

Implication of Neuronal Versus Microglial P2X4 Receptors in Central Nervous System Disorders

Alexia Duveau, Éléonore Bertin, Eric Boué-Grabot

▶ To cite this version:

Alexia Duveau, Éléonore Bertin, Eric Boué-Grabot. Implication of Neuronal Versus Microglial P2X4 Receptors in Central Nervous System Disorders. Neuroscience Bulletin, 2020, 10.1007/s12264-020-00570-y . hal-02997812

HAL Id: hal-02997812 https://hal.science/hal-02997812v1

Submitted on 10 Nov 2020

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers. L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

This is an accepted pre-copyedit version of an article published in Neurosci. Bull.

The final authenticated version is available online Neurosci Bull. 2020;10.1007/

s12264-020-00570-y. doi:10.1007/s12264-020-00570-y

Implication of neuronal versus microglial P2X4 receptors in central nervous system disorders

Alexia Duveau^{1,2}, Eléonore Bertin^{1,2} and Eric Boué-Grabot^{1,2*}

¹Univ. de Bordeaux, Institut des Maladies Neurodégénératives, UMR 5293, F-33000 Bordeaux, France

² CNRS, Institut des Maladies Neurodégénératives, UMR 5293, F-33000 Bordeaux, France

*: Corresponding author: Eric Boué-Grabot (EB-G) Institut des Maladies Neurodégénératives (IMN) CNRS UMR 5293, Université de Bordeaux, 146 rue Léo Saignat, 33076 Bordeaux cedex, France; phone: 33 (0)5 33 51 47 95; email: eric.boue-grabot@u-bordeaux.fr

Key words : purinergic signaling / P2X / ATP / ligand-gated ion channels / chronic pain / brain trauma / ischemia / neurodegenerative diseases /alcohol / neuropsychiatric disorders

Abstract

The P2X4 receptor (P2X4) is an ATP-gated cation channel that is highly permeable to calcium and widely expressed in neuronal and glial cell types throughout the CNS. A growing body of evidence indicates that P2X4 plays key roles in numerous central disorders. P2X4 trafficking is highly regulated and consequently in normal situations, P2X4 is present on the plasma membrane at low density and found mostly within intracellular endosomal/lysosomal compartments. An increase in *de novo* expression and/or surface density of P2X4 has been observed in microglia and/or neurons during pathological states. This review aims to summarize knowledge on P2X4 functions in CNS disorders and provide some insights into the relative contributions of neuronal and glial P2X4 in pathological contexts. However, determination of the cell-specific functions of P2X4 along with its intracellular and cell surface roles remain to be elucidated before the potential of P2X4 as a therapeutic target in multiple disorders can be defined.

Introduction

Initially discovered as one of the main sources of energy inside cells, adenosine 5' triphosphate (ATP) is now recognized as a ubiquitous extracellular cell-to-cell signaling molecule in the peripheral (PNS) and central nervous system (CNS), as well as in peripheral organs ^[1]. Although the release of ATP by sensory nerve cells was observed in the 1950s ^[2], Geoffrey Burnstock was the first to propose in 1972, the concept of purinergic signaling using ATP as a fast neurotransmitter ^[3]. ATP was later shown to mediate fast neurotransmission in the PNS and CNS alike, but is nowadays mostly considered as a neuromodulator co-released with other classical neurotransmitters such as GABA or glutamate at inhibitory or excitatory synapses in the CNS ^[4]. ATP is also a gliotransmitter released by glial cell types such as astrocytes or microglia ^[5-7]. Neuronal and glial released ATP exerts multiple actions in the neuromodulation of synaptic activity or plasticity, communication in glial networks and directly between glia and neurons via the activation of metabotropic P2Y and ionotropic P2X receptors, or after the conversion of ATP by ectonucleotidases into ADP and adenosine by the activation of P2Y and adenosine P1 receptors ^[4,8].

There is also growing evidence for the involvement of purinergic signaling in major CNS disorders including chronic pain, brain trauma, hypoxia/ischemia, epilepsy and neurodegenerative diseases associated with neuro-inflammation (for recent review, see ^[9]). In these noxious conditions, extracellular levels of ATP increase and serve as a danger signal along with activation of several purinergic receptor subtypes that are widely distributed both in neurons at pre-and post-synaptic sites and in glial cells ^[4, 6, 8, 10, 11]. Besides P2X7, P2X4, P2Y1 and A2_AR also P2Y6 (microglial

phagocytosis) and P2Y12 (microglial chemotaxis) participate in ATP danger signaling ^[9], increasing evidence indicates that microglial or neuronal P2X4 receptors are upregulated and play key roles in many CNS disorders. In this review, we focus on the function of neuronal and glial P2X4 function in physiological and pathological conditions.

Properties and function of P2X4 receptors

P2X receptors are trimeric ATP-gated cation channels that are formed by homo or heteromeric associations from 7 different subunits (P2X1 - P2X7) encoded by seven genes in mammals. Functional P2X receptors are non-selective cation channels permeable to sodium (Na⁺), potassium (K⁺) and calcium (Ca²⁺) and their activation by ATP binding leads to cell depolarization and Ca²⁺ influx. With the exception of P2X6, all homomeric P2X receptors are functional and display different ATP sensitivities, current kinetics and pharmacological properties. Moreover, the variety in expression pattern of the seven P2X subunits and the existence of heteromeric associations between these subunits contribute to the diversity and widespread action of ATP signaling ^[8]. Among the seven P2X subunits, P2X4 is the most widely distributed in various cell types throughout the body ^[1, 12]. All P2X subunits are expressed in the CNS, in a heterogeneous manner throughout the different structures, cell types and subcellular compartments ^[13], although P2X4 is one of the main subunits found in both neurons and glial cells such as microglia and more controversially in astrocytes, oligodendrocytes and Schwann cells (Table 1 and Figure 1)^[14]. However, the subunit composition of P2X receptors in most CNS cells is not well defined, and the physiological roles of P2X4 or P2X4-containing receptors is far from being deciphered due to the paucity of selective pharmacological tools. TNP-ATP a weak and non-selective P2X4 antagonist or

Neuroscience Bulletin

5-BDBD a selective allosteric P2X4 antagonist, have been commonly used to show the involvement of P2X4 in rodent CNS tissue ^[12] *in vitro* and *in vivo*. New potent antagonists of rodent P2X4 (BAY-1797 or NP-1815-PX) were recently identified ^[15, 16] as well as a highly selective human P2X4 antagonist (BX430) with no effect on rodent P2X4 ^[17]. Ivermectin, which is an agonist of invertebrate glutamate-gated chloride channels, is used as an antiparasitic drug in human medicine ^[18]. This compound is also a very potent positive modulator of mammalian P2X4. However, although highly selective for the P2X4 subunit among P2X subtypes, Ivermectin potentiates human P2X7 currents and can modulate other ligand-gated ion channels such as GABA_A or nicotinic receptors ^[19-22]. Ivermectin is a positive allosteric modulator of P2X4 that stabilizes this subunit in the open state ^[20], which consequently may also prevent P2X4 internalization ^[21, 22].

P2X4 trafficking

Indeed, in contrast to other P2X subunits, P2X4 is constitutively and highly internalized, and as a result, is found mainly in endosomal/lysosomal compartments thereby ensuring low surface expression of these receptors in basal states ^[23]. Recent work showed that intracellular P2X4 in these acidic compartments are functional and may have important functions in lysosome membrane fusion ^[24]. In various pathological states, such as trauma, ischemia, chronic pain, neurodegenerative processes and several neuropsychiatric disorders, *de novo* expression of P2X4 and/or an increase in cell surface P2X4 density was observed in microglia and/or neurons, thus suggesting possible key and multiple roles of neuronal and microglial P2X4 receptors in the establishment and/or maintenance of these pathologies ^[25]. Changes in the intracellular expression of P2X4 may also have important consequences in the pathophysiological context.

Similar to all seven P2X subunits, P2X4 has a topology with 2 transmembrane regions, linked by a large extracellular domain and two intracellular amino and carboxyl termini ^[26]. The first X-ray crystal structure of a P2X receptor resolved in a truncated form of the zebrafish P2X4 receptor in

its closed state ^[27] and then in complex with ATP ^[28] from crystallized human P2X3 in different states ^[29] confirmed the subunit topology and trimeric organization of P2X receptors. The structure of head domain of rat P2X4 has also been investigated ^[30]. These data also revealed the importance of a large extracellular disulfide-rich region packed with N-linked glycosylation moieties and the existence of three inter-subunit binding sites. They also showed that the binding of three molecules of ATP is required to induce conformational changes leading to the opening of the channel pore formed by the three transmembrane regions ^[28]. P2X subunits share several conserved motifs within the intracellular domains, such as a YXXXK in the C-terminus that regulates surface expression ^[31] and a TXK/R in the N-terminus which is a putative protein kinase C phosphorylation site regulating current desensitization ^[32]. However, in terms of sequence and length, the C-terminus domain is the most divergent between P2X subunits ^[12]. The mammalian P2X4 subunit displays two endocytic tyrosine-based and di-leucine motifs within the C- and Ntermini, respectively, which are responsible for its constitutive internalization and main localization in lysosome-related intracellular organelles ^[33]. This distribution is in contrast to other P2X receptors such as P2X2, which are mainly found at the level of the plasma membrane ^[34]. Interestingly, the clathrin-dependent constitutive internalization of P2X4 is mediated by an interaction between the non-canonic motif YXXG Φ and the μ 2 subunit of the adaptive protein 2 (AP2), whereas the canonic YXXΦ motif is not involved ^[34, 35]. Mutations of the endocytic motif YXXGL or blockade of clathrin-mediated internalization lead to an increase both in the number of P2X4 at the plasma membrane and associated ATP currents^[22, 34-36]. Being highly glycolyzed, the extracellular loop of P2X4 allows it to resist the very acidic intralysosomal environment and proteolysis. Intracellular P2X4 pools can thus remain intracellular or either recycle back to the cell surface and dynamically regulate the number of surface P2X4 receptors ^[23, 34, 37-39]. Several studies have shown that in macrophages and microglia, the surface trafficking of P2X4 is upregulated in response to stimuli promoting lysosomal exocytosis or altering endocytosis ^[33, 40, 41]. In numerous

 pathophysiological conditions, *de novo* expression of P2X4 and/or increased surface trafficking had been observed ^[42] suggesting an important role of the dynamic regulation of P2X4 surface trafficking in diseases (Figure 1).

intracellular P2X4 function

Finally, growing evidence suggests that lysosomal P2X4 may also have important physiological functions (for review, see ^[43]). In lysosomes, the extracellular domain of P2X4 containing the agonist binding sites faces the luminal compartment, which contains high concentrations of ATP ^[44, 45] transported by the vesicular nucleotide transporter VNUT/SLC17A9 present in the lysosomal membrane. Alkalinization of the intralysosomal pH may lead to P2X4 activation and participate in endolysosomal fusion and vacuolation ^[43]. Indeed, Ca²⁺ flux through P2X4 activates calmodulin which subsequently associates with the receptor channel and promotes vesicular fusion ^[46].

