

## The distribution of Silicon in soil is influenced by termite bioturbation in South Indian forest soils

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- 1 The distribution of Silicon in soil is influenced by termite bioturbation in South
- 2 Indian forest soils

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#### **ABSTRACT**

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Si is one of the most abundant element on earth and an abundant literature shows its beneficial effects on plant growth and resistance. We here question the influence of termites, as key soil bioturbators, on the distribution of Si in a tropical soil. The abundance and forms of Si in termite mounds build by Odontotermes obesus (TM) or in the soil eroded from TM but redistributed on the ground surface (EROD) were compared to those measured in the 0-5 (Ctrl<sub>0-5</sub>) and 70-120 cm soil layers (Ctrl<sub>70-120</sub>). Although termites use the soil from Ctrl<sub>70-120</sub> for building their mounds, we found that TM and EROD had intermediate soil physical, chemical and mineralogical properties between Ctrl<sub>0-5</sub> and Ctrl<sub>70-120</sub>. Clay content was not significantly different between soil materials. However, the lower variability measured in TM than in the soil suggested that termites used soil layers with higher amounts of clay fraction and with a preference especially for layers enriched in 2:1 clay minerals (smectite) most likely because they provide better physical properties in terms of plasticity and water retention than kaolinite. Finally, phytoliths and bioavailable Si (Si<sub>CC</sub>) contents were increased in TM in comparison with Ctrl<sub>70-120</sub>, suggesting an incorporation of phytoliths in termite construction through their saliva and/or an increasing availability of Si<sub>CC</sub> from the minerals. In conclusion, this study highlights how termites, through their feeding and building activities, impact Si distribution in tropical soils.

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41 *Keywords*: Si, clay, mineralogy, phytoliths, bioturbation, soil profile

1. Introduction

Silicon (Si) is the second most abundant element in the earth's crust after oxygen, with approximately 28 and 47% of its mass, respectively (Singer and Munns, 1999). It is not included in the list of elements essential for plants but there is an abundant literature showing its beneficial effects on the resistance of plants to parasites and drought (Liang et al., 2015; Datnoff et al., 2001).

In soil Si is a major constituent of parent rock minerals (such as quartz, feldspars) and of secondary minerals originated from pedogenesis, such as clay minerals and amorphous silica. Biogenic amorphous silica particles can also be found in soil as plant and microorganism remains, i.e., phytoliths (Cornelis and Delvaux, 2016) and diatom frustules or thecamoebian tests (Sommer et al., 2006). These amorphous silica particles can play an important role in the cycling of Si because they constitute a pool of highly reactive silica (Fraysse et al., 2009). Large

fauna like grazers can also affect the cycling of Si through the accelerating transfer of plant Si

to aquatic environments (Schoelynck et al., 2019) or through the increase production of

et al., 2019), and there is a dearth of information on how soil macrofauna impacts clay mineralogy and the availability of Si for plants.

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Amongst the soil macrofauna, organisms from the soil engineering group (sensu Lavelle et al., 1997) impact soil functioning at different spatial and temporal scales (Bottinelli et al., 2015). This role is played by termites in tropical dryland ecosystems, and especially by termites from the fungus-growing functional group (Macrotermitinae subfamily) in Africa and Asia (Jouquet et al., 2016). Termites are key decomposers of the litter in tropical environments (Amadou Issoufou et al., 2019) and their building activity is associated to the redistribution of soil organic matter and elements in their biomass and organo-mineral constructions. Studies carried out in Africa and Asia showed that fungus-growing termites impact the distribution, the type of clay minerals (kaolinite vs. smectite) and the mineralogical properties of phyllosilicates (Jouquet et al., 2002, 2007). Termites are also known to impact the dynamic of Si at the profile scale through the selective use of material coming from the deep soil layers within their nests, then resulting in the redistribution of clay minerals and elements (Holt and Lepage, 2000; Abe et al., 2012; Mujinya et al., 2010, 2011, 2013; Erens et al., 2015). However, the depth at which termites fetch the clay particles remains mostly unknown and termite bioturbation and the upward translocation of soil from the deeper layers to the surface are likely to be associated with a redistribution of Si<sub>CC</sub> and Si<sub>AA</sub> pools in termite mounds and thereafter in the surrounding soil environment with the erosion of termite mounds.

In Southern India, the termite species *Odontotermes obesus* is known to play an important role in the decomposition of litter (Shanbhag and Sundararaj, 2013) and its mounds are conspicuous features of the landscapes (Shanbhag et al., 2017). These mounds are especially abundant in protected areas such as in the deciduous forests of the Bandipur Tiger Reserve. In this environment termite mounds have specific soil physical and chemical properties (Jouquet et al., 2017a) and a recent study showed that they are mainly made of soil collected between 70

to 120 cm depth (Jouquet et al., 2017b). The aim of this study was therefore to determine the impact of *O. obesus* bioturbation on the distribution of Si in soil. Our hypothesis was that termite feeding and building activities (i.e., the construction of mounds from material selected in the deep soil layers) participate in a selective redistribution of clay minerals, phytoliths and available Si (Sicc.), which are thereafter redistributed to the soil surface.

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#### **Material and Methods**

100 Study site

Soils were sampled in the Mule Hole experimental watershed (4.1 km<sup>2</sup>). This dry deciduous forest is located in the Bandipur Tiger reserve in Southern India at 11°44'N and 76°26'E (Chamarajanagar district, Karnataka state). The Mule Hole watershed is located in the subhumid zone of the sharp climatic gradient induced by the Western Ghats. As a result of the short-term variability of the South-West Monsoon, the experimental watershed is characterized by rainfall ranging from 1000 to 1500 mm yr<sup>-1</sup>, usually concentrated during the monsoon from June to August and from October to November. The mean annual temperature is 27°C (Braun et al., 2009). The elevation of the watershed ranges from 820 to 910 m a.s.l. The relief is mostly undulating with gentle slopes. The soil cover of the watershed has been mapped by Barbiero et al. (2007) based on the FAO terminology (IUSS-Working-Group-WRB, 2015). Parent rocks consist of peninsular gneiss intermingled with much less abundant mafic to ultramafic rocks, mostly amphibolite. Soil depth is variable but ranging from 1 to 3 m in average (Barbiero et al., 2007; Braun et al., 2009). The main soil types are Ferralsols and Chromic Luvisols, with clay mineralogy dominated by kaolinite (1:1 clay) and characterized by an accumulation of iron and aluminum oxides. The soils have less than 20 g kg<sup>-1</sup> of C in the first soil layer and < 10 g kg<sup>-1</sup> below 20 cm depth. The clay content ranges from 30 to 40% and clays minerals are also dominated by 1:1 clays (kaolinite) (Jouquet et al., 2016b). The vegetation is a dry deciduous forest. In this environment, cathedral-shaped termite mounds are built by O. obesus (Chhotani

and Bose, 1979; Jouquet et al., 2016a,b, 2017a) (Figure 1). The soil of termite mounds has similar clay contents but it is impoverished in C (0.54-fold) in comparison with the surrounding topsoil (Jouquet et al., 2016, 2017a,b).

