



HAL
open science

The Problem of Aspartimide Formation During Protein Chemical Synthesis Using SEA-Mediated Ligation

Jennifer Bouchenna, Magalie Sénéchal, Hervé Drobecq, Jérôme Vicogne, Oleg Melnyk

► **To cite this version:**

Jennifer Bouchenna, Magalie Sénéchal, Hervé Drobecq, Jérôme Vicogne, Oleg Melnyk. The Problem of Aspartimide Formation During Protein Chemical Synthesis Using SEA-Mediated Ligation. Peptide and Protein Engineering, pp.13-28, 2020, 978-1-0716-0720-6. 10.1007/978-1-0716-0720-6_2. hal-02995426

HAL Id: hal-02995426

<https://hal.science/hal-02995426>

Submitted on 13 Nov 2020

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

The problem of aspartimide formation during protein chemical synthesis using SEA-mediated ligation

Jennifer Bouchenna, Magalie Sénéchal, Hervé Drobecq, Jérôme Vicogne & Oleg Melnyk

CNRS, Pasteur Institute of Lille, University of Lille, UMR CNRS 8204, Lille, France. Correspondence should be addressed to O.M. (oleg.melnyk@ibl.cnrs.fr).

Summary

Aspartimide formation often complicates the solid phase synthesis of peptides. Much less discussed is the potential occurrence of this side-reaction during the coupling of peptide segments using chemoselective peptide bond forming reactions such as the native chemical ligation and extended methods. Here we describe how to manage this problem using *bis*(2-sulfenylethyl)amido (SEA)-mediated ligation.

Key words: Aspartimide, *bis*(2-sulfenylethyl)amido (SEA)-mediated ligation, SPPS

1. Introduction

Aspartimide formation is a well-known side-reaction, which often complicates the solid phase peptide synthesis (SPPS⁽¹⁾). This side-reaction is due to the carboxylic acid group of the aspartic acid side-chain, which can react with the α -nitrogen of the next residue to form a succinimide moiety called aspartimide. The aspartimide can eventually be opened by nucleophiles, typically water or hydroxide ion, to produce a peptide having a natural backbone or an isopeptidic structure depending on which aspartimide carbonyl is attacked by the nucleophile. Note that the opening of an aspartimide yields usually the isopeptide are the major product. Note also that aspartimides are prone to epimerization. Therefore, aspartimide formation can ultimately result in the formation of several byproducts. Several tools are now available for minimizing this side reaction during SPPS such as optimized Fmoc deprotection cocktails, side-chain protecting groups for aspartic acid or backbone protecting groups for the residue following Asp.⁽²⁾

Aspartimide byproduct formation can also spontaneously occur in aqueous solution at neutral pH and as such is one of the mechanisms that lead to the degradation of proteins in vivo with aging.⁽³⁾ The

occurrence of this side-reaction in proteins,**(3)** peptide therapeutics**(4)** and model peptides**(5-7)** featuring Asn or Asp residues has been the subject of numerous studies. The occurrence of Asp/Asn-Gly dipeptides in proteins, which are particularly prone to aspartimide formation, is on average 2.7 units per protein (1.2 for Asp-Gly, 1.5 for Asn-Gly, data extracted from UniRef50**(8)**). Therefore, such aspartimide-prone sites are frequent and can considerably complicate the synthesis of most proteins (see **Note 1**), regardless of the type of ligation used.**(9)**

The chemical synthesis of SUMO-2 and SUMO-3 proteins has been described in a few works using either ketoacid-hydroxylamine ligation**(10)** or optimized SPPS protocols.**(11)** In each case, the SPPS of the peptide segments was complicated by aspartimide byproduct formation due to the presence of two Asp-Gly units in SUMO-2 and 3 domains (Figure 1). Aspartimide byproduct formation during SPPS could be suppressed by using Fmoc-Asp(OtBu)-(Dmb)Gly-OH dipeptide unit for introducing the Asp and Gly residues.

During our investigations on the chemical synthesis of SUMO-2 and 3 proteins by SEA-mediated ligation,**(12)** we also found that the use of Fmoc-Asp(OtBu)-(Dmb)Gly-OH dipeptide unit during the SPPS of the peptide segments was mandatory for suppressing aspartimide byproduct formation. Performing the ligation of the peptide segments under classical experimental conditions (pH 5.5, 4-mercaptophenylacetic acid (MPAA),**(13)** 37 °C) resulted in the concomitant formation of the target SUMO protein and of a -18 uma contaminant (~ 20% by MALDI-TOF), which could not be separated by HPLC. Interestingly, lowering the temperature of the ligation mixture to 25 °C enabled greatly minimizing the side-reaction and the isolation of the target proteins in good yield and purity.**(14,15)**

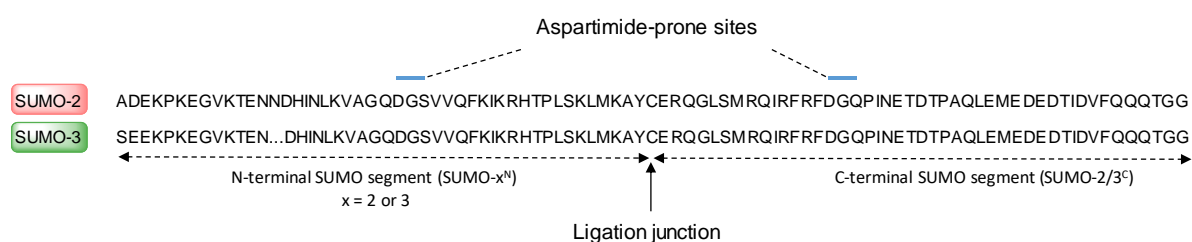


Figure 1. Sequences of SUMO-2 and 3 proteins showing the presence of an Asp-Gly dipeptide unit on both sides of the ligation site.

