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Review Article

The emerging potential of cold atmospheric plasma in skin biology

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ABSTRACT

The maintenance of skin integrity is crucial to ensure the physiological barrier against exogenous compounds, microorganisms and dehydration but also to fulfill social and aesthetic purposes. Besides the development of new actives intended to enter a formulation, innovative technologies based on physical principles have been proposed in the last years. Among them, Cold Atmospheric Plasma (CAP) technology, which already showed interesting results in dermatology, is currently being studied for its potential in skin treatments and cares. CAP bio-medical studies gather several different expertise ranging from physics to biology through chemistry and biochemistry, making this topic hard to pin. In this review we provide a broad survey of the interactions between CAP and skin. In the first section, we tried to give some fundamentals on skin structure and physiology, related to its essential functions, together with the main bases on cold plasma and its physicochemical properties. In the following parts we dissected and analyzed each CAP parameter to highlight the already known and the possible effects they can play on skin. This overview aims to get an idea of the potential of cold atmospheric plasma technology in skin biology for the future developments of dermo-cosmetic treatments, for example in aging prevention.

1. Introduction

The skin is the largest organ of the human body and ensures several distinct functions because of its particular position, in connection between the outside and the inside of the body. This keratinized tegument envelops the whole body and protects it from environmental aggressions and from massive water loss [1]. Although the physical barrier against environmental and pathogen insults is the main role of this organ, the skin possesses several other functions such as vitamin D production [2], humidity, temperature and mechanical sensing [3–5], temperature regulation [6], molecule absorption [7], excretion and secretion [8–11] and some immunological functions [12]. In view of the above, daily cares of this organ are necessary to preserve its integrity and functions. Skin care practices are not only needed for the whole body health but also for social and aesthetic purposes. Cosmetic skin care was practiced from the dawn of time by the ancient civilizations. In the 21st century, with the increase of span life, people from all walks of life ask to live healthy and look younger. Consequently, the global consumer demand of cosmetic products is rapidly expanding today. In 2018, the value of the global cosmetics market was 508 billion U.S. dollars. The market is

projected to value at about 758 billion U.S. dollars by 2025 [13]. Modern skin treatment-offer ranges from chemical product application to physical treatments. Among the treatments, creams, sera and oils are commonly used as at-home beauty treatments while skin peeling treatments are often administered by professional beauticians. Physical treatments are also administered in beauty centers although today some devices can be bought for domestic use. Among them, LED lights and lasers are often used for rejuvenation purposes. These light sources stimulate skin renewal by physically removing the external layers of the skin (resurfacing) thus activating skin cell metabolism [14,15]. For a deeper skin rejuvenation, more invasive and expensive techniques such as aesthetic surgery are required. Currently, a physicochemical approach, based on ionized gases, is joining the skin non-surgical treatments. This technology, named Cold Atmospheric Plasma (CAP), was already used in dermatology to promote wound healing. Today, CAP is entering into the cosmetic field, thus providing a new challenge. In reason of their unique ability to generate a complex chemical mix and thanks to their physical properties, CAPs could be a promising alternative in non-invasive treatment of skin. However, the scientific bases of cold plasma effects on skin and the identification of their exact

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mechanisms of action, both at the cellular and at the molecular levels, are still lacking and they constitute a new active field of investigation.

In the present review, based on skin fundamental notions and on chemo-physical properties of plasmas, we described the possible beneficial interactions between CAPs and skin and how they could participate in improving skin wellness and regeneration.

2. Fundamentals on skin and its microenvironment

To understand how CAP could be used for skin treatments, it is important to take into consideration all skin parameters contributing to skin health that could be potentially modified by plasma treatment.

2.1. Skin structure

The skin covers and isolates the whole body. In humans its thickness can vary from 0.5 to 4 mm. Skin thickness strictly correlates with the collagen content of the dermis and it is gender dependent [16]. The different cell types composing the skin form a complex structure that can be divided in three main layers: the epidermis, the dermis and the hypodermis (Fig. 1). The epidermis is the layer that takes contact directly with the external environment. Its thickness can change at the different body sites and it is also age, gender and phototype dependent [17,18]. The epidermis (Fig. 1A) is mainly made up of keratinocytes and melanocytes, these latter confer the typical color tone of the skin. Some other cells can be found in this part of the skin, the immune system Langerhans cells and the mechanosensors Merkel cells. Human epidermis can be further divided in four or five functional layers. The deeper layer of the epidermis is called *stratum basale* or *germinativum* and is made of non-differentiated keratinocytes, melanocytes and Merkel cells laying on the basal lamina (*lamina basalis*). The presence of keratinocyte stem cells permit the epidermis turnover [19–21]. Once divided, one of the keratinocyte daughter cells moves and differentiates toward the upper layers of the epidermis forming, in sequence, the *strata spinosum*, *granulosum*, *lucidum* (found only in the palms of the hands and soles of the feet) and the *corneum*. Epidermis in healthy and young humans forms digitations that penetrate the dermis (papillary dermis). The epidermis is separated from the dermis thanks to a thin layer of extracellular matrix called basal lamina. This latter is mainly made up of collagen IV,

laminins and glycosaminoglycans (GAGs) and ensures the mechanical link between the basal layer of the epidermis and the underlying connective tissue, the dermis. This middle layer between the epidermis and the hypodermis (Fig. 1B), is made up of a network of fibrillar proteins, collagens, elastin and an amorphous ground substance rich in GAGs such as dermatan sulfate and hyaluronic acid. These negatively charged macromolecules attract water molecules keeping the skin hydrated. Few cell types can be found in the dermis, essentially papillary and reticular fibroblasts with some mast cells. The dermis possesses a sensitivity to mechanic stimuli thanks to the presence of mechanosensors such as the Ruffini's, Meissner's and Pacini corpuscles. The hypodermis is the third deeper layer of the skin. It is an adipose tissue made up of fibroblasts, adipose cells and macrophages. The main function of this tissue is to store fat for thermal insulation and as a source of energy. Besides its three horizontal layers, mammalian skin possesses typical appendages crossing this structure. During the embryogenesis some cells of the ectoderm forming the epidermis invaginate into the dermis creating cutaneous annexes like hair follicles, nails, sweat and sebaceous glands (Fig. 1).

The skin possesses a heterogeneous vascularization (Fig. 1). Vessels coming from the hypodermis enter the dermis and form the deep vascular plexus in the reticular dermis, supplying nutrients and oxygen to hair bulbs and glands. The whole dermis layer is vascularized thanks to the ascending arterioles and descending collecting venules coming from the deeper layers of the skin. In the upper dermis, these vessels form the superficial vascular plexus from which originate the capillary loops extending in dermal papilla [22]. Unlike the other cutaneous layers the epidermis is not vascularized at all.

2.2. Skin oxygenation and antioxidant status

A consequence of the typical skin vascularization is a non-uniform oxygenation. Indeed oxygen diffuses inside the skin from two sources: the atmosphere (21% O₂) and the blood stream circulating in the dermis (5–9% O₂) [23,24]. This results in two oxygen gradients that reach a point of minimum (1–3% O₂) at the basal lamina where keratinocytes stem cells reside (Fig. 1D). Although oxygen has a vital importance for aerobic cells, during the respiratory metabolism a small part of this diatomic molecule is converted in reactive oxygen species (ROS). While

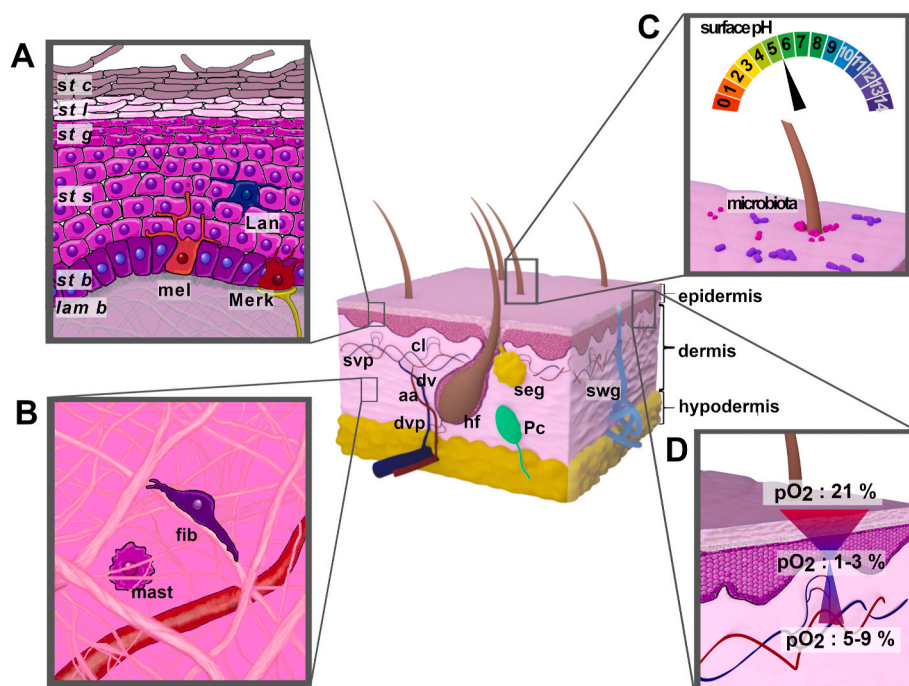


Fig. 1. The skin and its microenvironment. In the middle, the 3D structure of the whole skin with its vascular system: superficial vascular plexus (svp), capillary loops (cl), ascending arteriole (aa), descending collecting venule (dv), deep vascular plexus (dvp). Typical skin appendages: hair and hair follicles (hf) with the associated sebaceous gland (seg) and a sweat gland (swg). Sensory organs such as the Pacini corpuscle (Pc) permit to feel the external stimuli. A) Keratinocytes forming the epidermis layers: *stratum corneum* (st c), *stratum lucidum* (st l) only present in the palms of the hands and soles of the feet, *stratum granulosum* (st g), *stratum spinosum* (st s) and *stratum basale* (st b). The epidermis is separated from the dermis by the *lamina basalis* (lam b). Some other cell types found in this epidermis: melanocytes (mel), Langerhans cells (Lan) and Merkel cells (Merk). B) The vascularized dermis layer made up of extracellular matrix fibers and few cell types such as: fibroblasts (fib) and mast cell (mast). C) The acidic surface of the skin and the associated microbiota living on the *stratum corneum* and inside the hair follicles. D) Skin oxygenation with the two gradients created by oxygen coming on one side from the atmosphere and on the other side from dermis blood vessels.

in low doses ROS play an essential role in cell signaling and in the maintaining of cell homeostasis, in case of overproduction, these free radicals can induce the so-called oxidative stress [25]. Being exposed to different percentage of oxygen, the different skin layers are likely to possess a different sensitivity to oxidative stress. However, to avoid ROS-induced damages, skin cells are able to eliminate the excess of these reactive species keeping their concentration at a physiological level. This task is performed by the antioxidant system that is made up of both small molecules and enzymes such as glutathione peroxidases, catalase and superoxide dismutases allowing the conversion of ROS into harmless molecules [26].

2.3. Skin surface and its physiological pH

A healthy epidermis is relatively acidic. Its outer layer, the *stratum corneum*, possesses an “acidic mantle” that confers it a pH ranging between 4 and 6 (Fig. 1C). Keeping an acidic pH fulfills several physiological roles [27]. At birth, the skin is exposed to a non-sterile environment and it is rapidly colonized by microorganism. The acidic mantle inhibits the growth of pathogen microorganisms and promotes the growth of the physiological microflora [28]. The acidic mantle is generated thanks to endogenous factors but also to exogenous ones such as bacteria. Among the endogenous factors, the relatively-low pH seems to be induced by the generation of urocanic acid [29], and free fatty acids [30]. The maintaining of this acidic mantle seems to be due to the activity of the sodium-proton exchanger (NHE1) [31]. The low pH contributes also to preserve the skin barrier. Indeed, *stratum corneum* enzymes, involved in the synthesis of the ceramide barrier, possess an optimal activity at acidic pH [32]. In addition, a dysregulated acidic mantle can have a negative impact on the skin barrier [33]. Moreover, a high pH of the *stratum corneum* is correlated with skin pathologies such as atopic dermatitis [34,35]. To keep the pH at physiological values, the skin possesses its own buffering system permitting to avoid rapid pH variations induced by external insults [36].

2.4. The skin as a semipermeable barrier of the body

As already described, the *stratum corneum* creates a protective, semipermeable barrier [37–41]. The first representation of the *stratum corneum* was made in 1975 by Michaels and collaborators who simplistically described the outer layer of the skin as a *brick and mortar like structure* [42]. Although the skin isolates and protects the body from external aggressions, this cornified organ is not a sealed barrier. Skin actively communicates and interacts with the external environment. The organ can expel, in a controlled way, water containing organic and inorganic molecules but can also absorb exogenous compounds from the environment. Molecules can penetrate the skin in three ways: through the cells (intracellular way), via the inter-cellular space (intercellular way) or through the skin pores, glands and hair follicles (trans-appendageal way) [43]. In order to understand how molecules can enter the skin, several models have been proposed. In 1992, Auton tried to construct a mathematical model of skin penetration considering either diffusion or metabolism of the compound passing through the skin layers [44]. He considered two main barriers in his skin model 1) the external barrier of the skin, the *stratum corneum* - this physical barrier lets lipophilic compound to pass more easily than hydrophilic molecules -, 2) the second barrier made up of living cells in the epidermis and the dermis where exogenous molecules are subjected to enzymes such as esterase and oxygenase. In 2001 a more complex model, called “single gel phase” was proposed by Norlén and colleagues to describe the *stratum corneum* of the skin [45]. This model is in accordance with the Michaels’ “brick and mortar” one. Other approaches, focused on how different chemical compounds can penetrate the *stratum corneum*, were developed by Trommer and Neubert [46]. Mathematical models of skin permeability have been summarized in a complete review by Mitragotri and collaborators [47].

2.5. The skin microbiota

Thanks to the previously described structure, the skin possesses a unique and complex microenvironment. Because of the low water content of the *stratum corneum*, the low physiological pH, lysozyme and RNase production, skin is a poor substrate for microbial colonization. However, even in healthy conditions, the skin is the host of commensal or symbiotic bacteria [48] at the epidermal but also at the dermal level [49]. After birth, the skin is directly exposed to environmental microorganisms and rapidly colonized by some of them leading to a balance that is unique to each individual. In adults, considering the skin appendages such as follicles and sebaceous glands, the total exposed surface is estimated to be 30 m². This makes our external organ the largest epithelial surface for interacting with microorganisms [50]. Different types of microorganisms can be found in different sites of the skin, depending on humidity, sweat production and local temperature. Commensal bacteria not only protect skin from pathogen bacteria colonization, avoiding thus skin diseases, but play also a symbiotic role with this organ [51,52] that is currently under investigation [53,54].

2.6. Skin homeostasis and aging

The whole skin wellness resides on the physiological equilibrium of all the mentioned biochemical, bio-physical and symbiotic parameters. Although the skin is a powerful organ able to rapidly regenerate, the continuous biological, chemical and physical aggressions weaken its structure over time.

Skin aging is a natural process driven by our genes (chronological aging), that can be drastically accelerated by our way of life, sun and pollutant exposures [55] more largely named exposome [56]. This aging process is mainly mediated by the oxidative stress, an overproduction of ROS able to damage biological molecules such as lipids, proteins and nucleic acids and inducing the alteration of their activity. Typical signs of time in skin are the loss of elasticity and hydration, the slowing down of cell metabolism and the alteration of melanocyte activity. All these changes lead to a decrease of the complexion radiance and the occurrence of light or dark spots and the appearance of wrinkles.

Since the dawn of humanity, people tried to find methods to hide the signs of senescence. Cosmetic treatments have always been the answer to slow down skin deterioration process and ameliorate its aesthetic appearance. To achieve such goal, besides cosmetic products, new physical technologies such as LED- or ultrasounds-based devices have recently been developed. Among these, cold atmospheric plasma may be an innovative and interesting approach in this field. To know CAPs characteristics and how they can interact with the skin, the following chapter will provide some fundamentals about these ionized gases.

3. Plasma bases and biomedical applications

Understanding the physicochemical aspect of plasmas, and the mechanisms involved in the interactions between them and biological tissues, is primordial in order to use them on skin in efficient and secure conditions.

3.1. Introduction to plasmas

The stars, the sun, the lightning and the aurorae have always fascinated human being. All of these wonders of the nature share a common feature, they are forms of plasma: the fourth state of the matter. From the first attempt to artificially generate and master these ionized gases, plasmas have been exploited in different fields. The first artificial plasma was generated by the German physicist Johann Heinrich Wilhelm Geissler who created the ancestor of the fluorescent tube in 1857. Based on the Geissler tube, William Crookes built the first cathode tube in 1879. In his high vacuum tube, Crookes observed that the rarefied gas, exposed to the high voltage between two electrodes, emitted a weak

light at the cathode. Since he did not know that the light was due to the electrons flowing from the cathode, he called them “cathode rays”. Crookes understood that the rarefied gas inside the tube, exposed to the high voltage, was in a particular state that he described as a fourth state of the matter or “radiant matter”. Gases possess almost no interatomic or intermolecular forces. Since they are formed by neutral atoms or molecules, gases are perfect electric insulators. In 1889, Friedrich Paschen studied the loss of the insulating property of the gases exposed to a high electric field between two planar electrodes. Paschen called “breakdown voltage” the minimum voltage needed to generate a discharge in the gas. This voltage is depending on the gas pressure and the distance between the two electrodes [57]. Arc generation is due to the ionization of the gas with consequently a loss of the insulating property. It’s only in 1928 that the chemist Irving Langmuir described the state of a rarefied gas exposed to a high voltage between two electrodes. In his paper, Langmuir wrote that *except near the electrodes, the ionized gas was made up of an equal mix of ions and electrons* and proposed to use the term of plasma for this state of the matter [58].

