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ARTICLE

A novel codrug made of the combination of ethionamide and its potentiating booster: Synthesis, self-assembly into nanoparticles and antimycobacterial evaluation

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Ethionamide (ETH) is one of the most widely used second-line chemotherapeutic drugs for the treatment of multi-drug-resistant tuberculosis. The bioactivation and activity of ETH is dramatically potentiated by a family of molecules called “boosters” among which BDM43266 is one of the most potent. However, the co-administration of these active molecules is hampered by their low solubility in biological media and by the strong tendency of ETH to crystallize. A novel strategy is proposed here consisting in synthesizing a codrug able to self-associate into nanoparticles prone to be taken up by infected macrophages. This codrug is designed by tethering *N*-hydroxymethyl derivatives of both ETH and its booster through a glutaric linker. This codrug self-assembles into nanoparticles of around 200 nm, stable upon extreme dilution without disaggregating as well as upon concentration. The nanoparticles of the codrug can be intranasally administered overcoming the unfavorable physico-chemical profiles of the parent drugs. Intrapulmonary delivery of the codrug nanoparticles to *Mtb* infected mice *via* intranasal route at days 7, 9, 11, 14, 16 and 18 post-challenge reduces bacterial load in the lungs by a factor of 6.

Introduction

The Tuberculosis (TB) is a leading infectious cause of death worldwide. About one-third of the world's population is infected with *Mycobacterium tuberculosis* (*Mtb*), the bacterium that causes TB. Ethionamide (ETH, **1**) is one of the most widely used second-line chemotherapeutic drugs for the treatment of multi-drug-resistant TB (MDR-TB).^{1–3} ETH inhibits the mycobacterial enoyl-acyl carrier protein reductase *InhA*, thus impairing the biosynthesis of mycolic acids, essential components of the bacterial wall.⁴ Although the emergence of MDR-TB makes second line TB drugs become increasingly important, ETH suffers from a low therapeutic index and high rates of dose-dependent drug adverse effects, including nausea, abdominal pain, liver and thyroid complications.⁵ ETH is actually a prodrug which is converted into its active nicotinamide adenine dinucleotide (NAD) adduct inside the

mycobacteria by EthA, a flavin-containing Bayer-Villiger mono-oxygenase enzyme that oxidizes ETH into its *S*-oxide derivative and then into reactive radicals.⁶ The production of EthA is controlled by the transcriptional repressor EthR. The latter limits the expression of EthA and thereby the bioactivation of ETH.⁷ The sensitivity of *Mtb* to ETH can be enhanced by using small-molecule inhibitors of EthR that allosterically inactivate its DNA-binding activity. The discovery of efficient EthR inhibitors that when used in combination with ETH allowed to dramatically increase the *in vitro* and *in vivo* potency of ETH and to reduce its side effects was a major advance.^{8,9} Among them, BDM43266 has been identified as a potent ligand of EthR allowing at low concentration (80 nM) to increase by 10-fold the *Mtb* growth inhibition by ETH in macrophages (Fig. 1).¹⁰ However the co-administration of ETH/BDM43266 combination regimen is hampered by the poor water solubility of both compounds accentuated by the propensity of ETH to crystallize.^{11,12} The simultaneous delivery of two drugs, while maintaining an appropriate concentration, is a difficult challenge because when compounds, with dissimilar pharmacokinetics, are administered together as a physical mixture they are transported to the site of action with different efficiencies. As a result the expected additive therapeutic effects are not optimal while concurrently the risk of adverse effects increases.¹³

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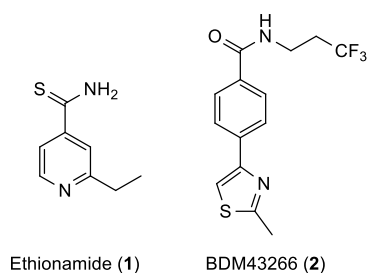


Fig. 1 Structure of ethionamide and of its BDM43266 booster

The design of a codrug is thought to offer a potential and effective implement for addressing this issue. Codrugs consist of two different synergetic drugs joined together in a single entity directly or through a linker designed to release the parent drugs by metabolic bio-transformations.^{14,15} In the present case the ETH/BDM43266 codrug would optimally embed self-assembling properties allowing direct pulmonary delivery. Direct administration of anti TB drugs into lungs is a promising approach to improve the efficiency of the treatments as it increases drug concentration at the infection site while minimizing systemic toxic effects.¹⁶ It must be emphasized that the high crystallinity of ETH added to the low solubility of BDM43266 completely impaired the co-administration by nebulization of a physical mixture of the two drugs directly into the lungs. To address this problem, cyclodextrin-based nanocarriers able to encapsulate simultaneously ETH and BDM43266 with a 5 wt% drug loading have recently been developed.^{17,18} The nanocarriers of around 10 nm were found non-toxic and suitable for pulmonary administration by nebulization directly into the lungs. In this study, we attempted to go steps further by elaborating nanocarriers practically made of pure drugs. The challenge was to couple ETH and its booster in such a way that the novel bioconjugate would self-assemble under the form of nanoparticles prone to be taken up by infected macrophages. We report herein a full account of the synthesis of a mutual prodrug of ETH and its BDM43266 potentiating booster bound together through a potentially cleavable linker and its formulation as stable nanoparticles through spontaneous self-assembling of the codrug leading to a total of 78 wt% drug content. Moreover, we disclose a first *in vivo* proof of concept of the therapeutic efficacy of the codrug nanoparticles against TB after intranasal administration in a model of *Mtb*-infected mice.

Results and Discussion

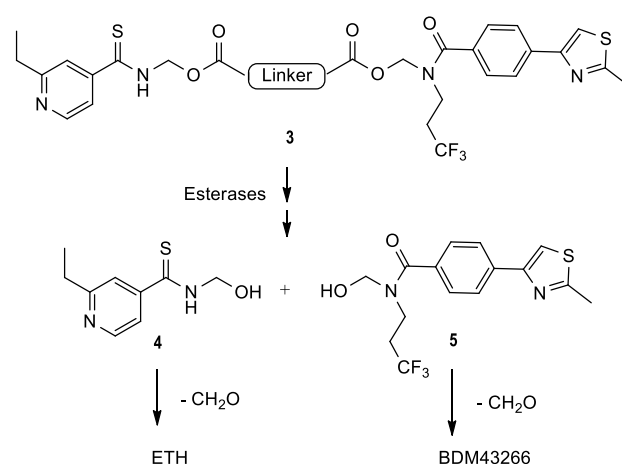
Synthesis of the ethionamide-booster codrug (ETH-B)

