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## Fabrication of Robust Spatially-Resolved Photochromic Patterns on Cellulose Papers by Covalent Printing for Anti-Counterfeiting Applications

Guillaume Bretel,<sup>‡</sup> Erwan Le Grognec,<sup>‡</sup> Denis Jacquemin,<sup>‡</sup> Takashi Hirose,<sup>†</sup> Kenji Matsuda,<sup>†</sup> François-Xavier Felpin\*<sup>‡</sup>

<sup>‡</sup>Université de Nantes, CNRS UMR 6230, CEISAM, F-44000 Nantes, France

<sup>†</sup> Department of Synthetic Chemistry and Biological Chemistry, Graduate School of Engineering, Kyoto University, Katsura, Nishikyo-ku, Kyoto 615-8510, Japan

E-mail: fx.felpin@univ-nantes.fr

## ■ ABSTRACT GRAPHIC



## ■ ABSTRACT

Despite its millennial age, cellulose paper remains the preferred material for domestic and professional printings, covering applications from simple office paperwork to fiducial solutions such as bills, passports, and head letters. The creation of robust photochromic patterns on cellulose papers for anti-counterfeiting applications is an important and still partially unaddressed challenge. In this contribution, we report the covalent printing of dibenzothienylethenes as photochromic compounds through a spatially-controlled light mediated thiol-X ligations. Photophysical and theoretical studies provide evidences for a reversible photochromism behavior, not affected significantly by the polar environment of the cellulose matrix, and demonstrates a high fatigue resistance over 18 successive write-erase cycles. The strong coloration-discoloration switch can be easily followed by a direct naked-eye readout.

#### KEYWORDS

Cellulose paper; Photochromic pattern; Thiol-yne reaction; Covalent printing; Dibenzothienylethene, Photoswitch

#### ■ INTRODUCTION

While electronic devices flood our everyday life, paper prints paradoxically remain a modern media of communication and storage in both domestic and professional usages. Indeed, despite its millennial age, cellulose paper is more than ever a modern material as its low cost, disposability, stability, recyclability, and biocompatibility, are all major benefits in the development of sustainable societies.

The fabrication of functional papers, expanding the traditional usages of cellulose paper, is a particularly active field of research with promising applications in area as diverse as medicine,<sup>1-4</sup> catalysis,<sup>5-9</sup> analytical,<sup>10-13</sup> and environmental sciences.<sup>14-16</sup> In this framework, the incorporation of photochromic dyes within the cellulose matrix allows the fabrication of photochromic papers as potential anti-counterfeiting devices or rewritable materials.<sup>17</sup> Creating photochromic patterns on cellulose papers for anti-counterfeiting applications is an ambitious objective needing important requirements: (*i*) a spatial control of the cellulose functionalization; (*ii*) a reversible photochromism, not affected by the cellulose matrix; (*iii*) a robust patterning, insensitive to washing processes; and (*iv*) a good contrast for a naked-eye readout.

The vast majority of published works reports the preparation of photochromic papers using merocyanine dyes as photoactive compounds without any spatial control of the cellulose functionalization, excluding any molecularly imprinted patterns.<sup>18-25</sup> Moreover, when covalently anchored to the material surface, the photochromism of merocyanine dyes is frequently affected by the polar environment of the cellulose matrix as the colored open form is strongly stabilized within the hydrogen bond network of cellulose.<sup>19-20</sup> The irreversibility of the photochromism can be prevented through the impregnation of merocyanine dyes with a polymer carrier within the cellulose matrix,<sup>18, 22, 26</sup> but the creation of photochromic patterns through a spatial control of the functionalization is still not demonstrated through these studies.

Diarylethenes are well-known photochromic compounds, derived from stilbene, which exist under a poorly conjugated open-form, absorbing in the UV region, and a strongly conjugated closed-form presenting a red-shifted absorption, typically in the 500-600 nm

domain.<sup>27-28</sup> Upon UV-light irradiation the open-ring isomer undergoes a  $6\pi$ electrocyclization to furnish the closed-ring isomer which can be back converted upon exposure to visible light. Most diarylethenes are thermally irreversible (P-type) compounds, although, thermally unstable (T-type) diarylethenes have been reported as well.<sup>28</sup> Koga et al. prepared a remarkable photochromic paper by impregnation of cellulose fibers with a solution of diarylethenes in ionic liquids.<sup>29</sup> The absence of charges in both the open- and closed-ring isomers prevents the polar environment of cellulose to excessively stabilize one form with respect to the other and induce an unwanted irreversible photochromism behavior. Again, such strategy excludes any creation of photochromic patterns as paper fibers are all impregnated with the photochromic solution and physisorbed dyes could suffer from a limited stability as they are susceptible to be washed off from the material upon contact with liquids.<sup>30</sup>

Therefore, robust photochromic papers remain to be developed through an original strategy allowing a spatial control of the cellulose functionalization. In this work, we report the first strategy allowing the creation of robust photochromic patterns on cellulose paper through our concept of covalent printing,<sup>31-32</sup> which employs spatially-controlled thiol-X ligations. Our strategy exploits the photophysical properties of dibenzothienylethenes (DBE)<sup>33</sup> and features: (*i*) an efficient and fast coloration-discoloration process upon appropriate light irradiation; (*ii*) a thermal stability of the DBE isomers as shown by both theory and experiment; (*iii*) a good color contrast; and (*iv*) a high fatigue resistance, allowing many cycles to be performed without significant loss of performances.

