

Multi-scale analysis and modelling of collective migration in biological systems

Andreas Deutsch, Peter Friedl, Luigi Preziosi, Guy Theraulaz

▶ To cite this version:

Andreas Deutsch, Peter Friedl, Luigi Preziosi, Guy Theraulaz. Multi-scale analysis and modelling of collective migration in biological systems. Philosophical Transactions of the Royal Society B: Biological Sciences, 2020, 375 (1807), pp.20190377. 10.1098/rstb.2019.0377. hal-02988844

HAL Id: hal-02988844 https://hal.science/hal-02988844v1

Submitted on 4 Nov 2020

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

PHILOSOPHICAL TRANSACTIONS B

royalsocietypublishing.org/journal/rstb

Introduction



Cite this article: Deutsch A, Friedl P, Preziosi L, Theraulaz G. 2020 Multi-scale analysis and modelling of collective migration in biological systems. *Phil. Trans. R. Soc. B* **375**: 20190377. http://dx.doi.org/10.1098/rstb.2019.0377

Accepted: 11 June 2020

One contribution of 14 to a theme issue 'Multiscale analysis and modelling of collective migration in biological systems'.

Subject Areas:

behaviour, biophysics, cellular biology, cognition, computational biology, developmental biology

Keywords:

collective migration, collective behaviour, selforganization, multi-scale analysis, mathematical biology, cancer

Author for correspondence:

Guy Theraulaz

e-mail: guy.theraulaz@univ-tlse3.fr

Multi-scale analysis and modelling of collective migration in biological systems

Andreas Deutsch¹, Peter Friedl^{2,3,4}, Luigi Preziosi⁵ and Guy Theraulaz^{6,7,8}

(D) AD, 0000-0002-9005-6897; PF, 0000-0002-0119-4041; GT, 0000-0002-3062-4215

Collective migration has become a paradigm for emergent behaviour in systems of moving and interacting individual units resulting in coherent motion. In biology, these units are cells or organisms. Collective cell migration is important in embryonic development, where it underlies tissue and organ formation, as well as pathological processes, such as cancer invasion and metastasis. In animal groups, collective movements may enhance individuals' decisions and facilitate navigation through complex environments and access to food resources. Mathematical models can extract unifying principles behind the diverse manifestations of collective migration. In biology, with a few exceptions, collective migration typically occurs at a 'mesoscopic scale' where the number of units ranges from only a few dozen to a few thousands, in contrast to the large systems treated by statistical mechanics. Recent developments in multi-scale analysis have allowed linkage of mesoscopic to micro- and macroscopic scales, and for different biological systems. The articles in this theme issue on 'Multi-scale analysis and modelling of collective migration' compile a range of mathematical modelling ideas and multi-scale methods for the analysis of collective migration. These approaches (i) uncover new unifying organization principles of collective behaviour, (ii) shed light on the transition from single to collective migration, and (iii) allow us to define similarities and differences of collective behaviour in groups of cells and organisms. As a common theme, self-organized collective migration is the result of ecological and evolutionary constraints both at the cell and organismic levels. Thereby, the rules governing physiological collective behaviours also underlie pathological processes, albeit with different upstream inputs and consequences for the group.

This article is part of the theme issue 'Multi-scale analysis and modelling of collective migration in biological systems'.

1. Introduction

Collective migration, the coordinated movement of groups of biological units, is observed across a multitude of scales in living systems (figure 1). At the cellular scale, it plays an essential role in biological development and the progression of cancer [2–4]. In particular, the collective migration of cohesive cell groups is observed during embryogenesis and is key to the formation of complex tissues and organs. Here, multicellular dynamics form the basis of epithelia, vascular and neuronal structures, with very different resulting shapes and functions. Similar collective cell behaviour is displayed by many invasive cancer types,

¹Department of Innovative Methods of Computing, Center for Information Services and High Performance Computing, Technische Universität Dresden, Dresden, Germany

²Department of Cell Biology, Radboud Institute for Molecular Life Sciences, Radboud University Medical Centre, Nijmegen, The Netherlands

³Cancer Genomics Center, Utrecht, The Netherlands

⁴Department of Genitourinary Medicine, University of Texas MD Anderson Cancer Center, Houston, TX, USA ⁵Department of Mathematical Sciences, Politecnico di Torino, Torino, Italy

⁶Centre de Recherches sur la Cognition Animale, Centre de Biologie Intégrative, Université de Toulouse, CNRS, UPS, Toulouse, France

⁷Centre for Ecological Sciences, Indian Institute of Science, Bengaluru, India

⁸Institute for Advanced Study in Toulouse, Toulouse, France

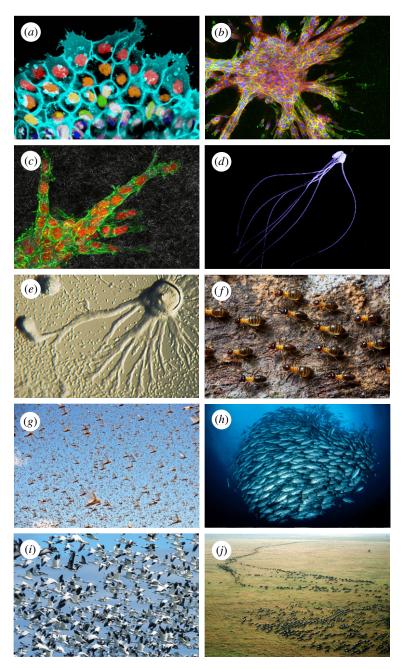


Figure 1. Collective migration in biological systems. (*a*) Collectively migrating neural crest cells in *Xenopus* embryos; membrane in cyan, nucleus pseudocoloured to distinguish leader (red) and trailing (blue) cells, photograph by Roberto Mayor; (*b*) E-cadherin negative MMT cells invading three-dimensional fibrillar collagen, photograph by Olga Ilina and Peter Friedl; (*c*) collective migration of cancer cells *in vitro* (red, nuclei; green, cytoskeleton; grey, collagen), photograph by Antoine Khalil and Peter Friedl; (*d*) electronmicrograph showing the aggregate formed by seven sperm cells of the dear mouse (*Peromyscus maniculatus*), modified from [1]; (*e*) collective migration and aggregation by chemotaxis in the social amoeba *Dictyostelium discoideum*; (*f*) a colony of termites (*Nasutitermes* sp.) on a march for food, following, and leaving, trail pheromones, photograph by Paul Bertner; (*g*) a migratory swarm of locusts (*Schistocerca gregaria*) in Isalo National Park, photograph by Tiphaine Desjardin; (*h*) a school of bigeye trevally (*Caranx sexfasciatus*), photograph by Ludovic Galko-Rundgren; (*i*) a flock of greater snow goose (*Chen caerulescens atlantica*), photograph by Will MacGregor; (*j*) the great wildebeest migration in the Serengeti National Park (*Connochaetes gnou*), photograph by Pete Aighton. (Online version in colour.)

where detrimental tissue disruption and collective metastasis consequently arise [5]. Collective migration at the organismic scale is observed in animal species that typically move over long distances and in a periodic manner implying a regular return to the region of departure [6]. The Natal sardine run is, without any doubt, one of the most spectacular examples of collective migration observed in the wild [7]. It is the second most important animal migration on the planet after that of wildebeest in the Serengeti [8], and it takes place every year from the beginning of May to the end of July along the east coast of South Africa. Millions of sardines

(Sardina pilchardus) leave the cold waters of the Cape region to go up north, following the 'Benguela' stream, which moves up along the coast, to join more temperate waters. Schools of sardines can reach sizes of more than 7 km length, 1.5 km width and 30 m depth. Collective migrations may be seasonal, but also irruptive and linked to the particular context. Thus, in locusts (Schistocerca gregaria), individual insects adapt their physiology, morphology and behaviour to gregarious life when environmental conditions (rain, abundance of food) become favourable and when their density exceeds the threshold of about 65 winged adults per square

metre [9–11]. The adults gather in gigantic swarms of several million insects, which can travel thousands of kilometres, fall on the crops and devastate everything on their path. That was one of the largest outbreaks of the last century and it is an ongoing problem in East Africa.

In each of these scenarios, collective movements require a tight behavioural coordination of individual units, based on the direct, proximate or indirect, mid- or long-range exchange of information between the units [12-15]. Information exchange then influences the behaviour of other individuals of the group at a later point in time, in support of collective coordination or dispersion. Physical and mechanical interactions are also involved in coordinated movements at the cellular level in combination with topographic cues [16,17]. The fine decoding of the mechanisms governing such collective phenomena has been facilitated by the development of new tracking techniques making it possible to reconstruct with an increasing level of precision the trajectories of cells and organisms in moving groups and over increasingly long periods of time [18-21]. High-precision tracking combined with mathematical modelling has enabled us to understand how the non-trivial properties of collective dynamics emerge at the macroscopic scale from the combined interactions between the units at the microscopic level [12,22,23]. For a long time, conceptually similar research fields evolved independently, i.e. theorizing on how individual behaviour influences collective migration [24-28] or identifying similar collective phases in migrating lymphocyte clusters and fish schools [27,29,30]. Recent developments in parameterization, modelling and multi-scale analysis now permit us to extract common principles as well as differences between these phenomena at cellular and organismic levels.

