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Copper (II)-based halogen -substituted chromone antitumor drug entities: Studying biomolecular interactions with ct-DNA mediated by sigma hole formation and cytotoxicity activity.

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Abstract

Copper-based antitumor drug entities **1-3** derived from substituted (F⁻, Br⁻, -CH₃) 3-formylchromone pharmacophore were synthesized and thoroughly characterized by spectroscopic and single X-ray crystallographic studies. These complexes show structural novelty due to presence of the X-bonds in chromone scaffold which could facilitate higher propensity for nucleic acids via sigma σ -hole interactions. Therefore, structure-activity relationship of **1-3** was studied by performing ct-DNA binding, pBR322 cleavage and cytotoxicity activity to validate their potential to act as chemotherapeutic drug entities. The binding studies of **1-3** with ct- DNA were carried out employing many biophysical techniques and the corroborative results of these experiments showed intercalation mode of binding and the order of binding was found to be **2** > **1** > **3**. The structure of drug entities could facilitated strong halogen bonding interaction (in case of **1** & **2**) and stability of X bond was rationalized by sigma hole region of positive electrostatic potential on the surface of C-X covalent bond, as determined by gas phase B3LYP computational DFT studies. Interestingly, **2** exhibited most avid binding affinity due to presence of Br⁻ electron withdrawing and polarizable group. Further, cleavage studies of **1-3** with pBR322 plasmid DNA were performed which demonstrated significant cleavage activity, the supercoiled form (Form

I) of plasmid DNA was converted to nicked form (Form II) with the appearance of linearized form (Form III) in between two, implicating lethal double strand breaks of DNA. **2** showed predominantly higher cleavage activity following the similar trend as observed for binding studies. The cytotoxicity of the complexes **1-3** was evaluated by MTT assay against the human liver carcinoma (Huh-7) and prostate cancer (DU-145) cell lines; complex **2** exhibited specific and selective cytotoxicity for the DU-145 cancer cell line with LC_{50} value of $1.6\mu M$.

Introduction

Copper is an endogenously biocompatible trace element which plays a key role in many vital enzymes in biological system [1]. Copper complexes exhibit interesting chemical and biological properties viz., strong Lewis acid nature, redox active behavior with one electron $Cu(II)/Cu(I)$ shuttle, high binding propensity for DNA, DNA cleaving ability *via* an oxidative pathway, antitumor and apoptosis inducing character [2-5]. Copper complexes have emerged as potent class of future generation anticancer agents, primarily due to their differential response towards cancerous and normal cells. Copper was found to be a co-factor for several angiogenesis mediators including vascular endothelial growth factor (VEGF) [6], basic fibroblast growth factor (bFGF) [7], interleukin-1 (IL-1) and IL-8 [8]. Tumors are dependent on angiogenesis for their growth, invasion and metastasis [9], since tumor cells are in a state of hypoxia due to altered metabolism and rapid growth. Furthermore, copper content is usually much higher in cancerous cells as compared to normal cells. Many copper complexes are also known to induce oxidative stress by the production of ROS or the depletion of GSH due to cellular accumulation of the redox active copper ions [10].

Chromones- benzoannelated γ -pyrone heterocycles are considered as versatile privileged scaffolds in medicinal chemistry owing to their diverse pharmacological applications [11, 12]. They are

excellent templates for the design of novel pharmacologically important compounds due to ease of substitution and structure modification. Therefore, many substituted derivatives of 3-formyl chromones have been synthesized and explored for medicinal properties for treating chronic neurodegenerative, inflammatory, and infectious diseases as well as diabetes and cancer [13-15]. Chromone based compounds have been incorporated in cancer- targeted therapeutic drug molecules which were capable of inhibiting different types of cancer targets, viz., TNF α -signaling and protein kinases [16]. Copper (II) chromone complexes have shown cytotoxicity against different phenotypes of cancer cell lines with varied potencies [17-19]. The cytotoxic activity of Cu(II) chromone Schiff base complexes on cell proliferation of chemoresistant lung cancer (A549) and breast (MCF-7) cancer cell line was evaluated and the complexes were capable of effectively overcoming cisplatin resistance in these cell lines which was attributed to chelation of the chromone ligand with metal ion [20]. In our laboratory, the cytotoxic potential of chromone-Cu(II) drug candidates was evaluated on human breast (MCF-7) and liver cancer (HepG2) cell lines which revealed good cytotoxic response at significantly lower IC_{50} values of (5-10 $\mu\text{g mL}^{-1}$) [21].

Recently, halogen bonding has fueled much interest in the subdomain area of small molecule-nucleic acid interactions in medicinal inorganic chemistry [22]. Halogen bonding is a type of charge transfer interaction which is a qualitative analogue of hydrogen bonding [23, 24]. The halogen bonds take place between a halide (X) in one molecule with neighboring electron-rich Lewis base site, similar to a hydrogen bonds. DNA is most amenable therapeutic drug target for many antitumor drugs in clinical use which possesses several easily accessible electron rich reactive sites such as nucleobase pairs and ribose sugar- phosphate backbone of double helix. Interaction of small molecules with X-bond with DNA double helix could be interpreted on the

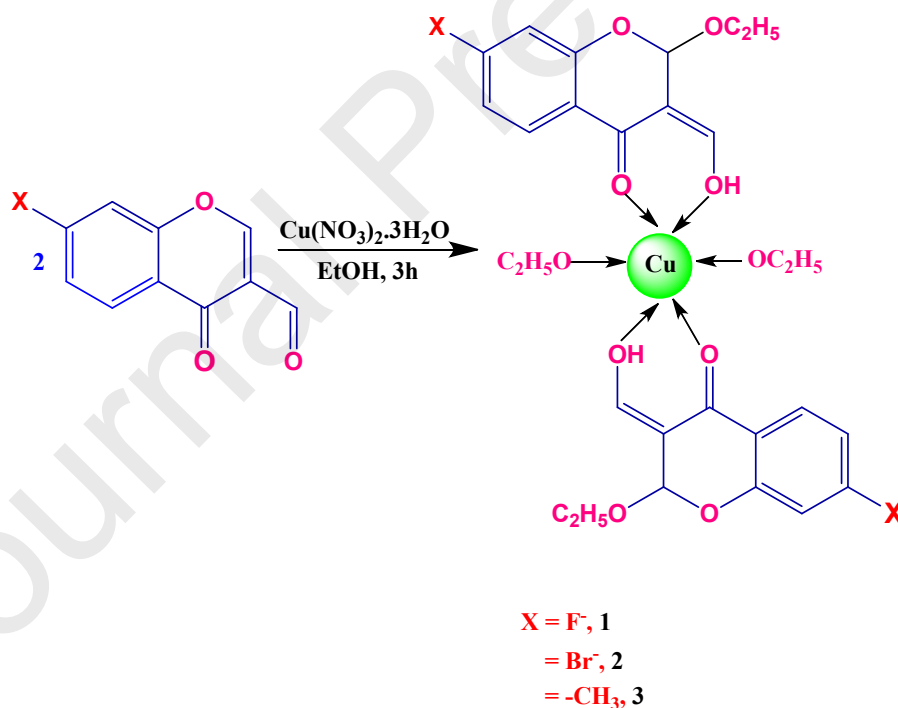
basis of halogen bonding, as formation of the halogen bond is thermodynamically more favorable and can be thus, be used to predict non-covalent interactions such as electrostatic interactions, groove binding and intercalation. Halogens in organic and inorganic compounds are known to polarize their covalent bonds to generate electropositive crown where the halogen acts as a Lewis acid to pair with Lewis bases, including oxygen and nitrogen atoms of biomacromolecules. These electrostatic pairs known as halogen bonds or X-bonds could contribute immensely to the understanding of drug-nucleic acid interactions [25]

We describe herein, new copper (II) based antitumor drug candidates **1-3** derived from substituted chromone scaffold where X= F⁻, Br⁻, and -CH₃. The substitution of halogens and alkyls in chromone scaffold drastically alters the physiochemical and pharmacological properties of drug entities in following ways: i) They can affect the polarity and lipophilicity which enable the facile transportation/permeation via the cell membrane into the cells thus preventing off-target toxicity ii) prolong the lifetime of active metabolite iii) show cytotoxic response at low submicromolar dosage iv) inhibit cell proliferation by targeting TNF α -signaling and protein kinases. Besides, copper(II) embedded in bioactive chromone pharmacophore also plays a significant role by inducing ROS oxidative stress and mediating angiogenesis. We have evaluated the DNA binding affinity of Cu(II) chromone complexes **1-3** by a battery of biophysical methods. Since these complexes possess the X-bonds which could facilitate sigma σ -hole interactions, it was considered worthwhile to study their *in vitro* differential response with ct- DNA and cytotoxicity in cancerous cells. The present Cu(II) complexes- DNA model system demonstrated that among all X-bonds, bromine substituted chromone Cu(II) complex **2** exhibited avid binding propensity with DNA and exhibited selective cytotoxicity for the DU-145 cancer cell line at a low intracellular concentration.

Results and discussion

Synthesis and characterization

The copper (II) complexes **1–3** derived from substituted (F-, Br-, -CH₃) 3-formylchromone were synthesized and thoroughly characterized by elemental analysis and various other spectroscopic techniques *viz.*, UV-vis, FT-IR, EPR and ESI-MS (Scheme 1). The molecular structure of complexes **1–3** was confirmed by single X-ray diffraction studies. The synthesis of the complexes involved formation of an *in situ* intermediate (3-hydroxymethylene-2-ethoxychroman-4-one) via ethoxylation at the 2- position of the chromone derivatives (F-, Br-, -CH₃). The synthesized complexes were air-stable and soluble in organic solvents such as EtOH, DCM, acetone, DMF and DMSO.