#### EXPERIMENTAL SECTION

**General remarks**. All commercial solvents and reagents were used as received from Sigma-Aldrich, Fischer Scientific Ltd or Alfa Aesar. Whatman grade 6 filter paper was used as the cellulose source. <sup>1</sup>H and <sup>13</sup>C NMR spectra were recorded at 300 or 400 MHz and 75 or 100 MHz, respectively. Proton chemical shifts were internally referenced to the residual proton resonance in CDCl<sub>3</sub> (7.26 ppm). Carbon chemical shifts were internally referenced to the deuterated solvent signals in CDCl<sub>3</sub> (77.16 ppm). Melting points were recorded on a Stuart Scientific 7SMP3 apparatus. FT-IR spectra were recorded on a Bruker Tensor 27 spectrometer by the ATR technique. HRMS spectra were recorded on a Xevo G2-XS Qtof. UV-visible absorption spectra were recorded using a Shimadzu UV-2501 PC

spectrophotometer for all compounds before the grafting onto cellulose and a Perkin Elmer lambda 1050 was used for the study of papers.

General procedure for the pre-treatment of cellulose filters. A piece of paper (ca. 140 mg) was immersed in an aqueous solution of NaOH (40 mL, 10 wt%) and the mixture was shaken for 24 h on an orbital agitator. The paper was washed 6 times with EtOH and stored in EtOH.

Synthesis of dithiodiglycolic acid. A round-bottom flask was charged with 1% H<sub>2</sub>O<sub>2</sub> aqueous solution (120 mL), tetrabutylammonium iodide (200 mg, 0.54 mmol) and thioglycolic acid (5 g, 54.3 mmol). The resulting mixture was stirred for 1 h. The crude product was extracted with ethyl acetate (3 × 50 mL). The combined organic layers were washed with water (5 × 40 mL), dried with MgSO<sub>4</sub> and concentrated under vacuum to give the corresponding product as a red powder (9.88 g, 99%) which was used without further purification.

**Grafting of dithiodiglycolic acid onto cellulose paper.** A pre-treated cellulose paper was washed 3 times with anhydrous toluene and placed in a flame-dried round-bottom flask under argon with dry toluene (20 mL). A solution of BF<sub>3</sub>.Et<sub>2</sub>O in toluene (1% in 1.1 mL) and dithiodiglycolic acid (471.9 mg, 2.59 mmol) was added to the reaction mixture. The resulting solution was stirred for 24 h at 110 °C on an orbital agitator, protected from light and under argon. Then, the paper was washed successively with EtOH, MeOH, acetone and CH<sub>2</sub>Cl<sub>2</sub>, dried under vacuum and stored under argon.

**2-Methylbenzothiophene (1)**. A four necked flask was charged with benzothiophene (5 g, 37.26 mmol) in dry THF (40 mL) and cooled to -78 °C. *n*-BuLi (25.6 mL, 40.99 mmol, 1.6 M in hexane) was added slowly and the mixture was stirred for 1 h. Then, methyl iodine (2.6 mL, 40.99 mmol) was added and the solution was stirred for 40 minutes at -78 °C. The resulting mixture was quenched with cold water (100 mL), extracted with ether (2 × 50 mL), washed with brine (100 mL) and concentrated to give **1** as a white powder (5.46 g, 99%). mp 50-51 °C [Lit.<sup>34</sup> 51–52 °C]. IR (ATR) *v* 1430, 1196, 1134, 1016, 865, 828, 743, 721, 562, 488 cm<sup>-1</sup>. <sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz)  $\delta$  7.76 (d, 1H, *J* = 7.9 Hz), 7.66 (d, 1H, *J* = 7.4 Hz), 7.34–7.23 (m, 2H), 6.98 (s, 1H), 2.60 (s, 3H). <sup>13</sup>C NMR (CDCl<sub>3</sub>, 100 MHz)  $\delta$  141.0, 140.6, 139.9, 124.2, 123.5, 122.7, 122.1, 121.7, 16.3. HRMS (ASAP<sup>+</sup>) *m*/*z* [M + H]<sup>+</sup> Calcd for C<sub>9</sub>H<sub>9</sub>S 149.0425; Found 149.0427.

**3-Bromo-2-methylbenzothiophene**. In a round-bottom flask, a solution of 2methylbenzothiophene (5.45 g, 36.82 mmol) in AcOH/CHCl<sub>3</sub> (1/1, 100 mL) was stirred with N-bromosuccinimide (7.21 g, 40.49 mmol). The reaction mixture was stirred for 17 h. Then, a part of chloroform and acetic acid was evaporated (*ca.* 80 mL) and a saturated aqueous Na<sub>2</sub>S<sub>2</sub>O<sub>3</sub> solution (60 mL) was added and stirred for 5 min. The resulting mixture was extracted with CH<sub>2</sub>Cl<sub>2</sub> and washed with a saturated aqueous NaHCO<sub>3</sub> solution and H<sub>2</sub>O. The organic layer was dried over MgSO<sub>4</sub> and concentrated under vacuum. Purification by flash chromatography (pentane) afforded the titled compound as a white solid (7.83 g, 95%). mp 42–43 °C [Lit.<sup>35</sup> 43–45 °C]. IR (ATR) *v* 1432, 1300, 1252, 916, 748, 719 cm<sup>-1</sup>. <sup>1</sup>H NMR (CDCl<sub>3</sub>, 300 MHz)  $\delta$  7.75–7.70 (m, 2H), 7.44–7.31 (m, 2H), 2.57 (s, 3H). <sup>13</sup>C NMR (CDCl<sub>3</sub>, 75 MHz)  $\delta$  138.6, 137.3, 135.4, 125.0, 124.9, 122.7, 122.3, 106.7, 15.65. HRMS (ASAP<sup>+</sup>) *m/z* [M]<sup>+</sup> Calcd for C<sub>9</sub>H<sub>7</sub>SBr 225.9452; Found 225.9455.