The design of prodrug from molecules possessing hydroxyl, amino, carboxylic acid groups or other available acidic functions is well established. In the present case the two drugs display only amide and thioamide groups respectively. Most known prodrug forms for amide drugs are N-acyl derivatives that suffer from the low regioselectivity of the cleavage of the

corresponding imide.¹⁹ Alternatively, Mannich bases or N-hydroxyalkyl derivatives have been proposed but in spite of the fact that a number of potentially useful prodrugs were prepared, very few compounds have been marketed to date.²⁰⁻²² N-acyloxyalkyl prodrugs appear as the most promising strategy, combining chemical stability and easy release upon exposure to esterase via the corresponding hydroxyalkyl derivative.²³

Thus, we initially chose to synthesize the bis-N-acyloxyalkyl prodrug **3**, anticipating that esterases should release the hydroxyalkyl derivative of both drugs (**4** and **5**) that should then decompose into ETH and BDM43266 by release of formaldehyde at physiological pH according to a base catalysis mechanism. Formaldehyde is produced endogenously by various oxidative demethylation reactions catalyzed by cytochromes, low levels of formaldehyde exists *in vivo* and exogenous formaldehyde is produced by currently marketed drugs such as pivampicillin, methenamine or rolitetracycline.²⁴ Considering the hydrophobic character of the BDM43266 bearing a trifluoropropyl appendage we may expect its coupling with the more polar ethionamide through a polyoxygenated spacer to give an amphiphilic conjugate embedding self-assembling properties. Furthermore by modulating the diacid linker between the two drugs it will be possible to adjust the amphiphilic balance and the distance between both entities (Scheme 1).

To start our synthesis, we first examined the formation of the N-hydroxymethyl derivative **4** from ETH (**1**). In the event, treatment of **1** by a large excess of formalin (25 equiv. of formaldehyde 37 wt. % in H₂O) as reported by Böhme and Hotzel^[25,26] provided the desired hemiaminal **4** but accompanied by many side products and formaldehyde polymers. Upon reducing the amount of formalin to 4 equiv. a 56% yield of **4** was achieved after chromatographic purification.

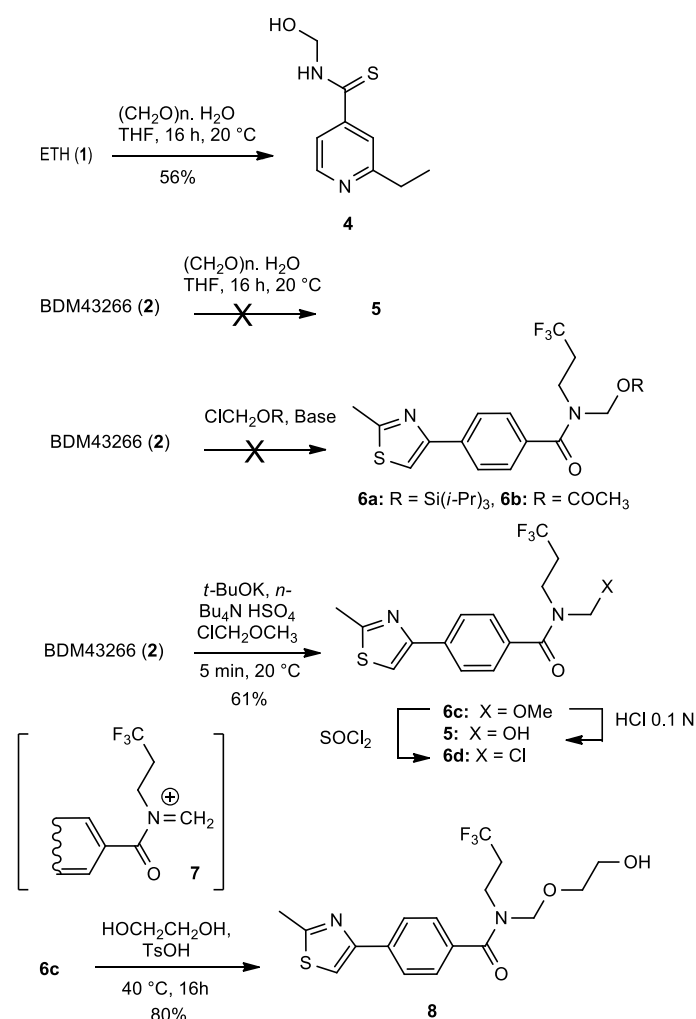


Scheme 1 General structure of the ethionamide-BDM43266 codrug **3** and hypothetical mechanism of the release of the parent drugs.

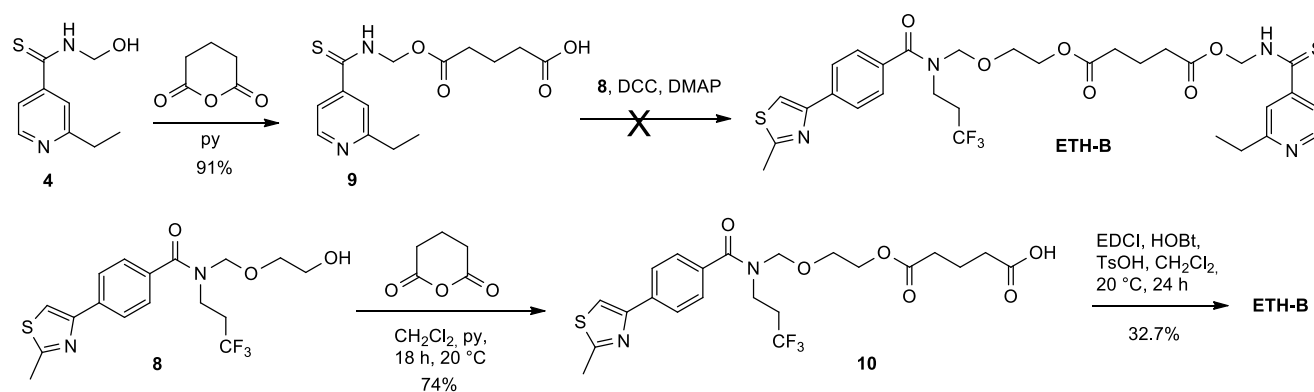
Unfortunately, the same procedure failed to furnish the corresponding hydroxymethyl derivative **5** from the BDM43266 amide **2** whatever the conditions tested. Likewise, reaction of **2** with paraformaldehyde and Me_3SiCl according to the method of Majumdar et al.²⁷ did not give the expected *N*-chloromethyl derivative **6d**. We therefore turned our attention to *N*-alkylative procedures. Unfortunately, alkylation of the anion derived from amide **2** upon treatment with potassium bis(trimethylsilyl)amide (KHMDs) and various electrophiles including (chloromethoxy)tris(propan-2-yl)silane or acetic acid chloromethyl ester failed to provide the expected derivatives **6a**, or **6b**.^{28,29} It was finally found that exposure of amide **2** to chloromethyl methyl ether and *t*-BuOK in solid-liquid phase transfer conditions as reported by Danikiewicz et al. delivered **6c** in 61% yield.³⁰ Interestingly, although a complete conversion was observed after a few minutes reaction, reversion to the starting material occurred upon longer reaction times. This method allowed the functionalization of the booster amide group of BDM43266 but did not provide any handle to tether the ETH moiety. To address this problem we speculated that treatment of **6c** with a Lewis or a Brønsted acid should deliver the *N*-acyliminium ion **7** that might further react with a suitable nucleophile. To check this idea compound **6c** was treated with succinic acid in the presence of TsOH as catalyst, but only reversion of the starting material to **2** was observed. Likewise, all attempts to acylate the *N*-hydroxymethyl derivative **6d**, available in low yield along amide **2** by direct hydrolysis with 0.1 N HCl of the methoxymethyl derivative **6c**, with various reagent induced only reversion to starting amide **2**. Reversing the polarity of the two partners was also fruitless. For example SOCl_2 treatment of **6c** provided the chloromethyl derivative **6d** in moderate yield together with some starting amide **2**. However, $\text{S}_\text{N}2$ displacement of the chloride **6d** with cesium propionate used as a simple carboxylate model did not afford the expected acyloxymethyl adduct but once again reversion to the starting amide **2**, while the same method was found successful on a simple *N*-chloromethylbenzamide model (Scheme 2).