3.2. Hot and cold plasmas

To generate a plasma, a gas must be supplied with enough energy to ionize it. The gas could be heated at very high temperatures in order to permit the external electrons to escape the atoms. The electron and the ionized atoms can interact with other atoms creating a cascade reaction. Thus the generated plasma is a mix of electrons, ions and neutral species. Although it is formed by charged species, the plasma is neutral on its whole. As described previously, a gas can be ionized and form plasma also under a strong electric field generated between two electrodes. Plasmas can be divided in two main categories: thermal and non-thermal plasmas. Most of the natural occurring plasmas belong to the thermal category. These hot plasmas possess electrons and heavy particles at the same temperature. Artificial hot plasmas are produced for some applications such as hard material cutting and toxic waste destruction [59, 60]. In non-thermal plasmas (NTPs), free electrons possess a higher temperature than ions and neutral gas molecules, these latter conferring a relatively low temperature to the whole plasma. Thanks to this characteristic, NTPs are used in a plethora of industrial applications: micro-electronics, lighting, surface treatment and functionalization. Most NTPs conceived for industrial purposes are produced using low pressure gases inside reactors. The controlled atmosphere inside the reactors results in a well-controlled plasma generation. NTPs, can also be generated at atmospheric pressure and are called Cold Atmospheric Plasmas (CAPs).

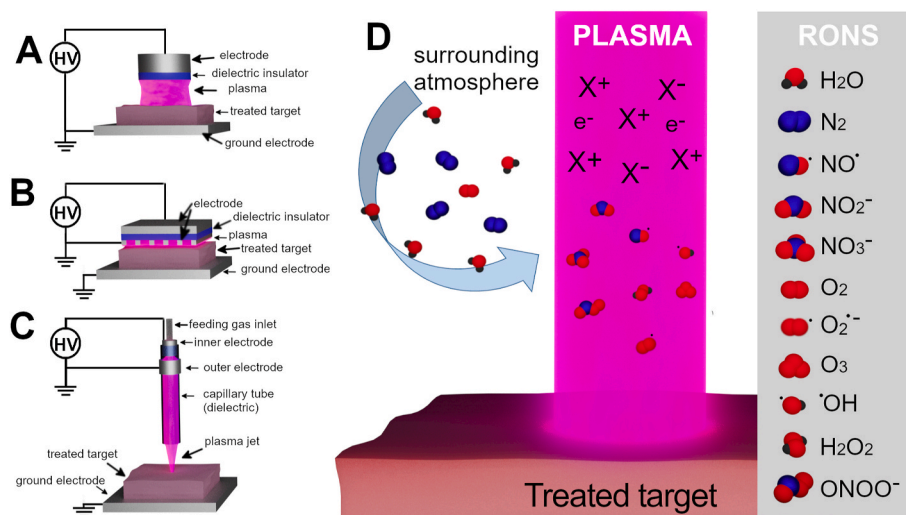


Fig. 2. Cold atmospheric plasma devices and plasma chemistry. Most commonly used CAP-setup for bio-medical applications: A) Volume dielectric barrier discharge reactor; B) Surface dielectric barrier discharge reactor; C) Plasma jet reactor. (HV) High Voltage generator. D) The plasma chemistry: the ionized gas forming a plasma, with its negative and positive ions (X^+ , X^-) and free electrons (e^-), generates a mix of reactive oxygen and nitrogen species (RONS) from the atmospheric gases and water vapor.

3.3. Cold atmospheric plasmas generation and characterization

CAPs are mainly generated by means of energy coming from electric alternate or direct currents, radiofrequencies or microwaves. Several methods and configurations are today used to produce these ionized gases: volume and surface Dielectric Barrier Discharge (DBD) [61,62], Atmospheric Pressure Plasma Jets [63], plasma needles and Plasma Pencils [64] (Fig. 2A, B, C). These different configurations, together with the choice of the feeding gas, result in a plasma having peculiar characteristics. Commonly used gases are atmospheric air, pure nitrogen, noble gases or custom gas mixtures of these latter. Once the gas is ionized to plasma, the charged species can react with the matter (target) with which they enter in contact. In the case of CAPs the first encountered medium is the atmospheric air. Reacting with the nitrogen and oxygen composing the air, CAPs produce the so-called Reactive Oxygen and Nitrogen Species (RONS) (Fig. 2 D). Plasma treatment can be performed on several types of targets, either solids or liquids. When CAPs interact with a solid surface, the generated reactive species can modify, charge or ablate the external layers of the target. These modifications can change the physical characteristics of the surface modifying for example its wettability [65].

Aqueous media exposed to a CAP are enriched of new chemical species. Once generated in the surrounding atmosphere, RONS can be carried and solubilized in the liquid medium [66–68]. Some species such as hydrogen peroxide (H_2O_2) are produced directly from water vapor entering in contact with the plasma effluents [69]. Oxygen already dissolved in the liquid medium also participate to RONS generation [70, 71]. The amount of generated and solubilized species is proportional to the plasma working frequencies [70,72], the voltage waveform, the CAP exposure time [72,73] and the distance from the treated target. Gases passing to the state of plasma emit electromagnetic radiations in the visible and infra-red spectrum. Low amount of UV are commonly generated together with other visible radiations whose wavelength depends on the feeding gas used to produce the plasma. Since CAPs are generally ignited by a strong electric field, once the plasma is generated, the movement of the charged species produces itself an electric field. Lastly, although these plasmas are named cold, they generate thermal energy and develop temperatures between 30° and 100 °C. While temperatures higher than 40 °C are not suitable for the treatment of mammalian tissues, tuning the gas flow rate and the distance of the CAP source from the target permits to avoid overheating effects. The complex interplays between CAP and the treated target are summarized on Fig. 3. In view of the above mentioned characteristics CAPs can be compatible with applications on biological tissues.

Taking into account all the above-mentioned parameters and

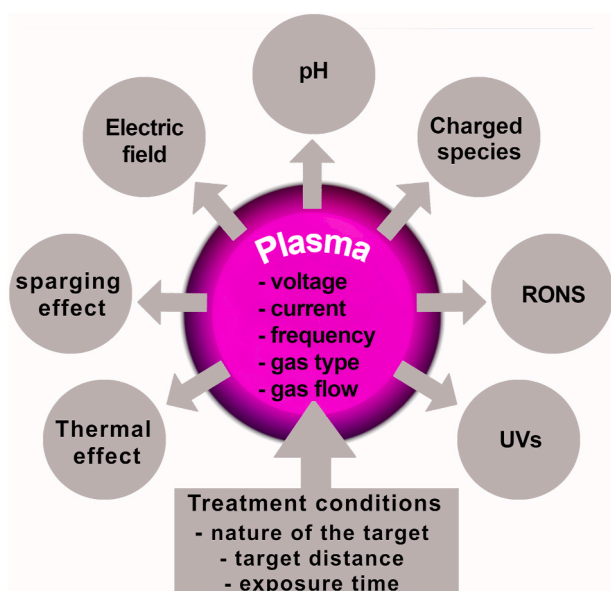


Fig. 3. The interplays between CAP and the treated target. CAP characteristics intrinsically depend on the electrical setup, the feeding gas and its flow rate. The fine tuning of these parameters influences the amount of the produced charged species and RONS, the CAP-generated electric field, the acidification and de-gasification (sparging) of the target. Although marginal, the emitted UVs and the small thermal increase can play a role in the administered treatment. In addition, the treatment conditions such as the nature of the target, the plasma-target distance and the exposure duration also strongly influence the CAP behavior and performances.

knowing that each research laboratory developed its own CAP device, the plethora of data obtained with these sources are not easy to compare. Moreover, depending on the selected technical parameters, such as voltage, frequency, carrier gas composition and gas flow, as well as at different temperatures and humidity, a single individual device also produces very different plasmas inducing different effects. Conscious of this heterogeneity, in the following paragraphs we will make use of the generic term of CAP to indicate different devices and settings employed to generate a Cold Atmospheric Plasma.

3.4. Biomedical applications of plasmas

Plasma application in the bio-medical field is called plasma medicine [74]. In the last decade this field has rapidly expanded. CAPs have shown promising results in cancer therapy either *in vitro* or in clinical case studies [75–80]. Being more sensitive to RONS than normal cells, cancer cells can be selectively killed by plasma exposure [81,82]. Non-thermal plasmas for biomedical applications can be used in indirect or direct configuration. In indirect configuration, the long lived reactive species generated between the electrodes are carried to the target thanks to the gas flow. In direct configuration, the biological target is one of the two electrodes and actively participates to plasma generation [83,84]. Although CAP effluents can now be brought inside the body by means of catheters for *in situ* treatments [85,86], at the beginning plasma medicine was conceived for surface, non-invasive applications. Most of the *in vivo* studies are performed by treating directly the external tissues of the body. Besides cancer applications, CAPs are also used in dermatology with promising effects on chronic wound disinfection and healing [87–94]. Lately some CAP devices have been developed for skin regeneration [95,96]. These plasma sources, operating with atmospheric air, nitrogen or argon, can reach temperatures higher than 60 °C and are mostly used in skin resurfacing. Like laser resurfacing, this type of cosmetic plasma treatment is mostly used to burn the outer layers of the skin and force its renewal [97–100]. Although plasma resurfacing is

a less invasive alternative to facial plastic surgery, the high temperatures, the long recovery after treatment and the limited trial data on long term side effects suggest a careful use of these treatments [101]. Whereas the above mentioned plasma treatments work by a mechanic ablation of the dead outer skin layers, some recent evidences suggest that mild CAP treatments may actually stimulate the deeper layer of the skin and play an anti-age role on skin cells [102]. Hence, plasma applications in dermatology and in cosmetics are, today, hot topics.

4. Cold atmospheric plasma activity on skin

As previously mentioned, CAP features are under the dependency of several parameters of the device such as the voltage and the pulse frequency. The characteristics of the generated plasma are modified by the environment and the gas composition, the nature of the target, the distance from the target and the treatment time (Fig. 3). This results in differences in terms of RONS nature and amount, in the generated electric field and so in a different effect on the treated biological tissue. This is why it is important to understand how plasma chemistry and physics can interact with the skin and how the different parameters can be adapted to achieve the desired effect.

4.1. Effects of CAP-generated RONS on biological components

4.1.1. On biomolecules

Liquid physiological media treated by plasma are enriched in RONS species and have been shown to carry out interesting biological activities on cells and tissues [103,104].

Among the CAP-produced RONS, nitrites (NO_2^-), nitrates (NO_3^-), nitric oxide (NO^\bullet), hydroxyl radical ($^\bullet\text{OH}$), superoxide anion ($\text{O}_2^{\bullet-}$), singlet oxygen ($^1\text{O}_2$), hydrogen peroxide (H_2O_2) and ozone (O_3) are of great interest in biology [105,106].

Biomolecules, in contact with the plasma-generated RONS, can be chemically changed, inactivated or irreversibly damaged [107,108]. Takai and collaborators analyzed the chemical structure of water-dissolved amino acids after CAP exposure and demonstrated that most of the treated amino-acids can be oxidized and acquire new chemical groups by hydroxylation or nitration [109]. The amino acid cysteine, involved in the redox activities of enzymes and abundant in the keratin, has been shown to be oxidized by CAP exposure [110]. Since amino acid can be modified by cold plasma treatment, the whole proteins exposed to CAP can be altered either at their primary amino-acid sequence or at their secondary and tertiary structure. These modifications can change the activity or induce a loss of function of the protein [111–114].

Lipids have also been shown to be oxidized by CAP exposure [115–117]. Phospholipids composing cell membranes can directly be peroxidized by a non-thermal plasma treatment [118]. This oxidation can temporarily change the permeability of the cell membrane facilitating the penetration of exogenous molecules [119]. Lipid oxidation is a phenomenon that naturally occurs during oxidative stress and chronic inflammation [120–122].

CAP-induced modifications and damages were observed equally in nucleic acids [123,124] although strong DNA damages have been observed after long plasma exposure [125]. Taken together, these CAP-induced effects on biological molecules underpin the toxic impact that plasma can have on living matter. Although cold plasmas are a source of potentially toxic reactive species, CAPs delivery can induce a hormesis effect [126] since a proper use of these ionized-gas source, with adapted and controlled conditions, can bring beneficial effects on the treated biological tissue.

4.1.2. On cell metabolism

As already mentioned, a proper CAP treatment can, for example, accelerate wound healing or selectively kill cancer cells with little or no impact on normal cells [81,82,127,128]. CAPs have been largely used to

treat easily accessible tissues and organs and therefore they are suitable to treat the skin, the external organ of our body. Being exposed to the external environment, the skin is continuously stressed by physical and chemical insults. Every day the epidermis is exposed to UV radiations, ozone, cigarette smoke, pollutants, organic solvents or alcohols that directly generate or induce the formation of large amount of ROS [129]. Beside these exogenous sources, cellular metabolism also generates in a lesser extend endogenous free radicals [130]. Human and animal tissues produce reactive oxygen species such as O_2^- , 1O_2 , H_2O_2 and O_3 [131]. These ROS are mainly produced in mitochondria during cell respiration [132]. Although these highly reactive species can be harmful, the cells are able to neutralize free radicals in excess using their anti-oxidant system. Moreover our body can adapt to chronic exposure of toxic compounds. Human adaptation to lethal poisons by ingestion of sub-lethal doses was known thousands of years ago thanks to Mithridates and his Antidotum Mithridaticum [133]. In mitochondria the adaptation to chronic exposure of small doses of ROS is a phenomenon called mitohormesis [134,135].

4.1.3. On skin cellular signaling

Besides their damaging role, well known as oxidative stress when in large amount, ROS are required molecules for cell viability and activity. In fact, right amounts of ROS play some physiological effects as they can act as second messengers [136–138], stimulators for stem cells proliferation [138,139], and as a booster for the immune system [140,141]. In injured skin, the immune system cells entering in contact with pathogens, start to produce large amounts of RONS. This phenomenon called “respiratory burst” helps wound disinfection. Correct wound healing depends on the equilibrium between ROS generation and the activity of the antioxidant system [142]. Among the reactive nitrogen species (RNS), the small gaseous molecule NO^* possesses several biological activities [143] acting as a signaling molecule [144], a vasodilator [145], an angiogenesis modulator [146], an immune system stimulator [147] and a melanogenesis enhancer [148]. In skin NO^* is also involved in hair growth, in the proliferation and differentiation of epidermal cells and in wound healing [149,150]. Nitrates present in sweat can be converted in nitrites and then in NO^* thanks to the skin bacterial microflora [28] or directly from the photo-decomposition of nitrites [151,152]. NO^* derived from photo-decomposition of nitrites has been shown to protect human skin cells from lipid peroxidation and thus apoptosis induced by UVA exposure [152,153]. However an over production or a high exposure to these reactive species, can be harmful [154,155]. Indeed high doses of ROS can contribute to the development of several pathologies such as psoriasis [156–158]. Moreover the oxidative stress in skin can accelerate the natural process of ageing [159,160]. As cold plasmas generate a heterogeneous mix of RONS and considering the “Janus effect” of most of the above mentioned reactive species, the final effect on the treated tissues will depend on the amount of the released reactive species. CAP treatment has been shown to induce an oxidative stress in human keratinocytes [161] but the same cell line exposed for short periods to a helium-fed CAP with a low flow rate increases its viability [126]. Moreover, Schmidt and collaborators observed a hormesis-like increase of the antioxidant system in human keratinocytes cells exposed to an argon fed CAP [162]. In light of the biphasic effect of CAP treatment on skin, mastering the delivery of CAP-produced RONS is a challenge in dermatology and in cosmetics where CAP could be a powerful tool.

4.2. CAP influence on skin proliferation and motility

Cutaneous cell proliferation is important for epidermis turnover and to ensure the healing process in case of skin injuries. When correctly administered, CAP treatment can stimulate these processes. Short exposures to a helium-fed CAP were shown to stimulate HaCaT human keratinocytes proliferation and motility [126,163]. A beneficial effect of plasma treatment on HaCaT cells was also observed with the use of

argon-DBD. Choi et al. showed that CAP treatment inhibits the E-cadherin-mediated intercellular junctions and activates a β -catenin-mediated proliferative signal. This plasma-activated proliferative pathway accelerates *in-vivo* re-epithelialization in mice wound [164]. Furthermore, short treatments (1–3 min) with argon-based plasma jet were shown to increase the proliferation of the *stratum basale* keratinocytes in human, intact skin explants [165]. On dermal fibroblasts very short treatments were shown to increase the proliferation [163,166] while treatments of few minutes exert a toxic effect [166,167]. The higher sensitivity of fibroblasts to CAP treatment, when compared to keratinocytes, can be explained by the fact that these cells reside in a deep layer of the skin. Being protected in the deep dermis, fibroblasts are less equipped to endure an external oxidative stress. *In vivo*, skin fibroblasts can be directly exposed to CAP only in case of wounds. Moreover, in wound treatments, the CAP anti-proliferative effect can be advantageous to avoid the anti-aesthetic side effects of the healing process such as excessive scarring [168].