**3,3'-(Perfluorocyclopent-1-ene-1,2-diyl)bis(2-methylbenzothiophene) (4).** To a solution of 3-bromo-2-methylbenzothiophene (6.23 g, 27.7 mmol) in dry THF (60 mL) under N<sub>2</sub> atmosphere, *n*-BuLi (19.04 mL, 30.5 mmol, 1.6 M in hexane) was added dropwise at -78 °C. The mixture was stirred for 30 minutes and octafluorocyclopentene (1.66 mL, 12.47 mmol) was added dropwise to the mixture with a cooled syringe. The resulting mixture was stirred for 2 h at -78 °C, warmed to room temperature and quenched with cold water (100 mL). The organic phase was extracted with ether (3 × 50 mL) and the organic layers were collected, washed with brine (2 × 50 mL), dried over MgSO4, filtered and evaporated under vacuum. The resulting mixture was purified by flash column chromatography (100% hexane) to give **4** as a yellow solid (3.62 g, 62%) as mixture of parallel (p) and antiparallel (ap) isomers in a ratio of 36:64. mp 153–154 °C [Lit.<sup>36</sup> 154–156 °C]. IR (ATR) *v* 1437, 1337, 1274, 1102, 962, 755, 735, 574, 565 cm<sup>-1</sup>. <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>)  $\delta$  7.72–7.58 (m, 4H), 7.42–7.16 (m, 4H), 2.52 (s, 6H × 0.36, p), 2.25 (s, 6H × 0.64, ap). <sup>13</sup>C NMR (CDCl<sub>3</sub>, 75 MHz)  $\delta$  142.8, 142.3, 138.5, 138.3, 124.8, 124.6, 124.5, 122.2, 122.3, 122.0, 119.3, 15.29. HRMS (ASAP<sup>+</sup>) *m/z* [M + H]<sup>+</sup> Calcd for C<sub>23</sub>H<sub>15</sub>F<sub>6</sub>S<sub>2</sub> 469.0519; Found 469.0514.

**Dibenzothienylethenes (5) and (6).** A solution of 3,3'-(perfluorocyclopent-1-ene-1,2diyl)bis(2-methylbenzothiophene) **4** (2.65 g, 5.66 mmol) in acetic acid (200 mL) was treated with water (9.4 mL), sulfuric acid (4 mL), iodine (1.72 g, 6.79 mmol) and orthoperiodic acid (554 mg, 2.43 mmol) at room temperature. The mixture was stirred for 3 h at 70 °C and the solution was poured into water (500 mL). The organic phase was extracted with AcOEt ( $2 \times$ 100 mL). The organic extracts were washed with water (100 mL), saturated aqueous NaHCO<sub>3</sub> solution (100 mL) and saturated aqueous Na<sub>2</sub>S<sub>2</sub>O<sub>3</sub> solution (100 mL). The resulting solution was dried over MgSO<sub>4</sub>, filtered and concentrated. The residue was purified flash chromatography (100% hexane) to afford a white solid (2.77 g, 72%) as a mixture of 5/6 in a ratio of 33/66 which was directly used in the next step.

**Dibenzothienylethenes (7) and (8).** Under nitrogen atmosphere, a solution of a mixture of 5/6 in a ratio of 33/67 (760 mg, 0.374 mmol 5 and 0.747 mmol 6), Pd(PPh<sub>3</sub>)<sub>2</sub>Cl<sub>2</sub> (137 mg, 0.195 mmol), CuI (411 mg, 2.16 mmol) and trimethylsilylacetylene (1.42 g, 2.16 mmol) in Et<sub>3</sub>N (11 mL) was heated at 80 °C for 15 h. The crude mixture was hydrolyzed with water (10 mL). Et<sub>3</sub>N was removed under vacuum and the solution was extracted with CH<sub>2</sub>Cl<sub>2</sub> ( $3 \times 50$  mL). The organic extracts were washed with brine (50 mL), dried with MgSO<sub>4</sub> and concentrated under reduced pressure. The residue was purified with short cake of celite followed by a flash chromatography on silica gel (100% hexane) to afford 7 (200 mg, 95%, based on 5) and 8 (449 mg, 91%, based on 6) as yellow solids.

**Dibenzothienylethene** 7 was isolated as a mixture of parallel (p) and antiparallel (ap) isomers in a ratio of 38/62. mp 121–122 °C. IR (ATR) *v* 1272, 1250, 1139, 1110, 897, 841, 756 cm<sup>-1</sup>. <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>)  $\delta$  7.83–7.19 (m, 7H), 2.49 (s, 6H × 0.38, p), 2.23 (s, 3H x 0.62, ap), 2.22 (s, 3H × 0.62, ap), 0.30 (s, 9H × 0.62, ap), 0.27 (s, 9H × 0.38, p). <sup>13</sup>C NMR (CDCl<sub>3</sub>, 75 MHz)  $\delta$  144.6, 144.1, 142.8, 142.4, 138.4, 138.3, 138.2, 128.5, 128.3, 125.9, 125.7, 124.8, 124.6, 122.3, 122.0, 121.9, 119.5, 119.2, 104.9, 95.2, 94.9, 88.2, 86.1, 15.4, 15.2, 0.1. HRMS (ES<sup>-</sup>) *m/z* [M - H]<sup>-</sup> Calcd for C<sub>28</sub>H<sub>21</sub>F<sub>6</sub>S<sub>2</sub>Si 563.0758; Found 563.0748.