Scheme 1 Synthetic route for complexes **1**, **2** and **3**

Spectroscopic characterization

In the IR spectra of complexes **1–3**, the characteristic bands assigned to $\nu(\text{HC=O})$ and $\nu(\text{-C=O})$ of 3-formylchromone derivatives observed at 1694 and 1671 cm^{-1} , respectively were shifted to lower wave number by 15-25 cm^{-1} indicating the complexation of these groups with metal ions (Fig. S1) [26]. The bands which appeared at 592 cm^{-1} and 1263 cm^{-1} were assigned to $\nu(\text{C-Br})$ and $\nu(\text{C-F})$ stretching vibrations, respectively. In complex **3**, two characteristic bands at 2907 cm^{-1} and 3069 cm^{-1} corresponded to sp^2 and sp^3 hybridized carbon atoms, respectively. The band at 1060 cm^{-1} was attributed to $\nu(\text{C-O})$ stretch, indicating the ethoxylation of the 3-formylchromone derivative at 2-position. In the far-IR of complexes **1–3**, the metal coordination was substantiated by the bands at 426-500 cm^{-1} .

The solid state X-band EPR spectra of complexes **1–3** were recorded at room temperature on 9.1 GHz frequency under the magnetic field strength of 3000 G (Fig. S2). The EPR spectra of **1–3** displayed the anisotropic band centered at g values as shown in Table S4. These values are in agreement with tetragonally elongated octahedral geometry of Cu(II) ion with d^9 configuration [27] i.e. $(e_g)^4 \cdot (a_{1g})^2 \cdot (b_{2g})^2 \cdot (b_{1g})^1$. The trend of g -tensor values is $g_{\parallel} > g_{\perp} > 2.0023$ suggested that unpaired electron mainly resides in dx^2-y^2 orbital. Furthermore, the complexes possess $g_{\parallel} < 2.3$, revealing an appreciable metal-ligand covalent character of the complexes [28, 29].

Electronic spectra of complexes **1–3** were recorded in DMSO at room temperature which revealed two well resolved absorption maxima in the range 260 to 290 nm and 300-315 nm attributed to π - π^* and n - π^* electronic transitions, respectively. Broad d-d bands observed in the visible part of spectra in the range 400-642 nm are typical for tetragonally distorted octahedral environment around the Cu(II) center [30].

Description of the X-ray crystal structures

The complex **1** crystallizes in the monoclinic space group $P21/n$ possessing the lattice parameters $a = 9.9291(4) \text{ \AA}$, $b = 5.1435(2) \text{ \AA}$, $c = 27.7256(10) \text{ \AA}$ and $\alpha = \gamma = 90^\circ$, $\beta = 98.1990(10)^\circ$. The complex shows slight distorted octahedral geometry CuO_6 coordination environment with ligation from four carbonyl oxygen [$\text{Cu-O}=1.9325(12)$] of fluoro substituted chromone ligand scaffold and two oxygen [$\text{Cu-O}=1.958(13)$] from solvate ethanol moiety. The bond lengths of Cu-O at axial positions were slightly larger due to static Jahn-Teller distortion. Further extension of the crystal structure occurs through π - π stacking interactions, resulting in the 3D supramolecular architecture as shown in Fig. 1.

The ORTEP view of the complex **2** is shown in Fig. 1. The O-O donor bromine substituted formylchromone ligand scaffold binds to the Cu (II) ion to form 6-membered chelate rings. Due to static Jahn-Teller distortion, there was considerable variations in equatorial and axial bond lengths. The complex crystallized in the triclinic system in $P-1$ space group. The optimized lattice parameters are $a = 5.1134(5) \text{ \AA}$, $b = 9.7980(9) \text{ \AA}$, $c = 14.9650(14) \text{ \AA}$ and $\alpha = 85.415(3)^\circ$, $\beta = 83.430(3)^\circ$, $\gamma = 89.988(4)^\circ$.

Single crystal X-ray study of complex **3** revealed that it crystallized in the triclinic $P-1$ space group possessing the lattice parameters, $a = 5.1349(4) \text{ \AA}$, $b = 11.6066(9) \text{ \AA}$, $c = 12.9630(11) \text{ \AA}$ and $\alpha = 105.733(3)^\circ$, $\beta = 97.741(3)^\circ$, $\gamma = 90.384(3)^\circ$ per unit cell. An ellipsoid view of complex **3** along with atom numbering scheme is depicted in fig. 1. The structure of complex **3** represent a centrosymmetric Cu(II) ion coordinated to a methyl substituted chromone ligand scaffold through O,O-atoms with a solvate ethoxy group at 2-position along with two ethanol moieties at the axial position. All the Cu–O bond distances in complexes **1-3** were observed within the range reported for other Cu (II) octahedral complexes [31].

Other crystallographic data *viz.*, selected bond lengths and bond angles for **1-3** are summarized in Table S1-S3.

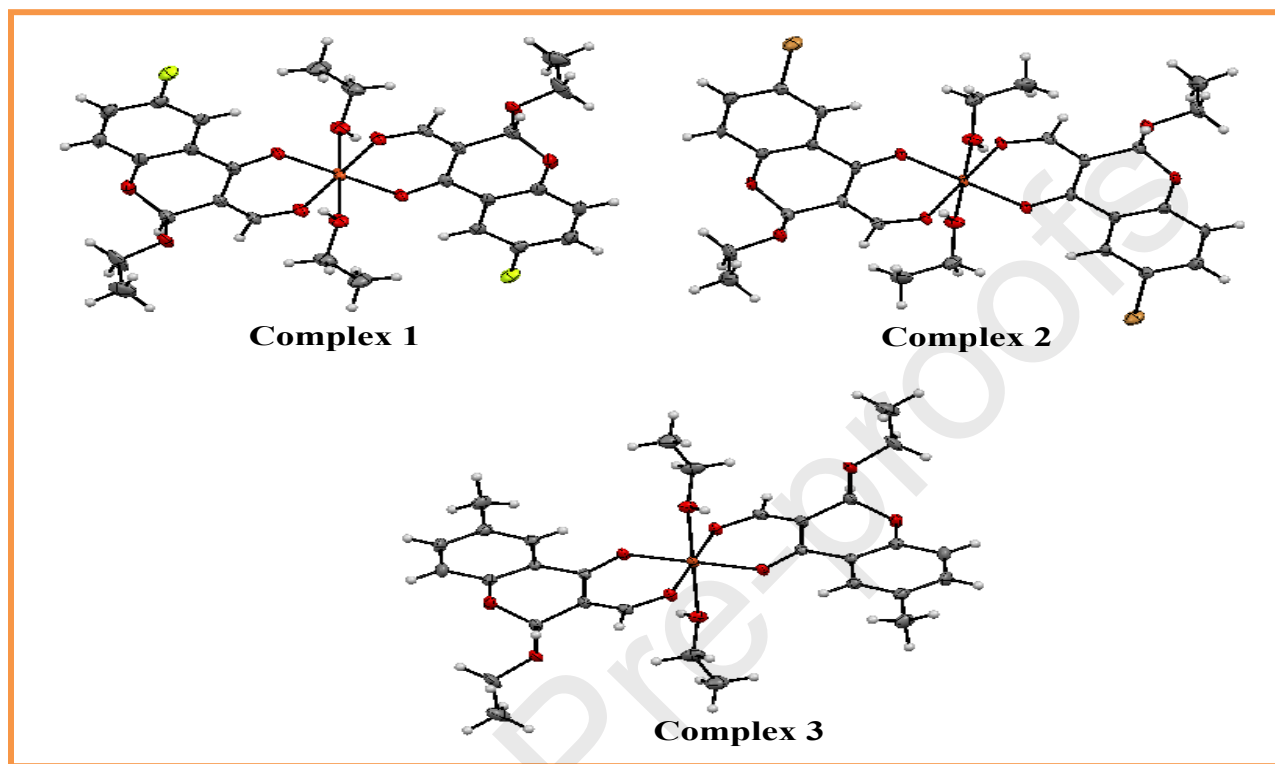


Fig. 1. Single crystal X - ray structure of complexes **1 – 3** (50 % probability ellipsoids).

DNA binding studies

Metal-based drug candidates can uncoil the helical structure of ct-DNA, undrape the embedded electron rich nucleobases to the exterior helix leading to an efficacious binding with the drug entities either in covalent or non- covalent binding mode *viz.*, electrostatic, groove binding and intercalation [29, 32]. The binding of copper(II)-based halogen substituted-chromone derivatives **1-3** with ct-DNA was followed by a battery of different complementary techniques such as UV-vis absorption, florescence spectroscopy, EPR spectroscopy, circular dichorism and cyclic voltammetry.

Electronic absorption titration of complexes **1-3** was carried out in presence and in the absence of ct-DNA at room temperature (Fig. 2). Upon addition of increasing concentration ($0.00-3 \times 10^{-5}$ M) of ct-DNA to a fixed concentration (2×10^{-4} M) of **1-3**, a sequential decrease of the absorption intensity was observed at the intra-ligand band at 250-300 nm exhibiting “hypochromism” of 43% in case of **1**, 54% for **2** and 41% for **3**, respectively. There was a pronounced red shift of 7-8 nm in complexes **1** and **2**, suggesting intercalative mode of binding while a moderate shift of 1-2 nm was observed in case of **3** which implicated groove binding mode [33]. The rationale for intercalation mode of binding in complexes **1** & **2** is presence of aromatic π - π stacking chromophore which could easily “slip in” or insert between the nucleobases while **3** articulates groove binding due to the non-planarity of the methyl groups, therefore prefers to bind by electrostatic interaction to the negatively charged phosphate oxygen of DNA helix. The binding strengths of **1-3** with ct-DNA was quantified by the intrinsic binding constant K_b , using the Wolfe-Shimer equation (1) [34].

$$[\text{DNA}]/\epsilon_a - \epsilon_f = [\text{DNA}]/\epsilon_b - \epsilon_f + 1/K_b (\epsilon_b - \epsilon_f) \quad (1)$$

Where [DNA] is the ct-DNA concentration and ϵ_a , ϵ_f , and ϵ_b are apparent ($A_{\text{abs}}/[\text{Cu (II) complex}]$), free and bound complex molar absorptivity coefficients, respectively. From the plot of $[\text{DNA}]/\epsilon_a - \epsilon_f]$ vs [DNA] with a slope of $1/(\epsilon_b - \epsilon_f)$ and intercept of $1/[K_b (\epsilon_a - \epsilon_f)]$, the values of K_b were derived by the ratio of slope to intercept.

