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Late Ventilator-Induced Diaphragmatic Dysfunction After Extubation

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Objectives: Mechanical ventilation is associated with primary diaphragmatic dysfunction, also termed ventilator-induced diaphragmatic dysfunction. Studies evaluating diaphragmatic function recovery after extubation are lacking. We evaluated early and late recoveries from ventilator-induced diaphragmatic dysfunction in a mouse model.

Design: Experimental randomized study.

Setting: Research laboratory.

Subjects: C57/BL6 mice.

Interventions: Six groups of C57/BL6 mice. Mice were ventilated for 6 hours and then euthanatized immediately ($n = 18$), or 1 ($n = 18$) or 10 days after extubation with ($n = 5$) and without S107 ($n = 16$) treatment. Mice euthanatized immediately after 6 hours of anesthesia ($n = 15$) or after 6 hours of anesthesia and 10 days of recovery ($n = 5$) served as controls.

Measurements and Main Results: For each group, diaphragm force production, posttranslational modification of ryanodine receptor, oxidative stress, proteolysis, and cross-sectional areas were evaluated. After 6 hours of mechanical ventilation, diaphragm force production was decreased by 25–30%, restored to the control levels 1 day after extubation, and secondarily decreased by 20% 10 days after extubation compared with controls. Ryanodine receptor was protein kinase A-hyperphosphorylated, S-nitrosylated, oxidized, and depleted of its stabilizing subunit calstabin-1 6 hours after the onset of the mechanical ventilation, 1 and 10 days after

extubation. Post extubation treatment with S107, a Rycal drug that stabilizes the ryanodine complex, did reverse the loss of diaphragmatic force associated with mechanical ventilation. Total protein oxidation was restored to the control levels 1 day after extubation. Markers of proteolysis including calpain 1 and calpain 2 remained activated 10 days after extubation without significant changes in cross-sectional areas.

Conclusions: We report that mechanical ventilation is associated with a late diaphragmatic dysfunction related to a structural alteration of the ryanodine complex that is reversed with the S107 treatment. (*Crit Care Med* 2020; XX:00–00)

Key Words: mechanical ventilation; ryanodine receptor calcium release channel; Rycal; ventilator-induced diaphragmatic dysfunction; weaning

Diaphragm weakness is associated with the loss of diaphragm force production and is observed up to 64% of patients upon ICU admission (1). Sepsis and mechanical ventilation (MV) are the two main known risk factors of diaphragm weakness (1, 2). The majority of patients with limb muscle weakness do have diaphragm weakness but up to 67% of patients without limb muscle weakness may be diagnose with diaphragm weakness which contributes to under-recognition of diaphragm weakness in ICU patients (3).

Diaphragm weakness is associated with poor outcome, that is, difficult weaning (4, 5) and prolonged ICU stay (1, 4, 5). Since it may affect respiratory function and dependence on ventilatory support, and it may participate in late reintubation, recovery from diaphragm weakness deserves particular attention. However, recovery from diaphragm weakness in the ICU has been sparsely studied. In one cohort of 43 patients, 23 presented diaphragm weakness at admission, where nine of them showed signs of recovery during the ICU stay (6). Along with sepsis and shock, the use of MV by itself leads to diaphragm weakness within a few hours (7, 8): 12 hours in rats (9) and 18 hours in critically ill patients (10). This specific weakness linked with MV has been coined ventilator-induced diaphragmatic dysfunction (VIDD) by Vassilakopoulos and Petrof (11).