## Soil sampling

Termite mounds built by *O. obesus* vary considerably in size (Shanbhag et al., 2017) and only large mounds from 1.5 to 1.8 m high and > 0.5 m<sup>3</sup> were considered in this study. Samples were collected from the outer-wall of three termite mounds (TM) and from their periphery in a distance of about 15 cm (impacted by mound erosion, EROD). Soils were also sampled in the surrounding soil environment (control) in a distance > 5–10 m ahead of the mounds in taking into account the gentle slope of the watershed (not impacted by mound erosion) at several soil depths down to 4.0 m in three different locations (n = 39). Data were grouped according to their depths (Ctrl<sub>0-5</sub>: 0–5 cm, Ctrl<sub>70-120</sub>: 70-120 cm) with the assumption that termites collect the soil mainly from the 70-120 cm soil layer for building their mounds (Jouquet et al., 2017b).

#### Analysis of soil properties

The soil physical properties measured included the particle-size distribution, after destruction of organic matter and dispersion with sodium hexametaphosphate (AFNOR, NFX 31 107): clay (< 2  $\mu$ m) and silt (2–50  $\mu$ m) were obtained by pipette method and sand (50–2000  $\mu$ m) by sieving. To assess Soil Organic Matter (SOM) content, the C and N concentrations were measured using an elemental analyzer Flash 2000 HT. The cationic exchange capacity and the content of exchangeable cations (Ca, Mg, Na, K, Fe, Mn and Al) were determined at soil pH by exchange with cobaltihexamine (AFNOR, NF ISO 23470). The amount of SiO<sub>2</sub>, Fe<sub>2</sub>O<sub>3</sub> and Al<sub>2</sub>O<sub>3</sub> were measured by ICP-OES after fusion with lithium metaborate LiBO<sub>2</sub> and subsequent dissolution in a mixture of HNO<sub>3</sub> (1 mol L-1), H<sub>2</sub>O<sub>2</sub> (0.5 % v/v) and glycerol (10 % v / v) at the

CNRS Service d'Analyse des Roches et des Minéraux (SARM) (Carignan et al., 2001). All these analyses were carried out on soil aggregates >2 mm that were ground before analyses to homogenize the material.

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Si pools and phytoliths

Plant available Si was measured in comparing the pools of readily soluble Si extracted by 0.01 M CaCl<sub>2</sub> (Si<sub>CC</sub>, Haysom and Chapman, 1975) and the pool of adsorbed Si extracted by 0.5 M acetate-acetic acid (Si<sub>AA</sub>, Korndörfer and Lepsch, 2001; Narayanaswamy and Prakash, 2009, 2010). Phytoliths were extracted using HCl (33%) to remove carbonates, 2/3 nitric plus 1/3 perchloric acid to remove organic matter. Soil clays were removed by repeated decantations, and separation of biogenic silica particles from heavy minerals was achieved by densimetric separation using  $ZnBr_2$  set at d = 2.3 g cm<sup>-3</sup>. The extracts were weighted and phytoliths were mounted on slides for observation under optical microscope. More than 250 phytoliths were counted per sample. Morphotypes with clear taxonomical identification were silica short cell phytoliths (rondel, bilobate, polylobate and cross) from Poaceae (grasses), spheroid psilate and decorated from woody dicotyledons (considered forest indicator phytoliths), and spheroid echinate from Arecaceae (palms). Other phytoliths included bulliform flabellate, acute bulbosus (silicified hair cells), and various elongate and blocky morphotypes which were mostly nondiagnostic (International Code for Phytolith Nomenclature, 2019). The ratio of spheroid decorated versus the sum of grass silica short cells (or D/P° index) increases with the increasing abundance of trees and shrubs versus grasses and is commonly used to trace the tropical forest/savanna boundary (Bremond et al. 2008). On the same slides, but in a separate counting session we measured the proportion of corroded vs. plain phytoliths to identify the origin of phytoliths with the assumption that phytoliths at depth likely underwent prolonged dissolution

- than the more recent phytoliths located at the soil surface following Sommer et al. (2006).
- Phytoliths relative abundance were calculated based on the total phytolith sums.

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- 171 Clay mineralogy
- Clay mineralogy from termite mound soil samples (TM and EROD) was compared to the 172 mineralogy of the soil at 0-5 and 70-120 cm depths ( $Ctrl_{0-5}$  and  $Ctrl_{70-120}$ ) (n = 3 per treatments). 173 Clay mineralogy was determined after extraction of the clay size fraction (particles < 2 µm) 174 from the bulk soil samples. The fractions < 50 µm were obtained by wet sieving and then 175 dispersed into 1 mol L<sup>-1</sup> NaCl (five saturations) and ultrasonication. The < 2 µm fractions were 176 obtained by sedimentation and were subsequently flocculated and saturated with  $0.5 \text{ mol } \text{L}^{-1}$ 177 CaCl<sub>2</sub> (Caner et al., 2014). XRD patterns of oriented preparations were recorded in the air-dried 178 state (Ca-AD) and after saturation with ethylene glycol (Ca-EG) to identify the clay minerals 179 (Caner et al., 2014). Since no additional information was obtained from the saturation with 180 ethylene-glycol, only results from the Ca-AD preparation was shown. The X-Ray diffraction 181 patterns were decomposed into elementary peaks (Lanson, 1997) using Fityk software over the 182 3-14°20 range. The procedure included removal of baseline and decomposition of the signal 183 into Gaussian elementary curves. The diffractograms were fitted with the minimum curves to 184 obtain a reliable fit of the experimental pattern. The parameters of the elementary curves 185 (position, full-width at half maximum – FWHM and maximum intensity, surface area) were 186 used to identify the soil clay mineral (kaolinite, illite, smectite, mixed layers illite/smectite) 187 species according to Righi et al. (1995) and Velde (2001). A semi-quantitative phase analysis 188 was performed using the relative surface peak area of each peak of clay minerals over the sum 189 of the peak surface areas. The relative surface peak area of each peak does not correspond to 190 the proportion of the respective mineral, however, the variations of relative surface areas 191 correspond to variations in mineral quantities if the samples comes from the same sequence 192

(Lanson 1997). Peaks at ~7.20 and 7.30-7.40 Å were attributed to kaolinite (K). The peak of illite at ~10 Å was decomposed into well crystallized illite (WCI) at 10.0 Å and poorly crystallized illite (PCI) at 10.2-10.4 Å. The peaks at ~15 Å in AD state that displace to ~17 Å were attributed to smectite (S) and the peaks between 10 and 15 Å in AD state that displaced between 10 and 17 Å in EG were attributed to mixed layers illite/smectite with variable proportion of the two components.

