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Seasonal variation in telomere dynamics in African striped mice

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ABSTRACT

Telomere shortening has been used as an indicator of aging and is believed to accelerate under harsh environmental conditions. This can be attributed to the fact that telomere shortening has often been regarded as non-reversible and negatively impacting fitness. However, studies of laboratory mice indicate that they may be able to repair telomere loss to recover from environmental harshness, as indicated by recent studies in hibernating rodents. We studied seasonal variation in telomere dynamics in African striped mice (Rhabdomys pumilio) living in a highly seasonal environment. In our annual species, individuals born in the moist spring (high food availability) need to survive the harsh dry summer (low food availability) to be able to reproduce in the following spring. We studied the effect of the harsh dry vs. the benign moist season on telomere dynamics. We also tested whether telomere length and changes in telomere length were associated with the probability that individuals disappeared form the population during the dry season. Male but not female stripped mice showed age-related telomere erosion. Telomeres were longer at the beginning of the dry season compared to the rest of the year. Telomeres increased significantly in length during the moist season. Neither telomere length at the onset of the dry season nor telomere loss over the dry season predicted whether or not individuals disappeared. In conclusion, our data suggest that seasonal attrition and restoring of telomeres also occurs in non-hibernating wild rodents living in hot food restricted environments.

1 INTRODUCTION

2 Telomeres are DNA sequences that cap the end of linear chromosomes and protect genome integrity 3 (Chan and Blackburn 2004). Telomeres erode progressively with cell replication and accumulation of 4 stress-derived damages (Ilmonen et al. 2008; Kotrschal et al. 2007; Olonikov 1973), which is considered 5 as one main indicator of the ageing rate of cells and of organisms. Hence, a straightforward initial 6 telomere-ageing hypothesis was that telomere length is shortened progressively with age, and that 7 faster shortening rate actually reflects senescence, and reduced individual fitness (Dantzer and Fletcher 8 2015; Salomons et al. 2009). Accordingly, both telomere length and telomere shortening have been 9 suggested to predict survival and/or longevity in different taxa (Fairlie et al. 2016; Heidinger et al. 2012; 10 Whittemore et al. 2019).

11

12 A potential causal role of telomeres on life history trajectories was supported by studies showing that 13 telomere length / shortening may reflect life stress effects like the costs of reproduction (Ilmonen et al. 14 2008; Kotrschal et al. 2007), and are related to immune status and response (Criscuolo et al. 2018b). 15 However, contradictory results have been published challenging the idea of continuous telomere 16 shortening with age or its link with higher mortality (Cerchiara et al. 2017; Martin-Ruiz et al. 2005; Seeker 17 et al. 2018). This highlighted a key element for our understanding of telomere role in animal ageing, 18 which is that the telomere – fitness relationships are indeed more complex than initially thought, 19 possibly because telomere dynamics evolved differently under distinct environmental pressures (Tian et 20 al. 2018). Whether telomeres are causal in ageing or rather just a biomarker of somatic maintenance is 21 still an open question (Simons 2015; Young 2018). This seems to be especially true for small mammals in 22 which telomere shortening is often not part of the cell replicative ageing process (Gorbunova et al. 2008; 23 Tian et al. 2018). For example, in laboratory mice, telomeres do not always shorten with cell divisions, 24 (Gomes et al. 2010; Gorbunova et al. 2008). However, other recent studies in laboratory mouse strains

25 (e.g. C57BL/6 mice housed under ad libitum in standard laboratory conditions), showed some shortening 26 in telomere length as mice aged, at rates predictive of individual longevity (Vera et al. 2012). In addition, 27 sex differences exist in telomere length (Barrett and Richardson 2011), resulting in additional complexity 28 related to sex-specific telomere length inheritance or aging. Field studies of telomere dynamics on small 29 mammals to test these contrasting results obtained from controlled laboratory strains are mostly absent. 30 Even in other taxa than rodents, some studies found that telomeres may either be stable (cows: 31 (Seeker et al. 2018)) or even increase over time (great tits: (Atema et al. 2018)). In other mammals, such 32 as wild Soay sheep, telomere lengthening over an individual's lifetime was described (Fairlie et al. 2016) 33 (for a bird example, see also (Spurgin et al. 2018)). Remarkably, a recent study on hibernating rodents 34 have highlighted compensatory telomere lengthening when environmental conditions are becoming 35 favorable after winter (Hoelzl et al. 2016a). Three explanations have been proposed for an increase of 36 telomere length over time. Although (1) measurement errors or statistical adjustment in telomere length 37 assessment need to be checked (Nettle and Bateson 2017; Simons et al. 2014; Verhulst et al. 2013), 38 biological explanations have to be considered (Bateson and Nettle 2016), such as (2) variation in cell 39 population on which telomere lengths are measured, or (3) in activated telomere maintenance processes 40 (e.g. telomerase activity or alternative lengthening pathway) leading to actual elongation of telomere 41 ends (Blackburn 1991; Cesare and Reddel 2010; Criscuolo et al. 2018a), a process which may be 42 energetically costly (Hoelzl et al. 2016a; Nowack et al. 2019). If telomere size elongation in juvenile or 43 adult somatic cells is more common than initially believed (Fairlie et al. 2016; Haussmann et al. 2004; 44 Olsson et al. 2018), this would have important consequences for our understanding of how telomeres 45 interact with fitness-related traits. For instance, it may highlight why long telomeres correlate better 46 with individual fitness than short telomeres (Bauch et al. 2014). Importantly, we should test whether 47 there is further evidence for an increase in telomere length in individuals living under different harsh 48 environmental conditions than studied so far and whether this is related to fitness.

49 In wild-derived house mice strains (Mus musculus) kept under laboratory conditions, individuals 50 showed accelerated telomere loss with immune, reproductive or social stress but non-significant 51 relationships with mortality (Ilmonen et al. 2008; Kotrschal et al. 2007). Thus, one important emerging 52 question is whether the idea that the shortening of telomeres under environmentally harsh conditions is 53 long-lasting (De Punder et al. 2018; Reichert et al. 2014), or whether shortened telomeres can remain 54 stable in size or even increase when environmental conditions become favorable. Whether telomere 55 lengthening is a selected maintenance process improving individual fitness via increased lifespan, for 56 example in hibernating mammals (Hoelzl et al. 2016b; Turbill et al. 2012), remains an unanswered 57 question. To our knowledge, only four published studies described seasonal variation in telomere length 58 (one on small wild mammals) (Fairlie et al. 2016; Hoelzl et al. 2016a; Quirici et al. 2016b; Stier et al. 59 2014). Two studies approached this question indirectly through cross-sectional sampling, assessing 60 differences in telomere length between individuals experiencing different environments during growth 61 or reproduction (Quirici et al. 2016a; Stier et al. 2014). Studies in Soay sheep and edible dormice 62 provided some evidence that changes in telomere length are more related to seasonal variation in 63 environmental conditions (like high population densities and feeding conditions before and/or during the 64 winter) than to age (Fairlie et al. 2016; Hoelzl et al. 2016a).

