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# Selectivity via Cooperativity: Preferential Stabilization of the p65/14-3-3 Interaction with Semisynthetic Natural Products

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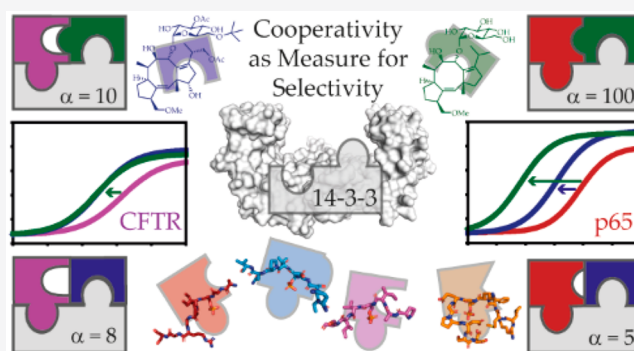
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**ABSTRACT:** Natural compounds are an important class of potent drug molecules including some retrospectively found to act as stabilizers of protein–protein interactions (PPIs). However, the design of synthetic PPI stabilizers remains an understudied approach. To date, there are limited examples where cooperativity has been utilized to guide the optimization of a PPI stabilizer. The 14-3-3 scaffold proteins provide an excellent platform to explore PPI stabilization because these proteins mediate several hundred PPIs, and a class of natural compounds, the fusicocanes, are known to stabilize a subset of 14-3-3 protein interactions. 14-3-3 has been reported to negatively regulate the p65 subunit of the NF- $\kappa$ B transcription factor, which qualifies this protein complex as a potential target for drug discovery to control cell proliferation.

Here, we report the high-resolution crystal structures of two 14-3-3 binding motifs of p65 in complex with 14-3-3. A semisynthetic natural product derivative, DP-005, binds to an interface pocket of the p65/14-3-3 complex and concomitantly stabilizes it. Cooperativity analyses of this interaction, and other disease relevant 14-3-3-PPIs, demonstrated selectivity of DP-005 for the p65/14-3-3 complex. The adaptation of a cooperative binding model provided a general approach to characterize stabilization and to assay for selectivity of PPI stabilizers.



## INTRODUCTION

The need to develop new strategies that address the challenges of today's most important diseases has motivated efforts to modulate protein–protein interactions (PPIs).<sup>1–4</sup> PPI stabilizers are increasingly attractive given the potential for extending the druggable proteome, exploiting the endogenous-ligand uncompetitive mode of binding, and harnessing cooperativity as the driving force to enhance physiological, regulatory mechanisms instead of their artificial disruption.<sup>1–4</sup> Promising progress has been made on both the inhibition and the stabilization of PPIs, although the ab initio design of PPI stabilizers in particular is still an exception.<sup>3</sup> Nature itself provides the most potent and selective stabilizers of PPIs, which validate the concept of stabilization as a valuable strategy for targeting PPIs.<sup>5,6</sup> The mode of action of known PPI stabilizers was mostly discovered retrospectively, as, for example, with rapamycin (Rapamune, Pfizer), cyclosporin (Sandimmun, Novartis Pharmaceuticals), or FK506 (Prograf, Astellas Pharma).<sup>5–7</sup>

To achieve PPI stabilization, the binding equilibrium of two target proteins needs to be shifted to the complexed state by binding of a third interaction partner, preferably a drug-like small molecule. This underlying binding improvement can be described via cooperative binding models whereby the

increased binding affinity of the interaction partners, due to the formation of a ternary complex, is quantified via the cooperativity factor  $\alpha$ .<sup>8–11</sup> Adapting the cooperativity model to PPI drug discovery projects has been an underexplored opportunity, which has however started to gain more attention since PROTACs (proteolysis-targeting chimeras) were shown to induce cooperative binding of their protein binding partners.<sup>12–14</sup> In this work, using the interactions of 14-3-3 proteins as an example, we show that cooperativity analysis not only provides a description of binding properties but can also be used as a direct entry to establishing the selectivity of compound-stabilized PPIs.

The 14-3-3 family of proteins pertains to the adaptor proteins, which represent one of the most important protein classes in the PPI field, because they mediate interactions between proteins and thereby regulate the function of their partner proteins.<sup>15,16</sup> The 14-3-3 proteins recognize and bind

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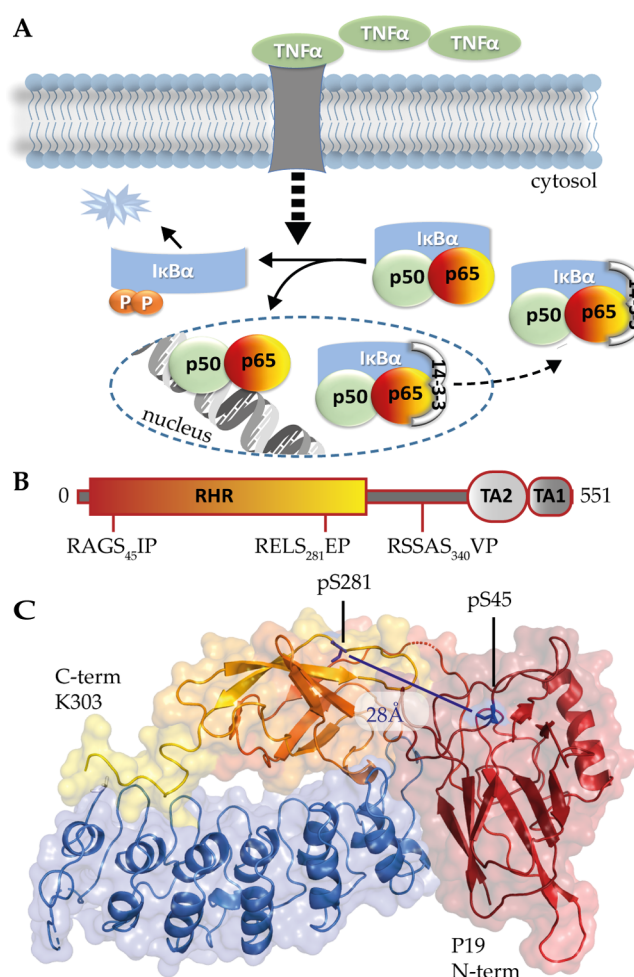
phosphoserine/threonine motifs of hundreds of protein partners in eukaryotic cells, and subsequently alter the catalytic activity, subcellular localization, or interactional preference of their partners.<sup>17,18</sup> 14-3-3 is functionally present as a w-shaped dimer featuring two highly conserved, amphipathic grooves, where the phosphorylated residues are bound.<sup>19</sup> A key element for the promising development of 14-3-3 proteins as potential drug targets is the availability of multiple members of a class of natural products, the fusicocanes (FCs), that have been demonstrated to stabilize the binding of 14-3-3 proteins to a number of partner proteins.<sup>20–24</sup> Protein crystallography studies have shown, for example, how such FCs stabilize 14-3-3 binding to protein targets such as p53,<sup>20</sup> c-Raf,<sup>21</sup> ER $\alpha$ ,<sup>22</sup> Gab2,<sup>23</sup> and CFTR.<sup>24</sup> These natural compounds bind to protein pockets delineated by the interface of the complex of 14-3-3 and its partner protein. In this way, FCs establish contacts to both protein partners simultaneously and act as a molecular glue.<sup>5,25</sup>

Among the 14-3-3 partners are at least two proteins that crucially participate in the NF- $\kappa$ B pathway.<sup>26–28</sup> The NF- $\kappa$ B signaling pathway has raised considerable attention as a therapeutic target, because of its intimate involvement in cell proliferation, apoptosis, immunity, and inflammation via the expression of several hundred genes.<sup>29–31</sup> NF- $\kappa$ B proteins are dimeric transcription factors, which are sequestered out of the nucleus into the cytosol by inhibitor proteins. Upon activation of the NF- $\kappa$ B pathway, for example, by TNF $\alpha$ , NF- $\kappa$ B translocates to the nucleus and activates transcription (Figure 1A). Inhibition of this transcriptional activity has been the goal of numerous drug discovery campaigns; however, only a limited number of inhibitors of the NF- $\kappa$ B pathway are currently marketed drugs, none of which directly target NF- $\kappa$ B.<sup>32–34</sup> The NF- $\kappa$ B subunit p65 and its inhibitor I $\kappa$ B $\alpha$  bind to 14-3-3, which favors localization of p65 in the cytosol.<sup>26</sup> Only limited information about the p65/14-3-3 interaction is available, even though downregulation of 14-3-3 leads to increased transcriptional activity of p65 in both a breast cancer model<sup>27</sup> and studies on Ischemia-Reperfusion.<sup>28</sup> Stabilization of the p65/14-3-3 complex might therefore inhibit the transcriptional activity of p65, opening a new and unique opportunity for p65-specific NF- $\kappa$ B inhibition.