Neuronal P2X4

P2X4 is expressed in various tissue cell types of peripheral organs including epithelial cells, kidney, pancreas, lung, liver, cardiovascular system or immune cells (for review, see ^[12]) in which it has important functions. In the CNS (Table 1), P2X4 along with P2X2 and P2X6, is the most widely expressed P2X subunit in both neurons and glial cells ^[14, 47]. P2X4 was first identified and cloned from brain tissues in 1996 by two independent groups ^[14, 48]. Their studies also showed the expression of P2X4 mRNA in the dentate gyrus, CA1/CA3 pyramidal cells of the hippocampus and in cerebellum Purkinje cells ^[14, 48]. Further immunohistochemical studies with anti-P2X4 antibodies confirmed the presence of P2X4 at the protein level in GABAergic interneurons of the hippocampus, cerebellum and olfactory bulb ^[13, 49], spiny neurons of the striatum and substantia nigra ^[49, 50], and in the hypothalamus and hypophysis ^[51]. Other studies reported P2X4 expression

in somatosensory cortical neurons ^[52], sensory nerves or ganglia ^[53-55] and the retina ^[56]. Electron microscopy has indicated that neuronal P2X4 is located both pre- and post-synaptically, mainly at the edge of the postsynaptic density in the peri- and extra-synaptic space ^[13]. This is consistent with the idea that activation of synaptic P2X receptors by ATP coreleased with other neurotransmitters such as glutamate or GABA did not mediate fast synaptic transmission but modulate excitatory or inhibitory synapses in the CNS ^[6]. The widespread distribution of P2X4 in CNS neurons was confirmed with P2X4 knockout mice expressing β-galactosidase instead of P2X4 ^[57] and recently with a transgenic reporter mice expressing tdTomato under the control of a P2X4 promotor and subsequently using a novel internalization-defective P2X4mcherryIN knockin mouse ^[58, 59].

Neuronal P2X receptors located at pre- and post-synaptic sites act mainly as neuromodulators. However, in most cases the specific contribution of P2X4 has been difficult to ascertain due to the paucity of appropriate pharmacological or genetic tools. In addition, the P2X4 subunit may not only form functional homomeric receptors, but can also assemble in heteromeric association with P2X6 (P2X4/6) or as heterotrimeric P2X2/4/6 ^[60, 61]. Activation of pre-synaptic P2X4 increases the release of neurotransmitters such as glutamate or GABA in neurons of the arcuate nucleus of the hypothalamus regulating food intake ^[58]. At the post-synaptic level, ATP released by neurons or glial cells acting on postsynaptic P2X4 receptors modulates activity and plasticity of excitatory and inhibitory synapses in several areas of the brain ^[10, 62]. P2X4 can modulate excitatory synaptic transmission and plasticity by acting on glutamatergic NMDA and AMPA receptors ^[63]. The influx of Ca² + *via* the opening of either P2X2 or P2X4 channel pores can trigger the internalization of AMPARs by phosphorylation of the GluA1 subunit ^[10, 11] *in vitro*, although in hippocampal neurons, this action seems to be mediated by P2X2 rather than P2X4, consistent with the low surface expression of hippocampal neuronal P2X4 in basal conditions ^[10]. P2X4 receptors can also

Neuroscience Bulletin

the hippocampus. Indeed, the pharmacological blockade of P2X4 facilitates the induction of NMDAR-dependent LTP ^[64], suggesting that P2X4 have a negative impact on LTP. However, another study has shown that potentiation of P2X4 by ivermectin (IVM), a positive modulator of P2X4, increases LTP in the hippocampal CA1 region ^[57]. This effect was not observed in P2X4KO mice. The results suggested that the Ca²⁺ influx *via* postsynaptic P2X4, by promoting the incorporation of synaptic NMDARs, would strengthen synaptic activity during LTP ^[65]. ATP originating from individual excitatory synapses or glial cells, can activate P2X4 in neocortical neurons and in turn reduce the currents induced by NMDAR activation. This effect is not observed in P2X4KO mice in which LTP is increased ^[62]. Altogether, these results point to the multiple neuromodulator actions of P2X4 at central excitatory synapses.

P2X4 is also capable of modulating GABA inhibitory synapses. The physical interaction between post-synaptic P2X4 and GABA_A receptors in the ventromedial nucleus of the hypothalamus leads to an inhibition of GABA-mediated postsynaptic currents and consequently enhances neuronal excitability ^[36, 66]. Interestingly, this effect was observed solely after blockade of P2X4 internalization using a competitive peptide, leading to the increase of surface expression of P2X4 ^[36]. P2X4-mediated attenuation of tonic and phasic GABAergic inhibition was also observed in cortical neurons. In this case the effect is apparently mediated by P2X4-induced Ca²⁺ influx and a PKC-dependent downregulation of GABA_A receptors ^[6, 67].

Glial P2X4

Glial cells are themselves capable of releasing ATP, which can play an important role in neuronglia and glial network signaling such as propagating glial calcium waves ^[68]. P2X4 is expressed in microglia with P2X7 and both receptors can interact with each other. The existence of P2X4/7 heteromers remains controversial and interaction may also occur by direct or indirect interaction between homomeric P2X4 and P2X7 ^[69]. A *de novo* expression of P2X4 and/or an increased

 surface trafficking of P2X4 observed in activated microglia ^[42] play key roles in many CNS diseases (see below). The upregulation of microglial P2X4 was revealed following LPS-induced inflammation in the hippocampus and selectively involving activated microglia using *dt*Tomato reporter mice and internalization-defective P2X4mCherryIN knockin mice ^[58, 59].

The P2X4 receptor has been revealed by immunostaining in S100β-positive astrocytes of the rat hippocampus in the CA1, CA3 regions and the dentate gyrus ^[70]. P2X4 expression was additionally detected in astrocytes from hippocampal cultures of P2X4mCherryIN mice, as well as in astrocyte end-feet contacting endothelial cells in brain slices of these mice solely by electron microscopy ^[59]. P2X4 were also detected in GFAP-positive astrocytes of the rat nucleus accumbens ^[71]. Nevertheless, P2X4 expression in astrocytes remains debated ^[72], although this discrepancy may merely reflect the heterogeneity of astrocytes across the different regions of the CNS. Indeed, a growing body of evidence suggests that astrocytes as well as microglia and oligodendrocytes display regional and contextual heterogeneities in terms of gene expression pattern, phenotypes and roles in controlling physiological and pathological brain functions ^[73-75].

Recent studies have shown the presence of P2X4 in satellite glial cells (SGCs) stained by astrocytic markers of dorsal root ganglia. This expression was observed solely during neuropathic pain ^[76] and diabetes-induced neuropathy ^[77] suggesting a pathology-induced upregulation of P2X4 in SGCs. The expression of several types of P2 receptors, including P2X4, has also been demonstrated in oligodendroglial precursor cells ^[78]. However, its function is not yet fully understood, in contrast to P2X7 receptors which contribute to ATP excitotoxicity in oligodendrocytes and the pathogenesis of experimental autoimmune encephalomyelitis^[79].

P2X4 has also been observed in Schwann cells, mainly localized intracellularly in lysosomes, under normal physiological conditions. After peripheral nerve injury, an overexpression of P2X4

receptors and their increase on the surface of Schwann cells was shown to promote, through the release of BDNF, motor and sensory functional recovery and ameliorate nerve remyelination ^[80].

P2X4 in CNS diseases

Neuropathic Pain

Acute pain serves a protective function in promoting reflex withdrawal from painful stimuli and the avoidance of injury or damage. Acute pain is initiated by an activation of nociceptive neurons in peripheral dorsal root ganglia (DRG), with information being processed and integrated within the spinal cord (SC) before transmission to the brain ^[81]. Whereas acute pain is a normal physiological function of the CNS, chronic pain results from pathological and persistent alterations in the peripheral and central nervous systems ^[82]. Chronic pain can be induced by nerve damage, so-called neuropathic pain, or by persistent inflammatory pain associated with the healing of damaged tissue^[83]. Chronic pain persists even after the expected period of healing and becomes disabling for the individual ^[84].

Peripheral nerve injury (PNI) was shown to induce an activation of microglia in the spinal cord dorsal horn ^[85, 86] by several pathways involving interferon- γ (IFN- γ) or platelet-derived growth factor (PDGF) ^[87, 88]. Activation of spinal microglia causes tactile allodynia, a symptom of neuropathic pain , and several studies following the pioneered work of Tsuda *et al.* ^[89] showed that a specific microglial response phenotype characterized by the *de novo* expression of P2X4 is critical for the pathogenesis of neuropathic pain ^[67]. The expression of P2X4 increases exclusively in the reactive microglia after PNI and is confined to the ipsilateral spinal dorsal horn (Figure 2). Moreover, the genetic ablation or downregulation of P2X4 using either P2X4KO mice or RNA

antisense as well as the blockade of P2X4 by the antagonist TNP-ATP dramatically reduces tactile allodynia after PNI ^[89-91]. Interestingly, an activation of microglia is also observed in P2X4KO mice after PNI ^[91] indicating that P2X4 is not required for microglial activation but is necessary and sufficient for the development of allodynia by neuro-glial communication. Indeed, activation by ATP of microglial P2X4 reduces GABA or Glycine receptor-mediated inhibitory currents within the spinal cord. This disinhibition results from an increase in the intracellular concentration of chloride that becomes sufficient to block GABA or Glycine mediated-inhibitory responses or to eventually convert inhibition into excitation, thereby inducing the hypersensitivity of spinal neurons and tactile allodynia^[92, 93]. Changes in chloride concentration is due to the downregulation of the potassium-chloride co-transporter KCC2, which is mediated by the extracellular release of BDNF and the activation of Trk-B receptors in inhibitory neurons. Several studies have established the key role played by BDNF in this signaling cascade and the resulting disinhibition and painrelated behavior ^[92, 94]. Also established is the requirement for ATP-mediated activation of P2X4 receptors to elicit BDNF release by microglia^[91], although how P2X4 actually triggers the release of BDNF remains to be determined. In the dorsal horn of the spinal cord, ATP can be released by the terminals of primary sensory neurons ^[95], dorsal horn neurons ^[96], or glial cells ^[7].

A recent study showed that PNI increased the expression of the vesicular nucleotide transporter (VNUT) and induced a slight increase in the extracellular concentration of ATP in the spinal cord. By cell-specific deletion of VNUT, the authors elegantly showed that ATP released by spinal cord neurons and astrocytes, but not by sensory neurons, contributes to neuropathic pain^[97] very likely by activating microglial P2X4-mediated signaling cascades. However, ATP is co-released with GABA in spinal dorsal horn neurons that can activate simultaneously postsynaptic P2X and GABA_A receptors^[96]. A cross-talk between neuronal P2X, including P2X4 and GABA_A receptors,

Neuroscience Bulletin

 may downregulate GABA-mediated currents and thus contribute to disinhibition in neuropathic pain ^[36, 96, 98].

Several mechanisms are involved in the increase of P2X4 expression in microglia following PNI. Two transcription factors of the Interferon Regulatory Factor (IRF) family, IRF8 and IRF5, that are expressed in immune cells including microglia ^[99, 100] are upregulated after PNI in dorsal horn microglia and silencing IRF8 was shown to suppress PNI-induced tactile allodynia^[100]. IRF8 promotes several actors involved in chronic pain ^[100] and IRF5 expression which in turn controls directly P2X4 transcription by binding to the promoter region of p2rx4 gene ^[100]. Fibronectin also appears to be a key regulator of P2X4 expression through several pathways. The observed increase in extracellular fibronectin after PNI could induce *de novo* P2X4 expression through several fibronectin/integrin signaling pathways ^[90, 101], as well as promoting P2X4 trafficking from the lysosome to the cell surface through the release of a chemokine CCL2 and subsequent activation of microglia CCR2 receptors ^[102].

An upregulation of P2X4 in spinal microglia is likely to play a key role in tactile allodynia induced by PNI, although interestingly, recent work has suggested that this mechanism occurs only in males ^[94, 103]. In contrast to male mice following PNI, no increase in P2X4 expression was observed in dorsal horn microglia of post-PNI female mice and tactile allodynia was not modified by the pharmacological blockade of P2X4 with TNP-ATP ^[94]. Results suggest that immune cells contribute to pain hypersensitivity in female mice ^[94, 104], although other studies have indicated that microglial P2X4 and BDNF/TrkB signaling contribute to bone cancer ^[105] and herpetic pain ^[16] models in female rodents, thus demonstrating the need for additional studies to clarify this sexual dimorphism.