All attempts to introduce the desired *N*-(acyloxymethyl) carboxamide group having failed, we thus turned our attention to the introduction of a simple hemiaminal linker that might ultimately be hydrolyzed in vivo as it was shown for other alkoxyethyl prodrugs.^{31,32} Thus, treatment of **6c** with ethylene glycol in the presence of a catalytic amount of TsOH was attempted. However, no reaction was observed. This apparent lack of reactivity was attributed to the significant basicity of the thiazole ring [pKa 2.54 (Marvin Sketch)]. Accordingly, when the same reaction was conducted in the presence of 1.1 equiv of TsOH the desired alcohol **8** was obtained in 80% yield (Scheme 2). Introduction of the diacid linker was next examined. To this aim, the hemiaminal **4** was condensed with glutaric anhydride to give acid **9** in 91% yield. Unexpectedly, when acid **9** was reacted with the booster derivative **8** using dicyclohexylcarbodiimide (DCC) as coupling agent, the ethionamide-booster conjugate (ETH-B) was not observed. Instead, a complex mixture was obtained in which

the only identified products were the hemiglutarate ester of alcohol **8** and some starting amide **2**. The same result was obtained with a large range of coupling agents. Suspecting some instability of the activated form of the acid **9**, we decided to reverse the partners in the final coupling. To this aim, the alcohol **8** was first reacted with glutaric anhydride to give **10** in 74% yield. Condensation of acid **10** with the ETH hemiaminal **4** using *N*-(3-dimethylaminopropyl)-*N'*-ethylcarbodiimide.HCl /hydroxy-benzotriazole (EDCI/HOBt) as coupling agent, provided the desired codrug ETH-B but in minute amount. A large variety of coupling agents (DCC, 4-(4,6-dimethoxy-1,3,5-triazin-2-yl)-4-methylmorpholinium chloride (DMTMM), hexafluorophosphate azabenzotriazole tetramethyluronium (HATU), mixed anhydride using ClCO_2Et , acid chloride obtained upon ClCOCl or $(\text{CH}_3)_2\text{C}=\text{CCINMe}_2$ treatment, etc.) was tested but improving only marginally this result. It was ultimately found that reaction of the thiazole ring with the activated species was the main cause of the problem.



Scheme 2 Synthesis of *N*-hydroxymethyl derivatives of ethionamide and functionalization of the BDM43266 booster



Scheme 3 Attempted synthesis of the ETH-B codrug by tethering the glutaric spacer on the *N*-hydroxymethyl ethionamide derivative **4** and final synthesis of the ETH-B codrug

We therefore decided to add a proton source to protonate the nitrogen atom of the thiazole during the coupling, keeping in mind that the hemiaminal function is strongly acid sensitive.

In the event when the standard EDCI coupling was conducted in the presence of 1.2 equiv. of TsOH the desired ester was obtained in 32.7% yield after chromatographic purification. Thus, although the yield of the final coupling step needs to be improved, this synthetic route represented an acceptable synthesis of the codrug ETH-B for formulation studies and preliminary biological evaluation (Scheme 3).

Nanoparticle formulation

ETH-B NPs were formed by simple nanoprecipitation in water and were shown to possess spherical shapes and uniform morphologies as determined by cryogenic transmission electron microscopy (cryo-TEM) (Fig. 2). They were found malleable, adapting their shape to the copper grid used for cryo-TEM, but did not stick to each other, possibly because the surface of the NPs was covered by polyvinyl alcohol (PVA), which was added in the aqueous solution (1 wt. %) as emulsifying agent for nanoprecipitation. The NPs prepared at ETH-B 1 mg/mL concentration were further characterized by Nanoparticle Tracking Analysis (NTA) showing a mean diameter of 195 ± 28 nm. NTA is a method of choice as it allows detecting individual NPs in their Brownian motion so that mean diameters can be calculated from NPs trajectories. In this study, around 5000 NPs were detected allowing obtaining statistically relevant data.

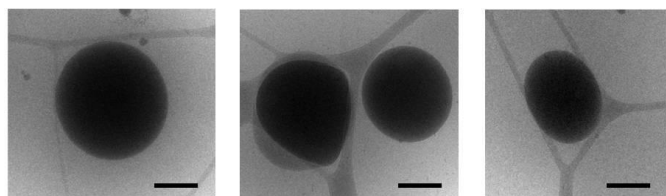


Fig. 2 Typical cryoTEM images of EthB NPs deposited on a copper grid. Scale bar represents 100 nm.