The overarching goal of this theme issue is to bring together biologists, physicians, physicists and mathematicians in order to meet this challenge. The articles span central facets of studying collective migration phenomena in both cellular and behavioural biology. This includes methodological advances in data analysis to reconstruct the physical and biological interactions between units; integrating strategies to parameterize and quantify the collective dynamics of cells, tissues and animal groups; and the development of discrete and continuous mathematical models of collective migration. It is our intention that this issue will become a resource for scientists wishing to learn about the methods and techniques used to investigate collective migration in biological systems, to identify the similarities and differences in the coordination mechanisms at work at the cellular and organismic levels, and to shape future interdisciplinary research agendas.

2. Methods and key issues

The general methodology used to study collective migration operates at two levels, to monitor and quantify (i) the behaviours of individual units and, in parallel, (ii) the collective organization and behaviours of the group, and then connect both levels at different scales (micro, individual; meso, group; and macro, whole population), by means of mathematical models [12,14,22,31–37]. Third, environmental heterogeneity guides and modulates the structures, dynamics and forms of collective behaviours [38,39]. Consequently, the experimental work performed under laboratory-controlled conditions

has to make it possible to identify and quantify the biological and physical interactions between units on the one hand and between units and the substrate on the other (i.e. the extracellular matrix in the case of cells, or the environment in case of animals), as well as the effects of these interactions on the behaviour of the units. To quantify such behaviour, the set of successive units' positions is recorded during a given period of time, usually at discrete time steps. Where possible, observations and measurements on individual and collective scales are also recorded in homogeneous environments, to decouple the modulation effects of these environmental parameters on individual interactions [40]. For studying collective movements of animal groups, experimental interventions include, for instance, precisely controlling temperature, brightness and humidity of the experimental room [41]. For monitoring cell groups, experimental variables include group size (from cell doublets to whole tissues and organisms spanning thousands of cells), chemical signals providing directionality (chemokines, cytokines), geometry (two-dimensional, three-dimensional), topology (linear, random), mechanical properties (stiffness, plasticity) and molecular organization of the extracellular environment [38]. From these experimental data, one can then reconstruct the trajectories of all units in a group and calculate different quantities, such as their instantaneous velocity and acceleration in response to directive cues present in the environment, as well as the group cohesion and polarization and, finally, reconstruct and model the functional form of interactions on an individual scale (e.g. [42,43] in this issue).

Understanding the non-trivial properties of collective dynamics that emerge at the macroscopic scale from interactions between units at the microscopic level requires the construction and use of mathematical models. The main function of a model is to predict or reproduce experimental results, including how individuals combine the information about the behaviour of their neighbours with environmental cues to coordinate their own motion and make collective decisions. Precise mathematical modelling (which is definitely not just fitting) is recognized as a major and often essential element of all research work in modern biology and especially in the research area of collective behaviour in biological systems (e.g. [12,44]. A model helps to identify the most important processes at play in the system. Thus, a model retains only a few, but highly characteristic and decisive, components of the real system. The model should consider as few parameters as possible so that, if a key component is missing in the model, it should be possible to extract particular experimental parameters that would invalidate the model. This strongly suggests that the retained minimal parameters jointly comprise and govern the key mechanisms underlying collective organization and decision-making. The qualitative and quantitative agreement between the model's predictions and experimental data contribute to our understanding of a system. A model failing at predicting or reproducing qualitatively or quantitatively (to some extent) basic experimental results is a sure sign that our understanding of the system is incomplete at best. In an iterative manner, new experiments are also commonly designed to try to put a model at fault, in order to test its domain of applicability/validity, and to find the minimal set of components that must be integrated to improve the model.

On the other hand, if the mathematical model is able to reproduce the experimental behaviour of interest in a satisfactory way, then one can modify the parameters to investigate what would happen in an experiment if a biological modification is introduced, prior to performing the experiments. Restricted to the collective dynamics this theme issue is focusing on, a key question one can ask the model aims at understanding how many changes at the individual level one should introduce in order to control the collective behaviour in the desired manner, how many leaders are necessary to control a group of uninformed followers, and how versatile and adjustable should the leadership role be to guarantee a collectively robust system.

Over the past years, a multitude of mathematical models for studying collective migration have been developed and are presented in this theme issue. Many of these models are derived from physical systems and adapted for the study of collective biological migration. For example, the cellular Potts model for interacting cell systems has been derived from a large-Q Potts model of interacting spins on a crystalline lattice, and the lattice-gas cellular automaton models for migrating cell populations have been adopted from lattice-gas models for incompressible fluids [45,46].

An important difference between living and non-living systems is the presence of stable heterogeneity in biological systems. For example, cells can differ strongly in speed and/or adhesivity. Therefore, it is important to quantify dynamic heterogeneity in experiments and in models. Only recently, mathematical models for collective dynamics have been developed to analyse the effects of heterogeneity [47].

To accommodate the increasing need of complexity and heterogeneity, various modelling and simulation platforms for the study of patterning and migration in multicellular systems have been introduced [48–50]. It will be important to make use of these platforms for the study of collective migration. Moreover, they might trigger the development of modelling and simulation platforms for animal populations too.

3. Overview of contributed papers

(a) Mathematical models and multi-scale analysis

After celebrated papers on collective migration of animal groups [51-55], the number of modelling approaches used to also describe the collective dynamics of biological cells has expanded considerably. Mathematical models look at collective phenomena from different levels: from the microscopic scale, e.g. by lattice-based and/or agent-based models, from the mesoscopic scale, e.g. by kinetic models, and from the macroscopic scale by systems of partial differential equations. From the mathematical point of view, such a burst of models operating at different scales has brought the need of connecting the models, highlighting not only similarities and differences, but also of clarifying how, in the various models, information at the smaller scale can be transferred to the larger scale. This requires the use and development of proper multi-scale methods [56]. Instead, staying on the same modelling level calls for a systematic comparison of different models and for the study of how different model terms affect the collective behaviour of the aggregates.

The most natural approach to describe the coordination of collective movements in groups of cells and organisms is the one that describes the behaviour of the single agents using individual-based or agent-based models. Such approaches can be split into two classes depending on whether a discrete lattice is used to describe space or not. Cellular and lattice-gas cellular automaton models are examples of on-lattice models [34]. Though on- and off-lattice models operate at the same spatial scale, the fact that space is described in different ways might lead to artefacts that need to be quantified. So, it is natural to ask whether such models yield similar results and to what extent. This is the aim of the contribution by Nava-Sedeño et al. [57]. Defining prototypic on- and off-lattice models of polar and apolar alignment, they show how to obtain an on-lattice from an off-lattice model for collective migration based on velocity alignment. For the purposes mentioned above, the macroscopic limits derived from the two model approaches are also compared, highlighting similarities and differences.

An important off-lattice model class for the study of collective migration is based on the idea of self-propelled particles (SPP), which describe autonomous agents converting energy from the environment into directed or persistent motion. Interacting SPP display fascinating collective phenomena. One of the most remarkable examples is the possibility of long-range orientational order in two-dimensional SPP models with continuum symmetry. The Mermin-Wagner theorem states that equilibrium systems with these characteristics cannot exhibit long-range order [58]. However, SPP with continuum symmetry and moving in a two-dimensional space, being out of equilibrium systems, can develop long-range orientational order, as shown for the first time in the seminal work by Vicsek et al. [54]. In this model, the velocity alignment of self-propelled agents is key for achieving different modes of collective migration. Meanwhile, various types of polar (ferromagnetic) and apolar (nematic) alignment have been studied with onand off-lattice models [59-61]. In their paper, Bernardi and Scianna [62] address the collective dynamics in groups of moving animals with a 'deterministic SPP approach'. They use a similar modelling approach as in the contribution by Colombi et al. [63] that focuses on a cellular system. The migration of the population is described by a set of firstorder ordinary differential equations for the animal positions, which can be formally derived from a second-order Newtonian approach under the assumption of an over-damped velocity response. Extending existing discrete models for collective migration that typically assume the constant speed of individual migration, the model by Bernardi & Scianna describes animals moving with changing speed and orientation, where the individual orientation response results from a set of behavioural stimuli. Model simulations capture different types of collective migration patterns depending on the particular choice of attractive, repulsive and alignment behavioural stimuli. The inclusion of an escape stimulus into the model allows the study of various hunting scenarios in a predator-prey system.

In agent-based and cellular automata models, it is straightforward to study the collective behaviours of cells or animals emerging from interactions with their spatial neighbours [27,64–67]. In fact, individual dynamics in such models explicitly take into account the interaction of particles located at different points in space. This is not the case in kinetic and continuous (e.g. reaction–diffusion) models that typically only represent reaction/interaction terms evaluated at the same space point. To deal with collective behaviours in

partial differential equation models that require us to take into account point-to-point interactions, one can include non-local operators over the region sensed by the individual that is characterized by a finite range and, e.g. a visual angle. The contribution by Chen et al. [68] summarizes how classical reaction-diffusion-transport models need to be transformed in order to take non-local effects into account. Modelling non-locality usually involves the introduction of convolutional operators in space. The focus of this contribution is to study the effect of the transport term that is typically in charge of modelling taxis-type migration phenomena. In fact, the literature has so far mainly focused on the transport aspect, though in principle a similar approach could be extended to other model components, to include non-local effects on growth and death. Moreover, the contribution also includes a discussion on how macroscopic reaction-diffusion-transport models can be obtained from microscopic individual-based models.