4.3. CAP influence on skin oxygenation

CAPs have been shown to influence the oxygen content in the treated target. In plasma jets, the gas flow rate exerts the so-called “sparging effect”, de-oxygenating the treated liquid [70,169] while *in vivo* CAP applied on mouse skin has been shown to enhance underneath tissues oxygenation [170]. An increase of the post capillary oxygen saturation after plasma treatment has also been observed in human skin [84,171]. The mechanism on how the plasma source increases *in vivo* skin oxygenation is still unclear. As previously mentioned CAPs are not really cold sources and the devices used in biology develop temperatures between 30° and 40 °C. The application of this relatively warm sources on the skin could induce a local vasodilation and so the increase of tissue oxygenation. Furthermore, since CAP directly or indirectly induces NO^* formation, this vasodilator molecule could be also responsible of the observed phenomenon. A correct oxygenation is fundamental for a proper cellular metabolism. Some skin cosmetic therapies are based on the increase of tissue oxygenation. Hyperbaric Oxygen Therapy, already used for decompression sickness, is currently used in skin-rejuvenation. The administration of pressurized oxygen seems to protect the skin from UVB-induced photoaging [172]. However skin oxygenation needs to be carefully controlled. Oxygen has a double face and its excess can generate an oxidative stress accelerating the ageing process. Moreover skin stem cells need a very low oxygen partial pressure to keep their stemness [23] and thus ensure the renewal of the epidermis.

4.4. CAP effect on skin vascularization and extracellular matrix

Skin oxygenation is directly related to the vascularization of the organ. A healthy skin possesses a well-organized vasculature. With the age, skin microcirculation reactivity such as vasodilation and vasoconstriction as well as vascular density are impaired [173]. The inability to handle ROS generation and the endothelial oxidative stress seem to be one of the reason of the loss of this age-related vessels functionality [174]. Although ROS are down regulators of vascular endothelial functions, CAP treatments, in right conditions, seem to play a stimulating action. Human Umbilical Vein Endothelial Cells (HUVEC) were shown to release pro-angiogenic factors and increase *in vitro* angiogenesis when exposed to an argon-fed CAP for 30s [175]. An increase of proliferation was also shown in porcine endothelial cells where a 30s treatment with an air-DBD induced the release of the fibroblast growth factor-2 (FGF2) [176]. The CAP-angiogenesis stimulatory effect was recently showed by Dzimitrowicz et al. who used a He-fed CAP on human endothelial cells for very short treatments (10s) [163]. *In vivo*, since endothelial cells are not directly affected by plasma treatment, the increase of angiogenesis could be induced via a paracrine mechanism mediated by keratinocytes exposed to plasma treatment [175,177]. The small molecule NO^* also plays a role in angiogenesis. NO^* can act as a

pro- or anti-angiogenic factor depending on its amount [146,178,179]. Moreover NO[•] can increase the synthesis of collagen IV and activate endothelial cells adhesion [180]. Duchesne et al. demonstrated that CAP treatment stimulates endogenous NO[•] synthesis in *in vitro* and *in vivo* models. Indeed, in a murine burn wound model, plasma-generated RONS was shown to increase the expression of the endothelial nitric oxide synthase (eNOS). The increase of the enzyme producing endogenous NO[•], together with proangiogenic factors, speeds up the healing process [181]. In wounds, the extracellular matrix (ECM) of the dermis is directly exposed to CAP treatment and the improvement of the healing process could depend on a direct modification of the ECM properties. Ring and collaborators showed that plasma-treated collagen-elastin matrix scaffolds implanted under mice skin induce an enhancement of neovascularization [182]. In addition, a well-organized ECM is also essential to keep a youthful looking skin. Murine fibroblasts directly exposed for 15s to an argon-CAP were shown to increase the expression of collagen I and III while slightly longer treatments exert an opposite effect [166]. Unlike wounds, in intact skin CAP cannot directly influence the extracellular matrix metabolism. However some *in vivo* studies suggested that CAP can indirectly induce dermal remodeling. An increase of dermal collagen content was observed in intact mouse skin exposed to an argon-DBD plasma [183]. Type I collagen is the major component of the dermis. While the neo-synthesis and a well-organized network of the fibrillary protein is a sign of a young and healthy skin, its overproduction is sometimes the expression of skin damages and diseases. Interestingly CAP is able to stimulate physiological collagenogenesis while inhibiting the pathological synthesis of the protein. *In vitro* and *in vivo* studies have shown this selective effect in dermo-aesthetic disorders such as scars and keloids [184,185].

5. Cold atmospheric plasma effect on skin surface

5.1. CAP-induced pH modification

In addition to their stimulating/oxidizing effects, CAP-generated RONS were shown to induce an acidification of the treated target. A decrease of the initial pH is commonly observed in non-buffered or weakly buffered treated liquids [186,187] and in hydrated 3D matrices [188]. The pH-drop can be mainly attributed to acidic species originating from the precursor NO[•] that generates nitric (HNO₃) and nitrous (HNO₂) acids in solution [187]. The induced acidification is proportional to the plasma exposure time. It has been shown that the pH of alkaline or neutral non-buffered media drops rapidly after few minutes of CAP treatment. The quick pH-decrease tends to stabilize at pH values between 3.5 and 2.5 thanks the transient formation of HONO/ONO⁻ buffer and to the generation of nitrous acid (pKa 3.3) [187–189]. In light of the above, CAP exposure can lower the pH of biological tissues.

Pork skin sebum and human lipids can be rapidly acidified when exposed to an air-fed CAP [190]. The CAP-induced acidification was also confirmed by clinical trials on intact human skin [84,150]. Thanks to these acidifying properties, CAP treatment can contribute to keep the skin in healthy conditions. Indeed, by lowering the pH, cold plasmas can stimulate and accelerate skin renewing. In acute skin wound, a physiological acidification was shown to enhance the proteases activity and stimulate fibroblast proliferation [191]. While skin pH higher than the physiological values can lead to pathologies, very acidic pH can burn the external tissues of the organ. In order to avoid chemical burns, skin exposure to CAP should be carefully controlled [150]. However, in cosmetic, chemical peel or chemexfoliation is used to gently ablate the external layer of the epidermis and force skin renewal. This approach uses often organic acids to lower the pH and peel off the epidermis layers [192]. A well administered plasma treatment could exert a similar noninvasive exfoliating effect. Moreover, as the pH of the acidic mantle increases with the aging process leading to a weakening of this barrier [193], CAP treatments could be beneficial to re-establish the physiological pH barrier and stimulate the regeneration in mature skin.

5.2. CAP effect on bacterial decontamination and skin disinfection

Cold atmospheric plasmas possess a well-known bactericidal activity [194–197]. In CAP-treated solutions such as plasma-activated water (PAW), short and long lived species react with each other and create a powerful anti-microbial mix. Some authors reported that the biocidal properties mostly derives from a combination of oxidative and nitrosative effects induced by the synergistic activity of H₂O₂ and NO[•] [198]. Zhou et al. demonstrated that the biocidal effect is achieved by the combination of H₂O₂ and NO₂⁻. The two molecules alone possess a very weak anti-bacterial activity while reacting together they can form peroxynitrite (ONOO⁻), an unstable isomer of NO₃⁻. Peroxynitrite was described as the key species in PAW-induced bacterial damages [199]. Moreover in PAW the protonated form of peroxynitrite, the peroxynitrous acid (ONOOH), can be further oxidized by H₂O₂ and produce the peroxynitric acid (O₂NOOH), a stronger bactericidal molecule [200]. Within the ROS, O₃, mainly produced by air-fed CAPs, also contributes to the plasma-induced biocidal effect [201]. While bacteria directly exposed to plasma can be quite easily killed by the chemical attack of short and long lived RONS, in real life most of these microorganisms are protected by biofilms. Biofilm is a complex consortium of various microorganisms growing on a substrate. Bacteria are embedded in a sort of clammy extracellular matrix made up of extracellular polymeric substances such as polysaccharides, lipids, protein and DNA that are meant to protect bacteria from dehydration and external environment insults. Thanks to this physical barrier, bacteria are more resistant to antibiotic treatments. *In vitro* tests have shown that CAP is able to destroy biofilms of skin pathogens such as the *candida albicans* yeast [202] and the *staphylococcus aureus* [203]. In direct CAP treatment, biofilms are not only exposed to the highly reactive species but also to an intense electric field that can destabilizes the physicochemical structure of the microorganisms [204]. Interestingly, the CAP ability to destroy biofilms has been demonstrated also *in vivo* in wounds that are not any more responsive to common antibiotics [93,205,206]. Although CAP treatment can be efficiently used to deteriorate biofilms, some bacterial strains could adapt and tolerate plasma treatment stress. Tailoring of CAP devices and treatment conditions are to date a big challenge for a safe and efficient biofilm decontamination [207]. It was already mentioned that skin microbiota plays a physiological role on the intact organ. However when the symbiotic equilibrium between the skin and the microorganisms is disrupted, these latter can become pathogenic [208]. The previously mentioned acidic mantle controls this equilibrium by inhibiting the growth of pathogen organisms and by promoting commensal skin bacteria proliferation. Low pH is unfavorable to the growth of several pathogens, and the already discussed CAP-induced acidification enhances the antimicrobial activity of these sources [187, 209–211]. Thanks to their anti-microbial activities, CAP devices can be used as an alternative and efficient method to sanitize intact skin [92, 212]. CAPs can inhibit the growth of the anaerobic pathogen *Cutibacterium (Propionibacterium) acnes* involved in the oily skin inflammation known as acne vulgaris and in other more severe pathologies [213]. A CAP patented-device was shown to inhibit, *in vitro*, the growth of *Malassezia restricta* and *Malassezia globosa*, yeasts responsible for dandruff [214]. Onychomycosis caused by the bacterium *Escherichia coli* and by the fungus *Trichophyton rubrum* was successfully treated by a helium CAP [215]. In most cases pathogens like the above proliferate in the deep skin appendages. The eradication of these microorganism with topic antibiotic applications is often difficult since the drugs fail to penetrate inside the deep structure of the skin. As will be explained later, CAP treatment can overcome this issue.

5.3. Plasma and skin hydration

A healthy and functional skin needs the right amount of water. In the dermis the hydration is guaranteed by the high hygroscopic GAGs such as hyaluronic acid. In the epidermis, water content varies between 70%

in the viable part and 15–30% in the external layers [216]. The *stratum corneum* is able to sense the environmental humidity and adapt its biochemical metabolism [217]. Corneocytes, the dead cells composing the *stratum corneum*, can keep water thanks to hygroscopic molecules composing the Natural Moisturizing Factor and keratins [218,219]. Corneocytes adhere tightly each other avoiding massive water loss. Furthermore, intercellular lipids such as ceramides create a hydrophobic barrier against desiccation [220]. Cold plasma treatment can exert a double effect on skin hydration. CAP can at first destabilize the skin barrier and desiccate the external layer of the epidermis. A small temporary water loss was observed *in vivo* in human *stratum corneum* after plasma exposure [221]. Epidermis desiccation is a desired effect in plasma skin resurfacing where the non-ablated dried epidermis protects the thermally damaged layers during the recovery process [222]. Paradoxically since CAPs can deposit charges on the treated surface, it is likely that after a plasma treatment skin could attract more water molecules. Human *stratum corneum* wettability rapidly increases in the first seconds of plasma treatment [223]. The increased hydrophilicity was also demonstrated in finger nails where plasma treatment ameliorates the adhesion of cosmetic nail varnish [224].

5.4. RONS skin penetration and CAP-mediated percutaneous absorption

During CAP treatment, RONS can directly influence the skin cell physiology or mediate the activation of downstream responses. One of the hot topic in dermocosmetic plasma therapies is to understand the ability of these reactive species to penetrate and diffuse inside the skin layers. While short lived species rapidly react and deactivate in contact with the *stratum corneum*, long lived molecules such as H_2O_2 , NO_3^- , NO_2^- and NO^\bullet can diffuse deeply. Knowing how these molecules penetrate inside the skin during CAP treatment is of crucial importance in treatment planning. Whether in dermatological therapies the deep penetration of these species is allowed, in pure cosmetic treatments the effect of RONS must be superficial and affects only the epidermis. Simple organic skin mimics such as agarose gel and gelatin have been used to quantify and measure the diffusion kinetics of the reactive species after CAP treatment [225]. These models permit to have an idea on the ability of RONS to cross the mesh of semi-solid gel but they overestimate the ability of these molecules to diffuse into the biological tissues. In real world, the physicochemical barriers of the skin can slow down the diffusion and neutralize most of the reactive species. Even using real skin, the exact quantification of RONS passing through the tissues can be distorted by the nature of the liquid medium used to collect and analyze them [226]. As already mentioned, molecules can cross the skin either through the cells, the intercellular space or via the external pores. Small species such as NO^\bullet can passively diffuse through cell membrane, whereas some others can pass via channel proteins. Indeed, the water channel aquaporins [227,228] were shown to facilitate the cellular diffusion of H_2O_2 produced upon CAP treatment [229,230]. In addition, as previously evoked, thanks to lipid peroxidation RONS can generate transient pores on the membrane and so facilitate their own penetration. Furthermore, by destabilizing the cellular membrane, the electric field generated during the plasma treatment participates to the pore formation [231,232].

Beside the direct role of CAP in skin treatment, cold plasma sources could be also used to ameliorate the effect of topic dermocosmetic cares. Thanks to their ability to destabilize the skin barrier, CAP were shown to facilitate the transdermal delivery of other molecules with a mechanism similar to electroporation [233–236]. This temporary loss of the epidermis barrier could permit the intercellular diffusion of drugs or cosmetic actives. Lastly, RONS and ointments can easily penetrate through the appendageal way whose density depends on the body zone.

6. CAP delivery and skin electrical parameters

CAPs can behave differently depending on the electrical nature of the

target [237–240]. Human organs possess different values of electrical conductivity and this latter depends on the biochemical and biophysical structure of the organ, on its water content and on its extracellular electrolyte composition. In skin, the conductivity varies greatly in the three different layers composing the organ. The *stratum corneum*, with its low water content and its lipid composition, acts as a real dielectric barrier, insulating the whole body from electric shocks. However as already mentioned in the previous paragraph, CAP can temporarily increase the conductivity of the epidermis promoting transdermal penetration of molecules. The electroporation induced by high-voltage pulses lets also the current flow deeper into the tissues. CAP-transported electric current, penetrating the skin layers, could have beneficial effects. High frequency electrotherapy was used since the early 20th century to treat several dermatological conditions and other diseases [241]. It is known that a certain electrical stimulation can speed up the wound healing process by promoting dermal fibroblast motility [242, 243]. In addition, direct and pulsed currents were shown to stimulate keratinocytes differentiation, epidermis proliferation, vascularization and new collagen deposition [244,245].

The interaction between CAPs and the skin is bidirectional, CAP can modify the initial physico-chemical parameters of the targeted tissue while the characteristics of this latter can influence the plasma performance (Fig. 3). The knowledge of the body electrical parameters is crucial for planning a safe, controlled and effective CAP treatment [240, 246]. We already mentioned that the thickness and the structure of the skin change along the body which is why the same plasma treatment can produce a different effect depending on the treated zone. Moreover, the conductivity of the skin itself can vary over the time according to its hydration and to the temporary psychological and hormonal state of the treated person [247,248]. To overcome these variations, CAP delivery should dynamically adapt to the electrical changes of the skin [249]. The applied electric field has to be carefully chosen and controlled and will depend on the desired effect on skin, either a stronger for dermatological use or a lighter for cosmetic applications.

7. Towards a potential role of CAP in skin biology and ageing prevention

In this review we tried to summarize the interactions between cold atmospheric plasmas and the skin, based on the current knowledge of their own properties and characteristics, highlighting the already known and some other possible effects of these sources on the integumentary system biology (Fig. 4). We described how the multiple physical parameters involved in cold plasma can individually or together impact the cutaneous microenvironment and skin cell activities. The coaction of all these effects tends towards a beneficial role of CAPs on skin biology and brings relevant arguments in favor of the use of cold plasma for restoring skin functional barrier and thus improving skin health and appearance.

In the light of these findings, one could ask: can CAPs be considered as a fountain of youth for the skin? Answering this question is not simple.