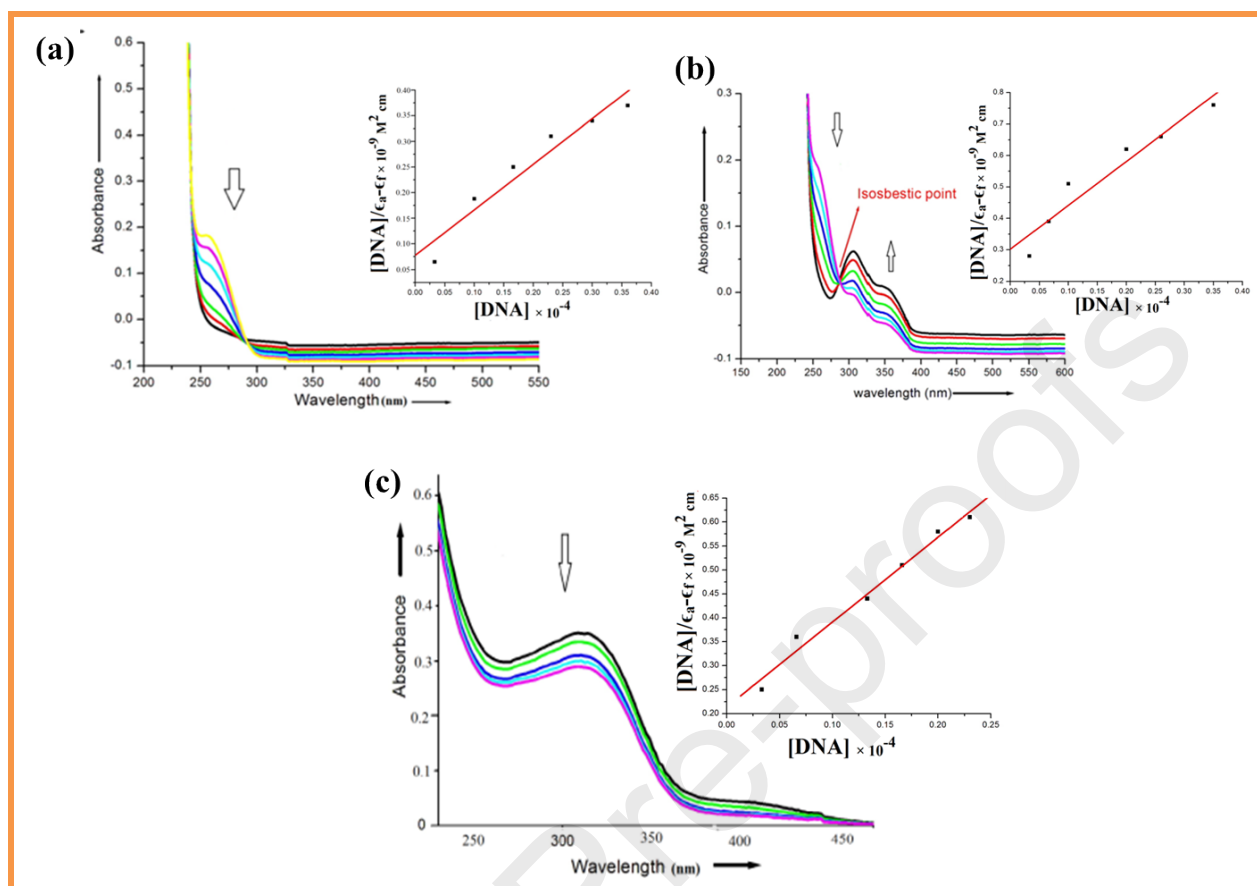


Fig. 2. Absorption spectra of complexes **1** (a), **2** (b) and **3** (c) in absence and presence of increasing concentrations of ct-DNA in Tris – HCl buffer (pH=7.3). Inset: plots of $[DNA]/(\epsilon_a - \epsilon_f) \times 10^{-9} \text{ M}^2 \text{ cm}$ vs $[DNA]$ for the absorption titration with **1-3**. The arrows specify the change (increase/decrease) in intensity of absorbance with increasing ct-DNA.

Table 2 The binding constant values for complexes **1-3** obtained upon interaction with ct-DNA.

Complex		Absorption spectra, $K_b (\times 10^4 \text{ M}^{-1})$	Emission spectra	
			$K (\times 10^4 \text{ M}^{-1})$	$K_{sv} (\times 10^4 \text{ M}^{-1})$
1	ct-DNA	6.46 (± 0.3)	6.97(± 0.05)	5.04(± 0.03)
2	ct-DNA	7.58 (± 0.1)	7.23(± 0.02)	6.13(± 0.05)
3	ct-DNA	4.33 (± 0.4)	4.93(± 0.07)	4.19(± 0.01)

The intrinsic binding constant, K_b values of the complexes **1-3** with ct-DNA are depicted in Table 2 and were found to be in the order of magnitude $2 > 1 > 3$. Interestingly, the K_b values of **1** & **2** were found to be two-fold higher in magnitude as compared to complex **3** which has been attributed to the presence of halogens on their ligand scaffolds as halogen substituents provide hydrophobic, polar and planar environment suitable for intercalation with ct-DNA [35]. Notably, bromine substituted complex **2** revealed highest binding affinity towards DNA, which can be envisaged by a model that shows that the electron density from DNA nucleobases could be directly transferred to polarizable C-Br bond of ligand scaffold *via* the sigma σ -hole (Fig. 3) with further “proof of concept” obtained by docking and computational B3LYP /DFT studies [36]. Albeit no such observation was seen in case of fluorine-substituted moiety of **1** while the complex **3** exhibited lowest K_b value with ct-DNA which can be explained by the lack of planarity and steric hindrance due to methyl group present on ligand moiety.

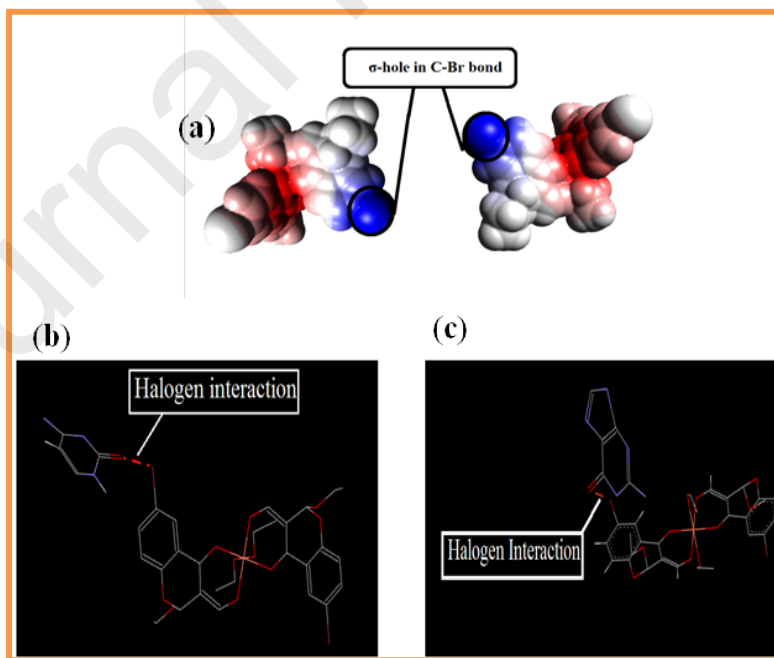


Fig. 3. Electrostatic potential surface depicting σ -hole in blue colour (a) in complex **2**. Selected halogen (Br) interactions of complex **2** with nucleobases, cytosine (a) and guanine (b). Hydrogen atoms have been omitted for clarity.

Similarly, binding constant (K) values of **1-3** were obtained from the Scatchard equation [37] with ct-DNA and the binding propensity further substantiated that complex **2** exhibited remarkable binding with ct-DNA as compared to complex **1** and **3** (Table 2).

The validation of binding mode of **1-3** was evaluated by performing competitive binding studies with ethidium bromide, EB. Ethidium bromide is a typical intercalator of ct-DNA possessing planar phenanthridine ring by which it intercalates in between adjacent base pairs of the ct-DNA double helix, thus emitting intense fluorescence at *ca.* 600 nm [38]. The enhanced fluorescence emission has been attributed to the formation EB-ct-DNA complex and may be quenched with the co-existence of motifs that intercalates ct-DNA equally or strongly than ethidium bromide. Upon addition of **1-3** to EB-ct-DNA system such that [complex]/[ct-DNA]=1:1 to 5:1, the significant quenching of the band at 600 nm was observed (Fig. 4), resulting fluorescence emission was 32% - 33% of the initial EB-ct-DNA fluorescence intensity in case of complex **1** & **2** while it was 50% for **3**. These experimental results demonstrate that **1** & **2** displaced EB from EB- ct-DNA system more efficiently than **3**, thus implicating their non-covalent intercalative mode of binding with ct-DNA and groove binding interaction in case of **3**. In an another fluorescence quenching experiment, Stern - Volmer constant, K_{sv} values for **1-3** were calculated (equation 2) which depicted quenching of fluorescence, in the order **2** > **1** > **3** well corroborated with other binding studies ascertaining strong affinity of complexes **1** and **2** for ct-DNA as compared to **3**. The calculated Stern-Volmer constant, K_{sv} values for **1**, **2** and **3** were found to be 5.04, 6.13 and 4.19, respectively.

