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Natural products originated from the oxidative coupling of tyrosine and tryptophan: biosynthesis and bioinspired synthesis

Shanyu Tang,^[a] and Guillaume Vincent^{*[a]}

Dedication ((optional))

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Abstract: The oxidative coupling of tyrosine and tryptophan units is a pivotal step in the total synthesis of some peptide-derived marine and terrestrial natural products such as the diazonamides, azonazine and tryptorubin A. This review details the biosynthesis and bioinspired synthesis of natural products with such structures. Through this review, we focused on the challenges of the synthesis of these natural products and the innovative solutions adopted by synthetic chemists.

1. Introduction

Amino acid-derived natural products are widespread in nature. Tyrosine and tryptophan are common amino acids and therefore usually appear in the structure of natural peptides. Apart to classical amide bonds, few of these natural products also display an additional linkage between the phenol part of a tyrosine (**1a**) and the indole part of a tryptophan (**2a**, Figure 1). Biosynthetically,

these additional bonds arise from an oxidative cyclization.^[1] They could be produced by marine organisms or by terrestrial fungus.

The synthesis of such molecules has indeed attracted widespread attention from synthetic chemists, not only because of the challenging structure, but also for the biological activity of some of these compounds.

Understanding their biosynthesis is important to help chemists designing simplified and efficient synthetic routes towards these natural products.

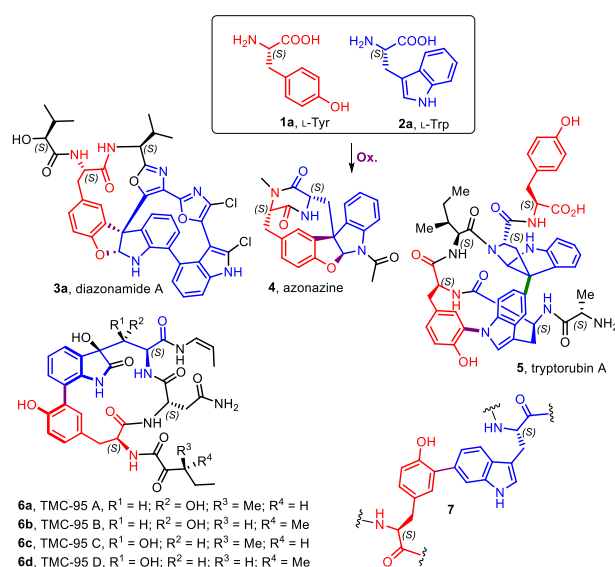
Therefore, we will review the biosynthesis of diazonamide A (**3a**), azonazine (**4**) and tryptorubin A (**5**) and their related bioinspired syntheses via oxidative couplings of tyrosine and tryptophan units involving the formation of a carbon-heteroatom bond.

Besides, the natural products presented in this minireview, it should be noted that other natural entities present a link between the benzene part of the indole of a tryptophan and the *ortho*-position of the phenol of tyrosine. It is the case of TMC-95 A-D^[2,3] (**6a-d**, produced by the fermentation broth of *Apioispora montagnei* Sacc Tc. 1093 isolated from soil samples) and of some proteins **7** that are composed of covalently crosslinked amino acid side-chains.^[4]

Shanyu Tang obtained her master degree in organic chemistry in 2019 under the supervision of Prof. Guanyu Yang from Zhengzhou University, China. She is currently working as a PhD student under the supervision of Dr. Guillaume Vincent at Université Paris-Saclay. Her research interests are focused on electrochemical synthesis.



Guillaume Vincent graduated from CPE Lyon (2002) with a one-year internship at Dupont Pharma with Dr. Patrick Y. S. Lam. He obtained a Master (2002) and a PhD (2005) degrees from Université Lyon-1 with Prof. Marco A. Ciufolini. After two postdoctoral experiences with Prof. Robert M. Williams at Colorado State University and with Prof. Louis Fensterbank and Prof. Max Malacria at Université Pierre et Marie Curie Paris-6, he was recruited as a CNRS researcher in 2007 at the Institut de Chimie Moléculaire et des Matériaux d'Orsay (ICMMO) of Université Paris-Sud which is now Université Paris-Saclay.



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Figure 1. Natural products biosynthetically derived from the oxidative coupling of L-tyrosine and L-tryptophan residues.

In the context of the total synthesis of the TMC-95 natural products, the C-C biaryl bond linking the tyrosine and tryptophan units are usually formed by a Suzuki coupling in the context of total synthesis.^[3]

2. Diazonamide A

Diazoamide A is a marine secondary metabolite which displays low nanomolar cytotoxicity against several tumoral cell lines. It was isolated in 1991 by Fenical from the ascidian *Diazona angulata* (originally attributed to *Diazona chinensis*) along with diazoamide B.^[6] Their original structures were determined by X-ray crystallography of a derivative of diazoamide B. The proposed structure **8** of diazoamide A included notably a linkage between the phenol of a tyrosine and a hydroxydihydrobenzofurane, as well as a sequence of two oxazoles and an indole (Figure 2). The antitumoral potential and the rare and particular structure of diazoamide A has attracted the wide attention of synthetic chemists. However, in 2001, when the group of Harran completed the first total synthesis of the proposed structure,^[6] it was found to be different from the natural product. After reinterpretation of the X-ray crystallography data and additional 2D-NMR experiments, Harran proposed the alternative structures **3a** and **3b** of diazoamides A and B, in which the difference with the original scaffold is the presence of a central benzofuroindoline instead of the 3-aryl-2-hydroxydihydrobenzofurane.^[7] The benzofuroindoline moiety is also found in other natural products such as bipheophylline, pleiocraline or pleiocorine.^[8,9] Moreover Diazoamide A possesses a hydroxyisovaleric acid unit instead of a valine at one end of the molecule. In 2008, Reyes isolated, from *Diazona* sp., diazoamides C-E (**3c-3e**) which possess the same backbone of diazoamides A and B.^[10]

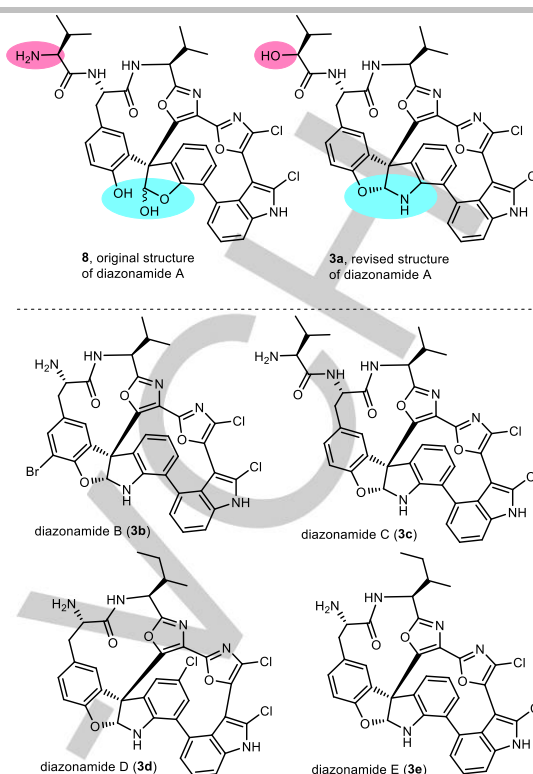


Figure 2. Structures of diazoamides A-E (**3a-e**).

After the correction of the general structure of the diazoamides, it seems plausible that they have for biosynthetic origin the assembly of four amino-acids: one tyrosine (**1a**), two tryptophanes (**1a**) and one valine (**9a**) as well as a (*S*)- α -hydroxyisovaleric acid **10** (Figure 2).