With a similar philosophy, in kinetic models, the tactic and kinetic responses of test individuals as a consequence of the non-local sensing of the surrounding environment and of the presence and behaviour of other individuals is considered by introducing non-local collision (or rather, interaction) operators [69-72]. In the kinetic framework, continuous models are recovered by diffusive or hydrodynamic limits, introducing, respectively, a parabolic or hyperbolic scaling of the kinetic model [71,73]. We mention that similar non-local operators should also be present in agent-based models when, in addition to other organisms, the individual needs to sense the environment for other important cues, typically chemo-attractants and substrate properties, which are usually described by continuous variables. In this theme issue, this feature is for instance present in the contribution of Colombi et al. [63] dealing with the migration of neural crest cells forming the posterior lateral line (PLL) of the zebrafish, which is a biological model system allowing the study of collective cell migration in embryonic development and pathological situations. PLL development involves the formation of a primordial cell cluster which collectively moves within the animal myoseptum. This process depends on the activity of specific diffusive chemicals, which trigger collective chemotactic cell migration and patterning. In this case, a multi-scale hybrid model is defined that integrates a discrete model for the cellular level and a continuous reaction-diffusion model for the molecular scale. With respect to previous mathematical models for PLL formation, the model developed by Colombi et al. [63] includes more molecular details, in particular, the two chemo-attractants FGF10 and SDF1a. Numerical model simulations can reproduce not only PLL development of wild-type embryos but also several pathological conditions induced by the fragmentation of the primordial cell cluster.

Another feature that can help classifying models for collective migration is whether they are deterministic or stochastic. Certainly, the former is a strongly simplifying assumption. Stochasticity is present in all behaviours of living organisms and cells. Cell and animal collective movement is no exception. Stochasticity might have a strong effect on the overall behaviour of the aggregate leading to what is called intrinsic noise at the group level. Theory predicts that the strength of intrinsic noise is not a constant but often depends on the collective state of the group. For this reason, it is also called a state-dependent noise or a

multiplicative noise. This effect is discussed in the contribution of Jhawar & Guttal [74]. By characterizing the role of stochasticity directly from high-resolution time-series data of collective dynamics, they argue that the group-level noise may encode important information about the underlying processes at the individual scale.

The difficult step in every model is usually to pass from qualitative to quantitative validation. Luckily, the recent development of new tracking techniques allows the provision, in principle, of large and accurate data sets that can be used for this purpose. Unfortunately, so far data are only available for a few systems. In this respect, the contribution by Escobedo *et al.* [43] presents a general method to extract interaction functions between individuals that are required to achieve collective migration. The method is then specifically applied to characterize social interactions in two species of shoaling fish, the rummy-nose tetra (*Hemigrammus rhodostomus*) and the zebrafish (*Danio rerio*), which both exhibit burst-and-coast motion. In principle, the method can be extended to other systems when data becomes available.

(b) Collective migration at the cellular level

The collective migration of cells as a cohesive group depends on both the interaction between neighbouring cells through mechanical cell–cell junctions and/or chemical information exchange [75]. Intercellular junctions can be weak and transient, as in swarming leucocytes or neural crest cells, or very tight and cohesive, as in moving epithelia or contracting muscle [76]. As a result of collective organization and dynamics, cells move as sheets, strands, clusters or ducts rather than individually, and use similar actin- and myosin-mediated protrusions. In the end, collective polarization, force generation and cell decision-making eventually result in complex tissue organization.

(i) Embryonic development

The migration of neural crest cells in the developing embryo has provided a rich resource for understanding the balance between adhesive and chemical signal integration between cells moving as a loose collective. In their contribution, Shellard & Mayor [77] provide an overview on how moving cells integrate mechanical cell-cell coupling, based on cellcell adhesion provided by cadherin molecules, that is counteracted by cell repulsion mediated by ephrins and semaphorins surface receptors (similarly to what is done in the contribution by Colombi et al. [63] mentioned above). Accordingly, a balance of adhesion and repulsion is critical in mediating cell alignment and group coordination. They further demonstrate the importance of chemotactic cell-to-cell signalling, which directs weakly adherent cells to align with neighbours and move along the joint trail. To recapitulate these principles, they summarize how computational modelling using Boïds, which simulates the flocking behaviour of birds [78], and robotics' algorithms to simulate the swarm intelligence [79] have contributed to identifying collective behaviours in the developing embryo as a self-organizing system.

In contrast to the developing embryo, sperm cell migration occurs in a fluid within body cavities, without direct adhesive sperm cell interaction. While only a single sperm cell will eventually fertilize the egg, the long-distance travelling of sperm cells towards the egg has recently been identified as collective dynamics. Schoeller *et al.* [80] here summarize quantitative

analyses on multicellular sperm dynamics, including wavelike patterns, multicellular coordination and directionality. Besides experimental observations, they describe mechanistic models that link the motion of individual sperm cells and their flagella to observed collective dynamics. As an emerging principle, multicellular coordination largely depends on the synchronization of the sperms' flagellae. Lastly, they discuss the importance of mucus, which provides a viscous extracellular environment for sperm propagation and multicellular coordination. Thus, as in multicellular organisms, an interplay between intercellular coordination and an integrating extracellular substrate jointly coordinate collective motion.

The development of a tree-like structure of epithelial branches is the central building block of several epithelial organs, including lungs, kidneys and the mammary gland. Their function is the secretion or absorption of molecules, coupled to a guiding tube-like duct structure for transport. Rens et al. [81] summarize the cellular mechanisms of branching morphogenesis, which include directed collective cell migration, oriented cell division, cell shape changes, cell differentiation and cell competition. Thereby, the moving epithelium interacts and is guided by the surrounding mesenchymal tissues, which ultimately dictates the geometry and function of the resulting organ. To extract the basic mechanism of autonomous branching morphogenesis, Rens et al. propose a simple mechanism, which is mediated by collective movement in the absence of proliferation and cross-talk with the surrounding mesenchymal tissue. Using combined cellular Potts and partial differential equation modelling, they show that cell-autonomous autocrine secretion of a morphogen (transforming growth factor-β, TGF-β) inhibits the formation of cell protrusions, which then leads to curvature-dependent inhibition of sprouting and duct formation. The outcome is consistent with the experimentally observed tissue geometrydependent determination of the branching sites, and it suffices for the formation of self-avoiding branching structures.

(ii) Cancer invasion and wound healing

Collective cell migration also plays a major role in wound repair and cancer invasion. It may lead to the formation of cell clusters, i.e. 'multicellular structures' that enable cells to better respond to chemical and physical cues, when compared with isolated cells. In particular, epithelial cells heal wounds via the migration of large sheets of cells with tight intercellular connections. Various mathematical models have been suggested that allow mechanistic insights which will benefit the clinical understanding of wound healing [82]. Moreover, cancer invasion is a hallmark of cancer progression [83]. Increasing evidence indicates that metastasis is driven by collective cell migration, where clusters of metastatic cells invade collectively into the vasculature and lymphatic system of cancer patients [5]. Meanwhile, mathematical models have been established to analyse potential mechanisms of invasion with a focus on glioma invasion [84].

The contribution by Kim *et al.* [85] focuses on glioblastoma multiforme (GBM) which is the most aggressive and malignant brain tumour. GBM is characterized by aggressive proliferation and cellular infiltration of healthy brain tissue. The primary treatment option is surgery followed by chemoradiation. Unfortunately, recurrence of the tumour is almost sure to occur within a rather short time span, the causes of which are largely unknown. It has been speculated that

surgery-mediated effects could play a major role. Kim et al. [85] define a mathematical model to study the implications of surgery on the dynamics of reactive astrocytes in the tumour microenvironment and test the following hypothesis: astrocyte injury from surgery induces a transition of reactive astrocytes into a stem cell-like phenotype which secretes the chemokine Cxcl5. This signal in turn promotes GBM proliferation and migration through the miR-451-LKB1-AMPK-OCT1mTor signalling pathway, which is known to regulate GBM proliferation and invasion. The resulting multi-scale mathematical model couples a differential equation model for the signalling pathway and a reaction-diffusion model for glucose, oxygen and the chemokine with a force-based model for the dynamics of tumour cells and astrocytes after surgery. The model shows how variations in glucose availability significantly affect the activity of signalling molecules and, in turn, lead to critical cell migration. The model also predicts that microsurgery of a primary tumour can induce phenotypical changes in reactive astrocytes and stem cell-like astrocytes promoting tumour cell proliferation and migration. Moreover, a novel anti-tumour strategy based on Cxcl5-targeting drugs was tested with the help of the model. It shows that the optimal use of anti-Cxcl5 drugs may slow down tumour growth and prevent cell invasion and recurrence.