$$I_0/I = 1 + K_{sv} \cdot r \quad (2)$$

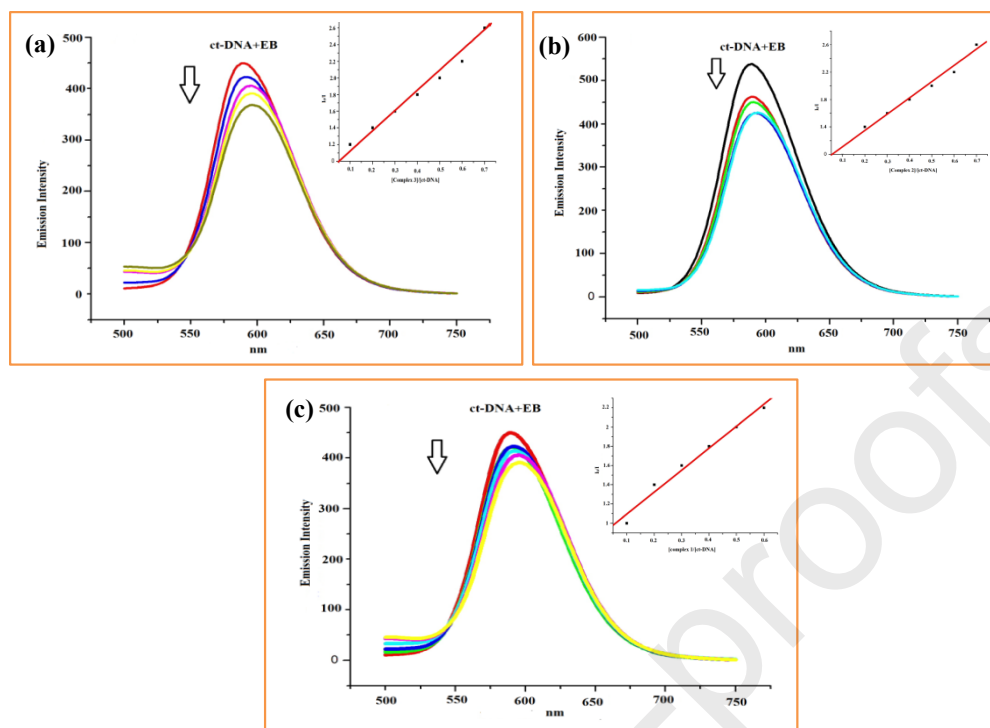


Fig. 4 Emission spectra of EB-ct-DNA in the absence and presence of complexes **1** (a), **2** (b) and **3** (c) in Tris-HCl buffer at pH 7.3. Arrow indicates change in intensity with increasing concentration of complexes **1–3**.

The liquid state X-band EPR spectra of complexes **1–3** were recorded in DMSO at LNT in the absence and presence of ct-DNA which revealed two g_{\parallel} and two g_{\perp} signals (Fig. 5). Upon addition ct-DNA, EPR spectra of complex **2** exhibited weak signatures with minor shift in g -values, indicating the binding of complex **2** with ct-DNA [39,40]. However, the spectra of complex-ct-DNA showed that the coordination number of the metal in **1–3** was not perturbed. Similar intensity pattern was observed in the complex- ct-DNA spectra of **1** and **3** (Fig. S3).

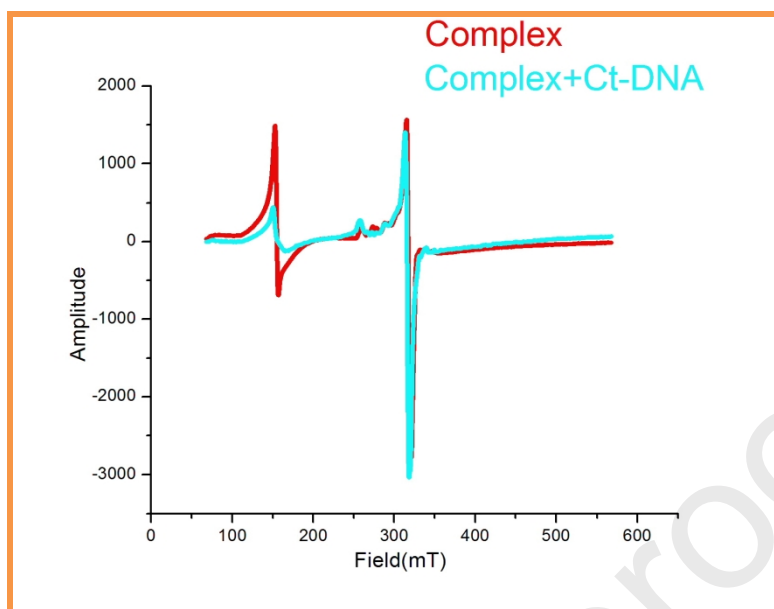


Fig. 5. X-band EPR (LNT) spectra of complex **2** (red), complex **2** + ct-DNA (blue).

Cyclic voltammetric studies of **1** & **2** in Tris-HCl buffer solution at a scan rate of 0.2 volts per second showed quasi-reversible one electron shuttle with Cu(II)/Cu(I) redox pairs and $E_{1/2} = 0.413$ volts and $\Delta E_p = 0.193$ volts in case of **1** and $E_{1/2} = 0.442$ volts and $\Delta E_p = 0.199$ volts in case of **2**, respectively. The ratio of anodic to cathodic (I_a/I_c) peak current was found to be 0.49. Upon interaction of **1** & **2** with ct-DNA, there was a positive potential shift in both $\Delta E_p = 0.199$ volts and 0.213 volts; $E_{1/2} = 0.461$ and 0.471 volts in both **1** and **2**, respectively demonstrating that **1** & **2** exhibited strong binding with ct-DNA (Fig.6). In the cyclic voltammogram of **1** & **2**, a substantial decrease in current was observed possibly due to the slow diffusion of ct-DNA-Cu complex motif. The anodic to cathodic peak current (I_a/I_c) of **1** & **2** also experienced a positive shift i.e., 0.53 and 0.56, respectively. The greater shift of **2** as compared to **1** in the magnitude of ΔE_p , $E_{1/2}$ and (I_a/I_c) suggests its strong binding affinity with ct-DNA [41].

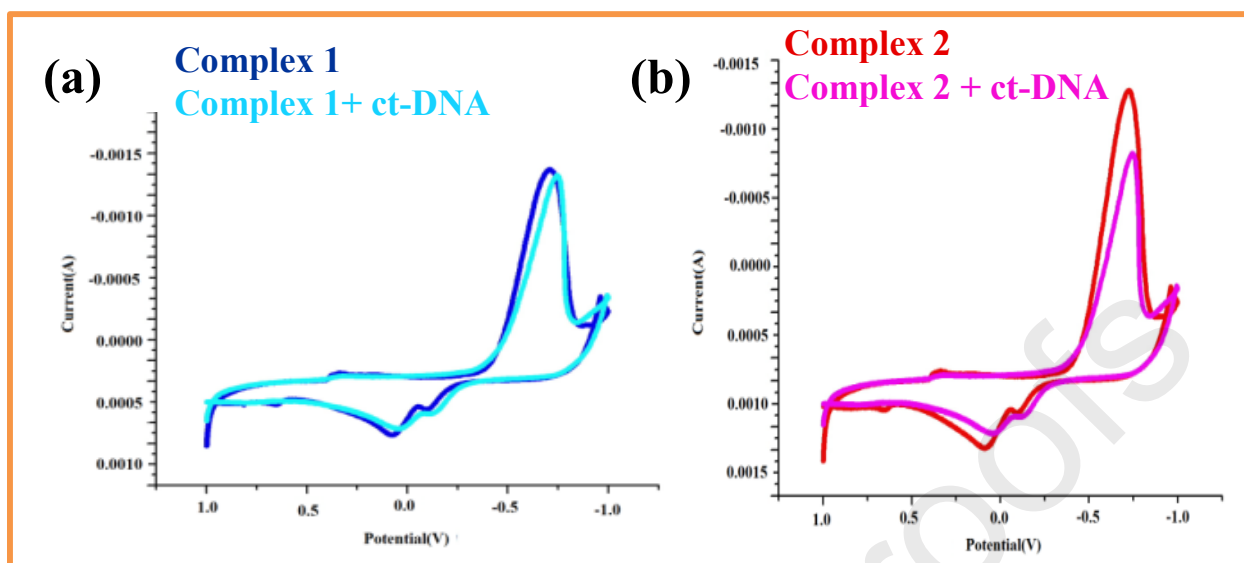


Fig. 6. Cyclic voltammograms (1 : 2 DMSO : Tris-HCl buffer solution, at 25 °C) of **1** and **2**

Circular dichroism is an efficient and powerful optical technique used for the study of chiral biopolymers to analyse the stereochemical and conformational changes upon interaction with small molecules/metallo-drugs. In the CD spectrum of the characteristic right-handed B-form DNA, a positive peak at 276nm is assigned to base stacking and a negative peak at 246nm corresponds to the helicity of B-form DNA [42, 43]. The CD spectra of complexes **1-3** in presence of ct-DNA are depicted in Fig. 7. The less perturbations in the positive and negative bands upon addition of complex **3** are indicative of groove or electrostatic mode of binding whereas the enhancement of both positive and negative signatures with a slight shift in wavelength upon addition of **1** & **2** validate their intercalative mode of binding.