This protocol details the chemical synthesis of SUMO-2 and 3 proteins by SEA-mediated ligation at 25 °C in the presence of MPAA. The presence in SUMO proteins of a Cys residue in central position enabled

their assembly by ligating two peptide segments of about 45 amino acids (Figure 1). The N-terminal peptide segments, i.e., SUMO- x^N ($x = 2$ or 3) were produced by 9-fluorenylmethyloxycarbonyl (Fmoc) SPPS using bis(2-sulfanylethyl)amino (SEA) ChemMatrix[®] resin (Fig. 2A).^(16,17) The C-terminal peptide segment SUMO-2/3^C is the same for both proteins. It was produced by Fmoc SPPS starting from 4-hydroxymethylphenoxyacetyl (HMPA) ChemMatrix[®] resin (Fig. 2B).

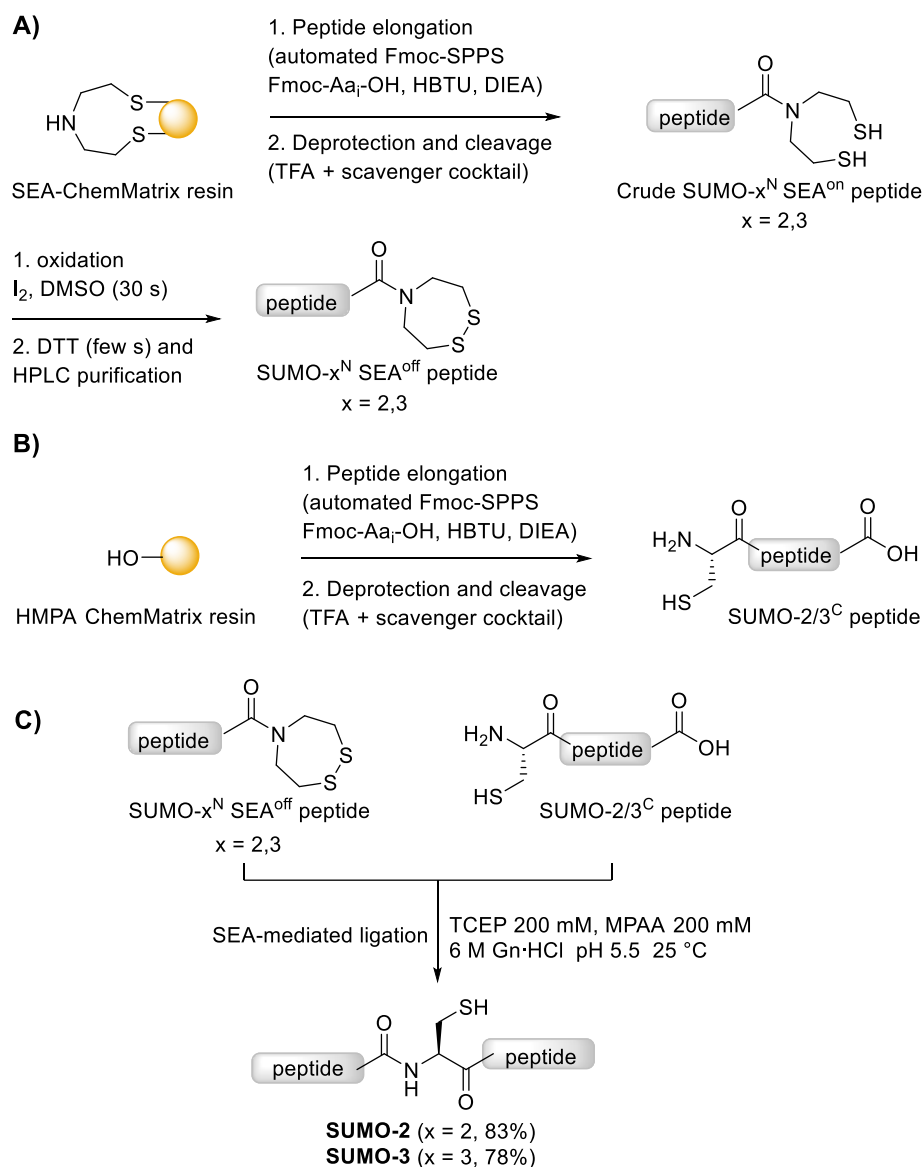


Figure 2. Synthesis of SUMO-2 and 3 proteins by SEA-mediated ligation. A) SPPS of the N-terminal segments using SEA-ChemMatrix[®] resin. B) SPPS of the C-terminal segment using 4-hydroxymethylphenoxyacetyl (HMPA) ChemMatrix[®] resin. C) Assembly of SUMO-2 and 3 proteins by SEA-mediated ligation at 25 °C.

2. Materials & Instruments

All organic solvents and chemicals used in this protocol should be handled inside a chemical fume hood with appropriate personal protective equipment (lab coat, gloves and protective glasses). TFA is strongly corrosive and toxic.