The contribution of Vishwakarma et al. [86] studies the dynamics of collective cell migration during epithelial wound closure which bear similarities with a jammingunjamming transition in dense granular matter [87,88]. In this process, cells known as 'leader cells' migrate at the tips of cellular cohorts and provide directional cues to the followers. Interfacial geometry and bulk mechanical activity play an important role in regulating leader cell-mediated migration and subsequent collective migration. As cell density increases, the spatial distribution of velocities and forces becomes more homogeneous. Vishwakarma et al. [86] suggest the four-point susceptibility to characterize the dynamic heterogeneity of the cell population. This susceptibility has been previously introduced to quantify correlations between relaxation processes at different points in non-biological systems. It is shown that varying the susceptibility, by changing cell density, alters the number of leader cells at the wound margin. At a low heterogeneity level, wound closure is delayed, with decreased persistence, reduced coordination and disruptive leader-follower interactions. Finally, a microscopic characterization of cell-substrate adhesions illustrates how heterogeneity influences orientations of focal adhesions, i.e. affecting coordinated cell movements. Together, these results demonstrate the importance of the newly defined four-point susceptibility as readout of dynamic heterogeneity in epithelial wound healing.

(c) Collective migration at the organismic level

From the point of view of collective migration of animal groups, a previous theme issue of the *Philosophical Transactions* has already been devoted to behavioural ecology and movement ecology [89]. For this reason, this theme issue is not intended to cover the whole range of collective migration phenomena observed in groups of organisms. In fact, the examples discussed here concern more specifically the experimental analyses of behavioural interactions that take place at the individual scale and their impact on collective behaviour. In this respect, today, behavioural biologists have access to large volumes of data that make it possible to

precisely measure social interactions involved in the coordination of collective movements in animal groups, thanks to methods such as those presented by Escobedo et al. [43] in their article. Therefore, by systematically applying such methods it becomes possible to build a map of the different forms of social interactions used by different species to coordinate their movements when travelling in groups, according to their mode of propulsion, the shape of their body, their sensory and cognitive modalities and their living environment. Establishing such a map could bring important insights about the impact of these parameters on the various forms of social interactions, and in particular, the effect of an explicit alignment on the direction of movement of neighbours as observed in the rummy-nose tetra (Hemigrammus rhodostomus) [42] or a simple combination of long-range attraction and short-range repulsion as observed in the mosquitofish (Gambusia holbrooki) [90]. This information is crucial to understand the evolution of coordination mechanisms in moving animal

Social interactions between organisms do not only allow the coordination of individuals' actions, but also they endow a group with emergent properties such as the ability to efficiently escape predator attacks and navigate up noisy and weak thermal or resource gradients, even if no individual is capable of estimating the local gradient. The animal group as a whole thus functions as an integrated self-organizing sensor network, which significantly increases the range of effective perception of individuals [91]. It is also well-known that collective foraging based on indirect interactions in social or pre-social insects or based on long-range direct interactions in vertebrates improves the detection and exploitation of food sources. In their article, Ding et al. [92] investigate the foraging strategies in Caenorhabditis elegans, a 1 mm long nematode which has very limited sensory modalities and very short-range social interactions. By combining a computational model and experiments carried out on two strains of C. elegans, one social and the other solitary, they show that very simple social interactions such as the detection by an individual of a nearby worm allows the social phenotype to detect more efficiently patchy food sources in the environment.

These results suggest that the amount of social information needed by each individual to ensure the coordination of collective actions is limited. Still, this question remains widely debated. In the case of starlings (Sturnus vulgaris), previous studies have suggested that each bird within a flock interacts on average with a limited number of neighbours (typically seven) rather than with all neighbours within a fixed metric distance [93]. However, several recent studies suggest that, in fish, each individual in a school would collect only a limited amount of information on its environment thus avoiding cognitive saturation [94-96]. In animal groups, it is particularly difficult to design experiments to investigate such complex issues. In this respect, the results reported in the article by Jayles et al. [97] provide some elements to answer these questions. Using an artificial sensory device that filters out and adapts the amount of information delivered in real time to the participants in simple segregation task experiments with human groups, it is shown that the amount of information collected at the individual scale deeply affects the collective separation dynamics. However, they also show that each individual must only acquire a minimum amount of information on their first seven nearest neighbours to get an optimal level of segregation. In fact, the results show that the acquisition of additional information by individuals does not improve the collective performance of the group.

4. A path forward: integration of the concepts and methods in future research

Collective migration phenomena observed in cells and animal groups share many similarities and their comparative study enriches both cellular and behavioural biology. It is, therefore, not surprising that not only the questions addressed, but also the concepts and tools used to study and understand these phenomena, are rather similar in these research fields.

Thus, in addition to the common use of tracking, trajectory analysis and modelling techniques, certain concepts and processes stemming from the analysis of collective behaviour in social insects such as stigmergy have been successfully applied to the description of the mechanisms underlying collective cell migration. Stigmergy is a process by which social insects such as ants or termites coordinate their foraging and nestbuilding activities through indirect interactions by depositing pheromone trails on the ground or impregnating building materials with chemical compounds [98,99]. These chemical traces constitute sources of information and stimulation that, once perceived by other insects, trigger the execution of specific behaviours leading to the self-organization of the activities in a colony [100]. It has been shown that the self-organized multicellular behaviours that occur during the collective migration of the social bacterium Myxococcus xanthus over a surfacesolidified nutrient are coordinated, at least in part, through stigmergic processes [101]. Indeed, the components of the extracellular matrix and the furrows produced by vanguard cells facilitate and guide subsequent cellular movements. This process leads to the formation of trails that physically confine cells and facilitate their motility, which further reinforce and deepen the furrows, leading to the continued maintenance of the trail network. A similar mechanism has also been described in Pseudomonas aeruginosa that coordinates the expansion of its interstitial biofilms through the creation and remodelling of an interconnected network within a semisolid substratum [15]. Thus, stigmergy appears as an important self-organizing principle in biological systems. The study of collective cell migration could, therefore, benefit from the numerous experimental and modelling studies that have been carried out on stigmergic processes in social insects over the past 30 years [102–107].

The last decade has also seen further improvement in our comprehension of coordination processes and the way information pervades biological systems during collective motion, which has led to the development of new common research themes. One concept that has united several works in recent years is that of criticality [108,109]. An increasing number of works have indeed shown that biological systems such as collectives of cells, swarms of insects, flocks of birds and herds of sheep behave as though they are near the 'critical point' of a phase transition, like correlated spins in a magnet on the verge of ordering [110-115]. In flocks of starlings, for example, one can observe 'scale-free correlations', which occur on all possible length scales in the flock [116]. This hallmark of criticality is characterized by the fact that the velocity fluctuations of two distant birds mutually influence each other. The same property has been recently discovered in the self-organized aggregation of the social amoeba Dictyostelium discoideum in response to starvation [114]. In this species the local coupling between cells via cAMP secretion is tuned to a critical value, resulting in emergent long-range communication between cells that span the whole group independently of its size. Here, criticality brings some adaptive advantage allowing cells to act as a single, coherent unit and to make informed decisions to achieve optimal aggregate sizes for the most effective spore dispersal. Criticality also endows a group of cells or organisms with an extreme sensitivity to changes in the behaviour of a small number of individuals within the group [117]. Thanks to the social interactions, the reaction of these individuals can spread to all the other group members, allowing them to react more efficiently to external disturbances such as a predator attack. In sheep, it has been shown that the intensity with which individuals imitate each other (i.e. the strength of the coupling between sheep) leads a flock to be in a critical state, while optimizing two conflicting needs at the individual scale: the need to explore the maximum area of space to avoid inter-individual competition when foraging and the need to keep contact with the other group members to ensure cohesion and protection in case of danger [113]. These few examples suggest that natural selection operates not only on the particular form of social interactions but also on their intensity so that groups of cells or organisms have collective adaptive capabilities. Future work should focus on confirming these collective properties in a larger number of biological systems.

Selection processes also drive cancer progression and support cancer cell heterogeneity, a hallmark of cancer. So far, cancer evolution and collective migration have been studied separately [118]. It will be interesting to include mutations and selection in mathematical models for collective cell migration that will shed light on the evolution of collective cancer cell migration.

Originally, the study of collective migration has focused on the implications of inter-individual interactions. More recently, the importance of the non-cellular environment has been studied in cellular systems. In a recent paper [119], the physical geometry and available space (confinement) have been identified as key regulators forcing cells together, irrespective of their cell-cell adhesion properties. Whereas highly adhesive epithelial cells retain strict cell-cell interactions and migrate with strong neighbour correlation (like a solid-like state), decreasing cell-cell adhesion provides increasing degrees of freedom with diminishing intercellular coordination until near-complete independence of individual movements is reached in the cell group (active fluid). This transition reflects a fundamental building principle to physically control unjamming transitions and cancer cell invasion irrespective of the composition and stability of cell-cell junctions. The new study highlights the importance of the non-cellular microenvironment while previous work focused on the role of cell-cell interactions. Beyond cancer cell invasion, jamming-unjamming transitions are likely to shed light on the mechanisms of multicellular cooperation and collective behaviours in a range of cell models, as well as animal migration and coordination such as those described in ants and sheep [120–122].