### Statistical analyses

Residues were tested for normality of variance using the Shapiro–Wilk test and log10-transformed if required. Soil chemical and physical properties were analyzed by analysis of variance (ANOVA) with treatments as independent variables. LSD tests were then performed to assess differences between means. Post-hoc planned pairwise comparisons were performed by Wilcoxon–Mann–Whitney U tests when parametric ANOVA were impossible to use, even after data transformation. Impact of the treatments on soil variables and soil mineralogy were analyzed using a principal component analysis (PCA). Correlations between variables and the amount in Si<sub>CC</sub>, Si<sub>AA</sub> and phytoliths were carried out using the spearman test. 95% confidence intervals associated with the D/P° ratios were calculated following Suchéras-Marx et al. (2019). All statistical parameters were calculated using R studio and R version 3.2.1 with ade4, factoextra, factoMineR and corrplot packages. Differences among treatments were declared significant at the 0.05 probability level.

#### Results

- Difference in soil chemical and physical properties
- The chemical properties of the 4 soil materials are shown in Table 1 and a PCA-projection of the samples from significant variables is shown in Figure 1. Soil materials were mainly

differentiated along the first axis while the variability within the samples was evidenced on the second axis, which explained 63.0 and 18.5% of the total variability, respectively. Samples varied from the soil surface (Ctrl<sub>0-5</sub>) to TM, EROD and finally Ctrl<sub>70-120</sub>. No significant difference in soil particle size fractions, in pH and in exchangeable Mg<sup>2+</sup> and K<sup>+</sup> contents could be measured between the four soil materials (P > 0.05 in all cases) (Table 1). Ctrl<sub>0-5</sub> was enriched in C and N and characterized by a greater CEC and exchangeable Ca<sup>2+</sup> content but lower exchangeable Al<sup>3+</sup> content in comparison with the other treatments. Conversely, Ctrl<sub>70-</sub> 120 were impoverished in C and N but enriched in exchangeable Na<sup>+</sup>, Al<sub>2</sub>O<sub>3</sub> and Fe<sub>2</sub>O<sub>3</sub> in comparison with the other soil materials. In general, TM and EROD had intermediate properties in comparison with the Ctrl<sub>0-5</sub> and Ctrl<sub>70-120</sub> (Figure 1) and no significant difference occurred between them, with the exception of the concentration in exchangeable Mn<sup>2+</sup> that was similar in TM and  $Ctrl_{70-120}$  but higher than in EROD and  $Ctrl_{0-5}$  (P > 0.05 between them, Table 1). Figure 2 shows the evolution of clay content in TM and EROD and at different soil depths. The amount of clay was highly variable (e.g. from < 10 to > 50% at 150 cm depth), especially in comparison with the clay content in TM and EROD. Despite this high variability, an increase in clay content was measured at about 50-150 cm depth.

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#### Si pools and phytoliths

Figure 3 shows that the proportion of  $SiO_2$  and the amount of  $Si_{CC}$  were the lowest in  $Ctrl_{70-120}$ , without any significant difference between the other treatments (P > 0.05). Conversely  $Si_{AA}$  content was the highest in  $Ctrl_{70-120}$  and the lowest in  $Ctrl_{0-5}$  while TM and EROD had intermediate positions. The proportion of phytoliths was low with percentages < 1%. The highest value was measured in  $Ctrl_{0-5}$  in comparison with  $Ctrl_{70-120}$  and TM (P < 0.05 in both cases). Conversely, the lowest value was measured in  $Ctrl_{70-120}$  in comparison with  $Ctrl_{0-5}$  and EROD (P < 0.05 in both cases). TM and EROD had a similar percentage of phytoliths with

intermediate values in comparison with the two control soils. Si<sub>CC</sub> was not correlated with the soil physical and chemical properties, with the exception of a positive correlation with phytoliths (P = 0.028) and Mg<sup>2+</sup> (P = 0.017). Conversely, Si<sub>AA</sub> was negatively correlated with the amounts in C, N, the CEC and Si:Al ratio (P = 0.048, 0.048, 0.049 and 0.013, respectively) but positively correlated with the total amount in  $Al_2O_3$  (P = 0.013). Moreover, the amount in phytoliths was positively correlated with the CEC and the amounts in C, N, Ca<sup>2+</sup>, Mg<sup>2+</sup> and  $Si_{CC}$  (P = 0.001, 0.008, 0.008, 0.001, 0.001 and 0.028, respectively) but negatively correlated to the amount in  $Fe^{3+}$  and  $Al^{3+}$  (P = 0.018 and 0.011, respectively).

Identification of the phytoliths shows that most of them came from grasses and woody dicotyledons (trees and shrubs), with little differences in the phytolith-inferred vegetation signal between Ctrl<sub>0-5</sub> and Crtl<sub>70-120</sub>, but strong difference with TM. In TM, phytolith pool was essentially coming from grasses (phytolith ratio Dicotyledons:Poaceae, D:P° < 0.7), while it was coming from woody plants in the Crtl samples (D: $P^{\circ} > 2$ ) (Figure 4 and see appendix A for the detailed phytolith counts). A gradient of corrosion was observed from the top-soil to the deeper soil layers with a dominance of non-corroded phytoliths in Ctrl<sub>0-5</sub>, a dominance of corroded phytoliths in the 90-100 cm soil layer and intermediate values in the 20-40 cm soil layer (Figure 5). In TM, phytoliths had intermediate proportion of corroded phytoliths than between the 0-5 and 20-40 cm soil layers (Figures 5 and 6).