2.1 General

1. Synthesis of *bis*(2-sulfanylethyl)aminotriyl ChemMatrix (SEA ChemMatrix) resin was carried out as described elsewhere.**(16,17)**
2. HMPA ChemMatrix resin (IRIS Biotech, Lot 08K11-20-03-043).
3. Dichloromethane RPE ACS stabilised with amylene (DCM; Carlo Erba, Cat. No. 463314).
4. *N,N*-Dimethylformamide for peptide synthesis (DMF; Carlo Erba, Cat. No. P0343521).
5. 1-Methyl-2-pyrrolidinone (NMP; Carlo Erba, Cat. No. P0873521)
6. Resins were conditioned in DCM (3 x 2 min, 3 mL) and then in DMF (3 x 2 min, 3 mL) in a manual SPPS glass reactor prior to their use.
7. *N* α -Fmoc protected amino acids were obtained from Iris Biotech GmbH: Fmoc-Ala-OH, Fmoc-Arg(Pbf)-OH, Fmoc-Asn(Trt)-OH, Fmoc-Asp(OtBu)-OH, Fmoc-Asp(OtBu)[Dmb-Gly]-OH, Fmoc-Gln(Trt)-OH, Fmoc-Glu(OtBu)-OH, Fmoc-Gly-OH, Fmoc-His(Trt)-OH, Fmoc-Ile-OH, Fmoc-Leu-OH, Fmoc-Lys(Boc)-OH, Fmoc-Met-OH, Fmoc-Phe-OH, Fmoc-Pro-OH, Fmoc-Ser(*t*Bu)-OH, Fmoc-Thr(*t*Bu)-OH, Fmoc-Tyr(*t*Bu)-OH, Fmoc-Val-OH, Fmoc-Cys(*S*tBu)-OH or Fmoc-Cys(Trt)-OH.
8. *N*-[(dimethylamino)-1*H*-1,2,3-triazolo-[4,5-*b*]pyridin-1-ylmethylene]-*N*-methylmethanaminium hexafluorophosphate *N*-oxide (HATU; Novabiochem, Cat. No. 01-62-0041).
9. *N*-[(1*H*-benzotriazol-1-yl)(dimethylamino)methylene]-*N*-methylmethanaminium hexafluorophosphate *N*-oxide (HBTU; IRIS Biotech, Cat. No. RL-1030).
10. *N,N*-Diisopropylethylamine (DIEA; SDS, Cat. No. 0403516).
11. Acetaldehyde (Aldrich, Cat. No. 110078), 2 vol-% in DMF.
12. *p*-Chloranil (Acros, Cat. No. 213561000), 2 wt-% in DMF.
13. 2,4,6-Trinitrobenzenesulfonic acid (TNBS, TIC Europe N.V, Cat. No. 2508-19-2)
14. *N,N'*-diisopropylcarbodiimide (DIC, Acros, Cat. No. 446181000)
15. 4-(dimethylamino)pyridine (DMAP, Aldrich chemie, Cat. No. 10 770-0)
16. Acetic anhydride (Ac₂O) was purchased from Biosolve and Fisher-Chemical.

17. UV spectrophotometer
18. Precision quartz cell (length 10 mm, quartz suprasil, Hellma, Cat. No. 117104-05)
19. Diethyl ether RPE stabilised with 2,6-bis(1,1-dimethylethyl)-4-methylphenol (Carlo Erba, Cat. No. 447523)
20. Piperidine for peptide synthesis (Biosolv, Cat. No. 16183302).
21. Thioanisole (Fluka, Cat. No. 88470).
22. Thiophenol (Alfa Aesar, Cat. No. 10201873)
23. Trifluoroacetic acid (TFA; Biosolv, Cat. No. 20233301).
24. Triisopropylsilane (TIS; Aldrich, Cat. No. 23378-1).
25. 1,2-Ethanedithiol, 99% (EDT, Aldrich, Cat. No. E360-0).
26. *n*-Heptane (Biosolv, Cat. No. 0805202).
27. Water was purified with a Milli-Q Ultra Pure Water Purification System.
28. Dithiothreitol (DTT; Acros Organics, Cat. No. 16568-0050).
29. Iodine (Acros Organics, Cat. No. 38705100).
30. 0.1% TFA in water, Optima LC/MS (Fisher Chemical, Cat. No. LS119-212).
31. 0.1% TFA in acetonitrile, Optima LC/MS (Fisher Chemical, Cat. No. LS119-212).
32. Sodium dihydrogen phosphate dihydrate ($\text{NaH}_2\text{PO}_4 \cdot 2\text{H}_2\text{O}$, Prolabo, Cat. No. 28015294).
33. Disodium hydrogen phosphate dihydrate ($\text{Na}_2\text{HPO}_4 \cdot 2\text{H}_2\text{O}$, Prolabo, Cat. No. 28015294).
34. Guanidine hydrochloride ($\text{Gn} \cdot \text{HCl}$; Aldrich, Cat. No. G4505).
35. Glacial acetic acid (AcOH) (Carbo Erba, Cat. No. 401422).
36. Nitrogen gas ($\text{O}_2 < 3$ ppm, Air Liquide).
37. Argon (Alphagaz Airliquide).
38. Hydrochloric acid (HCl; Sigma Aldrich, Cat. No. 32-0331-500 mL).
39. Sodium hydroxide (NaOH; VWR, Cat. No. 28240292).
40. *Tris*(2-carboxyethyl)phosphine hydrochloride (TCEP-HCl; Aldrich, Cat. No. C4706-10G).
41. 4-mercaptophenylacetic acid (MPAA; Alfa Aesar, Cat. No. H27658).
42. Conical tubes (15 mL, 50 mL).
43. Thermomixer comfort (Eppendorf) equipped with a block heater (25 °C)
44. A set of adjustable pipettes (0.5–10, 2–20, 10–50, 10–100, 100–1,000 μL , Eppendorf)
45. Pipette tips, 0.5–10, 2–20, 20–200, 100–1,000 μL (VWR).
46. pH meter
47. Vortex shaker.
48. Microfuge tubes (1.5 mL safe-lock tubes; Eppendorf).