5. Conclusion

One of the key properties of life is that it can switch between gas-, fluid- and solid-like states: cells or organisms move independently (gas state), form lose connections and migrate collectively (fluid state), or aggregate into immobile sheets and clusters (solid or jammed state). In the devastating example of malignant tumours, cells switch from a solid-like state to an invasive fluid or gas state, ultimately leading to deadly metastases. What are the decision-making mechanisms that underlie these transitions? Since the celebrated paper by Vicsek *et al.* [54], who suggested alignment interaction as a simple organization principle of collective migration, we have seen major progress in the study of collective phenomena. This theme issue highlights important developments: quantitative mathematical models and new multi-scale methods for their analysis have been introduced and new organization principles have been suggested.

A general tendency is that models become more and more individualized and heterogeneity and variability is considered more and more in various model extensions. Though some models presented in this theme issue take into account the variability of response to environmental cues, very little has been done in coupling the individual behaviour to the internal mechanisms determining that behaviour and the related stochastic response. In the context of cell migration, emerging questions focus on how the expression of a protein and the triggering of certain pathways determine the collective behaviour and, in particular, discriminate between an individual and a social behaviour, between a leading role and a follower role. Studying such questions would require nesting a microscopic model into an individual-based model with the ability to include the possibility that not all individuals may respond in the same way to the same environmental cues, introducing further stochastic aspects.

The presence of anomalous diffusion behaviour at the single-cell level with the transition from random walks to Levi's walks [123] and their social control by the group is also an interesting subject that deserves to be addressed in the future.

In this respect, also in view of applications to crowd and traffic management, what has been learned by studying the behaviour of cell and animal groups can be transferred to human groups, in particular by using the same research methods (e.g. [124,125]).

Looking at animals and cells from a modelling perspective highlights similarities but also important differences. Better knowledge about collective decision-making in cell populations is highly relevant for medical applications. New cancer therapy ideas will benefit from new insights into the control of collective migration that are also relevant for optimizing wound healing and regeneration since the controlled switching between single and collective migration is a key ingredient in these systems. New insights into collective behaviour will also be important for tissue engineering and optimization of organoid systems, and for explaining collective polarity, cell sorting and lumen formation. To this end, better knowledge of both single-cell behaviours and the resulting effects on the organism will identify currently unappreciated types of indirect and long-range information exchange beyond juxtacrine cellular mechano-coupling, which underlie collective coordination, including the deposition of information in tissues (chemical memory), swarm intelligence and jamming transitions [87,88].

Competing interests. We declare we have no competing interests.

Funding. The authors acknowledge the support of the Centre for Interdisciplinary Research (ZiF) at the University of Bielefeld, Germany, for funding the workshop 'Multiscale analysis and modelling of collective migration in biological systems' held on October 9–13, 2017.

The discussions and interactions spurred by the workshop have been very valuable in inspiring this special issue. L.P. acknowledges the support of the MIUR grant given to Dipartimenti di Eccellenza 2018-2022, project no. E11G18000350001. A.D. acknowledges support by the EU-ERACOSYS project no. 031L0139B. P.F. acknowledges the European Research Council (617430-DEEPINSIGHT), NIH-U54 CA210184-01 and the Cancer Genomics Center (CGC.nl).

Acknowledgements. We greatly appreciate the help of Helen Eaton, Senior Commissioning Editor at Philosophical Transactions B during all stages of preparation of this theme issue. G.T. gratefully acknowledges the Indian Institute of Science to serve as Infosys visiting professor at the Centre for Ecological Sciences in Bengaluru. A.D. acknowledges generous support by the Centre for Information Services and High Performance Computing, Technische Universität Dresden.

Authors' profile



Andreas Deutsch is head of the department 'Innovative Methods of Computing' at the Center for Information Services and High Performance Computing, Technische Universität Dresden, Germany. He holds a diploma in mathematics (University of Mainz), a PhD in biology (University of Bremen) and a habilitation in theoretical biology (University of Bonn). His research focuses on self-organization principles in multicellular systems, in particular collective migration, cancer dynamics and biological development. He pioneered multi-scale analysis for cellular automaton models of multicellular systems and co-authored the monograph 'Cellular automaton modelling of biological systems' (Birkhauser, Boston 2018).



Peter Friedl is director of the Microscopical Imaging Centre of the Radboud University Nijmegen Medical Centre, Nijmegen, The Netherlands, and holds a joint-faculty position at the University of Texas MD Anderson Cancer Center, Houston, TX. He received his MD from the University of Bochum in 1992 and PhD from the McGill University, Montreal in 1996, and is a board-certified clinical dermatologist and allergologist. His research interest is the mechanisms and plasticity of cell migration in immune regulation and cancer metastasis, with emphasis on cell-matrix adhesion, pericellular proteolysis and cell-cell communication during migration, with implications for molecular targeted and immunotherapy.



Luigi Preziosi is a member of the Accademia dei Lincei and full professor of Mathematical Physics at the Politecnico di Torino where he teaches Continuum Mechanics and Mathematical Models in Biomedicine. He received PhDs in Mechanics from the University of Minnesota (USA) and in Mathematics from the University of Naples, Italy. His research focuses on cancer modelling, cell migration and mechanical behaviour of cells and tissue using both continuum mechanics, kinetic models, and individual-based models. He has published ten books, mostly devoted to mathematical modelling of living systems.



Guy Theraulaz is a senior research fellow at the National Center for Scientific Research (CNRS) and an expert in the study of collective animal behaviour. He received a PhD in Neurosciences and Ethology in 1991 from Aix-Marseille University and a habilitation in quantitative behavioural biology from the University Paul Sabatier in Toulouse. His research focuses on the understanding of a broad spectrum of collective behaviours in animal societies by quantifying and then modelling the individual level behaviours and interactions, thereby elucidating the mechanisms generating the emergent, group-level properties. He has co-authored five books, among which *Swarm intelligence: From natural to artificial systems* (Oxford University Press, 1999) and *Self-organization in biological systems* (Princeton University Press, 2001) are considered as reference textbooks.

References

- Fisher HS, Giomi L, Hoekstra HE, Mahadevan L. 2014 The dynamics of sperm cooperation in a competitive environment. *Proc. R. Soc. B* 281, 20140296. (doi:10.1098/rspb.2014.0296)
- Friedl P, Gilmour D. 2009 Collective cell migration in morphogenesis, regeneration and cancer. *Nat. Rev. Mol. Cell Biol.* 10, 445–457. (doi:10.1038/nrm2720)
- Rørth P. 2009 Collective cell migration. *Annu. Rev. Cell Dev. Biol.* 25, 407–429. (doi:10.1146/annurev. cellbio.042308.113231)
- Weijer CJ. 2009 Collective cell migration in development. *J. Cell Sci.* 122, 3215–3223. (doi:10. 1242/jcs.036517)
- Cheung KJ, Ewald AJ. 2016 A collective route to metastasis: seeding by tumor cell clusters. *Science* 352, 167–169. (doi:10.1126/science.aaf6546)
- Dingle H. 1996 Migration: the biology of life on the move. New York, NY: Oxford University Press.
- Van der Lingen CD, Coetzee JC, Hutchings L. 2010 Overview of the KwaZulu-Natal sardine run. *Afr. J. Mar. Sci.* 32, 271–277. (doi:10.2989/ 1814232X.2010.501581)
- Estes R. 2014 The gnu's world: Serengeti wildebeest ecology and life history. Berkeley, CA: University of California Press.
- Kennedy JS. 1951 The migration of the desert locust (Schistocerca gregaria Forsk.). I. The behaviour of swarms. II. A theory of long-range migrations. Phil. Trans. R. Soc. Lond. B 235, 163–290. (doi:10.1098/ rsth.1951.0003)
- Anstey ML, Rogers SM, Ott SR, Burrows M, Simpson SJ. 2009 Serotonin mediates behavioral gregarization underlying swarm formation in desert locusts. Science 323, 627–630. (doi:10.1126/science. 1165939)
- Topaz CM, D'Orsogna MR, Edelstein-Keshet L, Bernoff AJ. 2012 Locust dynamics: behavioral phase change and swarming. *PLoS Comput. Biol.* 8, e1002642. (doi:10.1371/journal.pcbi.1002642)
- Camazine S, Deneubourg JL, Franks N, Sneyd J, Theraulaz G, Bonabeau E. 2001 Self-organization in biological systems. Princeton, NJ: Princeton University Press.
- Karsenti E. 2008 Self-organization in cell biology: a brief history. *Nat. Rev. Mol. Cell Biol.* 9, 255–262. (doi:10.1038/nrm2357)
- Deutsch A, Theraulaz G, Vicsek T. 2012 Collective motion in biological systems. *Interface Focus* 2, 689–692. (doi:10.1098/rsfs.2012.0048)
- Gloag ES, Javed MA, Wang H, Gee ML, Wade SA, Turnbull L, Whitchurch CB. 2013 Stigmergy: a key driver of self-organization in bacterial biofilms. Commun. Integr. Biol. 6, e27331. (doi:10.4161/cib. 27331)
- Trepat X, Wasserman MR, Angelini TE, Millet E, Weitz DA, Butler JP, Fredberg JJ. 2009 Physical forces during collective cell migration. *Nat. Phys.* 5, 426–430. (doi:10.1038/nphys1269)
- 17. Londono C, Loureiro MJ, Slater B, Lücker PB, Soleas J, Sathananthan S, Aitchison JS, Kabla AJ, McGuigan