The corresponding L-Tyr-L-Val-L-Trp-L-Trp tetrapeptide **11** would be involved in several oxidation events involving the tryptophan units: oxidation at the α -position of the indoles to form the two oxazoles and formation of the biaryl bond between the two indoles. Importantly, the oxidative macrocyclization between the tyrosine and one of the tryptophan would form the benzofuroindoline core **12** via formation of a C-C bond between the *ortho*-position of the phenol and the C3-position of the indole, as well as a C-O bond between the oxygen of the phenol and the C2-position of the indole. This reaction involves the union of two nucleophilic entities and from a mechanistic point of view several scenarios are possible: either the phenol or the indole could be oxidized into a cationic intermediate (**A** or **B**) on which the other nucleophile, respectively the indole or the phenol, could add. However, an enzymatic-mediated radical process seems more reasonable.^[11] Both the indole and the phenol could be oxidized via two single-electron transfers (SET) into radical species (**C**) that would be coupled. Alternatively, only the indolyl radical **D** or phenoxy radical **E** is formed and the second SET happens after the reaction of this radical with the other heteroaromatic unit.

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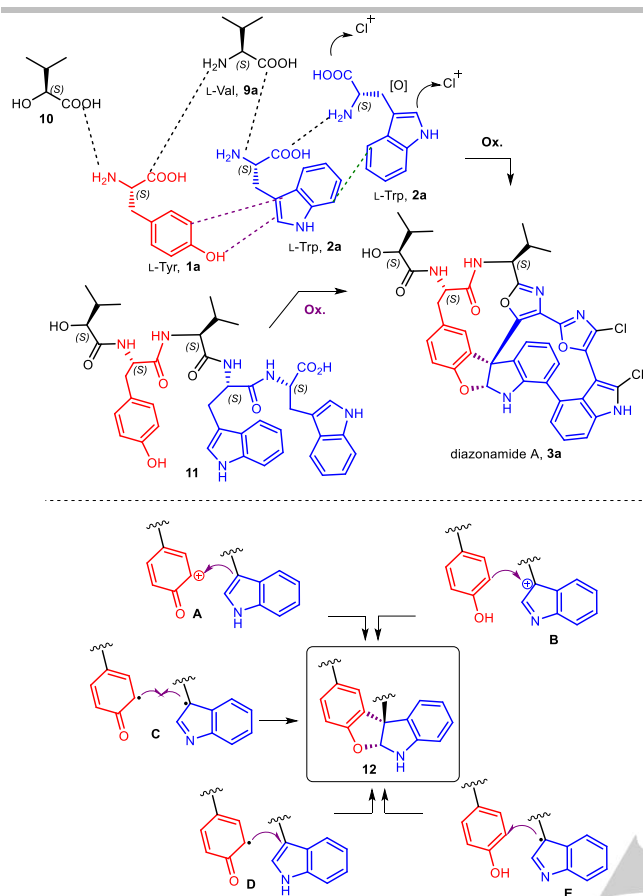
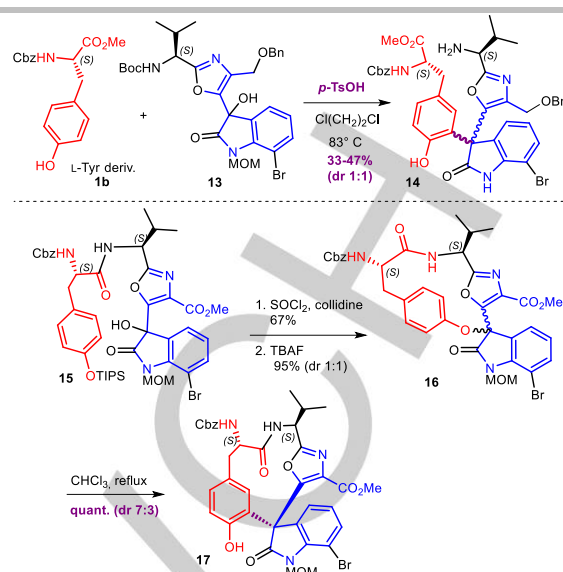


Figure 3. A plausible biosynthetic origin of diazonamide A (3a).

In 2002, the group of Nicolaou was the first to report the total synthesis of the revised structure of diazonamide A via the acid-mediated addition of protected tyrosine methyl ester **1b** to 3-hydroxy-3-oxazolyl-2-oxindole **13** which can be seen as a pre-oxidized tryptophan surrogate leading to **14** without diastereocontrol (Scheme 1).^[11] In 2007 Magnus reported a related intramolecular approach in which the tyrosine residue is attached to the valine unit in **15**.^[12] Conversion of the hydroxyl group on the oxindole into a chloride, followed by removal of the silyl group on the phenol of tyrosine, led to the formation of O-aryl ether **16** as a 1:1 mixture of two epimers. Upon heating the latter, O to C aryl migration occurred delivering overall 3-aryloxindole **17** in 70% along with 30% of its epimer. An intermediate of the Nicolaou synthesis was then intercepted from **17**.



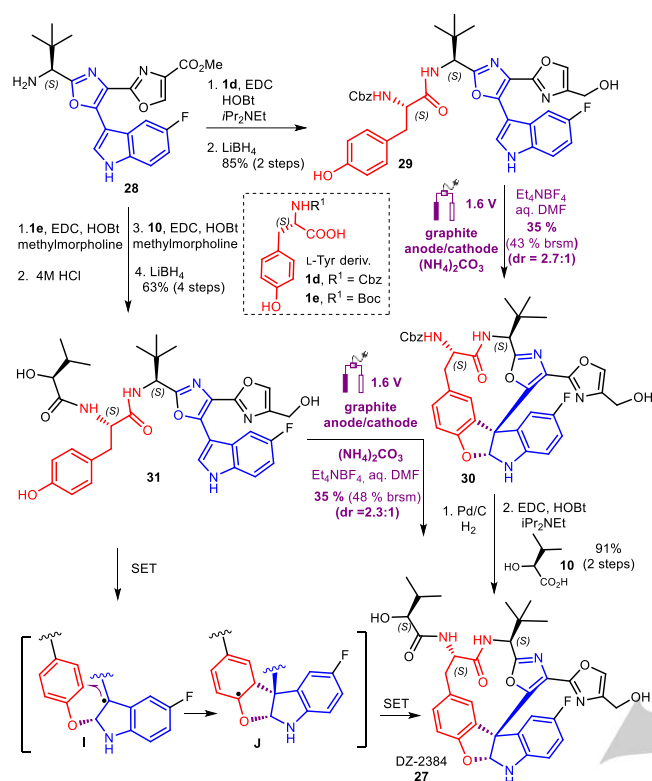
Scheme 1. Key steps of the first total synthesis of diazonamide A by Nicolaou (top, 2002) and the formal synthesis of Magnus (bottom, 2007).

Shortly after the synthesis of Nicolaou (and before the Magnus formal synthesis), Harran proposed a truly bioinspired synthetic route toward diazonamide A from amino-acid derivatives (Scheme 2).^[13] Dipeptide **18** obtained from racemic bromo-tryptophan **2b** with N-Cbz-L-valine **9b** was oxidized with DDQ to form the oxazole ring of **19**.^[14] Further peptidic coupling with N-sulfonyl-L-tyrosine **1c** afforded key precursor **20**.