It was also found that the ETH-B NPs were stable upon storage at 4 °C for one week with less than 8% diameter changes (210 ± 31 nm after one week storage). Proton NMR investigations were carried out to study NP integrity after preparation to detect any possible degradation of the codrug. For this, NP suspensions were centrifuged after their preparation, pellets and supernatant were recovered and submitted to ^1H NMR investigations. Less than 1 wt% of codrug was found in the supernatants, showing that the new hydrophobic ETH-B bioconjugate efficiently associated to form the NPs. The NPs recovered in the pellets have identical composition as the starting ETH-B conjugate, demonstrating that no degradation or hydrolysis occurred during NP preparation.

To reach the optimal therapeutic drug concentration, the increase in the concentration of NPs has been investigated. Thus, when NP suspensions, initially at 1 mg/mL ETH-B were concentrated up to 8 mg/mL under nitrogen flow, their size was only slightly modified, from 195 ± 28 nm to 208 ± 34 nm after concentration. This supported the fact that the ETH-B NPs did not aggregate upon high concentration. It was further interesting to study the stability of the NPs upon high dilution, in order to detect a possible disassembling effect. For this, NPs were diluted 1000 fold, from an initial concentration of 1 mg/mL ETH-B down to only 1 $\mu\text{g}/\text{mL}$, and their size distributions was analyzed by NTA. A typical size distribution is shown in Fig. 3A, indicating a mean diameter of around 195 ± 28 nm with narrow size distribution. In the concentration range suitable for NTA, it was found that the NP size remained constant whatever the dilution (Fig. 3B). Moreover, the number of NPs per volume unit of suspension media was calculated and related to the mass of ETH-B in the same volume. Remarkably the ratio NP number/mass of ETH-B remained constant, indicating that the NPs were not destabilized (divided or aggregated). These results supported the good stability of the ETH-B codrug NPs, even upon extreme dilution, suggesting their suitability for further *in vivo* studies.

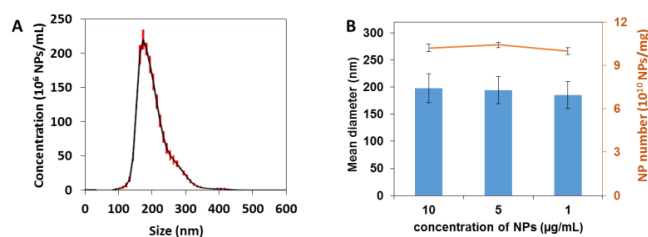


Fig. 3 Mean hydrodynamic diameters and concentration of ETH-B NP suspensions upon dilution. A. Size distribution of ETH-B NPs at a concentration of 1 µg/mL. Error bars are shown in red and were calculated as averages on 5 measurements. B. Mean hydrodynamic diameters (blue) and number (orange) of ETH-B NPs upon dilution. NP number corresponds to the number of NPs formed by a determined mass (1 mg) of ETH-B codrug and takes into account the dilution factor. It was directly determined by NTA by counting at least 5000 particles at concentrations of 1, 5 and 10 µg/mL.

Moreover, the ETH-B codrug NPs were found to be stable in physiological media complemented with a model protein, human serum albumin, at pH ranging from 1.2 to 7.4, suggesting their suitability for further *in vivo* studies (Table S1).

In vivo evaluation in a model of *Mtb* infected mice

ETH and BDM43266 could not be administered *in vivo* into the lungs of infected mice without the need of an appropriate carrier, in reason of the hydrophobicity of these compounds and of the strong tendency of ETH to crystallize.¹⁸ Herein, we address this challenge by using stable ETH-B NPs of around 200 nm. The NPs could be concentrated to ETH-B concentration of 8 mg/mL and administered by intranasal route to infected mice (Fig. 3). It is worth mentioning that in order to administer equivalent amounts of ETH, this drug needs to be solubilized and a mixture of DMSO: water containing around 40–50 vol % DMSO is needed.¹⁸ This elevated amount of organic solvent is poorly compatible with an *in vivo* administration. Keeping in mind that alveolar macrophages are reservoirs for *Mtb*, another advantage of the ETH-B NPs is their size of around 200 nm, prone to induce macrophage uptake to target the bacteria within host cell.³³ To study the efficacy of ETH-B NPs, mice were inoculated with *Mtb* H37Rv *via* the intranasal (i.n.) route and then mice received six administrations *via* gavage (gav.) or i.n. route at days 7, 9, 11, 14, 16 and 18 post-challenge (Fig. 4A). Contrary to gavage treatment which has no effect to contain *Mtb* burst, intranasal administration, allowed to reduce bacterial load in the lungs by a factor of 6 (Fig. 4B). This result demonstrated the benefit of using the pulmonary route of infection to target more specifically *Mtb* within reservoir cells. Using the same experimental design, we recently showed that ETH itself is ineffective when given intranasally in lungs during two weeks,³⁴ therefore a 6-fold reduction of the bacterial burden obtained after only 6 administrations of ETH-B NPs is encouraging. The next studies will aim at investigating whether an increase in the treatment duration or in the amount of administrated drugs could improve ETH-B efficacy. When compared with other strategies of ETH/BDM administration using nanocarriers, the use of ETH-B codrug

ensures simultaneous delivery of both drugs at the site of *Mtb* infection. Taken together, our results suggested that the pulmonary administration of ETH-B NPs could be a promising strategy to improve current anti-*Mtb* therapy in humans leading to a decrease of the antibiotics daily dose and the treatment duration. Such a non-invasive treatment could also be used to decrease the bacterial load in patients arriving at the hospital, in order to protect the staff and the family in contact.

Conclusions

In summary, an efficient synthesis of the ETH-B codrug of ETH and BDM43266 has been devised in which the key steps are the hydroxymethylation of both drugs. The obtained codrug was found able to self-assemble into NPs of around 200 nm which were remarkably stable either upon concentration (8 mg/mL) or dilution (1 µg/mL). Noticeably, the yield of NP formation was close to 100% in reason of the hydrophobicity of the new compound and no degradation occurred during NP preparation. The formed nanoparticle displayed an impressive drug loading of almost 80%. The suspension was suitable for administration directly to the lungs of infected mice, reducing the bacterial load by a factor of 6. We believe that this strategy can be extended to other synergistic drug combinations setting the way for promising therapeutic applications.