- AP. 2014 Nonautonomous contact guidance signaling during collective cell migration. *Proc. Natl Acad. Sci. USA* **111**, 1807–1812. (doi:10.1073/pnas. 1321852111)
- Oates AC, Gorfinkiel N, Gonzalez-Gaitan M, Heisenberg CP. 2010 Quantitative approaches in developmental biology. *Nat. Rev. Genet.* 10, 517–530. (doi:10.1038/nrq2548)
- Olivier N et al. 2019 Idtracker.ai: tracking all individuals in small or large collectives of unmarked animals. Nat. Methods 16, 179–182. (doi:10.1038/ s41592-018-0295-5)
- Dell Al et al. 2014 Automated image-based tracking and its application in ecology. *Trends Ecol. Evol.* 29, 417–428. (doi:10.1016/j.tree.2014.05.004)
- Débarre D, Bourgine P, Santos A, Peyriéras N, Beaurepaire E. 2010 Cell lineage reconstruction of early zebrafish embryos using label-free nonlinear microscopy. *Science* 329, 967–971. (doi:10.1126/ science.1189428)
- Vicsek T, Zafeiris A. 2012 Collective motion. *Phys. Rep.* 517, 71–140. (doi:10.1016/j.physrep.2012. 03 004)
- Méhes E, Vicsek T. 2014 Collective motion of cells: from experiments to models. *Integr. Biol.* 6, 831–854. (doi:10.1039/C4IB00115J)
- 24. Couzin ID, Krause J, Franks NR, Levin SA. 2005 Effective leadership and decision-making in animal groups on the move. *Nature* **433**, 513–516. (doi:10. 1038/nature03236)
- Poujade M, Grasland-Mongrain E, Hertzog A, Jouanneau J, Chavrier P, Ladoux B, Buguin A, Silberzan P. 2007 Collective migration of an epithelial monolayer in response to a model wound. *Proc. Natl Acad. Sci. USA* 104, 15 988–15 993. (doi:10.1073/pnas.0705062104)
- Petitjean L, Reffay M, Grasland-Mongrain E, Poujade M, Ladoux B, Buguin A, Silberzan P. 2011 Velocity fields in a collectively migrating epithelium.
 Biophys. J. 98, 1790–1800. (doi:10.1016/j.bpj.2010. 01.030)
- Calovi DS, Lopez U, Ngo S, Sire C, Chaté, H, Theraulaz G. 2014 Swarming, schooling, milling: phase diagram of a data-driven fish school model. *New J. Phys.* 16, 015026. (doi:10.1088/1367-2630/ 16/1/015026)
- Calovi DS, Lopez U, Schuhmacher P, Sire C, Chaté H, Theraulaz G. 2015 Collective response to perturbations in a data-driven fish school model. J. R. Soc. Interface 12, 20141362. (doi:10.1098/rsif. 2014.1362)
- Malet-Engra G, Yu W, Oldani A, Rey-Barroso J, Gov NS, Scita G, Dupré L. 2015 Collective cell motility promotes chemotactic prowess and resistance to chemorepulsion. *Curr. Biol.* 25, 242–250. (doi:10. 1016/j.cub.2014.11.030)
- Rey-Barroso J, Calovi DS, Combe M, Moreau M, Wang X, Sire C, Theraulaz G, Dupré L. 2018
 Switching between individual and collective motility in lymphocytes is controlled by cell-matrix adhesion

- and inter-cellular interactions. *Sci. Rep.* **8**, 5800. (doi:10.1038/s41598-018-24222-4)
- 31. Alt W, Deutsch A, Dunn G (eds). 1997 *Dynamics of cell and tissue motion*. Boston, MA: Springer.
- Cai D, Dai W, Prasad M, Luo J, Gov NS, Montell DJ. 2016 Modeling and analysis of collective cell migration in an in vivo three-dimensional environment. *Proc. Natl Acad. Sci. USA* 113, E2134—E2141. (doi:10.1073/pnas.1522656113)
- Chauvière A, Preziosi L, Verdier C (eds). 2010 Cell mechanics: from single scale-based models to multiscale modeling. London, UK: Chapman and Hall/CRC.
- Deutsch A, Dormann S. 2018 Cellular automaton modeling of biological pattern formation: characterization, examples, and analysis. Basel, Switzerland: Birkhauser.
- Hatzikirou H, Deutsch A. 2008 Cellular automata as microscopic models of cell migration in heterogeneous environments. *Curr. Top. Dev. Biol.* 81, 401–434. (doi:10.1016/S0070-2153(07)81014-3)
- Preziosi L, Tosin A. 2009 Multiphase and multiscale trends in cancer modelling. *Math. Model. Nat. Phenom.* 4, 1–11. (doi:10.1051/mmnp/20094301)
- Weitz S, Blanco S, Fournier R, Gautrais J, Jost C, Theraulaz G. 2012 Modeling collective animal behavior with a cognitive perspective: a methodological framework. *PLoS ONE* 7, e38588. (doi:10.1371/journal.pone.0038588)
- Te Boekhorst V, Preziosi L, Friedl P. 2016 Plasticity of cell migration in vivo and in silico. *Annu. Rev. Cell Dev. Biol.* 32, 491–526. (doi:10.1146/annurev-cellbio-111315-125201)
- Pinter-Wollman N, Fiore SM, Theraulaz G. 2017 The impact of architecture on collective behavior. *Nat. Ecol. Evol.* 1, 0111. (doi:10.1038/s41559-017-0111)
- Stonko DP, Manning L, Starz-Gaiano M, Peercy BE. 2015 A mathematical model of collective cell migration in a three-dimensional, heterogeneous environment. *PLoS ONE* 10, e0122799. (doi:10. 1371/journal.pone.0122799)
- Jost C, Verret J, Casellas E, Gautrais J, Challet M, Lluc J, Blanco S, Clifton, MJ, Theraulaz G. 2007 The interplay between a self-organized process and an environmental template: corpse clustering under the influence of air currents in ants. J. R. Soc. Interface 4, 107–116. (doi:10.1098/rsif.2006.0156)
- Calovi DS, Litchinko A, Lecheval V, Lopez U, Pérez Escudero A, Chaté H, Sire C, Theraulaz G. 2018
 Disentangling and modeling interactions in fish with burst and coast swimming reveal distinct alignment and attraction behaviors. *PLoS Comput. Biol.* 14, e1005933. (doi:10.1371/journal.pcbi. 1005933)
- Escobedo R, Lecheval V, Papaspyros V, Bonnet F, Mondada F, Sire C, Theraulaz G. 2020 A data-driven method for reconstructing and modelling social interactions in animal groups. *Phil. Trans. R. Soc. B* 375, 20190380. (doi:10.1098/rstb.2019.0380)

- 44. Sumpter DJT. 2010 *Collective animal behavior*. Princeton, NJ: Princeton University Press.
- Chopard B, Ouared R, Deutsch A, Hatzikirou H, Wolf-Gladrow D. 2010 Lattice-gas cellular automaton models for biology: from fluids to cells. *Acta Biotheor.* 58, 329–340. (doi:10.1007/s10441-010-9118-5)
- Graner F, Glazier JA. 1992 Simulation of biological cell sorting using a two-dimensional extended Potts model. *Phys. Rev. Lett.* 69, 2013–2016. (doi:10. 1103/PhysRevLett.69.2013)
- Reher D, Klink B, Deutsch A, Voß-Böhme A. 2017 Cell adhesion heterogeneity reinforces tumour cell dissemination: novel insights from a mathematical model. *Biol. Direct* 12, 18. (doi:10.1186/s13062-017-0188-z)
- Ghaffarizadeh A, Heiland R, Friedman SH, Mumenthaler SM, Macklin P. 2018 PhysiCell: an open source physics-based cell simulator for 3-D multicellular systems. *PLoS Comput. Biol.* 14, e1005991. (doi:10.1371/journal.pcbi.1005991)
- Starruß J, de Back W, Brusch L, Deutsch A. 2014 Morpheus: a user-friendly modeling environment for multiscale and multicellular systems biology. *Bioinformatics* 30, 1331–1332. (doi:10.1093/ bioinformatics/btt772)
- Swat MH, Thomas GL, Belmonte JM, Shirinifard A, Hmeljak D, Glazier JA. 2012 Multi-scale modeling of tissues using CompuCell3D. *Methods Cell Biol*. 110, 325–366. (doi:10.1016/B978-0-12-388403-9. 00013-8)
- Carrillo JA, Fornasier M, Toscani G, Vecil F. 2010 Particle, kinetic, and hydrodynamic models of swarming. In Mathematical modeling of collective behavior in socio-economic and life sciences (eds G Naldi, L Pareschi, G Toscani), pp. 297–336. Cambridge, MA: Birkhäuser Boston.
- Cucker F, Smale S. 2007 Emergent behavior in flocks. *IEEE Trans. Autom. Control* 52, 852–862. (doi:10.1109/TAC.2007.895842)
- D'Orsogna MR, Chuang Y-L, Bertozzi AL, Chayes LS. 2006 Self-propelled particles with soft-core interactions: patterns, stability, and collapse. *Phys. Rev. Lett.* 96, 104302. (doi:10.1103/PhysRevLett.96. 104302)
- Vicsek T, Czirok A, Ben-Jacob E, Cohen I, Shochet O.
 1995 Novel type of phase transition in a system of self-driven particles. *Phys. Rev. Lett.* 75, 1226. (doi:10.1103/PhysRevLett.75.1226)
- Cucker F, Smale S. 2007 On the mathematics of emergence. *Jpn. J. Math.* 2, 197–227. (doi:10.1007/ s11537-007-0647-x)
- 56. Lesne A. 2013 Multiscale analysis of biological systems. *Acta Biotheor.* **61**, 3–19. (doi:10.1007/s10441-013-9170-z)
- Nava-Sedeño JM, Voß-Böhme A, Hatzikirou H, Deutsch A, Peruani F. 2020 Modelling collective cell motion: are on- and off-lattice models equivalent? *Phil. Trans. R. Soc. B* 375, 20190378. (doi:10.1098/ rstb.2019.0378)
- 58. Mermin ND, Wagner H. 1966 Absence of ferromagnetism or antiferromagnetism in one- or