The stage was set to perform the biomimetic oxidative union of the phenol and indole moiety. Accordingly, oxidation of **20** with the hypervalent iodine reagent $\text{PhI}(\text{OAc})_2$ underwent macrocyclization to directly produce benzofuroindoline **21**. The free phenol of tyrosine would exchange with an acetate of $\text{PhI}(\text{OAc})_2$ to generate highly electrophilic intermediate **F**, which would be captured by the nucleophilic position of the indole to form the C-C bond.^[15] Then, cyclohexanediene would add to the adjacent iminium **G** to provide the hemiaminal of **21** via formation of the C-O bond. The stereocontrol induced by tyrosine and valine residues allows the formation of the quaternary carbon in a 3:1 ratio of diastereoisomers in favour of the desired configuration. The fact that it is the phenol that is oxidized rather than the indole is deduced from the formation of spirocyclic cyclohexanediene **22** as the major by-product via intramolecular addition of the carbonyl of tyrosine into electrophilic intermediate **F**.

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compound **29** via coupling between N-Cbz-L-tyrosine **1d** and compound **28** which was raised from oxidation of a tripeptide constituted of *l*-leucine, 5-fluorotryptophan and serine residues.



Scheme 4. Two concise syntheses of DZ-2384 employing an electrolytic macrocyclization by Harran, Joyant Pharmaceuticals and Paraza Pharma (2010-2015).

Substrate **29** was then electrolyzed in an undivided cell open to air surrounded by concentric rings of graphite rods which acts as anodes and cathodes at a constant cell potential of +1.6 V at ambient temperature in aqueous DMF with addition of solid (NH₄)₂CO₃ in batches and in presence of tetraethylammonium tetrafluoroborate as electrolyte. Impressively, the macrocyclization went smoothly on 60 grams of **29** and 21 grams of **30** were obtained in 35% yield (43% brsm) with a 2.7:1 diastereomeric ratio without the formation of the undesired spirocyclic dienone observed with PhI(OAc)₂. Cyclic voltammetry experiments suggested that it is the indole moiety which is oxidized into radical intermediate **I** via single electron transfer at the anode. **I** could then react with the phenol and a second SET event from **J** would produce benzofuroindoline. This distinct mechanism from the PhI(OAc)₂ oxidation explains that a spirodienone is not formed. The two diastereoisomers were separated after the subsequent removal of the Cbz group. Introduction of the 2-hydroxyisovaleric acid moiety then delivers 20 grams of DZ-2384 (**27**). Because it is necessary to perform a chromatography on silica gel after the electrolysis, the author hoped to perform the electrolysis at the last step to delay this purification. Therefore, the electrolysis was performed on 740 mg of **31** which already contains the 2-hydroxyisovaleric acid residue and was obtained in four steps without chromatography from **28**. Application of the same electrochemical conditions allowed to

obtain directly a 35% yield (48% brsm) of DZ-2384 (**27**) as a 2.3:1 mixture with its diastereoisomer. In summary, the authors were able to obtain drug candidate DZ-2384 in 13 steps with a total yield of 5.7 % from *tert*-leucine.

3. Azonazine

Azonazine (**4**) is another tyrosine-tryptophan-containing marine natural product that contains a benzofuroindoline core. In 2010, Crews and co-workers reported the isolation of this unique hexacyclic dipeptide from a Hawaiian marine sediment-derived fungus, *Aspergillus insulicola*.^[24] Structurally, the benzofuroindoline ring system bearing a quaternary center at its ring junction, indeed presents similarity to the core of the diazonamides (**3a-e**). Noteworthy, the tyrosine-derived nitrogen is methylated and the indolic nitrogen is acetylated. In 2013, following the total synthesis of *ent*-azonazine by Yao, the relative stereochemistry of azonazine was revised from **32** to **4** (Figure 4).^[25] The natural product arose from L-tryptophan and L-tyrosine instead of their D-enantiomers as initially proposed, while the absolute stereochemistry at the ring junction of the benzofuroindoline is the same as the proposed structure. This absolute stereochemistry of the benzofuroindoline part is opposed to the one of the diazonamides, while they both have L-tyrosine and L-tryptophan as starting points. Crews proposed that, biosynthetically, diketopiperazine **33** derived from tyrosine and tryptophan, would undergo an oxidative cyclization in a similar fashion as for diazonamide A.

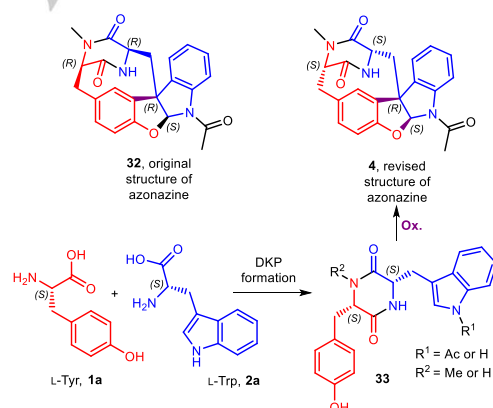
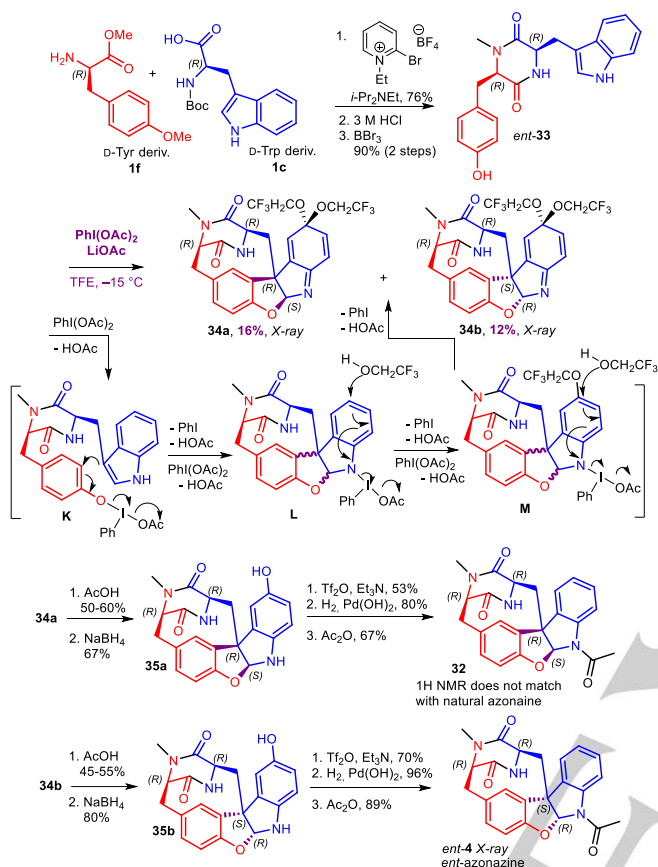


Figure 4. Azonazine and its plausible biosynthesis.

Inspired by the total synthesis of diazonamide A by Harran,^[13] the group of Yao thought to deploy a bioinspired oxidative cyclization between the indole and the phenol of diketopiperazine *ent*-**33** to construct the highly strained benzofuroindoline-containing 10-membered ring system. Diketopiperazine *ent*-**33** was obtained from D-tryptophan **2c** and D-tyrosine **1f** derivatives. After screening various conditions with hypervalent iodine(III) reagents, it was found that 2 equivalents of PhI(OAc)₂ in trifluoroethanol at -15°C allowed to obtain in modest yields the overoxidized hexacyclic skeleton of azonazine as two diastereoisomers **34a** and **34b**, which structures were secured by X-ray analysis. A first oxidative coupling happens between the indole and the phenol to deliver the benzofuroindoline via phenolate iodobenzene **K**. Then,

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exchange of an acetate of $\text{PhI}(\text{OAc})_2$ by the benzofuroindolinic nitrogen could lead to **L** which would be attacked by trifluoroethanol to produce a trifluoroethylarylether. A second addition of trifluoroethanol according to the same process via **M** delivers iminoquinone acetals **34a** and **34b**.



