

Liquid chromatography—tandem mass spectrometric assay for the quantification of CDK4/6 inhibitors in human plasma in a clinical context of drug-drug interaction

Fanny Leenhardt, Matthieu Gracia, Catherine Perrin, Claudia Muracciole-Bich, Bénédicte Marion, Céline Roques, Marie Alexandre, Nelly Firmin, Stephane Pouderoux, Litaty Mbatchi, et al.

▶ To cite this version:

Fanny Leenhardt, Matthieu Gracia, Catherine Perrin, Claudia Muracciole-Bich, Bénédicte Marion, et al.. Liquid chromatography—tandem mass spectrometric assay for the quantification of CDK4/6 inhibitors in human plasma in a clinical context of drug-drug interaction. Journal of Pharmaceutical and Biomedical Analysis, 2020, 188, pp.113438. 10.1016/j.jpba.2020.113438. hal-03003807v2

HAL Id: hal-03003807 https://hal.science/hal-03003807v2

Submitted on 10 Jun 2024

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

1 Liquid chromatography-tandem mass spectrometric assay for the quantification of CDK4/6 inhibitors in human plasma in a clinical context of drug-drug interaction. 2 3 Authors: 4 Fanny Leenhardt^{1,2,3}, Matthieu Gracia^{1,3}, Catherine Perrin⁴, Claudia Muracciole-Bich⁴, 5 Bénédicte Marion⁴, Celine Roques⁴, Marie Alexandre⁵, Nelly Firmin^{3,5}, Stephane Pouderoux⁵, 6 Litaty Mbatchi^{1,3,6}, Celine Gongora³, William Jacot^{3,5}, Alexandre Evrard^{1,3,6}. 7 8 1- Laboratoire de Pharmacocinétique, Université de Montpellier, Faculté de Pharmacie, 9 France 10 2- Service Pharmacie, Institut du Cancer de Montpellier, Université de Montpellier, 208 rue 11 des Apothicaires, 34298, Montpellier, France. 12 3- Institut de Recherche en Cancérologie de Montpellier (IRCM), INSERM U1194, Université 13 de Montpellier, Montpellier, France. 14 4- Institut des Biomolécules Max Mousseron (IBMM), UMR 5247-CNRS-UM-ENSCM, 15 16 Montpellier, France. 5- Département d'oncologie médicale, Institut du Cancer de Montpellier, Université de 17 18 Montpellier, 208 rue des Apothicaires, 34298, Montpellier, France 6- Laboratoire de Biochimie et Biologie moléculaire, Centre Hospitalier Universitaire Nîmes, 19 20 France 21 Corresponding author: Fanny Leenhardt, fanny.leenhardt@icm.unicancer.fr, Institut du 22 23 cancer de Montpellier, 208 avenue des Apothicaires, Montpellier, France. Phone number : 24 00 336 83 44 04 06 25 26 Credit authorship contribution statement: 27

Fanny Leenhardt: Conceptualization, Methodology, Investigation, Formal analysis, Writing -

Matthieu Gracia: Methodology, Investigation, Formal analysis, Writing - Original Draft

Original Draft, Supervision, Funding acquisition

28

29

- 31 Catherine Perrin: Methodology, Validation, Formal analysis, Writing Original Draft,
- 32 Supervision Claudia Muracciole-Bich: Methodology, Resources, Writing Original Draft
- 33 Bénédicte Marion : Methodology, Investigation, Resources
- 34 Celine Roques : Methodology, Investigation, Resources
- 35 Marie Alexandre : Investigation, Writing Original Draft
- 36 Nelly Firmin: Investigation, Writing Original Draft
- 37 Stephane Pouderoux : Investigation
- 38 Litaty Mbatchi: Writing Original Draft
- 39 Celine Gongora: Writing Original Draft, Project administration
- 40 William Jacot: Conceptualization, Investigation, Writing Original Draft, Supervision, Project
- 41 administration, Funding acquisition
- 42 Alexandre Evrard: Conceptualization, Methodology, Writing Original Draft, Supervision,
- 43 Project administration, Funding acquisition

