



Better off alone? New insights in the symbiotic relationship between the flatworm *Symsagittifera roscoffensis* and the microalgae *Tetraselmis convolutae*

Thibault Androuin, Christophe Six, François Bordeyne, Florian de Bettignies, Fanny Noisette, Dominique Davoult

► To cite this version:

Thibault Androuin, Christophe Six, François Bordeyne, Florian de Bettignies, Fanny Noisette, et al.. Better off alone? New insights in the symbiotic relationship between the flatworm *Symsagittifera roscoffensis* and the microalgae *Tetraselmis convolutae*. *Symbiosis*, 2020, 81, p.161-171. <10.1007/s13199-020-00691-y>. <hal-03019696>

HAL Id: hal-03019696

<https://hal.science/hal-03019696v1>

Submitted on 25 Nov 2020

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.



HAL Authorization

Better off alone? New insights in the symbiotic relationship between the flatworm *Symsagittifera roscoffensis* and the microalgae *Tetraselmis convolutae*

Thibault Androuin^{1,2}, Christophe Six², François Bordeyne², Florian de Bettignies², Fanny Noisette¹, Dominique Davoult².

¹ Institut des Sciences de la Mer, Université du Québec à Rimouski, 310 Allée des Ursulines, Rimouski, Québec, G5L3A1, Canada

² Sorbonne Université, Centre National pour le Recherche Scientifique, Station Biologique de Roscoff, UMR SU-CNRS 7144, 29680 Roscoff, France.

Corresponding author: Thibault Androuin, thibault.androuin01@gmail.com

ORCID: 0000-0001-9310-026X

Short title: Photobiology of *Symsagittifera roscoffensis* and *Tetraselmis convolutae*

Keywords: symbiosis, animal-plant, photobiology, *Symsagittifera roscoffensis*, *Tetraselmis convolutae*

Abstract

The acoel flatworm *Symsagittifera roscoffensis* lives in obligatory symbiosis with the microalgal chlorophyte *Tetraselmis convolutae*. Although this interaction has been studied for more than a century, little is known on the potential reciprocal benefits of both partners, a subject that is still controversial. In order to provide new insights into this question, we have compared the photophysiology of the free-living microalgae to the symbiotic form in the flatworm, both acclimated at different light irradiances. Photosynthesis – Irradiance curves showed that the free-living *T. convolutae* had greater photosynthetic performance (i.e., oxygen production rates, ability to harvest light) than their symbiotic form, regardless of the light acclimation. However, they were affected by photoinhibition under high irradiances, which did not happen for the symbiotic form. The resistance of symbiotic microalgae to photoinhibition were corroborated by pigment analyses, which evidenced the induction of photoprotective mechanisms such as xanthophyll cycle as well as lutein and β -carotene accumulation. These processes were induced even under low light acclimation and exacerbated upon high light acclimation, suggesting a global stress situation for the symbiotic microalgae. We hypothesize that the internal conditions in the sub-epidermal zone of the flatworm (e.g., osmotic and pH), as well as the phototaxis toward high light imposed by the worm in its environment, would be major reasons for this chronic stress situation. Overall, our study suggests that the relationship between *S. roscoffensis* and *T. convolutae* may be a farming strategy in favor of the flatworm rather than a symbiosis with mutual benefits.

1. Introduction

Symbiosis is an interspecific interaction where two or more organisms live in close association, often supported by a high level of integration of the physiology of both partners (Muscatine 1980). Depending on the benefit reciprocity, symbiosis range along a continuum from parasitism, where one partner benefits at the expense of the other(s), to mutualism, where all partners benefit (Tipton et al. 2019). Symbiosis with photosynthetic microalgae occurs in many groups of marine invertebrates such as Porifera, Cnidaria, Mollusca or Chordata (Venn et al. 2008; Rumpho et al. 2011; Melo Clavijo et al. 2018). Among the most studied animal-algal relationships are the reef-building corals, hosting photosynthetic dinoflagellates in their polyps. More discreet and less known, the flatworm *Symsagittifera roscoffensis* (Graff 1891) establishes an obligatory symbiosis with the green alga *Tetraselmis convolutae* (Parke and Manton 1967; Arora 2016; Arboleda et al. 2018). Their interaction is set up at each generation by worm juveniles ingesting microalgae which they will keep for the lifetime of the animal (Keebles 1910). The adult stage does not ingest any organic food and lives autotrophically in the light, in phosphate and nitrate-rich environments, thus depending entirely on the microalgal photosynthetic products (e.g., fatty acid, amino acid, organic acid) (Taylor 1974; Boyle and Smith 1975; Meyer et al. 1979). During the vacuolization process, microalgae lose their cell wall, flagella and eyespot and stay under the epidermis of the worm (Douglas, 1983a). Benefitting the symbiosis, *T. convolutae* uses the uric acid produced by the worm as an endogenous nitrogen source (Boyle & Smith, 1975; Douglas, 1983b). This association is not only biologically remarkable but also ecologically relevant, as the flatworm can form large populations on sandy shores, easily noticeable as dark green patches (Keebles 1910; Doonan and Gooday 1982). The exploitation of this ecological niche would generate an annual primary production rates of 872.9 g C m⁻² (Doonan and Gooday 1982), a value similar to those measured on coral reefs (Muscatine, 1990). Furthermore, the ability of the symbiotic algae to actively uptake nitrates makes *S. roscoffensis* a possible important *in situ* nitrate interceptor in the sediment at local scales (Carvalho et al. 2013).

In many animal-algal associations (i.e., symbiosis, keptoplasty), the fact that the photosynthetic activity of the algal partner is often increased compared to the free-living form, has led to the 'host-mediated enhanced photosynthetic performance' hypothesis (Johnson et al. 2006; Johnson 2011; Serôdio et al. 2014; Decelle et al. 2019). By providing a suitable environment regarding light and nutrient availability, the host may increase the ability of the algae to maximize photosynthetic carbon fixation. In the case of *S. roscoffensis* and *T. convolutae*, the flatworm can also adjust its position along the vertical light gradient of the sediment photic zone, therefore regulating light acclimation of its symbiont (Serôdio et al. 2011; Worley et al. 2019). It has also been suggested that the typical circular milling of the flatworm, which allow the formation of dense and stable biofilm, may promote the photosynthesis of the microalgae (Sendova-Franks et al. 2018). In their study, Serôdio et al. (2011) suggested that non photochemical quenching of chlorophyll fluorescence (i.e., dissipation of excess

energy by heat), a mechanism involved in photoprotection, could also play a role in the regulation of light utilization by the symbiotic algae. This ability to maximize the photobiology of the symbiont might constitute a real advantage for *T. convolutae*, in comparison to other free-living forms of non-motile microalgae in intertidal areas. On the other hand, the study of Nissen et al. (2015) demonstrated that the host's reaction to light stimuli was not adjusted to the symbionts' needs, which questions its behavioral capacity of photosynthetic regulation (Serôdio et al. 2011). Considering the lack of evidence for a true mutualistic relationship (*sensus* Mushegian and Ebert 2015) between *S. roscoffensis*-*T. convolutae* (Selosse 2000; Kiers and West 2016), and the somewhat contradictory results about the behavioral photosynthetic regulation of the flatworm (Serôdio et al. 2011; Nissen et al. 2015), more experiment are needed to addressed the benefit of the symbiosis for the algae and a potential exploitative host control by the flatworm.

In this context, the present study assessed the effect of the symbiosis on the photobiology of the microalga *Tetraselmis convolutae* by comparing the free-living and the symbiotic forms of the alga, acclimated to different light irradiance levels. To do so, we performed oxygen production measurement and pigment analyses in order to understand whether the photosynthesis is increased and/or regulated differently in the host compared to the free-living form.

2. Material and methods

2.1. Biological material and culture conditions

A strain of the green microalga *Tetraselmis convolutae* was isolated from one individual of the acoel flatworm *Symsagittifera roscoffensis* collected on a sandy shore of the French coast of the English Channel (48°44'N, 3°59'W, Roscoff, Brittany, France) on December 7th, 2007. A *T. convolutae* culture was then obtained by micropipette isolation of a single cell released from the specimen. The resulting monoclonal culture was maintained in filter-sterilized seawater supplemented with modified K/2 medium (-Tris, -Si; Keller *et al.*, 1975) at 17°C under a light irradiance of 70-80 $\mu\text{mol photons m}^{-2} \text{s}^{-1}$ in a 12:12 light: dark cycle. The culture was deposited in the Roscoff Culture Collection (<http://roscoff-culture-collection.org/>) as RCC1563.