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When spontaneous breathing is abolished, VIDD is the consequence of diaphragm unloading and ventilated patient biopsies show signs of sarcomeric disruptions (10), fatty infiltration (12), proteolysis, and autophagy (13), leading to atrophy (10, 14, 15) and mitochondrial oxidative stress (16). Atrophy has been associated with the loss of diaphragmatic force production in numerous animals and in a few human studies (10, 17). Loss of force production with no atrophy, a condition coined dynapenia, is observed in a 6 hours MV mouse model (7, 8) and has been suggested in humans (18, 19). Early diaphragmatic dynapenia during MV is linked to excitation-contraction coupling (ECC) impairment as a consequence of structural remodeling of the type 1 Ca^{2+} release channel/ryanodine receptor 1 (RyR1) (8, 16). Posttranslational modifications including protein kinase A (PKA)-hyperphosphorylation, cysteine oxidation, and nitrosylation dissociate the stabilizing subunit FK506 binding protein 12 (FKBP12) (e.g., calstabin 1) from the Ca^{2+} release channel. RyR1 remodeling, in turn, results in impaired Ca^{2+} handling due to pathologic Ca^{2+} leak from the sarcoplasmic reticulum to the cytosol and is associated with ECC impairment and muscle weakness (8, 16). Although the deleterious impact of MV on diaphragm structure and function has been established, only two studies have evaluated the diaphragmatic function recovery after MV discontinuation (9, 20). Two animal studies have reported a complete recovery in diaphragmatic force production 7 hours to 24 hours after the end of MV, with no data later in the course of recovery (9, 20).

Since prolonged diaphragmatic dysfunction may have an impact on respiratory function, reintubation, and ICU readmission, we aimed to study diaphragmatic structure and function early (1 d) and late (10 d) after MV discontinuation in a mouse model.

MATERIALS AND METHODS

Murine Model of VIDD

The **online supplement** (Supplemental Digital Content 1, <http://links.lww.com/CCM/F806>) contains details regarding ethics approval, diaphragm function evaluation, and the agents used. The present experimental work conforms to the guidelines laid out in the Guide for the Care and Use of Laboratory Animals (www.nap.edu/readingroom/books/labrats) and was approved by the local animal care ethic committee. We used the same study protocol that has been previously described by our group (7, 8). Six groups of adult C57/BL6 mice (12 wk old) were constituted: a control group (euthanasia after anesthesia $n = 15$), MV for 6 hours (MV, $n = 18$), 1 day of recovery following 6 hours of MV (MV + 24 hr, $n = 18$), 10 days of recovery following 6 hours of MV with and without S107 (MV + 10 d + S107, $n = 5$; MV + 10 d, $n = 16$), and a control group with 10 days recovery from anesthesia (control + 10 d, $n = 5$). All in vivo and ex vivo experiments were performed by investigators blinded to treatment groups.

Diaphragm Function

Mice were euthanized by exsanguination and the entire diaphragm was surgically excised. Isometric contractile properties

were assessed as described elsewhere (7). The excised diaphragm strip was mounted into jacketed tissue bath chambers filled with equilibrated and oxygenated Krebs solution. The muscles were supramaximally stimulated using square wave pulses (Model S48; Grass Instruments, Aurora Scientific Inc, Aurora, ON, Canada). The force-frequency relationship was determined by sequentially stimulating the muscles for 600 ms at 10, 20, 30, 50, 60, 80, 100, and 120 Hz with 1 minute between each stimulation train (7). Muscle fatigue was assessed by measuring the loss of force in response to repeated stimuli (30 Hz, 300 ms duration) during 5 minutes as previously reported (7). After measurement of contractile properties, muscles were measured at Lo (the length at which the muscle produced maximal isometric tension), dried, and weighted. For comparative purposes, muscle force production was normalized for total muscle strip cross-sectional area and expressed in N/cm^2 . The total muscle strip cross-sectional area was determined by dividing muscle weight by its length and tissue density ($1.056 \text{ g}/\text{cm}^3$).