65 To better understand how telomere dynamics are affected by environmental harshness, 66 longitudinal studies are needed, where individuals are sampled repeatedly under different 67 environmental conditions. Here, we investigated telomere dynamics in African striped mice (Rhabdomys 68 pumilio). Striped mice inhabit a highly seasonal environment with high food abundance in the moist 69 spring being followed by the food restricted harsh dry season in summer. We sampled individuals 70 repeatedly, at the onset of the dry season (January), after they experienced 3 months of harshness in the 71 middle of the dry season (March), 7 months after they experienced harshness at the end of the onset of 72 the moist season (July), and at the end of the favourable moist season with high food abundance

73 (October). We predicted telomere shortening to be faster during the harsh food-restricted dry season
74 than during the favourable moist season, and that telomere length at the onset of the dry season is
75 related to disappearance risk of individuals, which we used as proxy for dry season survival and thus
76 fitness.

78 MATERIAL AND METHODS

79 Study species

80 The striped mouse is an omnivorous annual species: individuals are born in the moist spring (August to 81 December) with peak food availability, then have to survive the following hot summer dry season 82 (January to May) with very low food availability, before reproducing in the following spring; less than 1% 83 of striped mice survive to reproduce in a second spring (Schradin et al. 2012). Depending on the 84 harshness of the dry season, 1% to 50% of the striped mice survive the dry season to reproduce, such 85 that dry season survival is a key determinant of individual fitness. In the breeding season, striped mice 86 can live solitarily but more often live in communally breeding groups with 2-4 breeding females and 1 87 breeding male, and adult offspring remain philopatric, such that all striped mice live in social groups 88 during the dry season (Schradin et al. 2010; Schradin et al. 2019).

89

90 Data collection

91 The study was conducted in Goegap Nature Reserve in South Africa (S 29 41.56, E 18 1.60). Goegap lies 92 within the semi-desert Succulent Karoo biome, which is characterized by cold and moist winters followed 93 by spring with high food abundance and hot dry summers with low food abundance. During the current 94 study, food availability measured as the number of food plant species per 4 m² monitoring plot changed 95 from 2.8 in January to 2.0 in March, 6.4 in August (after the July sampling), and 3.6 in October. Our field 96 site was 7 ha and contained 12 social groups. Because the population was monitored throughout the 97 year, we knew all individuals present on the field site, their life history, and their group affiliation 98 (Schradin et al. 2019). Trapping continued even after our study such that we knew which individuals 99 disappeared during the study and did not re-appear afterwards. During our normal monitoring, every 100 group was trapped for 3 days 2 to 4 times every month (i.e. 6 to 12 days per month).

101 Tail tissue can be used to evaluate somatic telomere dynamics and are representative of telomere 102 dynamics of overall body soma (Daniali et al. 2013). Collecting tail tissue has the advantage of being non-103 lethal and allowing repeated sampling, which is essential for longitudinal studies. We repeatedly 104 collected tissue samples (1-2mm tail snips) from all individuals present on the field site (n=80, age 11 – 105 737 days) during extensive trapping sessions (always for 7 days) in 2018 at: (i) the beginning of January 106 (onset dry season); (ii) March (middle dry season); (iii) End of July, beginning of August (onset of the 107 moist and breeding season); and (iv) October (end of the moist / breeding season). Individuals were born 108 in the moist season (August and December) in 2016 (n=9) or 2017 (n=48), with 14 individuals being born 109 between January and July 2018. Season of birth was unknown for 9 immigrants born in 2016 (n=2), 2017 110 (n=6) and undefined (n=1). Tail clips were stored in 95% ethanol and transported to Strasbourg, France, 111 for analysis.

For the onset of the dry season (January), we had samples from 54 individuals (29 females, 25 males, age 15-501 days). For the onset of the moist season (end of July), we had samples from 26 individuals (16 females, 10 males, age 9-652 days). In addition, intra-individual changes in telomere length was evaluated through repeated tissue sampling of the same individual in (i) January and March for 36 individuals (21 females, 15 males, individual variation in telomere dynamics over the dry season), (ii) July and October for 26 individuals (16 females, 10 males, individual variation in telomere dynamics over the moist season), and (iii) 6 individuals sampled monthly during all four months (Figure 1).

Fieldwork and tissue sampling were carried out under the necessary licenses. All data were collected following protocols approved by the Animal Ethics Screening Committee of the University of the Witwatersrand (AESC 2012/37/2A, AESC 2014/40/B).

122

123 **Telomere analysis**

Striped mice DNA was extracted from tail tissue using Nucleospin Tissue kit (Macherey-Nagel, Düren, Germany). DNA quality and concentration were assessed with a Nano-Drop spectrophotometer (NanoDrop Technologies, Thermo Fisher Scientific) and diluted to the concentration of 2.5 ng/mL. Relative telomere length was measured in striped mice on extracted DNA from tail tissue, using the methodology of real-time qPCR that originated from the work of Cawthon and collaborators on human samples (Cawthon 2002). For our study, the qPCR amplification protocol has been adapted as follows:

130 (i) qPCR gives us as a final results a ratio (T/S) of amplification cycles (Cq) of the telomere sequence (T) 131 and of a genomic gene sequence (S) that must be specific or preferably a non-variable copy number 132 (non-VCN) gene (Smith et al. 2011). Final T/S values were calculated by taking into account each plate 133 efficiencies, following (Pfaffl 2001). For the present study, we designed amplification primers for a region 134 using aligned sequences of PGK1 genes from other closely related organisms extracted from the NCBI 135 data base (Huchon et al. 2017). We used a 150 bp portion of the phosphoglycerate gene (PGK1), which 136 was found non-variable in copy number within our population of striped mice (Cq values ranged 137 between 24-25.6 for 90% of the samples). Primer sequences for the non-VCN gene were: Forward 138 primer: 5'-CGGAGACACCGCCACTTG-3' and Reverse primer: 5'-AAGGCAGGAAAATACTAAACAT-3'. The 139 5'forward and reverse primers the telomere for sequence were: 140 CGGTTTGTTTGGGTTTGGGTTTGGGTTTGGGTTTGGGTT-3' 5'and

141 GGCTTGCCTTACCCTTACCCTTACCCTTACCCTT-3', respectively.

(ii) Amplifications of T and S were done on distinct 96-wells plates due to differences in amplification
temperatures using an automated thermocycler (Master-cycler Realplex; Eppendorf, Hamburg,
Germany). The reactions conditions were set as follows: for the telomere amplification, 95°C for 2 min –
followed by 30 cycles of 95°C for 15 sec, 56°C for 15 sec, 72°C for 30 sec; for the PGK1 amplification, 95°C
for 2 min – followed by 30 cycles of 95°C for 15 sec, 60°C for 15 sec, 72°C for 30 sec. All amplification
runs were ended by a melting curve program (steady increase in amplification temperature from 56°C (T)

or 60°C (S)) to verify the absence of non-specific amplification events (*i.e.* lack of specificity of the primers or primer-dimer artefacts, see below). A negative control (without DNA) was also present on each plate to check for potential contamination.