Besides microglial P2X4 in the spinal cord, P2X4 expression in sensory neurons was recently linked to neuropathic pain. All P2X subunit mRNAs are expressed in sensory DRG neurons ^[106], but the functional expression and potential involvement in pain processing was only until very recently found to be limited to homomeric P2X3 or heteromeric P2X2/3 receptors expressed in nociceptive neurons ^[107]. Recent work showed that P2X4 is expressed at mRNA and protein levels in DRG sensory neurons ^[77, 108-110] and that its expression increases in several neuropathic or inflammatory pain models in rodents ^[111]. P2X4 receptor expression was also observed in satellite glial cells (SGC) which enwrap the cell bodies of sensory neurons within the ganglia ^[110, 112]. Although it can be difficult to distinguish P2X4 in the plasma membrane of sensory neurons or SCGs due to the apposition of both elements, several lines of evidence have established that P2X4 expression increases in SCGs under neuropathic pain conditions ^[77, 111, 113]. After PNI, the coupling between the SGCs themselves and between SGCs and neurons increases via the formation of gap junctions ^[114, 115] and activated SCGs release inflammatory mediators and other neuromodulators that contribute to neuronal hyperexcitability and pain hypersensitivity ^[115].

Chronic inflammatory pain

P2X4 receptors expressed in distinct cell types also contribute to inflammatory pain. For example, the presence of P2X4 in skin-resident macrophages contributes to the sensitization of peripheral sensory nerve termini and induces mechanical hypersensitivity due to the production of the inflammatory factor PGE2 ^[116]. P2X4 also plays a role in mechanical hypersensitivity following chronic inflammation by promoting the hyperexcitability of spinal dorsal horn neurons in chronic inflammation mouse models ^[116] (Figure 2). On the other hand, such hyperexcitability is downregulated in P2X4-deficient mice ^[117]. A recent study showed that the expression of P2X4 receptors in small diameter nociceptive neurons of DRGs is increased by twofold during chronic inflammation ^[109]. In addition, these sensory neurons express BDNF, a key factor in both

Neuroscience Bulletin

neuropathic and inflammatory pain. Indeed, in chronic inflammatory conditions, BDNF contributes to mechanical hypersensitivity using the similar pathway. However, unlike neuropathic pain, during chronic inflammation, BDNF is released in a P2X4-dependent manner by sensory neurons termini in the dorsal horn of the spinal cord, thereby leading *via* the TrkB/KCC2 cascade to spinal disinhibition ^[118]. P2X4 expressed in SGCs may also contribute to chronic inflammatory pain ^[112, 119] by releasing pro-inflammatory cytokines such as IL1 β and TNF α following activation of the inflammasome NLRP1 ^[116, 119].

Migraine

Chronic migraine is a neurological disorder characterized by repeated attacks that can be caused by an increase in the excitability of central neurons in the trigeminal nociceptive pathway, principally the trigeminal nucleus caudalis (TNC), leading to central sensitization ^[120]. Similar to neuropathic pain, microglia surrounding TNC neurons has been suggested to play direct and indirect roles in the establishment of central sensitization ^[121]. Indeed, microglial activation has been observed when nitroglycerin (NTG) is applied chronically in a model of chronic migraine, with a resultant effect on central sensitization ^[121]. As observed during neuropathic pain ^[89, 91], several studies suggested a role for BDNF expressed in the trigeminovascular system in migraine pathophysiology ^[122] and an increase in P2X4 and BDNF expression has been recently reported in the TNC after chronic intermittent administration of NTG [121, 123]. Moreover, P2X4 pharmacological inhibition has been very recently found to prevent hyperalgesia induced by NTG, associated with an inhibition of p-ERK phosphorylation and calcitonin gene-related peptide (CGRP) release in the TNC^[123]. This study also showed that in BV2 microglial cells, ATP triggers BDNF synthesis and release, which is in turn reduced by the P2X4 antagonists 5-BDBD or SB203580^[123]. In addition, a potentiation of P2X4 by ivermectin induced sustained hyperalgesia and significantly increased levels of p-ERK and CGRP release in the TNC ^[123]. Those results

therefore clearly indicate that microglial P2X4 contributes to TNC neuronal hyperexcitability involved in chronic migraine.

Post-ischemic inflammation

Brain ischemia is associated with post-ischemic inflammation, including glial cell activation along with an increase in extracellular ATP levels. Several studies have reported an upregulation of P2X4 receptors in activated microglial cells after ischemic brain injury ^[124-127] (Figure 3). During hypoxic conditions, ATP is released in large amounts from cells damaged by ischemia and the activation of both P2Y12 and P2X4 receptors present in a higher density in reactive microglia mediates, *via* the P13K signaling pathway, chemotaxis of motile microglia towards the lesion site as well as the release of proinflammatory cytokines ^[125, 128, 129]. More recently, a neuroprotective role of P2X4 during ischemia has also been reported ^[130]. Specifically, the activation of P2X4 expressed in vascular endothelial cells by ATP release consequently to pressure shocks mediated by vascular ischemia promotes the upregulation of osteopontin, a neuroprotective molecule ^[130]. These results therefore highlight a new mechanism whereby endothelial P2X4 induces an ischemic tolerance.

Epilepsy

Epilepsy is a chronic neurological condition characterized by recurrent seizures that spread to neighboring cortices, and lead to hippocampal neuron loss and microglia activation (Figure 3). To date, as for P2X7, the implication of P2X4 in epilepsy remains unclear, and whether the latter is up- or down-regulated in the hippocampus following status epilepticus (SE) is still debated. Some studies have reported a lack of change, or even a decrease in P2X4 immunoreactivity, 24 hrs after SE induced either by systemic pilocarpine or by intra-amygdala kainate injection . Kang et al.

Neuroscience Bulletin

reported similar findings in seizure-sensitive compared with seizure-resistant gerbils^[131], although the specificity of the antibody anti-P2X4 used in these studies has not been established.

Subsequently, in an epileptic mouse model using intraperitoneal injection of kainate, hippocampal P2X4 transcript and protein levels were found to increase 24 and 48 hrs after the SE, and particularly in activated microglia in the stratum radiatum ^[132]. In P2X4KO mice, a decrease in microglial activation as well as subsequent cellular death were observed after the induction of the SE^[132]. Even though epileptic seizures induced by kainate are not strongly inhibited in P2X4KO mice, there is still a significant reduction of cell death in the hippocampus as well as a decrease in the amplitude of hyperpolarizing outward current activated by microglia ^[132]. As for neuropathic pain, the increase in *de novo* P2X4 expression in activated microglia after the induction of epilepsy could facilitate BDNF release and the induction of hyperexcitability, thereby promoting epileptogenesis. Zicz

Alzheimer's disease

Alzheimer's disease (AD) is the most common neurodegenerative disorder and is characterized mainly by the accumulation of insoluble aggregates of beta-amyloid fragments (A β_{1-42}) that in turn lead to synaptic dysfunction and neuronal death causing a progressive loss of memory ^[133, 134]. Like other neurodegenerative diseases, the increase of extracellular nucleotides concomitantly with exacerbated neuroinflammatory events play a crucial role in AD pathogenesis ^[135].

It has been proposed that ATP, acting through P2X receptors, could participate directly in some of the toxic effects of A^β peptide observed in neurons, with a particularly important role of P2X4 receptors ^[136, 137]. Indeed, an accumulation of P2X4 has been observed in hippocampal neurons exposed to the Aβ peptide ^[138]. This expression increase was found to be restricted to the cell body after 6 hrs of treatment, before also reaching the neurites after 12 hrs ^[138]. In parallel, after exposure to an AB fragment, the authors revealed an accumulation of a smaller P2X4 fragment, which could

be prevented by exposure to a caspase inhibitor, thus suggesting occurrence of a proteolytic cleavage of P2X4 receptors (Figure 3). They concluded that the cleavage by caspase-3 slows down the closing time of the receptor's channel pore and prevents receptor agonist-dependent internalization. This leads to an increase in membranal surface trafficking of the neurons that can in turn induce neuron toxicity due to increased calcium influx ^[138]. In this study, moreover, the overexpression of neuronal P2X4 was found to promote the toxic effect of the A β fragment while inhibition of receptor expression has a significant positive effect on neuronal death after A β exposure. Consequently, these findings led to the conclusion that neuronal P2X4 receptors are directly implicated in neuronal death induced by A β ^[138]. Consistent with the implication of neuronal P2X4 in AD, an increased surface density of neuronal P2X4 was recently shown to induce synaptic deficits and alterations in learning and memory functions of P2X4 internalization-defective knock-in mice. Surprisingly, however, the authors also reported a decrease in P2X4 protein levels in the middle frontal and temporal gyri of AD patients with severe cognitive impairment ^[138].

P2X7 has been shown to play a key role in neuroinflammatory processes of AD and other neurodegenerative diseases, including Parkinson's disease (PD), amyotrophic lateral sclerosis (ALS), multiple sclerosis (MS) and Huntington disease (HD), in promoting proinflammatory cytokine production by microglia and astrocytes ^[139-141]. Like P2X7, P2X4 could also be implicated in neuroinflammation associated with AD ^[142] and the resultant potentiation of Aβ-mediated neuronal death, since similarities exist between these two receptor subtypes as well as a neighboring chromosomal localization ^[69, 143], and because PX4 has been correlated with several other inflammatory mechanisms ^[144, 145] and microglial activation processes ^[89].

Parkinson's disease

Page 19 of 51

Neuroscience Bulletin

PD is characterized by the progressive loss of dopaminergic nigrostriatal neurons specifically, causing impaired dopamine (DA) homeostasis and associated behaviors leading to motor troubles. A hallmark of PD is the presence of abnormal intracytoplasmic aggregates of α -synuclein, called Lewy bodies (LB), within neuronal cell bodies. It is well known that P2X4 modulates major neurotransmitter systems including glutamatergic and GABAergic functions ^[10, 11, 36, 57, 62, 64-67, 146-150]. Several early studies suggested that P2X4 receptors are indirectly involved in dopamine (DA) neurotransmission ^[151, 152] and behavioral deficits were recovered in P2X4 KO mice that may also express DA dysfunction ^[153, 154]. A recent study has confirmed the receptor's direct role in regulating DA homeostasis that is impaired in PD ^[155]. Specifically, using P2X4 KO mice, the authors showed that a *p2rx4* deficit affects DA synthesis and transport, and increases DA receptor levels that could in turn alter DA neuron function and neurotransmission, with a resulting impact on DA associated behaviors such as motor control and sensorimotor gating ^[155]. These findings further indicate that P2X4 could be implicated in multiple neurological disorders such as PD and psychiatric diseases that involve DA homeostasis impairment.

Neuroinflammation is also a main hallmark of PD ^[156-159] and microglia seems to be central in this neurodegenerative disease ^[160, 161]. As previously discussed, microglial P2X4 could be involved in those neuroinflammatory mechanisms that are common to neurodegenerative diseases, and for which, in the case of PD, P2X7 is known to play a role ^[162, 163]. Nevertheless, the implication of microglia in the physiopathology of PD is still unclear and it remains unknown whether microglial alterations are causal to, or a consequence of, DA neuron degeneration ^[164].

Amyotrophic lateral sclerosis (ALS)

ALS is a fatal neurodegenerative disease characterized by the progressive and selective loss of spinal motoneurons (MN) as well as cortical motor neurons (Figure 2). The causes of ALS are multifactorial and the mechanisms underlying the selective degeneration of MN are still unknown.