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Variation in clay abundance and mineralogy with soil depth

Differences in clay mineralogy between soil samples are shown in Figure 7. Soil materials were mainly differentiated by the second axis, which explained 30% of the total variability. Samples from the topsoil were enriched in smectite (S) and poorly crystallized illite (PCI) in comparison with soil sampled below 70 cm depth that were rich in kaolinite (K). TM and EROD had the

same signatures and intermediate properties between those of the soils sampled in  $Ctrl_{0-5}$  and  $Ctrl_{70-120}$ .

#### Discussion

Bioturbation and translocation of clay

Bioturbation and the edification of termite mounds by fungus-growing termites is associated with a translocation of soil material from the deeper soil layers to the surface. Usually evidenced by an enrichment in clay particles or the presence of minerals that are less abundant than in the topsoil, this process of translocation as well as the depth at which termites collect the soil remain poorly quantified. In a recent study, we showed that the soil from the outer-wall of *O. obesus* mounds mainly comes from the ~ 70 to 120 cm soil layer (Jouquet et al., 2017b). Here, measure of the percentage of clay fraction in soil did not evidence a significant difference between soil materials, mainly because of the high variability in clay content along the soil profile. This variability in clay content was reduced in termite mound soils, showing a possible regulation of the amount of clay fraction in soil by termites for strengthening cohesion between aggregate and ensuring the resistance of TM, as evidenced in laboratory conditions (Jouquet et al., 2002, Oberst et al., 2016) and for a better control of their habitat and resistance to predators (Jouquet et al., 2006).

#### Si available pools

The 0-5 cm soil layer was enriched in SiO<sub>2</sub> in comparison with the 70-120 cm soil layer, which was enriched in Al<sub>2</sub>O<sub>3</sub> content (see the supplementary file 1 showing the higher concentration in Al<sub>2</sub>O<sub>3</sub> at this soil depth). Phytoliths in the 70-120 cm soil layer were also less abundant and more corroded than in the topsoil in a good agreement with previous observations (Alexandre et al., 1997; Sommer et al., 2006). Indeed, the dynamic of phytoliths in soil results from burial, illuviation and bioturbation processes and phytoliths from deeper horizons are usually less

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abundant, older and more corroded than in the surface (Sommer et al. 2006; White et al., 2012). A similar amount of phytoliths was measured in TM and in the topsoil while EROD had intermediate phytolith contents between TM and Ctrl<sub>70-120</sub>. Difference in D:P° ratio between TM and Ctrl soils also shows that TM contained more phytoliths from grasses than control soil samples. Although this study does not allow to reject the assumption that phytoliths from deeper soil layers were selectively incorporated into TM, we consider this hypothesis unlikely since this would have resulted in higher representation of forest indicator phytoliths, as forest is the dominant vegetation at Mule Hole (Riotte et al., 2018). Moreover, phytolith morphotypes vary in size but grass silica short cell phytoliths and the forest indicator spheroids are in the same size range (8-30 microns). It is therefore unlikely that grass phytoliths were incorporated in TM because they were smaller and because they were differently associated to clay particles. Despite the fact that O. obesus can feed on a variety of litter components (e.g. elephant dung, wood and leaves) (Cheik et al., 2018, 2019; Shanbhag et al., 2019), grasses are mostly consumed by herbivores in Mule Hole catchment. The presence of grass phytoliths in TM therefore suggests that termites act as primary decomposers of herbivore dungs in the studied area. This hypothesis is in line with recent studies carried out in southern India which showed that elephant dung material is very attractive to termites which prefer this material over carton, wood or leaves (Cheik et al., 2018, 2019; Shanbhag et al., 2019). Indeed, despite inhabiting forested environments in Karnataka and the Nilgiri Region, elephants preferentially feed on Panicoideae grasses which constitute more than 84% of their annual bulk diet (Baskaran et al. 2010). A large amount of these grasses is only partially decomposed, making this material very attractive to termites. Therefore, if the consumption of grasses is associated with a partial digestion of organic matter by termites for building their biomass and ensuring their activity. the Si contained in the phytoliths is not used by termites. From these findings we suggest that termites incorporate phytoliths in their constructions most likely when they moisten the soil

aggregate pellets with saliva and potentially faeces. This hypothesis is consistent with the presence of monosaccharides from plant origin in the epigeous nest of fungus-growing termites (Sall et al., 2002) and the higher C content measured in TM in comparison with Ctrl<sub>70-120</sub> and the positive correlation between phytoliths and the C content. Moreover, since phytoliths were mainly non-corroded phytoliths in TM, we even suggest that phytoliths resist to termite and microbial (including *Termitomyces* sp.) metabolisms.

 $Si_{CC}$  and  $Si_{AA}$  were respectively higher and lower in the topsoil layer in comparison with Ctrl<sub>70-120</sub>. Interestingly, despite the fact that TM was mainly built from the soil at 70-120 cm depth, TM had a closer signature with the topsoil than with the deeper soil with similar amounts of  $SiO_2$ ,  $Si_{CC}$  and  $Si_{AA}$ . Although this study does not allow to disentangle the mechanisms associated to this change in Si pools, the positive correlation between  $Si_{AA}$  and the amount of  $Al_2O_3$ , but not the amount in clay, and the positive correlation between  $Si_{CC}$  and phytoliths suggest either (i) a total reorganization of the structure of soil pellets and a decrease in the interactions between Si and oxides and a shift from  $Si_{AA}$  to  $Si_{CC}$ , (ii) a production of  $Si_{CC}$  from phytolith dissolution, and/or (iii) a selection of particles from  $Ctrl_{70-120}$  that were impoverished in oxides (see supplementary file 2) but enriched in clay and  $Si_{CC}$ . Another explanation could be that the dissolution of 2:1 clay minerals, mainly found on the topsoil and TM, increases the  $Si_{CC}$  pool, even if this process is likely to be limited with a soil pH > 6. This explanation is in line with Violette et al. (2010) who found that the dissolution of smectite provides large amount of dissolved silica (i.e., 75% of the export of dissolved silica) in the study watershed.