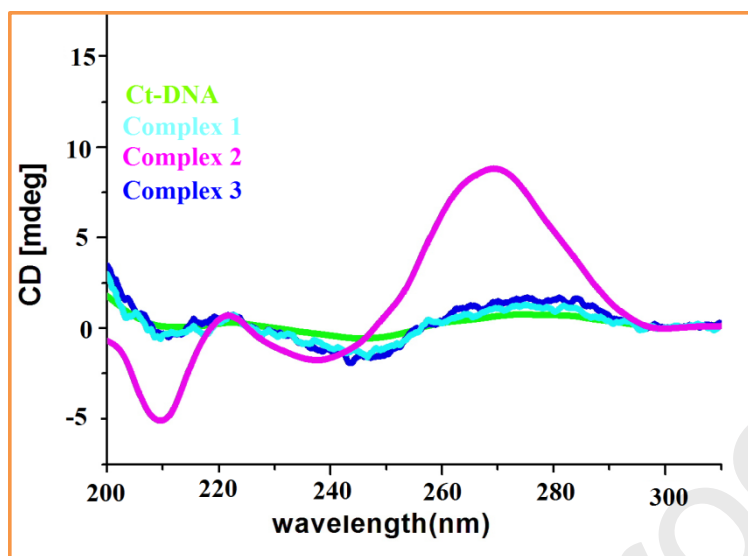


Fig. 7. CD spectra of complexes 1-3

Morphological studies

Due to the presence of various potential amenable binding sites in DNA viz., the phosphate group and electron donor atoms on nucleobases, metal ions carry out the dramatic condensation of DNA to a compact, rod like and toroidal structure, most commonly observed by electron microscopy. The condensation process has been attributed to diverse ionic interactions such as electrostatic and other forces viz., free energy of bending, mixing entropy, coulombic etc. [44]. The condensation or aggregation phenomena plays a critical role in many antitumor drugs. DNA condensation has become a dynamic useful technique as it can be used to transfer DNA containing genes of therapeutic interest from solution to target cells involving gene therapy [45, 46]. Since DNA possess negative charge density due to nucleobases present in the major and minor grooves, positively charged metal ions can easily intercalate within these nucleobases, thus showing molecular recognition and in vivo modification of DNA in the condensed state. In this study, condensation of ct-DNA was achieved to examine the morphological changes under scanning electron microscope (SEM) by evaporating an equimolar mixture of 1-3 in Tris-HCl buffer/ 50mM

NaCl at pH=7.3. The SEM micrographs of 1-3 showed single crystalline motifs in rod shapes (Fig. 8). On the contrary, the SEM micrographs of condensates of 1-3 with ct-DNA revealed the randomization and formation of globular structures, thus indicating substantial interaction of ct-DNA with complexes similar to earlier reports on copper-DNA condensates [47, 48].

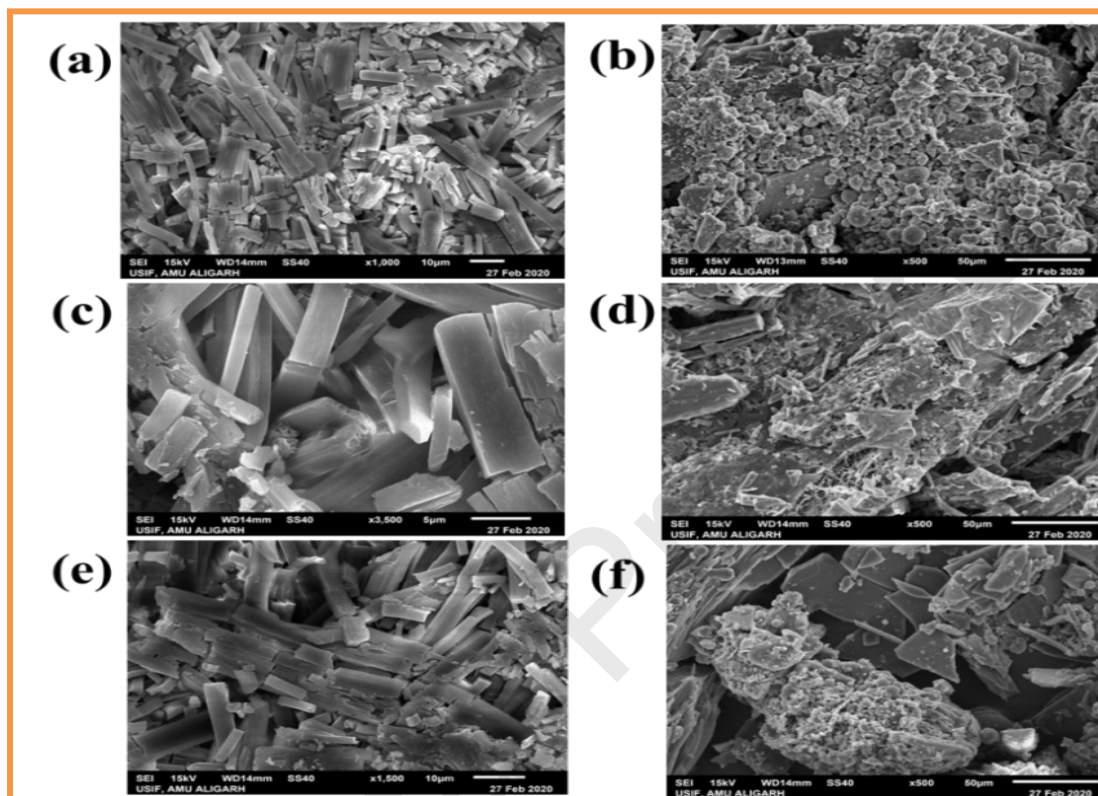


Fig. 8. SEM images of **1** (a), **2** (c), **3** (e) showing surface morphology upon condensation with ct-DNA (b, d & f, respectively).

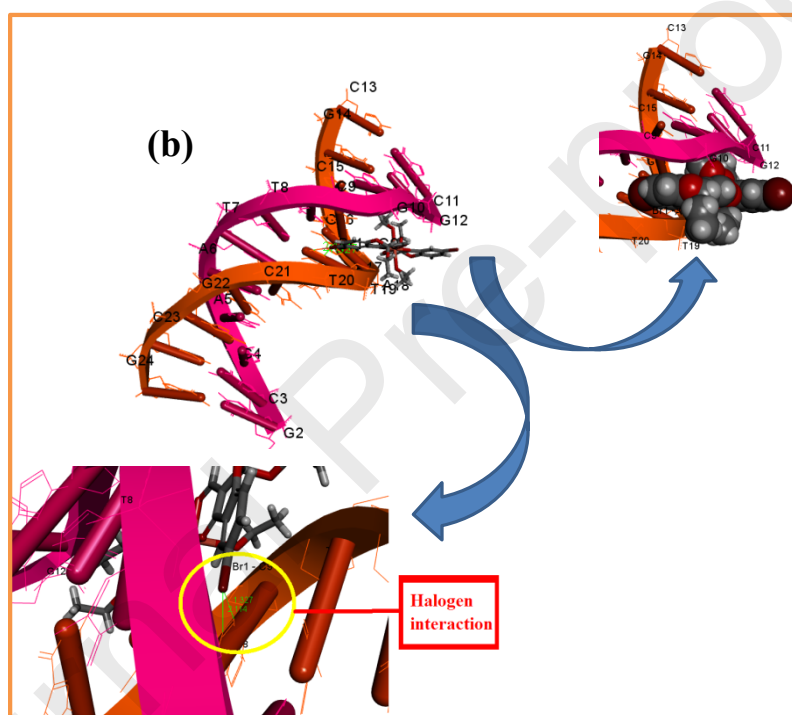
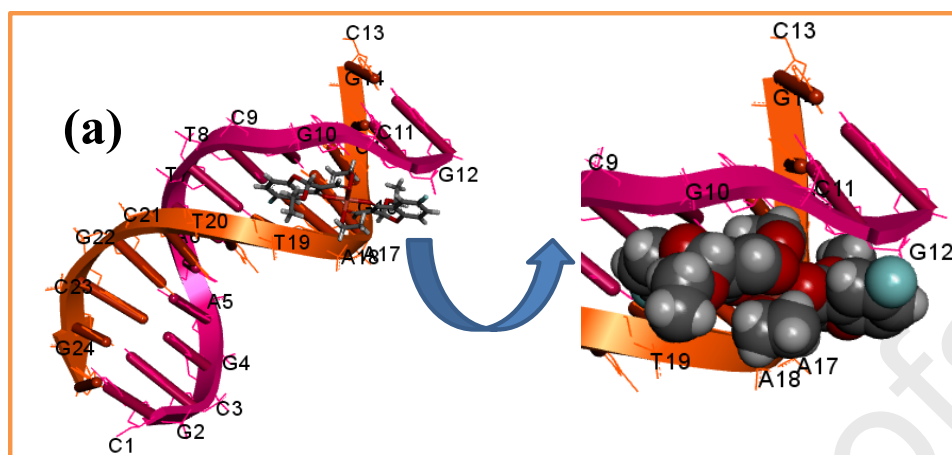
Molecular docking studies

The molecular docking study of **1-3** was carried out with the d(CGCGAATTCGCG)₂ (PDF ID : 1BNA) dodecamer sequence of DNA helix [49] to estimate the most probable binding affinity along with the sterically stable conformations (Fig.9). The binding energies for the best docked pose of complexes were quantified as -333.98 kJ/mole for **1**, -347.07 kJ/mole and -294.72 KJ/mole for **2** and **3**, respectively which revealed efficient DNA-drug interaction with the notably

higher binding energy of complex **2** than the rest of complexes. Moreover, the results of docking implicated that complexes **1-3** fit snugly in the intercalative sites of DNA helix and moderately deep in adenine-cytosine rich region stabilized by hydrogen bonding, van der Waals interactions and hydrophobic contacts of aromatic chromone ring motif with the minor groove of ct-DNA; as the minor groove of DNA is considered to be lucrative target for many small molecules [50].

These interactions are considered to be the main driving force of the intercalative binding mode in addition to electrostatic interactions between the polarized bromine and the lone pair electrons of the nitrogen and oxygen atom of a neighboring nucleobases of DNA helix.