44 45

46

47

58

<u>Abstract</u>

48 The CDK4/6 inhibitors palbociclib and ribociclib are kinase inhibitors used in association with 49 hormonal therapy for the management of patients with metastatic breast cancer. Like most 50 kinase inhibitors, therapeutic drug monitoring may be used for personalize their dosage. To this aim, we developed and validated a sensitive and specific HPLC-MS/MS method for 51 52 palbociclib and ribociclib quantification in blood samples. We then quantified exposure to 53 palbociclib (plasma trough concentration; C_{trough}) in a real-life cohort of patients with locally invasive or metastatic breast cancer (n=18) at day 15 of the first cycle of palbociclib 54 treatment to characterize palbociclib concentration at steady state (Clinicaltrials.gov 55 identifier NCT04025541). The geometric mean (± standard deviation [min-max]) of 56 57 palbociclib plasma C_{trough} was 88.58 ng/ml (± 26.4 [46.5 ng/mL – 133 ng/mL]) at day 15.

Some covariates, such as drug-drug interactions, could explain the concentration variations

observed in our Caucasian cohort. These first results in real-life settings obtained with our
HPLC-MS/MS method give important information on palbociclib monitoring and
pharmacokinetic variability.

Keywords: CDK4/6 inhibitor; HPLC-MS/MS; metastatic breast cancer; therapeutic drug
monitoring; drug-drug interactions

monitoring; drug-drug interactions

68

69

70

71

72

73

74

75

76

77

78

79

80

81

82

83

84

85

86

87

88

89

90

91

1. Introduction

The cyclin-dependent kinase 4/6 (CDK4/6) inhibitors palbociclib and ribociclib in combination with endocrine therapy are gradually becoming the first-line treatment for patients with locally advanced or metastatic hormone receptor-positive (HR+) /HER2-negative breast cancer. However, these combination therapies show specific adverse events, such as severe neutropenia (grade 3-4 for about 50% of patients), anemia, asthenia and liver toxicity [1]. Nevertheless, their safety profile is globally good, and the febrile neutropenia incidence is about 1% in the pivotal clinical trials. Moreover, they have shown a clear benefit in terms of progression-free survival compared with endocrine therapy alone [1]. As palbociclib and ribociclib require only a single, fixed oral dose per day, their administration is easy to manage but requires good patient's compliance for optimal drug exposure and therapeutic effect. However, neutropenia often leads to treatment interruption/delay or dosage modification, possibly linked to the lack of baseline dose adaptation in function, for instance, of age or body weight. In addition, targeted oral anti-cancer treatments, including CDK4/6 inhibitors, can display intra- and inter-individual pharmacokinetic variability that can influence their efficacy and tolerance. Drug-drug interactions (DDI), food intake and genetic polymorphisms in drug metabolizing enzymes are among the many factors that can influence drug-exposure variability. For instance, the observed variability concerning the Area Under the plasma concentration Curve (AUC) and the steady state trough concentration (Ctrough) of tyrosine kinase inhibitors are about 32% for imatinib, 34% for sunitinib, 24% for pazopanib, and 28% for vemurafenib [2]. For drugs with a linear pharmacokinetics, efficacy and tolerance are mostly related to plasma exposure. Therefore, therapeutic drug monitoring (TDM) is a useful tool to monitor this parameter. Practical recommendations on TDM use are based on pharmacokinetic data, availability of analytical techniques, and clinical trials that used TDM for dosage adjustments [3]. TDM data and recommendations are available for some oral targeted therapies used in solid tumors, such as sunitinib in renal carcinoma and imatinib in gastrointestinal stromal tumor and leukemia chronic myeloid [4]. Conversely, very few evidence-based data are available on TDM usefulness for oral targeted therapies used in breast cancer, such as everolimus and lapatinib. Some methods have been validated for the plasma quantification of CDK4/6 inhibitors, but they are less used in real-life settings [5–7]. Therefore, we developed a specific high performance liquid chromatography-tandem mass spectrometry (HPLC-MS/MS) method suitable for TDM of palbociclib and ribociclib in real-life settings. Here, we report the method validation and also preliminary results obtained in 18 patients with breast cancer at day 15 of the first cycle of palbociclib treatment.