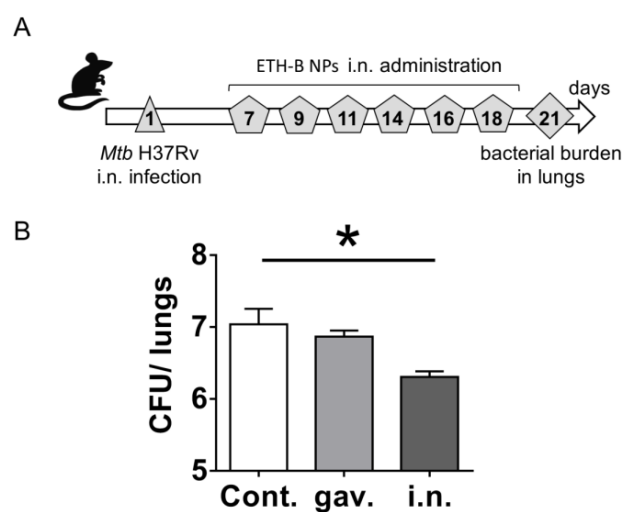


Fig. 4 Impact of ETH-B NPs on the *Mtb* bacterial load in the lungs of infected mice. (A) Experimental design; BALB/c mice were anesthetized and inoculated *via* intranasal (i.n.) route with 10^5 CFU of *Mtb* H37Rv strain contained in 20 µL of PBS. At days 7, 9, 11, 14, 16 and 18 post challenge, mice received administrations of 50 µL of ETH-B NPs suspension *via* gavage (gav.) or i.n. route (concentration of 8 mg/mL). At day 21 post infection, lungs were harvested for bacterial burden determination by colony forming unit (CFU) counting. (B) Comparison of bacterial burden (Log scale) in lungs of 21 days *Mtb* infected non-treated (cont.) mice and 21 days infected mice administrated six times with ETH-B NPs *via* intranasal (i.n.) route or gavage (gav.). * denote $p < 0.05$ (Mann–Whitney test).

Experimental

General information

IR spectra were obtained as solid or neat liquid on a Fourier Transform Bruker Vector 22 spectrometer. Only significant absorptions are listed. The ^1H and ^{13}C NMR spectra were recorded on Bruker Avance 300 (300 MHz and 75 MHz, for ^1H and ^{13}C , respectively) or Bruker Avance 400 (400 MHz and 100 MHz, for ^1H and ^{13}C , respectively) spectrometers. Recognition of methyl, methylene, methine, and quaternary carbon nuclei in ^{13}C NMR spectra rests on the J-modulated spin-echo sequence. Mass spectra were recorded on a Bruker Esquire-LC. Analytical thin-layer chromatography was performed on Merck silica gel 60F254 glass precoated plates (0.25 mm layer). Column chromatography was performed on Merck silica gel 60 (230-400 mesh ASTM). Diethyl ether and tetrahydrofuran (THF) were distilled from sodium/benzophenone ketyl. DMF, and CH_2Cl_2 were distilled from calcium hydride, under a nitrogen atmosphere. All reactions involving air- or water-sensitive compounds were routinely conducted in glassware which was flame-dried under a positive pressure of nitrogen or argon. Ethionamide and 4-acetylbenzoic acid were purchased from Sigma-Aldrich Chemical Co., France. 3,3,3-Trifluoropropylamine was purchased from Ugarit Chimie (France); PVA (88% hydrolysed, average M_w 31,000-50,000 Da) was obtained from Sigma-Aldrich (Saint Quentin Fallavier, France). Chemicals obtained from commercial suppliers were used without further purification. BDM43266 was synthesized according to B. Villemagne et al.¹⁰ pKa were calculated using the MarvinSketch 6.2.2 program (ChemAxon Ltd).

2-Ethyl-N-(hydroxymethyl)pyridine-4-carbothioamide (4)

To a solution of ethionamide (5.0 g, 30 mmol) in THF (90 mL) was added 37% aqueous formaldehyde (120 mmol, 4 equiv., 9 mL) and water (120 mL). The reaction mixture was stirred at room temperature for 16 h. After removing the solvent under reduced pressure the aqueous phase was extracted with CH_2Cl_2 (5 \times 20 mL). The combined organic layers were washed with brine, dried over magnesium sulfate and concentrated under reduced pressure to give the hemiaminal **4** as a yellow solid (3.30 g, 56%). The crude product was pure enough to be directly used in the next step. An analytical sample was obtained by silica gel column chromatography eluting with $\text{CH}_2\text{Cl}_2/\text{MeOH}$ 97:3. Mp 100-102 °C; ^1H NMR (300 MHz, DMSO- d_6) δ 10.90 (br s, 1H, NH), 8.52 (d, J = 5.1 Hz, 1H, H-6), 7.47 (s, 1H, H-3), 7.43 (d, J = 5.1 Hz, 1H, H-5), 6.26 (t, J = 6.9 Hz, 1H, OH), 5.11 (t, J = 5.4 Hz, 2H, HNCH_2OH), 2.79 (q, J = 7.6 Hz, 2H, CH_2CH_3), 1.23 (t, J = 7.6 Hz, 3H, CH_2CH_3) ppm; ^{13}C NMR (75 MHz, DMSO- d_6) δ 195.99 (C, C=S), 163.19 (C, C-2), 149.15 (CH, C-6), 148.00 (C, C-4), 119.16 (CH, C-3), 118.73 (CH, C-5), 69.39 (CH₂, OCH_2NCS), 30.71 (CH₂, CH_2CH_3), 13.71 (CH₃, CH_2CH_3) ppm; IR (neat): ν = 3200 (br), 3064, 3028, 2978, 2935, 2877, 1606, 1566, 1548, 1474, 1461, 1417, 1398, 1379, 1338, 1274, 1223, 1208, 1186, 1151, 1114, 1103, 963, 942, 882, 860, 824, 810, 730, 701 cm^{-1} ; MS (ESI+) m/z (%): 197.1 (100) $[\text{M} + \text{H}]^+$, 167.1 (80) $[\text{M} + \text{H} - \text{CH}_2\text{O}]^+$; (Fig. S1,S2)

N-(Methoxymethyl)-4-(2-methyl-1,3-thiazol-4-yl)-N-(3,3,3-trifluoropropyl)benzamide (6c)