A link has been established between ALS and P2X4 ^[165]. For example, a strong increase in P2X4 immunoreactivity in ventral horn MN immediately preceding their death has been reported in the SOD1-G93A ALS rodent model, suggesting an important role of neuronal P2X4 receptors in the pathogenesis of the disease ^[166]. P2X4 immunoreactivity has also been associated with a degeneration of other neuronal populations including noradrenergic neurons in the *locus coeruleus*, cerebellar Purkinje cells and serotoninergic neurons of raphe nucleus ^[166]. Surprisingly, the same authors subsequently showed, by using anti-P2X4 antibodies directed against the intracellular C-tail of P2X4 subunits, that SOD1-G93A misfolded proteins were detectable in MN but not in microglia ^[167]. This finding led to the suggestion that neuronal conformers of SOD1-G93A proteins that are reactive to anti-P2X4 antibodies could play a pathogenic role and induce neuroinflammation activating microglia and astrocytes when injected intracerebrally in normal animals ^[167].

An increase in P2X4 expression, both at mRNA and protein levels, was also reported in microglia of SOD1-G93A mice ^[168]. More recently, Volonté and colleagues demonstrated for the first time an increase in P2X4 and P2X7 expression in the peripheral nervous system of SOD1-G93A mice, particularly in the sciatic nerves ^[165].

Other work has shown that a potentiation of P2X4 function by ivermectin has a dual effect on MN survival depending on ATP concentrations, with a beneficial action at low ATP levels, but becoming deleterious at higher levels ^[146]. The authors deduced that ivermectin potentiates a neuroprotective effect of P2X4 on MN at low ATP levels, probably by increasing P2X4 function and/or the receptor's cell surface density ^[20, 22]. Moreover, the oral administration of ivermectin via the animal's drinking water from a presymptomatic stage (P50), caused a 10% increase in the lifespan of SOD1-G93A mice ^[146].

Neuroscience Bulletin

As for other neuroinflammatory diseases, P2X4 receptor is also likely to be involved in neuroinflammation processes associated with ALS and known to be implicated in MN degeneration mechanisms^[169]. Several studies have highlighted a role of P2X7 in this disease ^[170], notably via a modulation of autophagy ^[171] as well as of microglia-mediated neuroinflammation processes that may be both beneficial or deleterious according to the stage of the disease, ^[172, 173]. An interaction between P2X4 and P2X7 receptors signaling pathways could also be involved in microglial inflammatory actions, with one pathway activating signaling of the other. Such a cross-talk is already known for macrophages, where P2X4 receptors regulate the inflammatory functions of P2X7 receptors ^[174]. Although the implication of P2X4 in the pathogenesis of ALS remains to be clearly established, this receptor subtype appears to be a potential therapeutic target for confronting this disease.

Multiple Sclerosis

Multiple sclerosis (MS) is a chronic T-cell mediated auto-immune disease characterized by a massive infiltration of immune cells, demyelination and axonal loss. This neurodegenerative disease is also characterized by an early inflammation that is beneficial and initially delays the onset of neurodegeneration, which then evolves in a complex manner as the disease progresses ^[175]. The initial regenerative phase is due to the repairing actions of anti-inflammatory infiltrating macrophages and microglia.

Interestingly, P2X4 has been shown to be overexpressed in microglia and macrophages in a rat model of MS and experimental autoimmune encephalomyelitis (EAE) ^[127] (Figure 3). But in contrast to neuropathic pain, where P2X4 blockers have been proposed to be potential therapeutic drugs ^[16], P2X4 antagonists exacerbate neurological symptoms in MS animal models, whereas P2X4 potentiation by ivermectin or P2X4 expression upregulation are both beneficial in EAE ^[176]. Moreover, microglial P2X4 activation is known to induce BDNF release ^[92] and it has been shown

that BDNF release from microglia as a result of P2X4 potentiation or overexpression in Schwann cells promotes remyelination ^[80, 176].

In contrast, P2X4 potentiation by ivermectin increases myelin engulfment and degradation ^[176] and P2X4 activation in microglia induces endolysosomal membrane fusion and lysosome pH acidification, both of which are essential to phagocytic pathways ^[46, 176] that could control myelin phagocytosis. Lysosomes are also involved in secretion and myelin biogenesis ^[177, 178] and P2X4 has been shown to be implicated in lysosome exocytosis contributing to surfactant secretion in lung tissue ^[179]. Altogether these findings therefore suggest a potential role for lysosomal P2X4 receptors in secretion and phagocytosis in myelin disorders.

Alcohol-related disorders

Several studies have pointed to a role of P2X4 in alcohol-induced behavior ^[180-183]. P2X4 receptors are expressed in brain regions involved in the reinforcing properties of alcohol and other drugs ^[184]. Furthermore, a recent study suggested the involvement of P2X4 in regulating striatal dopamine homeostasis ^[155], supporting a role of P2X4 in the mesolimbic dopamine system implicated in the reward system of the brain (Figure 3).

An increase in ethanol intake is observed in P2X4 KO mice, suggesting that P2X4 could also be implicated in alcohol intake and/or preference ^[182, 185]. Other studies have demonstrated a lower P2X4 expression in rats that prefer alcohol, confirming the correlation between P2X4 expression and alcohol preference ^[183, 184]. Moreover, a direct link between P2X4 and the regulation of dopamine neurotransmission in the mesolimbic system has been established, further indicating that through its presence in dopaminergic neurons of the mesolimbic system, P2X4 plays a role in mediating alcohol drinking behavior ^[183].

Interestingly, low ethanol concentrations are able to inhibit P2X4 *in vitro*, probably by blocking opening of the channel without affecting its deactivation ^[186, 187]. Conversely, ivermectin is able to

Neuroscience Bulletin

antagonize the inhibition of P2X4 by ethanol, indicating that this positive modulator interferes with the linkage site of ethanol on the P2X4 receptor ^[188]. Altogether, these findings suggest that ethanol can reinforce alcohol intake via P2X4 inhibition and that pharmacological potentiation of P2X4 may reduce alcohol intake and preference.

Alcohol abuse is also known to promote neuroinflammation ^[189] and microglial P2X4 has been recently reported to play a role in this mechanism ^[190, 191]. Gofman and collaborators demonstrated that an alcohol-induced increase of P2X4 expression in microglia affects microglial migration and phagocytosis, and can be reversed by a P2X4 selective antagonist^[190]. Later, the same group found that via P2X4, alcohol decreases the phosphorylation of key regulatory proteins and increases CREB transcriptional activity^[191]. Cerebral ischemic events have also been reported in alcohol abuse, involving an upregulation of P2X4 in microglia following ischemia and hypoxia as 1.02 described above [128, 192].

Neuropsychiatric disorders

Depression, bipolar disorder, schizophrenia, attention deficit hyperactivity and anxiety are neuropsychiatric disorders where DA homeostasis is impaired. As mentioned above, P2X4 has been shown to be implicated in the regulation of DA homeostasis and sensory motor gating ^[155] and there is increasing evidence that P2X4 plays a critical role in psychiatric disorders [59, 126, 155, 193, 194]

Consistent with an involvement of P2X4 receptors, ivermectin has been found to produce anxiolytic-like and depressive-like behavior in mice ^[193]. Very interestingly and in line with these findings, data from a new conditional P2X4 internalization-defective knock-in mouse, namely P2X4mCherryIN, that display an increase in number of P2X4 receptors at the surface of targeted cells, further supported the link between neuronal P2X4 and anxiety-like behavior as well as in memory ^[59]. The anxiolytic effects of a selective increase in surface P2X4 density in mouse

excitatory forebrain neurons further underlined the role of neuronal P2X4 in anxiety ^[59]. Another study using P2X4KO mice on the role of P2X4 in ischemic stroke showed that P2X4 deletion predisposes animals to chronic depression-like behavior after stroke ^[126]. Significantly in this context, a recent work demonstrated that the potentiation of P2X4 by ivermectin can lead to DA hyperactivity and disruption of information processing because of a potential perturbation of the interaction between P2X4 and DA receptors ^[194]. Together these results suggest that P2X4 antagonists could serve as novel anti-psychotic treatments for psychiatric disorders arising from sensorimotor gating impairments linked to the disruption of DA homeostasis ^[194].

Concluding remarks

P2X4 is the most widespread P2X subunit expressed in several neuronal and glial cell types throughout the CNS. Homomeric or heteromeric P2X4-containing receptors are ATP-gated channels that are highly permeable to calcium and in basal conditions, are localized in the plasma membrane at low density and mostly in intracellular endosomal/lysosomal compartments. Growing evidence suggest that endo/lysosomal P2X4 receptors are functional and play important intracellular signaling roles. Intracellular P2X4 can also be rapidly mobilized to the cell surface and contribute, together with *de novo* expressed receptors, to the upregulation observed in specific cell types in many central disorders such as pain, neurodegenerative disorders and neuropsychiatric diseases. However, establishing the function of upregulated P2X4 expressed in glial *versus* neuronal cells remain elusive. We have recently developed conditional internalization-defective P2X4mCherryIN knockin-mice with increasing numbers of P2X4 at the surface of targeted cells and have demonstrated the role of surface neuronal P2X4 in synaptic plasticity and memory^[59]. The development of such a transgenic knockin mouse, along with the possibility of generating a conditional P2X4KO line that targets specific cell populations, represent major tools

Neuroscience Bulletin

to gaining in the near future a better understanding of the role of surface P2X4 in various physiological and pathological conditions and to evaluate the potential of P2X4 as a therapeutic target in these various disorders. Intracellular P2X4 can also play important roles and extensive studies are now needed to evaluate the respective functions of intracellular *versus* surface P2X4 receptors, which may differ between the various cell types expressing P2X4. It will also be important to identify mechanisms and regulation of P2X4 trafficking leading to the cell surface increase in pathological conditions, as well as determining the functional significance of the equilibrium between intracellular and surface P2X4 receptors.

Conflict of Interest

All authors declare that they have no competing interests

Acknowledgments

This work was supported by CNRS, University of Bordeaux, and grants from ARSLA, IdEx of Bordeaux and LabEx BRAIN ANR-10-LABX-43.

References

[1] North RA. P2X receptors. Philos Trans R Soc Lond B Biol Sci 2016, 371.

[2] Holton P. The liberation of adenosine triphosphate on antidromic stimulation of sensory nerves.

J Physiol 1959, 145: 494-504.

[3] Burnstock G. Purinergic nerves. Pharmacol Rev 1972, 24: 509-581.

[4] Khakh BS, North RA. Neuromodulation by extracellular ATP and P2X receptors in the CNS. Neuron 2012, 76: 51-69.

[5] Pankratov Y, Lalo U, Verkhratsky A, North RA. Vesicular release of ATP at central synapses.Pflugers Arch 2006, 452: 589-597.

[6] Boué-Grabot E, Pankratov Y. Modulation of Central Synapses by Astrocyte-Released ATP and Postsynaptic P2X Receptors. Neural Plast. 2017, 2017: 9454275.

[7] Imura Y, Morizawa Y, Komatsu R, Shibata K, Shinozaki Y, Kasai H, *et al.* Microglia release ATP by exocytosis. Glia 2013, 61: 1320-1330.

[8] Abbracchio MP, Burnstock G, Verkhratsky A, Zimmermann H. Purinergic signalling in the nervous system: an overview. Trends Neurosci 2009, 32: 19-29.

[9] Rodrigues RJ, Tome AR, Cunha RA. ATP as a multi-target danger signal in the brain. Front Neurosci 2015, 9: 148.

[10] Pougnet JT, Toulme E, Martinez A, Choquet D, Hosy E, Boue-Grabot E. ATP P2X receptors downregulate AMPA receptor trafficking and postsynaptic efficacy in hippocampal neurons. Neuron 2014, 83: 417-430.

[11] Pougnet JT, Compans B, Martinez A, Choquet D, Hosy E, Boue-Grabot E. P2X-mediated AMPA receptor internalization and synaptic depression is controlled by two CaMKII phosphorylation sites on GluA1 in hippocampal neurons. Sci Rep 2016, 6: 31836.