In this study, we provide the structural and biophysical data and analysis to substantiate the biological interaction studies of the p65/14-3-3 interaction. We tested several natural and semisynthetic FCs as potential small-molecule stabilizers of this intriguing PPI, which revealed the semisynthetic DP-005 as the most active compound. A detailed analysis of the cooperative binding of the FCs, and DP-005 in particular, to the p65/14-3-3 interaction and multiple other disease relevant 14-3-3 PPIs was performed. DP-005 was at least 10-fold more active in stabilizing the p65/14-3-3 interaction than any other tested combination of FC and 14-3-3 PPI. The analysis clearly demonstrates that the cooperativity factor  $\alpha$  can be used to quantify these stabilizing effects and is an underlying factor in achieving selectivity of stabilizer molecules on PPIs.

## RESULTS AND DISCUSSION

**Interaction of p65 and 14-3-3.** In p65, the sequence surrounding three serines, S45, S281, and S340, matches to 14-3-3 consensus binding motifs (Figure 1B).<sup>19</sup> Mutations of these serine residues to alanines were previously shown to reduce p65 binding to 14-3-3 in cells and to increase the p65 concentration in the nucleus.<sup>26</sup> The S45 and S281 sites are



**Figure 1.** Interaction of p65 and 14-3-3. (A) Schematic representation of the interaction of 14-3-3, p65, and I $\kappa$ B $\alpha$  in the context of the NF- $\kappa$ B pathway. Briefly, upon activation of the pathway (for example, by TNF $\alpha$ ), a signal cascade (represented by a thick dashed arrow) leads to the phosphorylation and degradation of I $\kappa$ B $\alpha$  and the nuclear translocation of NF- $\kappa$ B (here represented by p50/p65). Binding of I $\kappa$ B $\alpha$  and 14-3-3 to p65 is necessary for nuclear export or cytosolic retention (narrow dashed arrow). (B) Domain representation of the p65 protein, with the Rel Homology Region (RHR), the two transactivator domains (TA1, TA2), and amino acid sequences of the three conserved potential 14-3-3 binding sites. (C) Cartoon representation of the crystal structure of the complex of I $\kappa$ B $\alpha$  with the RHR domain of p65, with van der Waals' transparent surface. p65 (red to yellow) with I $\kappa$ B $\alpha$  (blue) (PDB ID: 1IKN; p50 hidden for clarity; S45 and S281 are highlighted for clarity).

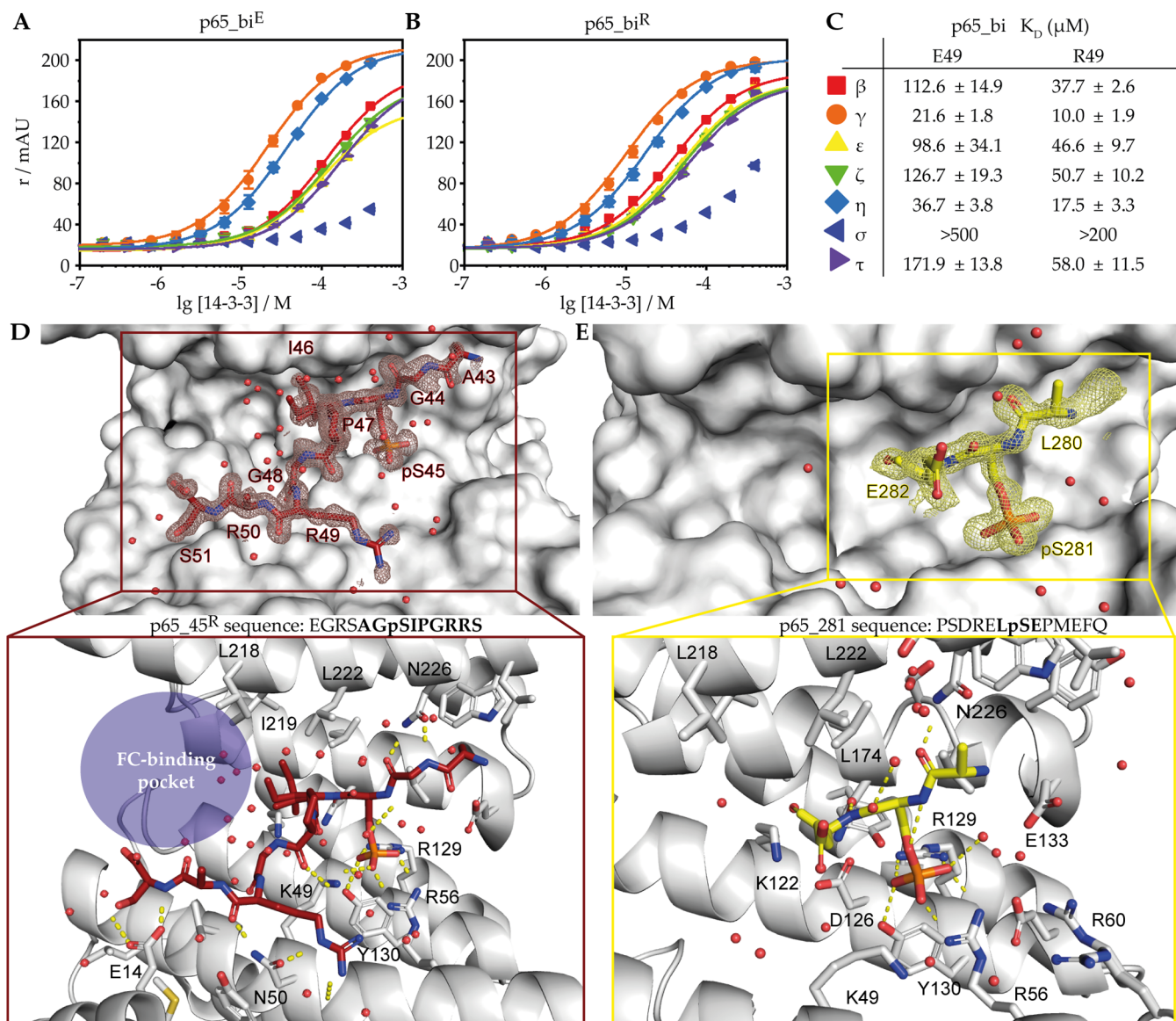
phosphorylated in response to TNF $\alpha$  treatment,<sup>35</sup> and both are highly conserved within mammals. No TNF $\alpha$ -dependent phosphorylation has been reported for the S340 site,<sup>35</sup> which is also less conserved (Figure S1). The pS45 and pS281 sites are located in unstructured loop regions of p65, and the folding of p65 brings these two residues into proximity with each other, which implies the possibility for a bivalent interaction with the 14-3-3 dimer (Figure 1C).<sup>24,36,37</sup>

To represent these interaction motifs, peptides of 13 amino acids were synthesized for the pS45 and pS281 sites (Table 1). Noteworthy, for the pS45 site, two different sequences are available: whereas the canonical sequence features a glutamate at position 49, a second sequence identified an arginine instead.<sup>38</sup> Because no biological or phenotypic relevance of



Table 1. Overview of Synthetic Peptides Representing the Proposed 14-3-3 Binding Motifs of p65

name	phosphosite	peptide sequence
p65_45 <sup>E</sup>	pS45	EGRSAGpS <sub>45</sub> IPGE <sub>49</sub> RS
p65_45 <sup>R</sup>	pS45	EGRSAGpS <sub>45</sub> IPGR <sub>49</sub> RS
p65_281	pS281	PSDRELpS <sub>281</sub> EPMEFQ
p65_bi <sup>E</sup>	pS45pS281	EGRSAGpS <sub>45</sub> IPGE <sub>49</sub> RSGSGGGSGPSDRELpS <sub>281</sub> EPMEFQ
p65_bi <sup>R</sup>	pS45pS281	EGRSAGpS <sub>45</sub> IPGR <sub>49</sub> RSGSGGGSGPSDRELpS <sub>281</sub> EPMEFQ