2.2 HPLC analysis & purification

1. TFA 10% (vol/vol) in water. Add carefully 10 mL of TFA to 90 mL of deionized water.
2. Eluent A for semi-preparative HPLC. TFA 0.1% (vol/vol) in deionized water. Add carefully 10 mL of 10% aqueous TFA (vol/vol) to 990 mL of deionized water.
3. Eluent B for semi-preparative HPLC. HPLC 80% (vol/vol) containing 0.1% (vol/vol) TFA. Add 10 mL of 10% aqueous TFA (vol/vol) to 990 mL of ACN 80% (vol/vol) in deionized water.
4. Analytical UPLC-MS system: Dionex UltiMate 3000/ LCQ Fleet Ion Trap, Dionex DA detector (215-280 nm), Corona Veo charged aerosol detector, ES+, m/z range 300–2000, capillary voltage 2.8 kV, cone voltage 10 V, tube lens 75 V, capillary voltage temperature 350 °C.
5. Analytical UPLC-MS column: Acquity UPLC peptide BEH300 C18 reverse-phase column (2.1 × 100 mm; pore size 300 Å; particle size: 1.7 μm).
6. Semi-preparative HPLC system (Waters 600 controller, UV 2487 Detector, 215 nm, TL 105 HPLC column heater).
7. Semi-preparative HPLC column: Waters XBridge BEH300 C18 reverse-phase column (10 x 250 mm; pore size 300 Å; particle size: 5 μm).

2.3 Lyophilization

1. Lyophilizer (Christ Gamma 2-20) equipped with a manifold.
2. Lyophilizer flasks (VWR, 300 mL, Cat. No. 88516; 600 mL, Cat. No. 88517).
3. Liquid nitrogen
4. Dewar

2.4 MALDI-TOF analysis

1. MALDI-TOF mass spectrometer (Autoflex Speed, Bruker).
2. 2,5-dihydrobenzoic acid (DHB, Aldrich, Cat. No. 490799).
3. A solution of DHB 20 mg/mL in 50% aqueous acetonitrile containing 0.1% TFA.

3. Solid phase peptide synthesis of the peptide segments

1. Column peptide synthesizer (Multipep Intavis AG Bioanalytical instrument equipped with a 6-column module)
2. Synthesis columns (Intavis, Cat. No. 35760)
3. Reaction columns (Intavis, Cat. No. 34274)
4. Manual peptide synthesis vessels
5. Fmoc amino acid solutions, Fmoc-AA-OH 0.54 M in DMF.
6. DIEA solution, DIEA 35% in NMP (vol/vol).

7. HBTU solution: HBTU 0.54 M in DMF.
8. Piperidine solution 20% in DMF (vol/vol), see **Note 2**.
9. Capping solution, acetic anhydride 10%/DIEA 5%/DMF 85% (vol/vol/vol), see **Note 3**.

3.1 Coupling of first amino acid to the SEA-Trt ChemMatrix® resin

1. Weigh the amino acid Fmoc-Tyr(OtBu)-OH (459 mg, 1.00 mmol, 10 equiv) in a plastic tube.
2. Weigh 377 mg (0.95 mmol, 9.5 equiv) of HATU in a plastic tube, see **Note 4**.
3. Add 5 mL of DMF to the tubes and agitate with a vortex shaker to dissolve the reagents.
4. Transfer the HATU/DMF solution to the tube containing the amino acid solution with an adjustable pipette.
5. Add 348 μ L (2.00 mmol, 20 equiv) of DIEA to the tube with an adjustable pipette and agitate with a vortex shaker for 1 min. The solution becomes yellow upon addition of the base.
6. Transfer the activated amino acid/DMF solution to the reaction column.
7. Agitate with a laboratory flask shaker for 1 h 30 min and drain the column.
8. Aspirate 5-10 mL of DMF, agitate for 2 min and then drain the column.
9. Repeat Step 8 twice.
10. Take a sample of the resin and perform the chloranil test which must be negative (colorless resin beads).
11. Acetylate remaining amino groups eventually present on the Fmoc-Tyr(OtBu)-SEA-Trt ChemMatrix resin by adding 5 mL of acetic anhydride/DIEA/DMF capping solution to the resin. Shake for 10 min and drain, see **Note 5**.
12. Repeat Step 11 twice.
13. Wash the resin with DMF (3 x 2 min, 5 mL).
14. Wash the resin with DCM (3 x 2 min, 5 mL).
15. Aspirate 5 mL of diethyl ether, shake for 2 min and then drain the column.
16. Repeat Step 15 twice.
17. Determine the loading of the Fmoc-Tyr(OtBu)-SEA Trt ChemMatrix resin by treating the resin with the 20% piperidine solution in DMF and UV quantification of the formed dibenzofulvene-piperidine adduct at 290 nm as described elsewhere. **(16)** Typical loading for Fmoc-Tyr(OtBu)-SEA-Trt ChemMatrix resin: 0.22 mmol/g.

3.2 Coupling of first amino acid to the HMPA ChemMatrix® resin

1. Weigh HMPA ChemMatrix® resin (500 mg).
2. Weigh the amino acid Fmoc-Gly-OH (297 mg, 1.00 mmol) in a plastic tube.
3. Weigh 12.2 mg (0.100 mmol) of DMAP.

4. Add 5 mL of DMF to the tube containing the amino acid to dissolve it.
5. Add 78.3 μL (0.500 mmol) of DIC to the tube containing the amino acid solution with an adjustable pipette.
6. Transfer the activated amino acid/DIC solution to the reaction column.
7. Add DMAP in one portion to the reaction column.
8. Agitate with a laboratory flask shaker for 2 h at room temperature.
9. Aspirate 5-10 mL of DMF, agitate for 2 min and drain the column.
10. Repeat Step 9 twice.
11. Wash the resin with DMF (3 x 2 min, 5 mL).
12. Wash the resin with DCM (3 x 2 min, 5 mL).
13. Add 5 mL of diethyl ether, shake for 2 min and drain the column.
14. Repeat Step 13 twice
15. Determine the loading of the Fmoc-Gly-HMPA ChemMatrix[®] resin after treating an aliquot with 20% piperidine solution in DMF and reading the absorbance of the supernatant at 290 nm. Typical loading for Fmoc-Gly-HMPA ChemMatrix[®] resin: 0.58 mmol/g.