[12] Suurvali J, Boudinot P, Kanellopoulos J, Ruutel Boudinot S. P2X4: A fast and sensitive purinergic receptor. Biomed J 2017, 40: 245-256.

[13] Rubio ME, Soto F. Distinct Localization of P2X receptors at excitatory postsynaptic specializations. J Neurosci 2001, 21: 641-653.

[14] Soto F, Garcia-Guzman M, Karschin C, Stuhmer W. Cloning and tissue distribution of a novelP2X receptor from rat brain. Biochem Biophys Res Commun 1996, 223: 456-460.

[15] Werner S, Mesch S, Hillig RC, Ter Laak A, Klint J, Neagoe I, *et al.* Discovery and Characterization of the Potent and Selective P2X4 Inhibitor N-[4-(3-Chlorophenoxy)-3-sulfamoylphenyl]-2-phenylacetamide (BAY-1797) and Structure-Guided Amelioration of Its CYP3A4 Induction Profile. J Med Chem 2019, 62: 11194-11217.

[16] Matsumura Y, Yamashita T, Sasaki A, Nakata E, Kohno K, Masuda T, *et al.* A novel P2X4 receptor-selective antagonist produces anti-allodynic effect in a mouse model of herpetic pain. Sci Rep 2016, 6: 32461.

[17] Ase AR, Honson NS, Zaghdane H, Pfeifer TA, Seguela P. Identification and characterization of a selective allosteric antagonist of human P2X4 receptor channels. Mol Pharmacol 2015, 87: 606-616.

[18] Twum-Danso NA. Serious adverse events following treatment with ivermectin for onchocerciasis control: a review of reported cases. Filaria J 2003, 2 Suppl 1: S3.

[19] Khakh BS, Proctor WR, Dunwiddie TV, Labarca C, Lester HA. Allosteric control of gating and kinetics at P2X(4) receptor channels. J Neurosci 1999, 19: 7289-7299.

[20] Silberberg SD, Li M, Swartz KJ. Ivermectin Interaction with transmembrane helices reveals widespread rearrangements during opening of P2X receptor channels. Neuron 2007, 54: 263-274.
[21] Stokes L. Rab5 regulates internalisation of P2X4 receptors and potentiation by ivermectin. Purinergic Signal 2012.

[22] Toulme E, Soto F, Garret M, Boue-Grabot E. Functional properties of internalizationdeficient P2X4 receptors reveal a novel mechanism of ligand-gated channel facilitation by ivermectin. Mol Pharmacol 2006, 69: 576-587.

[23] Robinson LE, Murrell-Lagnado RD. The trafficking and targeting of P2X receptors. Front Cell Neurosci 2013, 7: 233.

[24] Murrell-Lagnado RD. A role for P2X4 receptors in lysosome function. J Gen Physiol 2018, 150: 185-187.

[25] Burnstock G. Purinergic signalling and disorders of the central nervous system. Nat Rev Drug Discov 2008, 7: 575-590.

[26] Brake AJ, Wagenbach MJ, Julius D. New structural motif for ligand-gated ion channels defined by an ionotropic ATP receptor. Nature 1994, 371: 519-523.

[27] Kawate T, Michel JC, Birdsong WT, Gouaux E. Crystal structure of the ATP-gated P2X(4) ion channel in the closed state. Nature 2009, 460: 592-598.

[28] Hattori M, Gouaux E. Molecular mechanism of ATP binding and ion channel activation in P2X receptors. Nature 2012, 485: 207-212.

[29] Mansoor SE, Lu W, Oosterheert W, Shekhar M, Tajkhorshid E, Gouaux E. X-ray structures define human P2X(3) receptor gating cycle and antagonist action. Nature 2016, 538: 66-71.

[30] Igawa T, Abe Y, Tsuda M, Inoue K, Ueda T. Solution structure of the rat P2X4 receptor head domain involved in inhibitory metal binding. FEBS Lett 2015, 589: 680-686.

[31] Chaumont S, Jiang LH, Penna A, North RA, Rassendren F. Identification of a trafficking motif involved in the stabilization and polarization of P2X receptors. J Biol Chem 2004, 279: 29628-29638.

[32] Boue-Grabot E, Archambault V, Seguela P. A protein kinase C site highly conserved in P2X subunits controls the desensitization kinetics of P2X(2) ATP-gated channels. J Biol Chem 2000, 275: 10190-10195.

[33] Qureshi OS, Paramasivam A, Yu JC, Murrell-Lagnado RD. Regulation of P2X4 receptors by lysosomal targeting, glycan protection and exocytosis. J Cell Sci 2007, 120: 3838-3849.

[34] Bobanovic LK, Royle SJ, Murrell-Lagnado RD. P2X receptor trafficking in neurons is subunit specific. J Neurosci 2002, 22: 4814-4824.

[35] Royle SJ, Bobanovic LK, Murrell-Lagnado RD. Identification of a non-canonical tyrosinebased endocytic motif in an ionotropic receptor. J Biol Chem 2002, 277: 35378-35385.

Neuroscience Bulletin

3
4
5
c
6
7
8
9
10
11
11
12
13
14
15
16
17
10
18
19
20
21
22
23
23
24
25
26
27
28
20
29
30
31
32
33
34
25
22
30
37
38
39
40
Δ1
47
42
43
44
45
46
17
40
48
49
50
51
52
52
54
55
56
57
58
50
59
0U

[36] Jo YH, Donier E, Martinez A, Garret M, Toulme E, Boue-Grabot E. Cross-talk between P2X4 and gamma-aminobutyric acid, type A receptors determines synaptic efficacy at a central synapse.J Biol Chem 2011, 286: 19993-20004.

[37] Boumechache M, Masin M, Edwardson JM, Gorecki DC, Murrell-Lagnado R. Analysis of assembly and trafficking of native P2X4 and P2X7 receptor complexes in rodent immune cells. J Biol Chem 2009, 284: 13446-13454.

[38] Royle SJ, Qureshi OS, Bobanovic LK, Evans PR, Owen DJ, Murrell-Lagnado RD. Noncanonical YXXGPhi endocytic motifs: recognition by AP2 and preferential utilization in P2X4 receptors. J Cell Sci 2005, 118: 3073-3080.

[39] Xu J, Chai H, Ehinger K, Egan TM, Srinivasan R, Frick M, *et al.* Imaging P2X4 receptor subcellular distribution, trafficking, and regulation using P2X4-pHluorin. J Gen Physiol 2014, 144: 81-104.

[40] Toulme E, Garcia A, Samways D, Egan TM, Carson MJ, Khakh BS. P2X4 receptors in activated C8-B4 cells of cerebellar microglial origin. J Gen Physiol 2010, 135: 333-353.

[41] Stokes L, Surprenant A. Dynamic regulation of the P2X4 receptor in alveolar macrophages by phagocytosis and classical activation. Eur J Immunol 2009, 39: 986-995.

[42] Trang T, Salter MW. P2X4 purinoceptor signaling in chronic pain. Purinergic Signal 2012,8: 621-628.

[43] Murrell-Lagnado RD, Frick M. P2X4 and lysosome fusion. Curr Opin Pharmacol 2019, 47:126-132.

[44] Zhang Z, Chen G, Zhou W, Song A, Xu T, Luo Q, *et al.* Regulated ATP release from astrocytes through lysosome exocytosis. Nat Cell Biol 2007, 9: 945-953.

[45] Besnard A, Gautherot J, Julien B, Tebbi A, Garcin I, Doignon I, *et al.* The P2X4 purinergic receptor impacts liver regeneration after partial hepatectomy in mice through the regulation of biliary homeostasis. Hepatology 2016, 64: 941-953.

[46] Cao Q, Zhong XZ, Zou Y, Murrell-Lagnado R, Zhu MX, Dong XP. Calcium release throughP2X4 activates calmodulin to promote endolysosomal membrane fusion. J Cell Biol 2015, 209:879-894.

[47] Bo X, Kim M, Nori SL, Schoepfer R, Burnstock G, North RA. Tissue distribution of P2X4 receptors studied with an ectodomain antibody. Cell Tissue Res 2003, 313: 159-165.

[48] Seguela P, Haghighi A, Soghomonian JJ, Cooper E. A novel neuronal P2x ATP receptor ion channel with widespread distribution in the brain. J Neurosci 1996, 16: 448-455.

[49] Lê KT, Villeneuve P, Ramjaun AR, McPherson PS, Beaudet A, Seguela P. Sensory presynaptic and widespread somatodendritic immunolocalization of central ionotropic P2X ATP receptors. Neuroscience 1998, 83: 177-190.

[50] Amadio S, Montilli C, Picconi B, Calabresi P, Volonte C. Mapping P2X and P2Y receptor proteins in striatum and substantia nigra: An immunohistological study. Purinergic Signal 2007, 3: 389-398.

[51] Stojilkovic SS. Purinergic regulation of hypothalamopituitary functions. Trends Endocrinol Metab 2009, 20: 460-468.

[52] Lalo U, Verkhratsky A, Pankratov Y. Ivermectin potentiates ATP-induced ion currents in cortical neurones: evidence for functional expression of P2X4 receptors? Neurosci Lett 2007, 421: 158-162.

[53] Tan Y, Zhao B, Zeng QC, Shi CM, Zhao FB, Li ZW. Characteristics of ATP-activated current in nodose ganglion neurons of rats. Neurosci Lett 2009, 459: 25-29.

[54] Luo J, Yin GF, Gu YZ, Liu Y, Dai JP, Li C, *et al.* Characterization of three types of ATPactivated current in relation to P2X subunits in rat trigeminal ganglion neurons. Brain Res 2006, 1115: 9-15.

[55] Ito K, Chihara Y, Iwasaki S, Komuta Y, Sugasawa M, Sahara Y. Functional ligand-gated purinergic receptors (P2X) in rat vestibular ganglion neurons. Hear Res 2010, 267: 89-95.

[56] Wheeler-Schilling TH, Marquordt K, Kohler K, Guenther E, Jabs R. Identification of purinergic receptors in retinal ganglion cells. Brain Res Mol Brain Res 2001, 92: 177-180.

[57] Sim JA, Chaumont S, Jo J, Ulmann L, Young MT, Cho K, *et al.* Altered hippocampal synaptic potentiation in P2X4 knock-out mice. J Neurosci 2006, 26: 9006-9009.

[58] Xu J, Bernstein AM, Wong A, Lu XH, Khoja S, Yang XW, *et al.* P2X4 Receptor Reporter Mice: Sparse Brain Expression and Feeding-Related Presynaptic Facilitation in the Arcuate Nucleus. J Neurosci 2016, 36: 8902-8920.

[59] Bertin E, Deluc T, Pilch KS, Martinez A, Pougnet JT, Doudnikoff E, *et al.* Increased surfaceP2X4 receptor regulates anxiety and memory in P2X4 internalization-defective knock-in mice.Mol Psychiatry 2020.

[60] Le KT, Babinski K, Seguela P. Central P2X4 and P2X6 channel subunits coassemble into a novel heteromeric ATP receptor. J Neurosci 1998, 18: 7152-7159.

[61] Antonio LS, Stewart AP, Varanda WA, Edwardson JM. Identification of P2X2/P2X4/P2X6 heterotrimeric receptors using atomic force microscopy (AFM) imaging. FEBS Lett 2014, 588: 2125-2128.

[62] Lalo U, Palygin O, Verkhratsky A, Grant SG, Pankratov Y. ATP from synaptic terminals and astrocytes regulates NMDA receptors and synaptic plasticity through PSD-95 multi-protein complex. Sci Rep 2016, 6: 33609.

[63] Kessels HW, Malinow R. Synaptic AMPA receptor plasticity and behavior. Neuron 2009, 61: 340-350.