From a technical point of view, the interesting *in vitro* and *in vivo* data found in the literature come often from different cold-plasma devices. Indeed, worldwide researchers have developed devices with different designs, gases and characteristics for plasma generation. This makes the biological results hardly comparable. Whether one specific device is better for one desired biological effect remains to be determined. Moreover, for a single device, parameters such as voltage, frequency and distance from the target also strongly influence plasma behavior and composition. In one hand, while these technical variables add a new level of complexity, in the other hand they also bring the possibility to modulate the plasma production. Moreover CAP physics and chemistry also depend on the operating conditions, such as ambient temperature and humidity. Thus the device should automatically adapt to the environmental conditions in order to deliver the same performance at every use. Hence, for each treatment, a systematic diagnosis of the device

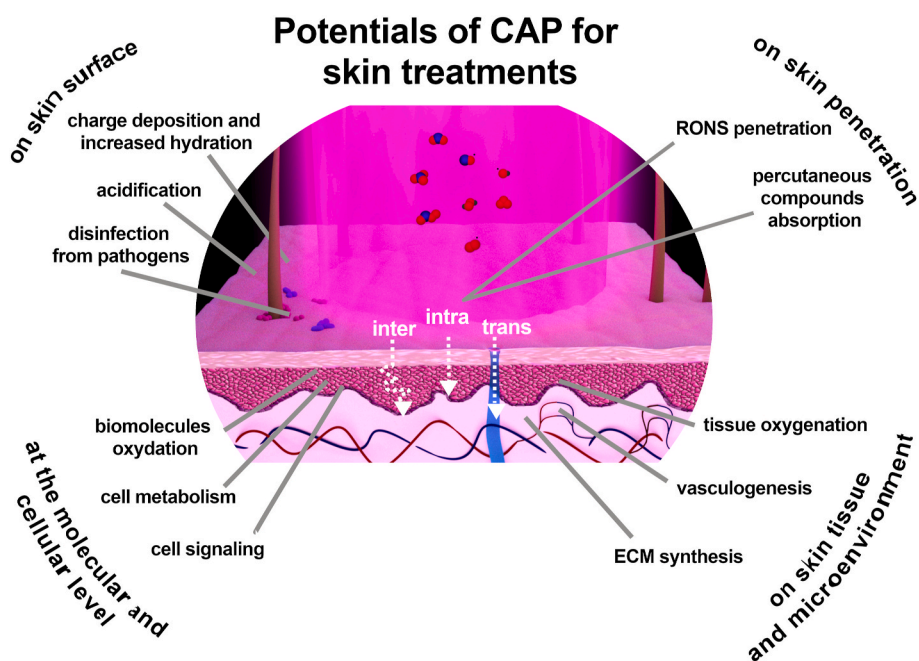


Fig. 4. Potentials of CAP in skin biology. CAP performs its activity at various levels of the skin. At a superficial level it promotes the hydration, acidification and decontamination of the stratum corneum. CAP-generated RONS can penetrate inside the skin via the intercellular way (**inter**), the intracellular way (**intra**) or via the transappendageal way (**trans**). By loosening the cutaneous barrier, CAP also promotes the absorption of other molecules such as drugs. At a molecular level, once penetrated into the skin, RONS can have a direct effect on skin biomolecule oxidation or activate cell metabolism and signaling. At the tissue level, CAP treatment lead to an increase in skin oxygenation, stimulates the vasculogenesis and the ECM remodeling or *de-novo* synthesis.

plasma composition should bring a new tool for the control and modulation of RONS generation. The development of means to finely analyze and control plasma delivery in interaction with skin with the support of modern plasma medicine studies should allow to determine the optimal conditions of use and contribute to the evaluation of this technology for skin wellbeing. Finally this CAP “plasticity” can become a strong point for specific applications.

From a biochemical point of view, the mechanisms involved in plasma effect on cells and tissues still remains to be determined. Progress in this field are currently ongoing by making a precise diagnosis of plasma composition with the characterization of RONS produced in liquid or in tissues. The next step is to understand how these species are able to induce a biological effect either alone or in synergy [250]. The last challenging step is to master all the other CAP components (charged particles, UV, electric field, thermal emission) that cooperate to the final biological effect. Such rational approaches should strengthen plasma effects understanding and bring new insight in plasma applications for skin but also more generally in plasma medicine domain [251].

From a biological point of view and according to the results reported till now in the literature and presented here, CAPs should be a promising technology to stimulate and/or regenerate skin. However, as noted in this work, relatively little is known about the molecular and cellular mechanisms involved in the plasma-induced biological effects and about the consequences of these treatments in the long term. Data obtained from *in vitro* experiments are essential to understand the cellular signaling activated by the CAP exposure. However, the biggest limit in the use of cell cultures in plasma treatments is that most of the time the cells are not directly exposed to the CAP. Cells are either treated through the physiological medium covering them or put in contact with plasma-pretreated medium. Thus, cells in culture receive only long-lived species diffusing from the surface of the medium or dissolved in the plasma-activated medium. Conversely, *in vivo*, the skin will be impacted by all the CAP-produced species since the *stratum corneum* of the epidermis is directly exposed to the atmosphere. To overcome this issue, the use of skin explants or more complex skin models such as Reconstructed Human Epidermis or Reconstructed Whole Skin could strengthen the understanding of the CAP-activated processes *in vitro* and allow to modulate the effects, define and correct the limits of this technology. Finally, once clear cause-and-effect mechanisms are defined in these studies, real life treatment must be adapted to each user as the skin

structure depends on the phototype but also on the age, sex and ethnicity. Indeed dermo-cosmetic CAP-based treatments should be considered as adapted and personalized therapies which are currently increasingly in demand. Thus a new generation of plasma-based treatments could emerge in the field of cosmetics and dermatology to improve skin aspect and health.

Taking into account the flexibility of plasma generation, thanks to its many parameters, CAPs could become new and promising treatments for skin care and regeneration. This will go through an optimization of devices, in terms of efficacy, control and safety for use as well as through the deciphering of mechanisms involved in plasma-induced effects on the skin. This innovative technology opens new areas of research at the interface between plasma physics and skin biology and should lead towards new applications in the dermo-cosmetic field.

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Declaration of competing interest

The authors declare that they have no competing interests.

Abbreviations

CAP	Cold atmospheric Plasma
ROS	Reactive Oxygen Species
RNS	Reactive Nitrogen Species
RONS	Reactive Oxygen and Nitrogen Species
DBD	Dielectric Barrier Discharge
ECM	Extracellular Matrix

References

- [1] G.K. Menon, A.M. Kligman, Barrier functions of human skin: a holistic view, *Skin Pharmacol. Physiol.* 22 (4) (2009) 178–189, <https://doi.org/10.1159/000231523>.
- [2] W.Z. Mostafa, R.A. Hegazy, Vitamin D and the skin: focus on a complex relationship: a review, *J. Adv. Res.* 6 (6) (2015) 793–804, <https://doi.org/10.1016/j.jare.2014.01.011>.
- [3] J.P. Kultz-Buschbeck, W. Andresen, S. Gobel, R. Gilster, C. Stick, Thermoreception and nociception of the skin: a classic paper of Bessou and Perl and analyses of thermal sensitivity during a student laboratory exercise, *Adv. Physiol. Educ.* 34 (2) (2010) 25–34, <https://doi.org/10.1152/advan.00002.2010>.
- [4] D. Filingeri, Neurophysiology of skin thermal sensations, *Comp. Physiol.* 6 (3) (2016) 1429, <https://doi.org/10.1002/cphy.c150040>.
- [5] D. Filingeri, Humidity sensation, cockroaches, worms, and humans: are common sensory mechanisms for hygro-sensation shared across species? *J. Neurophysiol.* 114 (2) (2015) 763–767, <https://doi.org/10.1152/jn.00730.2014>.
- [6] A.A. Romanovsky, Skin temperature: its role in thermoregulation, *Acta Physiol.* 210 (3) (2014) 498–507, <https://doi.org/10.1111/apha.12231>.
- [7] T.S. Poet, J.N. McDougal, Skin absorption and human risk assessment, *Chem. Biol. Interact.* 140 (1) (2002) 19–34, [https://doi.org/10.1016/s0009-2797\(02\)00013-3](https://doi.org/10.1016/s0009-2797(02)00013-3).
- [8] M. Gallagher, C.J. Wysocki, J.J. Leyden, A.I. Spielman, X. Sun, G. Preti, Analyses of volatile organic compounds from human skin, *Br. J. Dermatol.* 159 (4) (2008) 780–791, <https://doi.org/10.1111/j.1365-2133.2008.08748.x>.
- [9] H.J. Hurley, J. Witkowski, Dye clearance and eccrine sweat secretion in human skin, *J. Invest. Dermatol.* 36 (1961) 259–272, <https://doi.org/10.1038/jid.1961.44>.
- [10] K.R. Smith, D.M. Thiboutot, Thematic review series: skin lipids. Sebaceous gland lipids: friend or foe? *J. Lipid Res.* 49 (2) (2008) 271–281, <https://doi.org/10.1194/jlr.R700015-JLR200>.
- [11] Y. Peng, X. Cui, Y. Liu, Y. Li, J. Liu, B. Cheng, Systematic review focusing on the excretion and protection roles of sweat in the skin, *Dermatology* 228 (2) (2014) 115–120, <https://doi.org/10.1159/000357524>.
- [12] E. Guttman-Yassky, L. Zhou, J.G. Krueger, The skin as an immune organ: tolerance versus effector responses and applications to food allergy and hypersensitivity reactions, *J. Allergy Clin. Immunol.* 144 (2) (2019) 362–374, <https://doi.org/10.1016/j.jaci.2019.03.021>.
- [13] M. Shahbandeh, Value of the Cosmetics Market Worldwide from 2018 to 2025 (In Billion U.S. Dollars), 2019, 27 Nov 2019; Available from: <https://www.statista.com/statistics/585522/global-value-cosmetics-market/>.
- [14] E. Papadavid, A. Katsambas, Lasers for facial rejuvenation: a review, *Int. J. Dermatol.* 42 (6) (2003) 480–487, <https://doi.org/10.1046/j.1365-4362.2003.01784.x>.
- [15] F. Baez, L.R. Reilly, The use of light-emitting diode therapy in the treatment of photoaged skin, *J. Cosmet. Dermatol.* 6 (3) (2007) 189–194, <https://doi.org/10.1111/j.1473-2165.2007.00329.x>.
- [16] S. Shuster, M.M. Black, E. McVitie, The influence of age and sex on skin thickness, skin collagen and density, *Br. J. Dermatol.* 93 (6) (1975) 639–643, <https://doi.org/10.1111/j.1365-2133.1975.tb05113.x>.
- [17] J. Sandby-Moller, T. Poulsen, H.C. Wulf, Epidermal thickness at different body sites: relationship to age, gender, pigmentation, blood content, skin type and smoking habits, *Acta Derm. Venereol.* 83 (6) (2003) 410–413, <https://doi.org/10.1080/00015550310015419>.
- [18] S.C. Taylor, Skin of color: biology, structure, function, and implications for dermatologic disease, *J. Am. Acad. Dermatol.* 46 (2 Suppl Understanding) (2002) S41–S62, <https://doi.org/10.1067/mjd.2002.120790>.
- [19] P.R. Bergstresser, J.R. Taylor, Epidermal turnover time—a new examination, *Br. J. Dermatol.* 96 (5) (1977) 503–509, <https://doi.org/10.1111/j.1365-2133.1977.tb07152.x>.
- [20] C.S. Potten, C. Booth, Keratinocyte stem cells: a commentary, *J. Invest. Dermatol.* 119 (4) (2002) 888–899, <https://doi.org/10.1046/j.1523-1747.2002.00020.x>.
- [21] C. Pincelli, A. Marconi, Keratinocyte stem cells: friends and foes, *J. Cell. Physiol.* 225 (2) (2010) 310–315, <https://doi.org/10.1002/jcp.22275>.
- [22] I.M. Braverman, The cutaneous microcirculation, *J. Invest. Dermatol. Symp. Proc.* 5 (1) (2000) 3–9, <https://doi.org/10.1046/j.1087-0024.2000.00010.x>.
- [23] A. Carreau, B. El Hafny-Rahbi, A. Matejuk, C. Grillon, C. Kieda, Why is the partial oxygen pressure of human tissues a crucial parameter? Small molecules and hypoxia, *J. Cell Mol. Med.* 15 (6) (2011) 1239–1253, <https://doi.org/10.1111/j.1582-4934.2011.01258.x>.
- [24] N.T. Evans, P.F. Naylor, The systemic oxygen supply to the surface of human skin, *Respir. Physiol.* 3 (1) (1967) 21–37, [https://doi.org/10.1016/0034-5687\(67\)90020-5](https://doi.org/10.1016/0034-5687(67)90020-5).
- [25] H. Sies, C. Berndt, D.P. Jones, Oxidative stress, *Annu. Rev. Biochem.* 86 (1) (2017) 715–748, <https://doi.org/10.1146/annurev-biochem-061516-045037>.
- [26] F.A.S. Addor, Antioxidants in dermatology, *An. Bras. Dermatol.* 92 (3) (2017) 356–362, <https://doi.org/10.1590/abd1806-4841.20175697>.
- [27] S.M. Ali, G. Yosipovitch, Skin pH: from basic science to basic skin care, *Acta Derm. Venereol.* 93 (3) (2013) 261–267, <https://doi.org/10.2340/00015555-1531>.
- [28] R. Weller, S. Pattullo, L. Smith, M. Golden, A. Ormerod, N. Benjamin, Nitric oxide is generated on the skin surface by reduction of sweat nitrate, *J. Invest. Dermatol.* 107 (3) (1996) 327–331, <https://doi.org/10.1111/1523-1747.ep12363167>.
- [29] P.M. Krien, M. Kermici, Evidence for the existence of a self-regulated enzymatic process within the human stratum corneum—an unexpected role for urocanic acid, *J. Invest. Dermatol.* 115 (3) (2000) 414–420, <https://doi.org/10.1046/j.1523-1747.2000.00083.x>.
- [30] J.W. Fluhr, J. Kao, S.K. Ahn, K.R. Feingold, P.M. Elias, M. Jain, Generation of free fatty acids from phospholipids regulates stratum corneum acidification and integrity, *J. Invest. Dermatol.* 117 (1) (2001) 44–51, <https://doi.org/10.1046/j.0022-202x.2001.01399.x>.
- [31] M.J. Behne, J. Meyer, K.M. Hanson, N.P. Barry, S. Murata, D. Crumrine, R. R. Clegg, E. Gratton, W.M. Holleran, P.M. Elias, T.M. Mauro, NHE1 regulates the stratum corneum permeability barrier homeostasis: microenvironment acidification assessed with FLIM, *J. Biol. Chem.* (2002), <https://doi.org/10.1074/jbc.M204759200>.
- [32] J.-P. Hachem, D. Crumrine, J. Fluhr, B.E. Brown, K.R. Feingold, P.M. Elias, pH directly regulates epidermal permeability barrier homeostasis, and stratum corneum integrity/cohesion, *J. Invest. Dermatol.* 121 (2) (2003) 345–353, <https://doi.org/10.1046/j.1523-1747.2003.12365.x>.
- [33] M.H. Schmid-Wendtner, H.C. Korting, The pH of the skin surface and its impact on the barrier function, *Skin Pharmacol. Physiol.* 19 (6) (2006) 296–302, <https://doi.org/10.1159/000094670>.
- [34] D.J. Panther, S.E. Jacob, The importance of acidification in atopic eczema: an underexplored avenue for treatment, *J. Clin. Med.* 4 (5) (2015) 970–978, <https://doi.org/10.3390/jcm4050970>.
- [35] F. Rippe, V. Schreiner, T. Doering, H.I. Maibach, Stratum corneum pH in atopic dermatitis, *Am. J. Clin. Dermatol.* 5 (4) (2004) 217–223, <https://doi.org/10.2165/00128071-200405040-00002>.
- [36] H. Zhai, H.P. Chan, S. Farahmand, H.I. Maibach, Measuring human skin buffering capacity: an in vitro model, *Skin Res. Technol.* 15 (4) (2009) 470–475, <https://doi.org/10.1111/j.1600-0846.2009.00392.x>.
- [37] J.A. Bouwstra, M. Ponc, The skin barrier in healthy and diseased state, *Biochim. Biophys. Acta* 1758 (12) (2006) 2080–2095, <https://doi.org/10.1016/j.bbame.2006.06.021>.
- [38] Z. Nemes, P.M. Steinert, Bricks and mortar of the epidermal barrier, *Exp. Mol. Med.* 31 (1) (1999) 5–19, <https://doi.org/10.1038/emmm.1999.2>.
- [39] G.K. Menon, G.W. Cleary, M.E. Lane, The structure and function of the stratum corneum, *Int. J. Pharm.* 435 (1) (2012) 3–9, <https://doi.org/10.1016/j.ijpharm.2012.06.005>.
- [40] M.B. Murphrey, P.M. Zito, *Histology, Stratum Corneum, StatPearls*, Treasure Island (FL), 2018.
- [41] R.R. Wickett, M.O. Visscher, Structure and function of the epidermal barrier, *Am. J. Infect. Contr.* 34 (10, Supplement) (2006) S98–S110, <https://doi.org/10.1016/j.ajic.2006.05.295>.
- [42] A.S. Michaels, S.K. Chandrasekaran, J.E. Shaw, Drug permeation through human skin: theory and invitro experimental measurement, *AIChE J.* 21 (5) (1975) 985–996, <https://doi.org/10.1002/aic.690210522>.
- [43] C. Vitorino, J. Sousa, A. Pais, Overcoming the skin permeation barrier: challenges and opportunities, *Curr. Pharmaceut. Des.* 21 (20) (2015) 2698–2712, <https://doi.org/10.2174/1381612821666150428124053>.
- [44] T.R. Auton, Concentration-flux relations for a multicellular biological membrane with metabolism, *Math. Biosci.* 115 (1) (1993) 103–117, [https://doi.org/10.1016/0025-5564\(93\)90048-F](https://doi.org/10.1016/0025-5564(93)90048-F).
- [45] L. Norlen, Skin barrier structure and function: the single gel phase model, *J. Invest. Dermatol.* 117 (4) (2001) 830–836, <https://doi.org/10.1038/jid.2001.1>.
- [46] H. Trommer, R.H. Neubert, Overcoming the stratum corneum: the modulation of skin penetration. A review, *Skin Pharmacol. Physiol.* 19 (2) (2006) 106–121, <https://doi.org/10.1159/000091978>.
- [47] S. Mitragotri, Y.G. Anissimov, A.L. Bunge, H.F. Frasch, R.H. Guy, J. Hadgraft, G. B. Kasting, M.E. Lane, M.S. Roberts, Mathematical models of skin permeability: an overview, *Int. J. Pharm.* 418 (1) (2011) 115–129, <https://doi.org/10.1016/j.ijpharm.2011.02.023>.
- [48] E.A. Grice, J.A. Segre, The skin microbiome, *Nat. Rev. Microbiol.* 9 (4) (2011) 244–253, <https://doi.org/10.1038/nrmicro2537>.
- [49] L. Bay, C.J. Barnes, B.G. Fritz, J. Thorsen, M.E.M. Restrup, L. Rasmussen, J. K. Sorensen, A.B. Hesselvig, A. Odgaard, A.J. Hansen, T. Bjarnsholt, Universal dermal microbiome in human skin, *mBio* 11 (1) (2020), <https://doi.org/10.1128/mBio.02945-19.e02945-19>.
- [50] R.L. Gallo, Human skin is the largest epithelial surface for interaction with microbes, *J. Invest. Dermatol.* 137 (6) (2017) 1213–1214, <https://doi.org/10.1016/j.jid.2016.11.045>.
- [51] K. Chiller, B.A. Selkin, G.J. Murakawa, Skin microflora and bacterial infections of the skin, *J. Invest. Dermatol. Symp. Proc.* 6 (3) (2001) 170–174, <https://doi.org/10.1046/j.0022-202x.2001.00043.x>.
- [52] A.L. Cogen, V. Nizet, R.L. Gallo, Skin microbiota: a source of disease or defence? *Br. J. Dermatol.* 158 (3) (2008) 442–455, <https://doi.org/10.1111/j.1365-2133.2008.08437.x>.
- [53] H. Vallhov, C. Johansson, R.E. Veerman, A. Scheynius, Extracellular vesicles released from the skin commensal yeast *Malassezia sympodialis* activate human primary keratinocytes, *Frontiers in Cellular and Infection Microbiology* 10 (6) (2020), <https://doi.org/10.3389/fcimb.2020.00006>.
- [54] M.-A. Dagnelie, S. Corvec, A. Khammari, B. Dréno, Bacterial extracellular vesicles: a new way to decipher host-microbiota communications in inflammatory dermatoses, *Exp. Dermatol.* 29 (1) (2020) 22–28, <https://doi.org/10.1111/exd.14050>.
- [55] G.J. Fisher, S. Kang, J. Varani, Z. Bata-Csorgo, Y. Wan, S. Datta, J.J. Voorhees, Mechanisms of photoaging and chronological skin aging, *Arch. Dermatol.* 138 (11) (2002) 1462–1470, <https://doi.org/10.1001/archderm.138.11.1462>.