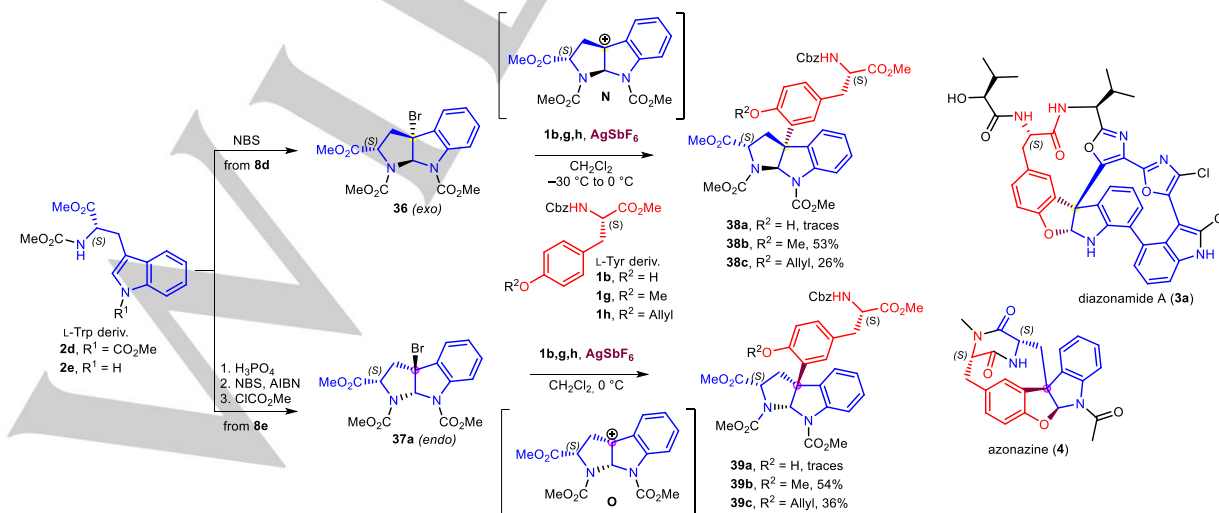
Scheme 5. Total synthesis of *ent*-azonazine (*ent*-**30**) by Yao (2013).

Hydrolysis of the acetal of **34a** and reduction of the quinone delivered phenol **35a** whose hydroxyl was removed by triflation

and hydrogenation. N-acetylation finally provided **32** whose structure was originally assigned to azonazine. However, NMR spectra of this compound did not match with the one of natural azonazine. The same sequence applied to **34b** via **35b** delivered a compound whose NMR spectra matched the ones of the natural product. However, the optical rotation was opposite to the one recorded for the natural product. Therefore, the authors have synthesized the enantiomer of azonazine (*ent*-**4**) and its structure was secured by X-ray analysis.

The diazonamides and azonazine both possess a similar benzofuroindoline structure which arose from the oxidative cyclization of a peptidic precursor containing tyrosine and tryptophan units. However, the configuration of the stereogenic centers at the junction of the benzofurane and indole parts are opposite, despite the fact that these two natural products are both thought to be produced from the proteogenic amino acids L-tyrosine and L-tryptophan.

In 2015, we reported a diastereodivergent intermolecular coupling procedure between L-tyrosine and L-tryptophan derivatives to access both the stereochemistry of the diazonamides and azonazine at the quaternary center linking the indole and the phenol units (Scheme 6).^[26] This strategy employs the stereospecific addition of nucleophiles including arenes,^[27] to bromopyrroloindoline derivatives. Both diastereoisomeric *exo*- and *endo*-bromopyrroloindolines **36** and **37a** were accessed by different bromination methods. Bisprotected L-tryptophan precursor **2d** could be converted diastereoselectively into *exo*-bromopyrroloindoline **36** with the use of NBS,^[28] while *endo*-bromopyrroloindoline **37a** was accessed by acid-mediated cyclization of mono-protected L-tryptophan derivative **2e** followed by a radical-mediated bromination with NBS in presence of AIBN followed by protection of the indoline nitrogen.^[29] In presence of soluble AgSbF_6 , pyrroloindolenium ions **N** and **O** are respectively formed from **36** and **37a** with precipitation of AgBr . Due to the existence of a stable two-membered *cis*-fused system in these cationic species, nucleophiles can add only on the open face and therefore this Friedel-Crafts-type reaction proceeds with retention of configuration. While free-phenol tyrosine derivative **1b** only led to traces of the desired coupling products, O-methyl and O-allyl tyrosine **1g** and **1h** were efficiently added to **36** and **37a** to respectively lead to **38b,c** with the stereochemistry of the diazonamides and **39b,c** related to azonazine.



Scheme 6. Diastereodivergent dearomative arylation process of L-tryptophan by L-tyrosine by Vincent (2015).

4. Tryptorubin A

Tryptorubin A (**5**) is of terrestrial origin unlike diazonamide A and azonazine. This natural product was isolated in 2017 by Clardy from a bacterial strain (*Streptomyces* sp. CLI2509) associated symbiotically to a fungus-growing ants (*Hymenochaete rubiginosa*).^[30] This bicyclic hexapeptide (Ala-Trp-Tyr-Ile-Trp-Tyr), which possesses two L-tryptophans (**2a**) and two L-tyrosines (**1a**), displays two unusual bonds: a C-N bond between one of the tyrosine and one of the tryptophan units and a C-C bond between the two tryptophan units (Figure 5). In contrast to the diazonamides and azonazine, the biosynthetic oxidative coupling between the tyrosine and tryptophan units of tryptorubin A (**5**) did not result in the formation of a benzofuroindoline scaffold.

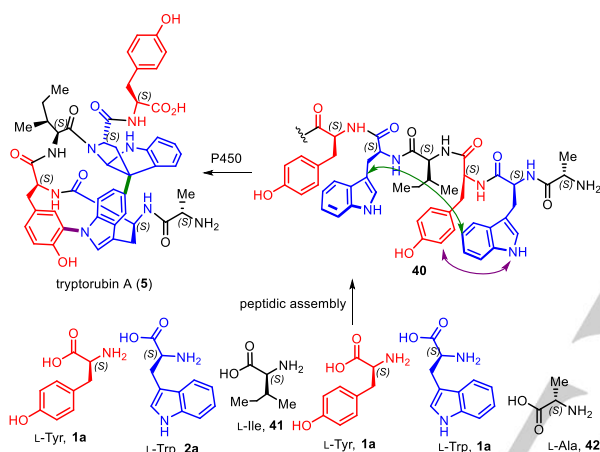


Figure 5. Structure and biosynthesis of tryptorubin A.

The tryptophan-tryptophan linkage connects the C3-position of one of the indole and the benzene ring of the other indole leading to a 3-arylated pyrroloindoline motif. Similar bonds between the C3 position and the benzene ring of two tryptophan units are found in dimeric diketopiperazine natural products such as naseseazines A-C, asperazine or pestalazine A (**43a-e**, Figure 6) which are biosynthetically produced by oxidative dimerisations mediated by P450 enzymes.^[31] More precisely, naseseazines A and B display a C3-C7' bond, while asperazine and pestalazine A present a C3-C8' bond. Only naseseazine C possess the C3-C6' connectivity encountered in tryptorubin A.

The linkage between the internal tyrosine and the tryptophan already connected by its benzene ring is characterized by a bond between the *ortho* position of the phenol and the indolic nitrogen. Among natural products, only, the dimeric ergot alkaloid, cividiclavine, feature a bond between the *ortho*-position of a phenol and an indolic nitrogen.^[32]

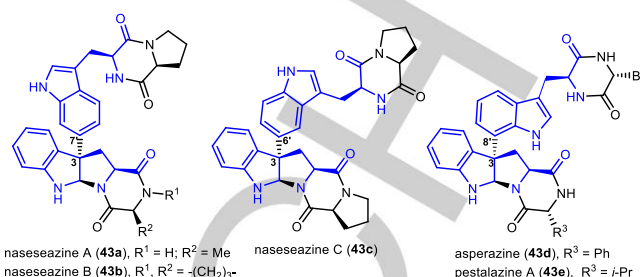


Figure 6. Structure of dimeric tryptophan-containing diketopiperazine related to tryptorubin A.