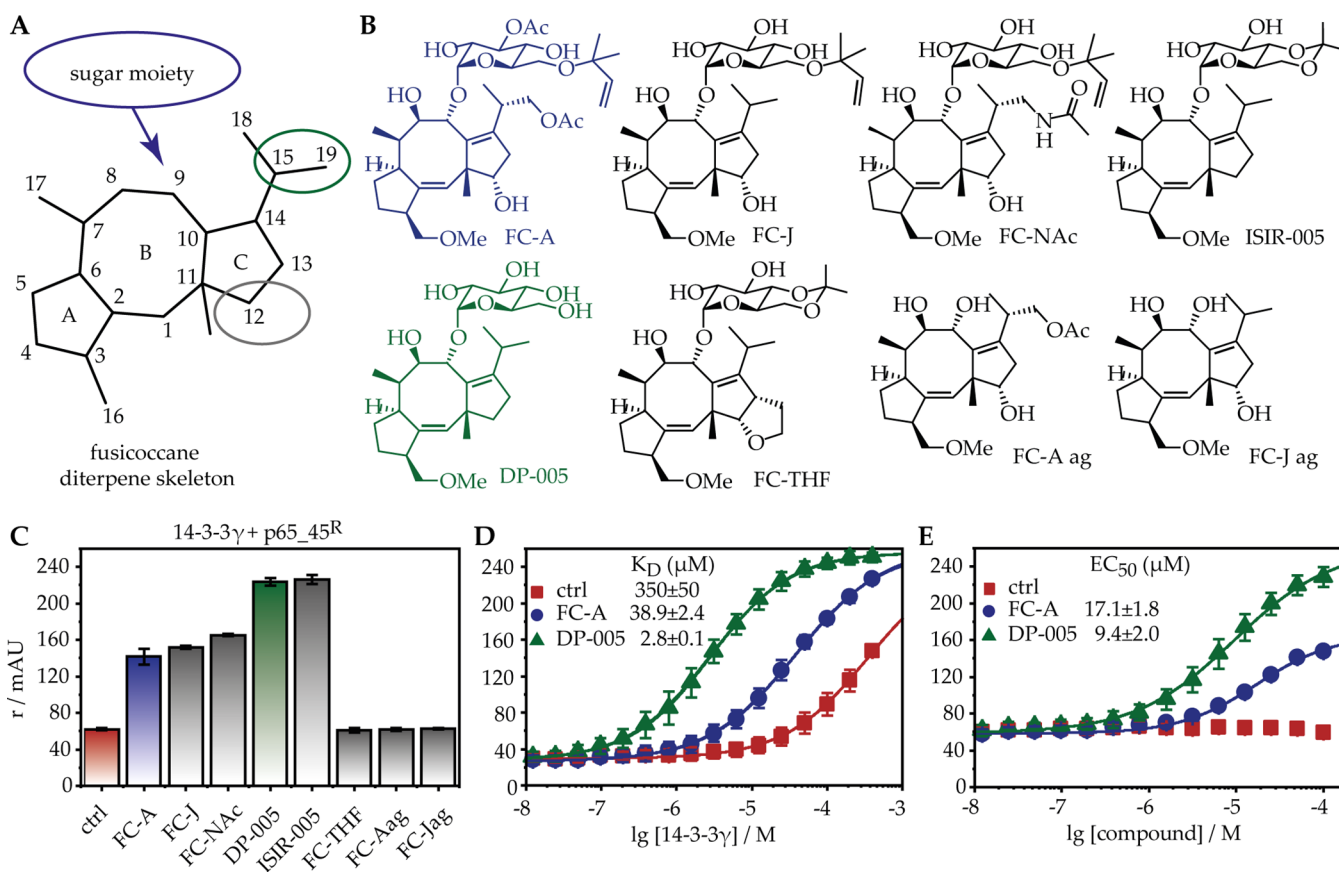
**Figure 2.** Binary complex of p65 peptides and 14-3-3. (A,B) The binding affinities of the indicated bivalent p65 peptides (100 nM) to all human 14-3-3 isoforms were measured via fluorescence anisotropy ( $r$  in mAU) assays. (C)  $K_D$  values in  $\mu$ M of the p65\_bi<sup>E</sup> and p65\_bi<sup>R</sup> to the different 14-3-3 isoforms as the mean  $\pm$  SD. (D) High-resolution X-ray crystal structures of p65\_45<sup>R</sup> (PDB ID: 6QHL). The peptide shows a C-terminal curved conformation, which leaves the typical FC binding pocket accessible (blue circle). (E) High-resolution X-ray crystal structures of p65\_281 (PDB ID: 6QHM) in complex with 14-3-3 $\Delta$ C. The proteins are displayed as either a white surface or a cartoon (close up); water molecules are shown as red spheres. The  $2mF_o - F_c$  electron density map is displayed with  $\sigma = 1$ . Polar contacts between protein and peptides are indicated as yellow dashed lines. Sequence of the p65\_45<sup>R</sup> peptide, EGRSAGpSIPGRRS; sequence of the p65\_281 peptide, PSDRELpSEPMEFQ (residues visible in the electron density are bold).

these variants is reported,<sup>38</sup> both sequences were tested for binding, hereafter termed p65\_45<sup>E</sup> and p65\_45<sup>R</sup> after the phosphorylation site S45 and the variant of amino acid 49. Each peptide is centered around the phosphorylated serine, flanked by six amino acids of the wild-type sequence on each side. To mimic the expected bivalent binding mode of the

p65/14-3-3 complex, double phosphorylated peptides were synthesized, which connected the two binding sites with a flexible linker.

**Elucidation of 14-3-3 Binding Motifs of p65.** In a first step, the binding affinities of the two conserved 14-3-3 binding motifs of p65 were measured. To this end, the p65-peptides





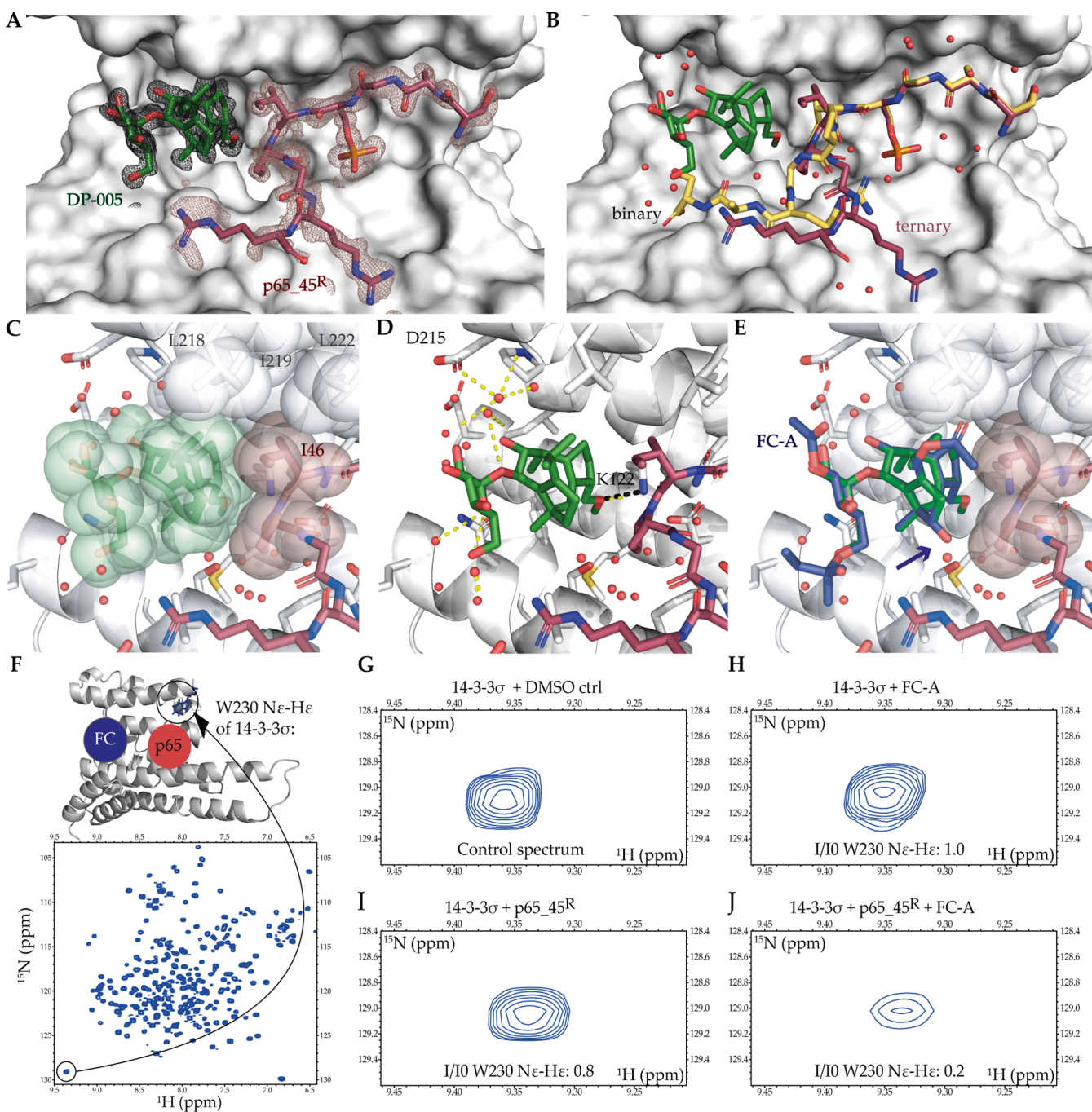
**Figure 3.** Stabilizing effect of fusicocanes on the p65<sub>45</sub>/14-3-3 interaction. (A) The diterpene skeleton of the fusicocane family. Encircled are the positions that are of interest in this study. (B) A collection of eight FCs used to screen for a stabilizing effect on the p65/14-3-3 complex. (C) FA-based screening results of the FC-collection shown in (B) on the p65<sub>45</sub><sup>R</sup>/14-3-3γ complex. The anisotropy (*r* in mAU) measurements were carried out with 100 μM compound, 50 μM 14-3-3γ, and 100 nM of p65<sub>45</sub><sup>R</sup>. Values represent the mean ± SD of two independent measurements performed in technical triplicates. (D) Titration of 14-3-3γ in the presence of 100 μM compound or DMSO control (ctrl) and 100 nM p65<sub>45</sub><sup>R</sup> measured with FA. Values and error bars represent the mean ± SD of three independent singlet measurements. (E) Titration of compound or DMSO control (ctrl) in the presence of 50 μM 14-3-3γ and 100 nM p65<sub>45</sub><sup>R</sup> measured with FA. Values and error bars represent the mean ± SD of three independent singlet measurements.