2. Material, patients and methods

2.1. Chemicals

Palbociclib, ribociclib, palbociclib ²H₈ and ribociclib ²H₆ were obtained from Alsachim (Illkirch, France). HPLC-grade acetonitrile (ACN) and methanol were purchased from Carlo Erba Reagents (Val De Reuil, France). Formic acid (FA) (98% pure) was obtained from PanReac AppliChem ITW Companies (Darmstadt, Germany). Dimethylsulfoxide (DMSO) was from Carlo Erba Reagents (Val De Reuil, France). Ultrapure water (H₂O) was produced with a Milli-Q® Simplicity apparatus (Millipore Corp., Burlington, MA, USA).

2.2. Control plasma and blood sample collection

Control plasma samples (blank samples) were obtained from Nîmes University Hospital Center (UHC) and were stored at -20°C. Patient blood samples were collected in EDTA tubes at the Montpellier Regional Institute of Cancer (ICM, Montpellier, France) and at Nîmes UHC, centrifuged and stored at -80°C till analysis. Patient blood samples were from patients with metastatic breast cancer treated with an aromatase inhibitor (letrozole or anastrozole) and CDK4/6 inhibitor (palbociclib or ribociclib) and enrolled in a multicenter prospective clinical trial (ALCINA 2, NCT04025541) initiated at ICM in 2018 to assess the clinical usefulness of various cancer biomarkers. Patients were included after signature of the informed consent. Cohort 1 (palbociclib) recruitment is already ongoing for patients received 125mg oral palbociclib as per therapeutic indication, once per day for 3 weeks followed by one week off. Gold standard of TDM for oral therapy in cancer is to assess the plasma concentration at the predose (C_{trough}) at the steady-state (one to two weeks after treatment start depending on elimination half-lives). Blood samples were collected at day 15 (steady-state reached) of the first and second treatment cycle before drug administration to estimate plasma exposure (C_{trough}). Cohort 2 (ribociclib) is active but not recruiting yet. Palbociclib plasma concentration at the first treatment cycle and at the predose was analyzed for the first twenty patient of cohort 1 (n=18; two patients excluded because blood samples were not collected at C_{trough}). Patients were classified according to their risk of DDI that might lead to inhibition of CYP3A4 and/or P-glycoprotein. Database search (e.g. DDI predictor®, Drugs.com®, Pubmed®) allowed the identification of the following drugs that could cause DDI: fluconazole, ivabradine, atorvastatin, amlodipine, losartan and nifedipine [8-12].

2.3. Stock and working solutions

113

114

115

116

117

118

119

120

121

122

123

124

125

126

127

128

129

130

131

132

133

Individual stock solutions (1 mg/mL) of palbociclib, ribociclib and internal standards (IS; palbociclib 2H_8 and ribociclib 2H_6) were prepared in DMSO and stored at -20°C. Mixed palbociclib, ribociclib and IS working solutions were then prepared in ACN/ H_2O (50:50, v/v) with 0.1% (v/v) FA by mixing the appropriate volumes of analyte and IS stock solutions. The palbociclib and ribociclib working solution concentrations were 78.12, 156.25, 312.5, 625, 1250, and 2500 ng/mL. They were used for the preparation of calibration standards and quality control (QC) samples. The final IS concentrations were 2500 ng/mL for all working solutions. Independent stock solutions were used for the preparation of the calibration and quality control standards. We also have performed quality control sample testing in two laboratories to ensure reproducibility of the method (data not shown).

2.4. Calibration standards, quality control, and patient plasma samples

To compute the calibration curve over a specific concentration range, calibration standards were prepared by spiking 200 μ L of blank plasma with 10 μ L of working solution that contained known concentrations (3.9, 7.8, 15.6, 31.5, 62.5, and 125 ng/mL) of palbociclib and ribociclib, respectively. For each calibration standard, the final IS concentration was 125 ng/mL. To determine the lower limit of quantification (LLOQ), medium, and upper limit of quantification (ULOQ), palbociclib and ribociclib QC samples were prepared at the concentrations of 3.9, 15.6, 50, and 100 ng/mL. All solutions were prepared according to the recommendations for bioanalytical method validation [13,14]. Plasma sample (100 μ L) was mixed with 100 μ L of blank plasma and 10 μ L of the final IS (palbociclib 2 H₈) working solution before sample extraction to avoid concentration measurements out of the limit of calibration range.