- [56] J. Krutmann, A. Bouloc, G. Sore, B.A. Bernard, T. Passeron, The skin aging exposome, *J. Dermatol. Sci.* 85 (3) (2017) 152–161, <https://doi.org/10.1016/j.jdermsci.2016.09.015>.
- [57] F. Paschen, Ueber die zum Funkenübergang in Luft, Wasserstoff und Kohlensäure bei verschiedenen Drucken erforderliche Potentialdifferenz, *Ann. Phys.* 273 (5) (1889) 69–96, <https://doi.org/10.1002/andp.18892730505>.
- [58] I. Langmuir, Oscillations in ionized gases, *Proc. Natl. Acad. Sci. U.S.A.* 14 (1928) 627–637, <https://doi.org/10.1073/pnas.14.4.627>.
- [59] P. Fauchais, A. Vardelle, Thermal plasmas, *IEEE Trans. Plasma Sci.* 25 (6) (1997) 1258–1280, <https://doi.org/10.1109/27.650901>.
- [60] J. Heberlein, A.B. Murphy, Thermal plasma waste treatment, *J. Phys. Appl. Phys.* 41 (5) (2008), 053001, <https://doi.org/10.1088/0022-3727/41/5/053001>.
- [61] G. Fridman, A. Friedman, A. Gutsol, A.B. Shekhter, V.N. Vasilets, A. Fridman, *Applied Plasma Medicine. Plasma Processes and Polymers* 5 (6) (2008) 503–533, <https://doi.org/10.1002/ppap.200700154>.
- [62] Y.-F. Li, T. Shimizu, J.L. Zimmermann, G.E. Morfill, Cold atmospheric plasma for surface disinfection, *Plasma Process. Polym.* 9 (6) (2012) 585–589, <https://doi.org/10.1002/ppap.201100090>.
- [63] M. Laroussi, T. Akan, Arc-free atmospheric pressure cold plasma jets: a review, *Plasma Process. Polym.* 4 (9) (2007) 777–788, <https://doi.org/10.1002/ppap.200700066>.
- [64] C. Hoffmann, C. Berganza, J. Zhang, Cold Atmospheric Plasma: methods of production and application in dentistry and oncology, *Med. Gas Res.* 3 (1) (2013) 21, <https://doi.org/10.1186/2045-9912-3-21>.
- [65] D. Rymuszka, K. Terpiłowski, P. Borowski, L. Holysz, Time-dependent changes of surface properties of polyether ether ketone caused by air plasma treatment, *Polym. Int.* 65 (7) (2016) 827–834, <https://doi.org/10.1002/pi.5141>.
- [66] W. Tian, A.M. Lietz, M.J. Kushner, The consequences of air flow on the distribution of aqueous species during dielectric barrier discharge treatment of thin water layers, *Plasma Sources Sci. Technol.* 25 (5) (2016), 055020, <https://doi.org/10.1088/0963-0252/25/5/055020>.
- [67] J. Chauvin, F. Judée, M. Yousfi, P. Vicendo, N. Merbahi, Analysis of reactive oxygen and nitrogen species generated in three liquid media by low temperature helium plasma jet, *Sci. Rep.* 7 (1) (2017) 4562, <https://doi.org/10.1038/s41598-017-04650-4>.
- [68] R. Thirumdas, A. Kothakota, U. Annappure, K. Siliveru, R. Blundell, R. Gatt, V. P. Valdramidis, Plasma activated water (PAW): chemistry, physico-chemical properties, applications in food and agriculture, *Trends Food Sci. Technol.* 77 (2018) 21–31, <https://doi.org/10.1016/j.tifs.2018.05.007>.
- [69] J. Liu, B. He, Q. Chen, J. Li, Q. Xiong, G. Yue, X. Zhang, S. Yang, H. Liu, Q.H. Liu, Direct synthesis of hydrogen peroxide from plasma-water interactions, *Sci. Rep.* 6 (2016) 38454, <https://doi.org/10.1038/srep38454>, <https://www.nature.com/articles/srep38454#supplementary-information>.
- [70] G. Busco, F. Fasani, S. Dozias, L. Ridou, C. Douat, J.-M. Pouvesle, E. Robert, C. Grillon, Changes in oxygen level upon cold plasma treatments: consequences for RONS production, *IEEE Transactions on Radiation and Plasma Medical Sciences* 2 (2) (2018) 147–152, <https://doi.org/10.1109/trpms.2017.2775705>.
- [71] E. Turrini, R. Laurita, E. Simoncelli, A. Stancampiano, E. Catanzaro, C. Calcebrini, G. Carulli, M. Rousseau, M. Gherardi, F. Maffei, V. Cocchi, M. Lenzi, V. Pellicioni, P. Hrelia, V. Colombo, C. Fimognari, Plasma-activated medium as an innovative anticancer strategy: insight into its cellular and molecular impact on in vitro leukemia cells, *Plasma Process. Polym.* (2020), <https://doi.org/10.1002/ppap.202000007> n/a(n/a): p. e2000007.
- [72] A. Valinataj Omran, G. Busco, L. Ridou, S. Dozias, C. Grillon, J.-M. Pouvesle, E. Robert, Cold atmospheric single plasma jet for RONS delivery on large biological surfaces, *Plasma Sources Sci. Technol.* 29 (10) (2020), 105002, <https://doi.org/10.1088/1361-6595/abaffd>.
- [73] E. Gjika, S. Pal-Ghosh, A. Tang, M. Kirschner, G. Tadvalkar, J. Canady, M. A. Stepp, M. Keidar, Adaptation of operational parameters of cold atmospheric plasma for in vitro treatment of cancer cells, *ACS Appl. Mater. Interfaces* 10 (11) (2018) 9269–9279, <https://doi.org/10.1021/acsami.7b18653>.
- [74] M.G. Kong, G. Kroesen, G. Morfill, T. Nosenko, T. Shimizu, J.v. Dijk, J. L. Zimmermann, Plasma medicine: an introductory review, *New J. Phys.* 11 (11) (2009) 115012, <https://doi.org/10.1088/1367-2630/11/11/115012>.
- [75] D. Yan, J.H. Sherman, M. Keidar, Cold atmospheric plasma, a novel promising anti-cancer treatment modality, *Oncotarget* 8 (9) (2017) 15977–15995, <https://doi.org/10.18632/oncotarget.13304>.
- [76] J. Schlegel, J. Körtzer, V. Boxhammer, Plasma in cancer treatment, *Clinical Plasma Medicine* 1 (2) (2013) 2–7, <https://doi.org/10.1016/j.cpm.2013.08.001>.
- [77] G. Isbary, J.L. Zimmermann, T. Shimizu, Y.F. Li, G.E. Morfill, H.M. Thomas, B. Steffes, J. Heinlin, S. Karrer, W. Stolz, Non-thermal plasma—more than five years of clinical experience, *Clinical Plasma Medicine* 1 (1) (2013) 19–23, <https://doi.org/10.1016/j.cpm.2012.11.001>.
- [78] A.M. Hirst, F.M. Frame, M. Arya, N.J. Maitland, D. O'Connell, Low temperature plasmas as emerging cancer therapeutics: the state of play and thoughts for the future, *Tumour Biol* 37 (6) (2016) 7021–7031, <https://doi.org/10.1007/s13277-016-4911-7>.
- [79] A. Dubuc, P. Monsarrat, F. Virard, N. Merbahi, J.-P. Sarrette, S. Laurencin-Dalieux, S. Cousty, Use of cold-atmospheric plasma in oncology: a concise systematic review, *Therapeutic advances in medical oncology* 10 (2018), <https://doi.org/10.1177/1758835918786475>, 1758835918786475–1758835918786475.
- [80] P.M. Metelmann, D.S. Nedrelow, M. Schuster, R. Rutkowski, C. Seebauer, Clinical studies applying physical plasma in head and neck cancer - key points and study design, *Int J Clin Res Trials* 1 (103) (2016) 2, <https://doi.org/10.15344/2456-8007/2016/103>.
- [81] R. Guerrero-Preston, T. Ogawa, M. Uemura, G. Shumulinsky, B.L. Valle, F. Pirini, R. Ravi, D. Sidransky, M. Keidar, B. Trink, Cold atmospheric plasma treatment selectively targets head and neck squamous cell carcinoma cells, *Int. J. Mol. Med.* 34 (4) (2014) 941–946, <https://doi.org/10.3892/ijmm.2014.1849>.
- [82] S.J. Kim, T.H. Chung, Cold atmospheric plasma jet-generated RONS and their selective effects on normal and carcinoma cells, *Sci. Rep.* 6 (2016) 20332, <https://doi.org/10.1038/srep20332>.
- [83] M. Kuchenbecker, N. Bibinov, A. Kaemling, D. Wandke, P. Awakowicz, W. Viöl, Characterization of DBD plasma source for biomedical applications, *J. Phys. Appl. Phys.* 42 (4) (2009), <https://doi.org/10.1088/0022-3727/42/4/045212>, 045212.
- [84] T. Borchardt, J. Ernst, A. Helmke, M. Tanyeli, A.F. Schilling, G. Felmerer, W. Viöl, Effect of direct cold atmospheric plasma (diCAP) on microcirculation of intact skin in a controlled mechanical environment, *Microcirculation* 24 (8) (2017), <https://doi.org/10.1111/micc.12399>.
- [85] M. Vandamme, E. Robert, S. Dozias, J. Sobilo, S. Lerondel, A. Le Pape, J. M. Poudmes, Response of human glioma U87 xenograft on mice to non thermal plasma treatment, *Plasma Med.* 1 (1) (2011) 27–43, <https://doi.org/10.1615/PlasmaMed.v1.i1.30>.
- [86] S. Mirpour, S. Piroozmand, N. Soleimani, N. Jalali Faharani, H. Ghomi, H. Fotovat Eskandari, A.M. Sharifi, S. Mirpour, M. Eftekhari, M. Nikkhab, Utilizing the micron sized non-thermal atmospheric pressure plasma inside the animal body for the tumor treatment application, *Sci. Rep.* 6 (2016) 29048, <https://doi.org/10.1038/srep29048>.
- [87] S. Kubinova, K. Zavisikova, L. Uherkova, V. Zablotskii, O. Churpita, O. Lunov, A. Dejneka, Non-thermal air plasma promotes the healing of acute skin wounds in rats, *Sci. Rep.* 7 (2017) 45183, <https://doi.org/10.1038/srep45183>, <https://www.nature.com/articles/srep45183#supplementary-information>.
- [88] B. Haertel, T. von Woedtke, K.D. Weltmann, U. Lindequist, Non-thermal atmospheric-pressure plasma possible application in wound healing, *Biomol Ther (Seoul)* 22 (6) (2014) 477–490, <https://doi.org/10.4062/biomolther.2014.105>.
- [89] J. Heinlin, J.L. Zimmermann, F. Zeman, W. Bunk, G. Isbary, M. Landthaler, T. Maisch, R. Monetti, G. Morfill, T. Shimizu, J. Steinbauer, W. Stolz, S. Karrer, Randomized placebo-controlled human pilot study of cold atmospheric argon plasma on skin graft donor sites, *Wound Repair Regen.* 21 (6) (2013) 800–807, <https://doi.org/10.1111/wrr.12078>.
- [90] S. Emmert, F. Brehmer, H. Hänßle, A. Helmke, N. Mertens, R. Ahmed, D. Simon, D. Wandke, W. Maus-Friedrichs, G. Däschlein, M.P. Schön, W. Viöl, Atmospheric pressure plasma in dermatology: ulcer treatment and much more, *Clinical Plasma Medicine* 1 (1) (2013) 24–29, <https://doi.org/10.1016/j.cpm.2012.11.002>.
- [91] G. Daeschlein, S. Scholz, R. Ahmed, A. Majumdar, T. von Woedtke, H. Haase, M. Niggemeier, E. Kindel, R. Brandenburg, K.D. Weltmann, M. Junger, Cold plasma is well-tolerated and does not disturb skin barrier or reduce skin moisture, *J Dtsch Dermatol Ges* 10 (7) (2012) 509–515, <https://doi.org/10.1111/j.1610-0387.2012.07857.x>.
- [92] G. Daeschlein, S. Scholz, R. Ahmed, T. von Woedtke, H. Haase, M. Niggemeier, E. Kindel, R. Brandenburg, K.D. Weltmann, M. Juenger, Skin decontamination by low-temperature atmospheric pressure plasma jet and dielectric barrier discharge plasma, *J. Hosp. Infect.* 81 (3) (2012) 177–183, <https://doi.org/10.1016/j.jhin.2012.02.012>.
- [93] G. Isbary, J. Heinlin, T. Shimizu, J.L. Zimmermann, G. Morfill, H.U. Schmidt, R. Monetti, B. Steffes, W. Bunk, Y. Li, T. Klaempfl, S. Karrer, M. Landthaler, W. Stolz, Successful and safe use of 2 min cold atmospheric argon plasma in chronic wounds: results of a randomized controlled trial, *Br. J. Dermatol.* 167 (2) (2012) 404–410, <https://doi.org/10.1111/j.1365-2133.2012.10923.x>.
- [94] J. Heinlin, G. Isbary, W. Stolz, G. Morfill, M. Landthaler, T. Shimizu, B. Steffes, T. Nosenko, J. Zimmermann, S. Karrer, Plasma applications in medicine with a special focus on dermatology, *J. Eur. Acad. Dermatol. Venereol.* 25 (1) (2011) 1–11, <https://doi.org/10.1111/j.1468-3083.2010.03702.x>.
- [95] M.J. Gonzalez, W.H. Sturgill, E.V. Ross, N.S. Uebelhoefer, Treatment of acne scars using the plasma skin regeneration (PSR) system, *Laser Surg. Med.* 40 (2) (2008) 124–127, <https://doi.org/10.1002/lsm.20617>.
- [96] S. Kilmer, N. Semchysyn, G. Shah, R. Fitzpatrick, A pilot study on the use of a plasma skin regeneration device (Portrait PSR3) in full facial rejuvenation procedures, *Laser Med. Sci.* 22 (2) (2007) 101–109, <https://doi.org/10.1007/s10103-006-0431-9>.
- [97] R. Fitzpatrick, E. Bernstein, S. Iyer, D. Brown, P. Andrews, K. Penny, A histopathologic evaluation of the plasma skin regeneration system (PSR) versus a standard carbon dioxide resurfacing laser in an animal model, *Laser Surg. Med.* 40 (2) (2008) 93–99, <https://doi.org/10.1002/lsm.20547>.
- [98] R.D. Gentile, J.D. McCoy, Pulsed and fractionated techniques for helium plasma energy skin resurfacing, *Facial Plastic Surgery Clinics of North America* 28 (1) (2020) 75–85, <https://doi.org/10.1016/j.fsc.2019.09.007>.
- [99] M.L. Elsaie, J.N. Kammer, Evaluation of plasma skin regeneration technology for cutaneous remodeling, *J. Cosmet. Dermatol.* 7 (4) (2008) 309–311, <https://doi.org/10.1111/j.1473-2165.2008.00411.x>.
- [100] M.A. Bogle, K.A. Arndt, J.S. Dover, Evaluation of plasma skin regeneration technology in low-energy full-facial rejuvenation, *Arch. Dermatol.* 143 (2) (2007) 168–174, <https://doi.org/10.1001/archderm.143.2.168>.
- [101] S.H. Bentkover, Plasma skin resurfacing: personal experience and long-term results, *Facial Plast. Surg. Clin.* 20 (2) (2012) 145–162, <https://doi.org/10.1016/j.fsc.2012.02.010>.
- [102] J.H. Choi, H.W. Lee, J.K. Lee, J.W. Hong, G.C. Kim, Low-temperature atmospheric plasma increases the expression of anti-aging genes of skin cells without causing cellular damages, *Arch. Dermatol. Res.* 305 (2) (2013) 133–140, <https://doi.org/10.1007/s00403-012-1259-8>.