Immunoprecipitation

RyR1 was immunoprecipitated from diaphragm using an anti-RyR1-specific antibody ($2 \mu\text{g}$) in 0.5 mL of a modified radioimmune precipitation assay buffer (50 mm Tris-HCl, pH 7.2, 0.9% NaCl, 5.0 mm NaF, 1.0 mm Na_3VO_4 , 1% Triton X-100, and protease inhibitors) overnight at 4°C . RyR1-specific antibody was RyR1-1327, an affinity-purified rabbit polyclonal antibody raised against a KLH-conjugated peptide with the amino acid sequence CAEPDITYENLRRS, corresponding to residues 1,327–1,339 of mouse skeletal RyR1, with an additional cysteine residue added to the amino terminus and affinity purified with the unconjugated peptide. The immune complexes were incubated with protein A-Sepharose beads (Sigma, St. Louis, MO) at 4°C for 1 hour, and the beads were washed three times with radioimmune precipitation assay buffer. The immunoprecipitates were size-fractionated on sodium dodecyl sulfate-polyacrylamide gel electrophoresis gels (4–20% for RyR1 and calstabin 1) and transferred onto nitrocellulose membranes for 2 hours at 200 mA. Immunoblots were developed using the following primary antibodies: anti-RyR1 (1:2,000; Affinity Bioreagents, Golden, CO), antiphospho-RyR-Ser(P) (1:5,000; Affinity Bioreagents), anti-calstabin (FKBP12 C-19, 1:1,000; Santa Cruz Biotechnology, Santa Cruz, CA), and antibody against cysteine nitrosylation (1:1,000; Sigma). To determine channel oxidation, the carbonyl groups in the protein side chains were derivatized to 2,4-dinitrophenol (DNP) by reaction with 2,4-dinitrophenylhydrazine. The DNP signal associated with RyR1 was determined using a specific anti-DNP antibody according to the manufacturer's instructions (Millipore, Billerica, MA). All immunoblots were developed using an Odyssey system (LI-COR Biosciences, Lincoln, NE) with infrared Dy-labeled anti-mouse and anti-rabbit IgG (1:10,000 dilution) secondary antibodies.

Western Blots

Immunoblot analysis was performed to evaluate the ratios of the active (cleaved) forms of calpain 1, 2 and caspase 3 to total

levels of calpain 1, 2 and caspase 3 isoforms, respectively. Total protein ubiquitination was evaluated using polyubiquitinated conjugates antibody as previously described (21). The ratio of LC3/2 to LC3/1 was evaluated as a marker of autophagy as previously reported (22).

Total protein oxidation was measured using an oxyblot kit (Millipore). The carbonyl groups in the protein side chains were derivatized to DNP by reaction with 2,4-dinitrophenylhydrazine. The DNP signal associated with oxidized proteins was determined using a specific anti-DNP antibody according to the manufacturer's instructions (Millipore). GAPDH was used as a loading control. We used primary antibodies as follows: caspase 3 (1:1,000; Abcam Ab13585, Cambridge, MA) and GAPDH (1:2,500; Abcam Ab9485). Calpain 1 (1:1,000; Abcam Ab108400), calpain 2 (1:1,000; Abcam Ab39165), LC3/1-2 (1:1,000; Abcam Ab51520), and ubiquitination (Abcam Ab7780). Secondary antibodies were anti-rabbit 800 nm and anti-mouse 700 nm (1:10,000).

Histology

Cross-sectional area was determined using cryosections (10 μm) and myosin ATPase histochemical staining as previously described (7, 8, 23). Muscle membrane was counterstained with dystrophin antibodies as previously reported (16). The muscle-fiber cross-sectional area was determined using the ImageJ analysis software (<https://imagej.nih.gov/ij/download.html>) and expressed in μm^2 (~900 fibers per group were analyzed). Muscle fibrosis was evaluated using Masson trichrome staining.

Statistical Analysis

Data were presented as means and *SEs* unless otherwise indicated. When more than two groups were compared, one- or two-way analysis of variances or Kruskal-Wallis tests (according to normality) were used. The α level was set at 0.05, corrected for multiple comparisons with the Holm-Sidak test, and was bilaterally tested. Statistical analysis was performed using GraphPad Prism 6.0 (San Diego, CA). The number of mice per group required to identify differences in major physiologic parameters was estimated based on previous experience with this model.