#### Modification in clay mineralogy

The 70-120 cm soil layer was characterized by a higher percentage of clay fraction and higher contents in kaolinite and Al<sub>2</sub>O<sub>3</sub> that are characteristics of kandic horizons (Brady and Weil, 1999; Soil Survey Staff, 2014). The presence of the kandic horizon was confirmed with the X-

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ray diffraction data that revealed the presence of smectites and poorly crystallized illite (i.e. 2:1 phyllosilicates) in the topsoil and on the contrary a mineralogy dominated kaolinite (1:1 phyllosilicate) in the kandic horizon. The presence of smectite on the topsoil was unexpected and suggests a high level bioturbation activity with the upward movement of soil and neoformation processes. The concentration of Mn<sup>2+</sup> in soil solutions is largely controlled by oxidation and reduction reactions and the higher concentration of this cation in the kandic horizon also confirms a process of accumulation at this soil depth. Therefore, similar concentration in Mn<sup>2+</sup> between the 70-120 cm soil layer and TM confirms the utilization of soil from the kandic horizon for building termite mounds. However, the lower concentration in Mn<sup>2+</sup> in EROD is likely to be explained by the high solubility of this cation in solution (Gilkes and McKenzie, 1988) and its rapid leaching from the soil eroded from the mound, despite the fact that this soil is poorly permeable in comparison with the surrounding topsoil (Traoré et al., 2019). Although TM were mainly made of soil coming from the kandic horizon, clay minerals in termite mounds had intermediate properties between those observed in the topsoil and those sampled in the deeper soil layers. Differences in clay minerals or in Si:Al ratio between TM and the surrounding topsoil have been evidenced in different environment (e.g., Wielemaker, 1984; Abe et al., 2012). They are usually explained by the use of specific clay minerals and/or the ability of termites to modify the properties of clay minerals (Schaefer, 2001; Jouquet et al., 2011). Since the modification of clay minerals by termites is limited to the swelling ability of 2:1 clay minerals (Jouquet et al., 2002; 2007), our results suggest that 2:1 clay minerals may be selected by O. obesus for building their nests than kaolinite, most likely because of their better physical properties in terms of plasticity and water retention (Andrade et al., 2011; Spagnoli et al., 2018). Another explanation could be 2:1 clay minerals found in TM could be produced from the dissolution of phytoliths and the interaction between Si<sub>CC</sub> and kaolinite (Sommer et al., 2006). In a highly weathered Oxisols in the Amazon forest, Lucas et al. (1993) showed that the

occurrence of kaolinite, instead of gibbsite, in the topsoil (or surface horizon) was attributed to the resilication of gibbsite by the soluble silica provided by phytolith dissolution. A similar process of resilication of kaolinite could be suggested for the formation of smectite in TM by the biological uplift of cations and highly reactive silica from the dissolution of phytoliths. Over longer period of time, the accumulation of 2:1 clay minerals in TM is likely to be important at the ecosystem scale. Indeed, *O. obesus* mound turnover rate has been estimated to reach a soil turnover of 3.1-31 mm ky<sup>-1</sup> in the same study site (Jouquet et al., 2017). Since clay minerals accumulated in termite mounds return to the surrounding soil environment after termite mound degradation, further research is needed to determine if the accumulation of this material contributes and/or explains the presence of 2:1 clay minerals in the topsoil.

#### Conclusion

Findings from this study show that *O. obesus* has a significant impact on the distribution of Si in soil. First, we explained the lower variability in clay content in comparison with the high variability measured along the soil profile by the need of termites to control the properties of their nests, i.e., the amount of clay fraction in TM, then highlighting the importance of this material for termites. The higher phytolith content in TM was unexpected but could be explained by termite foraging activity (i.e., the consumption of grasses) and the excretion of phytolith in termite saliva. Higher amount in Si<sub>CC</sub> was also surprising and raised the question of a possible impact of termites on the interaction between Si and minerals in soil. In perspective, we consider that more research is now needed to better understand the fate of the phytoliths in TM. Further studies are also needed to determine if the increase in Si<sub>CC</sub> content in TM contribute to explain the presence of 2:1 minerals in termite constructions (Jouquet et al., 2002) and the specific vegetation growing in the vicinity or on termite mounds in Africa (e.g.,

| 393 | Traoré et al., 2015), as well as the utilization of TM soil as amendment for increasing soil   |
|-----|--|
| 394 | fertility (Loko et al., 2017).   |
| 395 |  |
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| 401 | Recherche en Environnement - Bassin Versant Expérimentaux Tropicaux, <a href="http://bvet.omp.obs-">http://bvet.omp.obs-</a>           |
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| 407 |  |
| 408 | Appendix A. Supplementary data to this article can be found online at  |
| 409 | Bardoni, D.; Meunier, J-D.; Jouquet, P., 2019, Phytoliths in termite mounds and in the soil  |
| 410 | environment, India, 2017. <a href="https://doi.org/10.23708/G3HWON">https://doi.org/10.23708/G3HWON</a> ,                              |
| 411 | Jouquet, P.; Jamoteau, F.; Majumdar, S.; Nagabovanalli, P.; Caner, L.; Meunier, J-D., 2019,  |
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| 413 | and the soil environment in Mule Hole, Karnataka, India, <a href="https://doi.org/10.23708/MHUF7F">https://doi.org/10.23708/MHUF7F</a> |
| 414 |  |
| 415 |  |
| 416 |  |

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| <b>Figures</b> | and | table | captions |
|----------------|-----|-------|----------|
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Figure 1. Biplot showing the principal components analysis (PCA) from variables describing soil physical and chemical properties showed in Table 1 and significantly influenced by the different treatments: control soil sampled at 0-5 and 70-120 cm depth in blue and yellow, respectively, and the termite mound soil (TM, in red) or the soil eroded from the mound (EROD, in grey).

Figure 2. Clay content along the soil profile, in the outer part of termite mound (TM, arbitrary displayed at + 50cm depth) and in the soil eroded from TM (EROD, arbitrary displayed at + 5 cm depth). The three different soil profiles are given in three different soil colors.

Figure 3. Histograms showing the percentages in  $SiO_2$  and phytoliths (%), and the amounts in  $Si_{CC}$  and  $Si_{AA}$  (ppm) in the outer part of termite mound (TM), in the soil eroded from TM (EROD) and in the control soils at 0-5 and 70-120 cm depths (Ctrl<sub>0-5</sub> and Ctrl<sub>70-120</sub>, respectively). Bars represent standard errors and histograms with the same letters are not significantly different.

