

#### Impact of nitrogen sources on the fermentative kinetic and aroma compounds production by non-Saccharomyces yeasts in alcoholic fermentation

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#### ▶ To cite this version:

Pauline Seguinot, Ying Su, Isabelle Sanchez, Audrey Bloem, Pascale Brial, et al.. Impact of nitrogen sources on the fermentative kinetic and aroma compounds production by non-Saccharomyces yeasts in alcoholic fermentation. International Conference in Wine Sciences Macrowine 2018, May 2018, Saragosse, Spain. hal-02958317

HAL Id: hal-02958317

https://hal.science/hal-02958317

Submitted on 8 Oct 2020

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# IMPACT OF NITROGEN SOURCES ON THE FERMENTATIVE KINETIC AND AROMA COMPOUNDS PRODUCTION BY NON-**SACCHAROMYCES YEASTS**

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Over the past 50 years, the completion of wine fermentation and the formation of undesirable off-compounds have been mainly controlled through the use of selected starter strains of S. cerevisiae. Little attention was paid to the non-Saccharomyces yeasts, predominant in grape juice but rapidly outcompeted by S. cerevisiae during fermentation. However, the potential of some of these species, thank to their ability to produce fermentative aromas, to improve the sensory quality of wines is now recognized even if the metabolic specificities of non-Saccharomyces yeasts remain ill-characterised. To fill this shortcoming that is a hindrance to an efficient use of non-Saccharomyces strains during wine fermentation, the aim of this study was to investigate to impact of the nature of nitrogen source on the fermentation performances and the production of volatile molecules of 2 non-Saccharomyces species.

### **Material and Methods** Fermentation in synthetic media with only one Study of 3 yeasts stains, naturally present nitrogen source (200 mgN/L) among 27: on grapes: Ade, Ala, Arg, Asp, Asn, Cit, Cys, Cyt, GABA, Gln, Glu, Torulaspora delbrueckii (TD 291) Gly, His, Ile, Leu, Lys, Met, NH4, Orn, Phe, Pro, Ser, Metschnikowia pulcherrima (MP 346) Thr, Trp, Tyr, Urea, Val Saccharomyces cerevisiae (SC 09) Fermentation in 15 mL-tubes – 1 AA per media PhenoFerm: automatic weighting device > Point each 4 h per tube to accurate determination of fermentation rate **Fermentation parameters** Offline determination of metabolites Measure of concentrations of 35 aroma compounds at the end of fermentation by GCMS Determination of characteristic parameters : - Tlag: lag phase (h) Residual nitrogen at the end of fermentation Rmax: maximal fermentation rate (g/L/h) determined by NOPA analysis (amino acids) or Trmax: time to reach Rmax (h)

## Production of aroma compounds

with only valine:

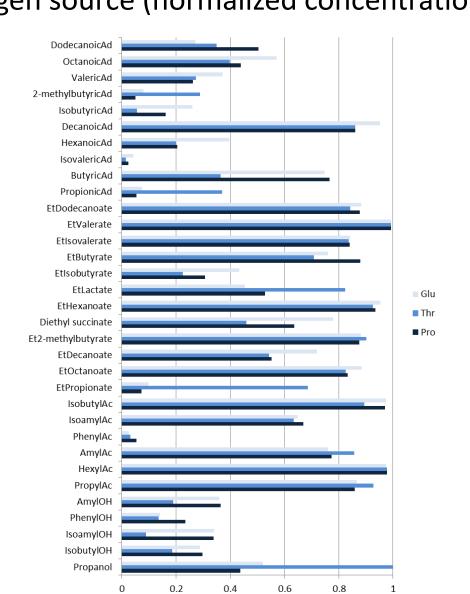
Ethyl isobutyrate

from  $\alpha$ -keto-isovalerate.

←→ α-keto-isovalerate

Production of volatile compounds during fermentation using a sole nitrogen source

Aromatic profiles of TD 291 in fermentation on sole nitrogen source (normalized concentrations):



propyl acetate

Percentage difference between the production of aroma

compounds by TD 291 in complete medium vs on medium

Pyruvate – → TCA cycle – → Aspartate – – > Threonine

**TD 291** 

α-keto-butyrate

ethyl propionate

+1723%

Most of studied aroma compounds are produced irrespective of the nitrogen source Compounds synthetized from