For the experiment, *T. convolutae* was grown in pyrex erlenmeyer flasks in autoclaved seawater supplemented with 1% Provasoli solution (Provasoli et al. 1968), in a thermostat-water bath at 19-20°C. The cultures were acclimated during 8 days to low (30 $\mu\text{mol photons m}^{-2} \text{s}^{-1}$) and high (350 $\mu\text{mol photons m}^{-2} \text{s}^{-1}$) white light irradiance upon a 12:12 light: dark cycle, provided by an iodine metal halide light source (400 W, Sylvania, Wilmington, USA). The acclimation time was chosen since it is assumed that light acclimation requires one generation time, which varies between 2 hours and 6 days depending on species and physiology (Reynolds 1984; Grobbelaar et al. 1992; MacIntyre et al. 2002). The irradiance levels were adjusted using a quantameter (Quantum LI-190SA, LiCor, Lincoln, USA) and the distance from the light source. The algal cultures were maintained in exponential growth phase by

diluting the cultures with fresh medium every two days and monitoring the optical density at 665 nm with a spectrophotometer (Jenway 6305, JENWAY, Staffordshire, UK). *S. roscoffensis* were collected on a sandy beach in Roscoff in July 2015 (48°44'N, 3°59'W, Brittany, France) and maintained in the same conditions as described above for the cultures of the free-living microalgae.

2.2. Oxygen production measurements

We used a custom-made light chamber to perform oxygen measurements under different light intensities in order to obtain Photosynthesis-Irradiance (P-I) curves. Briefly, this system consisted in a marine water-bath in which the water was maintained at a constant temperature (19-20°C) with continuously running water. The same white light source as described above was placed above the device and a set of 5 neutral density filters (LEE Filters, Andover, UK) placed between the light source and the aquarium, enabling us to set 8 different irradiance levels (from 0 to $\approx 1200 \mu\text{mol photons m}^{-2} \text{ s}^{-1}$). Cultures of *Tetraselmis convolutae* in their growth exponential phase and flatworms (40-50 individuals), along with two controls containing autoclaved seawater, were placed in 2 mL vials (3 and 4 replicates, respectively) and incubated at the different light irradiances. Preliminary experiments allowed the determination of the proper incubation time (30-40 minutes depending on light irradiance) to ensure linear oxygen production/consumption and to avoid Warburg effect (Turner and Brittain 1962).

Oxygen production/consumption rate, translated as net primary production (in the light) and respiration rates (in the dark), were calculated by measuring the dissolved oxygen molar concentration (then converted in mass concentration) at the beginning and the end of the incubation period with a non-invasive optical fiber system (FIBOX 3, PreSens, Regensburg, Germany). The system was calibrated prior to the experiment using sodium dithionite (0 %) and air bubbling (100 %). Net oxygen production, respiration and gross oxygen production rates were corrected from controls and calculated as:

$$R = \frac{\Delta O_2 \times V}{\Delta t \times \text{chl } a} \quad NP = \frac{\Delta O_2 \times V}{\Delta t \times \text{chl } a} \quad GP = NP - R$$

where NP is the net oxygen production rate ($\mu\text{g O}_2 \text{ h}^{-1} \mu\text{g chl } a^{-1}$), R the respiration rate ($\mu\text{g O}_2 \text{ h}^{-1} \mu\text{g chl } a^{-1}$), GP the gross oxygen production rate ($\mu\text{g O}_2 \text{ h}^{-1} \mu\text{g chl } a^{-1}$), ΔO_2 the difference between initial and final O_2 concentrations ($\mu\text{g O}_2$), V the volume of the chamber (mL), Δt the incubation time (h), and chl *a* the chl *a* mass (μg).

At the end of the experiment, the cultures of *T. convolutae* free-living cells were centrifuged at $4\,500 \times g$ and the supernatant was discarded. For the symbiotic forms, the medium was carefully removed with a micropipette. All samples were then stored at -20°C until further analysis. Pigments were fully extracted in the dark at 4°C, during 6 h in 100 % methanol for the free-living cells and 2 h in 100 % ethanol for the symbiotic forms. Absorbance measurements were made using a spectrophotometer, and chl *a* was quantified using the Ritchie equations (Ritchie 2008).

2.3. HPLC pigment analysis

A volume of 3 mL of exponentially growing culture of *T. convolutae* was centrifuged at $4\,500 \times g$ and a total of 60 *Symsagittifera* flatworms by sample were isolated in glass tubes. The cell pellets or the flatworms were resuspended in 100 % acetone for an hour at $-20\text{ }^{\circ}\text{C}$ and sonicated during 5 rounds of 10 sec. The extracts were then centrifuged at $20\,000 \times g$, the supernatants collected and centrifuged again to ensure total removal of particles and cell debris. All sample preparations were carried out under subdued light at 4°C .

The extracts were brought to 25 % Milli-Q water to avoid peak distortion (Zapata and Garrido 1991) and a volume of 100 μL of the pigment extract was immediately injected into a High Pressure Liquid Chromatography system (Hewlett-Packard HPLC 1100 Series, AGILENT, Santa Clara, USA), equipped with a quaternary pump and diode array detector. Pigment separations were performed using a Waters Symmetry C_8 column ($150 \times 3 \times 4.6\text{ mm}$, $3.5\text{ }\mu\text{m}$ particle size) for 38 min at a flow rate of 1 mL min^{-1} , according to procedures previously published (Garrido et al. 2009; Lopes dos Santos et al. 2016). The modified protocol of Garrido *et al.*, (2009) was used in order to allow the separation of neoxanthin and loroxanthin derivatives, with the solutions A = methanol:acetonitrile:aqueous pyridine solution in the proportions 50:40:10 (v/v/v), and B = acetonitrile:acetone (80:20 v/v), following the gradient (min, %B): 0 min, 0 %; 22 min, 40 %; 28 min, 95 %; 38 min, 95 %. In order to separate the zeaxanthin from the lutein peak, the classical procedure of Zapata *et al.*, (2000) was also applied for each sample, with the mobile phases A = methanol:acetonitrile:aqueous pyridine solution (0.025 M) in the proportions 50:25:25 (v/v/v), and B = acetonitrile:acetone (80:20 v/v), following the same solvent gradient.

Chlorophylls and carotenoids were detected by their absorbance at 440 nm and identified by diode array spectroscopy. Pigments were identified and, when possible, the relative quantities were estimated using standards derived from macroalgae and phytoplankton cultures by preparative HPLC (Repeta and Bjørland 1997), using previously compiled extinction coefficients (Roy et al. 2011). The deepoxydation state (DES) of the xanthophyll cycle was calculated for each sample as:

$$\text{DES} = [\text{Zeaxanthin}] / ([\text{Violaxanthin}] + [\text{Antheraxanthin}] + [\text{Zeaxanthin}])$$

2.4. Statistical analyses

Oxygen production measurements were normalized to chl *a* and plotted versus irradiance. The gross oxygen production was preferred to the net oxygen production to take into account the respiration of the flatworm and be able to compare the rates between free-living and symbiotic microalgae forms. Non-linear regression models of photosynthesis considering photoinhibition were fitted to each P-I curve (Platt et al. 1980). Classical photosynthetic parameters were then calculated from the model, including P_{MAX} (maximal oxygen production rate), α (initial slope of the curve), I_K (light intensity at the

onset of light saturation) and β (photoinhibition parameter). P_{MAX} was interpreted as the maximum, potential, light-saturated, oxygen production rate under light condition without photoinhibition (Platt et al. 1980).

Two-way factorial analyses of variance (ANOVA) were used to assess differences in model parameters between *T. convolutae* forms (two levels: free-living and symbiotic) and light condition (two levels: low and high light acclimation). When significant, *post hoc* multiple comparisons were carried out using Tukey HSD. Normality and homocedasticity of the residuals were graphically assessed. When those conditions were not fulfilled, simple factor effect was verified using the non-parametric Wilcoxon test based on ranks. Changes in pigment concentrations were not statistically assessed because only duplicates for free-living cells were available. Statistical analyses were performed in R version 3.3.0 (R Core Team 2016).

3. Results

3.1. Oxygen production measurements

P-I curves showed clear different shapes between the free-living and the symbiotic forms of *Tetraselmis convolutae*, for both light irradiance acclimation (Figure 1a & b). No significant interaction was found between the factors “Living-form” and “Light acclimation” (Table 2).