- two-dimensional isotropic Heisenberg models. *Phys. Rev. Lett.* **17**, 1307. (doi:10.1103/PhysRevLett.17. 1307)
- Bär M, Grossmann R, Heidenreich S, Peruani F. 2020 Self-propelled rods: insights and perspectives for active matter. *Annu. Rev. Condens. Matter Phys.* 11, 441–466. (doi:10.1146/annurev-conmatphys-031119-050611)
- 60. Peruani F, Deutsch A, Bär M. 2006 Nonequilibrium clustering of self-propelled rods. *Phys. Rev. E* **74**, 030904. (doi:10.1103/PhysRevE.74.030904)
- Peruani F, Klauss T, Deutsch A, Voß-Böhme A. 2011 Traffic jams, gliders, and bands in the quest for collective motion. *Phys. Rev. Lett.* **106**, 128101. (doi:10.1103/PhysRevLett.106.128101)
- Bernardi S, Scianna M. 2020 An agent-based approach for modelling collective dynamics in animal groups distinguishing individual speed and orientation. *Phil. Trans. R. Soc. B* 375, 20190383. (doi:10.1098/rstb.2019.0383)
- Colombi A, Scianna M, Preziosi L. 2020
 Collective migration and patterning during early development of zebrafish posterior lateral line. *Phil. Trans. R. Soc. B* 375, 20190385. (doi:10.1098/ rstb.2019.0385)
- 64. Aoki I. 1982 A simulation study on the schooling mechanism in fish. *Bull. Jpn Soc. Sci. Fish* **48**, 1081–1088. (doi:10.2331/suisan.48.1081)
- 65. Huth A, Wissel C. 1992 The simulation of the movement of fish schools. *J. Theor. Biol.* **156**, 365–385. (doi:10.1016/S0022-5193(05)80681-2)
- Couzin ID, Krause J, James R, Ruxton GD, Franks NR. 2002 Collective memory and spatial sorting in animal groups. *J. Theor. Biol.* 218, 1–11. (doi:10. 1006/jtbi.2002.3065)
- Gautrais J, Ginelli F, Fournier R, Blanco S, Soria M, Chaté H, Theraulaz G. 2012 Deciphering interactions in moving animal groups. *PLoS Comput. Biol.* 8, e1002678. (doi:10.1371/journal.pcbi.1002678)
- 68. Chen L, Painter K, Surulescu C, Zhigun A. 2020 Mathematical models for cell migration: a non-local perspective. *Phil. Trans. R. Soc. B* **375**, 20190379. (doi:10.1098/rstb.2019.0379)
- 69. Eftimie R. 2012 Hyperbolic and kinetic models for self-organized biological aggregations and movement: a brief review. *J. Math. Biol.* **65**, 35–75. (doi:10.1007/s00285-011-0452-2)
- Carrillo J, Homann F, Eftimie R. 2015 Non-local kinetic and macroscopic models for self-organised animal aggregations. *Kinet. Relat. Models* 8, 413–441. (doi:10.3934/krm.2015.8.413)
- Loy N, Preziosi L. 2020 Kinetic models with non-local sensing determining cell polarization and speed according to independent cues. *J. Math. Biol.* 373–421. (doi:10.1007/s00285-019-01411-x)
- 72. Loy N, Preziosi L. 2020 Modelling physical limits of migration by a kinetic model with non-local sensing. *J. Math. Biol.* **80**, 1759–1801. (doi:10. 1007/s00285–020–01479-w)
- 73. Arlotti L, Bellomo N, de Angelis E. 2002 Generalized kinetic (Boltzmann) models: mathematical structures and applications. *Math. Models Methods*

- *Appl. Sci.* **12**, 567–591. (doi:10.1142/ S0218202502001799)
- Jhawar J, Guttal V. 2020 Noise-induced effects in collective dynamics and inferring local interactions from data. *Phil. Trans. R. Soc. B* 375, 20190381. (doi:10.1098/rstb.2019.0381)
- 75. Friedl P, Locker J, Sahai E, Segall JE. 2012 Classifying collective cancer cell invasion. *Nat. Cell Biol.* **14**, 777–783. (doi:10.1038/ncb2548)
- Friedl P, Mayor R. 2017 Tuning collective cell migration by cell-cell junction regulation. *Cold Spring Harb. Perspect. Biol.* 9, a029199. (doi:10. 1101/cshperspect.a029199)
- Shellard A, Mayor R. 2020 Rules of collective migration: from the wildebeest to the neural crest. *Phil. Trans. R. Soc. B* 375, 20190387. (doi:10.1098/ rstb.2019.0387)
- Reynolds CW. 1987 Flocks, herds and schools: a distributed behavioural model. In SIGGRAPH'87: Proc. 14th Annu. Conf. on Computer Graphics and Interactive Techniques, Anaheim, CA, pp. 25–34.
 New York, NY: ACM Press.
- Bonabeau E, Dorigo M, Theraulaz G. 1999 Swarm intelligence: from natural to artificial systems.
 Oxford, UK: Oxford University Press.
- 80. Schoeller SF, Holt WV, Keaveny EE. 2020 Collective dynamics of sperm cells. *Phil. Trans. R. Soc. B* **375**, 20190384. (doi:10.1098/rstb.2019.0384)
- Rens EG, Zeegers MT, Rabbers I, Szabó A, Merks RMH. 2020 Autocrine inhibition of cell motility can drive epithelial branching morphogenesis in the absence of growth. *Phil. Trans. R. Soc. B* 375, 20190386. (doi:10.1098/rstb.2019.0386)
- 82. Jorgensen SN, Sanders JR. 2016 Mathematical models of wound healing and closure: a comprehensive review. *Med. Biol. Eng. Comput.* **54**, 1297–1316. (doi:10.1007/s11517-015-1435-z)
- 83. Hanahan D, Weinberg RA. 2011 Hallmarks of cancer: the next generation. *Cell* **144**, 646–674. (doi:10.1016/j.cell.2011.02.013)
- 84. Alfonso JCL, Talkenberger K, Seifert M, Klink B, Hawkins-Daarud A, Hatzikirou H, Swanson KR, Deutsch A. 2017 The biology and mathematical modelling of glioma invasion: a review. *J. R. Soc. Interface* **14**, 20170490. (doi:10.1098/rsif.2017.0490)
- Kim Y, Lee D, Lawler S. 2020 Collective invasion of glioma cells through OCT1 signalling and interaction with reactive astrocytes after surgery. *Phil. Trans.* R. Soc. B 375, 20190390. (doi:10.1098/rstb.2019.0390)
- Vishwakarma M, Thurakkal B, Spatz JP, Das T. 2020 Dynamic heterogeneity influences the leader follower dynamics during epithelial wound closure. *Phil. Trans. R. Soc. B* 375, 20190391. (doi:10.1098/ rstb.2019.0391)
- Angelini TE, Hannezo E, Trepat X, Marquez M, Fredberg JJ, Weitz DA. 2011 Glass-like dynamics of collective cell migration. *Proc. Natl Acad. Sci. USA* 108, 4714–4719. (doi:10.1073/pnas.1010059108)
- 88. Bi D, Yang X, Marchetti MC, Manning ML. 2016 Motility-driven glass and jamming transitions in biological tissues. *Phys. Rev. X* **6**, 021011. (doi:10. 1103/PhysRevX.6.021011)