MCC metabolites, regulated by

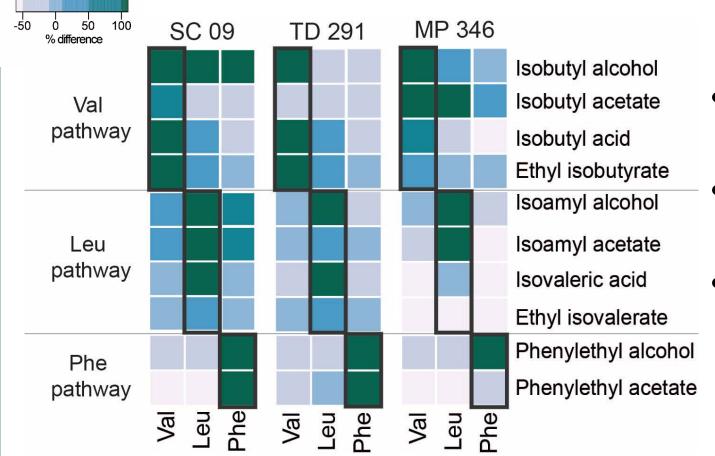
total nitrogen availability

- Moderate increase of volatile compounds not derivated from valine metabolism, as propanol
  - > Impact of MCC

Huge increase in volatiles molecules that directly derive

Impact of the nitrogen source on the production of some aroma through Ehrlich pathway

Percentage differences between the production on complete medium and on sole nitrogen source medium



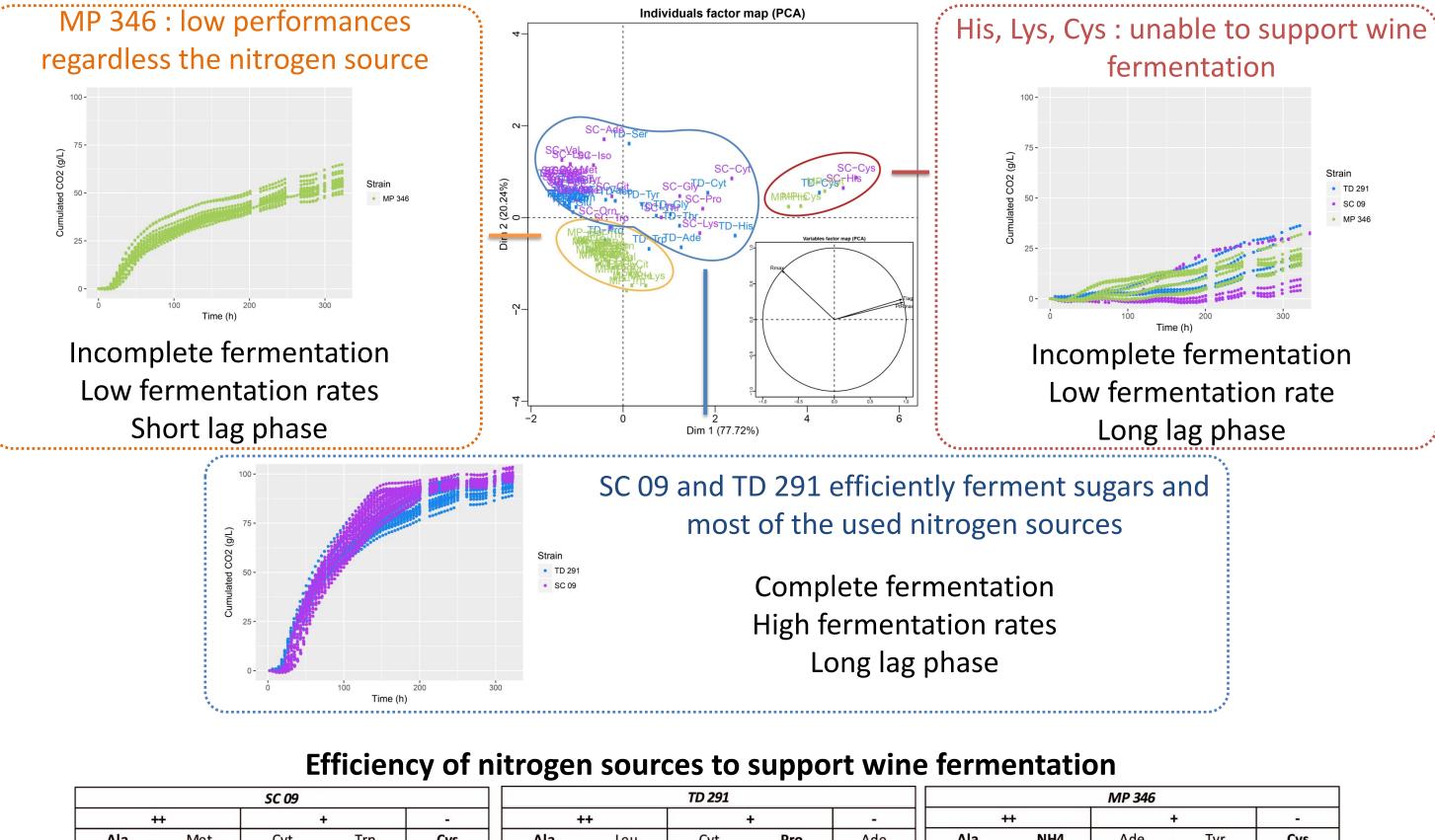
- Overall, increase in the production of compounds derived from the nitrogen source
- Low ability of TD 291 to produce acetate esters except phenylethyl acetate
- In contrast, with MP 346 increase in the production of acetate esters except phenylethyl acetate. Low producer of acids and their ethyl esters derivatives
- > Similar responses to the nature of the nitrogen sources for the 3 species
- > However, some species specificities, likely related to their genetic background