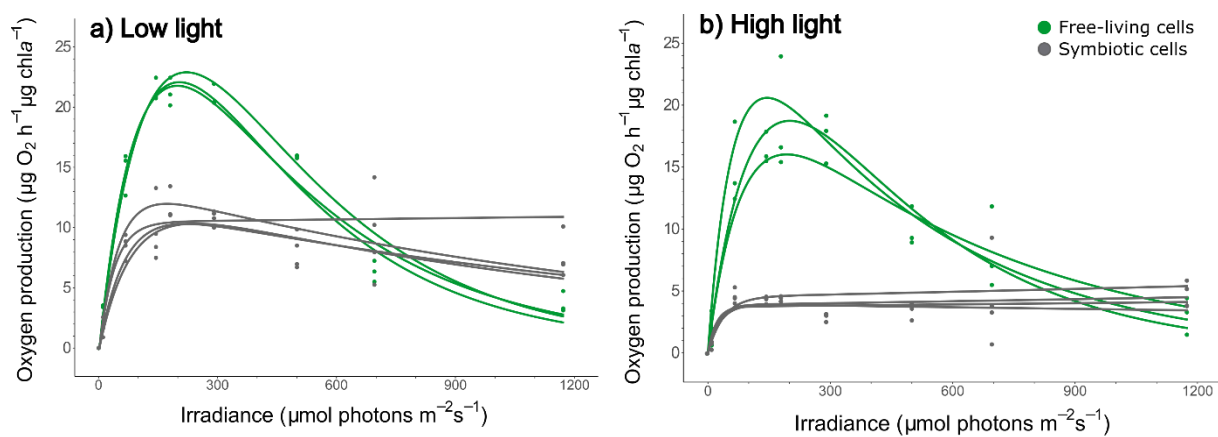


Fig. 1 Photosynthesis-irradiance (P-I) curve of *Tetraselmis convolutae* in the free-living (green) and symbiotic (grey) forms acclimated under low (a; 30 $\mu\text{mol photons m}^{-2} \text{s}^{-1}$) and high (b; 350 $\mu\text{mol photons m}^{-2} \text{s}^{-1}$) irradiance.

Overall, the free-living cells exhibited significantly higher values than symbiotic cells for all the photosynthetic parameters calculated: oxygen production rate (P_{MAX} was 3-8 fold higher in free-living cells than symbiotic ones) while the initial slope of the curve (α), and the light intensity at the onset of light saturation (I_K) were in average 2-fold greater in free living cells. Conversely the free-living cells showed strong photoinhibition, as evidenced by the significant higher values of photoinhibition index (β : 8-17 fold) (Table 1 & 2). Indeed, the oxygen production rate of the free living cells cultures strongly

decreased from about 100 $\mu\text{mol photons m}^{-2} \text{s}^{-1}$ down to values lower than those of the symbiotic algae. No significant change in respiration rate were detected.

Table 1 Mean (\pm SD) of maximal oxygen production rate (P_{MAX}), initial slope (α), saturation irradiance (I_K), photoinhibition index (β) and respiration rate (R), in the free-living and symbiotic forms of *Tetraselmis convolutae*, acclimated to low and high light. P_{MAX} , α , I_K and β were retrieved from the model of Platt *et al.*, (1980) whereas R was experimentally measured.

	Low light		High light	
	Free living cells (n=3)	Symbiotic cells (n=4)	Free living cells (n=3)	Symbiotic cells (n=4)
P_{MAX} ($\mu\text{mol O}_2 \text{h}^{-1} \mu\text{g chl } a^{-1}$)	36.1 \pm 3.3	12.4 \pm 1.4	30.9 \pm 3.9	4.0 \pm 0.3
α ($\mu\text{mol O}_2 \text{h}^{-1} \mu\text{g chl } a^{-1} \mu\text{mol photons m}^{-2} \text{s}^{-1}$)	0.31 \pm 0.01	0.22 \pm 0.06	0.33 \pm 0.1	0.15 \pm 0.03
I_K ($\mu\text{mol photons m}^{-2} \text{s}^{-1}$)	117.1 \pm 5.3	59.5 \pm 19.8	99.3 \pm 24.2	27.9 \pm 7.9
β ($\mu\text{mol O}_2 \text{h}^{-1} \mu\text{g chl } a^{-1} \mu\text{mol photons m}^{-2} \text{s}^{-1}$)	0.17 \pm 0.05	0.01 \pm 0	0.08 \pm 0.04	0 \pm 0
R ($\mu\text{mol O}_2 \text{h}^{-1} \mu\text{g chl } a^{-1}$)	-0.7 \pm 0.4	-1.2 \pm 0.2	0.1 \pm 1.1	-0.2 \pm 0.2

In terms of light acclimated, only the I_K parameter and the respiration varied significantly, regardless of the living form (Table 2). Mean I_K was 84.2 $\mu\text{mol photons m}^{-2} \text{s}^{-1}$ for low light acclimated algae compared to 58.5 $\mu\text{mol photons m}^{-2} \text{s}^{-1}$ for high light acclimated ones. Respiration was 14-fold higher in low light acclimated cell than high-light acclimated ones (-0.96 and -0.07 $\mu\text{g O}_2 \text{h}^{-1} \mu\text{g chl } a^{-1}$ respectively).

Table 2 Factorial two-way ANOVA followed by Tuckey HSD post hoc test showing the effect of light condition (low vs. high light acclimation) and living form (free-living vs. symbiotic cells) of the microalgae *Tetraselmis convolutae* on the initial slope (α), the saturation irradiance (I_K) and the respiration rate (R). The effect on the maximal oxygen production rate (P_{MAX}), and the photoinhibition index (β) was assessed by two distinct Wilcoxon tests. Bold numbers indicate significant level < 0.05 . LL: low light, HL: high light, FL: free-living, S: symbiotic.

	P_{MAX} ($\mu\text{mol O}_2 \text{h}^{-1} \mu\text{g chl } a^{-1}$)			α ($\mu\text{mol O}_2 \text{h}^{-1} \mu\text{g chl } a^{-1} \mu\text{mol photons m}^{-2} \text{s}^{-1}$)			I_K ($\mu\text{mol photons m}^{-2} \text{s}^{-1}$)			β ($\mu\text{mol O}_2 \text{h}^{-1} \mu\text{g chl } a^{-1} \mu\text{mol photons m}^{-2} \text{s}^{-1}$)			R ($\mu\text{mol O}_2 \text{h}^{-1} \mu\text{g chl } a^{-1}$)		
	df	W	P	df	F	P	df	F	P	df	W	P	df	F	P
	Light	Form	Light x Form	Light	Form	Light x Form	Light	Form	Light x Form	Light	Form	Light x Form	Light	Form	Light x Form
	1	13	0.17	1	1.01	0.338	1	8.89	0.01*	1	15	0.26	1	8.51	0.02*
	1	48	< 0.001***	1	15.2	0.002**	1	54.93	< 0.001***	1	48	< 0.001***	1	1.48	0.25
	-	-	-	1	1.87	0.2	-	0.63	0.44	-	-	-	1	0.26	0.62
Tuckey HSD post hoc test															
	Light	-	-	Light	-	-	LL > HL	-	-	LL > HL	-	-	LL > HL	-	-
	Form	FL > S	FL > S	Form	FL > S	FL > S	Form	FL > S	Form	FL > S	Form	FL > S	Form	FL > S	Form

* $P < 0.05$; ** $P < 0.01$; *** $P < 0.001$

3.2. HPLC pigment analysis

The pigments of the free-living form of *T. convolutae*, *i.e.* in culture, were difficult to extract, even though we carried out a strong extraction procedure. It displayed a particularly complex pigmentation with the major pigments being typical of Chlorophyta microalgae (similar to the so-called PRASINO-2A group; Roy *et al.*, 2011), notably including chlorophyll *b*, lutein, violaxanthin, *cis*-neoxanthin and loroxanthin (Figure 2a & b, Table 3, Table S1). *T. convolutae* contain several acyl-loroxanthin compounds, with the two major ones probably being loroxanthin-decenoate and loroxanthin-dodecenoate as previously reported in several other *Tetraselmis* species (Garrido *et al.*

2009) and in *Pyramimonas parkae* (Kohata and Watanabe 1989). We also detected a high number of more or less minor pigments. Using the modified method of Garrido et al. (2009), one of them eluted early and showed absorbance properties identical to *trans*-neoxanthin (peak 1 in Table S1). Two species of carotene were detected in the cell extracts, β,β - and β,ϵ -carotene but it is probable that other forms co-eluted with them and/or with phaeophytin *a*.