were labeled with a fluorophore, and fluorescence anisotropy assays (FA) were performed. Both single site binding epitopes showed an increase in FA upon addition of 14-3-3 proteins, although not with full binding saturation (Figure S2). This confirms weak binding of both sites to all human 14-3-3 isoforms. The bivalent peptides featured a significantly stronger binding affinity to all 14-3-3 isoforms, with the p65<sub>bi</sub><sup>R</sup> binding roughly 2-fold stronger than the p65<sub>bi</sub><sup>E</sup> peptide (Figure 2A–C). The strongest binding was observed with 14-3-3γ ( $K_{E49}^D = 21.6 \pm 1.8 \mu\text{M}$ ;  $K_{R49}^D = 10.0 \pm 1.9 \mu\text{M}$ ), directly followed by 14-3-3η ( $K_{E49}^D = 36.7 \pm 3.8 \mu\text{M}$ ;  $K_{R49}^D = 17.5 \pm 3.3 \mu\text{M}$ ). The dissociation constants of 14-3-3β, ε, ζ, and τ with p65<sub>bi</sub><sup>R</sup> ranged from 38 to 127 μM, while 14-3-3σ showed the weakest binding (Figure 2C). The range of binding affinities for the 14-3-3 isoforms is not atypical and seems to be independent of the peptide sequence.<sup>24,39,40</sup> The peptide–protein binding data thus confirm a physical interaction between p65 and 14-3-3 and argue in favor of a bivalent binding event.

The binding between p65 and 14-3-3 was further elucidated with structural data; cocrystallization of the single phosphorylated peptides with 14-3-3σΔC (last 17 unstructured residues on the C-terminus are truncated) resulted in crystals with the highest resolution of 1.2 and 1.25 Å for the p65<sub>45</sub><sup>R</sup> and

p65<sub>281</sub> complexes, respectively (Table S1). The phosphorylated S45 makes polar contacts with R56, R129, and Y130, the general phospho-accepting pocket of 14-3-3 (Figure 2D; PDB-ID: 6QHL). Upstream of pS45 only A43 and G44 are visible in the electron density map, whereby the backbone of G44 makes polar contacts with N226 of the 14-3-3 protein. At the +1 position of pS45, the isoleucine points into a hydrophobic pocket of the 14-3-3 binding groove to make hydrophobic contacts with L174, I219, and L222. The presence of a proline residue at the +2 position results in a curved conformation in the binding groove, which potentially creates a ligandable interface pocket (Figure 2D). In comparable crystal structures of 14-3-3 with other interaction partners, a similar pocket accommodates FCs and their derivatives.<sup>20–24</sup> The side chain of R49 makes two polar contacts with N50 of 14-3-3 as well as with several water molecules. The C-terminus of the p65<sub>45</sub><sup>R</sup> peptide makes additional polar contacts with E14 of 14-3-3. The direct comparison of the two variants of the p65<sub>45</sub> peptide only shows small differences in binding affinity, and structural information could only be obtained for the p65<sub>45</sub><sup>R</sup> peptide; hence, this variant was selected for the next experiments.

The electron density of the p65<sub>281</sub> peptide in complex with 14-3-3 revealed only the backbone of two additional



**Figure 4.** Structural analysis of the ternary DP-005/p65/14-3-3 and FC-A/p65/14-3-3 complexes. (A) Crystal structure of DP-005 (green sticks), p65<sub>45R</sub> (red sticks), and 14-3-3 $\Delta$ C (white surface) (PDB ID: 6NV2). (B) Binding of DP-005 induces a reorientation of the p65<sub>45R</sub> peptide (original conformation of binary complex, yellow sticks; ternary complex with DP-005, red sticks). (C) Hydrophobic contacts are indicated with spheres. (D) Polar contacts of DP-005 displayed as yellow dashed lines with the polar contact of the methoxy group of DP-005 and K122 highlighted with black dashed lines. (E) Overlay of FC-A (blue sticks) and DP-005 (green sticks). An arrow indicates the hydroxyl group at position 12, which causes the difference in affinity between both compounds. Hydrophobic residues of the peptide/protein complex are shown as spheres. (F) The resonance corresponding to the W230 N $\epsilon$ -H $\epsilon$  bond of 14-3-3 $\sigma$  was monitored to detect the stabilization of this PPI. This resonance is circled in black on the  $^1\text{H}$ - $^{15}\text{N}$  TROSY-HSQC spectrum of  $^{15}\text{N}^{13}\text{C}^2\text{H}$ -labeled 14-3-3 $\sigma$  (shown on the right), and the corresponding residue W230 is represented as sticks, colored in blue and circled in black on the crystal structure (shown on the left, represented as a white surface). This residue is close to the peptide-binding site and distant from the FC-A binding site. (G–J) The enlarged spectral region of the  $^1\text{H}$ - $^{15}\text{N}$  TROSY-HSQC containing the resonance corresponding to the W230 N $\epsilon$ -H $\epsilon$  bond of 14-3-3 $\sigma$  (125  $\mu\text{M}$ ) is shown in the presence of DMSO 4% (v/v), present in all samples (G), FC-A 125  $\mu\text{M}$  (H), p65<sub>45R</sub> peptide 625  $\mu\text{M}$  (I), and p65<sub>45R</sub> peptide 625  $\mu\text{M}$  and FC-A 125  $\mu\text{M}$  (J).

amino acids besides that of the phosphorylated serine (Figure 2E; PDB ID: 6QHM). Additional electron density on the N-terminus of the peptide could not be fully interpreted, which indicated multiple conformations of these amino acids. The

phosphoserine is trapped in the binding groove via polar contacts with R59, R129, and Y130 of 14-3-3 $\Delta$ C. Additionally, contacts could also be observed with the backbone nitrogen of E282, at the +1 position of p65<sub>pS281</sub>, and N175



of 14-3-3 and the carbonyl of L280 of the peptide and N226 of the protein.

**Fusicocanes Stabilize the p65/14-3-3 Interaction.** The structures of p65<sub>45</sub><sup>R</sup> and p65<sub>281</sub> provide the structural basis to investigate possible molecular strategies to stabilize these binary complexes. Because the 14-3-3 binding groove is highly conserved, the p65-peptide/14-3-3 $\sigma\Delta$ C interface displays the key molecular details of the primary p65/14-3-3 interface.<sup>19</sup> The p65<sub>45</sub><sup>R</sup> peptide is bending in a way that the typical FC binding pocket remains accessible and the conformation of the p65<sub>281</sub> is unknown (Figure 2D,E). This leaves the possibility that both complexes may be stabilized by FCs or synthetic derivatives thereof. The FCs share a diterpene core with a 5(A)-8(B)-5(C) ring structure (Figure 3A),<sup>41</sup> modifications that include additional sugar moieties, hydroxylation, or acetylation have introduced considerable chemical diversity into this compound family (Figure 3B).<sup>42</sup>

The effect of FCs on the p65/14-3-3 complex was investigated with a small collection of eight natural FCs and their semisynthetic derivatives (Figure 3B). The tested FCs have a FC-A-like structure with a hydroxylation at position 8, a methoxy group at position 16, and a sugar moiety at position 9. The main variations within the collection used here are on the sugar moiety, position 12 and/or 19. FC-A contains an additional acetyl on the 19 position, while the natural product FC-J only has an isopropyl on the C-ring. A recent study showed that replacing the acetyl with an acetamide (FC-NAc) improves the affinity to 14-3-3 with various partners.<sup>25</sup> The 12-position is hydroxylated in the natural compounds, but this position is not hydroxylated in the semisynthetic derivatives DP-005 and ISIR-005,<sup>23,43</sup> and a bulky group was introduced for FC-THF.<sup>44</sup> For FC-A-aglycon (FC-A-ag)<sup>44</sup> and FC-J-aglycon (FC-J-ag),<sup>44</sup> the sugar moiety was removed. Initially, a single dose of compound and the protein concentration were tested using FA. The single phosphorylated peptides were used so that a potential stabilizing effect could be correlated to a specific binding motif. 14-3-3 $\gamma$  was used because of the highest binding affinity of all peptides to this isoform.