2.5. Sample extraction procedure

Sample (calibration standards, QC, patient plasma samples) extraction was performed by Solid Phase Extraction (SPE) with Oasis Hydrophilic-Lipophilic Balance (HLB) columns (1cc; 30gr) (Waters®) on a vacuum support. $100~\mu$ L of methanol/ H_2O (50:50, v/v) and 1mL of FA 28% (v/v) were added to each sample (final volume: 1.310 mL). SPE columns were first conditioned with 2 mL of methanol and 2 mL of H_2O . Then, samples were loaded on the SPE column followed by two rinses (1 mL) with water. Samples were eluted with 1 mL of methanol. Eluted samples were dried and concentrated under a nitrogen stream using a TurboVap® device (Air Liquid, France) coupled with a $37^{\circ}C$ water bath. Dried extracts were reconstituted with $200~\mu$ L ACN/ H_2O (50:50, v/v) and 0.1% (v/v) FA and analyzed ($10~\mu$ L) by HPLC-MS/MS. We also compared SPE to liquid-liquid extraction in order to use the most cost-effective and reliable extraction technique. SPE has proven to be the technique with the highest repeatability and reproducibility, as well as the best performance (data not shown).

2.6. HPLC-MS-MS equipment

- An Agilent 1100 HPLC instrument linked to a triple quadrupole mass spectrometer (MS/MS)
- 173 (API3000, PE Sciex) with a turbo ion spray interface was used for all analyses.
- 174 Chromatographic separations were carried out on a Waters Symmetry® C18 column (4.6 µm
- x 75mm; 3.5 μ m). Data were treated with the Analyst 1.5.2 software.

2.7. HPLC-MS/MS conditions

The column and autosampler temperatures were maintained at 25 °C and 4°C, respectively. Eluent A was 0.1% (v/v) of FA in water and eluent B was ACN with 0.1% (v/v) of FA. Complete separation of palbociclib, ribociclib and IS was obtained using a carefully optimized 22min stepwise gradient with a flow rate of 0.5mL/min: 100% A (0-2 min), 0-50% B (2-8min), 50% B

182 (8-11 min, end of analytical run), 50-100% B (11-14min), 100% B (14-17 min), 100-0% B (17-19 min), and 100% A (19-22 min). The ion source temperature was set to 450°C and the ion 183 spray voltage at 5000 V. The nebulizer, curtain and collision gas pressures were 8, 8, and 4 184 psi, respectively. MS/MS transitions and optimal potential settings were determined for 185 each analyte/IS and are listed in Figure 1. 186 187 2.8. Method validation 188 189 The HPLC-MS/MS method was validated according to the FDA and EMA recommendations [13,14]. 190 191 2.9. Statistical analysis 192 193 The Wilcoxon rank-sum test was used to compare the distribution of quantitative variables 194 (palbociclib plasma exposure and risk of DDI). 195 3. Results 196 197 3.1 Method development Precursor and product ions were obtained for the investigated analytes and IS with collision 198 energy values of 39 V and 45 V for palbociclib and its IS and for ribociclib and its IS,

energy values of 39 V and 45 V for palbociclib and its IS and for ribociclib and its IS,

respectively (Figure 1). Precursor ions (Q1, m/z) for palbociclib and its IS was 448 and 456

respectively, and ribociclib and its IS was 435 and 441. Product ions (Q3, m/z) obtained were

379.9, 388, 322 and 322 for palbociclib, its IS, ribociclib and its IS respectively.

After HPLC gradient optimization, the method specificity was evaluated by replicating

204

205

206

and selective detection, identification and quantification of the components of interest in a mixture, based on monitoring the precursor-fragment ion pair. Figure 1 shows the

analyses using the multiple reaction monitoring (MRM) mode. This mode allows the specific

chromatogram and retention times obtained for the ribociclib and palbociclib working solutions at 125 ng/mL and their IS. Elution step was maintained up to 22 minutes in order to limit the risk of sample carry-over.

210

211

207

208

209

- 3.2 Method validation
- 212 3.2.1 Selectivity
- The selectivity of the method (i.e. the ability to differentiate between palbociclib, ribociclib and IS) for endogenous plasma matrix components was evaluated in four different batches of blank plasma. Selectivity was good with no interference observed between analytes and plasma components (Figure 1). Indeed, the signal of the blank matrix was <2% of that

obtained for palbociclib and ribociclib at the LLOQ (and <0.2% for the IS).