Tryptorubin A is believed to be a ribosomal synthesized and post-translationally modified peptide (RiPP). Biosynthetically the unusual C-C and C-N bonds were probably formed by two intramolecular oxidative coupling mediated by a cytochrome P450 enzyme after assembly of the linear hexapeptidic chain **40** (Figure 4).^[33] Interestingly the terminal tyrosine is not involved in any oxidative coupling while one of the tryptophan is central to the two cyclisation events.

During their synthetic studies towards tryptorubin A, Baran and co-workers discovered that two non-canonical atropoisomers **4** and **44** could exist with the same connectivity and central-stereochemistry (Figure 7).^[33] The natural isomer **4** of tryptorubin A is in the "bridge above" conformation, and its non-natural isomer **44** is in the "bridge below" conformation. The two compounds did not convert to each other at elevated temperature, indicating that the energy barrier between these two non-canonical atropoisomers is high. Theoretically, the interconversion of the two isomers **4** and **44** requires a non-physical, inside-out flip of the molecule, allowing one macrocycle to pass through the other. If it is assumed that the bond can be extended beyond the covalent limit, then **4** and **44** can be interconverted. In other words, without breaking and reformatting the covalent bond, this conversion can hardly happen. Overall, it is in stark contrast to canonical atropisomerism and topoisomerism.

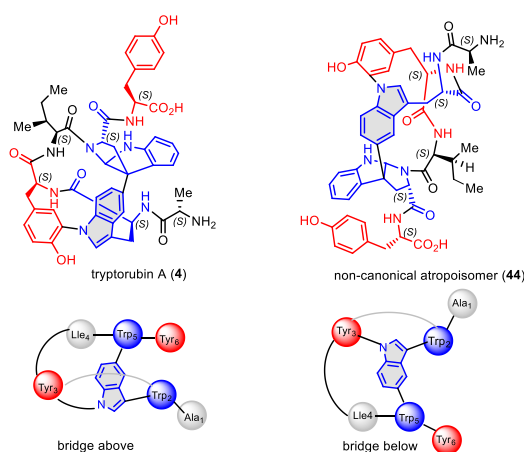
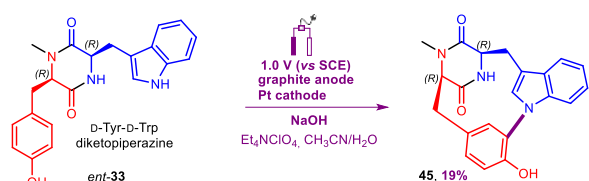


Figure 7. The conformation of the non-canonical atropoisomers of tryptorubin A.

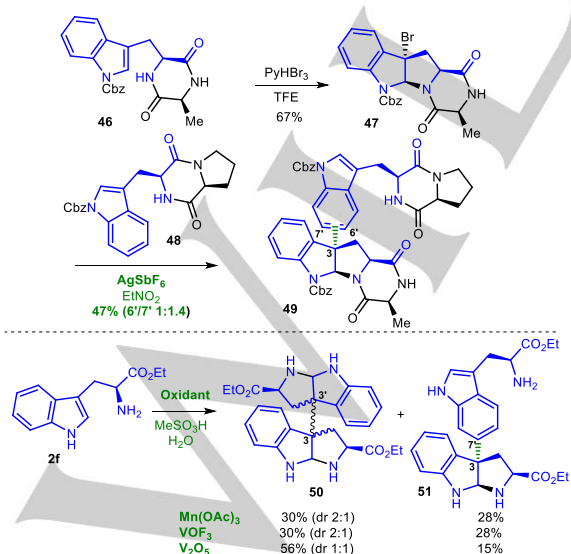
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Interestingly, before the discovery of tryptorubin A, the formation of a related tyrosine-tryptophan linkage was reported in 2014. During its synthetic studies toward azonazine, Yao investigated the use of electrochemistry to form the benzofuroindoline motif. Subjecting diketopiperazine **ent-33** to a constant current (1.0 V vs SCE) in presence of sodium hydroxide as a base resulted unexpectedly to the formation of the C(sp²)-N bond between the *ortho*-position of the phenol and the indolic nitrogen to generate a new 12-membered ring **45** similar to the one encountered in tryptorubin A (Scheme 7).^[25b] The reaction presumably proceeds via single electron transfers from the indole and/or the phenol into radical intermediates.



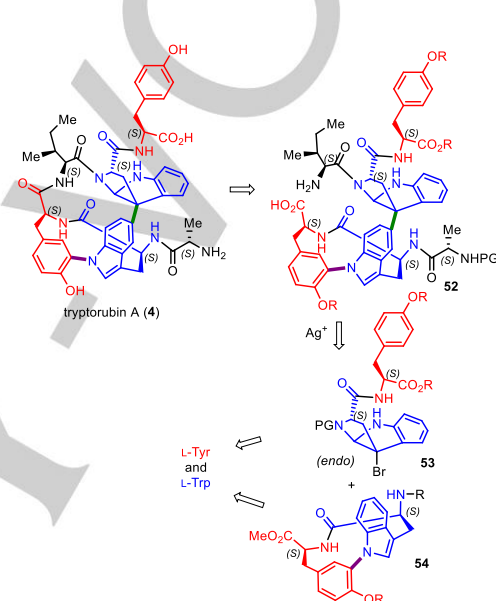
Scheme 7. Formation of a C-N bond by electrochemical oxidative cyclization of diketopiperazine **ent-33** (2014).

In the other hand, bioinspired oxidative dimerizations of tryptophan derivatives have been developed in the context of the total synthesis of naseeseazines A,B, asperazine or pestalazine A (Scheme 8).^[27a,27b,34-36] For instance, Movassaghi described a two steps process in which diketopiperazine **46** is converted into bromopyrroloindoline **47** that is then involved in a Friedel-Crafts reaction in presence of AgSbF₆ with another tryptophan-containing diketopiperazine **48** to deliver a mixture of regioisomeric arylated-pyrroloindolines **49**.^[27a,b] The regioselectivity could be controlled by the presence of a tetrafluoroborate on the nucleophilic entity but requires prefunctionalization (not shown). Alternatively, Ishikawa reported a direct biomimetic oxidation of L-tryptophan ester **2f** with Mn(OAc)₃, VOF₃ or V₂O₅ in acidic conditions that deliver C3-C3' dimer **50** as well as the C3-C7' **51** dimer.^[34]



Scheme 8. C-C bond formation via bioinspired oxidative dimerizations of tryptophan derivatives by Movassaghi (top, 2011) and Ishikawa (bottom, 2013).

