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Increased internal resistance to CO₂ diffusion impairs Nicotiana sylvestris mitochondrial mutant acclimation to growth light irradiance.

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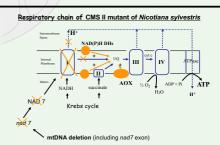
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The CMS II mutant of Nicotiana sylvestris lacking the complex I of the mitochondrial electron transport chain (1) is impaired in mitochondria/chloroplasts crosstalks as highlighted by lower net carbon assimilation rates under ambiant CO2 and O2 conditions and higher dark respiration rates. As mitochondrial function interacts with photosynthetic metabolism, this mutant was used as a tool to investigate the importance of mitochondria in photosynthetic apparatus acclimation to growth light irradiance.

Plants were grown under two irradiances: 80 (LL) and 350 (HL) µmol PAR m-2 s-1. Our first results demonstrated an impairment in net carbon assimilation and initial Rubisco activity in HL-grown CMSII plants as compared to HL-grown WT in spite of equivalent total chloroplastic electron transport and enzyme capacity in both plants (2). The questions raised about the reasons of such impaired acclimation observed in HL-CMSII plants as compared to HL-WT plants and about the fate of the excess electrons, not used for carbon assimilation, in HL-CMSII plants (3).



Mutant leaves are not able to acclimate photosynthesis to higher growth light irradiance (2).

The differences in net carbon assimilation previously observed between WT and CMSII plants depend on growth light irradiance and disappear under low light (80 µmol PAR m⁻² s⁻¹).



Mitochondrial function is involved in net carbon assimilation adjustment to an higher growth light irradiance.

Our first results show that HL-CMSII leaves are not affected in primary photochemistry (ETR) but have lower Rubisco and SPS activities.

Lower A_n but similar ETR

Where go excess electrons in HL-CMSII plants?

The mismatch of ETR, carbon assimilation and initial activity of Rubisco is further investigated in HL-CMSII leaves to characterize alternative electron sinks and identify limiting steps for light acclimation in HL-CMSII plants.

Oxygen as the major sink of chloroplastic electrons in excess in HL-CMSII plants?

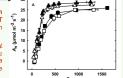
CO2 diffusion limitations : stomatal and/or internal ?

disapear under non-photorespiratory conditions (high light,

high CO2 and low O2)

This suggests higher photorespiration rates in HL-CMSII plants as compared to HL-WT confirmed by higher Glycolate Oxidase activity in HL-CMSII plants.

Both plants show a similar apparent specificity factor of Rubisco, suggesting a lower CO₂ concentration in HL-CMSII chloroplasts, so a limitation at the level of CO₂ diffusion from the atmosphere to the chloroplasts. → See 2

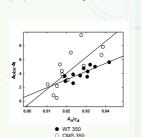


1 Despite a lower stomatal conductance in HL-CMSII plants as compared to HL-WT, for a same external CO₂ concentration, HL-WT and HL-CMSII plants show sir intercellular CO₂ concentration.

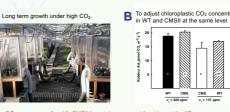
■ III accordance with calculations from models, experimental determinations of chloroplastic CO₂ concentration (Cc) by coupling isotopic discrimination and gas exchange measurements reveal a two times lower g_m in HL-CMSII than in HL-WT. Under the equations are represented potential discriminating steps during CO₂ diffusion from the atmosphere to the chloroplasts and during CO₂ fibation by Rubisco, first, considering Farquhar's model (1962) and second considering experimental determination.

 $\Delta_{(i)} = a \cdot (c_a - c_i) / c_a + b \cdot (c_i / c_a)$ $\Delta_{obs} = a \cdot (c_s - c_i) / c_a + a_i \cdot (c_i - c_c) / c_a + b \cdot c_c / c_a - d$

 $\Delta_{abs} - \Delta_i = (b - a_i) (g_n \cdot A_n / c_a + d$



Experiments performed to confirm the involvement of ${\rm CO_2}$ diffusion limitations in CMSII responses.



lants, in terms of net carbon assimilation, growth rate and Rubisco activation, confirm vement of the CO_2 supply for Rubisco activity in the impaired acclimation of HL-CMSII

These results show that mitochondrial function is necessary for photosynthetic acclimation and CO2 internal diffusion determination to growth light irradiance in Nicotiana sylvestris.

Indeed, CMSII plants do not acclimate to higher growth light irradiance as the WT (2). This is shown by a lower net CO2 assimilation under ambient air conditions and lower activation state of photosynthetic enzymes (2). These differences disappear after an exposure to an atmosphere either enriched in CO2 and/or depleted in O2, or when plants are grown under low-light or high CO2 conditions. Our results show a lower leaf internal conductance for CO2 which limits CO2 supply to the carboxylase activity of Rubisco. This could explain the lower net CO2 assimilation rates, the lower initial Rubisco activity and higher rates of photorespiration in HL-CMSII as compared to WT, photorespiration constituting the major sink of excess chloroplastic electrons non used for net carbon assimilation in HL-CMSII plants.

Lower g_m Increased photorespiration (specificity factor, CO₂/O₂ ratio) Less Rubisco activation Lower CO₂ fixation