**Dibenzothienylethene 8** was isolated as a mixture of parallel (p) and antiparallel (ap) isomers in a ratio of 41/59. mp 198–199 °C ; IR (ATR) *v* 1462, 1343, 1271, 1146, 1106, 840, 820 cm<sup>-1</sup>. <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>)  $\delta$  7.81 (s, 2H × 0.59, ap), 7.73 (s, 2H × 0.41, p), 7.53 (d, 2H × 0.59, *J* = 8.3 Hz, ap), 7.47–7.38 (m, 2H), 7.24 (s, 2H × 0.41, p), 2.46 (s, 6H × 0.41, p), 2.18 (s, 6H × 0.59, ap), 0.26 (s, 18H × 0.59, ap), 0.24 (m, 18H × 0.41, p). <sup>13</sup>C NMR (CDCl<sub>3</sub>, 75 MHz)  $\delta$  144.6, 144.1, 138.1, 128.4 128.4, 125.9, 125.8, 121.9, 121.6, 119.5, 119.3, 104.8, 95.2, 15.4, 0.1. HRMS (ES<sup>+</sup>) *m/z* [M + Na]<sup>+</sup> Calcd for C<sub>33</sub>H<sub>30</sub>F<sub>6</sub>NaS<sub>2</sub>Si<sub>2</sub> 683.1129; Found 683.1145.

**Dibenzothienylethene (9(0)).** A solution of 7 (100 mg, 0.177 mmol) in a mixture of CH<sub>2</sub>Cl<sub>2</sub> (20 mL) and MeOH (10 mL) was treated with K<sub>2</sub>CO<sub>3</sub> (600 mg, 4.35 mmol) under nitrogen. The resulting mixture was stirred for 1 h at room temperature and quenched with NH<sub>4</sub>Cl (50 mL). The organic phase was extracted with CH<sub>2</sub>Cl<sub>2</sub> ( $3 \times 50$  mL) and washed with water ( $2 \times 20$  mL). The resulting solution was dried over MgSO<sub>4</sub>, filtered and concentrated to give **9(0)** as a yellow solid (83.6 mg, 96%) as a mixture of parallel (p) and antiparallel (ap) isomers in a ratio of 37/63. mp: 163–164 °C. IR (ATR) *v* 3302, 2108, 1345, 1273, 1101, 965, 725, 646 cm<sup>-1</sup>. <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>)  $\delta$  7.88–7.12 (m, 7H), 3.12 (s, 1H × 0.63, ap), 3.06

(s, 1H × 0.37, p), 2.49 (s, 6H × 0.37, p), 2.23 (s, 3H × 0.63, ap), 2.21 (s, 3H × 0.63, ap). <sup>13</sup>C NMR (CDCl<sub>3</sub>, 75 MHz)  $\delta$  144.8, 144.2, 142.9, 142.4, 138.5, 138.4, 138.2, 138.1, 128.6, 128.4, 126.1, 125.9, 124.8, 124.7, 124.6, 122.3, 122.1, 122.1, 119.4, 119.2, 118.4, 83.5, 77.9, 15.4, 15.3. HRMS (ASAP<sup>+</sup>) m/z [M]<sup>+</sup> Calcd for C<sub>25</sub>H<sub>14</sub>F<sub>6</sub>S<sub>2</sub> 492.0441; Found 492.0435.

**Dibenzothienylethene (10(o)).** A solution of **8** (231 mg, 0.35 mmol) in a mixture of CH<sub>2</sub>Cl<sub>2</sub> (20 mL) and MeOH (10 mL) was treated with K<sub>2</sub>CO<sub>3</sub> (1.38 g, 10 mmol) under nitrogen. The resulting mixture was stirred for 1 h at room temperature and quenched with saturated aqueous NH4Cl solution (50 mL). The organic phase was extracted with CH<sub>2</sub>Cl<sub>2</sub> (3 × 50 mL) and washed with water (2 × 50 mL). The organic extracts were dried over MgSO4, filtered and concentrated under vacuum to give **10(o)** as a yellow solid (179 mg, 99%) as mixture of parallel (p) and antiparallel (ap) isomers in a ratio of 40/60. mp: 194–195 °C. IR (ATR)  $\nu$  3302, 2111, 1461, 1336, 1269, 1106, 965, 876, 816 cm<sup>-1</sup>. <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>)  $\delta$  7.84 (s, 2H × 0.6, ap), 7.75 (s, 2H × 0.4, p), 7.57 (d, 2H × 0.6, *J* = 8.3 Hz, ap), 7.48 (m, 2H), 7.30 (m, 2H × 0.4, p), 3.12 (s, 2H × 0.6, ap), 3.08 (s, 2H × 0.4, p), 2.48 (s, 6H × 0.4, p), 2.20 (s, 6H × 0.6, ap). <sup>13</sup>C NMR (CDCl<sub>3</sub>, 75 MHz)  $\delta$  144.8, 138.4, 138.2, 128.6, 128.5, 126.2, 126.0, 122.0, 121.8, 119.3, 118.5, 83.4, 77.9, 15.4. HRMS (ASAP<sup>+</sup>) *m/z* [M]<sup>+</sup> Calcd for C<sub>27</sub>H<sub>14</sub>F<sub>6</sub>S<sub>2</sub> 516.0441; Found 516.0432.