218

217

- 219 3.2.2 Calibration curve
- Linearity was assessed using calibration standards at increasing concentrations: 3.9 (LLOQ),
 7.8, 15.6, 31.2, 62.5, and 129 (ULOQ) ng/mL of palbociclib and ribociclib, respectively. Five
 calibration curves were analyzed on different days for each analyte (i.e. palbociclib and
 ribociclib). All correlation coefficients were >0.998 for palbociclib and ribociclib. All backcalculated concentrations were within 15% of the nominal concentrations (± 20% for the
 LLOQ). For both analytes, the LLOQ signals were >5 times greater than the signal of the blank
 sample.

- 3.2.3 Accuracy and precision
- Within-run and between-run accuracy and precision were studied using calibration standards with the following concentration levels: 3.9 ng/mL (LLOQ), 15.6 ng/mL (about 3 times the LLOQ), 50 ng/mL (about 40% of the calibration curve), and 100 ng/mL (about 78%

of the calibration curve). Accuracy was expressed as the difference (%) between the mean measured concentration and the nominal concentration (bias). Precision was expressed as the coefficient of variation (CV) (%). ANOVA was used to assess the between-run precision. Within-run and between-run accuracy and precision were within the acceptance criteria: ≤ 15% for low, medium and high QC and ≤20% for the LLOQ of the QC (Table 1). Accuracy and precision measurements were analyzed by pooling repeated results using statistical methods (ANOVA). The standard deviation and the common coefficient correlation were checked (Table 1).

3.2.4. Carry-over

Blank samples were injected after analysis of high-concentration QC samples to evaluate the carry-over. For palbociclib, interference was <5% of the peak area observed at the LLOQ. For ribociclib, interference was <2% of the peak area observed at the LLOQ. For both IS, interference was <0.5% of the IS peak area. These results are fully compliant with the required limits: $\leq 20\%$ of peak areas at the LLOQ for the analytes and $\leq 5\%$ of IS area for IS.

3.2.5 Matrix effect

The matrix effect was evaluated at the LLOQ and ULOQ. For each analyte, the normalized matrix effect was estimated by dividing the analyte matrix factor (i.e. analyte peak area in plasma divided by the analyte peak area in the water/ACN mixture) by the IS matrix factor (i.e. IS peak area in plasma divided by IS peak area in the water/ACN mixture). For palbociclib and ribociclib, the normalized matrix effect varied between 0.83 and 1.14. The overall CV of the normalized matrix effect was <15%.

3.2.6 Stability

Plasma samples were stable at ambient temperature for at least 6 hours with observed variations <5% compared with freshly prepared samples for both palbociclib and ribociclib. Samples stored at -20°C were stable for at least one month with <15% of difference with the nominal value.

3.3 Clinical application

The plasma concentration of palbociclib was assessed using our HPLC-MS/MS assay in the first 18 patients treated with palbociclib and aromatase inhibitor in the framework of the ALCINA 2 trial (see Table 2 for their description). The C_{trough} (geometric mean ± standard deviation [min-max]) was 88.58 ng/mL ± 26.4 [46.5 ng/mL – 133 ng/mL] at day 15 of the first cycle of palbociclib treatment, with a CV(%) of 29.8 (Figure 2).

No correlation between plasma concentration and body weight or area was found. DDI-linked pharmacokinetic variabilities, such as drug absorption or metabolism, can modulate palbociclib plasma exposure. Besides palbociclib and aromatase inhibitor treatment, the 18 patients were taking other drugs (mean number: 3.22 per day; min-max: 0-7). Therefore, they were divided in two groups based on the presence (n=7) or not (n=11) of potential DDI (CYP3A4 and P-glycoprotein inhibitors listed in 2.2). Palbociclib C_{trough} at day 15 was

4. Discussion and conclusion

Here, we described a specific, accurate and sensitive HPLC-MS/MS method that we developed and validated for the simultaneous estimation of ribociclib and palbociclib

significantly different in patients with and without potential DDI (p<0.01) (Figure 2).