The greater binding energy of complex **2** was particularly supported by halogen donation *via* σ -hole of C-Br covalent bond. This highly directional attraction between halogen atom (bromine in our case) and an electron donor nucleobase (adenine) helps in modulating the physicochemical properties of the molecular scaffolds; they affect the polarity and lipophilicity, which in turn could change membrane and blood brain barrier permeation of the molecule. Furthermore, the difficulty in metabolization of carbon-halogen, specifically C-Br bond of **2** could prolong the lifetime of active motif [51]. The electron donor nucleobase to halogen interaction in complex **2** was found to be nitrogen atom of adenine wherein nitrogen atoms are seen to transfer its electron density directly into the σ -hole of C-Br covalent bond, thus stabilizing DNA-drug interactions. Also the increasing stabilization was attributed to the aromatic character of nucleobase which allows them to act as electron donors via their π -electrons above and below their rings.



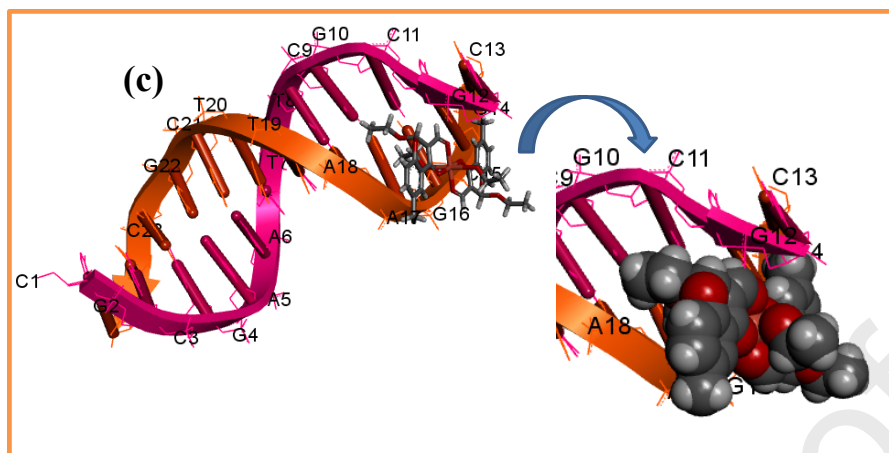


Fig. 9 Molecular docked model of complex **1** (a) complex **2** (b) and complex **3** (c) fitted into the G-C region of the minor groove of the ct-DNA dodecamer duplex of sequence d(CGCGAATTCGCG)₂, PDB ID: 1BNA.

B3LYP/DFT Computational studies

The electrostatic potential mapped structure was obtained to model the σ -hole region on the surface of C-Br covalent bond which was shown by the emergence of electropositive region (Fig. 3a; blue) along the C-Br axis in complex **2**, consistent with the presence of more polarizable Br⁻ atom. This region is surrounded by an electroneutral ring (Fig.3a; white) and farther out, an electronegative region (Fig.3a; red). The presence of this σ -hole region indicates that complex is **2** more likely to serve as halogen bond donor to nucleobases *via.*, its Br⁻ atom, hence giving an indication of the specific and enhanced binding affinity of **2** as compared to **1** and **3** with ct-DNA.

Moreover, B3LYP/DFT calculations were performed on **1-3** to get an insight onto the frontier molecular orbitals and their energy gap values which help to shed light on the chemical reactivity, kinetic stability and electrical transport properties of a molecule. The HUMO-LUMO energy gap is also used to check bioactivity from intramolecular charge transfer and correlated with various biological aspects like antibacterial, antioxidant and DNA binding aspects [52, 53]. The negative values of HOMO and LUMO indicates the stability of the complexes (Fig. 10) .The higher value

of E_{HOMO} and lower values of E_{LUMO} in **1-3** indicates their electron donating ability and electron accepting ability respectively. The orbital energy level analysis for complexes **1-3** shows that E_{HOMO} values are -5.605 eV, -5.566 eV and -5.337 eV, respectively while E_{LUMO} values are -2.228 eV, -2.262 eV and -1.880 eV, respectively. The lower value of E_{LUMO} in complex **2** defines that it has more electron acceptance ability than **1** and **3**; hence giving another evidence of the presence of σ -hole in C-Br bond which supports the higher binding affinity of complex **2**. Also the calculated ΔE values of **1-3** are 3.337 eV, 2.943 eV and 3.457 eV, respectively. The lower value HOMO-LUMO energy gap in **2** suggested its greater stability as compared to **1** and **3**.

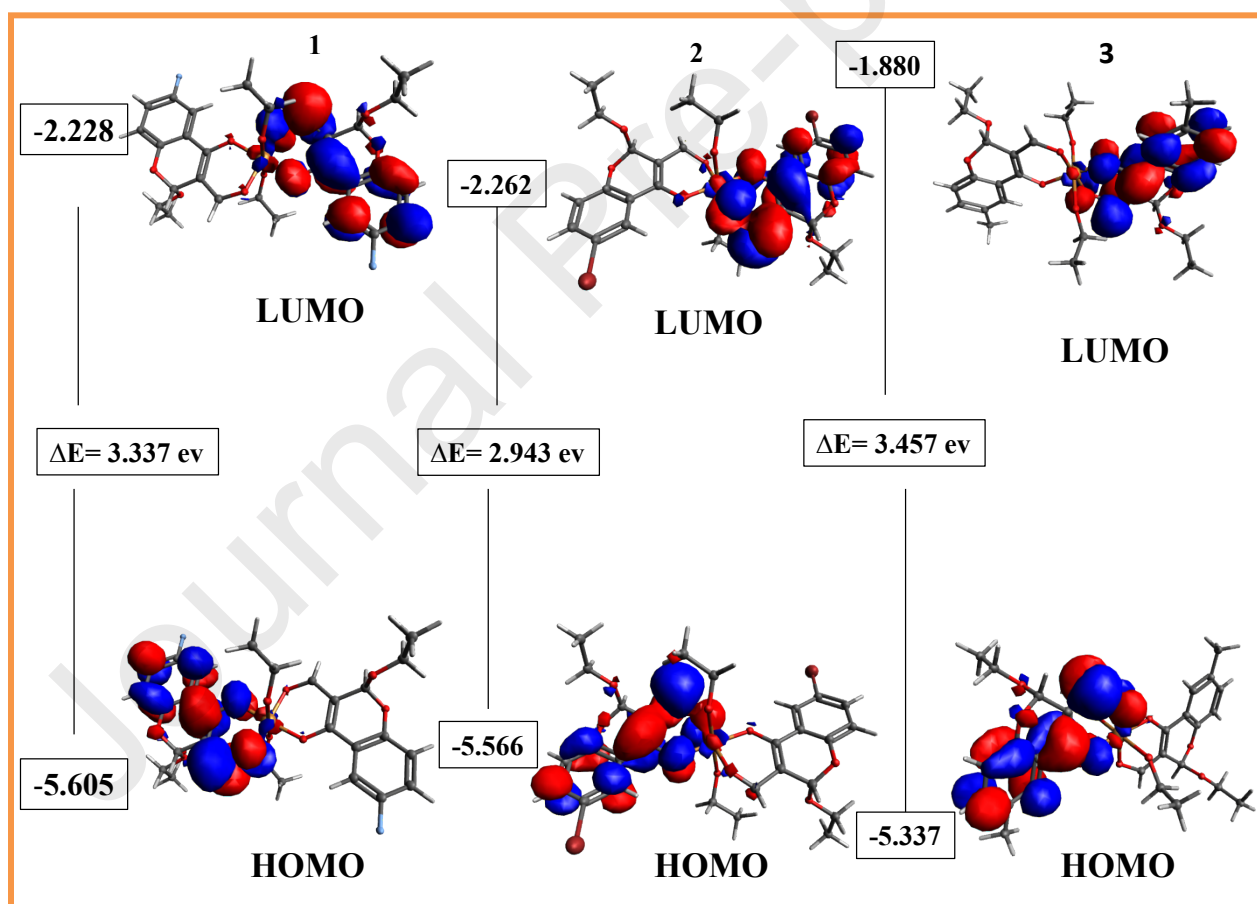


Fig. 10. Frontier molecular orbitals of complexes **1-3** using the B3LYP/DFT method.

***In vitro* cytotoxic activity**

The *in vitro* cytotoxic activity of complexes **1-3** was evaluated in terms of LC_{50} (the lethal concentration of the drug required to kill 50% of the cells) values by colorimetric 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide (MTT) assay [54] on the panel of two cancer lines viz., human liver carcinoma (Huh-7), prostate cancer (DU-145) and one normal prostate epithelial (PNT1) cell lines. Complexes **1-3** (2.5-10 μ M) were treated for 48 hours which resulted in potential dose-dependent growth inhibition of both Huh-7 and DU-145 cancer cell lines irrespective of origin of the cells (Fig. 11), but complex **2** showed higher inhibition than **1** and **3** suggesting that the presence of Br⁻ is lucrative in complex **2**. Interestingly, no significant cell death was observed in the normal PNT1 cell lines upon treatment with complexes **1-3**.