**Dibenzothienylethene (11).** A solution of **8** (300 mg, 0.454 mmol) in CH<sub>2</sub>Cl<sub>2</sub> (15 mL) under inert atmosphere was treated with *m*-CPBA (673 mg, 2.73 mmol, 70 % in water) at 0 °C and the resulting mixture was stirred for 48 h at 25 °C. The organic phase was extracted with CH<sub>2</sub>Cl<sub>2</sub> (3 × 50 mL) and washed with water (2 × 10 mL). The organic extracts were dried over MgSO4, filtered and concentrated under vacuum to give **11** as a yellow solid (329 mg, 89%) as a mixture of parallel (p) and antiparallel (ap) isomers in a ratio of 52/48. mp 151–153 °C. IR (ATR) *v* 2158, 1697, 1317, 1145, 842 cm<sup>-1</sup>. <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>)  $\delta$  7.81 (s, 1H), 7.77 (s, 1H), 7.63 (d, *J* = 7.7 Hz, 1H), 7.47 (d, *J* = 7.7 Hz, 1H), 7.04 (m, 2H), 2.18 (s, 3H), 2.03 (s, 3H), 0.25 (s, 18H) ; <sup>13</sup>C NMR (CDCl<sub>3</sub>, 75 MHz)  $\delta$  145.0, 144.3, 137.0, 136.8, 135.5, 128.6, 126.7, 126.5, 126.2, 123.6, 122.2, 122.1, 102.0, 100.9, 9.1, -0.22. HRMS (ES<sup>-</sup>) *m/z* [M - H]<sup>-</sup> Calcd for C<sub>33</sub>H<sub>29</sub>O4F6S<sub>2</sub>Si<sub>2</sub> 723.0950; Found 723.0969.

Covalent printing on cellulose paper with dibenzothienylethenes 9(o) - Cell-DBE-1. A piece of Cell-Dis (12 mg), wedged between a photomask and an aluminium plate (2 x 2.5 cm), was immersed in the solution of dibenzothienylethenes 9(o) (135 mg, 0.27 mmol) and 2,2-dimethoxy-2-phenylacetophenone (DMPA, 5 mg, 0.02 mmol) in DMSO (6 mL). The solution, under vigorous stirring, was irradiated with a Xenon lamp (50 W) for 150 minutes.

The paper was washed successively with EtOH, MeOH, acetone and CH<sub>2</sub>Cl<sub>2</sub>, dried under vacuum and stored under argon.

Covalent printing on cellulose paper with dibenzothienylethenes 10(o) - Cell-DBE-2. A piece of Cell-Dis (12 mg), wedged between a photomask and an aluminium plate (2 x 2.5 cm), was immersed in the solution of dibenzothienylethenes 10(o) (140 mg, 0.27 mmol) and 2,2-dimethoxy-2-phenylacetophenone (DMPA, 5 mg, 0.02 mmol) in DMSO (6 mL). The solution, under vigorous stirring, was irradiated with a Xenon lamp (50 W) for 150 minutes. The paper was washed successively with EtOH, MeOH, acetone and CH<sub>2</sub>Cl<sub>2</sub>, dried under vacuum and stored under argon.

**Computational details.** All theoretical calculations have been performed with the Gaussian 16 program.<sup>37</sup> The ground-state geometries of the different dyes have been optimized at the PBE0/6-31G(d) level,<sup>38</sup> taking into account dispersion effects through the use of the D3-BJ approach.<sup>39</sup> These geometry optimization were made using a *tight* convergence threshold and the so-called *ultrafine* DFT integration grid. Next, the minimum nature of all optimized geometries was confirmed by a frequency calculation that returned no imaginary frequency. The excited-state were explored using TD-DFT with the same PBE0 functional but a diffuse containing atomic basis set, namely 6-31+G(d). The thermal transition state between the open and closed forms was determined at the PBE0/6-31G(d) level using the broken-symmetry (BS-DFT) approach and it was verified that the final vibrational mode indeed corresponds to the cyclization process. During all calculations, solvent effects (CH<sub>2</sub>Cl<sub>2</sub>) were accounted fro using the Polarizable Continuum Model (PCM),<sup>40</sup> in its linear-response *non-equilibrium* form for the TD-DFT part of the calculation.

#### RESULTS AND DISCUSSION

**Synthesis of dibenzothienylethenes**. We recently reported the concept of covalent printing onto cellulose paper.<sup>31</sup> The covalent printing strategy uses photoresponsive papers functionalized with dithiodiglycolic ester which upon light irradiation allows thiol-X ligations of chromophoric inks with a powerful spatio-temporal control. The application of the covalent printing strategy to the fabrication of photochromic papers required the synthesis of dibenzothienylethenes bearing reactive alkyne groups for thiol-yne ligations onto the surface of cellulose paper.

The dibenzothienylethene backbone **4** was prepared in two steps from commercially available 2-methylbenzothiophene **1** by adaptation of literature procedures (Scheme 1).<sup>41</sup>

Bromination of 2-methylbenzothiophene 1 with NBS followed by lithium-halogen exchange with *n*-Buli furnished the corresponding lithiated benzothiophene derivative 2 which reacted with octafluorocyclopentene 3 at -78 °C to give 1,2-bis-(2-methylbenzo[b]thiophen-3yl)perfluorocyclopentene 4 in 62% yield over two steps. Iodination of the benzothiophene units furnished an inseparable mixture of mono- and diiodinated dibenzothienylethenes 5/6, in a ratio of 1/2, respectively, and 72% yield. The subsequent introduction of alkyne functional groups through a Sonogashira reaction with trimethylsilylacetylene was carried out on the iodinated mixture 5/6, furnishing the corresponding dibenzothienylethenes 7 and 8 which were separated by flash chromatography. The cleavage of the trimethylsilyl protecting group on 7 and 8 with K<sub>2</sub>CO<sub>3</sub> in CH<sub>2</sub>Cl<sub>2</sub>/MeOH proceeded smoothly to give the expected terminal alkynes 9(0) and 10(0) which were immediately engaged for the covalent printing of cellulose paper due to their rapid degradation upon storage. We also envisaged the preparation of the sulfonyl derivative 12 since it has been reported that the oxidation of benzothienyl moieties significantly increase the fluorescence quantum yield of the closed form, as compared to the corresponding unoxidized dibenzothienylethenes.<sup>42</sup> The oxidized photochromic compound **12** was tentatively prepared in two steps by sulfur oxidation of 8 with *m*-CPBA, furnishing 11 as intermediate, followed by trimethylsilyl cleavage. Unfortunately, the oxidized derivative 12 proved to be unstable and could not be obtained in pure form for further covalent printing on cellulose paper.