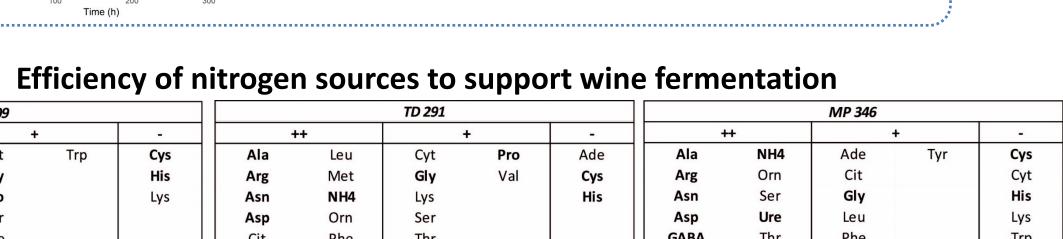
## Principal component analysis of fermentation parameters: lag phase (Tlag), maximal fermentation rate (Rmax), and time to reach the Rmax (Trmax).

Fermentation performances

Fermentation profiles and kinetics parameters depend on both the strain and the nitrogen source

enzymatic kit (NH<sub>4</sub>)





++: best nitrogen sources +: average sources +: poor sources. In bold, nitrogen sources in the same category for the 3 strains

- In contrast to MP 346, TD 291 and SC 09 are able to achieve fermentation using most of amino acids as sole nitrogen source

• Many similarities in nitrogen sources preferences between non-Saccharomyces strains and S. cerevisiae

- Some differences:
  - Phe supports more efficiently TD 291 and SC 09 fermentation than MP 346 fermentation
  - Trp can not be used by MP 346 to carry out fermentation unlike SC 09 and TD 291
  - Lys sustains a moderate fermentation by TD 291, but not for SC 09 and MP 346

## Consumption of nitrogen sources

Percentage of residual nitrogen at the end of fermentation: Citruline **Fryptophane** Glutamine Glutamine

- SC 09: exhaustion of most of the nitrogen sources except Ser (15%), Lys (11%), His (85%), Glu (9%) and Cys (39%). TD 291: complete consumption of Phe, Glu, Arg and Ala. Limited ability (residual nitrogen > 80%) to assimilate His and Gly. Higher consumption of Cys, Glu ans Ser than SC 09.
- MP 346 : consumption between 47 and 98% of initial nitrogen. Higher consumption of Cys, His and Ser than SC 09.
- > Correlation between the amount of consumed nitrogen and fermentation performances. In particular, inability to sustain fermentation on poorly consumed nitrogen sources.

Distinct fermentative profiles were observed depending on the nitrogen source and the yeast strain. Yeasts had their own preferences in term of nitrogen sources that have shown diverse efficiency to sustain fermentative activity. Analysis of aroma compounds production according to the nitrogen source revealed a redistribution of carbon and nitrogen thought the entire metabolism network, as the presence of a sole amino acids allow the production of all aroma compounds and not only those directly produced by the amino acid. Differences between the species have been highlighted, especially concerning the production of acetate esters. Nitrogen consumption is consistent with fermentation kinetics. These results provide new insights for the management of co-cultures of non-Saccharomyces and S. cerevisiae, emphasizing the importance of managing the nitrogen nutrition.



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### Macrowine 2018 - Zaragoza

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