RESULTS

Six Hours of MV in Mice Is Associated With 10 Days Late VIDD

MV is associated with diaphragm dysfunction in a murine model of VIDD. To investigate the mechanisms involved in the recovery from VIDD, sarcoplasmic reticulum fractions from ventilated diaphragm during 6 hours, ventilated diaphragm during 6 hours followed by 1 or 10 days recovery and controls were purified to analyze the composition of RyR1 macromolecular complex and posttranslational modifications known to be associated with channel leak as a proximal mechanism involved in VIDD (8, 16). MV diaphragms exhibit PKA-hyperphosphorylation, oxidation, and cysteine nitrosylation and were depleted of calstabin compared with controls (EFig. 1A–E, Supplemental

Digital Content 2, <http://links.lww.com/CCM/F807>; legend, Supplemental Digital Content 1, <http://links.lww.com/CCM/F806>). Indeed, this called “biochemical signature of leaky RyR1” was persistent 1 and 10 days after MV (EFig. 1 A–E, Supplemental Digital Content 2, <http://links.lww.com/CCM/F807>; legend, Supplemental Digital Content 1, <http://links.lww.com/CCM/F806>). Indeed, diaphragm function, evaluated by measuring the ex vivo-specific force production, was significantly decreased after 6 hours of MV and 10 days after MV compared with the controls (EFig. 1F–J, Supplemental Digital Content 2, <http://links.lww.com/CCM/F807>; legend, Supplemental Digital Content 1, <http://links.lww.com/CCM/F806>). Of note, transitory enhancement of the diaphragm force production 1 day after MV was observed (EFig. 1 H–J, Supplemental Digital Content 2, <http://links.lww.com/CCM/F807>; legend, Supplemental Digital Content 1, <http://links.lww.com/CCM/F806>). Diaphragm fatigue resistance was significantly decreased by 6 hours of MV and 10 days after extubation (EFig. 1K, Supplemental Digital Content 2, <http://links.lww.com/CCM/F807>; legend, Supplemental Digital Content 1, <http://links.lww.com/CCM/F806>).

Late Diaphragm Dysfunction Is Associated With Activation of Ca^{2+} Proteases (Calpain 1 and 2) Without Atrophy or Fibrosis

In order to investigate the mechanism responsible of the late diaphragm dysfunction observed in our model beside the impairment of the Ca^{2+} homeostasis, we evaluated the activity of the two major Ca^{2+} activated proteases, calpain 1 and 2. Calpain 1 was significantly increased 1 and 10 days after extubation compared with the controls while calpain 2 was significantly increased only 10 days after extubation (EFig. 2A–C, Supplemental Digital Content 3, <http://links.lww.com/CCM/F808>; legend, Supplemental Digital Content 1, <http://links.lww.com/CCM/F806>). Indeed, a transitory activation of caspase 3 (EFig. 2A–D, Supplemental Digital Content 3, <http://links.lww.com/CCM/F808>; legend, Supplemental Digital Content 1, <http://links.lww.com/CCM/F806>), autophagy (EFig. 2A–E, Supplemental Digital Content 3, <http://links.lww.com/CCM/F808>; legend, Supplemental Digital Content 1, <http://links.lww.com/CCM/F806>), and proteins oxidation (EFig. 2A–G, Supplemental Digital Content 3, <http://links.lww.com/CCM/F808>; legend, Supplemental Digital Content 1, <http://links.lww.com/CCM/F806>) were observed only after 6 hours of MV. Total proteins ubiquitination was unchanged (EFig. 2A–F, Supplemental Digital Content 3, <http://links.lww.com/CCM/F808>; legend, Supplemental Digital Content 1, <http://links.lww.com/CCM/F806>).

To investigate the histological changes related to Ca^{2+} release impairment and calpains activation, we measured cross-sectional areas and indices of fibrosis among our groups using immunohistochemistry and Masson trichrome staining on 10 μm diaphragm cryosections. Diaphragm cross-sectional areas were comparable among the group without significant changes of the fibers subtype (EFig. 3A–C, Supplemental Digital Content 4, <http://links.lww.com/CCM/F809>; legend, Supplemental Digital Content 1, <http://links.lww.com/CCM/F806>).