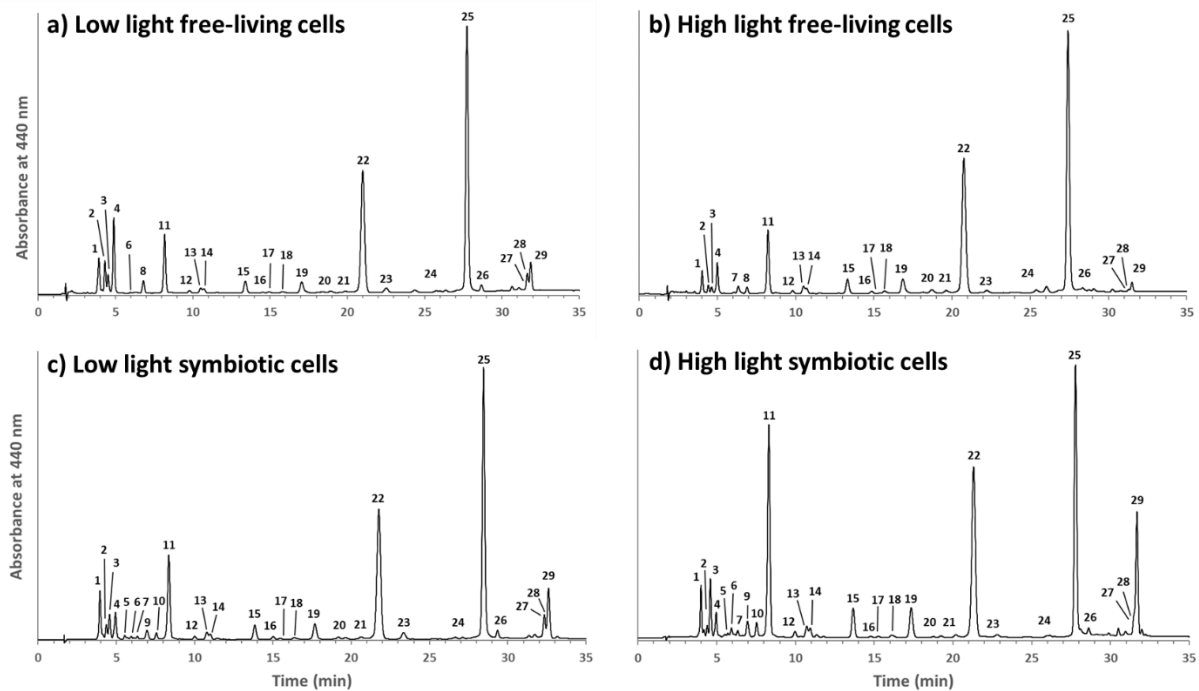


Fig. 2 Chromatographic analysis of *Tetraselmis convolutae* pigmentation (Garrido et al. 2009) as free-living cells acclimated to low (a) and high (b) light, and as a symbiont of *Symsagittifera roscoffensis* acclimated to low (c) and high (d) light. The retention times and wavelength maxima are mentioned in the supplementary material (Table S1). 1: *trans*-neoxanthin like; 2: loroxanthin; 3: *cis*-neoxanthin; 4: violaxanthin; 5: neochrome like; 6: Auroxanthin like; 7: Antheraxanthin; 8: *cis*-neoxanthin like; 9: Auroxanthin like; 10: Dihydrolutein like; 11: Lutein (+ zeaxanthin); 12: unidentified; 13: unidentified; 14: unidentified; 15: Loroxanthin decenoate; 16: Acyl Loroxanthin like; 17: Unidentified; 18: Unidentified; 19: Loroxanthin dodeceonate; 20: Unidentified; 21: Unidentified; 22: Chlorophyll *b*; 23: Chlorophyll *b* derivative; 24: Chlorophyll *a* derivatives; 25: Chlorophyll *a*; 26: Chlorophyll *a* derivative; 27: β,ϵ -carotene; 28: Phaeophytin *a*; 29: β,β -carotene.

In contrast to the cells in culture, we had no difficulty to achieve complete extraction of the pigments for the symbiotic forms. The pigmentation was globally similar to the one of the free-living forms (Figure 2c & d, Table 3). However, a higher proportion of the minor pigments neochrome and auroxanthin was measured in *hospite* microalgae, whereas these pigments were hardly detectable in the free-living cells. In addition, two other pigments showing wavelength maxima quasi identical to *cis*-neoxanthin and dihydrolutein eluted at about 4.5 and 7.5 min, respectively, only in the symbiotic form (Figure 2c & d).

Table 3 Mean (\pm SD) of the major pigments relatively to chlorophyll *a* (mg/g) in the free-living and symbiotic forms of *Tetraselmis convolutae*, acclimated to low and high light. We were not able to quantify all the detected pigments because many of them were in too low amounts to allow purification for calibration of the HPLC system.

	Low light		High light	
	Free living cells (n=2)	Symbiotic cells (n=3)	Free living cells (n=2)	Symbiotic cells (n=3)
Loroxanthin	9.2 \pm 0.8	4.2 \pm 0.6	2.7 \pm 2.7	2.9 \pm 1.3
<i>cis</i> -neoxanthin	4.8 \pm 2.2	5.1 \pm 2.7	3.3 \pm 3.3	15.4 \pm 2.8
Chlorophyll <i>b</i>	479.5 \pm 18.7	499.3 \pm 67.2	549.4 \pm 32.6	628.5 \pm 32.7
Loroxanthin decenoate	6.9 \pm 0.1	7.4 \pm 1.4	11.1 \pm 1.0	16.2 \pm 0.6
Loroxanthin dodecenoate	7.8 \pm 0.2	9.3 \pm 1.2	11.6 \pm 1.1	18.4 \pm 0.8
Lutein	29.5 \pm 2.1	33.4 \pm 4.2	36.6 \pm 2.1	91.0 \pm 8.1
β , β -carotene	7.6 \pm 1.0	12.9 \pm 2.8	3.1 \pm 1.0	43.9 \pm 4.6
Violaxanthin	25.5 \pm 1.3	9.5 \pm 0.1	15.7 \pm 3.7	7.3 \pm 1.1
Antheraxanthin	0.3 \pm 0.1	1.8 \pm 0.4	5.3 \pm 1.8	2.7 \pm 0.2
Zeaxanthin	0.2 \pm 0.3	4.1 \pm 1.6	2.5 \pm 0.3	11.4 \pm 1.7

Acclimation to high light levels induced similar changes in the pigmentation for both free-living and symbiotic forms of *T. convolutae* (Table 3). Several pigments decreased relatively to chlorophyll *a* in response to high light, including *trans*-neoxanthin, loroxanthin, *cis*-neoxanthin (only in the free-living cells), while the chlorophyll *b*:*a* ratio remained fairly stable in all conditions. By contrast, the two major acyl-loroxanthin compounds increased in response to high light acclimation. Photoprotective mechanisms such as the xanthophyll cycle were clearly activated (Demmig-Adams and Adams 1996; Jahns and Holzwarth 2012), as shown by the successive deepoxydation of violaxanthin into antheraxanthin and zeaxanthin in both the free-living and symbiotic forms of *T. convolutae* (Figure 3). This process was accompanied by a significant lutein synthesis (Table 3).

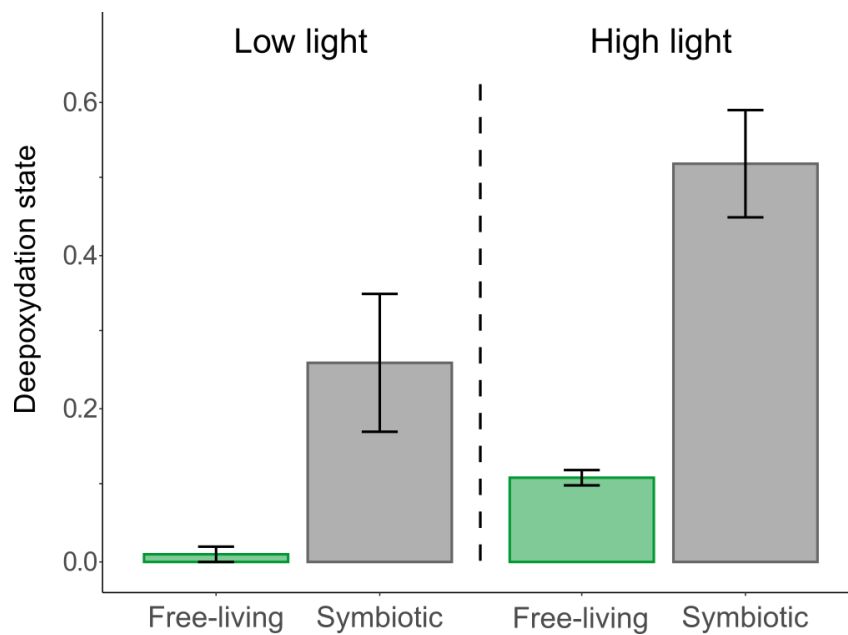


Fig. 3 Deepoxydation state of the xanthophyll cycle in the free-living (green) and symbiotic (grey) forms of *Tetraselmis convolutae*, acclimated to low and high light (mean \pm SD).