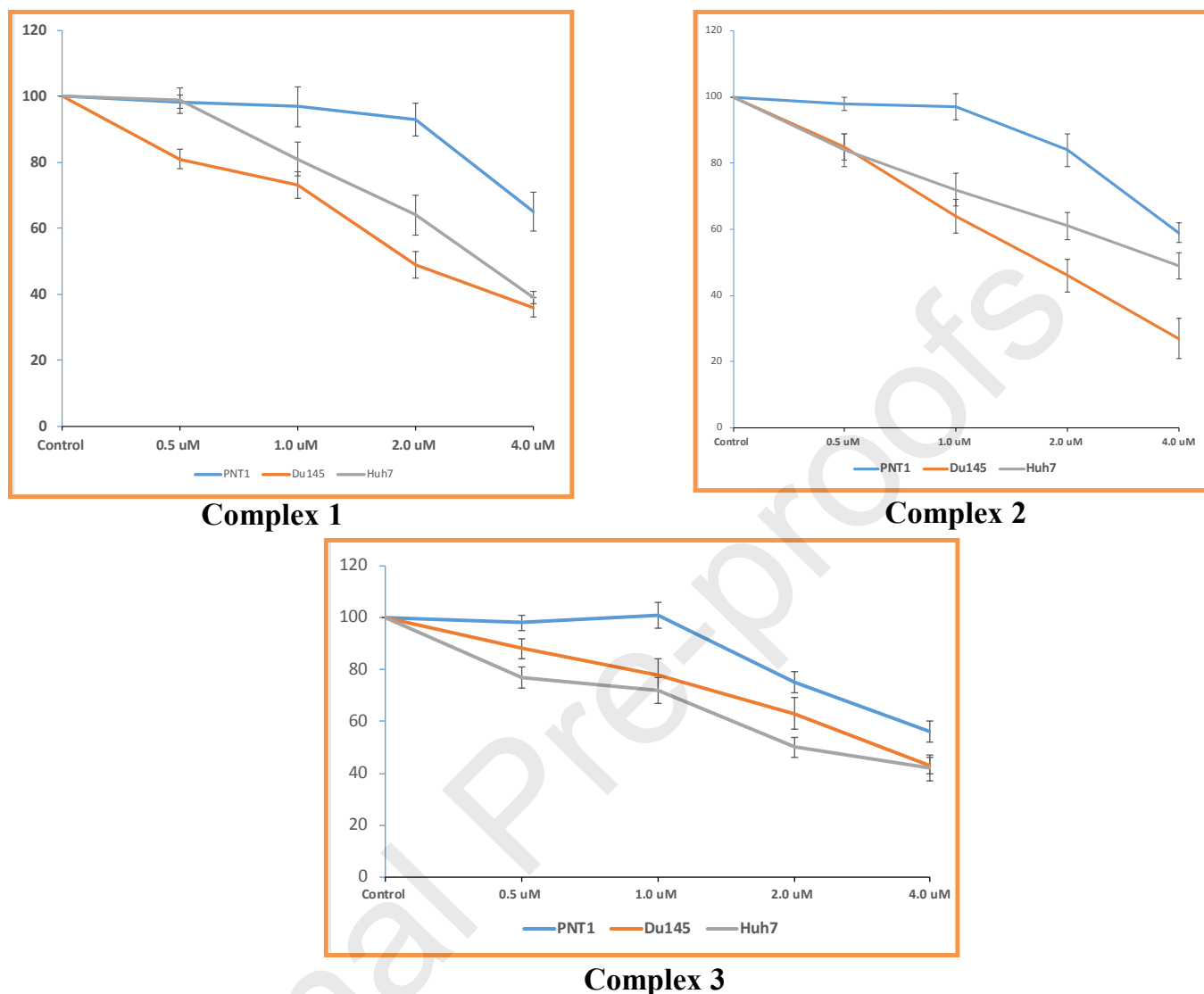


Fig. 11. Survival of tumor cells as a function of different concentrations of **1**, **2** & **3** determined by MTT test, after 48 h incubation.

Cleavage activity

The cleavage reaction of complexes **1-3** was monitored by agarose gel electrophoresis employing supercoiled pBR322 DNA as a substrate in a medium of 5 mM Tris-HCl/50 mM NaCl buffer at pH 7.2 under the influence of applied electric field. Upon gel electrophoresis of pBR322 DNA, the fastest migration results in the conversion of a supercoiled form (Form I) to a relaxed nicked form (Form II) and appearance of linearized form of DNA (Form III) in between the two,

implicating efficient double stranded cleavage of the DNA [54]. Concentration-dependent (5-25 μM) plasmid DNA cleavage experiments of **1-3** depicted the conversion of SC DNA (Form I) to NC DNA (Form II) and upon increasing the concentration of **1-3**, there was the appearance of linearized form (Form III) which suggested double-stranded cleavage of plasmid DNA induced by **1-3**, with the significantly higher cleavage exhibited by complex **2** at 20 μM (Fig. 12 a and S4). Furthermore, the plasmid DNA cleavage activity of **2** was investigated to examine mechanistic pathway in the presence of additives viz., DMSO and EtOH as hydroxyl radical scavengers, NaN_3 as singlet oxygen scavenger ($^1\text{O}_2$) and SOD as superoxide anion radical (Fig. 12 b) . During the reaction, it was ascertained that there was no significant quenching of plasmid DNA cleavage efficiency in presence of DMSO and EtOH, hence excluding the possibility of hydrolytic cleavage pathway, involving diffusible hydroxyl radicals in the cleavage reaction. However, in the presence of SOD and NaN_3 significant cleavage was observed, suggesting the oxidative cleavage pathway of complex **2** and demonstrated that ROS, singlet oxygen and superoxide anion radical could be responsible for cleavage activity [55] The further prediction of cleavage activity of complex **2** was discerned in presence of activators such as H_2O_2 , glutathione (GSH), 3-mercaptopropionic acid (MPA) and methyl green (MG) which revealed the significant enhancement of DNA cleavage activity. The activity efficacy for complex **2** followed the order as $\text{H}_2\text{O}_2 > \text{GSH} > \text{MPA} > \text{MG}$.

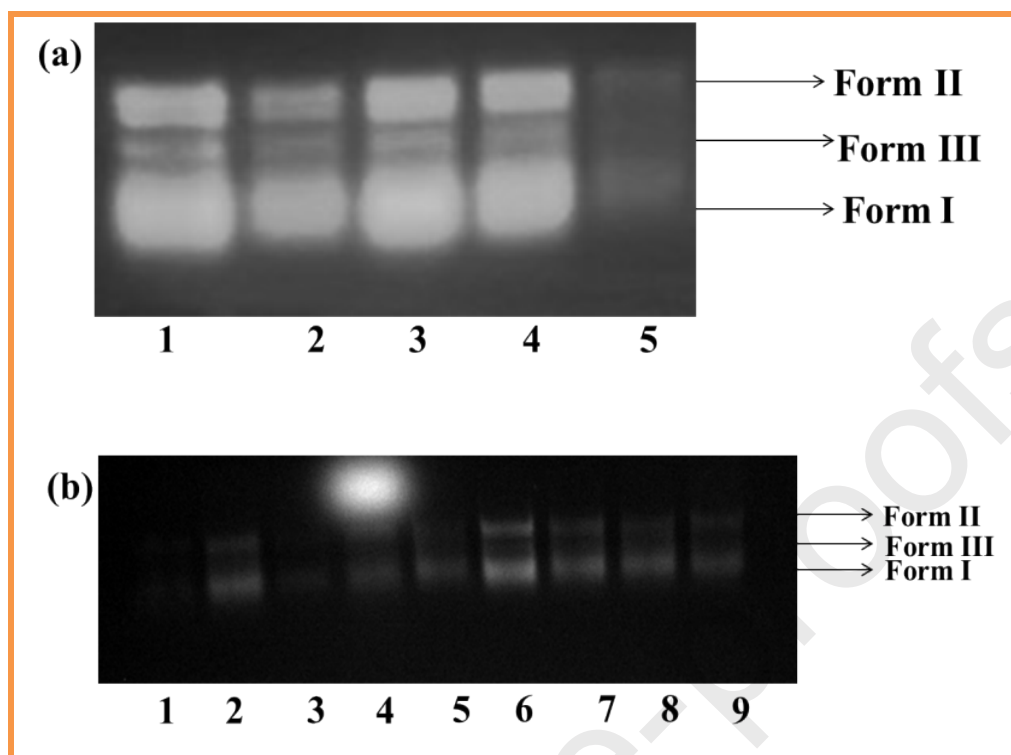


Fig. 12. Agarose gel electrophoresis demonstrating the cleavage of plasmid pBR322 DNA (300 ng) by complex 2 at 37 °C in the presence of different exogenous agents. (a) Lane 1, DNA control; lane 2, DNA + **2** (5 μM); lane 3, DNA + **2** (10 μM); lane 4, DNA + **2** (15 μM); lane 5, DNA + **2** (20 μM); (b) lane 1, DNA control, lane 2, DNA + **2** (20 μM) + H₂O₂ (0.4 μM), lane 3, DNA + **2** (20 μM) + NaN₃ (0.4 μM), lane 4, DNA + **2** (20 μM) + SOD (0.4 μM), lane 5, DNA + **2** (20 μM) + MPA (0.4 μM), lane 6, DNA + **2** (20 μM) + EtOH (5 μL); lane 7, DNA + **2** (20 μM) + DMSO (5 μL); lane 8, DNA + **2** (20 μM) + GSH (20 μM); lane 9, DNA + **2** (20 μM) + Methyl green (0.4 μM).

Experimental

Materials and instrumentation

6-Methyl-3-Formylchromone, 6-Bromo-3-Formylchromone, 6-Chloro-3-Formylchromone were purchased from Sigma Aldrich. All the solvents reagents were of best commercial grade and were used without any further purification. Elemental analysis were employed on Carlo Erba Analyzer Model 1106. Fourier-transform infrared (FT-IR) spectrum were recorded on an Interspec 2020 and Spectrum Two (Perkin Elmer) FT-IR spectrometers. The EPR spectrum of copper complexes **1–3** were carried out on a Varian E 112 ESR spectrometer using X-band frequency (9.5 GHz) at liquid

nitrogen temperature. ESI-MS spectra were recorded on Micromass Quattro II triple quadrupole mass spectrometer. Electronic spectrum were recorded on UV-1700 PharmaSpec UV-vis spectrophotometer (Shimadzu) while as emission spectrum were assessed on RF-5301PC Spectrofluorophotometer (Shimadzu). CD spectra were procured on Jasco J-815-CD spectropolarimeter. All experiments involving the interaction of complexes **1–3** with ct-DNA were carried out in aerated buffer (5 mM Tris-HCl, 50 mM NaCl, pH = 7.3). The concentrations per base pairs for ct-DNA were obtained spectrophotometrically by assuming ϵ_{260} nm values to be $6600 \text{ M}^{-1} \text{ cm}^{-1}$, respectively. Cleavage experiments were performed with the help of Axygen electrophoresis supported by a Genei power supply with a potential range of 50–500 V, visualized and photographed by a Vilber-INFINITY gel documentation system.