exposure in patient plasma samples. Most efficient extraction and interlaboratory control have allowed, between quality criteria, a useful and effectiveness method in TDM clinical use. In this first study, the method was also used to monitor palbociclib exposure in a reallife cohort of 18 patients with locally advanced metastatic breast cancer treated with palbociclib and an aromatase inhibitor. Like for most kinase inhibitors used in oncology, the relationship between pharmacokinetics and pharmacodynamics of CDK4/6 inhibitors assessed by TDM could help to improve the treatment efficacy and reduce toxicities. However, with the exception of the data from the PALOMA trials or from small cohorts, clinicians do not have much information on palbociclib plasma exposure [7]. Our preliminary analysis in 18 patients showed a mean C_{trough} of 88.58 ng/mL, similar to what reported in the PALOMA trials. In the PALOMA 1 trial, the geometric mean C_{trough} was 88.5 ng/mL (n=6) [15]. In the PALOMA 2 trial, the mean C_{trough} of palbociclib (%CV) at the steady state was 61 ng/mL (42%), with a mean C_{max} at 116 ng/mL (28%) [16]. In the PALOMA 2 subgroup, ethnicity influenced plasma exposure. Specifically, palbociclib Ctrough was higher in Japanese patients (n=27) than in non-Asian patients (n=142) (95.4 ng/mL versus 61.7 ng/mL) [17]. Our cohort included only Caucasian patients treated with palbociclib (125mg per day; full dose). DDI mediated by drug metabolizing enzymes and transporters are a major source of pharmacokinetic variability. In our subgroup with potential DDI, interactions between palbociclib and CYP3A4 or P-glycoprotein inhibitors could explain the palbociclib concentration variability (74.15 ng/mL vs 111.26 ng/mL with DDI). Additional studies are needed to characterize palbociclib pharmacokinetic variabilities that could explain plasma concentration variations between patients, and their clinical impact. These preliminary data must to be confirmed in both cohorts of the ALCINA 2 study (once enrollment will be

282

283

284

285

286

287

288

289

290

291

292

293

294

295

296

297

298

299

300

301

302

303

completed) in order to evaluate the interest of individualizing CDK4-6 inhibitor dosage according to pharmacokinetic and pharmacogenetic data integrated in a decision algorithm.

307

308

- 309 Declaration of competing interest:
- No competing interests to declare for all authors/provided financial support for the conduct
- 311 of the research
- 312 Acknowledgments: Benoit Blanchet (Hopital de Cochin, Paris, France) and Laboratoire de
- 313 Mesures Physiques (Université de Montpellier, France) for quality control sample testing.
- Funding source: This project was supported by INCa-Cancéropôle GSO.

- 316 [1] Thill M, Schmidt M. Management of adverse events during cyclin-dependent kinase
- 4/6 (CDK4/6) inhibitor-based treatment in breast cancer. Ther Adv Med Oncol
- 318 2018;10:175883591879332. https://doi.org/10.1177/1758835918793326.
- 319 [2] Lucas CJ, Martin JH. Pharmacokinetic-Guided Dosing of New Oral Cancer Agents. J
- 320 Clin Pharmacol 2017;57:S78–98. https://doi.org/10.1002/jcph.937.
- 321 [3] Widmer N, Bardin C, Chatelut E, Paci A, Beijnen J, Levêque D, et al. Review of
- 322 therapeutic drug monitoring of anticancer drugs part two Targeted therapies. Eur J Cancer
- 323 2014;50:2020–36. https://doi.org/10.1016/j.ejca.2014.04.015.
- 324 [4] Herviou P, Thivat E, Richard D, Roche L, Dohou J, Pouget M, et al. Therapeutic drug
- monitoring and tyrosine kinase inhibitors. Oncol Lett 2016;12:1223–32.
- 326 https://doi.org/10.3892/ol.2016.4780.
- 327 [5] Martínez-Chávez A, Rosing H, Hillebrand M, Tibben M, Schinkel AH, Beijnen JH.
- 328 Development and validation of a bioanalytical method for the quantification of the CDK4/6
- 329 inhibitors abemaciclib, palbociclib, and ribociclib in human and mouse matrices using liquid
- chromatography-tandem mass spectrometry. Anal Bioanal Chem 2019;411:5331–45.
- 331 https://doi.org/10.1007/s00216-019-01932-w.
- 332 [6] Janssen JM, de Vries N, Venekamp N, Rosing H, Huitema ADR, Beijnen JH.
- 333 Development and validation of a liquid chromatography-tandem mass spectrometry assay
- for nine oral anticancer drugs in human plasma. J Pharm Biomed Anal 2019;174:561–6.
- 335 https://doi.org/10.1016/j.jpba.2019.06.034.

