://www.R-project.org/>
- Saenz-Agudelo, P., Jones, G.P., Thorrold, S.R. & Planes, S. (2011). Detrimental effects of host anemone bleaching on anemonefish populations. *Coral Reefs*, 30, 497-506.
- Salles, O.C., Pujol, B., Maynard, J.A., Almany, G.R., Berumen, M.L., Jones, G.P. *et al.* (2016). First genealogy for a wild marine fish population reveals multi-generational philopatry. *Proc. Natl. Acad. Sci. USA*, 46, 13245-13250.
- Shama, L.N. (2015). Bet hedging in a warming ocean: predictability of maternal environment shapes offspring size variation in marine sticklebacks. *Glob. Chang. Biol.*, 21, 4387-4400.
- Spiegelhalter, D.J., Best, N.G., Carlin, B.P., & van der Linde, A. (2014). The deviance information criterion: 12 years on. *J. R. Stat. Soc. B.*, 76, 485-493.
- van Hooideonk, R., Maynard, J., Tamelander, J., Gove, J., Ahmadi, G., Raymundo, L. *et al.* (2016). Local-scale projections of coral reef futures and implications of the Paris Agreement. *Sci. Rep.*, 6, 39666.

1
2
3
4
5
6
7
8
9
10
11
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

599 Wheelwright, N.T., Keller, L.K. & Postma, E. (2014). The effect of trait type and strength of selection
600 on heritability and evolvability in an island bird population. *Evolution* 68-11, 3325-3336.
601 Wilson, A.J., Réale, D., Clements, M.N., Morrissey, M.M., Postma, E., Walling, C.A. *et al.* (2010).
602 An ecologist's guide to the animal model. *J. Anim. Ecol.*, 79, 13-26.

For Review Only

Table 1. Sources of variation in Lifetime Reproductive Success (LRS) for the Kimbe Island orange clownfish.

Here we reported variance component estimates quantified by using the animal model approach: Additive genetic (V_A), Maternal (V_M), Natal Habitat (V_{NH}), Resident Habitat (V_{RH}), Permanent Environmental (V_{PE}) and Residual (V_R) Variances. We also report size effects as proportions of explained phenotypic variance: narrow-sense heritability (h^2), maternal effects (m^2) and the mean standardized additive genetic variance: evolvability (I_A) for biennial LRS. Measures are expressed on a latent-scale (direct *MCMCglmm* R results) and the observed data-scale (*QGglmm* R back-transformation). 95% credible intervals (CI) are reported for each estimate.

	LRS	LRS
	Latent scale	Observed data-scale
V_A	0.046	0.030
(CI)	(1.381×10^{-3} to 0.146)	(4.94×10^{-4} to 0.060)
V_M	0.067	0.046
(CI)	(2.000×10^{-3} to 0.211)	(8.822×10^{-3} to 0.287)
V_{NH}	0.516	0.450
(CI)	(0.015 to 1.529)	(0.126 to 1.524)
V_{RH}	0.184	0.135
(CI)	(0.264 to 0.473)	(0.038 to 0.457)
V_{PE}	0.737	0.726
(CI)	(0.496 to 0.952)	(0.203 to 2.460)
V_R	0.894	0.963
(CI)	(0.717 to 1.105)	(0.270 to 3.264)
V_{LRS}	2.44	2.35

1
2
3
4
5
6
7
8
9
10
11
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

615

(CI)	(1.71 to 3.65)	(0.65 to 8.05)
<i>h</i>²	0.019	0.013
(CI)	(6.827×10 ⁻⁴ to 0.057)	(4.951×10 ⁻⁵ to 1.227×10 ⁻²)
<i>m</i>²	0.027	0.019
(CI)	(9.157×10 ⁻⁴ to 0.083)	(2.966×10 ⁻⁵ to 2.044×10 ⁻²)
<i>I</i>_A	0.154	0.103
(CI)	(4.643×10 ⁻⁴ to 0.492)	(1.661×10 ⁻³ to 0.511)