Interestingly, there were clear differences in the amplitude of these acclimation processes between the free-living and the symbiotic cells. Under low light, the symbiotic *T. convolutae* showed much higher levels of the photoprotective pigments lutein, zeaxanthin and β -carotene. These marked differences were exacerbated under high light acclimation, upon which the symbiotic cells developed high xanthophyll deepoxydation state and a pronounced induction of *cis*-neoxanthin, lutein along with a strong β -carotene accumulation.

4. Discussion

The symbiosis between the flatworm *S. roscoffensis* and the chlorophyte *T. convolutae* is a remarkable and rare model system that allows studying the level of reciprocity of such interactions (Serôdio et al. 2011; Dupont et al. 2012; Bailly et al. 2014; Nissen et al. 2015; Arboleda et al. 2018). Defining the status of this symbiosis has remained nevertheless challenging. In particular, a number of questions have been recently raised about egalitarian partnerships in nature, with some authors arguing that there is still a lack of evidence for a mutualistic relationship for the *S. roscoffensis*-*T. convolutae* couple, especially for the algae, and that the relationship might be considered as a form of imprisonment of the symbiotic partner (Selosse 2000; Kiers and West 2016). In order to give new insights into these questions, we studied the influence of the symbiotic lifestyle on the algal photobiology.

4.1 The microalgal lifestyle is associated with distinct net oxygen production

The oxygen production curves measured for both algal forms acclimated to two light irradiance levels, unambiguously showed oxygen production was higher in algal free-living form compared to the symbiotic one, at most of the tested irradiances. The only published data on the oxygen production rate of *T. convolutae* (Nozawa et al. 1972), using comparable light acclimation (i.e., $73 \mu\text{mol photons m}^{-2} \text{ s}^{-1}$) and under similar irradiance levels (i.e., $100\text{-}200 \mu\text{mol photons m}^{-2} \text{ s}^{-1}$), indicated 3-4 fold lower values than in our study, for both algal living-forms. Since both light during growing and measurement period were overall comparable (estimated by converting the published data, in foot-candle to units of irradiance), the difference could be attributed to the measurement technique (polarographic electrode vs. optical fiber). Interestingly, the difference between the two living forms was almost identical. The maximal oxygen production rate modelled (i.e., P_{MAX}) in free-living cells, which occurred between 100 and $200 \mu\text{mol photons m}^{-2} \text{ s}^{-1}$, was 3 and 8 fold higher at low light and high light, respectively, than in the symbiotic forms, while Nozawa *et al.* (1972) found a factors of 3 between the two living forms at low light acclimation. To explained this difference, authors proposed that the lower net oxygen production rate observed for the symbiotic form might be due to the host respiration, an increased symbiont photo-respiration, shading effect thanks to the arrangement of the algae within the host, and/or a reduced or host-regulated photosynthetic rate.

Although it is quite possible that the flatworm respired some of the oxygen produced by the alga, it is unlikely that the large differences in oxygen production we observed originated only in the

worm respiration, especially considering the small differences in the calculated respiration rates. Our data suggest that, although it was not statistically significant, the symbiotic system displays slightly higher respiration rates than the free-living algae alone because of the flatworm respiration, and that the microalgae respired at similar rate in any condition and lifestyle. In addition, the P-I curves of the free-living and symbiotic alga showed different shapes that are typical of differences in photosynthesis regulation (see below), which would thus be induced by the alga itself.

Our results thus suggest a different perspective to the idea that algal photosynthetic activity would be enhanced in the host (Johnson 2011; Serôdio et al. 2014; Decelle et al. 2019), as the symbiosis would leads here to a decrease of the global photosystem II activity of the microalgal symbiont.

4.2 Different photoacclimation capacities in different lifestyles

The different shapes of the oxygen production rate *vs.* irradiance curves strongly suggest that the algae regulate the photosynthesis process differently depending on the acclimation irradiance and its lifestyle. The symbiotic microalgae showed lower α and I_K values than the free-living ones, indicating lower efficiency to harvest light and a saturation at lower irradiance level, respectively (Platt et al. 1980; MacIntyre et al. 2002). This is often associated with a reduced functional photosynthetic antenna which, in green algae, is composed of the so-called light harvesting complex proteins. Even though the chl *b:a* ratio did not change, our results thus suggest a downregulation of at least some of these proteins in the alga in the symbiotic form.

In addition, the symbiotic microalgae were much less sensitive to high light irradiances than the free-living ones, as indicated by the photoinhibition phenomenon. This photoinhibition suggests a strong photosystem II photoinactivation at irradiances higher than 200 $\mu\text{mol photons m}^{-2} \text{s}^{-1}$ in the free-living cells. These differences between the two lifestyles were exacerbated upon high light acclimation. In particular, when acclimated to 350 $\mu\text{mol photons m}^{-2} \text{s}^{-1}$, the symbiotic algae were able to maintain a constant photosystem II oxygen evolving up to the highest irradiance tested (i.e. $\approx 1200 \mu\text{mol photons m}^{-2} \text{s}^{-1}$). These observations suggest that, in contrast to the free-living form, the symbiotic *T. convolutae* may chronically induce photoprotection mechanisms that prevent both photoinhibition and photodamages.

Pigment analyses showed clear differences between the two light irradiance acclimation states in both lifestyles and are globally in agreement with a previous study on *Tetraselmis* photoacclimation (Garrido et al. 2009). The reduction of some of the pigments in response to high light in both algal forms may be related to a reduction of the functional size of the photosynthetic antenna (Melis et al. 1999).

The most obvious changes concerned the photoprotection mechanisms. High light acclimation resulted in an increase of the photoprotective xanthophyll cycle activity (Demmig-Adams and Adams 1996), indicating an activation of the violaxanthin deepoxydase enzyme in the symbiont plastids. In addition, lutein accumulation was observed in response to high light pre-exposure in both lifestyles. Although the mechanism has not been yet extensively described, this pigment has been shown to be

induced in response to high light in a number of planktonic chlorophytes (see e.g. Six *et al.*, 2008; Garrido *et al.*, 2009; Lopes dos Santos *et al.*, 2016) and is thought to be involved in photosystem photoprotection (Niyogi *et al.* 1997; Horton and Ruban 2005; Jahns and Holzwarth 2012). Noteworthy, these photoprotective mechanisms were activated much more intensively in the symbiotic lifestyle than in the free-living microalgae. Indeed, the deepoxydation state of the xanthophyll cycle of the symbiotic algae acclimated to 30 $\mu\text{mol photons m}^{-2} \text{ s}^{-1}$ was twofold higher than the one of free-living algae grown under 350 $\mu\text{mol photons m}^{-2} \text{ s}^{-1}$. Similarly, the lutein accumulation was particularly pronounced in the symbiotic microalgae acclimated to high light. These photoprotection mechanisms were likely associated with non-photochemical quenching of fluorescence, although we did not carry out pulse-amplitude modulation (PAM) fluorimetry measurements. At last, the symbiotic microalgae accumulated large amounts of β -carotene. This is a well-known phenomenon in chlorophytes, which triggers this photoprotective and antioxidant mechanism in response to various types of stresses (Ahmed *et al.* 2015).

Our results thus suggest that the symbiotic lifestyle is associated with a stress signal in the alga. The fact that such protective mechanisms are induced even under low light suggests that they can be considered as a global stress, which is not necessarily directly linked to light. It is indeed well known that photoprotection can be related and induced by other environmental factors than light, for instance in temperature and osmotic stress situations, with the aim of decreasing the intraplastidial production of reactive oxygen species originating in photosystem II activity (Foyer and Shigeoka 2011). Considering that free-living cells of *T. convolutae* strain RCC1563 were grown since 2007 at a controlled temperature of 17°C and that symbiotic cells naturally experienced temperature ranging from 16-22°C at the moment of the collection (Doonan and Gooday 1982), the temperature of 20°C in our experiment is not considered as stressful.