3.3 First elongation using the automated peptide synthesizer

Elongate Fmoc-Tyr(OtBu)-SEA-Trt ChemMatrix resin and Fmoc-Gly-HMPA ChemMatrix[®] resin using the automated peptide synthesizer, see **Note 6**.

Elongated sequence using Fmoc-Tyr(OtBu)-SEA-Trt ChemMatrix resin (SUMO-2^N and SUMO-3^N): SVVQFKIKRHTPLSKLMKA

Elongated sequence using Fmoc-Gly-HMPA ChemMatrix[®] resin (SUMO-2/3^C): QPINETDTPAQLEMEDEDTIDVFQQQTG

3.4 Manual coupling of Fmoc-Asp(OtBu)[Dmb]Gly-OH

1. Weigh Fmoc-Asp(OtBu)[Dmb]Gly-OH (186 mg, 0.300 mmol, 3 equiv) in a plastic tube.
2. Weigh 108 mg of HATU (108 mg, 0.284 mmol, 2.85 equiv) in the plastic tube.
3. Add 3 mL of DMF to the tube containing HATU and agitate with a vortex shaker to dissolve the reagent.
4. Transfer the HATU/DMF solution to the tube containing the amino acid solution with an adjustable pipette.
5. Add 104 μL of DIEA (0.600 mmol, 6 equiv) to the tube with an adjustable pipette and agitate with a vortex shaker for 1 min. The solution becomes yellow upon addition of the base.
6. Transfer the activated amino acid/DMF solution to the reaction column.

7. Agitate with a laboratory flask shaker for 2 h and then drain the column.
8. Aspirate 5-10 mL of DMF, agitate for 2 min and drain the column.
9. Repeat Step 8 twice
10. Take a sample of the resin and perform a TNBS test that must be negative (colorless resin beads), see **Note 7**.
11. Optional: analysis of the elongated peptide by MALDI-TOF after cleavage and deprotection, see Steps 12-14.
12. Treat an aliquot of the peptidyl resin with 200 μ L of 20% piperidine to remove the N-terminal Fmoc group.
13. Remove the piperidine solution with 1 mL of diethyl ether using a Pasteur pipette and dry the resin under a gentle flow of argon.
14. Perform the TFA cleavage for 1 h by adding 70 μ L of the cleavage mixture to the resin aliquot: TFA/triisopropylsilane (TIS)//thioanisole/H₂O/thiophenol: 87.5/5/2.5/2.5/2.5 by vol (1 mL) for SEA ChemMatrix[®] resin, or TFA/TIS/H₂O/ethanedithiol (EDT): 90/5/2.5/2.5 by vol (1 mL) for HMPA-ChemMatrix[®] resin.
Take 1 μ L of the cleavage mixture, dilute with water (50 μ L) and analyze by MALDI-TOF to confirm the successful elongation of the peptide chain including the coupling of the dipeptide unit. Elongated sequence using Fmoc-Tyr(OtBu)-SEA-Trt ChemMatrix resin (SUMO-2^N and SUMO-3^N): DGSVVQFKIKRHTPLSKLMKAY [M+H]⁺ calc. 2663.45, found 2664.86.
Elongated sequence using Fmoc-Gly-HMPA ChemMatrix[®] resin (SUMO-2/3^C): DGQPINETDTPAQLEMEDEDTIDVFQQQTGG [M+H]⁺ calc. 3421.48, found 3422.10. See **Note 8**.
15. Continue the synthesis by acetylating unreacted amino groups eventually present on the HMPA or SEA ChemMatrix[®] peptidyl resins by adding 5 mL of acetic anhydride/DIEA/DMF capping solution to the solid support. Shake for 10 min and drain
16. Repeat Step 15 twice
17. Add 5 mL of DMF to the column shake for 2 min and drain the column.
18. Repeat Step 17 twice

3.5 Second elongation using automated peptide synthesizer

Elongate the peptidyl resins using the automated peptide synthesizer.

Elongated sequence for SEA-Trt ChemMatrix resin: ADEKPKEGVKTENNDHINLKVAGQ for SUMO-2^N, SEEKPEGVKTENDHINLKVAGQ for SUMO-3^N.

Elongated sequence for HMPA ChemMatrix[®] resin: CERQGLSMRQIRFRF for SUMO-2/3^C.