For Review Only

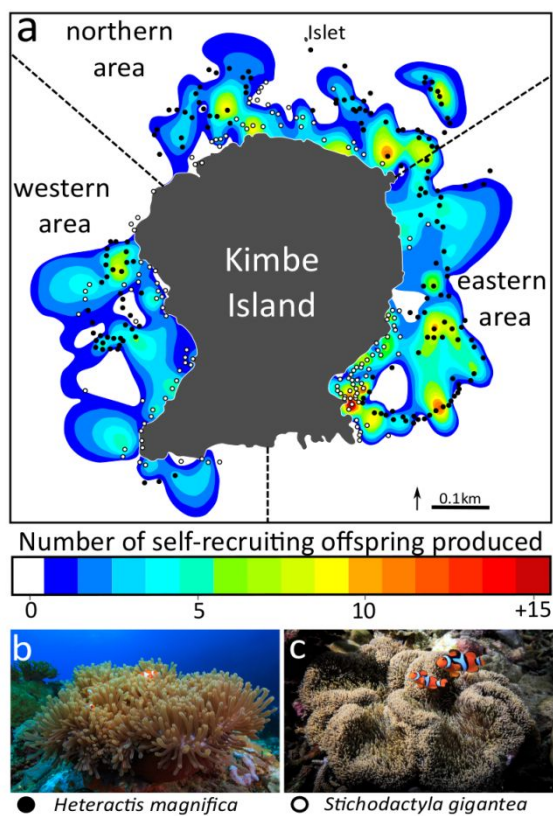


Figure 1. Variation of the total number of offspring orange clownfish produced on each anemone around Kimbe Island between 2003 and 2013.

(a) The studied region was divided in three areas (northern, eastern and western areas). Colors correspond to the variation of the total number of juveniles locally self-recruited that were produced on each anemone (varying from 0 to 27) over a 10-year period. The expected value is interpolated from those around it (using default algorithms implemented in *Origin* software). Dots correspond respectively to the location of the two host anemones species: (b) *Heteractis magnifica* (black dots) and (c) *Stichodactyla gigantea* (white dots). Photos by Tane Sinclair-Taylor.

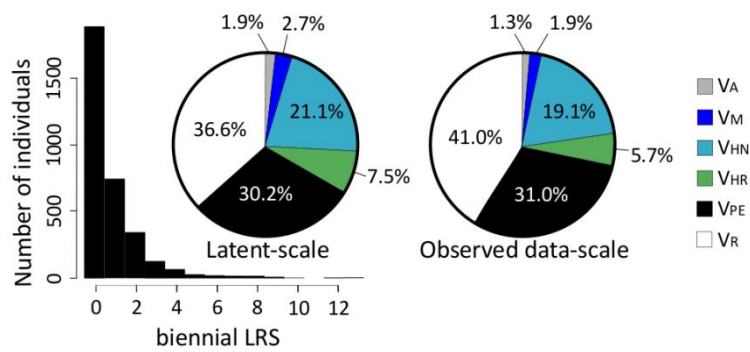


Figure 2. Sources of variation in the biennial estimate of the local Lifetime Reproductive Success (LRS) of the Kimbe Island orange clownfish.

Distribution of the biennial estimate of the LRS (histograms). Variance components on both latent-scale and observed data-scale (pie charts) for the biennial estimate of the LRS explained by Additive genetic (V_A), Maternal (V_M), Natal Habitat (V_{NH}), Resident Habitat (V_{RH}), Permanent Environmental (V_{PE}) and Residual (V_R) variances. These proportions were calculated from the values of the posterior modes of a quantitative genetics generalized linear mixed model analysis (for details see Table 1).