4.3 The flatworm as a stressful environment for the symbiotic alga

As recently observed in planktonic symbiotic Acantharians, the algal symbiont may undergo profound ultrastructural and physiological changes upon inclusion in the host (Decelle *et al.* 2019). It has been shown that symbiotic *Tetraselmis* have a different cell structure compare to the free-living cells (Oschman 1966; Provasoli *et al.* 1968). In particular, the inclusion in the flatworm is associated with the loss of a part of the cell envelope, leading to an intimate contact between the algal plasma membrane and the flatworm internal medium (Douglas 1983b). The large differences in the pigment extraction rates we observed are probably related to the presence of the cell envelope, since the free-living cells were much more difficult to extract. The internal medium of the flatworm might thus constitute a stressful environment for *T. convolutae*. As already suggested for kleptoplasts in sea slugs (Jesus *et al.* 2010), the direct contact between the symbiotic algae (then devoid of cell envelope) and the flatworm intern media, which is very different from the seawater, can dramatically change the acido-basic balance of the algal cells. Such drastic medium modification might be considered as a chronic osmotic stress that might activate protective mechanisms in the alga such as, for example, the violaxanthin

deepoxydase gene and those involved in the lutein synthesis. The accumulation of β -carotene is also very well-known to occur in osmotic stress situations, notably in *Dunaliella salina* (Oren 2005). Interestingly, we detected minor amounts of the pigments neochrome and auroxanthin only in the worm samples, whereas these pigments were hardly detectable in the free-living cells. In prasinophytes, these two pigments are often derived from the acidic catalyse of *cis*-neoxanthin and violaxanthin, respectively (Haugan and Liaaen-Jensen 1994). Even though these two carotenoids might have been synthesized during the pigment extraction, their formation support the idea that the flatworm constitutes a very acidic medium for the marine microalga. In addition, it is worth noting that a previous study also suggested that the flatworm may be a stressful environment for the alga. Indeed, Van Bergeijk and Stal (2001) showed that *in hospite* algae contain a 6-fold higher concentration of the algal molecule DMSP than in the free-living cells. The induction of this protectant molecule suggests a stress situation for the algae in the flatworm (Dickson and Kirst 1986; Sunda et al. 2002; Archer et al. 2010).

4.4 From a planktonic to a benthic light niche

Our results show that, in response to the inclusion in the flatworm, the algae induce permanent light-dissipating photoprotection mechanisms that are typical of a pronounced stress situation. The consequence is that the alga in its symbiotic lifestyle is quite resistant to light stress and is able to maintain photosystem II activity up to high irradiances in spite of a lower photosynthetic efficiency than free-living form. This is likely associated with a lower synthesis of photosynthetic compounds available for the host (Hoogenboom et al. 2006).

The chronic induction of protective mechanism appears to be crucial for the survival of the alga in the flatworm because the symbiosis may be associated, for the alga, to dramatic changes in the osmolarity of its environment but also of its light niche. Indeed, in its planktonic form, *T. convolutae* deals with light irradiances more or less attenuated in the water column, depending on the depth. By contrast, in the symbiotic lifestyle, the acclimation to light is dependent of *S. roscoffensis* migrations. The flatworms are often observed at the surface of the sediments at low tide and show a positive phototaxis (Doonan and Gooday 1982; Serôdio et al. 2011). This behavior, associated with a shading effect due to the arrangement of the algae within the host (Nozawa et al. 1972), may explain why the symbiotic cells were acclimated to lower light levels than the free-living ones. However, other studies showed that the flatworm could move in areas where the irradiance was higher than optimal for the algae, likely harming their photosynthetic apparatus (Nissen et al. 2015). It is worth noting that during our 8 days of acclimation, the flatworm did not have sediment to potentially escape from the high irradiance provided. However we did observe migration pattern toward less illuminated areas, which is in agreement with the photoaccumulation experiment of Serôdio et al. (2011). The microalgae must then cope with “abnormally” high light irradiances and it is probable that they could not survive without strong photoprotection. Thus, the symbiotic system *S. roscoffensis* – *T. convolutae* appears to use a similar photophysiological strategy as the intertidal diatom-dominated microphytobenthos, which show

adaptation traits to high irradiances (Cartaxana et al. 2013), while using motility to select the optimal light acclimation (Ezequiel et al. 2015). Our study thus highlights the physiological consequences of a change from a planktonic to a benthic light niche.

4.5 The flatworm as a parasite of the microalga?

The traditional ideas behind mutualism are reciprocal benefits and partner cooperation. However, this concept has been revisited for a few years toward a more parsimonious explanation known as exploitative host control (Wooldridge 2010; Lesser et al. 2013; Keeling and McCutcheon 2017). For instance, in *Paramecium bursaria* and its algal symbiont *Chlorella* sp., the photosynthetic efficiency of the chlorophyte is higher in the free-living microalgae than in the symbiotic ones. The *Chlorella* symbionts can escape their host, but only under conditions where the latter does not benefit from symbiosis, and there are no conditions where this nutritional symbiosis is mutually beneficial (Lowe et al. 2016). In the case of the *S. roscoffensis* - *T. convolutae* relationship, there is still no observation or evidence for possibility for the microalgae to escape from the flatworm (Muscatine et al., 1974), even though once free (mechanically during our isolation process), the symbiotic algae could recover cell envelope, flagella and eyespot (Ian Probert, person. com.). In addition, our data suggest that the microalgae undergo chronic stress in the flatworm, which induce the locking of the photosynthetic apparatus in photoprotective mode, while the flatworm assimilates photosynthetic compounds from the algae (Taylor 1974; Boyle and Smith 1975; Meyer et al. 1979).

The vertical transmission of symbionts is one important feature that can help understand whether a relationship is mutualistic or parasitic (Garcia and Gerardo 2014). Vertical transmission (i.e., symbionts inherited from parents) is expected to lead to a certain level of phylogenetic homogeneity of the symbionts and to increase symbiont effectiveness for the host (Herre et al. 1999). In the case of *T. convolutae*, algae are ingested as free-living cells by juveniles of *S. roscoffensis* and are therefore replaced at each generation (i.e., horizontal transmission), impeding any transgenerational selection. Overall, these results question the mutualistic nature of this relationship (Selosse 2000).

The symbiotic association between *S. roscoffensis* and *T. convoluta* could thus be a farming strategy where the flatworm would use the algae to its own needs, without mutualism retribution (Kiers and West 2016; Sørensen et al. 2019). Further research on the molecule exchanges between both partners and on the consequences of the association on the fitness of *T. convolutae* would allow a deeper understanding of the nature of the relationship.

5. Acknowledgments

This work was supported by the French program Investissement d'Avenir IDEALG. We thank the Roscoff Aquarium Services, especially Sébastien Henry and Ronan Garnier for their technical help and Xavier Bailly for his advices regarding *Symsagittifera roscoffensis* ecology and physiology (Research Federation 2424). We thank the Roscoff Culture Collection, in particular Ian Probert, for the

isolation and maintenance and the *Tetraselmis convoluta* culture. We also thank Alexandra Michiels for help with R coding, and Francisco Rodriguez and José Luis Garrido for the interesting discussions on the HPLC results. Finally, we are grateful to the anonymous reviewer whose suggestions greatly improved the manuscript.

Authors declare no conflict of interest.

6. References

- Ahmed F, Fanning K, Netzel M, Schenk PM (2015) Induced carotenoid accumulation in *Dunaliella salina* and *Tetraselmis suecica* by plant hormones and UV-C radiation. *Appl Microbiol Biotechnol* 99:9407–9416. <https://doi.org/10.1007/s00253-015-6792-x>
- Arboleda E, Hartenstein V, Martinez P, et al (2018) An emerging system to study photosymbiosis, brain regeneration, chronobiology, and behavior: the marine acoel *Symsagittifera roscoffensis*. *BioEssays* 40:1800107. <https://doi.org/10.1002/bies.201800107>
- Archer SD, Ragni M, Webster R, et al (2010) Dimethyl sulfoniopropionate and dimethyl sulfide production in response to photoinhibition in *Emiliania huxleyi*. *Limnol Oceanogr* 55:1579–1589. <https://doi.org/10.4319/lo.2010.55.4.1579>
- Arora M (2016) *Tetraselmis*, an introduction. *The Botanica* 66:155–175
- Bailly X, Laguerre L, Correc G, et al (2014) The chimerical and multifaceted marine acoel *Symsagittifera roscoffensis*: from photosymbiosis to brain regeneration. *Front Microbiol* 5:498 <https://doi.org/10.3389/fmicb.2014.00498>
- Boyle JE, Smith DC (1975) Biochemical interactions between the symbionts of *Convoluta roscoffensis*. *Proc R Soc Lond [Biol]* 189:121–135. <https://doi.org/10.1098/rspb.1975.0046>
- Cartaxana P, Domingues N, Cruz S, et al (2013) Photoinhibition in benthic diatom assemblages under light stress. *Aquat Microb Ecol* 70:87–92. <https://doi.org/10.3354/ame01648>
- Carvalho LF, Rocha C, Fleming A, et al (2013) Interception of nutrient rich submarine groundwater discharge seepage on European temperate beaches by the acoel flatworm, *Symsagittifera roscoffensis*. *Mar Pollut Bull* 75:150–156. <https://doi.org/10.1016/j.marpolbul.2013.07.045>
- Decelle J, Stryhanyuk H, Gallet B, et al (2019) Algal remodeling in a ubiquitous planktonic photosymbiosis. *Curr Biol* 29:968–978.e4. <https://doi.org/10.1016/j.cub.2019.01.073>
- Demmig-Adams B, Adams WW (1996) The role of xanthophyll cycle carotenoids in the protection of photosynthesis. *Trends Plant Sci* 1:21–26. [https://doi.org/10.1016/S1360-1385\(96\)80019-7](https://doi.org/10.1016/S1360-1385(96)80019-7)
- Dickson DMJ, Kirst GO (1986) The role of β -dimethylsulphoniopropionate, glycine betaine and homarine in the osmoacclimation of *Platymonas subcordiformis*. *Planta* 167:536–543
- Doonan S, Gooday G (1982) Ecological studies of symbiosis in *Convoluta roscoffensis*. *Mar Ecol Prog Ser* 8:69–73. <https://doi.org/10.3354/meps008069>
- Douglas AE (1983a) Uric acid utilization in *Platymonas convolutae* and symbiotic *Convoluta roscoffensis*. *J Mar Biol Ass* 63:435–447. <https://doi.org/10.1017/S0025315400070788>