(iii) Amplifications were performed with 5 ng of DNA (2 uL) per well, in a final volume of 10 uL containing
5 uL of GoTaq qPCR mix (Promega, Madison, WI, USA), 0.5 uL of each primers at an initial concentration
of 5 nM and 2 uL of ultra-pure water.

154 (iv) Samples were amplified in duplicates, on 4 different runs (each comprising 1 telomere plate and 1 155 PGK1 plate). Each 96-wells plate included a dilution curve of the same randomly-chosen reference 156 sample, serially diluted from 10 to 0.625 ng/mL of DNA, to check for consistency in amplification 157 efficiency among plates (r^2 of the dilution curves were comprised between 0.931 – 0.998 (telomere) and 158 0.995 - 0.998 (PGK1)). In addition, 5 samples were repeated over all plates. Mean amplification 159 efficiencies were of 96.0 \pm 0.44 % for PGK1 and 97.8 \pm 0.97 % for telomere. We did not include any plate 160 effect in our models since there was no indications for an inter-plate variation in the T/S ratio values 161 (Linear model, estimates -0.002 ± 0.017 , t = -0.13, P = 0.896).

162

163 qPCR repeatabilities and effect of storage time

164 As suggested by (Nettle et al. 2019), we evaluated the repeatability of our measures using intra-class 165 correlation coefficients (ICC, R package ICC, (Wolak et al. 2012)) both for intra-plate (duplicates) and 166 inter-plates variability in T/S ratio (see also (Eisenberg 2016)). Intra-plate ICC was of 0.864 and inter-plate 167 ICC was of 0.836 (Cicchetti 1994). We also checked whether ICC varied with time of storage, i.e. time of 168 sampling (from January to October): all samples were analysed together in May 2018, such that samples 169 from January had the longest storage time and those samples from October had shortest storage time. 170 Measurement repeatability (intra-individual) did not change with time from January (ICC = 0.851), March 171 (0.835), July (0.769) and October 2018 (0.901). In addition, we found that T/S ratios were correlated

within individuals measured both in January and March (n=35, $t_{1, 34}$ = 2.413, *P* = 0.021), and both in July and October (n=30, $t_{1, 29}$ = 2.198, *P* = 0.036; see Figure1 in ESM 1). Additional qPCR methodological details are presented in ESM1.

175

176 Statistical analysis

177 Data were analysed using R version 3.5.1 (R Foundation for statistical Computing Platform, 2018), using 178 the packages Ime4 (function Imer) (Bates et al. 2014) and Factominer (Lê et al. 2008). We tested for 179 normality with both a Shapiro-Wilk's test and QQ plots distribution (*fitdistrplus* package, (Delignette-180 Muller and Dutang 2015)). We used a Linear Mixed Model (LMM) to analyse the relationship between 181 telomere length (n=154 measures) and the following explanatory variables: Age, sex, and season and all 182 interactions. Season was coded in 4 categories : beginning of the dry season (January), middle of the dry 183 season (March), beginning of the moist season (July) and middle of the moist season (October). So, our 184 statistical analysis considers the 4 sampling points and tested for differences among 4 season categories. 185 Posthoc multiple comparisons among the 4 season categories were conducted using the function *alht* 186 from the *multcomp* package (Hothorn et al. 2008). Individual ID was included as random factor (to 187 control for repeated sampling). Because age, body mass and body size are interdependent, we also used 188 a principal component analysis (PCA) to merge those variables into one PC axis (ESM2). PCA1 turned out 189 to be non-significant in all models, so we kept only Age as a fixed factor in the final analysis. The date of 190 birth was initially used as a fixed factor when testing variance in individual relative telomere length, but 191 it was not significant (LMM, estimates 0.002 ± 0.001 , t = 1.556, P = 0.124) and not considered in our final 192 models. Individual changes in telomere length over the dry and moist seasons were tested using paired t-193 tests. To test the predictive value of relative telomere length on disappearance risk of individual striped 194 mice from the population (Vuarin et al. 2019), we used a linear mixed model with binomial distribution 195 of the survival variable (0 and 1, the latter indicating the presence of the individual). Individuals were 196 regarded as not having disappeared when they were still trapped in July or afterwards (monthly trapping 197 of groups continued for 1.5 years). Of the 54 individuals sampled in January, 19 survived until July and 35 198 disappeared. July was chosen as the censoring date, since this is when breeding dispersal peaks, such 199 that most cases of disappearance that occur before this time can be assumed to be due to mortality 200 (Vuarin et al. 2019). Explanatory variables included in the models were telomere lengths measured in 201 January, March and July (n=52 measures, with Individual ID as random factor). In addition, telomere 202 length rate of change (*i.e.* the within individual differences in telomere lengths measured within the dry 203 season or TROC, n=92 measures) was tested separately. Using the correction for the regression to the 204 mean (RtM) effect ((Verhulst et al. 2013), ESM4) or using, as we present in the main text, uncorrected 205 TROC with the initial telomere length as a covariate (Bateson et al. 2019; Hoelzl et al. 2016b) did not 206 change the output of the models . We choose to present models obtained after model selection using 207 the Akaike Information Criterion corrected for small sample sizes (Mumin R package, (Barton 2020), 208 ESM3).

210 **Results**

211 There was an overall significant variation in relative telomere length over the seasons (Table 1, Figure 2). 212 Posthoc Tukey contrasts indicated that, overall, relative telomere lengths were longest at the start of the 213 dry season in January and shortest in March and July. Relative telomere lengths were significantly longer 214 in October than in July (P = 0.035; while there was no significant difference between March and October, 215 P = 0.994). Individual telomere dynamics (Figure 3) demonstrated that, during the dry season, telomere 216 length of most individuals declined (paired t-test, $t_{35} = 3.486$, P = 0.001, mean difference in relative 217 telomere length = -0.238 T/S ratio) whereas during the moist season, telomere lengths of most 218 individuals increased (paired t-test, t_{31} = 4.373, P < 0.001, mean difference in relative telomere length = 219 0.170 T/S ratio).