- [103] A. Khlyustova, C. Labay, Z. Machala, M.-P. Ginebra, C. Canal, Important parameters in plasma jets for the production of RONS in liquids for plasma medicine: a brief review, *Front. Chem. Sci. Eng.* 13 (2) (2019) 238–252, <https://doi.org/10.1007/s11705-019-1801-8>.
- [104] S.A. Norberg, W.H. Tian, E. Johnsen, K.M. J. Atmospheric pressure plasma jets interacting with liquid covered tissue: touching and not-touching the liquid, *J. Phys. Appl. Phys.* 47 (47) (2014) 475203, <https://doi.org/10.1088/0022-3727/47/47/475203>.
- [105] D.B. Graves, Reactive species from cold atmospheric plasma: implications for cancer therapy, *Plasma Process. Polym.* 11 (12) (2014) 1120–1127, <https://doi.org/10.1002/ppap.201400068>.
- [106] A. Weidinger, A.V. Kozlov, Biological activities of reactive oxygen and nitrogen species: oxidative stress versus signal transduction, *Biomolecules* 5 (2) (2015) 472–484, <https://doi.org/10.3390/biom5020472>.
- [107] P. Attri, N. Kumar, J.H. Park, D.K. Yadav, S. Choi, H.S. Uhm, I.T. Kim, E.H. Choi, W. Lee, Influence of reactive species on the modification of biomolecules generated from the soft plasma, *Sci. Rep.* 5 (2015) 8221, <https://doi.org/10.1038/srep08221>. <https://www.nature.com/articles/srep08221#supplementary-information>.
- [108] C. Bernard, A. Leduc, J. Barbeau, B. Saoudi, L.H. Yahia, G.D. Crescenzo, Validation of cold plasma treatment for protein inactivation: a surface plasmon resonance-based biosensor study, *J. Phys. Appl. Phys.* 39 (16) (2006) 3470–3478, <https://doi.org/10.1088/0022-3727/39/16/s04>.
- [109] E. Takai, T. Kitamura, J. Kuwabara, S. Ikawa, S. Yoshizawa, K. Shiraki, H. Kawasaki, R. Arakawa, K. Kitano, Chemical modification of amino acids by atmospheric-pressure cold plasma in aqueous solution, *J. Phys. Appl. Phys.* 47 (28) (2014) 285403, <https://doi.org/10.1088/0022-3727/47/28/285403>.
- [110] J.W. Lackmann, K. Wende, C. Verlackt, J. Golda, J. Volzke, F. Kogelheide, J. Held, S. Bekechus, A. Bogaerts, V. Schulz-von der Gathen, K. Stapelmann, Chemical fingerprints of cold physical plasmas – an experimental and computational study using cysteine as tracer compound, *Sci. Rep.* 8 (1) (2018) 7736, <https://doi.org/10.1038/s41598-018-25937-0>.
- [111] E. Takai, K. Kitano, J. Kuwabara, K. Shiraki, Protein inactivation by low-temperature atmospheric pressure plasma in aqueous solution, *Plasma Process. Polym.* 9 (1) (2012) 77–82, <https://doi.org/10.1002/ppap.201100063>.
- [112] S.K. Pankaj, N.N. Misra, P.J. Cullen, Kinetics of tomato peroxidase inactivation by atmospheric pressure cold plasma based on dielectric barrier discharge, *Innovat. Food Sci. Emerg. Technol.* 19 (2013) 153–157, <https://doi.org/10.1016/j.ifset.2013.03.001>.
- [113] N.N. Misra, S.K. Pankaj, A. Segat, K. Ishikawa, Cold plasma interactions with enzymes in foods and model systems, *Trends Food Sci. Technol.* 55 (2016) 39–47, <https://doi.org/10.1016/j.tifs.2016.07.001>.
- [114] Y. Han, J.-H. Cheng, D.-W. Sun, Activities and conformation changes of food enzymes induced by cold plasma: a review, *Crit. Rev. Food Sci. Nutr.* 59 (5) (2019) 794–811, <https://doi.org/10.1080/10408398.2018.1555131>.
- [115] M. Gavahian, Y.-H. Chu, A. Mousavi Khaneghah, F.J. Barba, N.N. Misra, A critical analysis of the cold plasma induced lipid oxidation in foods, *Trends Food Sci. Technol.* 77 (2018) 32–41, <https://doi.org/10.1016/j.tifs.2018.04.009>.
- [116] J. Van Durme, A. Nikiforov, J. Vandamme, C. Leys, A. De Winne, Accelerated lipid oxidation using non-thermal plasma technology: evaluation of volatile compounds, *Food Res. Int.* 62 (2014) 868–876, <https://doi.org/10.1016/j.foodres.2014.04.043>.
- [117] P. Kulawik, C. Alvarez, P.J. Cullen, R. Aznar-Roca, A.M. Mullen, B. Tiwari, The effect of non-thermal plasma on the lipid oxidation and microbiological quality of sushi, *Innovat. Food Sci. Emerg. Technol.* 45 (2018) 412–417, <https://doi.org/10.1016/j.ifset.2017.12.011>.
- [118] J. Van der Paal, E.C. Neyts, C.C.W. Verlackt, A. Bogaerts, Effect of lipid peroxidation on membrane permeability of cancer and normal cells subjected to oxidative stress, *Chem. Sci.* 7 (1) (2016) 489–498, <https://doi.org/10.1039/C5SC02311D>.
- [119] M. Leduc, D. Guay, R.L. Leask, S. Coulombe, Cell permeabilization using a non-thermal plasma, *New J. Phys.* 11 (11) (2009) 115021, <https://doi.org/10.1088/1367-2630/11/11/115021>.
- [120] G.O. Fruhwirth, A. Loidl, A. Hermetter, Oxidized phospholipids: from molecular properties to disease, *Biochim. Biophys. Acta (BBA) - Mol. Basis Dis.* 1772 (7) (2007) 718–736, <https://doi.org/10.1016/j.bbadis.2007.04.009>.
- [121] A. Reis, C.M. Spickett, Chemistry of phospholipid oxidation, *Biochim. Biophys. Acta* 1818 (10) (2012) 2374–2387, <https://doi.org/10.1016/j.bbamem.2012.02.002>.
- [122] C.M. Spickett, A.R. Pitt, Oxidative lipidomics coming of age: advances in analysis of oxidized phospholipids in physiology and pathology, *Antioxidants Redox Signal.* 22 (18) (2015) 1646–1666, <https://doi.org/10.1089/ars.2014.6098>.
- [123] A.M. Hirst, F.M. Frame, N.J. Maitland, D. O'Connell, Low temperature plasma causes double-strand break DNA damage in primary epithelial cells cultured from a human prostate tumour, *IEEE Trans Plasma Sci IEEE Nucl Plasma Sci Soc* 42 (10) (2014) 2740–2741, <https://doi.org/10.1109/TPS.2014.2351453>.
- [124] M. Dezest, L. Chavatte, M. Bourdens, D. Quinton, M. Camus, L. Garrigues, P. Descargues, S. Arbault, O. Burlet-Schiltz, L. Casteilla, F. Clement, V. Planat, A. L. Bulteau, Mechanistic insights into the impact of Cold Atmospheric Pressure Plasma on human epithelial cell lines, *Sci. Rep.* 7 (2017) 41163, <https://doi.org/10.1038/srep41163>.
- [125] E.J. Szili, N. Gaur, S.-H. Hong, H. Kurita, J.-S. Oh, M. Ito, A. Mizuno, A. Hatta, A. J. Cowin, D.B. Graves, R.D. Short, The assessment of cold atmospheric plasma treatment of DNA in synthetic models of tissue fluid, tissue and cells, *J. Phys. Appl. Phys.* 50 (27) (2017) 274001, <https://doi.org/10.1088/1361-6463/aa7501>.
- [126] E.J. Szili, F.J. Harding, S.-H. Hong, F. Herrmann, N.H. Voelcker, R.D. Short, The hormesis effect of plasma-elevated intracellular ROS on HaCaT cells, *J. Phys. Appl. Phys.* 48 (2015), <https://doi.org/10.1088/0022-3727/48/49/495401>.
- [127] M. Wang, B. Holmes, X. Cheng, W. Zhu, M. Keidar, L.G. Zhang, Cold atmospheric plasma for selectively ablating metastatic breast cancer cells, *PLoS One* 8 (9) (2013), e73741, <https://doi.org/10.1371/journal.pone.0073741>.
- [128] J. Duan, X. Lu, G. He, The selective effect of plasma activated medium in an in vitro co-culture of liver cancer and normal cells, *J. Appl. Phys.* 121 (1) (2017), 013302, <https://doi.org/10.1063/1.4973484>.
- [129] G. Valacchi, C. Sticozzi, A. Pecorelli, F. Cervellati, C. Cervellati, E. Maioli, Cutaneous responses to environmental stressors, *Ann. N. Y. Acad. Sci.* 1271 (1) (2012) 75–81, <https://doi.org/10.1111/j.1749-6632.2012.06724.x>.
- [130] A. Phaniendra, D.B. Jestadi, L. Periyasamy, Free radicals: properties, sources, targets, and their implication in various diseases, *Indian J. Clin. Biochem.* 30 (1) (2015) 11–26, <https://doi.org/10.1007/s12291-014-0446-0>.
- [131] A.N. Onyango, Endogenous generation of singlet oxygen and ozone in human and animal tissues: mechanisms, biological significance, and influence of dietary components, *Oxid Med Cell Longev* 2016 (2016) 2398573, <https://doi.org/10.1155/2016/2398573>.
- [132] A.A. Starkov, The role of mitochondria in reactive oxygen species metabolism and signaling, *Ann. N. Y. Acad. Sci.* 1147 (2008) 37–52, <https://doi.org/10.1196/annals.1427.015>.
- [133] A.C. Celsus, W.G.W.G. Spencer, *De Medicina*, Harvard university press, London, 1935 (W. Heinemann, Ltd: Cambridge, Mass).
- [134] J. Yun, T. Finkel, Mitohormesis, *Cell Metabol.* 19 (5) (2014) 757–766, <https://doi.org/10.1016/j.cmet.2014.01.011>.
- [135] M. Ristow, K. Schmeisser, Mitohormesis: promoting health and lifespan by increased levels of reactive oxygen species (ROS), *Dose-Response* 12 (2) (2014) 288–341, <https://doi.org/10.2203/dose-response.13-035>. Ristow.
- [136] T. Finkel, Signal transduction by reactive oxygen species, *J. Cell Biol.* 194 (1) (2011) 7–15, <https://doi.org/10.1083/jcb.201102095>.
- [137] P.D. Ray, B.-W. Huang, Y. Tsuji, Reactive oxygen species (ROS) homeostasis and redox regulation in cellular signaling, *Cell. Signal.* 24 (5) (2012) 981–990, <https://doi.org/10.1016/j.cellsig.2012.01.008>.
- [138] C.R. Reczek, N.S. Chandel, ROS-dependent signal transduction, *Curr. Opin. Cell Biol.* 33 (2015) 8–13, <https://doi.org/10.1016/j.cob.2014.09.010>.
- [139] J.E. Le Belle, N.M. Orozco, A.A. Paucar, J.P. Saxe, J. Mottahefeh, A.D. Pyle, H. Wu, H.I. Kornblum, Proliferative neural stem cells have high endogenous ROS levels that regulate self-renewal and neurogenesis in a PI3K/Akt-dependant manner, *Cell Stem Cell* 8 (1) (2011) 59–71, <https://doi.org/10.1016/j.stem.2010.11.028>.
- [140] X. Chen, M. Song, B. Zhang, Y. Zhang, Reactive oxygen species regulate T cell immune response in the tumor microenvironment, *Oxidative Medicine and Cellular Longevity* (2016) 10, <https://doi.org/10.1155/2016/1580967>, 2016.
- [141] C. Kohchi, H. Inagawa, T. Nishizawa, G. Soma, ROS and innate immunity, *Anticancer Res.* 29 (3) (2009) 817–821.
- [142] U. auf dem Keller, A. Kumin, S. Braun, S. Werner, Reactive oxygen species and their detoxification in healing skin wounds, *J. Invest. Dermatol. Symp. Proc.* 11 (1) (2006) 106–111, <https://doi.org/10.1038/sj.jidsymp.5650001>.
- [143] D.A. Wink, J.B. Mitchell, Chemical biology of nitric oxide: insights into regulatory, cytotoxic, and cytoprotective mechanisms of nitric oxide, *Free Radic. Biol. Med.* 25 (4) (1998) 434–456, [https://doi.org/10.1016/S0891-5849\(98\)00092-6](https://doi.org/10.1016/S0891-5849(98)00092-6).
- [144] N.R. Prabhakar, NO and CO as second messengers in oxygen sensing in the carotid body, *Respir. Physiol.* 115 (2) (1999) 161–168, [https://doi.org/10.1016/S0034-5687\(99\)00019-5](https://doi.org/10.1016/S0034-5687(99)00019-5).
- [145] K. Chen, R.N. Pittman, A.S. Popel, Nitric oxide in the vasculature: where does it come from and where does it go? A quantitative perspective, *Antioxidants Redox Signal.* 10 (7) (2008) 1185–1198, <https://doi.org/10.1089/ars.2007.1959>.
- [146] A. Carreau, C. Kieda, C. Grillon, Nitric oxide modulates the expression of endothelial cell adhesion molecules involved in angiogenesis and leukocyte recruitment, *Exp. Cell Res.* 317 (1) (2011) 29–41, <https://doi.org/10.1016/j.yexcr.2010.08.011>.
- [147] P. Tripathi, P. Tripathi, L. Kashyap, V. Singh, The role of nitric oxide in inflammatory reactions, *Pathogens and Disease* 51 (3) (2007) 443–452, <https://doi.org/10.1111/j.1574-695X.2007.00329.x>.
- [148] Y. Dong, H. Wang, J. Cao, J. Ren, R. Fan, X. He, G.W. Smith, C. Dong, Nitric oxide enhances melanogenesis of alpaca skin melanocytes in vitro by activating the MITF phosphorylation, *Mol. Cell. Biochem.* 352 (1) (2011) 255–260, <https://doi.org/10.1007/s11010-011-0761-1>.
- [149] J. Liebmann, J. Scherer, N. Bibinov, P. Rajasekaran, R. Kovacs, R. Gesche, P. Awakowicz, V. Kolb-Bachofen, Biological effects of nitric oxide generated by an atmospheric pressure gas-plasma on human skin cells, *Nitric Oxide* 24 (1) (2011) 8–16, <https://doi.org/10.1016/j.niox.2010.09.005>.
- [150] K. Heuer, M.A. Hoffmanns, E. Demir, S. Baldus, C.M. Volkmar, M. Rohle, P. C. Fuchs, P. Awakowicz, C.V. Suschek, C. Oplander, The topical use of non-thermal dielectric barrier discharge (DBD): nitric oxide related effects on human skin, *Nitric Oxide* 44 (2015) 52–60, <https://doi.org/10.1016/j.niox.2014.11.015>.
- [151] A.N. Paunel, A. Dejam, S. Thelen, M. Kirsch, M. Horstjann, P. Gharini, M. Mürtz, M. Kelm, H. de Groot, V. Kolb-Bachofen, C.V. Suschek, Enzyme-independent nitric oxide formation during UVA challenge of human skin: characterization, molecular sources, and mechanisms, *Free Radic. Biol. Med.* 38 (5) (2005) 606–615, <https://doi.org/10.1016/j.freeradbiomed.2004.11.018>.
- [152] C.V. Suschek, P. Schroeder, O. Aust, H. Sies, C. Mahotka, M. Horstjann, H. Ganser, M. Mürtz, P. Hering, O. Schnorr, K.-D. Kröncke, V. Kolb-Bachofen, The presence