3.6 Cleavage step

1. In a graduated cylinder add 8.75 mL of TFA, 0.25 mL of deionized water, , 0.25 mL of thiophenol, 0.25 mL of thioanisole and 0.50 mL of TIS for peptides SUMO- x^N ($x = 2, 3$).
2. In a graduated cylinder add 9 mL of TFA, 0.25 mL of deionized water, 0.25 mL of EDT and 0.50 mL of TIS for peptide SUMO-2/3^C.
3. Transfer each peptidyl resin in a different manual peptide synthesis vessel.
4. Add the TFA solution to the manual peptide synthesis vessel and agitate at room temperature for 2 h
5. In the meantime, prepare three 300 mL Erlenmeyer flasks, each containing a mixture of 150 mL of diethyl ether and 150 mL of *n*-heptane. Add magnetic bars and place the Erlenmeyer flasks in an ice bath.
6. Precipitate the peptides by adding the peptide solutions dropwise to the ice-cold diethyl ether/*n*-heptane mixture with magnetic stirring.
7. Add 5 mL of TFA to each peptide reactor, shake the bead suspensions for 2 min and then add this solution dropwise to the Erlenmeyer flask.
8. Repeat Step 7, see **Note 9**.
9. Transfer the suspensions to centrifuge tubes and centrifuge at $1,900 \times g$ for 10 min, then pour carefully the supernatant into an Erlenmeyer flask.
10. Add 40 mL of cold diethyl ether 50%/n-heptane 50% (vol/vol) to the peptide, triturate the peptide pellet and centrifuge again at $1,900 \times g$ for 10 min, then pour carefully the supernatant into the Erlenmeyer flask.
11. Carefully flush the tubes with nitrogen to evaporate the residual solvent.
12. Dissolve the crude peptides in TFA 0.1% (vol/vol) in deionized water and place the peptide solutions immediately at -20°C overnight, see **Note 10**.
13. Lyophilize for 2 days.
14. Analyze the crude peptide by UPLC-MS and MALDI-TOF mass spectrometry

3.7 Oxidation SEA^{on} peptides. Purification of SUMO- x^N SEA^{off} peptides ($x = 2,3$)

1. Prepare the semi-preparative HPLC system, which should be ready before starting the oxidation of the peptides.
2. Rapidly weigh 100-150 mg of iodine in a 5 mL volumetric flask. Immediately add the DMSO up to 5 mL. Calculate the concentration of iodine, see **Note 11**.
3. Prepare a solution of DTT 10 mg/mL in deionised water.
4. Dissolve the crude peptide in phosphate buffer/Gn-HCl 6 M at room temperature (peptide concentration ~ 0.25 mM), see **Note 12**.

5. Add 2 equivalents of iodine in DMSO to the peptide solution. The solution becomes yellow indicating that an excess of iodine has been added to the peptide solution.
6. Vortex the solution for 30 s, see **Note 13**.
7. Then immediately add a few drops of the DTT solution with a Pasteur pipette until the yellow color disappears, indicating that the excess of iodine has been decomposed.

3.8 HPLC purification of SUMO-2/3^C and SUMO-2/3^N peptides.

1. Purify immediately by semi preparative HPLC.
2. Gradient used for the HPLC purification of peptide **SUMO-2^N**: eluent A water containing 0.1% of TFA, eluent B CH₃CN/water : 4/1 by vol containing 0.1% of TFA, gradient: 0-25% B in 5 min, then 25-35% B in 40 min, flow rate 6 mL/min, 50 °C, UV detection at 215 nm.
3. Yield for peptide **SUMO-2^N**: 34.4 mg of crude product furnished (11.2 mg) of **SUMO-2^N**, Calculated for M (average mass) 5281.17, observed 5281.13 by LC-MS after deconvolution.
4. Gradient used for the HPLC purification of peptide **SUMO-3^N**: eluent A water containing 0.1% of TFA, eluent B CH₃CN/water : 4/1 by vol containing 0.1% of TFA, gradient: 0-25% B in 5 min, then 25-35% B in 40 min, flow rate 6 mL/min, 50 °C, UV detection at 215 nm.
5. Yield for peptide **SUMO-3^N**: 30.8 mg of crude product furnished (11.2 mg) of **SUMO-3^N**, Calculated for M (average mass) 5197.09, observed 5197.17 by LC-MS after deconvolution.
6. Gradient used for the HPLC purification of peptide **SUMO-2/3^C**: eluent A water containing 0.1% of TFA, eluent B CH₃CN/water : 4/1 by vol containing 0.1% of TFA, gradient: 0-20% B in 5 min, then 20-40% B in 40 min, flow rate 6 mL/min, 50°C, UV detection at 215 nm
7. Yield for peptide **SUMO-2/3^C**: 31.3 mg of crude product furnished (10.6 mg) of **SUMO-2/3^C**, Calculated for M (average mass) 5331.82, observed 5331.95 by LC-MS after deconvolution.
8. Analyze the collected fractions by analytical UPLC-MS and pool the pure fractions in a 50 mL plastic tube.
9. Place the tubes in liquid nitrogen to freeze the peptide solutions and lyophilize for two or three days to obtain the purified **SUMO-2^N**, **SUMO-3^N**, **SUMO-2/3^C** peptides segments, see **Note 10**.

4. Synthesis of SUMO-2 and 3

1. Weigh 5.38 mg (1 equiv, 0.825 μmol, 4 mM) of SUMO-x^N SEA^{off} peptide (x = 2 or 3) in a 1.5-mL microfuge tube.
2. Weigh 4.94 mg (1 equiv, 0.825 μmol, 4 mM) of cysteinyl peptide SUMO-2/3^C in a 1.5-mL microfuge tube.
3. Weigh 13.42 mg (79.77 μmol, 200 mM final concentration) of MPAA in a 1.5-mL microfuge tube.
4. Weigh 31.6 mg (110 μmol, 200 mM final concentration) of TCEP in a 1.5-mL microfuge tube.