The difference in relative telomere length between sexes depended on age (interaction term Age x Sex: P = 0.031, Table 1). Telomeres of males shortened significantly during ageing while female telomeres did not (Figure 4, linear regression, males: df = 62, t = -2.91, P = 0.005; females: df = 88, t = -0.09, P = 0.930).

Individual disappearance risk during the dry season (*i.e.* from January to July) was not significantly predicted by the relative telomere length measured at the beginning of the dry season (n = 52, Table 2A) nor by TROC within the dry season (January to March, Table 2B). TROC corrected for the regression to the mean stayed non-significant for predicting disappearance risk (0.956 ± 0.866, z = 0.110, P = 0.912). None of the other explanatory variables (sex, age, and interaction terms, all P > 0.308) predicted individual disappearance.

231 Discussion

232 Here we showed that (i) the degree of telomere length degradation differed between the harsh dry and 233 the favorable moist season in African striped mice, and (ii) that this effect appeared independent of an 234 age effect which was only evident in males. In fact, on average, telomere length even increased in 235 individuals from July to October (more so in females), demonstrating that the telomere loss during the 236 preceding dry season cannot be simply explained by age (Figure 3). This indicates that environmental 237 harshness influences telomere dynamics: telomeres degrade under harsh conditions but can increase 238 afterwards, at least in females. Thus, our study indicates that a small mammal living under harsh 239 conditions characterized by heat and low food abundance shows similar telomere dynamics as small 240 hibernating mammals after winter but can be subsequently restored, at least partially, during periods of 241 higher food availability (Hoelzl et al. 2016a).

242 Telomere shortening might indicate energetic costs (Criscuolo et al. 2018b; Ilmonen et al. 2008; 243 Nettle et al. 2015; Reichert et al. 2014). In our study, food was two times more abundant during the peak 244 of the moist season than during the dry season, such that the dry season was much harsher than the 245 moist season. Accordingly, telomeres shortened significantly during the dry season. Physiological factors 246 that are suspected to negatively affect telomeres in other species include high metabolic rate via 247 increased oxidative stress ((Reichert and Stier 2017) but see (Boonekamp et al. 2017) and (Turbill et al. 248 2012)) and corticosterone levels ((Haussmann et al. 2012; Tissier et al. 2014) but see (Gil et al. 2019)). 249 Also, in striped mice, the food restricted dry season triggers several physiological mechanisms (Rimbach 250 et al. 2016; Schradin 2008a; Schradin 2008b; Schradin et al. 2014), enabling them to reduce both resting 251 metabolic rate and daily energy expenditure (Rimbach et al 2018a). Metabolic rate and corticosterone 252 levels in striped mice are increased during the moist season (Rimbach et al. 2018a; Rimbach et al. 2018b; 253 Schradin 2008c) which would predict a decrease in telomere length, while in opposite we observed an 254 increase. Instead, the low energy availability in the dry season might lead to a decreased investment in cell maintenance and telomere degradation. In the harsh summer, striped mice might rather invest in mechanisms that improve immediate dry season survival while increased food abundance in the subsequent spring favours energy investment into telomere maintenance.

258 We found age-related telomere erosion in males but not in females. Males also tended to have 259 slightly longer telomeres than females (Table 1, P = 0.076). If males start their yearly cycle with longer 260 telomeres (Figure 3), this may give a mechanistic explanation for faster shortening thereafter, since long 261 telomeres are expected to be more susceptible to damage and less to maintenance than short telomeres 262 (Bauch et al. 2014). A trade-off based effect due to sex-specific investment in reproduction vs. telomere 263 maintenance should be explored in the future, based, for instance, on an earlier reproductive investment 264 in males during the dry season (Schradin and Pillay 2005). Further, females may better protect their 265 telomeres or invest in soma maintenance in order to transfer longer telomeres to the next generation, as 266 suggested for another ageing-related mechanism, oxidative stress, in two distinct mammal species 267 (Mungos mungo and Urocitellus columbianus, (Viblanc et al. 2018; Vitikainen et al. 2016). Future studies 268 should investigate sex specific telomere dynamics depending on sex specific investment into 269 reproduction and whether the shield hypothesis protecting offspring from maternal oxidative stress 270 (Blount et al. 2016) extends to telomere inheritance.

271 Only 37% of the individuals sampled at the onset of the dry season were still present on the field 272 site after the dry season, the rest having disappeared. In our current study, individuals with shorter 273 telomeres were not more likely to disappear during the food restricted dry season. This leads to the 274 hypothesis that telomere length in striped mice is uncoupled from short-term survival because of high 275 extrinsic mortality risks. An alternative may be that, like other small rodents (Seluanov et al. 2007), 276 striped mice maintain a high telomerase activity throughout their lives and consequently telomere 277 length is not a good cell ageing marker. However, in wild mice strains (Mus musculus (Criscuolo et al. 278 2018b; Kotrschal et al. 2007)) and in a laboratory study conducted on Sudanian grass rat (Arvicanthis

ansorgei, (Grosbellet et al. 2015)), the rate of telomere shortening increased with stress caused by circadian desynchronization. Telomere shortening in mice should then be more closely related to survival (Ilmonen et al. 2008), since a faster telomere loss may reflect a concomitant deterioration of the organism's functioning (*i.e.* as an intrinsic ageing process, (Harris et al. 2016; Whittemore et al. 2019)). We did not find such a relationship since individuals with increased telomere shortening during the dry season were not more likely to disappear. In future, we need more data to test whether telomere length is correlated with reproductive success in striped mice.