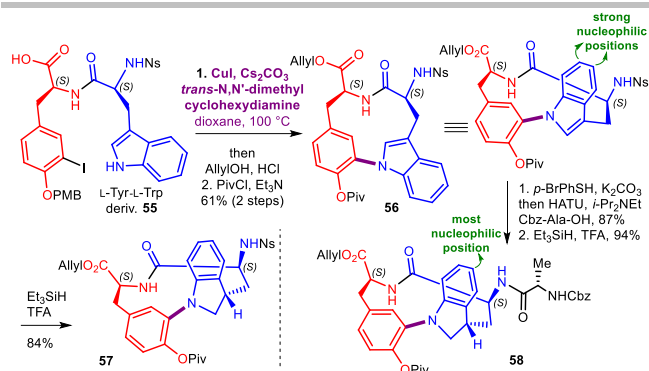
In 2020, Baran reported the first total synthesis of tryptorubin A (Scheme 9). When they started their synthetic endeavor, the authors were unaware of the possibility that the structure of tryptorubin A could exist as two non-canonical atropoisomers. Their retrosynthesis involved a late stage intramolecular peptidic coupling between the amine of the *iso*-leucine and the carboxylic acid of the internal tyrosine of **52**. Inspired by the Movassaghi's total synthesis of the naseeseazines,^[27a] the C-C bond between the two tryptophan residues would arise from an intermolecular Friedel-Crafts type reaction between tyrosine-containing *endo*-bromopyrroloindoline **53** and Tyr-Trp dipeptide **54** in which the C-N bond is already formed.



Scheme 9. Retrosynthesis of tryptorubin A by Baran (2020).

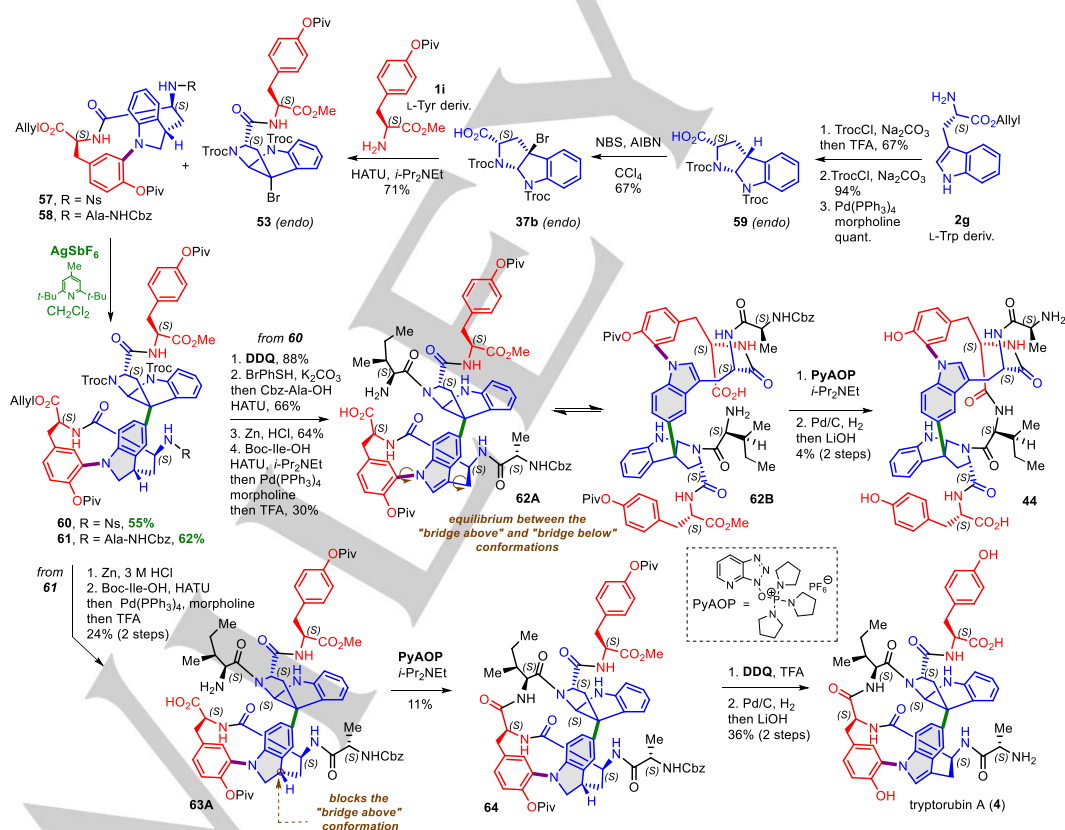
The authors adopted a non-biomimetic approach to form the C-N bond between the indole and the phenol (Scheme 10). An intramolecular Ullmann-Goldberg coupling with Buchwald-type conditions was applied to *ortho*-iodotyrosine-derived dipeptide **55** to yield **56** on a 20 g scale.^[37] To avoid 6'/7' regioselectivity issues during the forthcoming Friedel-Crafts reaction, as observed by Movassaghi for the naseeseazines (see Scheme 8), the indole ring was reduced into indoline **57** with triethylsilane in order to increase the nucleophilicity of the position in *para* of the nitrogen. The alanine residue could also be appended before the reduction of the indole into **58**.

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Scheme 10. C-N bond formation from a Tyr-Trp dipeptide derivative by Baran (2020).

Endo-Bromopyrroloindoline **53** was obtained by acid mediated-cyclization of tryptophan derivative **2g** followed by radical-bromination^[26,29] of **59** before coupling of **37b** with tyrosine methyl ester **1i**. Coupling of the latter with cyclic Tyr-Trp dipeptide **57** or **58** via the silver-mediated alkylation reaction established the key C-C bond of **60** and **61** in a regioselective and stereoselective manner (Scheme 11). Oxidative aromatization of indoline **60** with



Scheme 11. Total synthesis of tryptorubin A and its non-canonical atropisomer by Baran (2020).

5. Concluding remarks

DDQ allowed to restore the indole ring. Then installation of alanine and *iso*-leucine residues extended the peptidic chain and gave hexamer **62** after deprotection. Final cyclization was then performed which led after deprotection with a total yield of 4% to **44** whose NMR data did not match natural tryptorubin A. The structure of this compound was attributed to *atrop*-tryptorubin A.

The authors proposed that intermediate **62** have in equilibrium two conformers **62A** and **62B** via rotation around the enamine part of the indole. Since the second macrocycle has not been formed yet, this rotation is allowed. It is believed that the "ring-below" atropisomer **62B** is the most kinetically favourable conformation for the cyclisation process and therefore lead atroposelectively to **44**.

The authors circumvented this problem by performing the aromatisation of the indoline after the macrocyclisation. The central chirality of the stereocenter on the indoline locked the "bridge above" conformation by structurally excluding the "bridge below" conformational isomer. After introduction of *iso*-leucine, the macrocyclisation of "bridge above" *seco*-acid **63A** with PyAOP allowed to obtain atroposelectively isomer **64** which was converted to the natural product tryptorubin A (**4**) after oxidation of the indoline into the indole ring with DDQ and removal of the Cbz group and saponification of the methyl ester.

In summary, we analyzed the biomimetic synthetic strategies towards diazonamide A, azonazine and tryptorubin A via oxidative coupling of tyrosine and tryptophan residues of peptidic

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derivatives. It is apparent that accessing the benzofuroindoline core of diazonamide A and azonazine and its quaternary carbon has always been challenging. The development of phenol-indole oxidative coupling mediated by hypervalent iodine reagents allowed to overcome these difficulties and to perform the total synthesis of diazonamide A and azonazine. However, there is still room for innovation as demonstrated by the use of electrochemistry that improve the oxidative coupling process during the large scale production of DZ-2384, a simplified analog of diazonamide A with a high therapeutic potential as an antitumoral agent. Tryptorubin A differs from the diazonamides and azonazine by the fact that the biosynthetic oxidative coupling of tyrosine and tryptophan delivers a C-N bond between the indolic nitrogen and the carbon of in the *ortho*-position of the phenol instead of a benzofuroindoline. The recent synthesis of tryptorubin A highlighted the highly unusual non-canonical atropoisomerism of this bicyclic hexapeptide which may inspired chemists in the design and analysis of complex three dimensional structures.

