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Stereospecific Synthesis of Glycoside Mimics Through Migita–Kosugi–Stille Cross-Coupling Reactions of Chemically and Configurationally Stable 1-*C*-Tributylstannyl Iminosugars

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Abstract. An innovative and effective strategy for the synthesis of imino-*C*-glycosides is described. The methodology is based on the unprecedented reaction of 1-*C*-stannylated iminosugars with various electrophiles under the conditions of Migita–Kosugi–Stille cross-couplings, which gives 1-*C*-substituted iminosugar derivatives in a stereoretentive process. The required iminoglycosyl stannanes are obtained by way of the highly stereoselective addition of tributylstannyl lithium to (*S_R*)- or (*S_S*)-*N*-*tert*-butanesulfinyl glycosylamines, followed by an activation cyclization sequence.

Most interestingly, the methodology is tunable: the configuration of the tin adduct is controlled exclusively by the *tert*-butanesulfinyl auxiliary, thus giving access after ring formation to 'α'-configured or 'β'-configured iminoglycosyl stannanes. With the subsequent stereoretentive C–C bond-forming process, the methodology allows the synthesis of pseudo anomers of imino-*C*-glycosyl compounds in a controlled fashion.

Keywords: 1-Metalated Iminosugars; Stannanes; C–C coupling; Stereospecific Reactions; Catalysis

Introduction

Glycomimetics are carbohydrate analogs designed to interfere with biochemical pathways wherein carbohydrates play key roles and are associated with pathological disorders.^[1] As potential drug candidates, analogues of glycosides and glycoconjugates are gaining increased interest because of their ability to modulate the activity of carbohydrate-processing enzymes, or enhance their affinity for carbohydrate-recognizing proteins. One class of glycomimetics that is attracting currently a surge of interest is that of iminosugars (e.g., **1** and **2**, Fig. 1). These compounds exhibit remarkable activities as glycosidases and glycosyltransferases inhibitors, which bestow them high therapeutic value.^[2] Thus, iminosugar derivatives

have emerged as new drugs for example to control glucose uptake in non-insulin-dependent diabetes patients (Miglitol®, *N*-[2-hydroxy-ethyl]-1-deoxynojirimycin **1a**),^[3] to salvage mutant enzymes in lysosomal diseases such as Fabry disease (Galafold®, 1-deoxygalactonojirimycin,^[4] **1b**) or to treat type I Gaucher and Niemann–Pick diseases by inhibition of GlcCer synthase (Zavesca®, *N*-butyl-1-deoxynojirimycin **1c** (Fig. 1)).^[5–6]

However, these saccharide surrogates lack the structural and stereochemical substitution patterns that are commonly embedded within the aglycone of glycosides and, as a result, lack selectivity in their biological activities.

In that respect, 1-*C*-substituted iminosugars (**2**), which carry a C-linked substituent at the pseudo anomeric position in a well-defined configuration, i.e.

imino-*C*-glycosides, constitute highly relevant mimetics of glycosides or glycoconjugates, and enhance the biological and therapeutic potential of iminosugars.^{[2b],[2e],[7]} For example, certain imino-*C*-nucleosides have been identified as lead anticancer, antiviral and antibacterial compounds but also for the treatment of metabolic and genetic diseases.^[8] Mundesine® (Immucillin-H) **2a**, for example was approved in Japan for the treatment of relapsed/refractory peripheral T-cell lymphoma,^[9] and BCX4430 (Immucillin-A) **2b** has been demonstrated to protect against both Ebola and Marburg viruses in rodents and monkeys, even when administered up to 48 h after infection (Fig. 1).^[10]

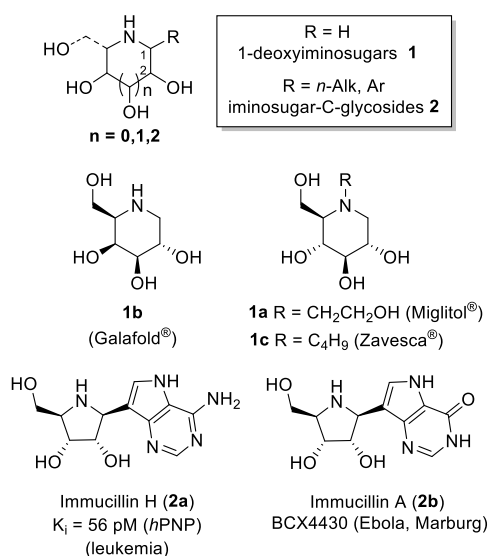


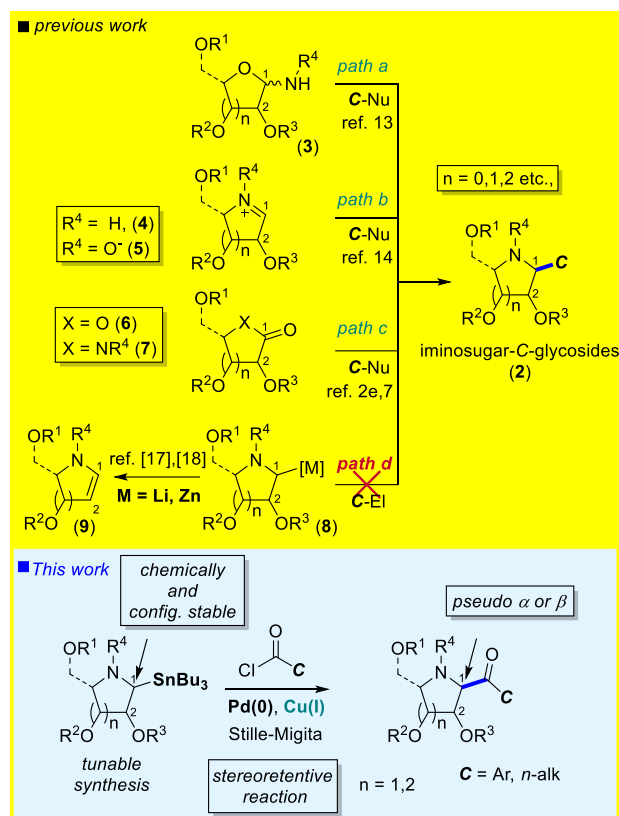
Figure 1. Chemical structures of iminosugar derivatives and imino-*C*-glycosides.