286 It is currently widely debated whether reported increases in telomere length represents 287 measurement errors or are of biological relevance (Bateson and Nettle 2016; Steenstrup et al. 2013). To 288 date, negative and positive variation in telomere length over life have been found in free-living mammals 289 and birds sampled longitudinally (Fairlie et al. 2016; Spurgin et al. 2018), illustrating the potential 290 biological relevance of telomere lengthening. In striped mice, telomeres even increased in many 291 individuals during the moist season (Figure 3). Although a long-term study with several dry and moist 292 seasons differing in harshness is needed to definitively show an effect of harshness on telomere 293 dynamics, the observed lengthening of telomeres raises questions about both its ultimate function (*i.e.* 294 link with reproductive success) and proximate mechanism (*i.e.* seasonal changes in telomerase activity or 295 changes in the rate of cell renewal). It is important to note that even though we were not able to 296 statistically test for true lengthening (which needs at least 3 repeated samples, (Simons et al. 2014), 297 because we used repeated tail sampling for telomere measurements, attributing telomere lengthening 298 to a change in cell population is less likely than in studies using repeated blood sampling. One ultimate 299 hypothesis may be related to breeding. Since the moist season is also the breeding season for striped 300 mice, the lengthening of somatic telomeres may improve reproductive success either because it is associated with enhanced adult physiological processes or because it ensures better offspring quality. 301 302 Therefore, elongating telomeres in breeding adults may be of key importance for fitness. Such an 303 increase in telomere length was previously found in the edible dormouse (Glis glis), also during the active 304 breeding season (Turbill et al. 2013). Our study sheds lights on the fact that telomere elongation is not 305 specific to hibernating rodents. Itsuggests that telomere dynamics change seasonally in a sex specific 306 way in free-living striped mice, and that, as previously found in hibernating small mammals (Hoelzl et al. 307 2016a; Nowack et al. 2019), telomeres can even lengthen after a harsh season when environmental 308 conditions become favourable. This indicates that telomere dynamics are plastic and future studies need 309 to focus on the ultimate function and proximate mechanisms of increasing telomere length in striped 310 mice specifically and free-living animals generally.

311

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315 assistants. Data are available online at figshare doi: xxxxxx.

317 FIGURES legends

318 Figure1

319 Schematic representation of the repeated sampling design of the study. In total, we collected 154 320 samples from 80 individuals (starting with 54 mice in January and adding 26 mice in July), but not all 321 individuals were sampled repeatedly. 36 individuals were sampled both at the start (January) and the 322 middle (March) of the dry season. 32 individuals were sampled both at the start (July) and the middle 323 (October) of the moist season. Altogether 6 individuals were sampled during all four months.

324 Figure 2

Year-round variation in relative telomere length in adult striped mice (156 samples from 80 individuals). Relative telomere length was repeatedly measured in 36 individuals over part of the dry season (two sampling sessions in January and March) and in 32 individuals over the moist season (July and October). indicates significant levels of posthoc Tukey contrasts: *** < 0.001, ** < 0.01, *< 0.05. Boxplots indicate minimum and maximum, median (black line), first and third quartile and mean (diamonds) values. See text for details.

331

332 Figure 3

Individual variation in relative telomere length in male (left panel) and female (right panel) adult striped mice over the beginning and middle of the dry season (January and February, black circles, 36 individuals) and beginning and middle of the moist season (July and October, grey circles, 26 individuals). Increases in telomere length were found in 24 of 32 individuals in the moist season. Numbers for the dry season was 11 of 36 individuals. Dashed lines indicate the 6 individuals that were sampled 4 times over the year.

339

340 **Figure 4**

Relative telomere length of striped mice in relation to sex over the entire study period. The Sex x Age interaction was significant (P = 0.031, Table 1), indicating that males underwent an age-related telomere erosion (linear regression, P = 0.005) while females did not (linear regression, P = 0.930). See text for details. Confidence intervals 95% are indicated in grey.

Table 1: Linear mixed model explaining the effects of season (coded using the 4 monthly sampling points, January / March for the dry season and July / October for the moist season), age, and sex on individual relative telomere length in wild striped mice. The model encompasses repeated telomere data from individuals sequentially sampled within seasons (individual ID as random factor) but includes all individual measurements (in total 154 telomere measurements among 80 individuals). Seasons are defined as dry (January and March) and moist (July and October, see figure 1). Significant fixed effects (in bold) are considered at P < 0.05 and tendencies (in italics) are considered at P < 0.1, P values being obtained using the anova R function. The model presented was selected based on AICc values. Posthoc comparisons were done using the *multcomp* R package.

Response variable: Relative telomere length	Estimates ± SE	d <i>f</i>	Z value	Р
Random factors Individual ID	0.019 ± 0.014			
Residual variance	0.063 ± 0.251			
Fixed factors				
Intercept	0.935 ± 0.057	128.2	16.275	< 0.001
Sex (M)	0.150 ± 0.084	100.6	1.794	0.076
Age	0.001 ± 0.001	114.4	0.436	0.664
Sex x Age	-0.001 ± 0.001	100.8	-2.192	0.031
Season	-0.199 ± 0.062	104.9	10.723	< 0.001
Tukey Contrast posthoc			Z value	Р
January - March	0.181 ± 0.056		3.263	0.006
January - July	0.334 ± 0.063		5.280	< 0.001
January - October	0.164 ± 0.063		2.593	0.047
March - July	0.152 ± 0.068		2.234	0.113
March - October	-0.018 ± 0.068		-0.259	0.994
July - October	-0.170 ± 0.063		-2.692	0.035

Table 2: Predictors of survival of striped mice until the beginning of the 2018 breeding season (*i.e.* July). A generalized linear model was used, following a binary distribution of individual survival (0 = disappearance) using (A) the individual telomere lengths measured at the start of the dry season (*i.e.* January) and (B) the rate of change in telomere length (TROC) over the dry season (*i.e.* January – March). Results were also not significant when using TROC corrected for the regression to the mean effect (see Results). Estimates ± SE are given for males *vs.* females. Significant fixed effects (in bold) are considered for P < 0.05. The models presented here were selected from AICc values.

A. Response variable: Survival	Estimates ± SE	Z value	Р
Random factors Individual ID	5276 ± 72.63		
Fixed factors			
Intercept	-18.899 ± 6.633	-2.849	0.004
Relative telomere length (T/S ratio)	0.664 ± 6.218	0.107	0.915
Sex (M)	-9.389 ± 23.168	-0.405	0.685
Age	-0.063 ± 2.902	-0.180	0.798
Sex x Age	0.039 ± 0.052	0.749	0.454
Relative telomere length x Sex	6.063 ± 14.284	0.424	0.671
B. Response variable: Survival	Estimates ± SE	Z value	Р
B. Response variable: SurvivalRandom factorsIndividual ID	Estimates ± SE NA	Z value	Р
B. Response variable: SurvivalRandom factorsIndividual IDFixed factors	Estimates ± SE NA	Z value	P
B. Response variable: SurvivalRandom factorsIndividual IDFixed factorsIntercept	Estimates ± SE NA -1.070 ± 1.703	Z value -0.629	P 0.530
B. Response variable: Survival Random factors Individual ID Fixed factors Intercept Telomere rate of change (T/S ratio)	Estimates ± SE NA -1.070 ± 1.703 0.390 ± 1.926	Z value -0.629 0.203	P 0.530 0.839
B. Response variable: SurvivalRandom factorsIndividual IDFixed factorsInterceptTelomere rate of change (T/S ratio)Age	Estimates ± SE NA -1.070 ± 1.703 0.390 ± 1.926 0.002 ± 0.003	Z value -0.629 0.203 0.781	P 0.530 0.839 0.435
B. Response variable: SurvivalRandom factorsIndividual IDFixed factorsInterceptTelomere rate of change (T/S ratio)AgeSex (M vs. F)	Estimates ± SE NA -1.070 ± 1.703 0.390 ± 1.926 0.002 ± 0.003 -2.791 ± 1.395	Z value -0.629 0.203 0.781 -2.001	P 0.530 0.839 0.435 0.045
B. Response variable: SurvivalRandom factorsIndividual IDFixed factorsInterceptTelomere rate of change (T/S ratio)AgeSex (M vs. F)Initial relative telomere length	Estimates ± SE NA -1.070 ± 1.703 0.390 ± 1.926 0.002 ± 0.003 -2.791 ± 1.395 1.144 ± 1.900	Z value -0.629 0.203 0.781 -2.001 0.602	P 0.530 0.839 0.435 0.045 0.547