- of nitrite during UVA irradiation protects from apoptosis, *Faseb. J.* 17 (15) (2003) 2342–2344, <https://doi.org/10.1096/fj.03-0359fj>.
- [153] C. Opländer, W. Wetzel, M.M. Cortese, N. Pallua, C.V. Suschek, Evidence for a physiological role of intracellularly occurring photolabile nitrogen oxides in human skin fibroblasts, *Free Radic. Biol. Med.* 44 (9) (2008) 1752–1761, <https://doi.org/10.1016/j.freeradbiomed.2008.01.030>.
- [154] S. Di Meo, T.T. Reed, P. Venditti, V.M. Victor, Role of ROS and RNS sources in physiological and pathological conditions, *Oxidative Medicine and Cellular Longevity* (2016) 1245049, <https://doi.org/10.1155/2016/1245049>, 2016.
- [155] V. Calabrese, C. Cornelius, E. Rizzarelli, J.B. Owen, A.T. Dinkova-Kostova, D. A. Butterfield, Nitric oxide in cell survival: a janus molecule, *Antioxidants Redox Signal.* 11 (11) (2009) 2717–2739, <https://doi.org/10.1089/ars.2009.2721>.
- [156] Q. Zhou, U. Mrowietz, M. Rostami-Yazdi, Oxidative stress in the pathogenesis of psoriasis, *Free Radic. Biol. Med.* 47 (7) (2009) 891–905, <https://doi.org/10.1016/j.freeradbiomed.2009.06.033>.
- [157] D.P. Kadam, A.N. Suryakar, R.D. Ankush, C.Y. Kadam, K.H. Deshpande, Role of oxidative stress in various stages of psoriasis, *Indian J. Clin. Biochem. : Indian J. Clin. Biochem.* 25 (4) (2010) 388–392, <https://doi.org/10.1007/s12291-010-0043-9>.
- [158] S.P. Cannavò, G. Riso, M. Casciaro, E. Di Salvo, S. Gangemi, Oxidative stress involvement in psoriasis: a systematic review, *Free Radic. Res.* 53 (8) (2019) 829–840, <https://doi.org/10.1080/10715762.2019.1648800>.
- [159] S. Pillai, C. Oresajo, J. Hayward, Ultraviolet radiation and skin aging: roles of reactive oxygen species, inflammation and protease activation, and strategies for prevention of inflammation-induced matrix degradation – a review, *Int. J. Cosmet. Sci.* 27 (1) (2005) 17–34, <https://doi.org/10.1111/j.1467-2494.2004.00241.x>.
- [160] M. Rinnerthaler, J. Bischof, M.K. Streubel, A. Trost, K. Richter, Oxidative stress in aging human skin, *Biomolecules* 5 (2) (2015) 545–589, <https://doi.org/10.3390/biom5020545>.
- [161] K. Wende, S. Strassenburg, B. Haertel, M. Harms, S. Holtz, A. Barton, K. Masur, T. von Woedtke, U. Lindequist, Atmospheric pressure plasma jet treatment evokes transient oxidative stress in HaCaT keratinocytes and influences cell physiology, *Cell Biol. Int.* 38 (4) (2014) 412–425, <https://doi.org/10.1002/cbin.10200>.
- [162] A. Schmidt, S. Dietrich, A. Steuer, K.D. Weltmann, T. von Woedtke, K. Masur, K. Wende, Non-thermal plasma activates human keratinocytes by stimulation of antioxidant and phase II pathways, *J. Biol. Chem.* 290 (11) (2015) 6731–6750, <https://doi.org/10.1074/jbc.M114.603555>.
- [163] A. Dzimitrowicz, A. Bielawska-Pohl, P. Jamroz, J. Dora, A. Krawczenko, G. Busco, C. Grillon, C. Kieda, A. Klimczak, D. Terefinko, A. Baszczynska, P. Pohl, Activation of the normal human skin cells by a portable dielectric barrier discharge-based reaction-discharge system of a defined gas temperature, *Plasma Chem. Plasma Process.* 40 (1) (2020) 79–97, <https://doi.org/10.1007/s11090-019-10039-0>.
- [164] J.H. Choi, Y.S. Song, K. Song, H.J. Lee, J.W. Hong, G.C. Kim, Skin renewal activity of non-thermal plasma through the activation of β -catenin in keratinocytes, *Sci. Rep.* 7 (1) (2017) 6146, <https://doi.org/10.1038/s41598-017-06661-7>.
- [165] S. Hasse, T. Duong, O. Hahn, S. Kindler, H.-R. Metelmann, T. von Woedtke, K. Masur, Induction of proliferation of basal epidermal keratinocytes by cold atmospheric-pressure plasma, *Clin. Exp. Dermatol.* (2015) 41, <https://doi.org/10.1111/ced.12735>.
- [166] X. Shi, J. Cai, G. Xu, H. Ren, S. Chen, Z. Chang, J. Liu, C. Huang, G. Zhang, X. Wu, Effect of cold plasma on cell viability and collagen synthesis in cultured murine fibroblasts, *Plasma Sci. Technol.* 18 (4) (2016) 353–359, <https://doi.org/10.1088/1009-0630/18/4/04>.
- [167] M. Bourdens, Y. Jeanson, M. Taurand, N. Juin, A. Carrière, F. Clément, L. Casteilla, A.-L. Bulteau, V. Planat-Bénard, Short exposure to cold atmospheric plasma induces senescence in human skin fibroblasts and adipose mesenchymal stromal cells, *Sci. Rep.* 9 (1) (2019), <https://doi.org/10.1038/s41598-019-45191-2>, 8671–8671.
- [168] J. Balzer, K. Heuer, E. Demir, M.A. Hoffmanns, S. Baldus, P.C. Fuchs, P. Awakowicz, C.V. Suschek, C. Opländer, Non-thermal dielectric barrier discharge (DBD) effects on proliferation and differentiation of human fibroblasts are primary mediated by hydrogen peroxide, *PLoS One* 10 (12) (2015), e0144968, <https://doi.org/10.1371/journal.pone.0144968>.
- [169] J.-S. Oh, X. Strudwick, R.D. Short, K. Ogawa, A. Hatta, H. Furuta, N. Gaur, S.-H. Hong, A.J. Cowin, H. Fukuhara, K. Inoue, M. Ito, C. Charles, R.W. Boswell, J. W. Bradley, D.B. Graves, E.J. Szili, How plasma induced oxidation, oxygenation, and de-oxygenation influences viability of skin cells, *Appl. Phys. Lett.* 109 (20) (2016) 203701, <https://doi.org/10.1063/1.4967880>.
- [170] G. Collet, E. Robert, A. Lenoir, M. Vandamme, T. Darny, S. Dozias, C. Kieda, J. M. Pouvesle, Plasma jet-induced tissue oxygenation: potentialities for new therapeutic strategies, *Plasma Sources Sci. Technol.* 23 (1) (2014), 012005, <https://doi.org/10.1088/0963-0252/23/1/012005>.
- [171] T. Kisch, A. Helmke, S. Schlessner, J. Song, E. Liodaki, F.H. Stang, P. Mailaender, R. Kraemer, Improvement of cutaneous microcirculation by cold atmospheric plasma (CAP): results of a controlled, prospective cohort study, *Microvasc. Res.* 104 (2016) 55–62, <https://doi.org/10.1016/j.mvr.2015.12.002>.
- [172] B. Asadamongkol, J. Zhang, The development of hyperbaric oxygen therapy for skin rejuvenation and treatment of photoaging, *Med. Gas Res.* 4 (2014) 7, <https://doi.org/10.1186/2045-9912-4-7>.
- [173] I. Bentov, M.J. Reed, The effect of aging on the cutaneous microvasculature, *Microvasc. Res.* 100 (2015) 25–31, <https://doi.org/10.1016/j.mvr.2015.04.004>.
- [174] A.J. Donato, I. Eskurza, A.E. Silver, A.S. Levy, G.L. Pierce, P.E. Gates, D.R. Seals, Direct evidence of endothelial oxidative stress with aging in humans, *Circ. Res.* 100 (11) (2007) 1659–1666, <https://doi.org/10.1161/01.RES.0000269183.13937.e8>.
- [175] S. Arndt, P. Unger, M. Berneburg, A.K. Bosserhoff, S. Karrer, Cold atmospheric plasma (CAP) activates angiogenesis-related molecules in skin keratinocytes, fibroblasts and endothelial cells and improves wound angiogenesis in an autocrine and paracrine mode, *J. Dermatol. Sci.* 89 (2) (2018) 181–190, <https://doi.org/10.1016/j.jdermsci.2017.11.008>.
- [176] S. Kalghatgi, G. Friedman, A. Fridman, A.M. Clyne, Endothelial cell proliferation is enhanced by low dose non-thermal plasma through fibroblast growth factor-2 release, *Ann. Biomed. Eng.* 38 (3) (2010) 748–757, <https://doi.org/10.1007/s10439-009-9868-x>.
- [177] H.S. Cui, Y.S. Cho, S.Y. Joo, C.H. Mun, C.H. Seo, J.B. Kim, Wound healing potential of low temperature plasma in human primary epidermal keratinocytes, *Tissue Eng Regen Med* 16 (6) (2019) 585–593, <https://doi.org/10.1007/s13770-019-00215-w>.
- [178] J.P. Cooke, D.W. Losordo, Nitric oxide and angiogenesis, *Circulation* 105 (18) (2002) 2133–2135, <https://doi.org/10.1161/01.CIR.0000014928.45119.73>.
- [179] S. Babaei, K. Teichert-Kuliszewska, Q. Zhang, N. Jones, D.J. Dumont, D. J. Stewart, Angiogenic actions of angiopoietin-1 require endothelium-derived nitric oxide, *Am. J. Pathol.* 162 (6) (2003) 1927–1936, [https://doi.org/10.1016/S0002-9440\(10\)64326-X](https://doi.org/10.1016/S0002-9440(10)64326-X).
- [180] J.A. Stewart Jr., T.A. West, P.A. Lucchesi, Nitric oxide-induced collagen IV expression and angiogenesis: FAK or fiction? Focus on "Collagen IV contributes to nitric oxide-induced angiogenesis of lung endothelial cells, *Am. J. Physiol. Cell Physiol.* 300 (5) (2011) C968–C969, <https://doi.org/10.1152/ajpcell.00059.2011>.
- [181] C. Duchesne, S. Banzet, J.-J. Lataillade, A. Rousseau, N. Frescaline, Cold atmospheric plasma modulates endothelial nitric oxide synthase signalling and enhances burn wound neovascularisation, *J. Pathol.* 249 (3) (2019) 368–380, <https://doi.org/10.1002/path.5323>.
- [182] A. Ring, S. Langer, A. Schaffran, I. Stricker, P. Awakowicz, H.-U. Steinau, J. Hauser, Enhanced neovascularization of dermis substitutes via low-pressure plasma-mediated surface activation, *Burns* 36 (8) (2010) 1222–1227, <https://doi.org/10.1016/j.burns.2010.03.002>.
- [183] B.B.R. Choi, J.H. Choi, J. Ji, K.W. Song, H.J. Lee, G.C. Kim, Increment of growth factors in mouse skin treated with non-thermal plasma, *Int. J. Med. Sci.* 15 (11) (2018) 1203–1209, <https://doi.org/10.7150/ijms.26342>.
- [184] S.U. Kang, Y.S. Kim, Y.E. Kim, J.-K. Park, Y.S. Lee, H.Y. Kang, J.W. Jang, J. B. Ryeo, Y. Lee, Y.S. Shin, C.-H. Kim, Opposite effects of non-thermal plasma on cell migration and collagen production in keloid and normal fibroblasts, *PLoS One* 12 (11) (2017), <https://doi.org/10.1371/journal.pone.0187978>, e0187978.
- [185] X.-F. Wang, Q.-Q. Fang, B. Jia, Y.-Y. Hu, Z.-C. Wang, K.-p. Yan, S.-Y. Yin, Z. Liu, W.-Q. Tan, Potential effect of non-thermal plasma for the inhibition of scar formation: a preliminary report, *Sci. Rep.* 10 (1) (2020) 1064, <https://doi.org/10.1038/s41598-020-57703-6>.
- [186] Z. Chen, L. Lin, X. Cheng, E. Gjika, M. Keidar, Effects of cold atmospheric plasma generated in deionized water in cell cancer therapy, *Plasma Process. Polym.* 13 (12) (2016) 1151–1156, <https://doi.org/10.1002/ppap.201600086>.
- [187] J.-L. Brisset, B. Benstaali, D. Moussa, J. Fanmoe, E. Njoyim-Tamungang, Acidity control of plasma-chemical oxidation: applications to dye removal, urban waste abatement and microbial inactivation, *Plasma Sources Sci. Technol.* 20 (3) (2011), 034021, <https://doi.org/10.1088/0963-0252/20/3/034021>.
- [188] G. Busco, A. Valinaj Orman, L. Ridou, J.M. Pouvesle, E. Robert, C. Grillon, Cold atmospheric plasma-induced acidification of tissue surface: visualization and quantification using agarose gel models, *J. Phys. Appl. Phys.* (2019), <https://doi.org/10.1088/1361-6463/ab1119>.
- [189] B. Benstaali, D. Moussa, A. Addou, J.-L. Brisset, Plasma treatment of aqueous solutes: some chemical properties of a gliding arc in humid air, *Eur. Phys. J. AP* 4 (2) (1998) 171–179, <https://doi.org/10.1051/epjap:1998258>.
- [190] A. Helmke, D. Hoffmeister, N. Mertens, S. Emmert, J. Schuette, W. Vioel, The acidification of lipid film surfaces by non-thermal DBD at atmospheric pressure in air, *New J. Phys.* 11 (11) (2009) 115025, <https://doi.org/10.1088/1367-2630/11/11/115025>.
- [191] L.A. Schneider, A. Korber, S. Grabbe, J. Dissemund, Influence of pH on wound-healing: a new perspective for wound-therapy? *Arch. Dermatol. Res.* 298 (9) (2007) 413–420, <https://doi.org/10.1007/s00403-006-0713-x>.
- [192] T. Soleymani, J. Lanoue, Z. Rahman, A practical approach to chemical peels: a review of fundamentals and step-by-step algorithmic protocol for treatment, *The Journal of clinical and aesthetic dermatology* 11 (8) (2018) 21–28.
- [193] E.-H. Choi, M.-Q. Man, P. Xu, S. Xin, Z. Liu, D.A. Crumrine, Y.J. Jiang, J.W. Fluhr, K.R. Feingold, P.M. Elias, T.M. Mauro, Stratum corneum acidification is impaired in moderately aged human and murine skin, *J. Invest. Dermatol.* 127 (12) (2007) 2847–2856, <https://doi.org/10.1038/sj.jid.5700913>.
- [194] M. Laroussi, D.A. Mendis, M. Rosenberg, Plasma interaction with microbes, *New J. Phys.* 5 (2003), <https://doi.org/10.1088/1367-2630/5/1/341>, 41–41.
- [195] L.F. Gaunt, C.B. Beggs, G.E. Georghiou, Bactericidal action of the reactive species produced by gas-discharge nonthermal plasma at atmospheric pressure: a review, *IEEE Trans. Plasma Sci.* 34 (4) (2006) 1257–1269, <https://doi.org/10.1109/TPS.2006.878381>.
- [196] M. Moreau, N. Orange, M.G.J. Feuillolay, Non-thermal plasma technologies: new tools for bio-decontamination, *Biotechnol. Adv.* 26 (6) (2008) 610–617, <https://doi.org/10.1016/j.biotechadv.2008.08.001>.
- [197] J. Guo, K. Huang, J. Wang, Bactericidal effect of various non-thermal plasma agents and the influence of experimental conditions in microbial inactivation: a