The synthesis of imino-*C*-glycosides represents a significant challenge and various elegant stereoselective methods have been developed for the *de novo* preparation of 1-*C*-substituted iminosugars,^[11] or more frequently, from sugar substrates.^[12] Most of the synthetic methods from sugars are based on the addition of carbon nucleophiles to glycosylamines (**3**) (*path a*),^[13] cyclic imines (**4**), or cyclic nitrones (**5**) (*path b*),^[14] lactones **6**, and lactams **7** (*path c*) (Scheme 1).^{[2b],[2e],[7],[15]} These tactics often require long reaction sequences and give low overall yields. In addition, the stereochemistry is governed by the inherent chirality of the saccharide core, thus giving in most cases moderate diastereoselectivities.

As a result, there are only two methodologies reported so far whereby it is possible to generate either (1*S*)- or (1*R*)-imino-*C*-glycosides (i.e., mimics of α - and β -glycosides) with high specificity in a tunable, and predictable manner.^{[11b],[12g],[16]} Furthermore, there is no report describing the more straightforward disconnection that would consist in functionalizing 1-deoxy iminosugars (**1**) at the C-1 position through a direct C(sp³)-H bond activation or by a metal-catalyzed reaction using a stable 1-metallated

iminosugar derivative and a carbon electrophile (*path d*).

Indeed, prior studies on the direct functionalization of saturated cyclic amines and on the preparation of 1-metallated iminoalditol derivatives **8** (e.g., $M = Li, Zn$) have highlighted the critical problems associated with these routes (e.g. formation of iminoglycals of type **9** through elimination of the alkoxy group at C-2 (Scheme 1), poor functional group tolerance, and difficulties in controlling the α -stereoselectivity etc.).^{[17],[18]}



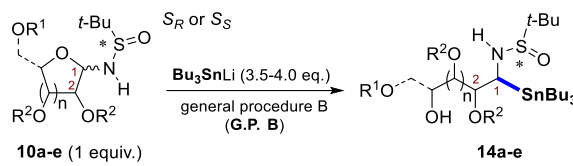
Scheme 1. Synthetic routes to iminosugar *C*-glycosides.

To be successful, the envisioned strategy would largely depend on the choice of the metal to be installed at the C-1 position. This problem was recently addressed by Walczak and co-workers in the sugar series, by capitalizing on stereospecific cross-coupling reactions of anomeric stannanes with aromatic iodides, diaryliodonium triflates and acyl donors in the presence of a palladium catalyst.^[19] However, these methods are based on the use of activated glycosyl donors as precursors and are not applicable to the synthesis of iminoglycosyl stannanes.

In our work, we have devised an efficient and stereoselective approach to iminoglycosyl stannanes whereby *path d* to imino-*C*-glycosides (Scheme 1) becomes feasible. In the course of our investigations on the reactivity of *N*-*tert*-butanesulfinyl glycosylamines (**10**), which are useful precursors of imino-*C*-glycosides by an addition-cyclisation strategy,^[13] we wondered whether it would be possible

generate the reactive imine by ring opening, the lithium ion coordinates to the liberated alkoxide (**15**). The tin-lithium reagent then coordinates to the lone pair of the sulfoxide moiety and the addition proceeds through an acyclic transition state of type **A**, following the Felkin–Ahn rules (Scheme 3).^[26]

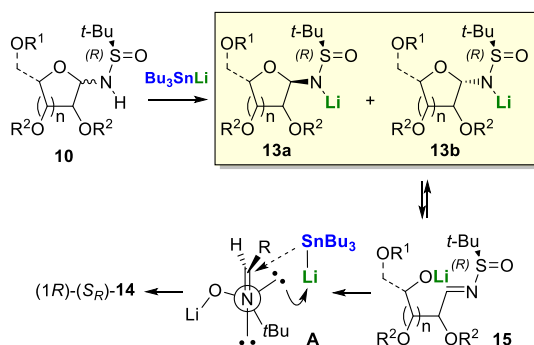
Table 1. Addition of Bu₃SnLi to glycosylamines **10a–e**.^{a)}



Entry	Substrate	Product	14 ratio (1 <i>R</i> :1 <i>S</i>) ^{b)}	14 yield (%) ^{c)}
1	(<i>S_R</i>)- 10a	(1 <i>R</i>)-(<i>S_R</i>)- 14a	> 98:2	89
2	(<i>S_S</i>)- 10a	(1 <i>S</i>)-(<i>S_S</i>)- 14a	> 2:98	81
3	(<i>S_R</i>)- 10b	(1 <i>R</i>)-(<i>S_R</i>)- 14b	> 98:2	85
4	(<i>S_S</i>)- 10b	(1 <i>S</i>)-(<i>S_S</i>)- 14b	> 2:98	87
5	(<i>S_R</i>)- 10c	(1 <i>R</i>)-(<i>S_R</i>)- 14c	> 98:2	84
6	(<i>S_S</i>)- 10c	(1 <i>S</i>)-(<i>S_S</i>)- 14c	> 2:98	81
7	(<i>S_R</i>)- 10d	(1 <i>R</i>)-(<i>S_R</i>)- 14d	> 98:2	81
8	(<i>S_S</i>)- 10d	—	—	nc
9	(<i>S_R</i>)- 10e	(1 <i>R</i>)-(<i>S_R</i>)- 14e	> 98:2	71

^{a)}Typical conditions (**G.P. B**): (1) *i*-Pr₂NH (3.5–4.0 eq.), *n*-BuLi (3.5–4.0 eq.), THF, 0 °C, 30 min; (2) Bu₃SnH (3.5–4.0 eq.), –78 °C, 15 min; (3) **10a–e**, THF, –78 °C; (4) The reaction mixture is allowed to reach a given temperature over 1.5–6 h. ^{b)}d.r determined on crude mixture using ¹H-NMR spectroscopy. ^{c)}Isolated yields (column chromatography). nc: no conversion.

The high stereoselectivity of the addition is quite remarkable and it is important to note that the potential chelating effect of the sugar functions does not divert the stereochemical mechanism away from the sulfinyl group, thus providing high stereocontrol in a tunable mode.