- Figure 4. Abundance and diversity of phytoliths.
- (a) Histograms showing the relative abundances of phytoliths in termite mound (TM) and control samples at 0-5 (Ctrl<sub>0-5</sub>) and 70-120 (Crtl<sub>70-120</sub>) cm depths. The differentiation was made into forest indicators, grasses, ND-elongate and ND-blocky phytoliths.
  - (b) Ratio of spheroid decorated versus the sum of grass silica short cells (or D/P° index).

| 626 |           |   |
|-----|-----------|---|
| 627 | Figure 5. | Proportion of corroded, non-corroded and undetermined phytoliths for termite  |
| 628 |           | mound (TM) and in the soil sampled at 0-5, 20-40 and 70-120 cm depths ( $Ctrl_{0-5}$ and  |
| 629 |           | $Ctrl_{70-120}$ ).  |
| 630 |           |   |
| 631 | Figure 6. | Examples of blocky and elongated non-corroded (a, b) and corroded phytoliths (c, d)   |
| 632 |           | obtained in the microscope. The scale is given with the horizontal bar which  |
| 633 |           | represents 40 mm.   |
| 634 |           |   |
| 635 | Figure 7. | Biplot showing the principal components analysis (PCA) from variables describing  |
| 636 |           | clay mineralogy of the different soil materials. Treatments are the soil at 0-5 and 70-   |
| 637 |           | 120 cm depth (in blue and yellow, respectively), the soil from termite mound (TM,   |
| 638 |           | in red) and the soil eroded from TM (EROD, in grey). Clays are kaolinite (K), well  |
| 639 |           | crystallized illite (WCI), poorly crystallised illite (PCI), inters-stratified (IS) and   |
| 640 |           | smectite (S).   |
| 641 |           |   |
| 642 | Suppleme  | entary file 1. Percentage in Fe <sub>2</sub> O <sub>3</sub> and Al <sub>2</sub> O <sub>3</sub> along the soil profile, in the outer part of |
| 643 |           | termite mound (TM, arbitrary displayed at + 50 cm depth) and in the soil eroded from  |
| 644 |           | TM (EROD, arbitrary displayed at + 5 cm depth).   |
| 645 |           |   |
| 646 | Suppleme  | entary file 2. Percentage of well crystallized oxides in soil (Fe <sub>DCB</sub> – Fe <sub>O</sub> and Al <sub>DCB</sub> –                  |
| 647 |           | $Al_{O}$ ) extracted by the DCB (Fe <sub>DCB</sub> and $Al_{DCB}$ ) and oxalate methods (Fe <sub>O</sub> and $Al_{O}$ )                     |
| 648 |           | (Mehra and Jackson, 1960; Schwertmann, 1964) along the soil profile, in the outer   |

part of termite mound (TM, arbitrary displayed at + 50 cm depth) and in the soil

eroded from TM (EROD, arbitrary displayed at + 5 cm depth).

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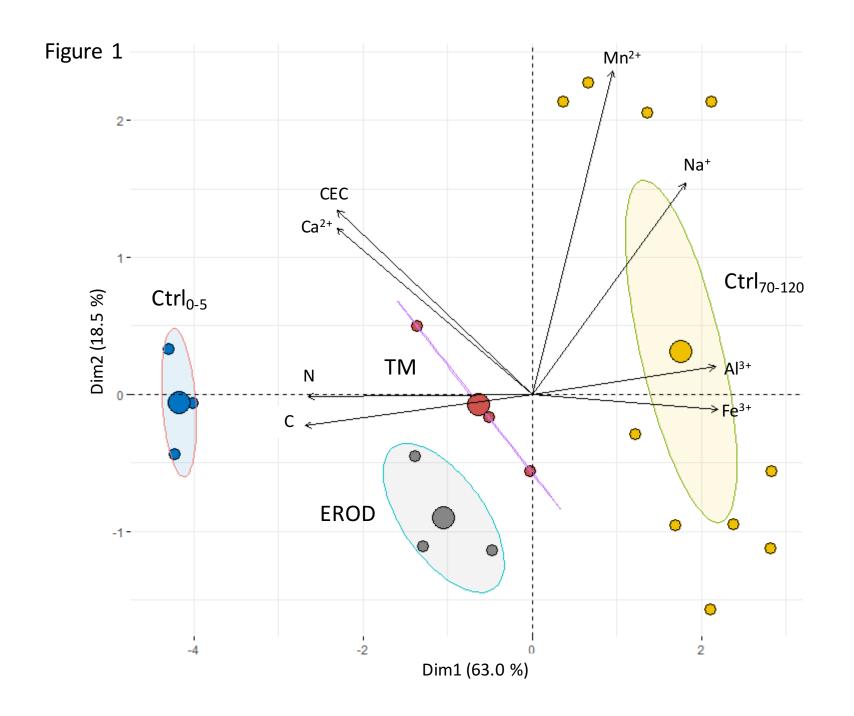