To a solution of BDM43266¹⁰ (447 mg, 1.42 mmol) in anhydrous THF (10 mL) were sequentially added *n*-tetrabutylammonium hydrogensulfate (48 mg, 0.14 mmol) and *t*-BuOK (480 mg, 4.26 mmol, 3 equiv.) and the resulting mixture was stirred for 15 min. Chloromethylmethyl ether (343 mg, 4.26 mmol, 3 equiv.) was then added dropwise to the suspension at room temperature. After 5 min. stirring, 0.5 N HCl (10 mL) was added and the THF was removed under reduced pressure. The aqueous residue was extracted with AcOEt (3 \times 10 mL). The combined organic layers were washed with brine, dried over magnesium sulfate and concentrated under reduced pressure. The residue was purified by silica gel column chromatography eluting with AcOEt/petroleum ether 4:1 to give methoxymethyl derivative **6c** as a colorless oil (312 mg, 61%). ^1H NMR (300 MHz, CDCl_3) δ 7.93 (d, J = 8.1 Hz, 2H, H-2_{Ar}, H-6_{Ar}), 7.53 (d, J = 8.1 Hz, 2H, H-3_{Ar}, H-5_{Ar}), 7.40 (s, 1H, $\text{HC}=\text{C}(\text{N})\text{Ar}$), 4.63 (br s, 2H, $\text{CONCH}_2\text{OCH}_3$), 3.78 (t, J = 7.0 Hz, 2H $\text{NCH}_2\text{CH}_2\text{CF}_3$), 3.19 (s, 3H, OCH_3), 2.79 (s, 3H, $\text{S}(\text{CH}_3)\text{C}=\text{N}$), 2.65-2.57 (m, 2H, $\text{NCH}_2\text{CH}_2\text{CF}_3$) ppm; ^{13}C NMR (75 MHz, CDCl_3) δ 172.03 (C, CON), 166.23 (C, $\text{CH}_3\text{C}(\text{S})=\text{N}$), 153.86 (C, $\text{HC}=\text{C}(\text{N})\text{Ar}$), 136.39 (C, C-4_{Ar}), 134.24 (C, C-1_{Ar}), 127.80 (2CH, C-2,6_{Ar}), 126.20 (2CH, C-3,5_{Ar}), 126.25 (q, J_{CF} = 275 Hz, CF_3), 113.75 (CH, $\text{HC}=\text{C}(\text{N})\text{Ar}$), 81.80 (CH₂, CONCH_2O), 55.08 (CH₃, OCH_3), 39.75 (CH₂, $\text{NCH}_2\text{CH}_2\text{CF}_3$), 32.31 (q, J_{CF} = 28.0 Hz, CH₂, $\text{NCH}_2\text{CH}_2\text{CF}_3$), 19.19 (CH₃, $\text{S}(\text{CH}_3)\text{C}=\text{N}$) ppm; ^{19}F NMR (188 MHz, CDCl_3) δ -65.28 ppm; IR (neat): ν = 3028, 2978, 1645, 1635, 1610, 1456, 1421, 1390, 1343, 1327, 1282, 1251, 1221, 1189, 1169, 1135, 1108, 1084, 1039, 1017, 990, 952, cm^{-1} ; MS (ESI+) m/z (%): 381.1 (100) $[\text{M} + \text{Na}]^+$, 358.10 (62) $[\text{M} + \text{H}]^+$; (Fig. S3,S4).

N-[(2-hydroxyethoxy)methyl]-4-(2-methyl-1,3-thiazol-4-yl)-N-(3,3,3-trifluoropropyl) benzamide (8)

To a solution of hemiaminal **6c** (312 mg, 0.87mmol) in ethane-1,2-diol (3 mL) at 45°C was added *p*-toluenesulfonic acid (165 mg, 1.1 equiv.) and the mixture was stirred at 40 °C for 16 h. The reaction mixture was neutralized with a saturated aqueous solution of sodium hydrogen carbonate, and the mixture was extracted with CH_2Cl_2 (3 \times 10 mL). The combined organic phases were washed with brine, dried over magnesium sulfate and concentrated under reduced pressure to give the title compound as a colorless oil (269 mg, 80%) which was directly engaged in the next step. ^1H NMR (300 MHz, CDCl_3) δ 7.93 (d, J = 8.3 Hz, 2H, H-2_{Ar}, H-6_{Ar}), 7.53 (d, J = 8.3 Hz, 2H, H-3_{Ar}, H-5_{Ar}), 7.39 (s, 1H, $\text{HC}=\text{C}(\text{N})\text{Ar}$), 4.77 (br s, 2H, CONCH_2O), 3.79 (t, J = 6.4 Hz, 2H, $\text{NCH}_2\text{CH}_2\text{CF}_3$), 3.72 (t, J = 4.1 Hz, 2H, $\text{OCH}_2\text{CH}_2\text{OH}$), 3.40 (br s, 2H, $\text{OCH}_2\text{CH}_2\text{OH}$), 2.79 (s, 3H, $\text{S}(\text{CH}_3)\text{C}=\text{N}$), 2.70-2.50 (m, 2H, $\text{NCH}_2\text{CH}_2\text{CF}_3$) 1.77 (br s, 1H, OH) ppm; ^{13}C NMR (75 MHz, CDCl_3) δ 172.24 (C, CON), 166.64 (C, $\text{CH}_3\text{C}(\text{S})=\text{N}$), 153.98 (C, $\text{HC}=\text{C}(\text{N})\text{Ar}$), 136.52 (C, C-4_{Ar}), 134.41 (C, C-1_{Ar}), 127.99 (2CH, C-2,6_{Ar}), 126.53 (2CH, C-3,5_{Ar}), 113.89 (CH, $\text{HC}=\text{C}(\text{N})\text{Ar}$), 80.95 (CH₂, CONCH_2O), 69.11 (CH₂, $\text{OCH}_2\text{CH}_2\text{OH}$), 61.77 (CH₂, $\text{OCH}_2\text{CH}_2\text{OH}$), 39.93 (CH₂, $\text{NCH}_2\text{CH}_2\text{CF}_3$), 32.46 (q,

$J_{CF} = 26.7$ Hz, CH_2 , $NCH_2CH_2CF_3$), 19.41 (CH_3 , $S(CH_3)C=N$) ppm (the CF_3 was not detected). ^{19}F NMR (188 MHz, $CDCl_3$) δ -65.19 ppm; IR (neat): $\nu = 3600$ -3400, 3092, 3028, 2962, 2919, 1635, 1610, 1479, 1462, 1429, 1398, 1346, 1328, 1257, 1200, 1169, 1137, 1098, 1083, 1008, 988, 910, 864, 858, 844, 810 cm^{-1} ; MS (ESI+) m/z (%): 411.1 (100) $[M + Na]^+$; (Fig. S5,S6).