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370 Literature cited

- 371 Atema E, Mulder E, Van Noordwijk AJ, Verhulst S (2018) Ultra-long telomeres shorten with age
- in nestling great tits but are static in adults and mask attrition of short telomeres.Molecular Ecology Resources 19:648-658
- Barrett ELB, Richardson DS (2011) Sex differences in telomeres and lifespan. Aging Cell 10:913 921. doi: 10.1111/j.1474-9726.2011.00741.x
- 376Barton K (2020) MuMIn: Multi-Model Inference.R package version 1.43.17. doi:377https://CRAN.R-project.org/package=MuMIn
- Bates D, Mächler M, Bolker B, Walker S (2014) Fitting linear mixed-effects models using lme4.
 arXiv preprint arXiv 1808.00864:1406.5823
- Bateson M, Eisenberg DTA, Nettle D (2019) Controlling for baseline telomere length biases
 estimates of the rate of telomere attrition. Royal Society Open Science 6:190937. doi:
 10.1098/rsos.190937 10.6084/m9.figshare.c
- Bateson M, Nettle D (2016) The telomere lengthening conundrum it could be biology. Aging
 Cell 16:312-319
- Bauch C, Becker PH, Verhulst S (2014) Within the genome, long telomeres are more informative
 than short telomeres with respect to fitness components in a long-lived seabird.
 Molecular Ecology 23:300-310. doi: 10.1111/mec.12602
- Blackburn EH (1991) Structure and function of telomeres. Nature 6319:569
- Blount JD, Vitikainen EIK, Stott I, Cant MA (2016) Oxidative shielding and the cost of
 reproduction. Biological Reviews 91:483-497. doi: 10.1111/brv.12179
- Boonekamp JJ, Bauch C, Mulder E, Verhulst S (2017) Does oxidative stress shorten telomeres?
 Biology Letters 13:20170164
- Cawthon RM (2002) Telomere measurement by quantitative PCR. Nucleic Acids Research 30:e47
- Cerchiara JA, Risques RA, Prunkard D, Smith JR, Kane OJ, BOERSMA PD (2017) Telomeres
 shorten and then lengthen before fledging in Magellanic penguins (*Spheniscus magellanicus*). Aging 9:487-493
- Cesare AJ, Reddel RR (2010) Alternative lengthening of telomeres: models, mechanisms and
 implications. Nature Reviews Genetics 11:319-330. doi: 10.1038/nrg2763
- Chan SRWL, Blackburn EH (2004) Telomeres and telomerase. Philosophical Transactions of the
 Royal Society B: Biological Sciences 359:109-122. doi: 10.1098/rstb.2003.1370
- 401 Cicchetti DV (1994) Guidelines, criteria, and rules of thumb for evaluating normed and
 402 standardized assessment instruments in psychology. Psychological Assessment 6:284 403 290. doi: 10.1037/1040-3590.6.4.284
- 404 Criscuolo F, Smith S, Zahn S, Heidinger BJ, Haussmann MF (2018a) Experimental manipulation of
 405 telomere length: does it reveal a corner-stone role for telomerase in the natural
 406 variability of individual fitness? Philosophical Transactions of the Royal Society B:
 407 Biological Sciences 373:20160440. doi: 10.1098/rstb.2016.0440 10.6084/m9
- 408 Criscuolo F, Sorci G, Behaim-Delarbre M, Zahn S, Faivre B, Bertile F (2018b) Age-related 409 response to an acute innate immune challenge in mice: proteomics reveals a telomere

- 410 maintenance-related cost. Proceedings of the Royal Society B: Biological Sciences
 411 285:20181877. doi: 10.1098/rspb.2018.1877
- Daniali L et al. (2013) Telomeres shorten at equivalent rates in somatic tissues of adults. Nature
 Communications 4:1597. doi: 10.1038/ncomms2602
- 414Dantzer B, Fletcher QE (2015) Telomeres shorten more slowly in slow-aging wild animals than in415fast-aging ones. Experimental Gerontology 71:38-47. doi: 10.1016/j.exger.2015.08.012
- De Punder K, Heim C, Przesdzing I, Wadhwa PD, Entringer S (2018) Characterization in humans
 of *in vitro* leucocyte maximal telomerase activity capacity and association with stress.
 Philosophical Transactions of the Royal Society B: Biological Sciences 373. doi:
 10.1098/rstb.2016.0441 10.6084/m9
- Delignette-Muller M-L, Dutang C (2015) Fitdistrplus: An R package for fitting distributions.
 Journal of Statistical Software 64:1-34
- Eisenberg DTA (2016) Telomere length measurement validity: the coefficient of variation is
 invalid and cannot be used to compare quantitative polymerase chain reaction and
 Southern blot telomere length measurement techniques. International Journal of
 Epidemiology:dyw191. doi: 10.1093/ije/dyw191
- 426Fairlie J, Holland R, Pikington JG, Pemberton JM, Harrington L, Nussey DH (2016) Lifelong427leukocyte telomere dynamics and survival in a free-living mammal. Aging Cell 15:140-148
- Gil D, Alfonso-Iñiguez S, Pérez-Rodríguez L, Muriel J, Monclús R (2019) Harsh conditions during
 early development influence telomere length in an altricial passerine: Links with
 oxidative stress and corticosteroids. Journal of Evolutionary Biology 32:111-125. doi:
 10.1111/jeb.13396
- 432 Gomes NMV, Shay JW, Wright WE (2010) Telomere biology in Metazoa. FEBS Letters 584:3741-433 3751. doi: 10.1016/j.febslet.2010.07.031
- Gorbunova V, Bozzella MJ, Seluanov A (2008) Rodents for comparative aging studies: from mice
 to beavers. Age 30:111-119. doi: 10.1007/s11357-008-9053-4
- 436Grosbellet E et al. (2015) Circadian desynchronization triggers premature cellular aging in a437diurnal rodent. The FASEB Journal 29:4794-4803. doi: 10.1096/fj.14-266817
- Harris SE et al. (2016) Longitudinal telomere length shortening and cognitive and physical
 decline in later life: The Lothian Birth Cohorts 1936 and 1921. Mechanisms of Ageing and
 Development 154:43-48. doi: 10.1016/j.mad.2016.02.004
- Haussmann MF, Longenecker AS, Marchetto NM, Juliano SA, Bowden RM (2012) Embryonic
 exposure to corticosterone modifies the juvenile stress response, oxidative stress and
 telomere length. Proceedings of the Royal Society B: Biological Sciences 279:1447-1456.
 doi: 10.1098/rspb.2011.1913
- Haussmann MF, Winkler DW, Huntington CE, Nisbet ICT, Vleck CM (2004) Telomerase
 Expression Is Differentially Regulated in Birds of Differing Life Span. Annals of the New
 York Academy of Sciences 1019:186-190. doi: 10.1196/annals.1297.029
- Heidinger BJ, Blount JD, Boner W, Griffiths K, Metcalfe NB, Monaghan P (2012) Telomere length
 in early life predicts lifespan. Proceedings of the National Academy of Sciences
 109:1742-1748. doi: 10.1073/pnas.1113306109
- Hoelzl F, Cornils JS, Smith S, Moodley Y, Ruf T (2016a) Telomere dynamics in free-living edible
 dormice (Glis glis): the impact of hibernation and food supply. The Journal of
 Experimental Biology 219:2469-2474. doi: 10.1242/jeb.140871