5. Prepare 2 mL of 0.1 M sodium phosphate buffer (pH 7.2) / 6 M Gn·HCl.
6. Transfer 1 mL of AcOH glacial, 1 mL of HCl 1 N and 1 mL of NaOH 6 N in three different 1.5-mL microfuge tubes.
7. Cap all the tubes and transfer them to the glove box under nitrogen atmosphere.
8. Add 0.551 mL of the sodium phosphate buffer from Step 5 to the microfuge tube containing TCEP (see Step 4). Vortex until dissolution of the TCEP and transfer 0.399 mL of this solution to the microfuge tube containing MPAA (see Step 3). Add NaOH 6 N (30 μ L) with the micropipette and vortex until dissolution of the MPAA. The final reagent concentration is 181 mM for MPAA and 181 mM for TCEP.
9. Measure the pH of the solution prepared in Step 8 and adjust to pH 5.5 by adding NaOH 6 N or HCl 1 N with the micropipette (2-20 μ L).
10. Add 0.205 mL of the solution prepared in Step 8 to the microfuge tube containing the SUMO-x^N SEA^{off} peptide (see Step 1). Vortex to dissolve the peptide and then add this solution to the microfuge tube containing the cysteinyl peptide SUMO-2/3^C (see Step 2). Vortex to dissolve the peptide.
11. Place the reaction tube from Step 10 in a thermomixer kept at 25 °C with agitation.
12. To monitor the ligation reaction (just after preparing the reaction mixture at Step 10 and every 6-7 hour until ~33 h of reaction, transfer 1.5 μ L of the reaction mixture to a microfuge tube and quench the reaction by acidifying the sample with 100 μ L of 10% aqueous AcOH (vol/vol). Vortex and remove the sample from the glove box.
13. Remove MPAA present in the reaction mixture by extracting the aqueous phase with 1 mL of diethyl ether. Remove the diethyl ether phase with a Pasteur pipette.
14. Repeat Step 13 twice.
15. Inject 10 μ L of the sample isolated in Step 14 into the analytical UPLC-MS system to confirm that the starting peptide segments have been consumed to produce the ligation product SUMO-2 or 3.
16. After 33 h, remove the reaction tube from the glove box.
17. Transfer the contents of the reaction tube into a 5-mL microfuge tube.
18. Add 0.500 mL of phosphate buffer / 6 M Gn·HCl to the reaction mixture and 70.5 μ L of glacial acetic acid to a final concentration of 5-10 % (vol/vol) with a micropipette to acidify the solution down to a pH of 2-3 (pH paper).
19. Extract the MPAA from the aqueous phase with 2 mL of diethyl ether as done in Step 13. Remove the diethyl ether phase with a Pasteur pipette.
20. Repeat Step 19 twice.
21. Remove the dissolved diethyl ether by flushing the tube with nitrogen (~2 min)

22. Purify the target SUMO-2 or 3 proteins using a Waters XBridge BEH300 C18 reverse-phase column (10 × 250 mm; pore size 300 Å; particle size : 5 μm) and a semi-preparative HPLC system; Gradient : 0 % eluent B in eluent A to 20 % (vol/vol) eluent B in eluent A over 5 min, and then 20 % (vol/vol) eluent B in eluent A to 40 % (vol/vol) eluent B in eluent A over 60 min (UV detection at 215 nm). Analyze the fractions using the MALDI-TOF spectrometer. Mix 1.2 μL of the DHB solution with 1 μL of the sample on the MALDI plate and let dry at room temperature in air.
23. Pool the pure fractions in a 50 mL plastic tube, freeze the solution with liquid nitrogen and lyophilize it for two or three days. The reaction yields 4.20 mg (78% yield) of SUMO-2 and 4.46 mg of SUMO-3 (83% yield).
24. Analyze the purified products by UPLC-MS. ACQUITY UPLC peptide BEH C18 300 Å 1.7 μm 2.1 mm × 150 mm, 50 °C. Flow 0.400 mL/min, eluent A 0.1% trifluoroacetic acid in water, eluent B 0.1% trifluoroacetic acid in 100% acetonitrile. Gradient from 0% eluent B to 70% eluent B in 20 min.
25. SUMO-2: Calculated for M (average mass) 10477.24, observed 10476.71 after deconvolution
26. SUMO-3: Calculated for M (average mass) 10393.66, observed 10393.84 after deconvolution.

Notes

Note 1. Aspartimide byproduct formation can occur during NCL.**(18)** For example, the assembly of Ras protein (166 amino acid residues) by NCL yielded a product having 18±2 uma less than expected.**(19)** The mass loss occurred during the last ligation and could be due to a loss of water or ammonia. The site of dehydration or deamination was not identified. For another example, see ref **(20)**.

Note 2. The solution can be stored or used for up to 2 weeks at room temperature.

Note 3. This solution can be used up to 24 h after preparation. For the automated peptide elongation step, this solution is renewed every day.

Note 4. HATU is an uronium-based coupling reagent with reacts efficiently with amines and thus can cause undesired peptide termination if present in excess. To avoid the presence of unreacted HATU during the coupling step, the amino acid is used in slight molar excess and the amino acid is activated prior to the coupling step.

Note 5. The capping step is mandatory even if the chloranil test is negative, because the sensitivity of the test for the secondary amino group of the SEA resin is lower than for α-amino groups.

Note 6. The synthesis of the ~50 amino acids SEA^{off} peptide segments **SUMO-x^N** is performed using the SEA-Trt-ChemMatrix® resin. The synthesis program must consider the large swelling volume of the starting SEA-Trt resin and the significant mass increase of the peptidyl resin that occurs during the peptide elongation step.

Note 7. We recommend to confirm the coupling of Fmoc-Asp(OtBu)[Dmb]Gly-OH by cleaving an aliquot in TFA and analyzing the cleaved peptide by MALDI-TOF.

Note 8. MALDI-TOF analysis of the cleaved peptides might show the expected peaks and also products of higher mass (+ 150 uma) due to the incomplete removal of the Dmb group.

Note 9. We recommend that the volume of the diethyl ether/*n*-heptane solution be at least 20 times that of the peptide solution in TFA.

Note 10. The peptide solution can be frozen quickly using liquid nitrogen. The tube must be checked before starting the lyophilization because it can crack during this step.

Note 11. Iodine is available as granules. Therefore, weighting a given amount of iodine precisely is difficult to achieve.