5-[2-({1-[4-(2-methyl-1,3-thiazol-4-yl)phenyl]-N-(3,3,3-trifluoropropyl)formamido}methoxy)ethoxy]-5-oxopentanoic acid (**10**)

To a solution of alcohol **8** (880 mg, 2.27 mmol) in CH_2Cl_2 (18 mL) was sequentially added anhydrous pyridine (730 mg, 4 eq.), glutaric anhydride (51 mg, 2 eq.) and 4-dimethylaminopyridine (28 mg, 0.1 eq.). The resulting mixture was stirred overnight at 20 °C. The CH_2Cl_2 was then removed under reduced pressure and the mixture was taken up into water. The mixture was extracted with AcOEt (4 × 20 mL) and the combined organic phases were washed with brine (2 × 3 mL), dried over magnesium sulfate and concentrated under reduced pressure. The residue was purified by silica gel column chromatography, eluting with petroleum ether/ ethyl acetate (80:20 to 50:50) to give compound **10** (845 mg, 74%) as a colorless oil; 1H NMR (300 MHz, $CDCl_3$) δ 7.89 (d, $J = 8.0$ Hz, 2H, H-2_{Ar}, H-6_{Ar}), 7.60 (d, $J = 8.0$ Hz, 2H, H-3_{Ar}, H-5_{Ar}), 7.40 (s, 1H, $S(CH_3)C=N$), 4.71 (s, 2H, $CONCH_2O$), 4.23 (t, $J = 4.0$ Hz, 2H, OCH_2CH_2OCO), 3.79 (t, $J = 6.7$ Hz, 2H, $NCH_2CH_2CF_3$), 3.53 (br s, 2H, OCH_2CH_2OCO), 2.82 (s, 3H, $S(CH_3)C=N$), 2.70-2.50 (m, 2H, $NCH_2CH_2CF_3$), 2.46 (t, $J = 7.4$ Hz, 2H, $O_2CCH_2CH_2CH_2CO_2H$), 2.41 (t, $J = 6.9$ Hz, 2H, CH_2CO_2H), 1.98 (quint, $J = 7.0$ Hz, 2H, $CH_2CH_2CO_2H$) ppm; ^{13}C NMR (75 MHz, $CDCl_3$) δ 176.86 (C, CO_2H), 172.87 (C, CO_2), 172.08 (C, $ArCO_2$), 167.58 (C, $CH_3C(S)=N$), 153.78 (C, $HC=C(N)Ar$), 136.18 (C, C-4_{Ar}), 134.39 (C, C-1_{Ar}), 128.09 (2CH, C-3,5_{Ar}), 126.54 (2CH, C-2,6_{Ar}), 114.15 (C, $HC=C(N)Ar$), 80.92 (CH_2 , $CONCH_2O$), 65.91 (CH_2 , $NCH_2OCH_2CH_2OCO$), 63.30 (CH_2 , $NCH_2OCH_2CH_2OCO$), 40.32 (CH_2 , $NCH_2CH_2CF_3$), 33.06 (2 CH_2 , $OCOCH_2CH_2CH_2CO_2H$), 32.37 (q, $J_{CF} = 27.0$ Hz, CH_2 , $NCH_2CH_2CF_3$), 20.17 (CH_2 , $O_2CCH_2CH_2CH_2CO_2$), 19.06 (CH_3 , $S(CH_3)C=N$) ppm, the CF_3 signal was masked by the aromatic carbons; ^{19}F NMR (188 MHz, $CDCl_3$) δ -65.24 ppm; IR (neat): $\nu = 3500$ -2500, 1738, 1708, 1645, 1610, 1567, 1531, 1455, 1421, 1393, 1340, 1332, 1291, 1251, 1223, 1172, 1138, 1111, 1087, 1052, 1027, 1016, 993, 861, 846, 791, 753, 745, 687, 655 cm^{-1} ; MS (ESI-) m/z (%): 501.2 (100) $[M - H]^-$; (Fig. S7,S8).

1-[(2-ethylpyridin-4-yl)methanethioamido]methyl 5-[2-({1-[4-(2-methyl-1,3-thiazol-4-yl)phenyl]-N-(3,3,3-trifluoropropyl)formamido}methoxy)ethyl] pentanedioate (ETH-B codrug)

To a solution of acid **10** (538 mg, 1.07 mmol) in CH_2Cl_2 (6 mL) was added at 0 °C, anhydrous TsOH (184 mg, 1.07 mmol) and the reaction mixture was stirred for 10 min. EDCI.HCl (328 mg, 1.71 mmol) and HOBt (231 mg, 1.71 mmol) were then successively added and the mixture was stirred at room temperature. After being stirred for 30 min, N-hydroxymethyl ethionamide (**4**) (420 mg, 2.14 mmol) was added at once. The