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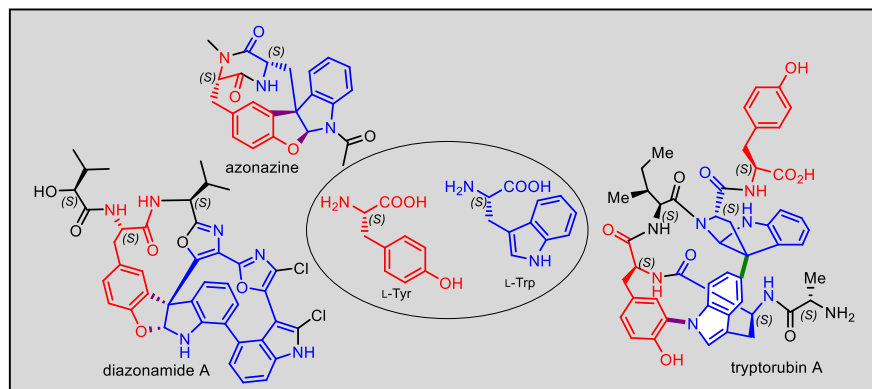
Keywords: tyrosine • tryptophan • oxidative coupling • natural products • bioinspired synthesis

- [1] M.-C. Tang, Y. Zou, K. Watanabe, C. T. Walsh, Y. Tang, *Chem. Rev.* **2017**, *117*, 5226-5333.
- [2] a) J. Kohno, Y. Koguchi, M. Nishio, K. Nakao, M. Kuroda, R. Shimizu, T. Ohnuki, S. Komatsubara, *J. Org. Chem.* **2000**, *65*, 990-995; for a review: b) A. Coste, F. Couty, G. Evano, *C. R. Chimie*, **2008**, *11*, 1544-1573.
- [3] a) S. Lin, S. J. Danishefsky, *Angew. Chem. Int. Ed.* **2002**, *41*, 512-515; b) M. Inoue, H. Sakazaki, H. Furuyama, M. Hiram, *Angew. Chem. Int. Ed.* **2003**, *42*, 2654-2657; c) B. K. Albrecht, R. M. Williams, *Org. Lett.* **2003**, *5*, 197-200.
- [4] a) Y. Yamada, T. Fujiwara, T. Sato, N. Igarashi, N. Tanaka, *Nat. Struct. Biol.* **2002**, *9*, 691-695; b) X. Carpena, S. Loprasert, S. Mongkolus, J. Switala, P. C. Loewen, I. Fita, *J. Mol. Biol.* **2003**, *327*, 475-489.
- [5] N. Lindquist, W. Fenical, G. D. VanDuyne and J. Clardy, *J. Am. Chem. Soc.* **1991**, *113*, 2303-2304.
- [6] J. Li, S. Jeong, L. Esser, P. G. Harran, *Angew. Chem. Int. Ed.* **2001**, *40*, 4765-4769.
- [7] J. Li, A. W. G. Burgett, L. Esser, C. Amezcua, P. G. Harran, *Angew. Chem. Int. Ed.* **2001**, *40*, 4770-4773.
- [8] bipleiophylline: a) T.-S. Kam, S.-J. Tan, S.-W. Ng, K. Komiyama, *Org. Lett.* **2008**, *10*, 3749-3752; pleiocraline: b) B. C. Das, J. P. Cosson, G. Lukacs, *J. Org. Chem.* **1977**, *42*, 2785-2786; pleiocorine: c) B. C. Das, J. P. Cosson, G. Lukacs, Potier, P., *Tetrahedron Lett.* **1974**, *15*, 4299-4302.
- [9] For reviews on the synthesis of benzofuroindolines: a) Ito, Y.; Ueda, M.; Miyata, O. *Heterocycles*, **2014**, *89*, 2029-2052; b) Beaud, R.; Tomakinian, T.; Denizot, N.; Pouilhès, A.; Kouklovsky, C.; Vincent, G. *Synlett*, **2015**, *26*, 432-440.
- [10] R. Fernández, M. J. Martín, R. Rodríguez-Acebes, F. Reyes, A. Francesch, C. Cuevas, *Tetrahedron Lett.* **2008**, *49*, 2283-2285.
- [11] a) K. C. Nicolaou, M. Bella, D. Y.-K. Chen, X. Huang, T. Ling, S. A. Snyder, *Angew. Chem. Int. Ed.* **2002**, *41*, 3495-3499; b) K. C. Nicolaou, D. Y.-K. Chen, X. Huang, T. Ling, M. Bella, S. A. Snyder, *J. Am. Chem. Soc.* **2004**, *126*, 12888-12896.
- [12] C.-M. Cheung, F. W. Goldberg, P. Magnus, C. J. Russell, R. Turnbull, V. Lynch, *J. Am. Chem. Soc.* **2007**, *129*, 12320-12327.
- [13] A. W. G. Burgett, Q. Y. Li, Q. Wei, P. G. Harran, *Angew. Chem. Int. Ed.* **2003**, *42*, 4961-4966.
- [14] The group of Moody was able to perform an oxidative cyclization of a tetrapeptide related to **11** to form the bis-oxazole moiety but without forming the benzofuroindoline and the biaryl bond: a) J. Sperry, C. J. Moody, *Chem. Commun.* **2006**, 2397-2399; b) J. Sperry, C. J. Moody, *Tetrahedron* **2010**, *66*, 6483-6495.
- [15] For other examples of synthesis of benzofuroindolines with hypervalent iodine reagents: a) Chan, C.; Li, C.; Zhang, F.; Danishefsky, S. J. *Tetrahedron Lett.* **2006**, *47*, 4839-4841; b) Nicolaou, K. C.; Majumder, U.; Roche, S. P.; Chen, D. Y.-K. *Angew. Chem. Int. Ed.* **2007**, *46*, 4715-4718; d) Li, L.; Yuan, K.; Jia, Q.; Jia, Y. *Angew. Chem. Int. Ed.* **2019**, *58*, 6074-6078; for other selected bioinspired synthesis of benzofuroindolines: e) W. Tian, L. Rao Chennamaneni, T. Suzuki, D. Y.-K. Chen, *Eur. J. Org. Chem.* **2011**, 1027-1031; f) R. Beaud, R. Guillot, C. Kouklovsky, G. Vincent, *Angew. Chem. Int. Ed.* **2012**, *51*, 12546-12550; g) L.; Liao, C.; Shu, M. Zhang, Y. Liao, X.-Y. Hu, Y. Zhang, Z. Wu, W. Yuan, X.-M. Zhang, *Angew. Chem. Int. Ed.* **2014**, *53*, 10471-10475; h) T. Tomakinian, R. Guillot, C. Kouklovsky, G. Vincent, *Angew. Chem. Int. Ed.* **2014**, *53*, 11881-11885; i) N. Denizot, A. Pouilhès, M. Cucca, R. Beaud, R. Guillot, C. Kouklovsky, G. Vincent, *Org. Lett.* **2014**, *16*, 5752-5755; j) D. Lachkar, N. Denizot, G. Bernadat, K. Ahamada, M. A. Beniddir, V. Dumontet, J.-F. Gallard, R. Guillot, K. Leblanc, E. O. N'ngang, V. Turpin, C. Kouklovsky, E. Poupon, L. Evanno, G. Vincent, *Nat. Chem.* **2017**, *9*, 793-798; k) Liu, K.; Tang, S.; Huang, P.; Lei, A. *Nat. Commun.* **2017**, *8*, 775.
- [16] a) K. C. Nicolaou, J. Hao, M. V. Reddy, P. B. Rao, S. A. Snyder, G. Rassias, X. Huang, D. Y.-K. Chen, *Angew. Chem. Int. Ed.* **2003**, *42*, 1753-1758; b) K. C. Nicolaou, J. Hao, M. V. Reddy, P. B. Rao, G. Rassias, S. A. Snyder, X. Huang, D. Y.-K. Chen, W. E. Brenzovich, N. Giuseppone, P. Giannakou, A. O'Brate, *J. Am. Chem. Soc.* **2004**, *126*, 12897-12906; c) C.-K. Mai, M. F. Sammons, T. Sammakia, *Angew. Chem. Int. Ed.* **2010**, *49*, 2397-2400; d) R. R. Knowles, J. Carpenter, S. B. Blakey, A. Kayano, I. K. Mangion, C. J. Sinz, D. W. C. MacMillan, *Chem. Sci.* **2011**, *2*, 308-311; e) N. David, R. Pasceri, R. R. A. Kitson, A. Pradal, C. J. Moody, *Chem. Eur. J.* **2016**, *22*, 10867-108776.
- [17] For a review: M. Lachia, C. J. Moody, *Nat. Prod. Rep.* **2008**, *25*, 227-253.
- [18] J. Lin, B. S. Gerstenberger, N. Y. T. Stessman, J. P. Konopelski, *Org. Lett.*, **2008**, *18*, 3969-3972;
- [19] a) Z. Cruz-Monserrate, H. C. Vervoort, R. Bai, D.J. Newman, S. B. Howell, G. Los, J. T. Mullaney, M. D. Williams, G. R. Pettit, W. Fenical, E. Hamel, *Mol. Pharmacol.* **2003**, *63*, 1273-1280; b) G. Wang, L. Shang, A. W. G. Burgett, P. G. Harran, X. Wang, *Proc. Natl. Acad. Sci. U.S.A.* **2007**, *104*, 2068-2073.
- [20] N. S. Williams, A. W. G. Burgett, A. S. Atkins, X. Wang, P. G. Harran, S. L. McKnight, *Proc. Natl. Acad. Sci. U.S.A.* **2007**, *104*, 2074-2079.
- [21] a) G. Hanson, C. Caldwell, P.G. Harran, S. Harran, Q. Wei, M. Zhou, *US 7,851,620 B2*, **2010**; b) Harran, P. G.; Williams, N.; Burgett, A. *US 7,538,129 B2*, **2009**.
- [22] a) M. Wiczorek, J.Tcherkezian, C. Bernier, A. E. Prota, S. Chaaban, Y. Rolland, C. Godbout, M. A. Hancock, J. C. Arezzo, O. Ocal, C. Rocha, N. Olieric, A. Hall, H. Ding, A. Bramoullé, M. G. Annis, G. Zogopoulos, P. G. Harran, T. M. Wilkie, R. A. Brekken, P. M. Siegel, M. O. Steinmetz, G. C. Shore, G. J. Brouhard, A. Roulston, *Sci. Transl. Med.* **2016**, *8*, 365-159; b) C. Bernier, A. Soliman, M. Gravel, M. Dankner, P. Savage, K. Petrecca, M. Park, P. M. Siegel, G. C. Shore, A. Roulston, *Anticancer. Drugs* **2018**, *29*, 774-785.
- [23] H. Ding, P. L. DeRoy, C. Perreault, A. Larivée, A. Siddiqui, C. G. Caldwell, S. Harran, and P. G. Harran, *Angew. Chem. Int. Ed.* **2015**, *54*, 4818-4822.
- [24] Q. X. Wu, M. S. Crews, M. Draskovic, J. Sohn, T. A. Johnson, K. Tenney, F. A. Valeriote, X. J. Yao, L. F. Bjeldanes, P. Crews, *Org. Lett.* **2010**, *12*, 4458-4461.
- [25] a) J. C. Zhao, S. M. Yu, Y. Liu, Z. J. Yao, *Org. Lett.* **2013**, *15*, 4300-4303; b) J. C. Zhao, S. M. Yu, H. B. Qiu, Z. J. Yao, *Tetrahedron*. **2014**, *70*, 3197-3210.
- [26] N. Denizot, R. Guillot, C. Kouklovsky, G. Vincent, *Chem. Eur. J.* **2015**, *21*, 18953-18956.
- [27] a) J. Kim, M. Movassaghi, *J. Am. Chem. Soc.* **2011**, *133*, 14940-14943; b) R. Loach, O. S. Fenton, M. Movassaghi, *J. Am. Chem. Soc.* **2016**, *138*, 1057-1064; c) Boyer, M. Movassaghi, *Chem. Sci.* **2012**, *3*, 1798-1803; d) A. Coste, J. Kim, T. C. Adams, M. Movassaghi, *Chem. Sci.* **2013**, *4*,