514 Douglas AE (1983b) Establishment of the symbiosis in *Convoluta roscoffensis*. J Mar Biol Ass
515 63:419–434. <https://doi.org/10.1017/S0025315400070776>

516 Dupont S, Moya A, Bailly X (2012) Stable photosymbiotic relationship under CO₂-induced
517 acidification in the acoel worm *Symsagittifera Roscoffensis*. PLoS ONE 7:e29568.
518 <https://doi.org/10.1371/journal.pone.0029568>

519 Ezequiel J, Laviale M, Frankenbach S, et al (2015) Photoacclimation state determines the
520 photobehaviour of motile microalgae: The case of a benthic diatom. J Exp Mar Biol Ecol
521 468:11–20. <https://doi.org/10.1016/j.jembe.2015.03.004>

522 Foyer CH, Shigeoka S (2011) Understanding oxidative stress and antioxidant functions to enhance
523 photosynthesis. Plant Physiol 155:93–100. <https://doi.org/10.1104/pp.110.166181>

524 Garcia JR, Gerardo NM (2014) The symbiont side of symbiosis: do microbes really benefit? Front
525 Microbiol 5:510 <https://doi.org/10.3389/fmicb.2014.00510>

526 Garrido JL, Rodríguez F, Zapata M (2009) Occurrence of loroxanthin, loroxanthin decenoate, and
527 loroxanthin dodecenoate in *Tetraselmis* species (Prasinophyte, Chlorophyta). J Phycol
528 45:366–374. <https://doi.org/10.1111/j.1529-8817.2009.00660.x>

529 Grobbelaar JU, Schanz F, Dubinsky Z, et al (1992) Photosynthetic characteristics of five high light
530 and low light exposed microalgae as measured with ¹⁴C-uptake and oxygen electrode
531 techniques. Mar Microb Food Webs 6:3–19

532 Haugan JA, Liaaen-Jensen S (1994) Blue carotenoids. Part 2. The chemistry of the classical colour
533 reaction of common carotenoid 5,6-Epoxides with acid. Acta Chem Scand 48:152–159

534 Herre E, Knowlton N, Mueller U, Rehner S (1999) The evolution of mutualisms: exploring the paths
535 between conflict and cooperation. Trends Ecol Evol 14:49–53. [https://doi.org/10.1016/S0169-](https://doi.org/10.1016/S0169-5347(98)01529-8)
536 [5347\(98\)01529-8](https://doi.org/10.1016/S0169-5347(98)01529-8)

537 Hoogenboom M, Anthony K, Connolly S (2006) Energetic cost of photoinhibition in corals. Mar Ecol
538 Prog Ser 313:1–12. <https://doi.org/10.3354/meps313001>

539 Horton P, Ruban A (2005) Molecular design of the photosystem II light-harvesting antenna:
540 photosynthesis and photoprotection. J Exp Bot 56:365–373. <https://doi.org/10.1093/jxb/eri023>

541 Jahns P, Holzwarth AR (2012) The role of xanthophyll cycle and of lutein in photoprotection of
542 photosystem II. Biochim Biophys Acta - Bioenergetics 1817:182–193.
543 <https://doi.org/10.1016/j.bbabi.2011.04.012>

544 Jesus B, Ventura P, Calado G (2010) Behaviour and a functional xanthophyll cycle enhance photo-
545 regulation mechanisms in the solar-powered sea slug *Elysia timida* (Risso, 1818). J Exp Mar
546 Biol Ecol 395:98–105. <https://doi.org/10.1016/j.jembe.2010.08.021>

547 Johnson MD (2011) The acquisition of phototrophy: adaptive strategies of hosting endosymbionts and
548 organelles. Photosynth Res 107:117–132. <https://doi.org/10.1007/s11120-010-9546-8>

549 Johnson MD, Tengs T, Oldach D, Stoecker DK (2006) Sequestration, performance, and functional
550 control of Cryptophyte plastids in the ciliate *Myrionecta rubra* (Ciliophora). J of Phycol
551 42:1235–1246. <https://doi.org/10.1111/j.1529-8817.2006.00275.x>

552 Keebles F (1910) Plant Animals, A Study in Symbiosis, Cambridge: University press. Cambridge,
553 UK

554 Keeling PJ, McCutcheon JP (2017) Endosymbiosis: The feeling is not mutual. *J Theor* 434:75–79.
555 <https://doi.org/10.1016/j.jtbi.2017.06.008>

556 Keller MD, Selvin RC, Claus W, Guillard RRL (1975) Media for the culture of oceanic ultraplankton.
557 *J Phycol* 23:633–638

558 Kiers ET, West SA (2016) Evolution: welcome to symbiont prison. *Curr Biol* 26:R66–R68.
559 <https://doi.org/10.1016/j.cub.2015.12.009>

560 Kohata K, Watanabe M (1989) Diel changes in the composition of photosynthetic pigments and
561 cellular carbon and nitrogen in *Pyramimonas parkeae* (Prasinophyte). *J Phycol* 25:377–385

562 Lesser MP, Stat M, Gates RD (2013) The endosymbiotic dinoflagellates (*Symbiodinium* sp.) of corals
563 are parasites and mutualists. *Coral Reefs* 32:603–611. [https://doi.org/10.1007/s00338-013-](https://doi.org/10.1007/s00338-013-1051-z)
564 1051-z

565 Lopes dos Santos A, Gourvil P, Rodríguez F, et al (2016) Photosynthetic pigments of oceanic
566 Chlorophyta belonging to prasinophytes clade VII. *J Phycol* 52:148–155.
567 <https://doi.org/10.1111/jpy.12376>

568 Lowe CD, Minter EJ, Cameron DD, Brockhurst MA (2016) Shining a light on exploitative host
569 control in a photosynthetic endosymbiosis. *Curr Biol* 26:207–211.
570 <https://doi.org/10.1016/j.cub.2015.11.052>

571 MacIntyre HL, Kana TM, Anning T, Geider RJ (2002) Photoacclimation of photosynthesis irradiance
572 response curves and photosynthetic pigments in microalgae and cyanobacteria. *J Phycol*
573 38:17–38. <https://doi.org/10.1046/j.1529-8817.2002.00094.x>

574 Melis A, Neidhardt J, Benemann JR (1999) *Dunaliella salina* (Chlorophyta) with small chlorophyll
575 antenna sizes exhibit higher photosynthetic productivities and photon use efficiencies than
576 normally pigmented cells. *J Appl Phycol* 10:515–525

577 Melo Clavijo J, Donath A, Serôdio J, Christa G (2018) Polymorphic adaptations in metazoans to
578 establish and maintain photosymbioses: Evolution of photosymbiosis. *Biol Rev* 93:2006–
579 2020. <https://doi.org/10.1111/brv.12430>

580 Meyer H, Provasoli L, Meyer F (1979) Lipid biosynthesis in the marine flatworm *Convoluta*
581 *roscoffensis* and its algal symbiont *Platymonas convolutae*. *Biochim Biophys Acta* 573:464–
582 480

583 Muscatine L (1980) Productivity of zooxanthellae. In: Primary productivity in the sea, Plenum. P.G.
584 Falkowski, New York

585 Muscatine L (1990) The role of symbiotic algae in carbon and energy flux in reef corals. In: Coral
586 Reefs, Elsevier. Z. Dubinsky, Amsterdam

587 Muscatine L, Elizabeth Boyle J, Smith DC (1974) Symbiosis of the acoel flatworm *Convoluta*
588 *roscoffensis* with the alga *Platymonas convolutae*. *Proc R Soc Lond [Biol]* 187:221–234

589 Mushegian AA, Ebert D (2015) Rethinking “mutualism” in diverse host-symbiont communities.
590 *BioEssays* 38:100–108. <https://doi.org/10.1002/bies.201500074>

591 Nissen M, Shcherbakov D, Heyer A, et al (2015) Behaviour of the plathelminth *Symsagittifera*
592 *roscoffensis* under different light conditions and the consequences for the symbiotic algae
593 *Tetraselmis convolutae*. *J Exp Biol* 218:1693–1698. <https://doi.org/10.1242/jeb.110429>