Indeed, indices of fibrosis were unchanged among the groups (EFig. 3, D and E, Supplemental Digital Content 4, <http://links.lww.com/CCM/F809>; legend, Supplemental Digital Content 1, <http://links.lww.com/CCM/F806>).

Rycals Drug S107 Prevents the Calstabin 1 Dissociation From RyR1 and Improves Diaphragmatic Force Production

In order to rescue the late diaphragm dysfunction observed 10 days after extubation, we treated our ventilated mice with S107, a Rycal drug which prevents calstabin 1 depletion from RyR1 and reduces pathologic Ca^{2+} leak from the sarcoplasmic reticulum. S107 significantly increased calstabin 1 binding to RyR1 despite the persistence of channel oxidation, phosphorylation, and nitrosylation (EFig. 4A–E, Supplemental Digital Content 5, <http://links.lww.com/CCM/F810>; legend, Supplemental Digital Content 1, <http://links.lww.com/CCM/F806>). S107 also prevented mice from late diaphragm dysfunction 10 days after extubation (EFig. 4F–J, Supplemental Digital Content 5, <http://links.lww.com/CCM/F810>; legend, Supplemental Digital Content 1, <http://links.lww.com/CCM/F806>).

DISCUSSION

We report for the first time in a murine model of VIDD that diaphragmatic dysfunction induced by 6 hours of MV is associated with early recovery, but with late diaphragmatic dysfunction observed 10 days after extubation. Late diaphragmatic dysfunction was associated with RyR1 complex structural remodeling and was reversed by the orally administered Rycal drug S107, a RyR1 complex stabilizer.

Delayed extubation failure leading to reintubation can occur up to 7–10 days after initial weaning success (24, 25), diaphragmatic dysfunction being one of the major mechanisms (26). During critical illness, diaphragmatic dysfunction can result from the consequences of sepsis, muscle unloading, medications, and any other condition associated with ICU acquired weakness (27). Primary diaphragmatic dysfunction can also be related to MV, a condition known as VIDD (11), which has deleterious impact on diaphragm structure, mass (10, 15, 23, 28), and force production (10, 13, 14). Different patterns of diaphragmatic function during critical illness have been described (6), and loss of force production is mitigated by spontaneous breathing (4, 19, 29). Despite two animal studies which have reported an early recovery from VIDD after MV discontinuation (9, 20), long-term follow up studies are scarce.

Our results suggest that even after early recovery from MV, intracellular events persist and are associated with late diaphragmatic dysfunction. We herein report that RyR1 structural remodeling combining oxidation, nitrosylation, inadequate phosphorylation, and calstabin unbound persist up 10 days after MV discontinuation (EFig. 1A–E, Supplemental Digital Content 2, <http://links.lww.com/CCM/F807>; legend, Supplemental Digital Content 1, <http://links.lww.com/CCM/F806>). Maladaptive cAMP-dependent protein kinase A mediated phosphorylation and redox-dependent modifications (nitrosylation and oxidation) of RyR1 have been linked to