The FA results showed that several FCs have a stabilizing effect on the p65<sub>45</sub><sup>R</sup>/14-3-3 $\gamma$  and p65<sub>45</sub><sup>R</sup>/14-3-3 $\gamma$  interaction, while they do not stabilize the p65<sub>281</sub>/14-3-3 $\gamma$  complex (Figure 3C and Figure S3). The fusicocanes FC-A, FC-J, and FC-NAc elicited about a 2-fold higher anisotropy signal of the p65<sub>45</sub><sup>R</sup>/14-3-3 $\gamma$  PPI as compared to the DMSO control. Modeling of cooperative complex stabilization of FC/14-3-3 PPIs indicated an intrinsic affinity of 300–500  $\mu$ M for FC-A and FC-J and about 10  $\mu$ M for FC-NAc. The higher affinity of FC-NAc to the apo14-3-3 does not translate into significantly higher stabilization of the p65<sub>45</sub><sup>R</sup>/14-3-3 complex.<sup>11</sup> These data show that the FC-induced stabilization is not directly connected to the intrinsic affinity of the FCs to 14-3-3. Interestingly, semisynthetic analogues DP-005 and ISIR-005 induced a stronger increase in the anisotropy signal (about 3-fold). Bulky extensions at the C-ring (FC-THF) and the absence of the sugar moiety (FC-A-ag and FC-J-ag) abrogated the ability to stabilize the p65<sub>45</sub><sup>R</sup>/14-3-3 $\gamma$  complex. A hydroxyl group at the 12-position or its ring-extension thus causes a loss in activity. The negligible activity of the aglycons is in line with previous work on these synthetic variants, which generally show lower activity.<sup>44</sup>

The stabilizing effects of FC-A and DP-005 were further investigated via 14-3-3 $\gamma$  protein titrations to the p65<sub>45</sub><sup>R</sup>

peptide at a set concentration of DP-005, FC-A, or the DMSO control (Figure 3D). The protein titrations confirmed the stronger stabilizing effect of DP-005 as compared to FC-A, with a clear left shift of the binding curve, to afford apparent  $K_D$  values of  $2.8 \pm 0.1$  and  $38.9 \pm 2.4$   $\mu$ M, respectively. Constraining the fitting for the DMSO control, while using the upper plateau of the DP-005 and FC-A binding curves, results in an apparent  $K_D$  of about 350  $\mu$ M for the p65<sub>45</sub><sup>R</sup>/14-3-3 $\gamma$  complex.

Hence, the improvement in peptide stabilization acquaints to approximately 10-fold (FC-A) and 100-fold (DP-005) as compared to the DMSO control. Finally, the effective concentrations of FC-A and DP-005 were determined by titrating the compounds to a fixed protein and peptide concentration of 50 and 0.1  $\mu$ M, respectively. This gave an  $EC_{50}$  value of  $9.4 \pm 2.0$   $\mu$ M for the semisynthetic compound DP-005, while the natural compound FC-A showed a weaker activity with an  $EC_{50}$  value of  $17.1 \pm 1.8$   $\mu$ M (Figure 3E).

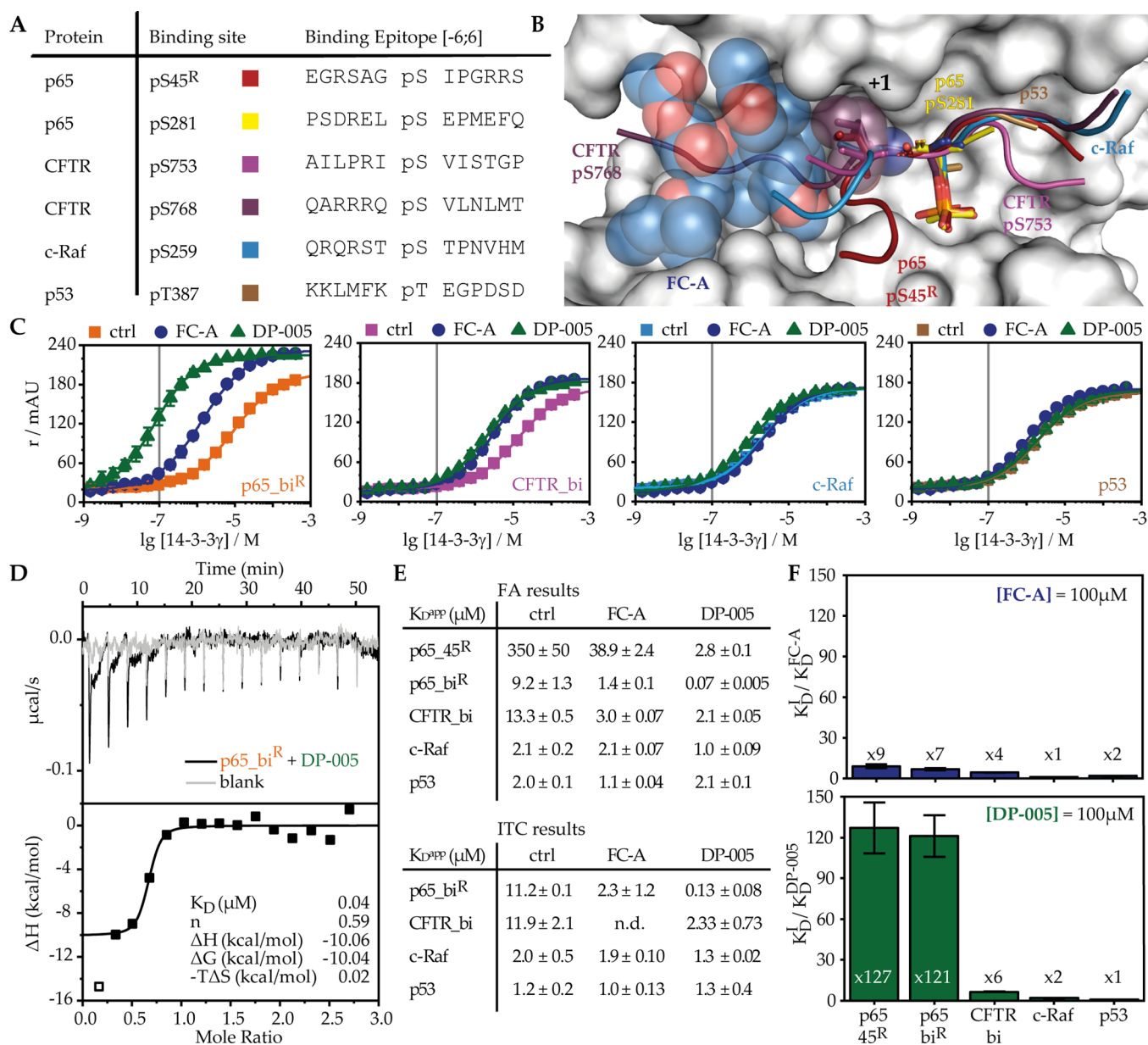
### Crystallography and NMR Provide Structural Confirmation of the FC Binding to p65/14-3-3.