594 Niyogi KK, Bjorkman O, Grossman AR (1997) The roles of specific xanthophylls in photoprotection.
595 Proc Natl Acad Sci USA 94:14162–14167. <https://doi.org/10.1073/pnas.94.25.14162>

596 Nozawa K, Taylor DL, Provasoli L (1972) Respiration and photosynthesis in *Convoluta roscoffensis*
597 Graff, infected with various symbionts. Biol Bull 143:420–430

598 Oren A (2005) A hundred years of Dunaliella research: 1905–2005. Saline Syst 1:1–14.
599 <https://doi.org/10.1186/1746-1448-1-2>

600 Oschman JL (1966) Development of the symbiosis of *Convoluta roscoffensis* Graff and *Platymonas*
601 sp. J Phycol 2:105–111

602 Parke M, Manton I (1967) The specific identity of the algal symbiont in *Convoluta roscoffensis*. J Mar
603 Biol Ass 47:445–464

604 Platt T, Gallegos CL, Harrison WG (1980) Photoinhibition of photosynthesis in natural assemblages
605 of marine phytoplankton. J Mar Res 38:103–111

606 Provasoli L, Yamasu T, Manton FRS (1968) Experiments on the resynthesis of symbiosis in
607 *Convoluta roscoffensis* with different flagellate cultures. J Mar Biol Ass 48:465–479

608 R Core Team (2016) R: a language and environment for statistical computing. R Foundation for
609 Statistical Computing, Vienna, Austria

610 Repeta DJ, Bjørland T (1997) Preparation of carotenoids standards. In: Phytoplankton Pigments in
611 Oceanography, UNESCO Publishing. Jeffrey SW, Mantoura RFC, Wright SW, Paris

612 Reynolds CS (1984) The ecology of freshwater phytoplankton., Cambridge University Press.
613 Cambridge, UK

614 Ritchie RJ (2008) Universal chlorophyll equations for estimating chlorophylls *a*, *b*, *c*, and *d* and total
615 chlorophylls in natural assemblages of photosynthetic organisms using acetone, methanol, or
616 ethanol solvents. Photosynthetica 46:115–126. <https://doi.org/10.1007/s11099-008-0019-7>

617 Roy S, Llewellyn CA, Egland ES, Johnsen G (2011) Phytoplankton Pigments, Characterization,
618 Chemotaxonomy, and Applications in Oceanography, Cambridge University Press.
619 Cambridge, UK

620 Rumpho ME, Pelletreau KN, Moustafa A, Bhattacharya D (2011) The making of a photosynthetic
621 animal. J Exp Biol 214:303–311. <https://doi.org/10.1242/jeb.046540>

622 Selosse M-A (2000) Un exemple de symbiose algue-invertébré à Belle-Isle-en-Mer: la planaire
623 *Convoluta roscoffensis* et la prasinophycée *Tetraselmis convolutae*. Acta Bot Gallica
624 147:323–331. <https://doi.org/10.1080/12538078.2000.10515864>

625 Sendova-Franks AB, Franks NR, Worley A (2018) Plant–animal worms round themselves up in
626 circular mills on the beach. R Soc open sci 5:180665. <https://doi.org/10.1098/rsos.180665>

627 Serôdio J, Cruz S, Cartaxana P, Calado R (2014) Photophysiology of kleptoplasts: photosynthetic use
628 of light by chloroplasts living in animal cells. Phil Trans R Soc B 369:20130242.
629 <https://doi.org/10.1098/rstb.2013.0242>

630 Serôdio J, Silva R, Ezequiel J, Calado R (2011) Photobiology of the symbiotic acoel flatworm
631 *Symsagittifera roscoffensis*: algal symbiont photoacclimation and host photobehaviour. J Mar
632 Biol Ass 91:163–171. <https://doi.org/10.1017/S0025315410001001>

- 633 Six C, Finkel ZV, Rodríguez F, et al (2008) Contrasting photoacclimation costs in ecotypes of the
634 marine eukaryotic picoplankter *Ostreococcus*. *Limnol Oceanogr* 53:255–265.
635 <https://doi.org/10.4319/lo.2008.53.1.0255>
- 636 Sørensen MES, Lowe CD, Minter EJA, et al (2019) The role of exploitation in the establishment of
637 mutualistic microbial symbioses. *FEMS Microbiol Lett* 366:fnz148.
638 <https://doi.org/10.1093/femsle/fnz148>
- 639 Sunda W, Kieber DJ, Kiene RP, Huntsman S (2002) An antioxidant function for DMSP and DMS in
640 marine algae. *Nature* 418:317–320. <https://doi.org/10.1038/nature00851>
- 641 Taylor DL (1974) Nutrition of algal-invertebrate symbiosis. I. Utilization of soluble organic nutrients
642 by symbiont-free hosts. *Proc R Soc Lond [Biol]* 186:357–368
- 643 Tipton L, Darcy JL, Hynson NA (2019) A developing symbiosis: enabling cross-talk between
644 ecologists and microbiome scientists. *Front Microbiol* 10:292.
645 <https://doi.org/10.3389/fmicb.2019.00292>
- 646 Turner JS, Brittain EG (1962) Oxygen as a factor in photosynthesis. *Biol Rev* 37:130–170
- 647 Van Bergeijk SA, Stal LJ (2001) Dimethylsulfoniopropionate and dimethylsulfide in the marine
648 flatworm *Convoluta roscoffensis* and its algal symbiont. *Mar Biol* 138:209–216
- 649 Venn AA, Loram JE, Douglas AE (2008) Photosynthetic symbioses in animals. *J Exp Bot* 59:1069–
650 1080. <https://doi.org/10.1093/jxb/erm328>
- 651 Wooldridge SA (2010) Is the coral-algae symbiosis really ‘mutually beneficial’ for the partners?
652 *Bioessays* 32:615–625. <https://doi.org/10.1002/bies.200900182>
- 653 Worley A, Sendova-Franks AB, Franks NR (2019) Social flocculation in plant–animal worms. *R Soc*
654 *open sci* 6:181626. <https://doi.org/10.1098/rsos.181626>
- 655 Zapata M, Garrido JL (1991) Influence of injection conditions in reversed-phase high-performance
656 liquid chromatography of chlorophylls and carotenoids. *Chromatographia* 31:589–594
- 657 Zapata M, Rodríguez F, Garrido JL (2000) Separation of chlorophylls and carotenoids from marine
658 phytoplankton; a new HPLC method using a reversed phase C₈ column and pyridine
659 containing mobile phases. *Mar Ecol Prog Series* 195:29–45

660

661

Supplementary material

Table S1: Chromatogram peak identification, retention time and online spectral characteristics of the pigments detected with the modified method of Garrido et al. (2009) in the free-living and symbiotic forms of *Tetraselmis convolutae*.

Peak #	Pigment	Retention time (min)	Wavelength maxima (nm)		
1	<i>trans</i> -neoxanthin like	3.85	418	442	471
2	Loroxanthin	4.21	(423)	447	474
3	<i>cis</i> -neoxanthin	4.48	414	438	466
4	Violaxanthin	4.86	414	441	470
5	Neochrome like	5.74	400	423	450
6	Auroxanthin like	5.94	381	402	427
7	Antheraxanthin	6.35	(423)	448	475
8	<i>cis</i> -neoxanthin like	4.48	413	437	465
9	Auroxanthin like	6.96	381	402	427
10	Dihydrolutein like	7.53	406	430	455
11	Lutein (+ zeaxanthin)	8.31	(422)	447	475
12	Unidentified	10	423	441	468
13	Unidentified	10.73	(422)	443	471
14	Unidentified	10.85	(421)	443	469
15	Loroxanthin decenoate	11.51	(424)	449	475
16	Acyl Loroxanthin like	12.56	(424)	449	475
17	Unidentified	15.49	(423)	444	472
18	Unidentified	16.41	(423)	445	470
19	Loroxanthin dodeceonate	17.81	(424)	449	476
20	Unidentified	19.25	(424)	448	477
21	Unidentified	20.59	(423)	444	472
22	Chlorophyll <i>b</i>	21.71	464	600	649
23	Chlorophyll <i>b</i> derivative	23.3	463	599	649
24	Chlorophyll <i>a</i> derivatives	24-27	430	617	663
25	Chlorophyll <i>a</i>	28.24	431	617	663
26	Chlorophyll <i>a</i> derivative	29.35	433	617	663
27	β,ϵ -carotene	32.33	(424)	448	476
28	Phaeophytin <i>a</i>	32.42	410	608	666
29	β,β -carotene	32.59	(427)	453	478