- Hoelzl F, Smith S, Cornils JS, Aydinonat D, Bieber C, Ruf T (2016b) Telomeres are elongated in
 older individuals in a hibernating rodent, the edible dormouse (Glis glis). Scientific
 Reports 6:36856. doi: 10.1038/srep36856
- 457 Hothorn T, Bretz F, Westfall P, Heiberger RM (2008) Multcomp: simultaneous inference for 458 general linear hypotheses. R package version, 1-0.
- Huchon D, Steppan SJ, Schenk JJ (2017) Muroid rodent phylogenetics: 900-species tree reveals
 increasing diversification rates. Plos One 12:e0183070. doi:
 10.1371/journal.pone.0183070
- 462 Ilmonen P, Kotrschal A, Penn DJ (2008) Telomere attrition due to infection. Plos One 3:e2143.
 463 doi: 10.1371/journal.pone.0002143.t001
- 464KotrschalA, Ilmonen P, Penn DJ (2007)Stress impacts telomere dynamics. Biology Letters4653:128-130. doi: 10.1098/rsbl.2006.0594
- Lê S, Josse J, Husson F (2008) FactoMineR: An R Package for Multivariate Analysis. Journal of
 Statistical Software 25:2-18
- Martin-Ruiz CM, Gussekloo J, Van Heemst D, Von Zglinicki T, Westendorp RGJ (2005) Telomere
 length in white blood cells is not associated with morbidity or mortality in the oldest old:
 a population-based study. Aging Cell 4:287-290
- 471 Nettle D, Bateson M (2017) Detecting telomere elongation in longitudinal datasets: analysis of a
 472 proposal by Simons, Stulp and Nakagawa. PeerJ 5:e3265. doi: 10.7717/peerj.3265
- 473 Nettle D, Monaghan P, Gillespie R, Brilot B, Bedford T, Bateson M (2015) An experimental
 474 demonstration that early-life competitive disadvantage accelerates telomere loss.
 475 Proceedings of the Royal Society B: Biological Sciences 282:20141610-20141610. doi:
 476 10.1098/rspb.2014.1610
- 477 Nettle D, Seeker LA, Nussey D, Froy H, Bateson M (2019) Consequences of measurement error
 478 in qPCR telomere data: A simulation study. PloS one 14:e0216118. doi: 10.1101/491944
- Nowack J, Tarmann I, Hoelzl F, Smith S, Giroud S, Ruf T (2019) Always a price to pay: hibernation
 at low temperatures comes with a trade-off between energy savings and telomere
 damage. Biology Letters 15:20190466. doi: 10.1098/rsbl.2019.0466
- 482 Olonikov AM (1973) A theory of marginotomy. J Theor Biol. 41:181-190
- Olsson M, Wapstra E, Friesen C (2018) Ectothermic telomeres: it's time they came in from the
 cold. Philosophical Transactions of the Royal Society B: Biological Sciences
 373:20160449. doi: 10.1098/rstb.2016.0449
- 486 Pfaffl MW (2001) A new mathematical model for relative quantification in real-time RT-PCR.
 487 Nucleic Acids Research 29:2003-2007
- Quirici V, Guerrero CJ, Krause JS, Wingfield JC, Vásquez RA (2016a) The relationship of telomere
 length to baseline corticosterone levels in nestlings of an altricial passerine bird in
 natural populations. Frontiers in Zoology 13. doi: 10.1186/s12983-016-0133-5
- Quirici V, Guerrero CJ, Krause JS, Wingfield JC, Vásquez RA (2016b) The relationship of telomere
 length to baseline corticosterone levels in nestlings of an altricial passerine bird in
 natural populations. Frontiers in Zoology 13:1. doi: 10.1186/s12983-016-0133-5
- 494 Reichert S, Stier A (2017) Does oxidative stress shorten telomeres in vivo? A review. Biology
 495 Letters 13:20170463. doi: 10.1098/rsbl.2017.0463
- 496 Reichert S et al. (2014) Increased brood size leads to persistent eroded telomeres. Frontiers in
 497 Ecology and Evolution 2. doi: 10.3389/fevo.2014.00009