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- 3191–3197; e) Y. Wang, C. Kong, Y. Du, H. Song, D. Zhang, Y. Qin, *Org. Biomol. Chem.* **2012**, *10*, 2793–2797.
- [28] C. S. Lopez, C. Perez-Balado, P. Rodriguez-Grana, A. R. de Lera, *Org. Lett.* **2008**, *10*, 77–80.
- [29] a) M. Movassaghi, O. K. Ahmad, S. P. Lathrop, *J. Am. Chem. Soc.* **2011**, *133*, 13002–13005; b) M. Bruncko, D. Crich, R. Samy, *J. Org. Chem.* **1994**, *59*, 5543–5549.
- [30] T. P. Wyche, A. C. Ruzzini, L. Schwab, C. R. Currie, J. Clardy, *J. Am. Chem. Soc.* **2017**, *139*, 12899–12902.
- [31] a) W. Tian, C. Sun, M. Zheng, J. R. Harmer, M. Yu, Y. Zhang, H. Peng, D. Zhu, Z. Deng, S.-L. Chen, M. Mobli, X. Jia, X. Qu, *Nat. Commun.* **2018**, *9*, 4428; b) H. Yu, S.-M. Li, *Org. Lett.* **2019**, *21*, 7094–7098.
- [32] L. C. Vining, A. G. McInnes, D. G. Smith, J. L. C. Wright, W. A. Taber, *FEMS Symp.* **1982**, *13*, 243–251.
- [33] S. H. Reisberg, Y. Gao, A. S. Walker, E. J. N. Helfrich, J. Clardy, P. S. Baran, *Science* **2020**, *367*, 458–463.
- [34] a) S. Tadano, Y. Mukaeda, H. Ishikawa, *Angew. Chem. Int. Ed.* **2013**, *52*, 7990–7994; b) S. Tadano, Y. Sugimachi, M. Sumimoto, S. Tsukamoto, H. Ishikawa, *Chem. Eur. J.* **2016**, *22*, 1277–1291.
- [35] For a review: S. Tadano, H. Ishikawa, *Synlett*, **2014**, *25*, 157–162.
- [36] For other selected syntheses: a) S. P. Govek, L. E. Overman, *J. Am. Chem. Soc.* **2001**, *123*, 9468–9469; b) S. P. Govek, L. E. Overman, *Tetrahedron* **2007**, *63*, 8499–8513; c) M. E. Kieffer, K. V. Chuang, S. E. Reisman, *J. Am. Chem. Soc.* **2013**, *135*, 5557–5560; d) X. Wang, G. Ma, Y. Peng, C. E. Pitsch, B. J. Moll, T. D. Ly, X. Wang, H. Gong, *J. Am. Chem. Soc.* **2018**, *140*, 14490–14497; e) L. Luo, X.-Y. Zhai, Ya-W. Weng, Y. Peng, H. Gong, *Chem. Eur. J.* **2019**, *25*, 989–992.
- [37] K. Kunz, U. Scholz, D. Ganzer, *Synlett* **2003**, *15*, 2428–2439.

Entry for the Table of Contents



We review the oxidative coupling of tyrosine and tryptophan units as a pivotal step in the total synthesis of some peptide-derived natural products such as diazonamide A, azonazine and tryptorubin A.

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