- Posocco B, Buzzo M, Poetto AS, Orleni M, Gagno S, Zanchetta M, et al. Simultaneous
- 337 quantification of palbociclib, ribociclib and letrozole in human plasma by a new LC-MS/MS
- method for clinical application. PloS One 2020;15:e0228822.
- 339 https://doi.org/10.1371/journal.pone.0228822.
- Tod M, Goutelle S, Bleyzac N, Bourguignon L. A Generic Model for Quantitative
- 341 Prediction of Interactions Mediated by Efflux Transporters and Cytochromes: Application to
- 342 P-Glycoprotein and Cytochrome 3A4. Clin Pharmacokinet 2019;58:503–23.
- 343 https://doi.org/10.1007/s40262-018-0711-0.
- 344 [9] Drugs.com. Drug Interaction Report: Palbociclib information from Drugs.com.
- DrugsCom n.d. https://www.drugs.com/interactions-check.php?drug_list=1717-0,3602-0
- 346 (accessed March 23, 2020).
- 347 [10] Holtzman CW, Wiggins BS, Spinler SA. Role of P-glycoprotein in Statin Drug
- Interactions. Pharmacother J Hum Pharmacol Drug Ther 2006;26:1601–7.
- 349 https://doi.org/10.1592/phco.26.11.1601.
- 350 [11] Wessler JD, Grip LT, Mendell J, Giugliano RP. The P-Glycoprotein Transport System
- and Cardiovascular Drugs. J Am Coll Cardiol 2013;61:2495–502.
- 352 https://doi.org/10.1016/j.jacc.2013.02.058.
- 353 [12] Bellet M, Ahmad F, Villanueva R, Valdivia C, Palomino-Doza J, Ruiz A, et al. Palbociclib
- and ribociclib in breast cancer: consensus workshop on the management of concomitant
- medication. Ther Adv Med Oncol 2019;11:175883591983386.
- 356 https://doi.org/10.1177/1758835919833867.
- 357 [13] FDA. Bioanalytical Method Validation Guidance for Industry 2018:44.
- 358 [14] EMA. Guideline on bioanalytical method validation 2011:23.
- 359 [15] Tamura K, Mukai H, Naito Y, Yonemori K, Kodaira M, Tanabe Y, et al. Phase I study of
- palbociclib, a cyclin-dependent kinase 4/6 inhibitor, in Japanese patients. Cancer Sci
- 361 2016;107:755–63. https://doi.org/10.1111/cas.12932.
- 362 [16] John K. Leighton,. Pharmacological review. Division of Hematology Oncology
- 363 Toxicology; 2015.
- 364 [17] Mukai H, Shimizu C, Masuda N, Ohtani S, Ohno S, Takahashi M, et al. Palbociclib in
- 365 combination with letrozole in patients with estrogen receptor-positive, human epidermal
- 366 growth factor receptor 2-negative advanced breast cancer: PALOMA-2 subgroup analysis of

Japanese patients. Int J Clin Oncol 2019;24:274-87. https://doi.org/10.1007/s10147-018-367 368 1353-9. 369 370 371 Figure: Figure. 1. Chemical structures of the analytes (palbociclib and ribociclib) and their IS 372 (palbociclib $^2\text{H}_8$ and ribociclib $^2\text{H}_6$) with their retention time (RT) and peak intensity. The 373 table specifies the precursor (Q1) and fragment (Q3) ions selected for each compound of 374 interest. The HPLC-MS/MS conditions are described in section 2.6. 375 376 Figure 2: Palbociclib plasma exposure at day 15 (n=18) in the whole cohort (n=18) and in the two subgroup with (n=7) and without (n=11) potential DDI. Black crosses represent the 377 378 subpopulation arithmetic mean values and open circles represent individual patient values.