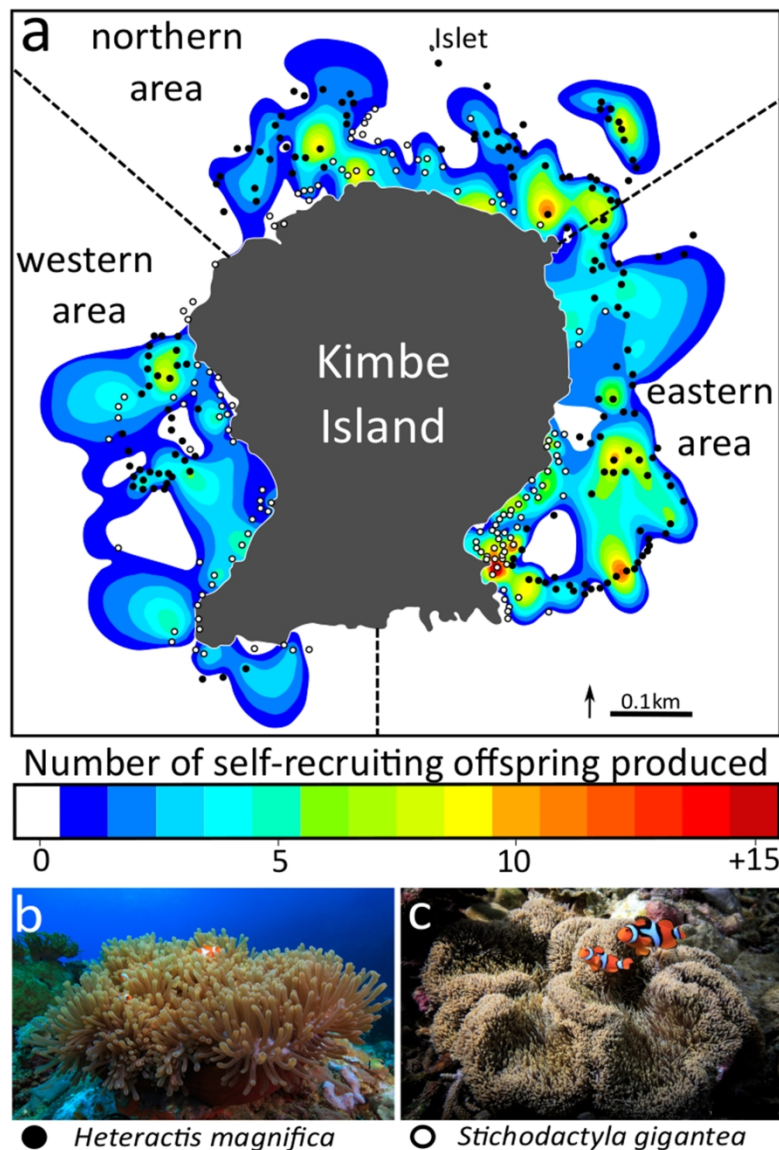


Figure 1. Variation of the total number of offspring orange clownfish produced on each anemone around Kimbe Island between 2003 and 2013.
 (a) The studied region was divided in three areas (northern, eastern and western areas). Colors correspond to the variation of the total number of juveniles locally self-recruited that were produced on each anemone (varying from 0 to 27) over a 10-year period. The expected value is interpolated from those around it (using default algorithms implemented in Origin software). Dots correspond respectively to the location of the two host anemones species: (b) *Heteractis magnifica* (black dots) and (c) *Stichodactyla gigantea* (white dots).
 Photos by Tane Sinclair-Taylor.

234x318mm (300 x 300 DPI)

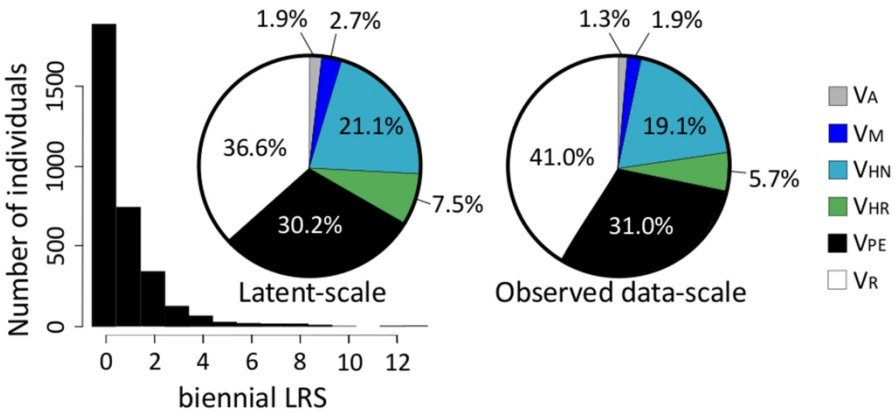


Figure 2. Sources of variation in the biennial estimate of the local Lifetime Reproductive Success (LRS) of the Kimbe Island orange clownfish. Distribution of the biennial estimate of the LRS (histograms). Variance components on both latent-scale and observed data-scale (pie charts) for the biennial estimate of the LRS explained by Additive genetic (V_A), Maternal (V_M), Natal Habitat (V_{VN}), Resident Habitat (V_{HR}), Permanent Environmental (V_{PE}) and Residual (V_R) variances. These proportions were calculated from the values of the posterior modes of a quantitative genetics generalized linear mixed model analysis (for details see Table 1).