SCHEME 1. Preparation of dibenzothienylethenes bearing alkyne functional groups

The covalent printing of 9(0) and 10(0) on photoactive cellulose paper (Cell-Dis), through a thiol-yne ligation, was carried out following an adaptation of our recently developed protocol.<sup>31</sup> Cell-Dis, prepared by grafting of dithiodiglycolic acid on mercerized cellulose paper, was inserted in a sample holder consisting of two aluminum plates; the top plate being machined with the letter "U". Upon immersion in a solution of 9(0) or 10(0) in DMSO, also containing 2,2-dimethoxy-2-phenylacetophenone (DMPA) as radical initiator, Cell-Dis was irradiated with a Xenon lamp (60 W) to print the letter "U" (Scheme 2).



SCHEME 2. General strategy for the covalent printing of photochromic compounds 9(0) and 10(0).

*Material characterization*. The covalent printing of 9(o) and 10(o) onto Cell-DiS was followed by Fourier-transform infrared spectroscopy (FTIR) and X-ray photoelectron spectroscopy (XPS) analyses (Figure 1). The appearance of a new band at *ca*. 1645 cm<sup>-1</sup> on the FTIR spectra of Cell-DBE-1 and Cell-DBE-2 characteristic of C=C stretching frequencies of the dibenzothienylethene backbone, successfully confirmed the grafting. The band at *ca*. 1725 cm<sup>-1</sup> accounts for the C=O stretching vibration of the ester groups confirming that the covalent printing conditions did not affect the thiomethylglycolate function. The remaining bands between 400 to 1500 cm<sup>-1</sup> are characteristic of the finger print of the cellulose backbone while the bands in the region of 3100-3400 cm<sup>-1</sup> are attributed to the stretching of O-H bonds. The XPS spectra of Cell-DBE-1 and Cell-DBE-2 also attested the succesful grafting of 9(o) and 10(o), respectively, through the presence of F1s peaks at *ca*. 688 eV.



**FIGURE 1**. (a) FTIR spectra of **Cell-DiS**, **Cell-DBE-1** and **Cell-DBE-2**. (b) Survey scan spectra of **Cell-DBE-1**. (c) Survey scan spectra of **Cell-DBE-2**. (d) High-resolution F1s spectrum of **Cell-DBE-1**. (e) High-resolution F1s spectrum of **Cell-DBE-2**.

The thermal stability of Cell-DBE-1 and Cell-DBE-2 were assessed using thermogravimetric analysis (TGA) experiments under  $N_2$  atmosphere (Figure 2) and

compared to pristine cellulose paper. The mass loss of ca. 4% below 200 °C for the three samples is attributed to the desorption of volatiles such as residual solvents and moisture. **Cell-DBE-1** and **Cell-DBE-2** display very similar stability profiles and are stable until 250 °C; Cell showing a slighly increased stability, up to 300. The sudden mass loss at 260 °C for **Cell-DBE-1** and **Cell-DBE-2** can be explained by the elimination of the grafted photochromic moieties. This result suggests that **Cell-DBE-1** and **Cell-DBE-2** are stable in a large range of temperatures from 25 to 250 °C.



Figure 2. Thermogravimetric analysis of Cell, Cell-DBE-1 and Cell-DBE-2.

The atomic composition of the patterned surface was determined by XPS and the degree of substitution (DS) was calculated to be ca. 1% for both paper. This value means that 1 glucose unit is functionalized with photochromic compounds every 100 glucose units. As the degree of substitution of **Cell-DiS** by dithiodiglycolic acid units was ca. 14%, the yield of the covalent printing was calculated to be ca. 7%. While the DS of the photochromic papers and the associated covalent printing yields could be hastily considered as unsatisfactory, the absorbance of the closed form at very long-wavelength allowed to follow the photochromic behavior of the patterned papers by naked eyes (*vide infra*). Moreover, a high concentration of the dyes onto the surface of cellulose paper might generate detrimental photochromic effects due to cross interactions.

*Photophysical properties.* In order to evaluate the influence of the cellulose matrix on the photophysical properties of dibenzothienylethenes **9(0)** and **10(0)**, we initially recorded their absorption spectra in solution for comparison with **Cell-DBE-1** and **Cell-DBE-2** (Figure 3). We also used theoretical chemistry to model their properties (*vide infra*).



FIGURE 3. (a) Absorbtion spectra of 9(0) in the ground state. Time evolution of the absorbance of 9(0) (b) under irradiation at 254 nm, (c) under visible light. (d) Absorbtion spectra of 10(0) in the ground state. Time evolution of the absorbance of 10(0) (e) under irradiation at 254 nm, (f) under visible light. All data obtained in CH<sub>2</sub>Cl<sub>2</sub> at  $4 \times 10^{-5}$  M and 25 °C.