Synthesis of Complexes 1-3

Synthesis of complex 1

Complex **1** was synthesized by stirring ethanolic solutions of copper (II) nitrate trihydrate (0.241 g, 1 mmol) with 6-fluoro-3-formylchromone (.348 g, 2 mmol) in the molar ratio 2:1 under reflux conditions for 5h. The resulting solution was filtered and allowed to evaporate slowly at room temperature. Green coloured crystals suitable for single X-ray crystallography were obtained after 2-3 days of slow evaporation of the reaction mixture which were found to be quite stable towards air and soluble in organic solvents such as EtOH, acetone, THF, DMF and DMSO. Yield: 78%, melting point: 214°C . CCDC no. 1970411. Elemental analysis calculated for $\text{C}_{28}\text{H}_{32}\text{CuF}_2\text{O}_{10}$: C, 53.37; H, 5.12. Found: C, 53.45; H, 5.15. FT-IR (KBr, $\nu_{\text{max}}/\text{cm}^{-1}$): 3069 $\nu(-\text{C}-\text{H})$, 1614 $\nu(-\text{C}=\text{O})$, 1060 $\nu(-\text{C}-\text{O})$, 1263 $\nu(-\text{C}-\text{F})$. UV-vis ($1 \times 10^{-3} \text{ M}$, DMSO, λ_{max} , nm): 282, 310, 341 and 620.

Synthesis of complex 2

Complex **2** was synthesized by stirring ethanolic solutions of copper (II) nitrate trihydrate (0.241 g, 1 mmol) with 6-bromo-3-formylchromone (.506 g, 2 mmol) in the molar ratio 2:1 under reflux conditions for 4h. The resulting solution was filtered and allowed to evaporate slowly at room temperature. Very fine green coloured crystals suitable for single X-ray crystallography were obtained after 3-4 days of slow evaporation of the reaction mixture which were found to be quite stable towards air and soluble in organic solvents such as EtOH, THF, DMF and DMSO. Yield: 76%, melting point: 209 °C. CCDC no. 1987454. Elemental analysis calculated for $C_{28}H_{32}Br_2CuO_{10}$: C, 44.73; H, 4.29. Found: C, 44.78; H, 4.35. FT-IR (KBr, $\nu_{\max}/\text{cm}^{-1}$): 3089 $\nu(-C-H)$, 1656 $\nu(-C=O)$, 1045 $\nu(-C-O)$, 535 $\nu(-C-Br)$. UV-vis (1×10^{-3} M, DMSO, λ_{\max} , nm): 296, 315, 380 and 660.

Synthesis of complex 3

Complex **3** was synthesized by stirring ethanolic solutions of copper (II) nitrate trihydrate (0.241 g, 1 mmol) with 6-methyl-3-formylchromone (.376 g, 2 mmol) in the molar ratio 2:1 under reflux conditions for 6h. The resulting solution was filtered and allowed to evaporate slowly at room temperature. Green crystals suitable for single X-ray crystallography were obtained after 5-6 days of slow evaporation of the reaction mixture which were found to be quite stable towards air and soluble in organic solvents such as EtOH, THF, DMF and DMSO. Yield: 83 %, melting point: 223 °C. CCDC no. 1969139. Elemental analysis calculated for $C_{30}H_{38}CuO_{10}$: C, 57.91; H, 6.16. Found: C, 57.96; H 6.23. FT-IR (KBr, $\nu_{\max}/\text{cm}^{-1}$): 2974 $\nu(-C-H)$, 1606 $\nu(-C=O)$, 1060 $\nu(-C-O)$. UV-vis (1×10^{-3} M, DMSO, λ_{\max} , nm): 298, 335, 357 and 698.

Molecular docking studies

The molecular docking studies were performed by employing HEX 8.0 software. [56, 57] The B-DNA dodecamer d(CGCGAATTCGCG)₂ (PDB ID : 1BNA) crystal structure was downloaded

from the <http://www.rcsb.org/pdb> (protein data bank). Before employing docking experiments, the structure of complexes **1-3** were converted into PDB format and the visualization was done by using Discovery Studio molecular graphics program.

Computational studies

Single-point density functional theory (DFT) calculations and geometry optimization were carried out by the using hybrid B3LYP functional. The optimized structure was further calculated using def2-TZVP basis set for all atoms to calculate the HOMO and LUMO energies.

The def2-TZV/J main, and def2-SVP/J auxiliary, Coulomb fitting basis set were used as implemented in the ORCA, to accelerate the calculations.

***In vitro* binding and cleavage experiments**

The DNA binding experiments were performed in Tris-HCl/NaCl (5:50 mM) buffer at pH 7.3 by employing absorption spectroscopy, emission spectroscopy, cyclic voltammetry and circular dichroism following previously used standard methods adopted by our laboratory [58,59] The cleavage experiments of complexes **1-3** were carried out using agarose gel electrophoresis in Tris-HCl/NaCl (5:50 mM) buffer at pH 7.3 using supercoiled pBR322 DNA (300 ng).

Cytotoxic studies

In vitro antitumor screening of complexes **1-3** was performed on human cancer lines of two different histological origin viz., Du145 (prostate) and Huh7 (Liver). The effect of complexes **1-3** on cell viability was determined by MTT assay. Briefly, cells were seeded in 96-well dishes at a density of 3.5×10^3 cells per well. Cells were allowed to attach for 24h before incubation with fresh medium containing either complex **1/2/3** (2.5-10.0 mM) for 48h. After incubation for 48h at 37 °C, MTT assay was performed as described previously [60].

Description of X-ray crystal structures

Single crystal X-ray structural studies of complexes **1-3** were performed on a D8 VENTURE Bruker AXS diffractometer, employing graphite-mono-chromated Mo-K α radiation generated from a fine-focus sealed tube ($\lambda = 0.71073$ Å) at 150 K. The structure was solved by dual-space algorithm using the SHELXT program and further refined with full-matrix least-square methods based on F² (SHELXL). All non-hydrogen atoms were refined with anisotropic atomic displacement parameters. H atoms were finally included in their calculated position. The drawing of the complex was realized with PLATON [61] A summary of selected crystallographic information for complexes **1-3** is given in Table 2.

Table 2 Crystallographic data for complexes **1-3**.

Parameters	1	2	3
CCDC	1970411	1987454	1969139
Formula	C ₂₈ H ₃₂ CuF ₂ O ₁₀	C ₂₈ H ₃₂ Br ₂ CuO ₁₀	C ₃₀ H ₃₈ CuO ₁₀
Fw (g mol ⁻¹)	630.07	751.89	622.14
Crystal system	Monoclinic	Triclinic	Triclinic
Space group	<i>P2₁/n</i>	<i>P-1</i>	<i>P-1</i>
a (Å)	9.9291(4) Å	5.1134(5)	5.1349(4)
b (Å)	5.1435(2) Å	9.7980(9)	11.6066(9)
c (Å)	27.7256(10) Å	14.9650(14)	12.9630(11)
α (deg)	90 °	85.415(3) °	105.733(3) °
β (deg)	98.1990(10) °	83.430(3) °	97.741(3) °
γ (deg)	90 °	89.988(4) °	90.384(3) °
U (Å ³)	1401.48(9)	742.43(12)	736.10(10)
ρ_{calc} (g/cm ³)	1.493	1.682	1.403

μ (mm ⁻¹)	0.849	3.481	0.798
Temp (K)	150	150	150
Measured reflns	10339	3367	13894
Indep reflns	3218	3367	3385
GoF ^a	1.077	1.161	1.088
R ^b [I>2 σ (I)]	0.0337	0.0583	0.0266
wR ₂ ^b (all data)	0.0879	0.1267	0.0663

^a GoF is defined as $\{\sum [w(F_0^2 - F_c^2)]/(n-P)\}^{1/2}$ where n is the number of data and p is the number of parameters. ^bR = $\{\sum ||F_0| - |F_c|| / \sum |F_0|, wR^2 = \{\sum w(F_0^2 - F_c^2)^2 / \sum w(F_0^2)^2\}^{1/2}$.

Conclusions

New Cu (II) drug entities **1–3** derived from substituted (F⁻, Br⁻, -CH₃) bioactive chromone pharmacophore were synthesized as antitumor chemotherapeutic agents. The complexes **1–3** were thoroughly characterized by various spectroscopic techniques (IR, UV-vis, EPR and ESI-MS) and single X-ray crystallography. Interaction of complexes **1–3** with ct-DNA was carried out employing UV-vis, fluorescence, cyclic voltammetry, EPR and circular dichroism techniques. The corroborative results of these experiments with ct-DNA showed higher binding propensity of complex **2** as compared to complexes **1** & **3**. It was observed that complex **2** facilitates strong halogen bonding interaction via, Br and stability of halogen bond was rationalized by the presence of sigma hole region as observed by gas phase B3LYP computational DFT studies. Moreover, the cleavage activity of complexes **1–3** with pBR322 DNA suggested double strand cleaving ability of the complexes and the mechanism was mediated by oxidative cleavage pathway involving SOD and NaN₃. *In vitro* cytotoxic activity of complexes **1–3** was carried out on a panel of two human cancer cell lines by MTT assay and the growth curve data revealed greater potency of complex **2**

towards human liver carcinoma (Huh-7) and prostate cancer (DU-145) cell lines than the other two complexes **1** & **3**.

Conflicts of interest

There are no conflicts to declare.

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Highlights

- Substituted chromone Copper (II) complexes were synthesized and characterized.
- DFT computational studies were also performed.
- Electrophoretic studies revealed an oxidative cleavage pathway.
- Cytotoxicity was evaluated against the human liver carcinoma (Huh-7) and prostate cancer (DU-145) cell lines.
- Copper (II) complex **2** exhibited selective cytotoxicity against DU-145 cell line at GI_{50} = 1.6 μ M.