[64] Pankratov Y, Lalo U, Krishtal OA, Verkhratsky A. P2X receptors and synaptic plasticity. Neuroscience 2009, 158: 137-148.

[65] Baxter AW, Choi SJ, Sim JA, North RA. Role of P2X4 receptors in synaptic strengthening in mouse CA1 hippocampal neurons. Eur J Neurosci 2011, 34: 213-220.

[66] Jo YH, Boue-Grabot E. Interplay between ionotropic receptors modulates inhibitory synaptic strength. Commun Integr Biol 2011, 4: 706-709.

[67] Lalo U, Palygin O, Rasooli-Nejad S, Andrew J, Haydon PG, Pankratov Y. Exocytosis of ATP from astrocytes modulates phasic and tonic inhibition in the neocortex. PLoS Biol 2014, 12: e1001747.

[68] Guthrie PB, Knappenberger J, Segal M, Bennett MV, Charles AC, Kater SB. ATP released from astrocytes mediates glial calcium waves. J Neurosci 1999, 19: 520-528.

[69] Craigie E, Birch RE, Unwin RJ, Wildman SS. The relationship between P2X4 and P2X7: a physiologically important interaction? Front Physiol 2013, 4: 216.

[70] Kukley M, Barden JA, Steinhauser C, Jabs R. Distribution of P2X receptors on astrocytes in juvenile rat hippocampus. Glia 2001, 36: 11-21.

[71] Franke H, Grosche J, Schadlich H, Krugel U, Allgaier C, Illes P. P2X receptor expression on astrocytes in the nucleus accumbens of rats. Neuroscience 2001, 108: 421-429.

[72] Stokes L, Layhadi JA, Bibic L, Dhuna K, Fountain SJ. P2X4 Receptor Function in the Nervous System and Current Breakthroughs in Pharmacology. Front Pharmacol 2017, 8: 291.

[73] Tan YL, Yuan Y, Tian L. Microglial regional heterogeneity and its role in the brain. Mol Psychiatry 2020, 25: 351-367.

[74] Marques S, Zeisel A, Codeluppi S, van Bruggen D, Mendanha Falcao A, Xiao L, *et al.* Oligodendrocyte heterogeneity in the mouse juvenile and adult central nervous system. Science 2016, 352: 1326-1329.

[75] John Lin CC, Yu K, Hatcher A, Huang TW, Lee HK, Carlson J, *et al.* Identification of diverse astrocyte populations and their malignant analogs. Nat Neurosci 2017, 20: 396-405.

[76] Deng Z, Li C, Du E, Liu C, Xia B, Chen H, *et al.* Catestatin Enhances Neuropathic Pain Mediated by P2X4 Receptor of Dorsal Root Ganglia in a Rat Model of Chronic Constriction Injury. Cell Physiol Biochem 2018, 51: 812-826.

Neuroscience Bulletin

[77] Teixeira JM, Dos Santos GG, Neves AF, Athie MCP, Bonet IJM, Nishijima CM, *et al.* Diabetes-induced Neuropathic Mechanical Hyperalgesia Depends on P2X4 Receptor Activation in Dorsal Root Ganglia. Neuroscience 2019, 398: 158-170.

[78] Agresti C, Meomartini ME, Amadio S, Ambrosini E, Volonte C, Aloisi F, *et al.* ATP regulates oligodendrocyte progenitor migration, proliferation, and differentiation: involvement of metabotropic P2 receptors. Brain Res Brain Res Rev 2005, 48: 157-165.

[79] Matute C, Torre I, Perez-Cerda F, Perez-Samartin A, Alberdi E, Etxebarria E, *et al.* P2X(7) receptor blockade prevents ATP excitotoxicity in oligodendrocytes and ameliorates experimental autoimmune encephalomyelitis. J Neurosci 2007, 27: 9525-9533.

[80] Su WF, Wu F, Jin ZH, Gu Y, Chen YT, Fei Y, *et al.* Overexpression of P2X4 receptor in Schwann cells promotes motor and sensory functional recovery and remyelination via BDNF secretion after nerve injury. Glia 2019, 67: 78-90.

[81] Todd AJ. Neuronal circuitry for pain processing in the dorsal horn. Nat Rev Neurosci 2010,11: 823-836.

[82] Basbaum AI, Bautista DM, Scherrer G, Julius D. Cellular and molecular mechanisms of pain.Cell 2009, 139: 267-284.

[83] Ji RR. Recent Progress in Understanding the Mechanisms of Pain and Itch: the Second Special Issue. Neurosci Bull 2018, 34: 1-3.

[84] Inoue K. Role of the P2X4 receptor in neuropathic pain. Curr Opin Pharmacol 2019, 47: 33-39.

[85] Gilmore SA. Proliferation of non-neuronal cells in spinal cords of irradiated, immature rats following transection of the sciatic nerve. Anat Rec 1975, 181: 799-811.

[86] Gilmore SA, Skinner RD. Intraspinal non-neuronal cellular responses to peripheral nerve injury. Anat Rec 1979, 194: 369-387.

[87] Masuda J, Tsuda M, Tozaki-Saitoh H, Inoue K. Intrathecal delivery of PDGF produces tactile allodynia through its receptors in spinal microglia. Mol Pain 2009, 5: 23.

[88] Tanga FY, Nutile-McMenemy N, DeLeo JA. The CNS role of Toll-like receptor 4 in innate neuroimmunity and painful neuropathy. Proc Natl Acad Sci U S A 2005, 102: 5856-5861.

[89] Tsuda M, Shigemoto-Mogami Y, Koizumi S, Mizokoshi A, Kohsaka S, Salter MW, *et al.*P2X4 receptors induced in spinal microglia gate tactile allodynia after nerve injury. Nature 2003, 424: 778-783.

[90] Tsuda M, Toyomitsu E, Kometani M, Tozaki-Saitoh H, Inoue K. Mechanisms underlying fibronectin-induced up-regulation of P2X4R expression in microglia: distinct roles of PI3K-Akt and MEK-ERK signalling pathways. J Cell Mol Med 2009, 13: 3251-3259.

[91] Ulmann L, Hatcher JP, Hughes JP, Chaumont S, Green PJ, Conquet F, *et al.* Up-regulation of P2X4 receptors in spinal microglia after peripheral nerve injury mediates BDNF release and neuropathic pain. J Neurosci 2008, 28: 11263-11268.

[92] Coull JA, Beggs S, Boudreau D, Boivin D, Tsuda M, Inoue K, *et al.* BDNF from microglia causes the shift in neuronal anion gradient underlying neuropathic pain. Nature 2005, 438: 1017-1021.

[93] Trang T, Beggs S, Wan X, Salter MW. P2X4-receptor-mediated synthesis and release of brain-derived neurotrophic factor in microglia is dependent on calcium and p38-mitogen-activated protein kinase activation. J Neurosci 2009, 29: 3518-3528.

[94] Sorge RE, Mapplebeck JC, Rosen S, Beggs S, Taves S, Alexander JK, *et al.* Different immune cells mediate mechanical pain hypersensitivity in male and female mice. Nat Neurosci 2015, 18: 1081-1083.

[95] Nakatsuka T, Gu JG. ATP P2X receptor-mediated enhancement of glutamate release and evoked EPSCs in dorsal horn neurons of the rat spinal cord. J Neurosci 2001, 21: 6522-6531.

[96] Jo YH, Schlichter R. Synaptic corelease of ATP and GABA in cultured spinal neurons. Nat Neurosci 1999, 2: 241-245.

[97] Masuda T, Ozono Y, Mikuriya S, Kohro Y, Tozaki-Saitoh H, Iwatsuki K, *et al.* Dorsal horn neurons release extracellular ATP in a VNUT-dependent manner that underlies neuropathic pain. Nat Commun 2016, 7: 12529.

[98] Boue-Grabot E, Toulme E, Emerit MB, Garret M. Subunit-specific coupling between gammaaminobutyric acid type A and P2X2 receptor channels. J Biol Chem 2004, 279: 52517-52525.

[99] Masuda T, Tsuda M, Yoshinaga R, Tozaki-Saitoh H, Ozato K, Tamura T, *et al.* IRF8 is a critical transcription factor for transforming microglia into a reactive phenotype. Cell Rep 2012, 1: 334-340.

[100] Masuda T, Iwamoto S, Yoshinaga R, Tozaki-Saitoh H, Nishiyama A, Mak TW, *et al.* Transcription factor IRF5 drives P2X4R+-reactive microglia gating neuropathic pain. Nat Commun 2014, 5: 3771.

[101] Tsuda M, Toyomitsu E, Komatsu T, Masuda T, Kunifusa E, Nasu-Tada K, *et al.* Fibronectin/integrin system is involved in P2X(4) receptor upregulation in the spinal cord and neuropathic pain after nerve injury. Glia 2008, 56: 579-585.

[102] Toyomitsu E, Tsuda M, Yamashita T, Tozaki-Saitoh H, Tanaka Y, Inoue K. CCL2 promotes P2X4 receptor trafficking to the cell surface of microglia. Purinergic Signal 2012, 8: 301-310.

[103] Mapplebeck JC, Beggs S, Salter MW. Sex differences in pain: a tale of two immune cells.Pain 2016, 157 Suppl 1: S2-6.

[104] Tsuda M. Modulation of Pain and Itch by Spinal Glia. Neurosci Bull 2018, 34: 178-185.

[105] Jin XH, Wang LN, Zuo JL, Yang JP, Liu SL. P2X4 receptor in the dorsal horn partially contributes to brain-derived neurotrophic factor oversecretion and toll-like receptor-4 receptor activation associated with bone cancer pain. J Neurosci Res 2014, 92: 1690-1702.

[106] Collo G, North RA, Kawashima E, Merlo-Pich E, Neidhart S, Surprenant A, *et al.* Cloning OF P2X5 and P2X6 receptors and the distribution and properties of an extended family of ATP-gated ion channels. J Neurosci 1996, 16: 2495-2507.

[107] Toulme E, Tsuda M, Khakh BS, Inoue K. On the Role of ATP-Gated P2X Receptors in Acute, Inflammatory and Neuropathic Pain. Translational Pain Research: From Mouse to Man 2010.

[108] Usoskin D, Furlan A, Islam S, Abdo H, Lonnerberg P, Lou D, *et al.* Unbiased classification of sensory neuron types by large-scale single-cell RNA sequencing. Nat Neurosci 2015, 18: 145-153.

[109] Lalisse S, Hua J, Lenoir M, Linck N, Rassendren F, Ulmann L. Sensory neuronal P2RX4 receptors controls BDNF signaling in inflammatory pain. Sci Rep 2018, 8: 964.

[110] Kobayashi K, Yamanaka H, Noguchi K. Expression of ATP receptors in the rat dorsal root ganglion and spinal cord. Anat Sci Int 2013, 88: 10-16.

[111] Ying M, Liu H, Zhang T, Jiang C, Gong Y, Wu B, *et al.* Effect of artemisinin on neuropathic pain mediated by P2X4 receptor in dorsal root ganglia. Neurochem Int 2017, 108: 27-33.

[112] Kushnir R, Cherkas PS, Hanani M. Peripheral inflammation upregulates P2X receptor expression in satellite glial cells of mouse trigeminal ganglia: a calcium imaging study. Neuropharmacology 2011, 61: 739-746.

[113] Yuan H, Ouyang S, Yang R, Li S, Gong Y, Zou L, *et al.* Osthole alleviated diabetic neuropathic pain mediated by the P2X4 receptor in dorsal root ganglia. Brain Res Bull 2018, 142: 289-296.

[114] Zhang H, Mei X, Zhang P, Ma C, White FA, Donnelly DF, *et al.* Altered functional properties of satellite glial cells in compressed spinal ganglia. Glia 2009, 57: 1588-1599.