The absorption spectra of 9(0) and 10(0), recorded in CH<sub>2</sub>Cl<sub>2</sub> (4 × 10<sup>-5</sup> M) at 25 °C, only show absorption bands in the UV region (< 350 nm), as expected. Upon light irradiation of 9(0) and 10(0) at 254 nm, the corresponding colorless solutions turn deep pink and purple, respectively. Such color change indicates the successful  $6\pi$ -electrocyclization of the ring-open isomers 9(0) and 10(0) to the corresponding closed forms 9(c) and 10(c), respectively. The photocyclization, upon irradiation at 254 nm, is evidenced by the gradual decay of bands in the UV region ( $\lambda < 300$  nm) together with the appearance of strong absorption bands at 359, 371 and 535 nm for the closed-ring isomer 9(c), and 364, 379 and 546 nm for the closed-ring isomer 10(c). These results show that the presence of a second alkyne function in 10(c) with respect to 9(c) redshifts absorption bands due to the increased conjugation (see also calculations below). The photostationary state is reached after 20 minutes of irradiation for 9(c) and 5 min for 10(c). The cycloreversion gradually occurs under daylight (standard lightning in the laboratory) as indicated by the decay of the intensities of the bands in the region of 340-550 nm and the complete bleaching of the solution. The photostationary state is obtained after 5 min for 9(0) and 10 min for 10(0) under daylight. We also confirmed that 9(0) and 10(0) are both P-type photochromic compounds as thermal relaxation is not observed upon prolonged heating in the dark (refluxing CH<sub>2</sub>Cl<sub>2</sub> for 2 hours).

We have performed Density Functional Theory (DFT) calculations to characterize dyes 9(0), 10(0), 9(c), and 10(c) (see Experimental Section for computational details). For the open-ring structures, the relative free energies of the parallel and anti-parallel conformers are nearly equal (difference < 0.5 kcal.mol<sup>-1</sup>) indicating that, consistently with experiment, there is therefore only half of the DBE that can undergo photochromism, a usual outcome in diaryletheyne switches. The closed-ring forms 9(c) and 10(c) are respectively less stable by 7.7 and 8.0 kcal.mol<sup>-1</sup> as compared to the open 9(0) and 10(0), whereas the thermal 10(c)-to-**10(0)** barrier is as large as 36 kcal.mol<sup>-1</sup>, consistent with the observed P-type photochromism. We also carried out TD-DFT calculations to probe the nature of the excited-states. For the hallmark visible absorption of the closed forms, TD-DFT returns transitions at 567 nm (f=0.36) and 584 nm (f=0.47) for 9(c) and 10(c), respectively. These values are in good agreement with the experimental ones (see Figure 3), the redshift upon addition of a second ethynyl group being reproduced. As expected, these transitions are attributed to a HOMO-LUMO transition. These orbitals are displayed on Figure 4. As can be seen, both orbitals are significantly delocalized on the ethynyl segment(s), consistently with an increase of delocalization and hence a redshift of the absorption spectra.



FIGURE 4. Representation of the PCM (CH<sub>2</sub>Cl<sub>2</sub>)-PBE0-D3<sup>BJ</sup>/6-31+G(d) frontier orbitals (bottom: HOMO, top: LUMO) for dyes 9(c) (leftmost), 10(c) (center left) and the dyes corresponding to DBE anchored onto cellulose Cell-DBE-1 (9'(c), center right) and Cell-DBE-2 (10'(c), rightmost).

Reflective UV-Vis spectroscopy was used to follow the photochromic behavior of Cell-DBE-1 and Cell-DBE-2 (Figure 5). Due to the presence of whitening agents, absorbing between 250 to 350 nm, in commercially available cellulose paper, we restricted the absorbance recording at wavelengths >350 nm to follow the characteristic bands of the ringclosed isomers at ca. 540 nm. The time evolution of the absorption in the 400-750 nm window was followed upon irradiation at 254 nm. While the ground-state absorption spectra of Cell-DBE-1 and Cell-DBE-2 does not show any band between 400 and 750 nm, a new band centered at 534 and 548 nm for Cell-DBE-1 and Cell-DBE-2, respectively, appear after 10 seconds of irradiation. These measurements show that the  $\lambda_{max}$  of the absorption bands at ca. 540 nm are similar to the one of 9(0) and 10(0) as the band was blue-shifted by 1 nm on Cell-BDE-1 and red-shifted by 2 nm on Cell-DBE-2, respectively. We have also modeled at the TD-DFT level the properties of the DBE presenting a form equivalent to the one after grafting, that is, the DBE in which we replaced the ethynyl segment(s) by 2-methylthiovinyl moiety(ies), and we denote this species 9' and 10'. The thermodynamic properties were rather comparable to the one of the original switches with nearly equal free energies for the parallel and anti-parallel conformers and a closed form less stable by 6.4 and 4.9 kcal.mol<sup>-1</sup> than the open form for 9' and 10', respectively. The thermal 10'(c)-to-10'(o) barrier is as large as 37 kcal.mol<sup>-1</sup>, very close to the value obtained for **10** (*vide supra*). The PCM (CH<sub>2</sub>Cl<sub>2</sub>)-TD-DFT calculations predict that 9'(c) and 10'(c) have their lowest transition at 581 nm (f=0.66) and 617 nm (f=1.07) indicating significant bathochromic and hyperchromic shifts compared to 9(c) and 10(c) in the same conditions. The strong increase of the oscillator strength is a positive news as it means that even a small ratio of grafted photochrome could be detected. The predicted bathochromic displacements are a logical consequence of the stronger  $\pi$ - conjugation in double than triple bonds (see the extended HOMO and LUMO of 9'(c) and 10'(c) in Figure 4), but remains unseen experimentally when grafting the dyes on the paper, which hints that cellulose induces a small hypsochromic shift compared to solution. In any case, these theoretical and experimental results suggest that the conjugated  $\pi$ -electron systems were not strongly affected by the grafting, highlighting the high value of our thiol-X ligation for preserving the photophysical properties of photochromic compounds. Comparing Cell-DBE-1 and Cell-DBE-2 was also informative as the bi-functional photochromic compound grafted to Cell-DBE-2 redshifts the absorption band by 12 nm with respect to Cell-BDE-1 (534 *vs.* 548 nm). The photostationary state was reached after 20 and 60 minutes for Cell-BDE-1 and Cell-DBE-2, respectively. The reversible photochromic switches were evidenced upon subsequent exposure of the patterned paper to daylight. Then band centered at ca. 540 nm completely disappeared after 40 min for Cell-BDE-1 and 60 min Cell-DBE-2.