Scheme 3. Stereochemical model for the addition of Bu₃SnLi to compounds **10a–e**.

The lack of reactivity of a single glycosylamine, namely pyranosylamine (*S_S*)-**10e** is surprising and may be rationalized. In our DFT calculations on the

addition of [M]CF₂P(O)(OEt)₂ (M = Li, MgX) to compounds of type **10**, we assumed that internal complexation of the lithium ion by either the open-chain imine transient entity (**15**) and/or the various ether protecting groups would be entropically favored over solvent complexation.^[12g] In the (*S_S*)-D-xylopyranosylamine series, these interactions and possibly stereoelectronic factors might prevent opening of the metallated *N*-*tert*-butanesulfinyl-*N*-glycoside intermediate **13** (Scheme 3) to a reactive imine and therefore shut down the addition reaction.

Cyclization Reactions. As illustrated in Scheme 4, it was then possible to mesylate compound (1*R*)-(*S_R*)-**14a** (step a) through general procedure D-1 (**G.P. D-1**). As a bonus, the corresponding mesylated stannylsulfinamide (1*R*)-(*S_R*)-**16a** proved to be quite stable, surviving chromatography on silica gel and it was isolated in excellent yield (90%).

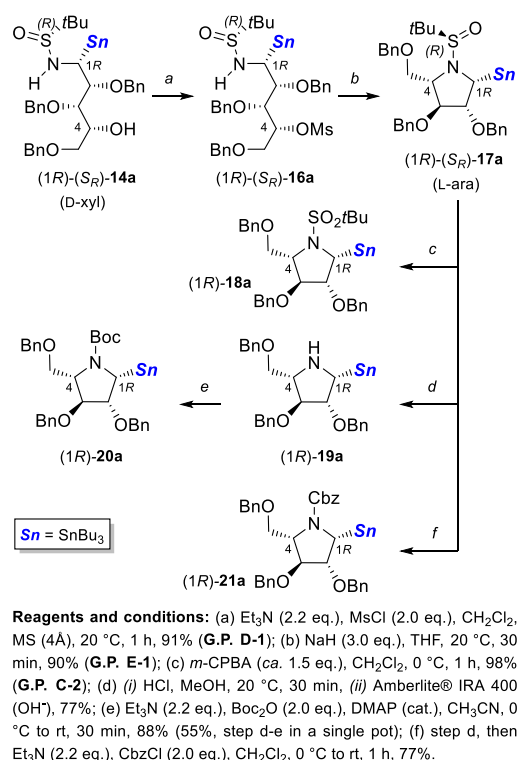
Our first attempts at cyclization using bulky bases such as *t*-BuOK or Et₃N in THF failed. Alternatively cleavage of the sulfinyl group (HCl in MeOH) and subsequent ring closure promoted by a basic resin was also unsuccessful.^{[12g],[13],[27]} The problem could be solved using an excess of sodium hydride (NaH, 3 equiv.) in THF at room temperature. From D-xylo (1*R*)-(*S_R*)-**16a**, the L-arabino (1*R*)-(*S_R*)-**17a** was obtained in good yield (90%, step b, **G.P. E-1**), resulting from an inversion of the configuration at C-4 upon ring closure. Oxidation of the labile sulfinyl group into a robust sulfonylamide was next carried out and compound (1*R*)-**18a** was obtained in almost quantitative yield (98%, step c) through general procedure C-2 (**G.P. C-2**).

One of the advantages of the *tert*-butanesulfinyl auxiliary is that it can be easily removed by treatment with an acid to provide free amines.^[28] Unlike Chong's investigations on α-sulfinylaminoorganostannanes,^[23b] we were able to remove the sulfinyl group in our sugar scaffold under acidic conditions (HCl, MeOH) without any noticeable degradation. Subsequent neutralization using a basic ion-exchange resin furnished (1*R*)-**19a** in good yield (77%, step d).

Compound (1*R*)-**19a** was thereafter reacted with Boc₂O in presence of DMAP in CH₃CN (step e) to provide the *N*-Boc derivative (1*R*)-**20a** (88%). Importantly, the formation of (1*R*)-**20a** could also be effectively promoted from (1*R*)-(*S_R*)-**17a**, carrying step d and e in a single pot, albeit with a slight decrease of the overall efficacy (55% vs. 68% overall). The *N*-Cbz protected iminosugar (1*R*)-**21a** was prepared similarly (see step f) and it was observed to exist as a 6:4 mixture of rotamers (77%).

the *tert*-butanesulfonyl (Bus) group was previously shown to be a useful protecting group, which could be used in Stille-type coupling reaction partners without notable degradation.^[29] In addition, it can be removed under acidic conditions,^[30] as shown in aminoacid

chemistry,^[31] and can be prepared from the *tert*-butanesulfinyl group via simple *m*-CPBA oxidation.^{[28],[30]} It was thus selected as key protecting group for the rest of the study.



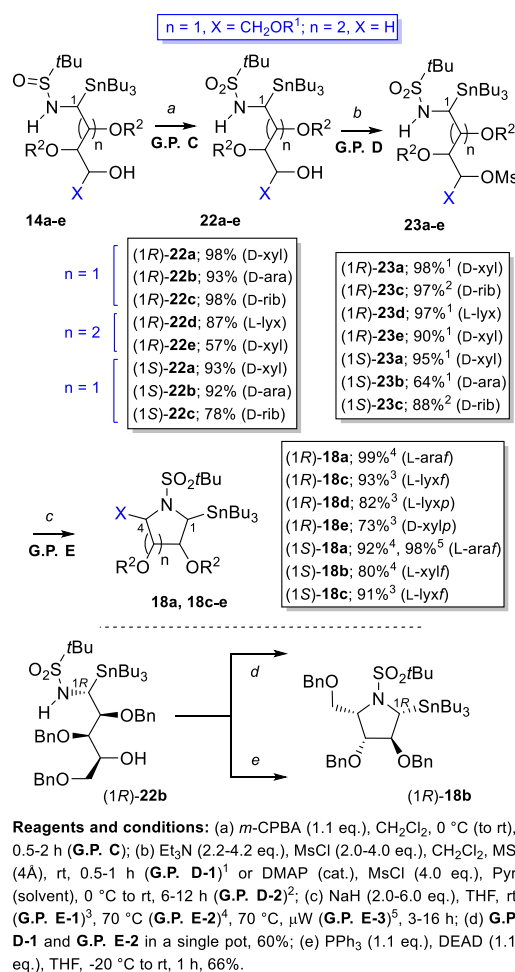
Scheme 4. Synthesis of anomeric iminosugar-1-tributylstannanes (1R)-(SR)-17a and (1R)-18-21a.