the depletion of the stabilizing unit calstabin 1 from the RyR1 channel complex (30, 31). RyR1 channel complex remodeling and instability result in impaired Ca^{2+} handling with a pathologic sarcoplasmic reticulum Ca^{2+} leak associated with diaphragmatic contractile dysfunction and other conditions such as muscle dystrophy, chronic muscle fatigue, and aging. RyR1 channel complex remodeling is the consequence of burst in oxidative stress (EFig. 2, Supplemental Digital Content 3, <http://links.lww.com/CCM/F808>; legend, Supplemental Digital Content 1, <http://links.lww.com/CCM/F806>) (12, 35–37) and adrenergic activation of the protein kinase A (EFig. 5, Supplemental Digital Content 6, <http://links.lww.com/CCM/F870>; legend, Supplemental Digital Content 1, <http://links.lww.com/CCM/F806>), which lead to actin-myosin uncoupling (8), proteolysis, and autophagy activation (EFig. 2, Supplemental Digital Content 3, <http://links.lww.com/CCM/F808>; legend, Supplemental Digital Content 1, <http://links.lww.com/CCM/F806>). Calpain 1 and 2, calcium-dependent proteases implicated in cleaving myofilament proteins, have been associated with diaphragmatic dysfunction (7, 27, 38) and long-term myocardial remodeling post myocardial infarction (39, 40). Calpain 1 and 2 levels were increased 10 days after extubation, possibly secondary both to the oxidative stress burst and the RyR1 sarcoplasmic reticulum calcium leak (EFig. 3, Supplemental Digital Content 4, <http://links.lww.com/CCM/F809>; and EFig. 4, Supplemental Digital Content 5, <http://links.lww.com/CCM/F810> [legend, Supplemental Digital Content 1, <http://links.lww.com/CCM/F806>]) (8).

In the present study, we observed a transient diaphragmatic force recovery 1 day after MV discontinuation despite the presence of RyR1 remodeling. One hypothesis to explain this transient recovery could be that the mitochondria network may have had the buffering capacity to prevent cytosolic Ca^{2+} overload at the early stage (1 d after extubation) (41) but that its capacity might have been insufficient at a later stage (10 d after extubation).

The 6 hours MV mouse model is not associated with histological changes (EFig. 4, Supplemental Digital Content 5, <http://links.lww.com/CCM/F810>; legend, Supplemental Digital Content 1, <http://links.lww.com/CCM/F806>), a finding in accordance with a previous study using the same model (8) and with the moderate correlation between force production and mass (42). Force production depends on muscle mass and also on other factors such as the fiber-to-muscle-length ratio, the intracellular calcium concentration, the amount of myosin heads available for interaction with actin filaments, or the ECC (43).

The persistence of RyR1 dysfunction after 10 days of recovery reinforces the possibility of a pharmacological approach with molecules such as Rycals that are small drug molecules stabilizing the RyR1 which have been previously reported to prevent VIDD in mice (8). In the present study, the use of S107, a Rycal administered in drinking water from MV discontinuation to day 10 was associated with calstabin rebind to RyR1 with no effect on oxidation, nitrosylation, and phosphorylation (EFig. 3, Supplemental Digital Content 4, <http://links.lww.com/CCM/F809>; legend, Supplemental Digital Content 1, <http://links.lww.com/CCM/F806>). The use

of S107 was moreover associated with a complete force production recovery 10 days after MV discontinuation (EFig. 3, Supplemental Digital Content 4, <http://links.lww.com/CCM/F809>; legend, Supplemental Digital Content 1, <http://links.lww.com/CCM/F806>) potentially paving the way toward a treatment for VIDD.

The present study has certain limitations. First, we have evaluated day 1 and day 10 time points and do not present more time points. Second, it would have been of interest to further explore the mechanisms associated with delayed diaphragmatic dysfunction following MV such as the mitochondrial calcium buffering capacity, but this was beyond the scope of this study, which focused on diaphragm function.

CONCLUSIONS

We report for the first time that 6 hours of MV secondarily impairs diaphragmatic function despite a transient recovery 24 hours after extubation. Significant diaphragmatic dysfunction occurs 10 days after MV discontinuation and extubation and is associated with a persistent RyR1 remodeling that is reversible with the use of the orally available S107, a Rycal drug. Further exploration of the mechanisms associated with late diaphragmatic failure and studies in humans and whether S107 would be a drug candidate to treat VIDD are warranted.

Drs. Dridi and Jung contributed equally to this work.

Drs. Dridi, Jung, Reiken, Marks, Lacampagne, and Jaber designed experiments, analyzed data, and edit/wrote the article. Drs. Yehya, Daurat, Reiken, and Moreau designed experiments and analyzed data.

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