To corroborate the stabilization and SAR obtained using FA, we sought a structural analysis of the ternary complex of DP-005 with p65<sub>45</sub><sup>R</sup>/14-3-3. For this purpose, DP-005 was soaked into crystals of the preformed p65<sub>45</sub><sup>R</sup>/14-3-3 $\sigma\Delta$ C complexes, to reveal extra electron density for the p65<sub>45</sub><sup>R</sup>/14-3-3 $\sigma\Delta$ C crystals. This electron density allowed for the complete refinement of DP-005 (Figure 4A, Table S1, PDB ID: 6NV2). The crystal structure provided detailed information about the orientation of the diterpenoid core and the sugar moiety. Of note, the orientation of the peptide changed significantly upon binding of DP-005 (Figure 4B). In particular, amino acid P47 changed from a *trans*- to a *cis*-conformation, and the residues from G48 to S51 moved away from the ligand. The most prominent interactions between DP-005 and the protein complex are the hydrophobic contacts with I46 and P47 of p65<sub>45</sub><sup>R</sup> as well as with L218, I219, and L222 of 14-3-3 $\sigma\Delta$ C (Figure 4C). Noteworthy is the contact of the methoxy group of DP-005 with the  $\epsilon$ -amino group of K122 of the protein, while the sugar moiety was surrounded by the water shell of 14-3-3 $\sigma\Delta$ C (Figure 4D). The fact that FC-aglycons showed only a negligible activity might be due to the importance of this sugar–water shell interaction.

An overlay of FC-A with the conformation of DP-005 points out the impact of the singular hydroxyl group at the 12-position of FC-A, not present in DP-005, on the binding to the protein complex (Figure 4E). The hydroxyl group would have an orientation that would induce a steric and polar clash with hydrophobic elements of the peptide. This indicates that the peptide would potentially require further rearrangements to accommodate the binding of FC-A, which potentially explains why the ternary structure of FC-A/p65<sub>45</sub><sup>R</sup>/14-3-3 $\sigma\Delta$ C could not be determined with X-ray crystallography.

As an alternative to obtaining structural information on the binding of FC-A to the binary p65/14-3-3 complex, NMR measurements were performed, on the basis of the assignments of 14-3-3 $\sigma$  signals.<sup>45</sup> Chemical shift perturbations in the resonances corresponding to specific residues along 14-3-3 $\sigma$  confirmed that FC-A binds in its previously reported binding pocket, in the presence and in the absence of the p65<sub>45</sub> peptide (Figure S4). To specifically monitor the binding event of the p65<sub>45</sub><sup>R</sup> to 14-3-3 $\sigma$ , we focused on the <sup>1</sup>H–<sup>15</sup>N TROSY-HSQC signature of the residue W230 side chain of <sup>15</sup>N<sup>13</sup>C<sup>2</sup>H labeled 14-3-3 $\sigma$ . This residue is specifically affected



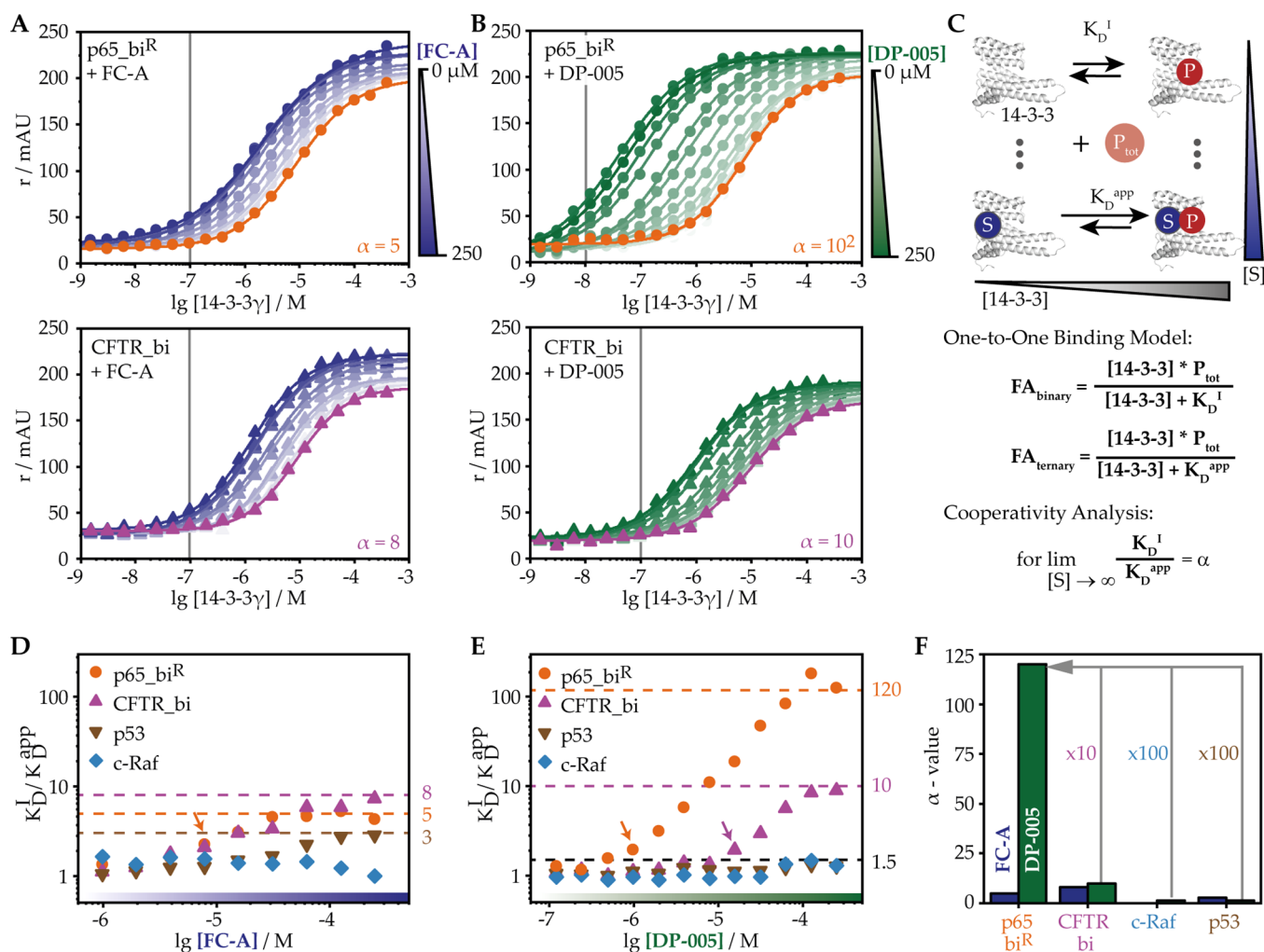


**Figure 5.** Comparison of FC-induced stabilization on 14-3-3 PPIs. (A) Overview of 14-3-3 binding epitopes. (B) Overlay of the crystal structures of the binding epitopes shown in (A) (cartoon representation, phosphorylated residue, and +1 amino acid shown as sticks). FC-A is shown as transparent spheres, while 14-3-3 is shown as white van der Waals surface. (C) Binding affinities of indicated peptides were measured for 14-3-3 $\gamma$  in the presence of DMSO as control (ctrl), 100  $\mu\text{M}$  FC-A, or 100  $\mu\text{M}$  DP-005 with FA ( $r$  in mAU). For peptide sequences, see Tables 1 and S3. (D) Apparent binding affinity of DP-005/p65<sub>bi</sub><sup>R</sup>/14-3-3 $\gamma$  complex formation measured with ITC ( $n = 1$ ). For the duplicate, see Figure S7C. Syringe, 150  $\mu\text{M}$  14-3-3 $\gamma$ , 200  $\mu\text{M}$  DP-005; cell, 10  $\mu\text{M}$  p65<sub>bi</sub><sup>R</sup>, 200  $\mu\text{M}$  DP-005. (E)  $K_D^{\text{DPP}}$  values in  $\mu\text{M}$  for the indicated peptides for the DMSO control (ctrl) or with FC-A or DP-005 as mean  $\pm$  SD measured with FA (upper panel,  $n = 3$ ) and ITC (lower panel,  $n = 2$ ). n.d. = not determined. (F) Increase in affinity of the binding partners due to FC-A or DP-005 shown as the ratio of the dissociation constant of the DMSO control ( $K_D^{\text{D}}$ ) divided by the dissociation constant in the presence of either FC-A ( $K_D^{\text{FC-A}}$ ) or DP-005 ( $K_D^{\text{DP-005}}$ ) based on FA results. The numbers indicate the factor with which the binding affinity is enhanced due to the FCs as compared to the control.

by the p65<sub>45</sub><sup>R</sup> peptide binding, but not by the FC-A binding, as can be expected from its remote position from the FC-A binding pocket (Figure 4F). Its side-chain  $\text{N}\epsilon\text{--H}\epsilon$  resonance has a clear signature because it is isolated in the spectrum (Figure 4F) and is unambiguously assigned (Figure S5). Monitoring the resonance intensity of  $\text{N}\epsilon\text{--H}\epsilon$  W230 revealed that the intensity of this resonance is, as expected, not affected by the presence of FC-A alone (Figure 4G,H). Addition of the p65<sub>45</sub><sup>R</sup> peptide alone results in resonance broadening due to the binding and an intensity drop to 80% as compared to the

reference spectrum (Figure 4I). Addition of FC-A together with the p65<sub>45</sub><sup>R</sup> peptide resulted in the most pronounced decrease of the intensity of the  $\text{N}\epsilon\text{--H}\epsilon$  W230 resonance, down to 20% (Figure 4J). The data thus orthogonally confirm the stabilization of the p65<sub>45</sub><sup>R</sup>/14-3-3 $\sigma$  complex by FC-A.