As described in Scheme 5, oxidation of aminoalditol derivatives **14a–e** was thus performed with *m*-CPBA (G.P. C) to give sulfonamides **22a–c** (in D-xylo, D-arabino and D-ribo series and with (1R)- and (1S)- configurations) in good to excellent yields (78–98%). In the case of the (1R)-L-lyxo and D-xylo derivatives, both featuring a primary hydroxyl group, compound (1R)-22d was obtained in good yield (87%), and (1R)-22e was formed in a non-optimized 57% yield. As expected, no erosion of the stereochemistry at C-1 could be detected. In the D-xylo, D-arabino and L-lyxo series, the same mesylation procedure as the one reported in Scheme 4 was then utilized to form compounds **23a–b**, (1R)-23d and (1R)-23e (G.P. D-1).

Surprisingly, for the D-ribo structure, the *tert*-butyldiphenylsilyl-protected pyrrole derivative **24** was isolated as a major by-product (see the Supporting Information (SI)). However, mesylation could be readily accomplished through G.P. D-2 (DMAP (cat.), excess MsCl) in pyridine to afford (1R)-23c and (1S)-23c in 97% and 88% yield respectively. Cyclisation of mesylates **23a–23e** could then be efficiently performed under the conditions of G.P.E, using NaH

at room temperature or at 70 °C, and in some cases under μ W irradiation (Scheme 5). Alternatively, cyclisation could also be performed under Mitsunobu conditions (*e.g.* (1R)-22b gave (1R)-18b in moderate yield (66%).

Gratifyingly, we found that cyclic, but also linear iminosugar stannanes **18a–e** and **22a–e** are chemically and configurationally stable. They can be purified by chromatography on silica gel, are stable against air and moisture, and retain anomeric configuration when exposed to light or when heated at 110 °C for several days. Furthermore, they can be stored at 0 °C under argon atmosphere for at least two years without erosion of the chirality at C-1.



Scheme 5. Synthesis of stereochemically-defined pseudo anomeric iminosugar stannanes.

Configuration Determination and J(C–Sn) as diagnostics. As summarized below in Fig. 2, the pseudo anomeric configuration of pyrrolidinyl iminosugar 1-stannanes was established based on the analysis of 2D nOesy spectra. The 1R configuration of L-arabinotols **17–21a** could be ascribed thanks to distinct nOe contacts between protons H-1 and H-4 in compounds (1R)-18a and (1R)-19a.

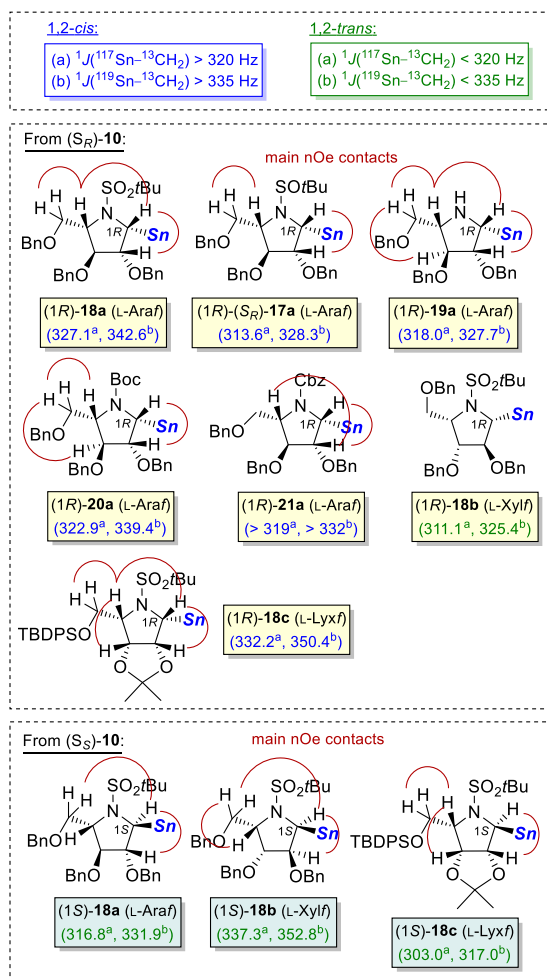


Figure 2. List of 1-stannyl pyrrolidinyl iminosugars, main nOe contacts and $^1J(\text{Sn}-\text{C})$ values.

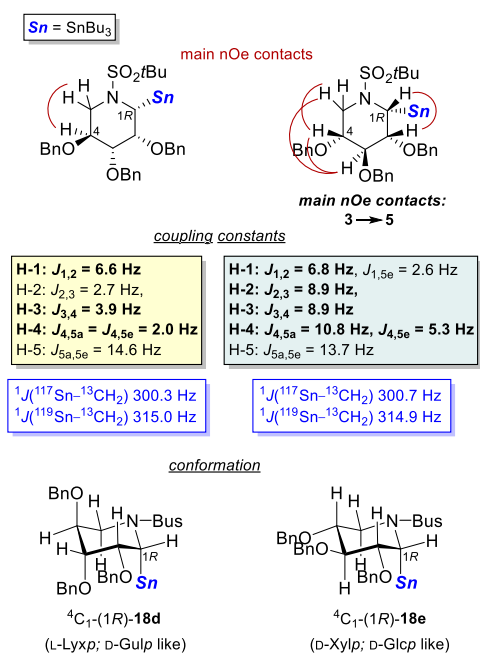


Figure 3. List of piperidinyl iminosugar 1-stannanes, main nOe contacts, coupling constants, conformations and $^1J(\text{Sn}-^{13}\text{C}-1)$ values.

The same relations were observed for L-arabinitol (1*R*)-**18c**. Regarding the L-xylf series, the 1*S* configuration of **18b** could in turn, be attributed owing to the main nOe contacts between protons H-1 and H-5.