- Rimbach R, Blanc S, Zahariev A, Gatta M, Pillay N, Schradin C (2018a) Seasonal variation in
 energy expenditure in a rodent inhabiting a winter-rainfall desert. Journal of
 Comparative Physiology B. doi: 10.1007/s00360-018-1168-z
- Rimbach R, Jäger J, Pillay N, Schradin C (2018b) Food Availability Is the Main Driver of Seasonal
 Changes in Resting Metabolic Rate in African Striped Mice (Rhabdomys pumilio).
 Physiological and Biochemical Zoology 91:826-833. doi: 10.1086/696828
- Rimbach R, Pillay N, Schradin C (2016) Both thyroid hormone levels and resting metabolic rate
 decrease in African striped mice when food availability decreases. The Journal of
 Experimental Biology. doi: 10.1242/jeb.151449
- Salomons HM, Mulder GA, van de Zande L, Haussmann MF, Linskens MHK, Verhulst S (2009)
 Telomere shortening and survival in free-living corvids. Proceedings of the Royal Society
 B: Biological Sciences 276:3157-3165. doi: 10.1098/rspb.2009.0517
- Schradin C (2008a) Differences in prolactin levels between three alternative male reproductive
 tactics in striped mice (*Rhabdomys pumilio*). Proceedings of the Royal Society of London
 B 275:1047-1052
- 513Schradin C (2008b) Seasonal changes in testosterone and corticosterone levels in four social514categories of a desert dwelling sociable rodent. Horm & Behav 53:573-579
- Schradin C (2008c) Seasonal changes in testosterone and corticosterone levels in four social
 classes of a desert dwelling sociable rodent. Hormones and Behavior 53:573-579. doi:
 10.1016/j.yhbeh.2008.01.003
- Schradin C, König B, Pillay N (2010) Reproductive competition favours solitary living while
 ecological constraints impose group-living in African striped mice. Journal of Animal
 Ecology 79:515-521
- 521 Schradin C et al. (2012) Social flexibility and social evolution in mammals: a case study of the 522 African striped mouse (*Rhabdomys pumilio*). Molecular Ecology 21:541-553. doi: 523 10.1111/j.1365-294X.2011.05256.x
- Schradin C, Pillay N (2005) Demography of the striped mouse (Rhabdomys pumilio) in the
 succulent karoo: a unique population in an extreme environment. Mammalian Biology
 70:84-92
- Schradin C, Pillay N, Bertelsmeier C (2019) Social flexibility and environmental unpredictability in
 African striped mice. Behavioral Ecology and Sociobiology 73:94
- Schradin C, Raynaud J, Arrivé M, Blanc S (2014) Leptin levels in free ranging striped mice
 (Rhabdomys pumilio) increase when food decreases: the ecological leptin hypothesis.
 General and Comparative Endocrinology 206:139-145. doi: 10.1016/j.ygcen.2014.06.024
- 532 Seeker LA et al. (2018) Longitudinal changes in telomere length and associated genetic 533 parameters in dairy cattle analysed using random regression models. Plos One 534 13:e0192864. doi: 10.1371/journal.pone.0192864
- Seluanov A et al. (2007) Telomerase activity coevolves with body mass, not lifespan. Aging Cell6:45-52
- 537 Simons MJP (2015) Questioning causal involvement of telomeres in aging. Ageing Research 538 Reviews. doi: 10.1016/j.arr.2015.08.002
- Simons MJP, Stulp G, Nakagawa S (2014) A statistical approach to distinguish telomere
 elongation from error in longitudinal datasets. Biogerontology 15:99-103. doi:
 10.1007/s10522-013-9471-2

- Smith S, Turbill C, Penn DJ (2011) Chasing telomeres, not red herrings, in evolutionary ecology.
 Heredity 107:372-373. doi: 10.1038/hdy.2011.14
- 544 Spurgin LG et al. (2018) Spatio-temporal variation in lifelong telomere dynamics in a long-term 545 ecological study. Journal of Animal Ecology 87:187-198. doi: 10.1111/1365-2656.12741
- Steenstrup T, Hjelmborg JvB, Kark JD, Christensen K, Aviv A (2013) The telomere lengthening
 conundrum--artifact or biology? Nucleic Acids Research 41:e131-e131. doi:
 10.1093/nar/gkt370
- 549Stier A et al. (2014) Starting with a handicap: phenotypic differences between early- and late-550born king penguin chicks and their survival correlates. Functional Ecology 28:601-611.551doi: 10.1111/1365-2435.12204
- Tian X et al. (2018) Evolution of telomere maintenance and tumour suppressor mechanisms
 across mammals. Philosophical Transactions of the Royal Society B: Biological Sciences
 373:20160443. doi: 10.1098/rstb.2016.0443 10.6084/m9
- 555Tissier ML, Williams TD, Criscuolo F (2014) Maternal Effects Underlie Ageing Costs of Growth in556the Zebra Finch (Taeniopygia guttata). Plos One 9:e97705. doi:55710.1371/journal.pone.0097705
- 558Turbill C, Ruf T, Smith S, Bieber C (2013) Seasonal variation in telomere length of a hibernating559rodent. Biology Letters 9:20121095-20121095. doi: 10.1098/rsbl.2012.1095
- Turbill C, Smith S, Deimel C, Ruf T (2012) Daily torpor is associated with telomere length change
 over winter in Djungarian hamsters. Biology Letters 8:304-307. doi:
 10.1098/rsbl.2011.0758
- Vera E, Bernardes de Jesus B, Foronda M, Flores Juana M, Blasco Maria A (2012) The Rate of
 Increase of Short Telomeres Predicts Longevity in Mammals. Cell Reports 2:732-737. doi:
 10.1016/j.celrep.2012.08.023
- Verhulst S, Aviv A, Benetos A, Berenson GS, Kark JD (2013) Do leukocyte telomere length
 dynamics depend on baseline telomere length? An analysis that corrects for 'regression
 to the mean'. European Journal of Epidemiology 28:859-866. doi: 10.1007/s10654-0139845-4
- 570 Viblanc VA et al. (2018) Maternal oxidative stress and reproduction: Testing the constraint, cost
 571 and shielding hypotheses in a wild mammal. Functional Ecology 32:722-735. doi:
 572 10.1111/1365-2435.13032
- 573 Vitikainen EIK et al. (2016) Evidence of Oxidative Shielding of Offspring in a Wild Mammal.
 574 Frontiers in Ecology and Evolution 4. doi: 10.3389/fevo.2016.00058
- Vuarin P, Pillay N, Schradin C (2019) Elevated basal corticosterone levels increase disappearance
 risk of light but not heavy individuals in a long-term monitored rodent population.
 Hormones and Behavior 113:95-102
- 578 Whittemore K, Vera E, Martínez-Nevado E, Sanpera C, Blasco MA (2019) Telomere shortening
 579 rate predicts species life span. Proceedings of the National Academy of Sciences
 580 116:15122-15127. doi: 10.1073/pnas.1902452116
- Wolak ME, Fairbairn DJ, Paulsen YR (2012) Guidelines for estimating repeatability. Methods in
 Ecology and Evolution 3:129-137. doi: 10.1111/j.2041-210X.2011.00125.x
- Young AJ (2018) The role of telomeres in the mechanisms and evolution of life-history trade-offs
 and ageing. Philosophical Transactions of the Royal Society B: Biological Sciences
 373:20160452. doi: 10.1098/rstb.2016.0452