- review, *Food Contr.* 50 (2015) 482–490, <https://doi.org/10.1016/j.foodcont.2014.09.037>.
- [198] V. Boxhammer, G.E. Morfill, J.R. Jokipii, T. Shimizu, T. Klämpfl, Y.F. Li, J. Körtzer, J. Schlegel, J.L. Zimmermann, Bactericidal action of cold atmospheric plasma in solution, *New J. Phys.* 14 (11) (2012), 113042, <https://doi.org/10.1088/1367-2630/14/11/113042>.
- [199] R. Zhou, R. Zhou, K. Prasad, Z. Fang, R. Speight, K. Bazaka, K. Ostrikov, Cold atmospheric plasma activated water as a prospective disinfectant: the crucial role of peroxydinitrate, *Green Chem.* 20 (23) (2018) 5276–5284, <https://doi.org/10.1039/C8GC02800A>.
- [200] S. Ikawa, A. Tani, Y. Nakashima, K. Kitano, Physicochemical properties of bactericidal plasma-treated water, *J. Phys. Appl. Phys.* 49 (42) (2016) 425401, <https://doi.org/10.1088/0022-3727/49/42/425401>.
- [201] M. Müller, T. Shimizu, S. Binder, P. Rettberg, J.L. Zimmermann, G.E. Morfill, H. Thomas, Plasma afterglow circulation apparatus for decontamination of spacecraft equipment, *AIP Adv.* 8 (10) (2018) 105013, <https://doi.org/10.1063/1.5040303>.
- [202] K. Fricke, I. Koban, H. Tresp, L. Jablonowski, K. Schröder, A. Kramer, K.-D. Weltmann, T. von Woedtke, T. Kocher, Atmospheric pressure plasma: a high-performance tool for the efficient removal of biofilms, *PLoS One* 7 (8) (2012), e42539, <https://doi.org/10.1371/journal.pone.0042539>.
- [203] P. Brun, G. Bernabè, C. Marchiori, M. Scarpa, M. Zuin, R. Cavazzana, B. Zaniol, E. Martines, Antibacterial efficacy and mechanisms of action of low power atmospheric pressure cold plasma: membrane permeability, biofilm penetration and antimicrobial sensitization, *J. Appl. Microbiol.* 125 (2) (2018) 398–408, <https://doi.org/10.1111/jam.13780>.
- [204] Y.W. Kim, S. Subramanian, K. Gerasopoulos, H. Ben-Yoav, H.-C. Wu, D. Quan, K. Carter, M.T. Meyer, W.E. Bentley, R. Ghodssi, Effect of electrical energy on the efficacy of biofilm treatment using the bioelectric effect, *npj Biofilms and Microbiomes* 1 (1) (2015) 15016, <https://doi.org/10.1038/npjbiofilms.2015.16>.
- [205] S.A. Ermolaeva, A.F. Varfolomeev, M.Y. Chernukha, D.S. Yurov, M.M. Vasiliev, A. A. Kaminskaya, M.M. Moiseyevich, J.M. Romanova, A.N. Murashev, I. I. Selezneva, T. Shimizu, E.V. Sysolyatina, I.A. Shaginyan, O.F. Petrov, E. I. Mayevsky, V.E. Fortov, G.E. Morfill, B.S. Naroditsky, A.L. Gintsburg, Bactericidal effects of non-thermal argon plasma in vitro, in biofilms and in the animal model of infected wounds, *J. Med. Microbiol.* 60 (1) (2011) 75–83, <https://doi.org/10.1099/jmm.0.020263-0>.
- [206] G. Isbary, T. Shimizu, J.L. Zimmermann, H.M. Thomas, G.E. Morfill, W. Stolz, Cold atmospheric plasma for local infection control and subsequent pain reduction in a patient with chronic post-operative ear infection, *New Microbes and New Infections* 1 (3) (2013) 41–43, <https://doi.org/10.1002/2052-2975.19>.
- [207] B.F. Gilmore, P.B. Flynn, S. O'Brien, N. Hickok, T. Freeman, P. Bourke, Cold plasmas for biofilm control: opportunities and challenges, *Trends Biotechnol.* 36 (6) (2018) 627–638, <https://doi.org/10.1016/j.tibtech.2018.03.007>.
- [208] Y.E. Chen, M.A. Fischbach, Y. Belkaid, Skin microbiota-host interactions, *Nature* 553 (7689) (2018) 427–436, <https://doi.org/10.1038/nature25177>.
- [209] K. Oehmigen, M. Hähnel, R. Brandenburg, C. Wilke, K.D. Weltmann, T. von Woedtke, The role of acidification for antimicrobial activity of atmospheric pressure plasma in liquids, *Plasma Process. Polym.* 7 (3–4) (2010) 250–257, <https://doi.org/10.1002/ppap.200900077>.
- [210] Q. Zhang, R. Ma, Y. Tian, B. Su, K. Wang, S. Yu, J. Zhang, J. Fang, Sterilization efficiency of a novel electrochemical disinfectant against *Staphylococcus aureus*, *Environ. Sci. Technol.* 50 (6) (2016) 3184–3192, <https://doi.org/10.1021/acs.est.5b05108>.
- [211] E.M. Jones, C.A. Cochrane, S.L. Percival, The effect of pH on the extracellular matrix and biofilms, *Adv. Wound Care* 4 (7) (2015) 431–439, <https://doi.org/10.1089/wound.2014.0538>.
- [212] G.A. Konesky, *COLD PLASMAJET HAND SANITIZER* C. Bovie Medical Corporation, FL (US) Editor, 2013. US 2013/0202496 A1: United States.
- [213] A. Ali, Y.H. Kim, J.Y. Lee, S. Lee, H.S. Uhm, G. Cho, B.J. Park, E.H. Choi, Inactivation of *Propionibacterium acnes* and its biofilm by non-thermal plasma, *Curr. Appl. Phys.* 14 (2014) S142–S148, <https://doi.org/10.1016/j.cap.2013.12.034>.
- [214] F. Woodland, F. Osolin, J.-Y. Legendre, G. Vic, in: *COSMETIC USE OF A COLD PLASMA*, L'OREAL, France, 2014.
- [215] Z. Xiong, J. Roe, T.C. Grammer, D.B. Graves, Plasma treatment of onychomycosis, *Plasma Process. Polym.* 13 (6) (2016) 588–597, <https://doi.org/10.1002/ppap.201600010>.
- [216] S. Verdier-Sévrain, F. Bonté, Skin hydration: a review on its molecular mechanisms, *J. Cosmet. Dermatol.* 6 (2) (2007) 75–82, <https://doi.org/10.1111/j.1473-2165.2007.00300.x>.
- [217] L. Cau, V. Pendaries, E. Lhuillier, P.R. Thompson, G. Serre, H. Takahara, M.-C. Méchin, M. Simon, Lowering relative humidity level increases epidermal protein deimination and drives human filaggrin breakdown, *J. Dermatol. Sci.* 86 (2) (2017) 106–113, <https://doi.org/10.1016/j.jdermsci.2017.02.280>.
- [218] A.V. Rawlings, C.R. Harding, Moisturization and skin barrier function, *Dermatol. Ther.* 17 (s1) (2004) 43–48, <https://doi.org/10.1111/j.1396-0296.2004.04S1005.x>.
- [219] E.H. Mojumdar, Q.D. Pham, D. Topgaard, E. Sparr, Skin hydration: interplay between molecular dynamics, structure and water uptake in the stratum corneum, *Sci. Rep.* 7 (1) (2017), <https://doi.org/10.1038/s41598-017-15921-5>, 15712–15712.
- [220] M.J. Choi, H.I. Maibach, Role of ceramides in barrier function of healthy and diseased skin, *Am. J. Clin. Dermatol.* 6 (4) (2005) 215–223, <https://doi.org/10.2165/00128071-200506040-00002>.
- [221] J.W. Fluhr, S. Sassning, O. Lademann, M.E. Darvin, S. Schanzer, A. Kramer, H. Richter, W. Sterry, J. Lademann, In vivo skin treatment with tissue-tolerable plasma influences skin physiology and antioxidant profile in human stratum corneum, *Exp. Dermatol.* 21 (2) (2012) 130–134, <https://doi.org/10.1111/j.1600-0625.2011.01411.x>.
- [222] K.W. Foster, R.L. Moy, E.F. Fincher, Advances in plasma skin regeneration, *J. Cosmet. Dermatol.* 7 (3) (2008) 169–179, <https://doi.org/10.1111/j.1473-2165.2008.00385.x>.
- [223] D.K. Athanasopoulos, P. Svarnas, A. Gerakis, Cold plasma bullet influence on the water contact angle of human skin surface, *J. Electrostat.* 102 (2019) 103378, <https://doi.org/10.1016/j.elstat.2019.103378>.
- [224] C. Kaemling, A. Kaemling, S. Tümmel, W. Viöl, Plasma treatment on finger nails prior to coating with a varnish, *Surf. Coating. Technol.* 200 (1) (2005) 668–671, <https://doi.org/10.1016/j.surfcoat.2005.01.065>.
- [225] J.-S. Oh, E.J. Szili, G. Nishtha, S.-H. Hong, H. Furuta, H. Kurita, A. Mizuno, A. Hatta, R.D. Short, How to assess the plasma delivery of RONS into tissue fluid and tissue, *J. Phys. Appl. Phys.* 49 (30) (2016) 304005, <https://doi.org/10.1088/0022-3727/49/30/304005>.
- [226] L. Nie, Y. Yang, J. Duan, F. Sun, X. Lu, G. He, Effect of tissue thickness and liquid composition on the penetration of long-lifetime reactive oxygen and nitrogen species (RONS) generated by a plasma jet, *J. Phys. Appl. Phys.* 51 (34) (2018) 345204, <https://doi.org/10.1088/1361-6463/aad427>.
- [227] M. Boury-Jamot, J. Daraspe, F. Bonte, E. Perrier, S. Schnebert, M. Dumas, J. M. Verbavatz, Skin aquaporins: function in hydration, wound healing, and skin epidermis homeostasis, *Handb. Exp. Pharmacol.* (190) (2009) 205–217, https://doi.org/10.1007/978-3-540-79885-9_10.
- [228] R. Patel, L. Kevin Heard, X. Chen, W.B. Bollag, in: *Aquaporins*, B. Yang (Eds.), *Aquaporins in the Skin*, Springer Netherlands, Dordrecht, 2017, pp. 173–191, https://doi.org/10.1007/978-94-024-1057-0_11.
- [229] Y. Dayun, X. Haijie, Z. Wei, N. Niki, Z. Lijie Grace, B. Ka, K. Michael, The role of aquaporins in the anti-glioblastoma capacity of the cold plasma-stimulated medium, *J. Phys. Appl. Phys.* 50 (5) (2017), 055401, <https://doi.org/10.1088/1361-6463/aa53d6>.
- [230] M. Yusupov, J. Razzokov, R.M. Cordeiro, A. Bogaerts, Transport of reactive oxygen and nitrogen species across aquaporin: a molecular level picture, *Oxidative medicine and cellular longevity* (2019), <https://doi.org/10.1155/2019/2930504>, 2019, 2930504–2930504.
- [231] M. Yusupov, J. Van der Paal, E.C. Neyts, A. Bogaerts, Synergistic effect of electric field and lipid oxidation on the permeability of cell membranes, *Biochim. Biophys. Acta Gen. Subj.* 1861 (4) (2017) 839–847, <https://doi.org/10.1016/j.bbagen.2017.01.030>.
- [232] V. Vijayarangan, A. Delalande, S. Dozias, J. Pouvesle, C. Pichon, E. Robert, Cold atmospheric plasma parameters investigation for efficient drug delivery in HeLa cells, *IEEE Transactions on Radiation and Plasma Medical Sciences* 2 (2) (2018) 109–115, <https://doi.org/10.1109/TRPMS.2017.2759322>.
- [233] J. Lademann, H. Richter, A. Kramer, O. Lademann, Enhancement of the Penetration of Topically Applied Substances by Tissue-Tolerable Plasma, 2018, pp. 441–447, https://doi.org/10.1007/978-3-319-67627-2_27.
- [234] M. Gelker, C.C. Müller-Goymann, W. Viöl, Permeabilization of human stratum corneum and full-thickness skin samples by a direct dielectric barrier discharge, *Clinical Plasma Medicine* 9 (2018) 34–40, <https://doi.org/10.1016/j.cpm.2018.02.001>.
- [235] K. Shimizu, K. Hayashida, M. Blajan, Novel method to improve transdermal drug delivery by atmospheric microplasma irradiation, *Biointerphases* 10 (2) (2015), <https://doi.org/10.1116/1.4919708>, 029517.
- [236] H.Y. Lee, J.H. Choi, J.W. Hong, G.C. Kim, H.-J. Lee, Comparative study of the Ar and He atmospheric pressure plasmas on E-cadherin protein regulation for plasma-mediated transdermal drug delivery, *J. Phys. Appl. Phys.* 51 (21) (2018) 215401, <https://doi.org/10.1088/1361-6463/aab8dc>.
- [237] T. Darny, J.M. Pouvesle, V. Puech, C. Douat, S. Dozias, R. Eric, Analysis of conductive target influence in plasma jet experiments through helium metastable and electric field measurements, *Plasma Sources Sci. Technol.* 26 (4) (2017), <https://doi.org/10.1088/1361-6595/aa5b15>, 045008.
- [238] O. Guaitella, A. Sobota, The impingement of a kHz helium atmospheric pressure plasma jet on a dielectric surface, *J. Phys. Appl. Phys.* 48 (25) (2015) 255202, <https://doi.org/10.1088/0022-3727/48/25/255202>.
- [239] K. Aboubakar, F.P. Saint, C. Muja, B. Caillier, P. Guillot, Investigation of the interaction between a helium plasma jet and conductive (Metal)/Non-Conductive (dielectric) targets, *Plasma Med.* 7 (2018), <https://doi.org/10.1615/PlasmaMed.2018019503>.
- [240] A. Stancampiano, T. Chung, S. Dozias, J. Pouvesle, L.M. Mir, E. Robert, Mimicking of human body electrical characteristic for easier translation of plasma biomedical studies to clinical applications, *IEEE Transactions on Radiation and Plasma Medical Sciences* 4 (3) (2020) 335–342, <https://doi.org/10.1109/TRPMS.2019.2936667>.
- [241] D.B. Graves, Lessons from tesla for plasma medicine, *IEEE Transactions on Radiation and Plasma Medical Sciences* 2 (6) (2018) 594–607, <https://doi.org/10.1109/TRPMS.2018.2866373>.
- [242] S. Snyder, C. DeJulius, R.K. Willits, Electrical stimulation increases random migration of human dermal fibroblasts, *Ann. Biomed. Eng.* 45 (9) (2017) 2049–2060, <https://doi.org/10.1007/s10439-017-1849-x>.
- [243] M. Rouabha, H. Park, S. Meng, H. Derbali, Z. Zhang, Electrical stimulation promotes wound healing by enhancing dermal fibroblast activity and promoting myofibroblast transdifferentiation, *PLoS One* 8 (8) (2013), e71660, <https://doi.org/10.1371/journal.pone.0071660>.

- [244] J. Dube, O. Rochette-Drouin, P. Levesque, R. Gauvin, C.J. Roberge, F.A. Auger, D. Goulet, M. Bourdages, M. Plante, V.J. Moulin, L. Germain, Human keratinocytes respond to direct current stimulation by increasing intracellular calcium: preferential response of poorly differentiated cells, *J. Cell. Physiol.* 227 (6) (2012) 2660–2667, <https://doi.org/10.1002/jcp.23008>.
- [245] A. Golberg, S. Khan, V. Belov, K.P. Quinn, H. Albadawi, G. Felix Broelsch, M. T. Watkins, I. Georgakoudi, M. Papisov, M.C. Mihm Jr., W.G. Austen Jr., M. L. Yarmush, Skin rejuvenation with non-invasive pulsed electric fields, *Sci. Rep.* 5 (2015) 10187, <https://doi.org/10.1038/srep10187>.
- [246] F. Judée, T. Dufour, Plasma gun for medical applications: engineering an equivalent electrical target of the human body and deciphering relevant electrical parameters, *J. Phys. Appl. Phys.* 52 (16) (2019), 16LT02, <https://doi.org/10.1088/1361-6463/ab03b8>.
- [247] A. Scavone, M.J. Kadziolka, C.J. Miller, State and trait mindfulness as predictors of skin conductance response to stress, *Appl. Psychophysiol. Biofeedback* (2020), <https://doi.org/10.1007/s10484-020-09467-y>.
- [248] M.E. Dawson, A.M. Schell, D.L. Filion, The electrodermal system, in: G. Berntson, J.T. Cacioppo, L.G. Tassinary (Eds.), *Handbook of Psychophysiology*, Cambridge University Press, Cambridge, 2016, pp. 217–243, <https://doi.org/10.1017/9781107415782.010>.
- [249] D. Gidon, D.B. Graves, A. Mesbah, Effective dose delivery in atmospheric pressure plasma jets for plasma medicine: a model predictive control approach, *Plasma Sources Sci. Technol.* 26 (8) (2017), 085005, <https://doi.org/10.1088/1361-6595/aa7c5d>.
- [250] P.-M. Girard, A. Arbabian, M. Fleury, G. Bauville, V. Puech, M. Dutreix, J. S. Sousa, Synergistic effect of H₂O₂ and NO₂ in cell death induced by cold atmospheric He plasma, *Sci. Rep.* 6 (1) (2016) 29098, <https://doi.org/10.1038/srep29098>.
- [251] T. Bernhardt, M.L. Semmler, M. Schäfer, S. Bekeschus, S. Emmert, L. Boeckmann, Plasma medicine: applications of cold atmospheric pressure plasma in dermatology, *Oxidative medicine and cellular longevity* (2019), <https://doi.org/10.1155/2019/3873928>, 2019, 3873928–3873928.