In the piperidine series, the 1*R* configuration and $^4\text{C}_1$ conformational arrangement of imino-L-lyxitol (1*R*)-**18d** and imino-D-xylitol (1*R*)-**18e** were unambiguously settled according to the value of the vicinal H,H-coupling constants between ring protons (Fig. 3). Noteworthy, both compounds exist in the conformation in which the stannyl group is axial, in spite of its bulkiness. This is consistent with the conformational properties of N-sulfonylated piperidines carrying a substituent on the alpha-carbon. For steric reasons ($\text{A}_{1,3}$ – type strain), the substituent adopts preferentially an axial position.^[32]

The ^{13}C NMR data of the 1-stannylated iminosugars (see SI and Fig. 2 and Fig. 3), were then analyzed. Interestingly, they revealed a trend in the heteronuclear coupling constants between the ^{13}C -1 nucleus in the butyl chain and the anomeric $^{117}\text{Sn}/^{119}\text{Sn}$ nuclei.

As a rule, for the **N-sulfonylated and N-acylated** 1-C-stannylated iminoalditols with a pyrrolidine structure, in which the tin group is in a *cis* relationship with the group at C-2, the $^1J(^{117}\text{Sn}-^{13}\text{C})$ coupling constants are above 320 Hz (335 Hz for $^1J(^{119}\text{Sn}-^{13}\text{C})$). In contrast, in the C-1 stannanes in which the tin group is in a 1,2-*trans* orientation, these coupling constants are below these cutoffs. This trend however was not observed for pyrrolidines carrying a N-sulfinyl or N-H group ((1*R*)-(*S_R*)-**17a** and (1*R*)-**19a**).

For the six-membered piperidine series, we made observations similar to those reported for 1-tributystannyl hexopyranoses: the values of $^1J(^{117}\text{Sn}-^{13}\text{C})$, when the tin group occupies the axial position in a $^4\text{C}_1$ pyranose conformer are smaller than 305 Hz (319 Hz for $^1J(^{119}\text{Sn}-^{13}\text{C})$).^[19e] Compounds (1*R*)-**18d** and (1*R*)-**18e** follow the rule, thus widening further this prediction tool.

At this point, we were also intrigued to see whether we could extend the tool and predict the C-1 configuration at an earlier stage of the synthesis. No such correlations emerged in compounds **14**, **22** and **23**, even when pushing the heteronuclear spin-spin coupling analysis through the whole *n*-butyl chain. Early investigations from Vasella on 1-stannylated glycosides have shown that $^1J(\text{Sn}-\text{C})$ between the anomeric carbon and the tin substituent could be of interest in structural assignments.^[33] Although no relationship between $^1J(\text{Sn}-^{13}\text{C}-1)$ values and the configuration at C-1 could be found in the N-sulfonylated 1-stannyl iminoalditols **14**, we were delighted to identify a correlation for the series of N-sulfonylated compounds **22**.

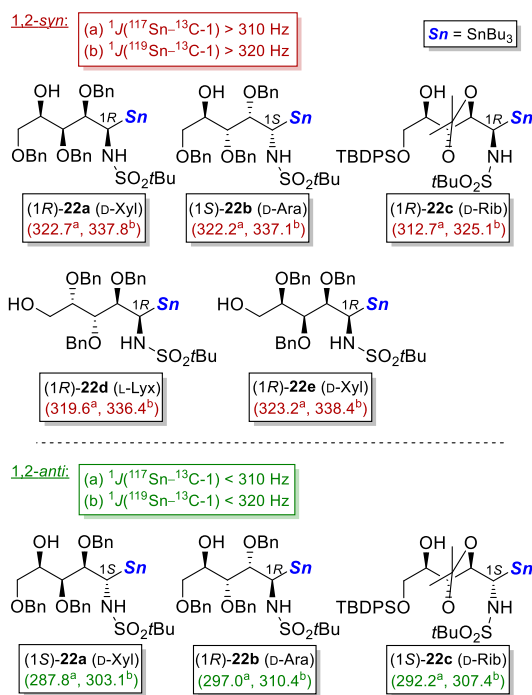


Figure 4. List of N-sulfonylated aminoalditol 1-stannanes **22**, and $^1J(\text{Sn}-^{13}\text{C}-1)$ trends.

As a rule, for the iminoalditols of type **22**, in which the amino group is in a 1,2-*syn* relationship with the alkoxy group at C-2, the $^1J(^{117}\text{Sn}-^{13}\text{C})$ coupling constants are above 310 Hz (320 Hz for $^1J(^{119}\text{Sn}-^{13}\text{C})$), whereas the C-1 stannylated iminoalditols with a 1,2-*anti* orientation have $^1J(\text{Sn}-^{13}\text{C})$ coupling constants below these cutoffs (see Fig. 4 and SI). Thus, it appears that the magnitude of this coupling constant is influenced by the configuration at the Sn-bearing carbon when steric effects are maximal and constitute a useful tool for configuration assignment.

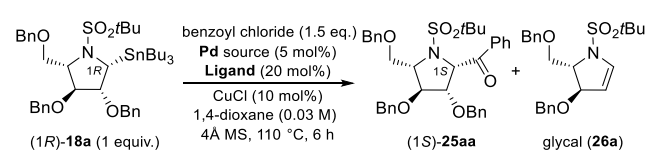
Migita–Kosugi–Stille Reaction Development. We next decided to investigate the Pd-catalyzed Migita–Kosugi–Stille cross-coupling reactions of the anomeric iminosugar stannanes: this reaction might unlock the synthesis of a diversity of iminoglycoside analogs, as suggested by the stereospecific Pd-mediated coupling reactions of chiral C(sp³) stannanes with aryl(aro)yl or alkyl(alkanoyl) halides^{[19],[34]}.

Since the pioneering work of Eaborn,^[35] Migita and Kosugi,^[36] in the 1970's and the extensive mechanistic studies by Stille,^[37] and others this reaction has progressively become one of the cornerstones in Pd-catalyzed C–C bond formation.^{[34],[38]} It is a mild process that tolerates a wide variety of functional groups, and which is frequently used in the synthesis of molecules of high complexity. Furthermore, organostannanes are relatively insensitive to moisture and oxygen, allowing for harsher reaction conditions.