FIGURE 5. (a) Absorbtion spectra of Cell-DBE-1 in the ground state. Time evolution of the absorbance of Cell-DBE-1 (b) under irradiation at 254 nm, (c) under visible light. (d)

Absorbtion spectra of **Cell-DBE-2** in the ground state. Time evolution of the absorbance of **Cell-DBE-2** (e) under irradiation at 254 nm, (f) under visible light.

The photochromic behavior of our patterned paper is illustrated on Figure 6. Pictures of patterned papers Cell-BDE-1 and Cell-DBE-2 under daylight (Figures 6a and 6c, leftmost) and after irradiation at 254 nm (Figures 6a and 6c, rightmost) show the strong colorationdiscoloration switch. An excellent color contrast and a good resolution of the printed letter "U" can be observed for both papers. The precision covalent printing allows patterns with line thickness of ca. 250-500 µM. Increasing the precision to line thickness <200 µM would not be relevant for cellulosic material due to a high heterogeneity of the surface composed of fibers with a diameter usually ranging from 10 to 100 µm. The grafting mostly occurs on the surface of the paper as the photochromic patterns are not visible on the opposite side of Cell-BDE-1 and Cell-DBE-2. The repeatability of the photochromic behavior was studied by applying 18 successive write-erase cycles and neither significant color change nor color intensity were observed. Indeed, as initially expected, both the photophysical and fatigue behaviors resistance studies demonstrate that the photochromic of grafted dibenzothienylethenes are much less affected by the polar environment of the cellulose matrix than merocyanine switches.<sup>20, 43</sup> The absence of charges in both the open- and closed forms of dibenzothienylethenes most probably prevents any negative interaction with cellulose that could produce unwanted irreversible photochromism behavior.



**FIGURE 6**. Photochromic behavior of patterned papers **Cell-BDE-1** (a) and **Cell-DBE-2** (c). Variation of absorbance intensity of **Cell-BDE-1** (b) and **Cell-DBE-2** (d) under UV (254 nm) and daylight cycles.

In order to compare the robustness of our covalent printing strategy with a simple adsorption process, we prepared the photochromic paper **Cell-DBE-A** which consists of the photochromic compound **8** adsorbed onto pristine cellulose paper (Figure 7). The high instability of **9(0)** and **10(0)** under the dry state precludes any adsorption test with these photochromic compounds on cellulose. While the photochromic behavior of **8** is visually preserved upon adsorption, a single washing of **Cell-DBE-A** in CH<sub>2</sub>Cl<sub>2</sub> completely washed-off the circular pattern, validating the better robustness of our covalent printing with respect to adsorption processes. The same washing process carried out on **Cell-DBE-2** (Figure 7d-f) does not affect the pattern, confirming that a covalent printing is much more robust than a simple adsorption.



**FIGURE 7**. Photochromic behavior of patterned papers **Cell-BDE-A** (a) under visible light, (b) after irradiation at 254 nm for 1 min, (c) after a single washing in CH<sub>2</sub>Cl<sub>2</sub> followed by an irradiation at 254 nm for 1 min and **Cell-BDE-2** (d) under visible light, (e) after irradiation at 254 nm for 1 min, (f) after a single washing in CH<sub>2</sub>Cl<sub>2</sub> followed by an irradiation at 254 nm for 1 min, (f) after a single washing in CH<sub>2</sub>Cl<sub>2</sub> followed by an irradiation at 254 nm for 1 min.

#### CONCLUSION

We have developed the first covalent patterning of cellulose paper with photochromic compounds. The strategy stands on the spatially-controlled light-mediated thiol-X ligation of dibenzothienylethenes (DBE) as photochromic compounds. The photophysical and theoretical studies are consistent with a reversible photochromic behavior of a thermally-stable DBE, not significantly affected by the hydrogen bond network of the cellulose matrix, and demonstrates the high fatigue resistance of the photochromic material over 18 successive write-erase cycles without decrease of the absorbance intensity. The robust covalent patterns are insensitive to successive washings in sharp contrast with patterns created through a non-covalent physisorbed process. We trust that the creation of robust molecularly imprinted photochromic patterns on cellulose paper is a promising technology that will find applications as anti-counterfeiting device in fiducial documents.

## ■ ASSOCIATED CONTENT

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NMR spectra of compounds: 1, 3-bromo-2-methylbenzothiophene, 4, 9, 7, 8, 9(0), 10(0) and 11 (PDF)

## ■ AUTHOR INFORMATION

## **Corresponding Author**

\*E-mail: fx.felpin@univ-nantes.fr

## ORCID

Erwan Le Grognec: 0000-0002-3351-7028 Denis Jacquemin: 0000-0002-4217-0708 Takashi Hirose: 0000-0002-5351-2101 Kenji Matsuda: 0000-0002-2420-4214 François-Xavier Felpin: 0000-0002-8851-246X **Notes** 

The authors declare no competing financial interest.

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