reaction mixture was stirred at 20 °C for 24 h and a saturated aqueous solution of $NaHCO_3$ was added (5 mL). The mixture was extracted with CH_2Cl_2 (3 × 10 mL). The combined organic phases were washed with brine, dried over magnesium sulfate and concentrated under reduced pressure. The oily residue was purified by silica gel column chromatography, eluting with CH_2Cl_2 and then with CH_2Cl_2 /MeOH (99:1) to give the title compound together some starting material. Further chromatographic purification eluting with Et_2O /EtOH (100:0 to 65:1) provided the ETH-B codrug as a yellow oil (238 mg, 32.7%). 1H NMR (400 MHz, $CDCl_3$) δ 9.15 (t, $J = 6.4$ Hz, 1H, $CONH$), 8.48 (d, $J = 5.6$ Hz, 1H, H-6_{ethio}), 7.88 (d, $J = 8.0$ Hz, 2H, H-2_{Ar}, H-6_{Ar}), 7.47 (d, $J = 8.0$ Hz, 2H, H-3_{Ar}, H-5_{Ar}), 7.44 (s, 1H, H-3_{ethio}), 7.40 (s, 1H, $HC=C(N)Ar$), 7.32 (d, $J = 5.6$ Hz, 1H, H-5_{ethio}), 5.74 (d, $J = 6.4$ Hz, 2H, CO_2CH_2NHCS) 4.70 (br s, 2H, $CONCH_2O$), 4.16 (br s, 2H, $NCH_2OCH_2CH_2OCO$), 3.77 (br s, 2H, $NCH_2CH_2CF_3$), 3.47 (br s, 2H, $NCH_2OCH_2CH_2OCO$), 2.81 (q, $J = 7.6$ Hz, 2H, CH_2CH_3), 2.75 (s, 3H, $S(CH_3)C=N$), 2.65-2.55 (m, 2H, $NCH_2CH_2CF_3$), 2.43 (t, $J = 7.2$ Hz, 2H, $O_2CCH_2CH_2CH_2CO_2$), 2.38 (t, $J = 7.2$ Hz, 2H, $O_2CCH_2CH_2CH_2CO_2$), 1.94 (quint, $J = 7.2$ Hz, 2H, $O_2CCH_2CH_2CH_2CO_2$), 1.27 (t, $J = 7.6$ Hz, 2H, CH_2CH_3) ppm; ^{13}C NMR (100 MHz, $CDCl_3$) δ 199.77 (C, C=S), 173.43 (C, CO_2CH_2), 172.71 (C, CO_2CH_2N), 172.29 (C, $ArCO_2$), 166.54 (C, $CH_3C(S)=N$), 164.19 (C, C-2_{ethio}), 153.88 (C, $HC=C(N)Ar$), 149.12 (CH, C-6_{ethio}), 148.36 (C, C-4_{ethio}), 136.68 (C, C-4_{Ar}), 134.07 (C, C-1_{Ar}), 127.87 (2CH, C-3,5_{Ar}), 126.45 (2CH, C-2,6_{Ar}), 119.66 (CH, C-3_{ethio}), 117.98 (CH, C-5_{ethio}), 114.05 (C, $HC=C(N)Ar$), 80.83 (CH_2 , $CONCH_2O$), 68.34 (CH_2 , CO_2CH_2NHCS), 65.76 (CH_2 , $NCH_2OCH_2CH_2OCO$), 63.23 (CH_2 , $NCH_2OCH_2CH_2OCO$), 39.97 (CH_2 , $NCH_2CH_2CF_3$), 33.01 (CH_2 , CH_2CO_2), 32.91 (CH_2 , CH_2CO_2), 32.40 (q, $J_{CF} = 28.3$ Hz, CH_2 , $NCH_2CH_2CF_3$), 31.10 (CH_2 , CH_2CH_3), 19.88 (CH_2 , $O_2CCH_2CH_2CH_2CO_2$), 19.40 (CH_3 , $S(CH_3)C=N$), 13.73 (CH_3 , CH_2CH_3) ppm (the CF_3 signal was not detected); ^{19}F NMR (188 MHz, $CDCl_3$) δ -65.2 ppm; IR (neat): $\nu = 3400$ -3200, 2980, 1737, 1610, 1598, 1530, 1475, 1454, 1339, 1252, 1223, 1190, 1172, 1141, 1127, 1110, 1088, 1050, 1016, 993, 964, 906, 859, 845, 820, 726, 687 cm^{-1} ; MS (ESI+) m/z (%): 411.1 (17) $[M + Na]^+$, 681.2 (100) $[M + H]^+$, 525.2 (15) $[M - H_2C=NHCS(C_7H_8N)]^+$; HRMS (ESI+) m/z : $[M + H]^+$ calcd for $C_{31}H_{36}F_3N_4O_6S_2$: 681.2023; found, 681.1991; (Fig. S9,S10).

ETH-B NPs preparation

ETH-B NPs were prepared according to a nanoprecipitation method. Briefly, the optimized procedure was as follows: 5.0 mg of codrug were accurately weighed and dissolved in 300 μ l acetone. This organic phase was added dropwise into an aqueous phase (5.0 mL MilliQ water containing 1 wt% PVA) and stirred magnetically at room temperature until complete evaporation of the organic solvent. The NP suspension was further concentrated up to 8.0 mg/mL under N_2 flow. All samples were prepared in duplicate.

Individual particle tracking and NPs concentration measurements by NTA

The NPs concentration and stability upon dilution were investigated by NTA analysis (NanoSight LM10 Instrument, Malvern Instruments Ltd, Orsay, France), combining a conventional optical microscope with a laser to illuminate the NPs in Brownian motion in a defined volume. Of main interest here, NPs could be visualized one by one, in a concentration range between 10^6 and 10^9 particles/mL. Mean hydrodynamic diameters, size distributions and concentration of ETH-B NPs were measured by NTA in the concentration range of 100-0.1 $\mu\text{g/mL}$. Each sample was measured 5 times for 60 seconds. Results were reported as mean diameter \pm SD.

Cryo-TEM imaging

5 μl of ETH-B NPs suspensions were deposited onto a 200 mesh copper grid and flash-frozen in liquid ethane cooled down at liquid nitrogen temperature. Cryo-TEM images were acquired on a JEOL 2200FS energy-filtered (20 eV) field emission gun electron microscope operating at 200 kV.

^1H NMR study of NPs after preparation

NPs were centrifuged at 30,000 g for 10 min, both the pellets and the supernatants were recovered and lyophilized. ^1H NMR studies confirmed that no degradation took place during NP preparation.

Stability test of NPs in different physiological media.

Prior to the stability tests, different physiological media were prepared, including 10 mM phosphate buffer saline (PBS) at pH 7.4 and 1.2. Human serum albumin (HSA, 10 mg/mL) which is the most abundant protein in the blood was added to PBS to better mimic the biological environment. NPs containing 1 mg/mL ETH-B were diluted in the PBS (pH 7.4 and pH 1.2) with or without HSA. The mean diameters of ETH-B NPs were characterized by dynamic laser scattering (DLS) after incubation at 37 °C for 0, 6, and 24 h.

In vivo efficacy of ETH-B NPs

6-Week old Balb/C female mice were purchased from Janvier (Le Genest-Saint-Isle, France) and were maintained in the animal facility of the Pasteur Institute of Lille, France (Agreement B59-350009). The project received ethical approval by French Committee on Animal Experimentation and the Ministry of Education and Research (00579.01 approved on December 2nd 2015 and APAFIS#10232-2017061411305485 v6 approved on September 2018) and all experiments were performed in accordance with relevant guidelines and regulations.

8-Week-old BALB/c mice (4 mice *per* group) were inoculated with *Mtb* H37Rv *via* the i.n. route (10^5 CFU/20 μl) as described.^[18] On day 7, 9, 11, 14, 16 and 18-post infection, mice were administrated *via* gavage or i.n. route with 50 μl of ETH-B NPs (concentration of 8 mg/mL). At day 21, mice were euthanized and lungs were homogenized in a tube containing

1 mL of 2.5-mm diameter glass beads and 1 mL of PBS using an MM300 bead beater apparatus (Qiagen) for 20 minutes. Ten-fold serial dilutions (from 10^{-2} to 10^{-9}) of each sample were plated onto 7H11 medium agar plates (Difco) supplemented with 10% oleic acid-albumin-dextrose-catalase (OADC, Difco). After a three-week growth at 37 °C, CFUs were determined at the appropriate dilution allowing optimal colony enumeration.

Conflicts of interest

There are no conflicts to declare.

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