In the sugar series, anomeric glycosylstannanes have been described by Sinaÿ,^[39] Kessler,^[40] and Vasella,^[33] and their first cross-coupling reactions with

benzoyl chloride and a range of chloro(thio)carbonates were investigated by Falck.^[41] The D-glucopyranosyl C-glycosides of interest were obtained in low to moderate yields, using excess electrophile, and with complete retention of configuration at the anomeric center. Walczak et al. recently demonstrated the scope and synthetic utility of the glycosylstannanes through coupling with aromatic iodides, diaryliodonium triflates and various thio- and selenoesters and generated an impressive array of mono- and oligo-saccharide-1-C-glycosides featuring free or diversely protected hydroxyl groups.^[19] However, no example of such reaction has been reported so far from iminosugars.

Table 2. Stille–Migita **reaction optimization**.^{a)}



Entry	Catalyst	Ligand	(1S)- 25aa (%) ^{b)}	26a (%) ^{b)}
1	Pd ₂ (dba) ₃	DPPE	7 ^{c)}	N.D.
2	Pd ₂ (dba) ₃	DPPF	16 ^{c)}	N.D.
3	Pd ₂ (dba) ₃	P(<i>o</i> -tol) ₃	7 ^{c)}	N.D.
4	Pd ₂ (dba) ₃	P(Cy) ₃	15 ^{c)}	N.D.
5	Pd ₂ (dba) ₃	P(Cy) ₃	35	N.D.
6	Pd ₂ (dba) ₃	PPh ₃	33	N.D.
7	Pd ₂ (dba) ₃	<i>R</i> -(+)-BINAP	0	N.D.
8	Pd(PPh ₃) ₄	–	31	N.D.
9	Pd(PPh ₃) ₂ Cl ₂	–	46	38
10	Pd ₂ (dba) ₃	P(Fu) ₃	36	N.D.
11	Pd ₂ (dba) ₃	AsPh ₃	18	N.D.
12	Pd ₂ (dba) ₃	TTMPP	22	N.D.
13	Pd ₂ (dba) ₃	P(<i>t</i> -Bu) ₃	40	N.D.
14	Pd ₂ (dba) ₃	JackiePhos	51	3

^{a)} Typical reaction conditions: Ar atmosphere, compound (1R)-**18a** (50 mg, 61.5 μmol, 1 equiv.), benzoyl chloride (1.5 equiv.), MS (4Å) (100 mg), Pd₂(dba)₃ (5 mol %), ligand (20 mol %), CuCl (10 mol %), heating at 110 °C for 6 h in 1,4-dioxane (0.03 M). ^{b)} Isolated yields (column chromatography). ^{c)} The reaction mixture was heated for 16 h. N.D. not determined.

Based on the single literature precedent by Chong et al. on the Stille coupling of stereochemically defined α-sulfonamido-organostannanes,^[29] and the aforementioned informations, optimization of the conditions for the Stille coupling of our iminoglycosyl stannanes with various electrophiles was thus started (Table 2). Robust L-*arabino* stannane (1R)-**18a** was selected as model substrate and the reaction was carried out with Pd₂(dba)₃ (5 mol%), 1,2-

bis(diphenylphosphino)ethane (dppe), copper(I)chloride (10 mol %) and benzoyl chloride (1.5 equiv.) in 1,4-dioxane for 16 h at 110 °C. A 7% yield of imino-*C*-glycoside **25aa** was obtained (entry 1).

Variation of the ligand was next envisaged with a careful choice of the phosphine, presenting various steric and electronic properties.^[42] With the chelating diphosphine 1,1'-bis(diphenylphosphino)ferrocene and the tertiary tricyclohexylphosphine (PCy₃), featuring high basicity (pK_a = 9.7) and large ligand cone angle (170°), the yield of the coupling product could be slightly increase to *ca.* 16% (entries 2 and 4). Interestingly with PCy₃, we noticed considerable decomposition and formation of an elimination product (an iminoglycal, **26a**). The yield of the coupling process could be increased to a decent 35% (isolated) yield by shortening the reaction time to 6 h (entry 5). Triphenylphosphine (PPh₃) and BINAP are some of the most often used ligands for palladium-catalyzed C–C bond-forming reactions. Thus, subsequent cross-coupling reactions were conducted with these phosphines, but a dramatic yield decrease or no improvement at all was noticed (entries 6 and 7).

Of note, in situ generation of tetrakis(triphenylphosphine)palladium(0) from Pd₂(dba)₃ and PPh₃ (entry 6) *vs.* direct addition of Pd(PPh₃)₄ (entry 8) did not produce any significant change of the reaction yield. With the square planar bis(triphenylphosphine)palladium(II) dichloride (Pd(PPh₃)₂Cl₂) as catalyst, the yield could be increased to a respectable 46%, although a large amount of iminoglycal **26a** (38%) was formed (entry 9).

As the rate-determining step in many Stille reactions is transmetalation, ligands of lower donicity, such as P(2-furyl)₃ and Ph₃As can dramatically enhance coupling reaction efficiencies.^[43] These two ligands were thus tested, but unsatisfactory 36% and 18% yields respectively were obtained with undetermined amounts of the iminoglycal derivative (entries 10 and 11). These disappointing results show that facilitating the Sn–Pd transmetalation step does not suppress the competitive elimination of the C-2 substituent from the transient anomeric palladium species nor facilitate reductive elimination.

The highly basic tris(2,4,6-trimethoxyphenyl)phosphine (TTMPP) used to perform the convenient synthesis of amino acids from α -sulfonamidoorganostannanes was thus tested thereafter,^[29] but a notable yield decrease was observed (22%, entry 12). The electron rich tri-*tert*-butylphosphine (P(*t*-Bu)₃) ligand was then examined and gave 1-*C*-benzoyl imino-*L*-arabinitol **27aa** in 40% yield (entry 13).