**Preferential Stabilization of the p65/14-3-3 Complex by DP-005.** The stabilization of p65/14-3-3 by DP-005 is significantly stronger than the effect of FC-A (vide supra), which raised the question about DP-005's impact on other FC responsive 14-3-3 PPIs. Crucial differences between FC-A and



**Figure 6.** Cooperativity analysis of FC-A and DP-005 on 14-3-3 PPIs. (A) 2D FA-results of FC-A with p65<sub>bi</sub><sup>R</sup>/14-3-3 $\gamma$  and CFTR<sub>bi</sub> ( $n = 1$ ). FC-A concentrations range from 0 to 250  $\mu$ M; the vertical gray line indicates the peptide concentration. Anisotropy ( $r$ ) given in mAU. (B) 2D FA-results of DP-005 with p65<sub>bi</sub><sup>R</sup>/14-3-3 $\gamma$  or CFTR<sub>bi</sub> ( $n = 1$ ). DP-005 concentrations range from 0 to 250  $\mu$ M; the vertical gray line indicates the peptide concentration. (C) Scheme of 2D titration in FA. 14-3-3 was titrated against 100 nM of peptide (P) varying fixed concentrations of stabilizer (S). The data analysis was based on a one-to-one binding model. For cooperativity analysis, the ratio of  $K_D$  of the binary ( $K_D^I$ ) and ternary ( $K_D^{app}$ ) complex was used to derive the cooperativity factor  $\alpha$  (for derivation of the equations and additional information, see the Supporting Information). (D)  $K_D^I/K_D^{app}$  ratio plotted against FC-A concentration for 2D titrations shown in (A) and Figure S9. The arrow indicates the minimal active concentration of the stabilizer, while the curve reaches saturation at the  $\alpha$ -value. (E)  $K_D^I/K_D^{app}$  ratio plotted against DP-005 concentration for 2D titrations shown in (B) and Figure S10. Analysis is in accordance with (D). (F) The cooperativity value  $\alpha$  for the ternary complexes with 14-3-3 $\gamma$  and either FC-A or DP-005 was plotted against the different target peptides. DP-005 has a 10-fold stronger effect on the p65/14-3-3 $\gamma$  interaction than does any other measured interaction.

DP-005 are the additional polar decorations of FC-A at positions 12 and 19 (Figure 3A,B). Hence, we hypothesized, on the basis of the crystal structure of the ternary complex, that the hydrophobic contacts of the Ile and Pro at the +1 and +2 positions of the p65<sub>45</sub><sup>R</sup> peptide would be complementary to the hydrophobic isopropyl of DP-005 and thus are a crucial driving force for DP-005-based stabilization. Diverse amino acid sequences can be found C-terminal from the phosphorylation site within FC responsive 14-3-3 PPIs.<sup>20–24</sup> We investigated the effects of FC-A and DP-005 on a small set of three different clinically relevant 14-3-3 interaction partners (CFTR,<sup>46,47</sup> c-Raf,<sup>37,48</sup> and p53<sup>49,50</sup>). These interaction partners differ in size and hydrophobicity of the +1 amino acid (Figure 5A). Apparent binding affinities were determined using FA and isothermal titration calorimetry (ITC) (Figures 5, S6, and S7).

The example of CFTR is particularly interesting because the binding behavior of the CFTR peptides shows similarities with the p65 peptides: for both interactions, a bivalent binding mode is suggested, whereby only one of the two binding sites is responsive to FCs (Figure S6A,B),<sup>24</sup> and both offer a hydrophobic amino acid at the +1 position (Figure 5A). Therefore, the bivalent p65<sub>bi</sub><sup>R</sup> and CFTR<sub>bi</sub> peptides were used to measure the effects of FC-A and DP-005 on complex formation with 14-3-3 $\gamma$  (Figure 5C–F). FC-A decreased the apparent dissociation constants of both complexes by a factor of 4–7 in FA. ITC measurements confirmed the FC-A-based stabilization of the p65<sub>bi</sub><sup>R</sup>/14-3-3 $\gamma$  complex, while FC-A induced a loss of heat signal for the FC-A/CFTR<sub>bi</sub>/14-3-3 $\gamma$  complex (Figure S7A–F). In the presence of DP-005, the apparent binding affinities shifted to about 2  $\mu$ M for the CFTR<sub>bi</sub>/14-3-3 $\gamma$  complex (6-fold stabilization) and 0.1  $\mu$ M for the p65<sub>bi</sub><sup>R</sup>/14-3-3 $\gamma$  complex (120-fold stabilization) with

FA and ITC (Figures 5 C–F and S7M,N). Overall, DP-005 has a 10× stronger stabilizing effect than does FC-A on the p65<sub>bi</sub><sup>R</sup>/14-3-3 $\gamma$  complex.

The preferential stabilization of the p65/14-3-3 $\gamma$  complex by DP-005 is remarkable, because the CFTR<sub>753</sub> peptide offers with a valine instead of an isoleucine at the +1 position a hydrophobic environment similar to that of the p65<sub>45</sub><sup>R</sup> peptide. However, the isoleucine of CFTR<sub>753</sub> at the +2 position is pointing toward the phosphorylated residue due to the alternating orientations of the amino acid side chains of unstructured motifs (Figure S6). In this orientation, the Ile is not able to establish favorable hydrophobic contacts with DP-005, unlike the proline at the +2 position of the p65<sub>45</sub><sup>R</sup> peptide (Figure 4C). Therefore, it can be suggested that the difference of the +2 position of the peptide sequences is crucial for the enhanced potency of DP-005 on the p65/14-3-3 $\gamma$  complex.

For c-Raf, two 14-3-3 binding sites are reported, pS233 and pS259, which both contain a more polar threonine at the +1 and a proline at the +2 positions (Figure 5A).<sup>21</sup> This interaction is not stabilized by FC-A;<sup>11</sup> therefore, it was interesting to analyze the stabilization achievable with DP-005 on the c-Raf<sub>259</sub>/14-3-3 complex. In the presence of DP-005, the c-Raf<sub>259</sub> peptide binds 2× stronger to 14-3-3 $\gamma$ , while the lack of stabilization for FC-A could be reproduced (Figure 5C–F). Besides the small effects, the observed stabilization was consistent in FA and ITC (Figures 5E and S7M,N). This shows that a small polar amino acid at the +1 position reduces the stabilizing effect of DP-005, which confirms that mostly hydrophobic contacts between this FC derivative, the peptide, and 14-3-3 are necessary to establish the cooperative binding.

p53 binds to 14-3-3 via its C-terminal domain, and one of multiple possible phosphorylated binding sites, the pT387 site, was cocrystallized with 14-3-3.<sup>49–51</sup> The binding sequence contains a polar glutamic acid at the +1 position, and the peptide is bound in a bent conformation within the 14-3-3 binding groove, which allows FCs to bind (Figure 5A,B).<sup>20</sup> However, FC-A shows only a stabilization of the p53/14-3-3 interaction by reducing the apparent  $K_D$  by a factor of 2 in FA, which could not be confirmed by ITC. DP-005 has no significant effect on the p53/14-3-3 complex (Figures 5C–F and S7M,N). DP-005 thus has a specific and strong stabilization effect on the p65 sequence exclusively.

**Cooperativity Factor as a Measure of Concentration-Independent Stabilization.** The relative effect of a set FC concentration on the examined 14-3-3 PPIs revealed major differences in stabilization potency. These measurements were performed at a single, fairly high concentration of FCs and give no information about the effective range of FC concentrations, the concentrations needed to reach saturation of the system, or the low threshold concentration necessary to achieve stabilization. Cooperativity analysis was done to gain a deeper understanding of the relevant concentrations and to quantify the preferential stabilization of p65 by DP-005, over other targets via the concentration-independent cooperativity factor  $\alpha$  (Figure S8 and Table S2). The  $\alpha$ -factor describes the enhancement of binding affinity of a ternary complex formation as compared to a binary complex.<sup>9,11,12</sup> To calculate this factor, 14-3-3 $\gamma$  titrations were performed in the presence of a variety of constant concentrations of DP-005 or FC-A stabilizer on the p65, CFTR, c-Raf, and p53/14-3-3 interactions (Figures 6A,B, S9, and S10). The highest FC concentration used was 250  $\mu$ M, which was reduced stepwise

in a 1:1 dilution series. At increasing concentrations of compound, the protein has an increasing partial occupancy of bound stabilizer during the titrations. Because of cooperative binding, this leads to more peptide binding at lower protein concentrations, and hence a left shift of the binding curves that resulted in a 2D stabilizing profile.<sup>11</sup> A binding event of FCs to the p65<sub>bi</sub><sup>R</sup> peptide in the absence of protein could not be detected (Figure S8C,D).