Figure 2

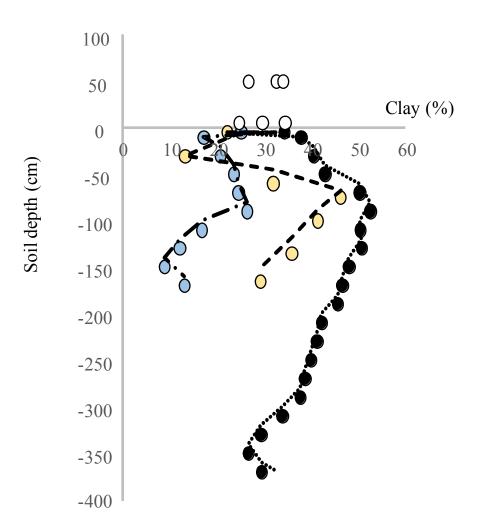


Figure 3

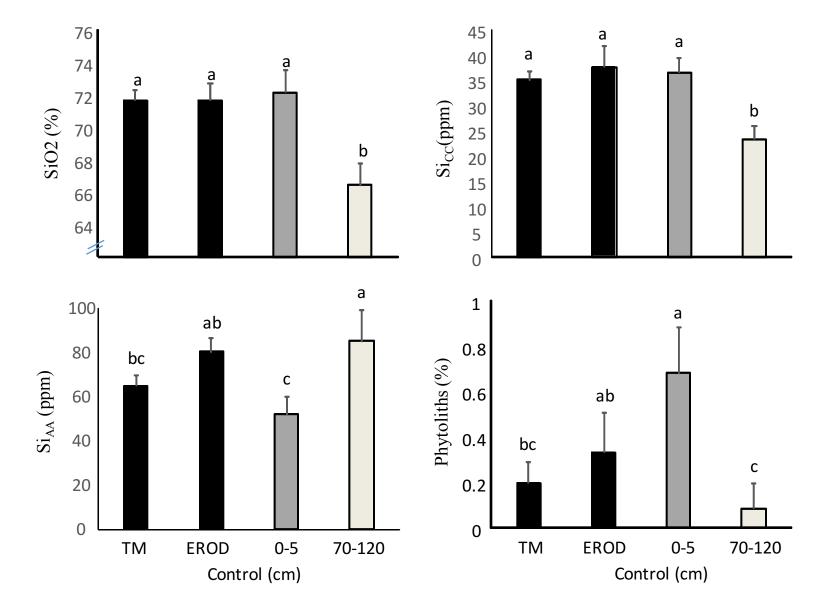


Figure 4

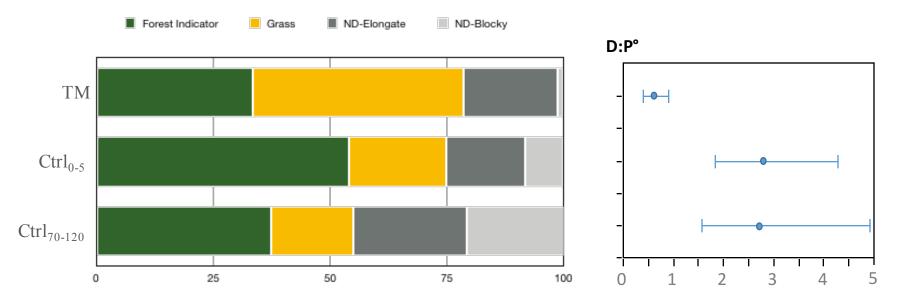


Figure 5

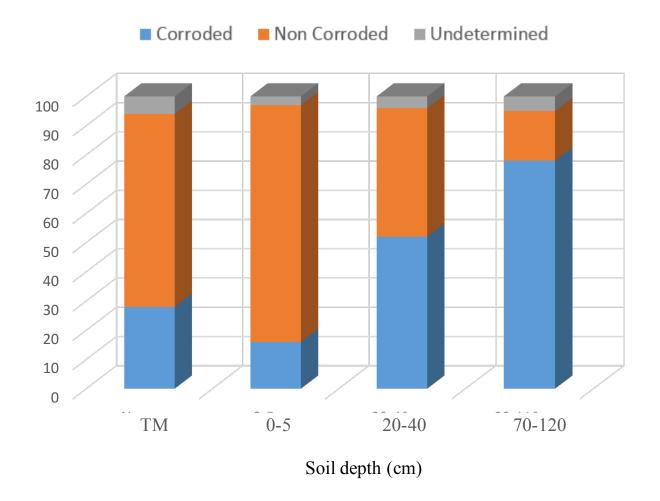


Figure 6

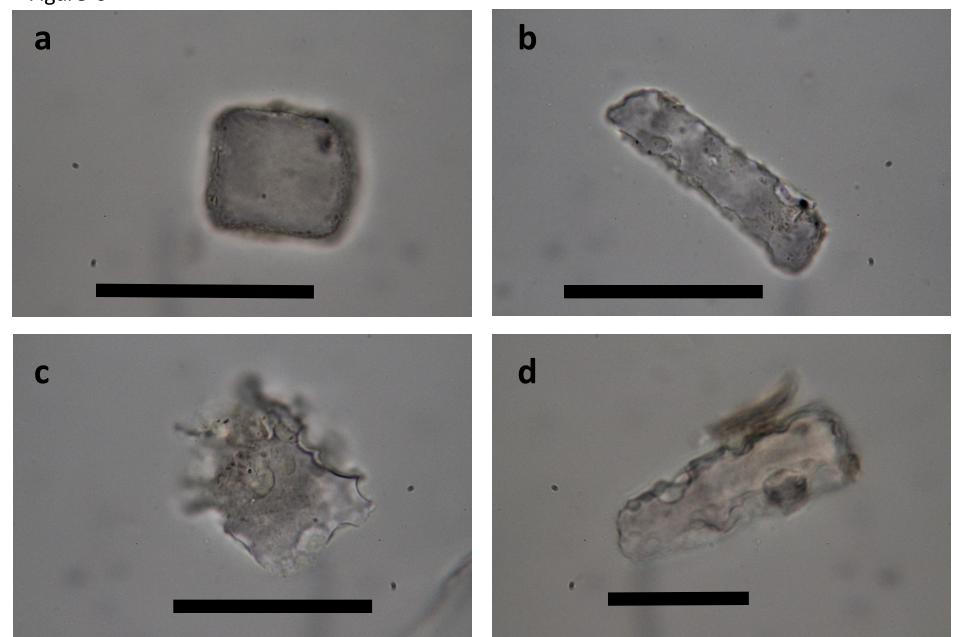
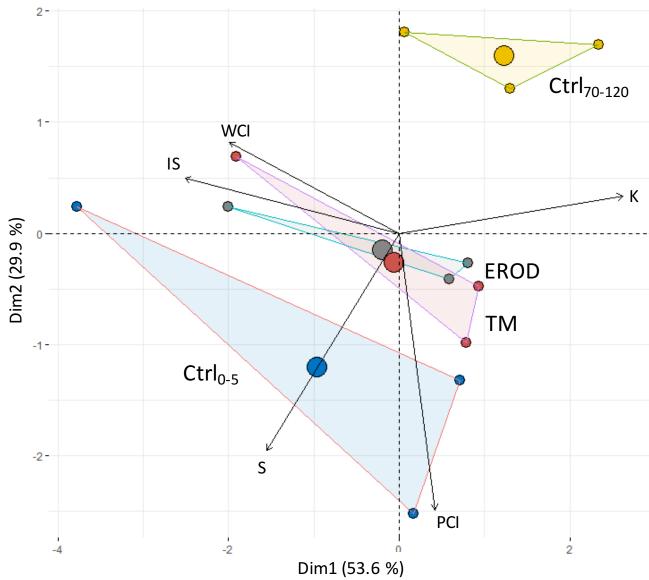
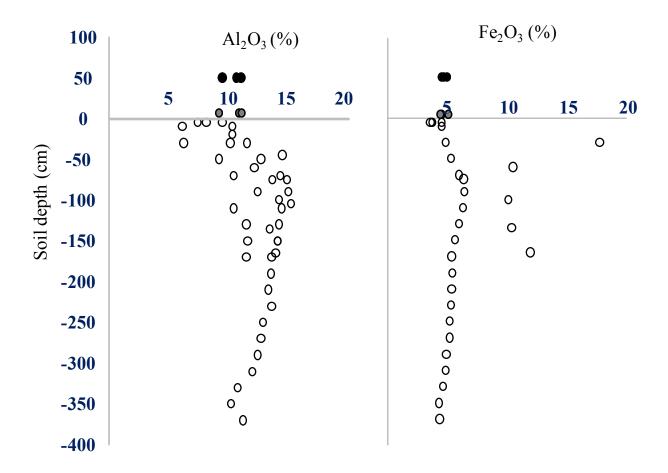