- Westley PAH, Berdahl AM, Torney CJ, Biro D. 2018
 Collective movement in ecology: from emerging
 technologies to conservation and management.
 Phil. Trans. R. Soc. B 373, 20170004. (doi:10.1098/rstb.2017.0004)
- Herbert-Read JE, Perna A, Mann RP, Schaerf TM, Sumpter DJT, Ward AJW. 2011 Inferring the rules of interaction of shoaling fish. *Proc. Natl Acad. Sci. USA* 108, 18 726–18 731. (doi:10.1073/pnas.1109355108)
- 91. Couzin I. 2007 Collective minds. *Nature* **445**, 715. (doi:10.1038/445715a)
- Ding SS, Muhle LS, Brown AEX, Schumacher LJ, Endres RG. 2020 Comparison of solitary and collective foraging strategies of *Caenorhabditis elegans* in patchy food distributions. *Phil. Trans. R. Soc. B* 375, 20190382. (doi:10.1098/rstb.2019.0382)
- 93. Ballerini M *et al.* 2008 Interaction ruling animal collective behavior depends on topological rather than metric distance: evidence from a field study. *Proc. Natl Acad. Sci. USA* **105**, 1232–1237. (doi:10. 1073/pnas.0711437105)
- 94. Hinz RC, de Polavieja GG. 2017 Ontogeny of collective behavior reveals a simple attraction rule. Proc. Natl Acad. Sci. USA 114, 2295—2300. (doi:10. 1073/pnas.1616926114)
- 95. Lei L, Escobedo R, Sire C, Theraulaz G. 2020 Computational and robotic modeling reveal parsimonious combinations of interactions between individuals in schooling fish. *PLoS Comput. Biol.* **16**, e1007194. (doi:10.1371/journal.pcbi.1007194)
- Rahmani P, Peruani F, Romanczuk P. 2020 Flocking in complex environments—attention trade-offs in collective information processing. *PLoS Comput. Biol.* 16, e1007697. (doi:10.1371/journal.pcbi. 1007697)
- Jayles B, Escobedo R, Pasqua R, Zanon C, Blanchet A, Roy M, Tredan G, Theraulaz G, Sire C. 2020 Collective information processing in human phase separation. *Phil. Trans. R. Soc. B* 375, 20190801. (doi:10.1098/rstb.2019.0801)
- Grassé PP. 1959 La reconstruction du nid et les coordinations interindividuelles chez *Bellicositermes* natalensis et Cubitermes sp. La théorie de la stigmergie: essai d'interprétation du comportement des termites constructeurs. *Insectes Soc.* 6, 41–83. (doi:10.1007/BF02223791)
- Khuong A, Gautrais J, Perna A, Sbaï C, Combe M, Kuntz P, Jost C, Theraulaz G. 2016 Stigmergic construction and topochemical information shape ant nest architecture. *Proc. Natl Acad. Sci. USA* 113, 1303–1308. (doi:10.1073/pnas.1509829113)

- Theraulaz G, Bonabeau E. 1999 A brief history of stigmergy. *Artif. Life* 5, 97–116. (doi:10.1162/ 106454699568700)
- 101. Gloag ES, Turnbull L, Javed MA, Wang HB, Gee ML, Wade SA, Whitchurch CB. 2016 Stigmergy coordinates multicellular collective behaviours during *Myxococcus xanthus* surface migration. *Sci. Rep.* 6, 26005. (doi:10.1038/srep26005)
- 102. Deneubourg JL, Goss S, Franks NR, Pasteels JM. 1989
 The blind leading the blind: modeling chemically mediated army ant raid patterns. *J. Insect Behav.* **2**, 719–725. (doi:10.1007/BF01065789)
- Calenbuhr V, Deneubourg JL. 1992 A model for osmotropotactic orientation (I). *J. Theor. Biol.* **158**, 359–393. (doi:10.1016/S0022-5193(05)80738-6)
- Calenbuhr V, Chretien L, Deneubourg JL, Detrain C.
 1992 A model for osmotropotactic orientation (II).
 J. Theor. Biol. 158, 395–407. (doi:10.1016/S0022-5193(05)80739-8)
- 105. Nicolis SC, Deneubourg JL. 1999 Emerging patterns and food recruitment in ants: an analytical study. *J. Theor. Biol.* **198**, 575–592. (doi:10.1006/jtbi.1999. 0934)
- 106. Theraulaz G *et al.* 2002 Spatial patterns in ant colonies. *Proc. Natl Acad. Sci. USA* **99**, 9645–9649. (doi:10.1073/pnas.152302199)
- 107. Detrain C, Deneubourg JL. 2006 Self-organized structures in a superorganism: do ants 'behave' like molecules? *Phys. Life Rev.* **3**, 162–187. (doi:10. 1016/j.plrev.2006.07.001)
- Bak P, Tang C, Wiesenfeld K. 1987 Self-organized criticality: an explanation of the 1/f noise. *Phys. Rev. Lett.* 59, 381–384. (doi:10.1103/PhysRevLett.59.381)
- 109. Muñoz MA. 2018 Colloquium: criticality and dynamical scaling in living systems. *Rev. Mod. Phys.* 90, 031001. (doi:10.1103/RevModPhys.90.031001)
- 110. Beggs JM. 2004 The criticality hypothesis: how local cortical networks might optimize information processing. *Phil. Trans. R. Soc. Lond. A* **366**, 329–343. (doi:10.1098/rsta.2007.2092)
- 111. Mora MT, Bialek W. 2011 Are biological systems poised at criticality? *J. Stat. Phys.* **144**, 268–302. (doi:10.1007/s10955-011-0229-4)
- 112. Bialek W, Cavagna A, Giardina I, Mora T, Pohl O, Silvestri E, Viale M, Walczak AM. 2014 Social interactions dominate speed control in poising natural flocks near criticality. *Proc. Natl Acad. Sci. USA* 111, 7212—7217. (doi:10.1073/pnas.1324045111)
- 113. Ginelli, F, Peruani F, Pillot MH, Chaté H, Theraulaz G, Bon R. 2015 Intermittent collective dynamics emerge from conflicting imperatives in sheep herds.

- *Proc. Natl Acad. Sci. USA* **112**, 12 729–12 734. (doi:10.1073/pnas.1503749112)
- 114. De Palo G, Yi D, Endres RG. 2017 A critical-like collective state leads to long-range cell communication in *Dictyostelium discoideum* aggregation. *PLoS Biol.* 15, e1002602. (doi:10.1371/journal.pbio.1002602)
- 115. Chialvo DR. 2010 Emergent complex neural dynamics. *Nat. Phys.* **6**, 744–750. (doi:10.1038/nphys1803)
- 116. Cavagna A, Cimarelli A, Giardina I, Parisi G, Santagati R, Stefanini F, Viale M. 2010 Scale-free correlations in starling flocks. *Proc. Natl Acad. Sci. USA* 107, 11 865–11 870. (doi:10.1073/pnas.1005766107)
- 117. Attanasi A *et al.* 2014 Finite-size scaling as a way to probe near-criticality in natural swarms. *Phys. Rev. Lett.* **113**, 238102. (doi:10.1103/PhysRevLett.113. 238102)
- 118. Michor F, Iwasa Y, Nowak MA. 2004 Dynamics of cancer progression. *Nat. Rev. Cancer* **4**, 197–205. (doi:10.1038/nrc1295)
- 119. Ilina O et al. In press. Cell-cell adhesion and 3D matrix confinement determine jamming transitions in breast cancer invasion. Nat. Cell Biol. (doi:10.1038/s41556-020-0552-6)
- Dussutour A, Fourcassié V, Helbing D, Deneubourg JL. 2004 Optimal traffic organization in ants under crowded conditions. *Nature* 428, 70–73. (doi:10. 1038/nature02345)
- 121. Gravish N, Gold G, Zangwill A, Goodismanb MAD, Goldman DI. 2015 Glass-like dynamics in confined and congested ant traffic. *Soft Matter* **11**, 6552. (doi:10.1039/C5SM00693G)
- 122. Zuriguel I, Olivares J, Pastor JM, Martin-Gomez C, Ferrer LM, Ramos JJ, Garcimartin A. 2016 Effect of obstacle position in the flow of sheep through a narrow door. *Phys. Rev. E* **94**, 032302. (doi:10.1103/ PhysRevE.94.032302)
- 123. Dieterich P, Klages R, Preuss R, Schwab A. 2008 Anomalous dynamics of cell migration. *Proc. Natl Acad. Sci. USA* **105**, 459–463. (doi:10.1073/pnas. 0707603105)
- 124. Moussaïd M, Garnier S, Theraulaz G, Helbing D. 2009 Collective information processing in swarms, flocks and crowds. *Top. Cogn. Sci.* 1, 469–497. (doi:10.1111/j.1756-8765.2009.01028.x)
- 125. Shiwakoti N, Sarvi M, Rose G, Burd M. 2011 Animal dynamics based approach for modeling pedestrian crowd egress under panic conditions. *Procedia Soc. Behav. Sci.* **17**, 438–461. (doi:10.1016/j.sbspro. 2011.04.526)