[115] Hanani M, Blum E, Liu S, Peng L, Liang S. Satellite glial cells in dorsal root ganglia are activated in streptozotocin-treated rodents. J Cell Mol Med 2014, 18: 2367-2371.

[116] Ulmann L, Hirbec H, Rassendren F. P2X4 receptors mediate PGE2 release by tissue-resident macrophages and initiate inflammatory pain. EMBO J 2010, 29: 2290-2300.

[117] Aby F, Whitestone S, Landry M, Ulmann L, Fossat P. Inflammatory-induced spinal dorsal horn neurons hyperexcitability is mediated by P2X4 receptors. Pain Rep 2018, 3: e660.

[118] Zhang W, Liu LY, Xu TL. Reduced potassium-chloride co-transporter expression in spinal cord dorsal horn neurons contributes to inflammatory pain hypersensitivity in rats. Neuroscience 2008, 152: 502-510.

[119] Zhao S, Zhou Y, Fan Y, Gong Y, Yang J, Yang R, *et al.* Involvement of purinergic 2X4 receptor in glycoprotein 120-induced pyroptosis in dorsal root ganglia. J Neurochem 2019, 151: 584-594.

[120] Dodick D, Silberstein S. Central sensitization theory of migraine: clinical implications. Headache 2006, 46 Suppl 4: S182-191.

[121] Long T, He W, Pan Q, Zhang S, Zhang Y, Liu C, *et al.* Microglia P2X4 receptor contributes to central sensitization following recurrent nitroglycerin stimulation. J Neuroinflammation 2018, 15: 245.

[122] Martins LB, Teixeira AL, Domingues RB. Neurotrophins and Migraine. Vitam Horm 2017, 104: 459-473.

[123] Long T, He W, Pan Q, Zhang S, Zhang D, Qin G, *et al.* Microglia P2X4R-BDNF signalling contributes to central sensitization in a recurrent nitroglycerin-induced chronic migraine model. J Headache Pain 2020, 21: 4.

[124] Cavaliere F, Florenzano F, Amadio S, Fusco FR, Viscomi MT, D'Ambrosi N, *et al.* Upregulation of P2X2, P2X4 receptor and ischemic cell death: prevention by P2 antagonists. Neuroscience 2003, 120: 85-98.

[125] Cheng RD, Ren JJ, Zhang YY, Ye XM. P2X4 receptors expressed on microglial cells in post-ischemic inflammation of brain ischemic injury. Neurochem Int 2014, 67: 9-13.

[126] Verma R, Cronin CG, Hudobenko J, Venna VR, McCullough LD, Liang BT. Deletion of the P2X4 receptor is neuroprotective acutely, but induces a depressive phenotype during recovery from ischemic stroke. Brain Behav Immun 2017, 66: 302-312.

[127] Vazquez-Villoldo N, Domercq M, Martin A, Llop J, Gomez-Vallejo V, Matute C. P2X4 receptors control the fate and survival of activated microglia. Glia 2014, 62: 171-184.

[128] Li F, Wang L, Li JW, Gong M, He L, Feng R, *et al.* Hypoxia induced amoeboid microglial cell activation in postnatal rat brain is mediated by ATP receptor P2X4. BMC Neurosci 2011, 12: 111.

[129] Ohsawa K, Irino Y, Nakamura Y, Akazawa C, Inoue K, Kohsaka S. Involvement of P2X4 and P2Y12 receptors in ATP-induced microglial chemotaxis. Glia 2007, 55: 604-616.

[130] Ozaki T, Muramatsu R, Sasai M, Yamamoto M, Kubota Y, Fujinaka T, *et al.* The P2X4 receptor is required for neuroprotection via ischemic preconditioning. Sci Rep 2016, 6: 25893.

[131] Kang TC, An SJ, Park SK, Hwang IK, Won MH. P2X2 and P2X4 receptor expression is regulated by a GABA(A) receptor-mediated mechanism in the gerbil hippocampus. Brain Res Mol Brain Res 2003, 116: 168-175.

[132] Ulmann L, Levavasseur F, Avignone E, Peyroutou R, Hirbec H, Audinat E, *et al.*Involvement of P2X4 receptors in hippocampal microglial activation after status epilepticus. Glia
2013, 61: 1306-1319.

[133] Murphy MP, LeVine H, 3rd. Alzheimer's disease and the amyloid-beta peptide. J Alzheimers Dis 2010, 19: 311-323.

[134] Sadigh-Eteghad S, Sabermarouf B, Majdi A, Talebi M, Farhoudi M, Mahmoudi J. Amyloidbeta: a crucial factor in Alzheimer's disease. Med Princ Pract 2015, 24: 1-10.

[135] Erb L, Woods LT, Khalafalla MG, Weisman GA. Purinergic signaling in Alzheimer's disease. Brain Res Bull 2019, 151: 25-37.

[136] Saez-Orellana F, Godoy PA, Bastidas CY, Silva-Grecchi T, Guzman L, Aguayo LG, *et al.* ATP leakage induces P2XR activation and contributes to acute synaptic excitotoxicity induced by soluble oligomers of beta-amyloid peptide in hippocampal neurons. Neuropharmacology 2016, 100: 116-123.

[137] Godoy PA, Ramirez-Molina O, Fuentealba J. Exploring the Role of P2X Receptors in Alzheimer's Disease. Front Pharmacol 2019, 10: 1330.

[138] Varma R, Chai Y, Troncoso J, Gu J, Xing H, Stojilkovic SS, *et al.* Amyloid-beta induces a caspase-mediated cleavage of P2X4 to promote purinotoxicity. Neuromolecular Med 2009, 11: 63-75.

[139] Martin E, Amar M, Dalle C, Youssef I, Boucher C, Le Duigou C, *et al.* New role of P2X7 receptor in an Alzheimer's disease mouse model. Mol Psychiatry 2019, 24: 108-125.

[140] Martinez-Frailes C, Di Lauro C, Bianchi C, de Diego-Garcia L, Sebastian-Serrano A, Bosca L, et al. Amyloid Peptide Induced Neuroinflammation Increases the P2X7 Receptor Expression in Microglial Cells, Impacting on Its Functionality. Front Cell Neurosci 2019, 13: 143.

[141] Sperlagh B, Illes P. P2X7 receptor: an emerging target in central nervous system diseases. Trends Pharmacol Sci 2014, 35: 537-547.

[142] McGeer PL, Rogers J, McGeer EG. Inflammation, Antiinflammatory Agents, and Alzheimer's Disease: The Last 22 Years. J Alzheimers Dis 2016, 54: 853-857.

[143] Weinhold K, Krause-Buchholz U, Rodel G, Kasper M, Barth K. Interaction and interrelation of P2X7 and P2X4 receptor complexes in mouse lung epithelial cells. Cell Mol Life Sci 2010, 67: 2631-2642.

[144] Li F, Guo N, Ma Y, Ning B, Wang Y, Kou L. Inhibition of P2X4 suppresses joint inflammation and damage in collagen-induced arthritis. Inflammation 2014, 37: 146-153.

[145] Layhadi JA, Turner J, Crossman D, Fountain SJ. ATP Evokes Ca(2+) Responses and CXCL5 Secretion via P2X4 Receptor Activation in Human Monocyte-Derived Macrophages. J Immunol 2018, 200: 1159-1168.

[146] Andries M, Van Damme P, Robberecht W, Van Den Bosch L. Ivermectin inhibits AMPA receptor-mediated excitotoxicity in cultured motor neurons and extends the life span of a transgenic mouse model of amyotrophic lateral sclerosis. Neurobiol Dis 2007, 25: 8-16.

[147] Boue-Grabot E, Emerit MB, Toulme E, Seguela P, Garret M. Cross-talk and co-trafficking between rho1/GABA receptors and ATP-gated channels. J Biol Chem 2004, 279: 6967-6975.

[148] Gordon GR, Baimoukhametova DV, Hewitt SA, Rajapaksha WR, Fisher TE, Bains JS.
Norepinephrine triggers release of glial ATP to increase postsynaptic efficacy. Nat Neurosci 2005, 8: 1078-1086.

[149] Gu JG, MacDermott AB. Activation of ATP P2X receptors elicits glutamate release from sensory neuron synapses. Nature 1997, 389: 749-753.

[150] Pankratov YV, Lalo UV, Krishtal OA. Role for P2X receptors in long-term potentiation. J Neurosci 2002, 22: 8363-8369.

[151] Krugel U, Kittner H, Franke H, Illes P. Purinergic modulation of neuronal activity in the mesolimbic dopaminergic system in vivo. Synapse 2003, 47: 134-142.

[152] Xiao C, Zhou C, Li K, Davies DL, Ye JH. Purinergic type 2 receptors at GABAergic synapses on ventral tegmental area dopamine neurons are targets for ethanol action. J Pharmacol Exp Ther 2008, 327: 196-205.

[153] Zhou FC, Zhang JK, Lumeng L, Li TK. Mesolimbic dopamine system in alcohol-preferring rats. Alcohol 1995, 12: 403-412.

[154] Ralph RJ, Paulus MP, Fumagalli F, Caron MG, Geyer MA. Prepulse inhibition deficits and perseverative motor patterns in dopamine transporter knock-out mice: differential effects of D1 and D2 receptor antagonists. J Neurosci 2001, 21: 305-313.

Neuroscience Bulletin

[155] Khoja S, Shah V, Garcia D, Asatryan L, Jakowec MW, Davies DL. Role of purinergic P2X4 receptors in regulating striatal dopamine homeostasis and dependent behaviors. J Neurochem 2016, 139: 134-148.

[156] Nagatsu T, Mogi M, Ichinose H, Togari A. Cytokines in Parkinson's disease. J Neural Transm Suppl 2000: 143-151.

[157] Imamura K, Hishikawa N, Sawada M, Nagatsu T, Yoshida M, Hashizume Y. Distribution of major histocompatibility complex class II-positive microglia and cytokine profile of Parkinson's disease brains. Acta Neuropathol 2003, 106: 518-526.

[158] Mount MP, Lira A, Grimes D, Smith PD, Faucher S, Slack R, *et al.* Involvement of interferon-gamma in microglial-mediated loss of dopaminergic neurons. J Neurosci 2007, 27: 3328-3337.

[159] Doorn KJ, Moors T, Drukarch B, van de Berg W, Lucassen PJ, van Dam AM. Microglial phenotypes and toll-like receptor 2 in the substantia nigra and hippocampus of incidental Lewy body disease cases and Parkinson's disease patients. Acta Neuropathol Commun 2014, 2: 90.

[160] Saijo K, Glass CK. Microglial cell origin and phenotypes in health and disease. Nat Rev Immunol 2011, 11: 775-787.

[161] Joers V, Tansey MG, Mulas G, Carta AR. Microglial phenotypes in Parkinson's disease and animal models of the disease. Prog Neurobiol 2017, 155: 57-75.

[162] Wang XH, Xie X, Luo XG, Shang H, He ZY. Inhibiting purinergic P2X7 receptors with the antagonist brilliant blue G is neuroprotective in an intranigral lipopolysaccharide animal model of Parkinson's disease. Mol Med Rep 2017, 15: 768-776.

[163] Crabbe M, Van der Perren A, Bollaerts I, Kounelis S, Baekelandt V, Bormans G, *et al.* Increased P2X7 Receptor Binding Is Associated With Neuroinflammation in Acute but Not Chronic Rodent Models for Parkinson's Disease. Front Neurosci 2019, 13: 799.

> [164] Le W, Wu J, Tang Y. Protective Microglia and Their Regulation in Parkinson's Disease. Front Mol Neurosci 2016, 9: 89.

> [165] Volonte C, Apolloni S, Parisi C, Amadio S. Purinergic contribution to amyotrophic lateral sclerosis. Neuropharmacology 2016, 104: 180-193.