Note 12. **SUMO-x^N SEA^{off} peptides (x = 2,3)** have a limited solubility and the use of Gn·HCl, which is a classical additive for improving the solubility of the peptide segments, was mandatory in this case. Iodine oxidation can be performed in the absence of Gn·HCl and at mM concentrations when solubility is not an issue.

Note 13. The duration of the iodine treatment should not exceed 30 s, otherwise iodine can react with some amino acid residues such as tyrosines. For the same reason, the amount of iodine which is added to the peptide solution must not exceed two equivalents. This oxidation procedure is not compatible with the presence of free cysteine thiols on the peptide chain.

Acknowledgements

This study was supported by the National Center for Scientific Research (CNRS), the University of Lille and the Pasteur Institute of Lille.

References

- (1) Merrifield RB Solid phase peptide synthesis. I. The synthesis of a tetrapeptide. *J Am Chem Soc* 1963;85:2149-54.
- (2) Behrendt R, White P, Offer J Advances in Fmoc solid-phase peptide synthesis. *J Pept Sci* 2016;22:4-27.
- (3) Voorter CE, de Haard-Hoekman WA, van den Oetelaar PJ, Bloemendal H, de Jong WW Spontaneous peptide bond cleavage in aging alpha-crystallin through a succinimide intermediate. *J Biol Chem* 1988;263:19020-3.
- (4) Joshi AB, Sawai M, Kearney WR, Kirsch LE Studies on the mechanism of aspartic acid cleavage and glutamine deamidation in the acidic degradation of glucagon. *J Pharm Sci* 2005;94:1912-27.
- (5) Geiger T, Clarke S Deamidation, isomerization, and racemization at asparaginyl and aspartyl residues in peptides. Succinimide-linked reactions that contribute to protein degradation. *J Biol Chem* 1987;262:785-94.
- (6) Stephenson RC, Clarke S Succinimide formation from aspartyl and asparaginyl peptides as a model for the spontaneous degradation of proteins. *J Biol Chem* 1989;264:6164-70.
- (7) Tyler-Cross R, Schirch V Effects of amino acid sequence, buffers, and ionic strength on the rate and mechanism of deamidation of asparagine residues in small peptides. *J Biol Chem* 1991;266:22549-56.
- (8) Suzek BE, Huang H, McGarvey P, Mazumder R, Wu CH UniRef: comprehensive and non-redundant UniProt reference clusters. *Bioinformatics* 2007;23:1282-8.
- (9) Agouridas V, El Mahdi O, Diemer V, Cargoet M, Monbaliu J-CM, Melnyk O Native chemical ligation and extended methods. Mechanisms, catalysis, scope and limitations. *Chem Rev* 2019;12:7328-443.
- (10) Wucherpennig TG, Pattabiraman VR, Limberg FRP, Ruiz-Rodríguez J, Bode JW Traceless preparation of C-terminal α -ketoacids for chemical protein synthesis by α -ketoacid-hydroxylamine ligation: Synthesis of SUMO2/3. *Angew Chem Int Ed* 2014;53:12248-52.
- (11) Mulder MPC, Merckx R, Witting KF, Hameed DS, El Atmioui D, Lelieveld L, Liebelt F, Neefjes J, Berlin I, Vertegaal ACO et al. Total chemical synthesis of SUMO and SUMO-based probes for profiling the activity of SUMO-specific proteases. *Angew Chem Int Ed* 2018;57:8958-62.
- (12) Ollivier N, Dheur J, Mhidia R, Blanpain A, Melnyk O *Bis*(2-sulfanylethyl)amino native peptide ligation. *Org Lett* 2010;12:5238-41.
- (13) Johnson EC, Kent SBH Insights into the mechanism and catalysis of the native chemical ligation reaction. *J Am Chem Soc* 2006;128:6640-6.
- (14) Bouchenna J, Sénéchal M, Drobecq H, Vicogne J, Melnyk O Total Chemical Synthesis of All SUMO-2/3 Dimer Combinations. *Bioconjugate Chem* 2019;30:2967-73.
- (15) Bouchenna J, Sénéchal M, Drobecq D, Stankovic-Valentin N, Vicogne J, Melnyk O The Role of the Conserved SUMO-2/3 Cysteine Residue on Domain Structure Investigated Using Protein Chemical Synthesis. *Bioconjugate Chem* 2019;30:2684-96.
- (16) Boll E, Drobecq H, Ollivier N, Blanpain A, Raibaut L, Desmet R, Vicogne J, Melnyk O One-pot chemical synthesis of small ubiquitin-like modifier (SUMO) protein-peptide conjugates using *bis*(2-sulfanylethyl)amido peptide latent thioester surrogates *Nat Protoc* 2015;10:269-92.
- (17) Boll E, Drobecq H, Ollivier N, Raibaut L, Desmet R, Vicogne J, Melnyk O A novel PEG-based solid support enables the synthesis of >50 amino-acid peptide thioesters and the total synthesis of a functional SUMO-1 peptide conjugate. *Chem Sci* 2014;5:2017-22.
- (18) Dawson PE, Muir TW, Clark-Lewis I, Kent SBH Synthesis of proteins by native chemical ligation. *Science* 1994;266:776-79.
- (19) Becker CF, Hunter CL, Seidel R, Kent SBH, Goody RS, Engelhard M Total chemical synthesis of a functional interacting protein pair: the protooncogene H-Ras and the Ras-binding domain of its effector c-Raf1. *Proc Natl Acad Sci U S A* 2003;100:5075-80.
- (20) Cowper B, Shariff L, Chen W, Gibson SM, Di W-L, Macmillan D Expanding the scope of N \rightarrow S acyl transfer in native peptide sequences. *Org Biomol Chem* 2015;13:7469-76.