For FC-A the apparent  $K_D$ 's of the p65<sub>bi</sub><sup>R</sup> and CFTR<sub>bi</sub> in complex with 14-3-3 $\gamma$  shift an order of magnitude from about 10 to 1  $\mu$ M for both peptides (Figure 6A). Increasing concentrations of FC-A also induce a shift in the upper plateau, most likely due to a reduced flexibility of the fluorescently labeled N-terminus of the peptide. This could be caused by a reduced flexibility of the “roof” of 14-3-3 comprised of the C-terminal helix of 14-3-3 upon binding of FCs (see its interactions with D215 and Ile218, Figure 3C,D).<sup>25</sup> Unlike FC-A, DP-005 induces about a 100-fold decrease of apparent  $K_D$ -values for the p65<sub>bi</sub><sup>R</sup>/14-3-3 $\gamma$  complex, while the curves for the DP-005/CFTR<sub>bi</sub>/14-3-3 $\gamma$  complexes are comparable to the curves for FC-A with a shift of only 1 order of magnitude (Figure 6B). DP-005 also causes an increase of anisotropy values for the upper plateau. These data were fitted using the Hill equation, and the resulting apparent  $K_D$ 's in the presence of the FCs ( $K_D^{\text{app}}$ ) were divided by the  $K_D$  of the binary complex ( $K_D^1$ ). It is expected that at a certain stabilizer concentration the system is saturated, which resulted in no further reduction of apparent  $K_D$ -values with increasing concentrations of stabilizer. This saturated system can be used to extrapolate the cooperativity factor  $\alpha$  (Figure 6C and Table S2). This value represents the maximum reduction of  $K_D$  induced via the stabilizer and can be derived from the ratio of the dissociation constant of the binary complex and the apparent  $K_D$  of the complex saturated with stabilizer (for more information, see the Supporting Information). By plotting the  $K_D^1/K_D^{\text{app}}$  ratio versus the FC concentration, not only the  $\alpha$ -factor but also a threshold concentration can be extracted (Figure 6D,E). This concentration is the minimal required FC concentration needed to induce cooperative formation of the ternary complex. The threshold concentration of FC-A was 10  $\mu$ M for both the p65<sub>bi</sub><sup>R</sup> and the CFTR<sub>bi</sub> (Figure 6D,E). The FC-A/p65/14-3-3 $\gamma$  complex reached saturation with 30  $\mu$ M FC-A, as higher concentrations do not change the ratio of  $K_D^1/K_D^{\text{app}}$  anymore (Figure 6D). At the point of saturation, the  $K_D^1/K_D^{\text{app}}$  gives an  $\alpha$ -value of 5 for the FC-A/p65/14-3-3 $\gamma$ , while for the FC-A/CFTR<sub>bi</sub>/14-3-3 $\gamma$  an  $\alpha$ -value of 8 is reached. An  $\alpha$ -value of 10 was determined for the DP-005/CFTR<sub>bi</sub>/14-3-3 $\gamma$  complex, also with the threshold concentration of 10  $\mu$ M to induce cooperative binding. In contrast, only 1  $\mu$ M of DP-005 is needed to induce cooperativity of the DP-005/p65/14-3-3 $\gamma$  complex with an  $\alpha$ -value of 120, at least 10× higher than any other measured  $\alpha$ -value. DP-005 revealed only a small enhancement of apparent  $K_D$  values for c-Raf and p53, even at the highest concentrations. The higher concentrations of FC-A led to a further decrease of  $K_D^{\text{app}}$  and resulted in an  $\alpha$ -value of 3 for the FC-A/p53/14-3-3 $\gamma$  complex. In contrast, the destabilizing effect of FC-A on the c-Raf/14-3-3 $\gamma$  complex becomes more obvious at higher concentrations, while the higher concentrations of DP-005 did not further decrease the  $K_D^{\text{app}}$ . An  $\alpha$ -value of about 1.5 was determined for the DP-005/c-Raf/14-3-3 $\gamma$  and the DP-005/p53/14-3-3 $\gamma$  complex.



## CONCLUSIONS

Cooperativity analysis has been shown to be a powerful tool to explain and compare the efficacy of PPI stabilizers and to optimize their affinity.<sup>11,12</sup> In this Article, the p65/14-3-3 interaction was structurally elucidated to provide fundamental molecular details of this interaction. These are urgently required, particularly to provide novel molecular entries for translational chemical biology targeting the NF- $\kappa$ B pathway. The biophysical characterization of the p65/14-3-3 complex was subsequently utilized for natural compound-based PPI stabilization studies and for the development of conceptual insights in the underlying cooperative mechanism. This has provided clear molecular directions to molecular optimization of both affinity and selectivity parameters of PPI stabilization drug development.

Natural compounds provide unique tools to probe and interrogate unique regions of chemical space,<sup>52</sup> which can be extended and brought to higher levels of control and specificity through their chemical modifications.<sup>25</sup> In the field of PPI stabilization, natural compounds are of unique importance because the first identified stabilizer molecules in clinical use belong to this compound class, for example, rapamycin or FK506.<sup>5–7</sup> A number of 14-3-3-based PPIs are stabilized by FCs with correlated positive effects on the underlying biological systems, such as tumor growth reduction by Cotylenin A<sup>21</sup> or improved neural regeneration with FC-A.<sup>53,54</sup> While DP-005 is generally a weaker stabilizer than FC-A for most 14-3-3 PPIs, here we revealed that, in contrast, DP-005 has the strongest stabilizing capability of all FCs for the p65/14-3-3 interaction, being 10-fold more potent than FC-A. The structural elucidation of the DP-005 binding mode identified the key elements required for highly cooperative binding, including specific hydrophobic contacts with the p65 partner epitope and 14-3-3 K122 as a polar anchor.

The high cooperativity of DP-005 for stabilizing the p65/14-3-3 interaction highlights that a relatively small modification to a small-molecule PPI stabilizer can have profound effects on the magnitude of the cooperativity. Specifically, the results revealed a strong increase in PPI selectivity by virtue of this cooperativity. The nondifferentiated stabilizing effects of FC-A, FC-J, and FC-NAC on the p65/14-3-3 complex demonstrate that higher intrinsic affinities of the stabilizer (FC-A and FC-J have similar binding affinities toward 14-3-3, while FC-NAC has a significantly higher affinity<sup>11,25</sup>) do not necessarily translate into better PPI stabilization. The higher cooperativity of DP-005 for the p65/14-3-3 PPI resulted in its most favorable stabilization, while simultaneously featuring similar or weaker capacity to stabilize other 14-3-3 PPIs, which thus leads to selectivity. PPI stabilization in general is an interplay of the intrinsic affinity of the binding partners and the cooperative binding. Increasing cooperativity rather than intrinsic affinity, during compound optimization, will simultaneously generate selectivity. FA-based cooperativity analysis, as described here, represents an excellent approach suitable to medium-throughput analysis with lower demand of compound as compared to ITC-based cooperative analysis. The 2D titration profiles enable an identification of the concentration regime under which a stabilizer might be useful and identify compounds likely to elicit good selectivity early in the drug discovery process.

In conclusion, this study provides the structural and biophysical basis of the p65/14-3-3 interaction as an important

step in the identification and rational design of small-molecule modulators. Specifically, the FC natural products and derivatives are shown to be highly promising tool compounds, with chemical entries to achieving PPI selective stabilization by means of cooperativity. The high cooperativity of the most promising compound DP-005, addressing the uniquely hydrophobic p65/14-3-3 interface pocket, also results in a low threshold concentration for realizing PPI stabilization. This study highlights the way toward selective PPI stabilization, with cooperativity analysis providing an analytical tool to guide compound or stabilizer optimization in PPI drug discovery projects.

## ASSOCIATED CONTENT

### Supporting Information

The Supporting Information is available free of charge at <https://pubs.acs.org/doi/10.1021/jacs.0c02151>.

p65 sequence homology, detailed fluorescence anisotropy data, peptide sequences, protein expression and purification, crystallography details, extensive clarification of the mass balance model, and materials and methods including compound synthesis and characterization (PDF)

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## Notes

The authors declare no competing financial interest.

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