[166] Casanovas A, Hernandez S, Tarabal O, Rossello J, Esquerda JE. Strong P2X4 purinergic receptor-like immunoreactivity is selectively associated with degenerating neurons in transgenic rodent models of amyotrophic lateral sclerosis. J Comp Neurol 2008, 506: 75-92.

[167] Hernandez S, Casanovas A, Piedrafita L, Tarabal O, Esquerda JE. Neurotoxic species of misfolded SOD1G93A recognized by antibodies against the P2X4 subunit of the ATP receptor accumulate in damaged neurons of transgenic animal models of amyotrophic lateral sclerosis. J Neuropathol Exp Neurol 2010, 69: 176-187.

[168] D'Ambrosi N, Finocchi P, Apolloni S, Cozzolino M, Ferri A, Padovano V, *et al.* The proinflammatory action of microglial P2 receptors is enhanced in SOD1 models for amyotrophic lateral sclerosis. J Immunol 2009, 183: 4648-4656.

[169] Liu J, Wang F. Role of Neuroinflammation in Amyotrophic Lateral Sclerosis: Cellular Mechanisms and Therapeutic Implications. Front Immunol 2017, 8: 1005.

[170] Cieslak M, Roszek K, Wujak M. Purinergic implication in amyotrophic lateral sclerosisfrom pathological mechanisms to therapeutic perspectives. Purinergic Signal 2019, 15: 1-15.

[171] Fabbrizio P, Amadio S, Apolloni S, Volonte C. P2X7 Receptor Activation Modulates Autophagy in SOD1-G93A Mouse Microglia. Front Cell Neurosci 2017, 11: 249.

[172] Apolloni S, Amadio S, Montilli C, Volonte C, D'Ambrosi N. Ablation of P2X7 receptor exacerbates gliosis and motoneuron death in the SOD1-G93A mouse model of amyotrophic lateral sclerosis. Hum Mol Genet 2013, 22: 4102-4116.

[173] Sluyter R, Bartlett R, Ly D, Yerbury JJ. P2X7 receptor antagonism in amyotrophic lateral sclerosis. Neural Regen Res 2017, 12: 749-750.

[174] Kawano A, Tsukimoto M, Mori D, Noguchi T, Harada H, Takenouchi T, *et al.* Regulation of P2X7-dependent inflammatory functions by P2X4 receptor in mouse macrophages. Biochem Biophys Res Commun 2012, 420: 102-107.

[175] Dendrou CA, Fugger L, Friese MA. Immunopathology of multiple sclerosis. Nat Rev Immunol 2015, 15: 545-558.

[176] Zabala A, Vazquez-Villoldo N, Rissiek B, Gejo J, Martin A, Palomino A, *et al.* P2X4receptor controls microglia activation and favors remyelination in autoimmune encephalitis.EMBO Mol Med 2018, 10.

[177] Feldmann A, Amphornrat J, Schonherr M, Winterstein C, Mobius W, Ruhwedel T, *et al.*Transport of the major myelin proteolipid protein is directed by VAMP3 and VAMP7. J Neurosci 2011, 31: 5659-5672.

[178] Chen G, Zhang Z, Wei Z, Cheng Q, Li X, Li W, *et al.* Lysosomal exocytosis in Schwann cells contributes to axon remyelination. Glia 2012, 60: 295-305.

[179] Fois G, Winkelmann VE, Bareis L, Staudenmaier L, Hecht E, Ziller C, *et al.* ATP is stored in lamellar bodies to activate vesicular P2X4 in an autocrine fashion upon exocytosis. J Gen Physiol 2018, 150: 277-291.

[180] Yardley MM, Wyatt L, Khoja S, Asatryan L, Ramaker MJ, Finn DA, *et al.* Ivermectin reduces alcohol intake and preference in mice. Neuropharmacology 2012, 63: 190-201.

[181] Wyatt LR, Finn DA, Khoja S, Yardley MM, Asatryan L, Alkana RL, *et al.* Contribution of P2X4 receptors to ethanol intake in male C57BL/6 mice. Neurochem Res 2014, 39: 1127-1139.

[182] Franklin KM, Asatryan L, Jakowec MW, Trudell JR, Bell RL, Davies DL. P2X4 receptors (P2X4Rs) represent a novel target for the development of drugs to prevent and/or treat alcohol use disorders. Front Neurosci 2014, 8: 176.

[183] Franklin KM, Hauser SR, Lasek AW, Bell RL, McBride WJ. Involvement of PurinergicP2X4 Receptors in Alcohol Intake of High-Alcohol-Drinking (HAD) Rats. Alcohol Clin Exp Res2015, 39: 2022-2031.

[184] Kimpel MW, Strother WN, McClintick JN, Carr LG, Liang T, Edenberg HJ, *et al.* Functional gene expression differences between inbred alcohol-preferring and -non-preferring rats in five brain regions. Alcohol 2007, 41: 95-132.

[185] Asatryan L, Nam HW, Lee MR, Thakkar MM, Saeed Dar M, Davies DL, *et al.* Implication of the purinergic system in alcohol use disorders. Alcohol Clin Exp Res 2011, 35: 584-594.

[186] Davies DL, Kuo ST, Alkana RL. Differential effects of propofol and ethanol on P2X4 receptors expressed in Xenopus oocytes. Int Congr Ser 2005, 1283: 285-287.

[187] Ostrovskaya O, Asatryan L, Wyatt L, Popova M, Li K, Peoples RW, *et al.* Ethanol is a fast channel inhibitor of P2X4 receptors. J Pharmacol Exp Ther 2011, 337: 171-179.

[188] Asatryan L, Popova M, Perkins D, Trudell JR, Alkana RL, Davies DL. Ivermectin antagonizes ethanol inhibition in purinergic P2X4 receptors. J Pharmacol Exp Ther 2010, 334: 720-728.

[189] Potula R, Haorah J, Knipe B, Leibhart J, Chrastil J, Heilman D, *et al.* Alcohol abuse enhances neuroinflammation and impairs immune responses in an animal model of human immunodeficiency virus-1 encephalitis. Am J Pathol 2006, 168: 1335-1344.

[190] Gofman L, Cenna JM, Potula R. P2X4 receptor regulates alcohol-induced responses in microglia. J Neuroimmune Pharmacol 2014, 9: 668-678.

[191] Gofman L, Fernandes NC, Potula R. Relative Role of Akt, ERK and CREB in Alcohol-Induced Microglia P2X4R Receptor Expression. Alcohol Alcohol 2016, 51: 647-654.

[192] Wixey JA, Reinebrant HE, Carty ML, Buller KM. Delayed P2X4R expression after hypoxiaischemia is associated with microglia in the immature rat brain. J Neuroimmunol 2009, 212: 35-43.

[193] Bortolato M, Yardley MM, Khoja S, Godar SC, Asatryan L, Finn DA, *et al.* Pharmacological insights into the role of P2X4 receptors in behavioural regulation: lessons from ivermectin. Int J Neuropsychopharmacol 2013, 16: 1059-1070.

[194] Khoja S, Asatryan L, Jakowec MW, Davies DL. Dopamine Receptor Blockade Attenuates Purinergic P2X4 Receptor-Mediated Prepulse Inhibition Deficits and Underlying Molecular Mechanisms. Front Cell Neurosci 2019, 13: 331.

Figure Legends

Figure 1: P2X4 expression in CNS diseases

Left, P2X4 in normal states is mostly intracellular with low surface expression at the surface of neurons and glia. P2X4 expression in astrocytes and oligodendrocytes precursors remain controversial (indicated by a question mark). Activation of P2X4 by extracellular ATP released by neurons or glial cells exerts mostly neuromodulatory actions in physiological state. Right, in pathological states, higher extracellular ATP concentration and *de novo* expression of P2X4 as well as increased surface trafficking of intracellular pools of P2X4 lead to the increase of the number of surface P2X4 in distinct cell types in numerous central disorders and exacerbation of purinergic signaling.

Figure 2: Implication of P2X4 in spinal cord disorders

de novo P2X4 expression and/or an increased surface trafficking of intracellular pools occur in glia or neurons of the spinal cord (inset) in various chronic pain conditions and amyotrophic lateral sclerosis (ALS). The contribution of P2X4 in spinal cord (as well as in sensory ganglia for pain) is indicated. P2X4 potentiation was induced by ivermectin.

Figure 3: Implication of P2X4 in brain disorders

de novo P2X4 expression and/or an increased surface trafficking of intracellular pools occur in various brain disorders in either microglia, neurons or in both cell types. The relative contributions of neuronal and glial P2X4 in brain disorders is indicated (MS, multiple sclerosis; AD, Alzheimer's disease; PD, Parkinson disease). P2X4 potentiation was induced by ivermectin. Increased P2X4 correspond to observation from P2X4mCherryIN knockin mice.

CNS regions	P2X4 mRNA ^a	P2X4 protein	P2rx4 reporter mice [82, 83]	P2X4 mCherryIN mice [84]	Cell types specificity	References
Amygdala	++	NA	NA	NA	NA	
Basal ganglia	++	+	NA	NA	Neurons, Astrocytes	[48, 49, 71]
Cerebellum	++	++	++	+	Neurons	[48, 49, 57, 58, 59,]
Cerebral cortex	+++	+	++	++	Neurons Astrocytes ^b	52, 58] [59]
Hindbrain	+	+++	++	NA	Neurons	[58]
Hippocampus	++	++	++	++	Neurons, Microglia, Astrocytes ^b	[13, 14, 57, 58, 59]
Hypothalamus	+	+	++	NA	Neurons	[51, 58]
Midbrain	+	+	++	NA	Neurons	[58]
Olfactory region	++	+++	+++	++	Neurons, Epithelial cells	[49, 57, 58, 59]
Spinal Cord	++	++	++ 2	NA	Motoneurons, Microglia	[58, 89, 109]
Thalamus	+	+	+	NA	Neurons	[58]

Table 1. Expression of P2X4 in the CNS

Expression levels are indicated on a semi-quantitative scale where (+) indicates the presence and relative abundance of P2X4 expression. (NA) no specific data set. (a) data are from the Allen brain atlas obtained by in situ hybridization and available at http://mouse.brain-map.org/experiment/show/75551474. (b) P2X4 expression was detected by electronic microscopy in astrocytes of P2X4mCherryIN mice using anti-RFP antibodies.





Figure 2: Implication of P2X4 in spinal cord disorders

118x96mm (300 x 300 DPI)

Brain	AD,PD, epilepsy, anxiety Neuror Post-isch Migraii Alcohol	ATP Atp Microglia Microglia Microglia Eunction of P2X4		
Post-ischemic inflammation	brain microglia, vascular endothelial cells	chemotaxis of microglia, release of cytokines upregulation of osteopontin (neuroprotective)		
AD	hippocampal neurons	promote the toxic effect of beta-amyloid fragments increased P2X4: induced alteration of synaptic plasticity and memory deficits		
Migraine	trigeminal nucleus caudalis microglia	microglial release of BDNF P2X4 potentiation: MN degeneration (high ATP), Increase p-ERK and CGRP release		
Epilepsy	hippocampal neurons, stratum radiatum microglia	release of BDNF, neuronal hyperexcitability, cell death		
MS	microglia, macrophages	release of microglial BDNF; Increase of myelin phagocytosis P2X4 potentiation: promotes remyelination		
Alcohol-related disorders	dopaminergic neurons, microglia	ethanol induced P2X4 inhibition, reinforce alcohol intake/preference		
PD	dopaminergic neurons	regulate DA homeostasis		
Anxiety	forebrain neurons	increased P2X4: induced anxiolytic effects		

Figure 3: Implication of P2X4 in brain disorders

114x123mm (300 x 300 DPI)