JackiePhos is a biaryl diphenylphosphine ligand, initially reported by Buchwald for the Pd-catalyzed amidations of aryl halides.^[44] It was recently shown by

Walczak to be quite efficient for the cross-coupling reactions of anomeric stannanes.^[19] It has two electron-rich methoxy and three large isopropyl groups to favor oxidative addition and possibly reductive elimination. Two electron-withdrawing 2,6-bis(trifluoromethyl)phenyl groups have also been set-up to counterbalance the trend towards transmetalation and reductive elimination. With this ligand, the iminoarabinitol-*C*-glycoside **25aa** was obtained in a respectable yield of 51%, with trace amounts (3%) of the elimination product (entry 14).

To fine-tune the reaction conditions solvent, additives, palladium and copper sources, stoichiometry of reagents, and reaction temperature were then screened (for details, see the Supporting Information). Based on the literature precedent that fluoride facilitates the Stille reaction,^[45] KF was selected as an additive for the optimization studies, but no beneficial effect could be observed. Given the accelerating role of Cu salts, additives such as CuCl (0.05, 0.1, 1, 2, 3 equiv.), *vs.* CuI, CuBr or CuCN in combination with Pd₂(dba)₃ as precatalyst were also studied. Of note, while CuCN and CuBr generated a yield comparable to CuCl, almost no conversion could be detected with CuI. The yield of the Stille reaction may also be improved by the use of several equivalents of the Cu(I) salt, but in this transformation, as low as 5–10 mol % of CuCl could be used without degradation of the reaction yield. It is important to mention that increasing the amount of CuCl gave rise to increased quantity of elimination product **26a**.

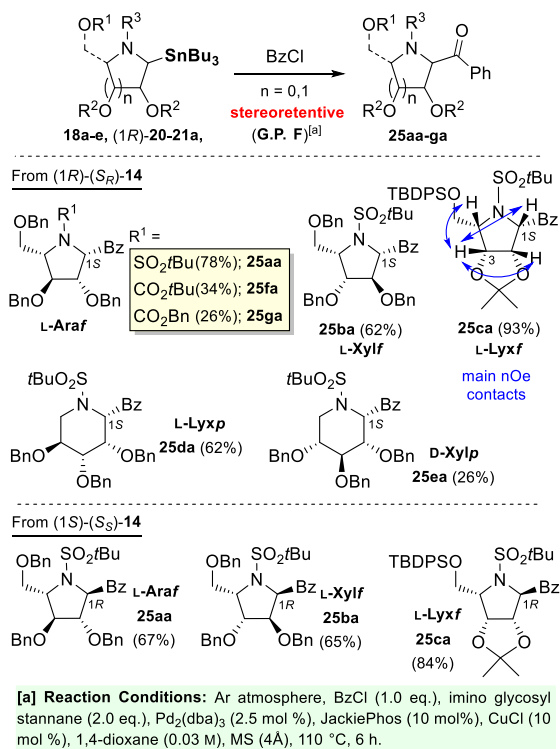
Likewise, in polar solvents (CH₃CN, NMP, DMF, DMSO, DMA) the 1-substituted iminoarabinitol derivative **25aa** was not obtained, the reaction providing again mainly by-product **26a** (19% in DMA and 72% in DMSO). Aromatic solvents may suppress glycal formation,^[19e] but in our case, the use of PhMe resulted in a lower yield of **25aa** (36%) and an increase of **26a** (18%). Other inorganic additives (ZnCl₂, FeCl₃, LiCl) were inefficient (typically 0% yield).

It is worth noting that all these results are consistent with a mechanism by a "closed" pathway (*vs.* the "open" pathway that leads to stereoinversion) involving a four-membered cyclic transition state and **complete retention of the configuration**. This also implies probable dissociation of one phosphine ligand prior to transmetalation. In addition, although no conversion was noted when the coupling reaction was performed without CuCl or Pd₂(dba)₃, at this point it is difficult to determine whether copper participates with palladium in a bimetallic catalytic cycle or if it is only Pd catalysis and Cu(I) acting as a ligand scavenger.^{[34],[46]}

Overall the optimized conditions were the following: stannane (1*R*)-**18a** (1 or 2 equiv.), benzoyl chloride (1.5 or 1 equiv.), MS (4Å), Pd₂(dba)₃ (2.5 mol %), ligand (10 mol %), copper(I) chloride (10

mol %), heating at 110 °C for 6 h in 1,4-dioxane (see general procedure F, **G.P. F**), **trace amounts (6%)** of the elimination product being obtained.

Migita–Kosugi–Stille Reaction Scope. The optimized iminoglycosyl-benzoyl cross-coupling conditions were thereafter tested in reactions with various iminoalditol derivatives **18a–e**, (1*R*)-**20a** and (1*R*)-**21a**, (Scheme 6).



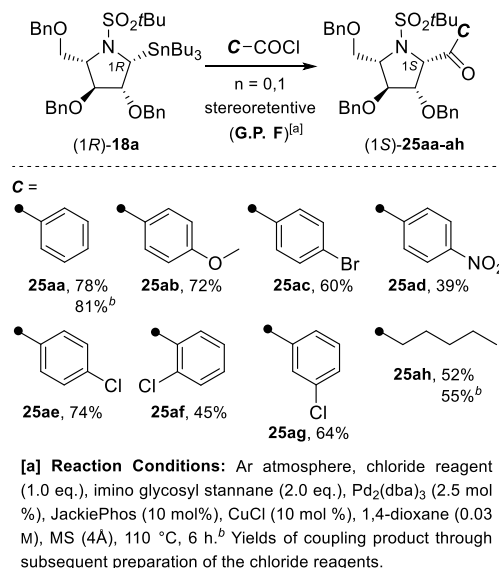
Scheme 6. Scope of C-glycosylation with anomeric iminoglycosyl tributylstannanes.

Pleasingly, we found that the general set of conditions is operational for the whole series of N-sulfonylated imino-L-arabino, L-xylitol and L-lyxose alditols derivatives, giving rise to the corresponding C-glycosides in moderate to good yields (62–93%) and stereospecifically. The 1*S* configuration of **25ca** could be assigned thanks to distinct nOe contacts between protons H-1 and H-3. One notes that lower yields of the coupling products were obtained from D-xylitol piperidine (1*R*)-**18e** (to give (1*S*)-**25ea**, 23% *e.g.*, 60% brsm) and from the two L-arabino pyrrolidines (1*R*)-**20** and **21a** ((1*S*)-**25fa** and (1*S*)-**25ga** formed in 34% and 26% yield respectively), which are not protected with a *tert*-butanesulfonyl group. For compounds **25fa** and **25ga**, the low yields can be explained by the formation of some deprotected products and some degradation.