Figure 7



## Supplementary file 1



# Supplementary file 2

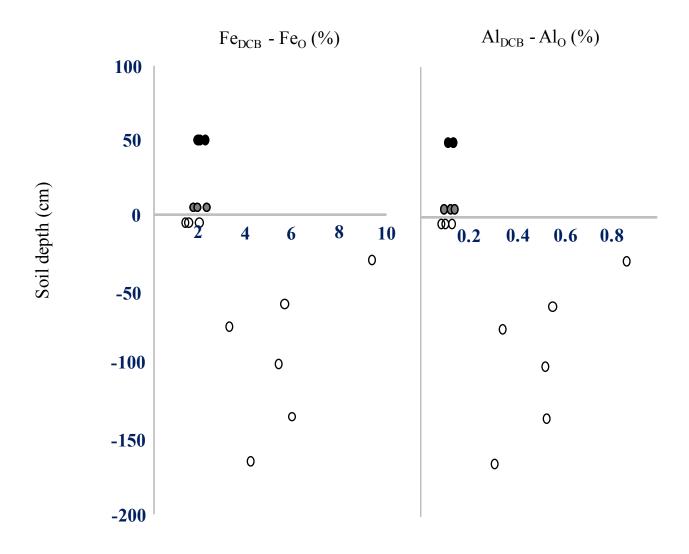


Table 1 Major physical (contents in clay, silt and sand, in %) and chemical properties (contents in C and N in %, CEC,  $Ca^{2+}$ ,  $Mg^{2+}$ ,  $Na^{+}$ ,  $K^{+}$ ,  $Fe^{3+}$ ,  $Mn^{2+}$  and  $Al^{3+}$  in  $cmol^{(+)}kg^{-1}$ ) in the control soils at 0-5 and 70-120 cm depth (Ctrl<sub>0-5</sub> and Ctrl <sub>70-120</sub>, respectively) and in termite mound (TM) or the soil eroded from TM (EROD). Values in parentheses are standard errors, values with the same letter are similar at P = 0.05. Results from the ANOVA (F and P-values) and Wilcoxon–Mann–Whitney U ( $Chi^{2}$  and P-values) tests are also given.

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|                        | C                 | N          | Clay    | Silt   | Sand   | CEC                | $Ca^{2+}$         | $Mg^{2+}$ | $Na^+$              | $K^{+}$                    | $\mathrm{Fe}^{3+}$ | $\mathrm{Mn}^{2^+}$ | $Al^{3+}$  | рН     |
|------------------------|-------------------|------------|---------|--------|--------|--------------------|-------------------|-----------|---------------------|----------------------------|--------------------|---------------------|------------|--------|
|                        |                   |            | (%)     |        |        |                    |                   |           | (cmol <sup>(-</sup> | $^{+)}$ kg <sup>-1</sup> ) |                    |                     |            |        |
| TM                     | 1.75 <sup>b</sup> | $0.13^{b}$ | 31.2    | 18.1   | 50.7   | 13.30 <sup>b</sup> | 9.25 <sup>b</sup> | 2.96      | $0.043^{b}$         | 0.27                       | $0.016^{ab}$       | $0.18^{a}$          | $0.09^{a}$ | 6.03   |
| (n = 3)                | (0.38)            | (0.03)     | (3.87)  | (0.6)  | (1.8)  | (0.61)             | (0.60)            | (0.05)    | (0.002)             | (0.04)                     | (0.001)            | (0.03)              | (0.01)     | (0.07) |
| EROD                   | 1.95 <sup>b</sup> | $0.14^{b}$ | 29.7    | 18.9   | 51.4   | $13.80^{b}$        | $9.68^{b}$        | 3.22      | $0.039^{b}$         | 0.40                       | $0.016^{ab}$       | $0.04^{b}$          | $0.09^{a}$ | 6.45   |
| (n = 3)                | (0.30)            | (0.02)     | (4.85)  | (0.2)  | (2.6)  | (0.25)             | (0.21)            | (0.12)    | (0.009)             | (0.06)                     | (0.002)            | (0.01)              | (0.01)     | (0.22) |
| $Ctrl_{0-5}$           | $3.43^{a}$        | $0.25^{a}$ | 26.6    | 17.4   | 56.0   | $19.50^{a}$        | $14.83^{a}$       | 3.40      | $0.025^{b}$         | 0.47                       | $0.010^{b}$        | $0.02^{b}$          | $0.05^{b}$ | 6.23   |
| (n = 3)                | (0.29)            | (0.03)     | (5.14)  | (0.3)  | (2.9)  | (0.78)             | (0.92)            | (0.21)    | (0.003)             | (0.11)                     | (0.003)            | (0.03)              | (0.01)     | (0.13) |
| Ctrl <sub>70-120</sub> | $0.49^{c}$        | $0.05^{c}$ | 37.23   | 18.06  | 44.72  | 11.78 <sup>b</sup> | $8.00^{b}$        | 3.36      | $0.111^{a}$         | 0.49                       | $0.022^{a}$        | $0.16^{a}$          | $0.13^{a}$ | 6.75   |
| (n = 10)               | (0.27)            | (0.03)     | (14.87) | (2.02) | (3.17) | (1.11)             | (0.89)            | (0.34)    | (0.01)              | (0.09)                     | (0.002)            | (0.04)              | (0.01)     | (0.90) |
| $F_{3,15}$             | 15.75             | 52.27      | 0.80    | 0.05   | 1.74   | 5.84               | 6.9               |           | 11.36               | 1.85                       | 5.34               | 5.62                | 5.94       | 1.88   |
| P                      | < 0.001           | < 0.001    | 0.511   | 0.986  | 0.202  | 0.007              | 0.004             |           | < 0.001             | 0.182                      | 0.010              | 0.008               | 0.007      | 0.222  |
|                        | ***               | ***        |         |        |        | **                 | **                |           | ***                 |                            | *                  | **                  | **         |        |
| $Chi^2$                |                   |            |         |        |        |                    |                   | 1.08      |                     |                            |                    |                     |            |        |
| P                      |                   |            |         |        |        |                    |                   | 0.781     |                     |                            |                    |                     |            |        |