Moreover, in case of the chemically stable imino-D-xylitol derivative (1*R*)-**25ea**, 24% of iminoglycosyl of type **26** (*e.g.*, **26e**, see SI) was isolated along with 53%

of unreacted 1-stannyl iminosugar (1*R*)-**18e**. For this particular substrate, the search for better ligands appear to be mandatory.

Importantly, N-sulfinylated compound (1*R*)-(1*S*)-**17a** was also tested in the Pd-catalyzed cross-coupling, but total decomposition was observed. The scope of the reaction was next further explored using anomeric stannane (1*R*)-**18a** and a range of substituted aryl chlorides and an *n*-alkanoyl reagent (Scheme 7).



Scheme 7. Scope of C-glycosylation with *n*-alkanoyl and aryl chlorides.

Gratifyingly, the yield of the coupling was quite good for most products. The performance of the acylation reactions did not seem to be correlated with the electronic properties of the aryl chloride reagents, except with a *p*-NO₂ group. However, sterics may be more important as it is observed that the yields of the reactions with *p*-Cl-, *m*-Cl- and *o*-Cl-benzoyl chloride decrease in this order (72%, 64% and 45% for **25ae**, **25ag** and **25af** resp.). Notably, with hexanoyl chloride, a significant 52% yield of **25ah** was achieved. Moreover, it is worth mentioning that the synthesis of the acyl chlorides from carboxylic acids (general procedure G, **G.P. G**), followed by cross-coupling with the anomeric iminosugar stannanes afforded **25aa** and **25ah** with an efficacy similar to commercial chlorides. These results thus open opportunities to incorporate acyl groups of greater complexity, such as for instance groups derived from amino acids or peptides for the synthesis of glycosyl aminoacids and glycopeptide mimics.^[47] Overall, it was demonstrated that the stereospecificity of the cross-coupling reactions is linked exclusively to the C-1 configuration of both iminoglycosyl stannanes anomers, with retention of the ‘anomeric’ configuration; furthermore, the problem of competitive β elimination of the

oxygen-based groups at C2 was addressed and the formation of unsaturated by-products **26** minimized. No epimerization of the chiral centers was noted.

Further data from literature, in particular from the work of Behr and Defoin on the configurational and conformational analysis of five-membered *N*-acyl-iminosugars, fully support our configurational assignments: it was shown that iminofuranosyl scaffolds tend to exhibit characteristic *J*(2,3) and *J*(4,5) coupling values for cis-relationships (*J* > 5.0 Hz) and *J* < 3 Hz for trans ones.^[48] As regards the stannyl iminosugar derivatives, this general trend was observed for all our pyrrolidine related to pentofuranoses (see SI): *J*(1,2) couplings when H-1 and H-2 are in cis-relationship were in the range of 4.3 to 6.0 Hz, whereas the coupling between H-1 and H-2 in 1,2-trans isomers is very small with H-1 appearing as a broad singlet. From the nOe analysis of L-lyxo derivative (1*S*)-**25ca** and ¹H NMR investigations on compounds of type **25**, it seems likely that the rule is conserved in the aroyl and hexanoyl imino-C-glycoside series (7.1 < *J* < 8.7 Hz for *J*(1,2)-cis and broad singlet for H-1 in 1,2-trans derivatives, see SI). As a result, based on literature precedents and on our investigations, anomeric stannanes undergo highly specific cross-coupling reactions with *n*-alkanoyl and aroyl chlorides through a **stereoretentive** mechanism.

Conclusion

In conclusion, we have developed an efficient approach for the preparation of 1-C-stannylated iminosugar anomers and their utilization in Stille cross-coupling reactions. The methodology demonstrated a good substrate scope and protecting group tolerance and has the great advantage of being tunable, i.e., the pseudoanomeric configuration of the 1-C-stannyl iminosugars and of the resulting 1-C-acylated iminosugar-C-glycosides can be chosen by selecting the configuration of the sulfinyl group in the starting *N*-tert-butanefulfinyl glycosylamines.

The iminoglycosyl stannanes were prepared by way of the stereospecific addition of tributylstannyl lithium to (*S_R*)- or (*S_S*)-*N*-tert-butanefulfinyl glycosylamines, followed by an oxidation and/or a mesylation-cyclization strategy. From *J*(C–Sn) data, a practical tool for the assessment of the chirality at C-1 was established. As a rule, (*S_S*)-glycosylamines [(*S_S*)-**10**] gave (1*S*)-configured stannylated iminosugar derivatives and (*S_R*)-glycosylamine [(*S_R*)-**10**], the (1*R*)-stereoisomer respectively. The iminosugar-derived organostannanes were then efficiently cross-coupled through Migita–Kosugi–Stille reactions with a range of variously substituted aroyl chlorides and an alkanoyl chloride, in a stereoretentive C–C bond forming process. Such compounds are desirable advanced intermediates for the synthesis of a diversity of glycoside mimics which could act as inhibitors of carbohydrate-processing enzymes. The generality of

the Stille coupling was investigated on 12 1-C-stannyl anomers, exhibiting furanose-like and pyranose-like skeletons. Eighteen examples of coupling products from the various stannylated iminosaccharides were obtained in moderate to excellent yields. Under the optimized coupling conditions, the β-elimination pathway was suppressed using a bulky phosphine ligand (JackiePhos) and a high stereoselectivity of the process was attained. Experimental evidence support a mechanistic rationale according to which the anomeric stannanes undergo stereoretentive cross-coupling reaction by way of a four-membered cyclic transition state. Taken together, these methodologies represent original and powerful strategies providing iminosugar-C-glycosides through an effective and hitherto unprecedented transformation. This synthetic tool allows for late stage coupling of iminosugar moieties with various aglycones and thus provides a means of performing the synthesis of imino-C-glycosyl compounds in a diversity-oriented approach. Further extension of the coupling reaction to other electrophiles such as aryl halides or heterocyclic halides should significantly expand the scope of the process and is currently under investigations.

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