

# A population pharmacokinetic model for escitalopram and its major metabolite in depressive patients during the perinatal period: Prediction of infant drug exposure through breast milk

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 depressive patients during the perinatal period: prediction of infant drug exposure
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#### 39 Abstract

Background and objectives: Escitalopram figures among the most frequently prescribed drugs for the treatment of depression in pregnant and breastfeeding women. Available information on exposure to escitalopram during the perinatal period and its excretion into breast milk is however based on heterogeneous and incomplete data. A population pharmacokinetic model was therefore developed using prospectively collected samples and clinical information to better characterize maternal and infant exposure to escitalopram and its metabolite, and predict the exposure variability under various conditions.

47 Methods: The study population was composed of women taking escitalopram or racemic 48 citalopram included in a multicenter prospective cohort study enrolling pregnant women taking 49 selective serotonin reuptake inhibitors (SSRI) and willing to breastfeed. A joined structural model was first built for escitalopram (SCIT) and S-desmethylcitalopram (SDCIT) in plasma 50 using NONMEM<sup>®</sup>. Then, drug breast milk concentrations were added to the basic model and 51 described by estimating milk-to-plasma ratios (MPR). The effect of different influential covariates 52 53 such as age, bodyweight, CYP2C19, 2D6 and 3A4/5 genotypes or milk fat content on drug and 54 metabolite pharmacokinetics was tested. Finally, drug exposure of a suckling child through breast milk was predicted under various conditions by simulation. 55

56 **Results:** The study enrolled 33 patients treated with escitalopram or racemic citalopram who 57 provided 80 blood and 104 milk samples. Mean SCIT clearance was 32.3 L/h with a betweensubject variability of 31% (CV%) and apparent volume of distribution was 1590 L. Poor 58 metabolizers of CYP2C19 showed a significant 51% decrease in SCIT clearance compared to 59 other phenotypes. Both parent drug and metabolite were characterized by a MPR of 1.9, while 60 61 an increased milk fat content was significantly associated with an increased drug transfer into 62 breast milk (+28% for SCIT and +18% for SDCIT when fat amount passes from 3.1 to 6.2 g/100 63 ml). Simulations suggested that an exclusively breastfed infant would ingest daily through breast milk 3.3% of the weight-adjusted maternal SCIT dose on average. 64

**Conclusion:** Escitalopram and its metabolite showed moderate between-subject variability in blood concentrations, partially explained by genetic polymorphisms in CYP2C19. Milk concentrations were similarly variable and mainly influenced by the milk fat content. The limited exposure to escitalopram through breast milk, as expected based on previous incomplete data, was confirmed by this population pharmacokinetic model approach. These findings provide reassurance with a good level of certainty for clinicians and patients successfully treated with escitalopram or racemic citalopram in the perinatal period.

- 73 **Keywords:** escitalopram, breastfeeding, exposure, population pharmacokinetics
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- 75

#### 76 **1. Introduction**

77 Worldwide about 10% of pregnant women and 13% of women who have just given birth 78 experience a mental disorder, primarily depression (http://www.who.int/mental health/maternal-79 child/en/). If left untreated, depression has been associated with adverse outcomes for the 80 mother, the infant and their family environment, such as reduced maternal sensitivity and 81 involvement in caregiving, and child emotional or behavioral difficulties [3-5]. Antidepressants, 82 mainly selective serotonin reuptake inhibitors (SSRI), are therefore commonly prescribed for 83 depression in pregnant and breastfeeding women, when a pharmacological treatment is 84 indicated.

Escitalopram (SCIT) belongs to the SSRI class and figures among the most frequently reported 85 maternal treatments for chronic conditions in pregnancy [6]. SCIT represents the 86 87 pharmacologically active S-(+) enantiomer of the racemic citalopram and is also marketed as the pure enantiomer escitalopram. After oral administration of doses of 10-20 mg of 88 escitalopram per day, SCIT is rapidly absorbed (T<sub>max</sub> of 3-4 h) and characterized by a volume of 89 90 distribution (V), oral clearance (CL) and elimination half-life (t<sub>1/2</sub>) of 18.3 L/kg, 0.48 L/h/kg and 91 about 30 h, respectively [7, 8]. SCIT is primarily metabolized in the liver by cytochrome P450 92 (CYP) enzymes into S-(+)-desmethylcitalopram (SDCIT), which shows a prolonged elimination half-life of 50 h [7, 8]. CYP2C19, CYP3A4 and to a lesser extent CYP2D6 have been reported 93 94 as the main metabolic pathways [9], and genetic polymorphisms in the enzyme-coding genes may be responsible for interpatient variations in drug plasma concentrations [10]. 95

96 Escitalopram doses ingested by a breastfed infant through milk have been reported to be low 97 (3-6% of the weight-adjusted maternal dose) and result very often in negligible drug 98 concentrations in infant plasma [11]. However, the extent of drug transfer into breast milk varies 99 widely between individuals in terms of milk-to-plasma concentration ratio (MPR, 1.7-2.7 for SCIT 100 and 1.8-3.1 for SDCIT) [12, 13]. Sampling procedure (e.g. single vs. several time points), milk

101 composition (foremilk vs. hindmilk, colostrum vs. mature milk), factors influencing maternal drug 102 disposition and sample size are possible sources of the observed variability. Based on these 103 heterogeneous data, health care professionals may in some cases advice mothers taking 104 escitalopram to discontinue breastfeeding due to the potential risk of adverse effects in the 105 suckling child, despite the numerous proven health benefits of breastfeeding [14].

The aim of this study was to describe for the first time the pharmacokinetics of escitalopram and its major metabolite in depressive women during the perinatal period using a population pharmacokinetic (popPK) modeling approach. The influence of genetic and environmental factors on drug concentrations and transfer into breast milk was also assessed. Finally, drug exposure of a suckling child was predicted under various conditions by simulation.

111

#### 112 2. Methods

#### 113 2.1. Study population

All relevant samples and data were collected from women included in the "SSRI-Breast Milk" 114 study (NCT01796132, more information on clinicaltrials.gov). This multicenter prospective 115 116 cohort study enrolls pregnant women willing to breastfeed and treated with a SSRI 117 antidepressant or a serotonin noradrenalin reuptake inhibitor (SNRI) in one of the maternity wards collaborating to the study (i.e. the maternities of the Lausanne University Hospital, 118 119 Geneva University Hospital, Ensemble hospitalier de la Côte in Morges, University Hospital of 120 Nancy and the Hospices Civils in Lyon). Study protocol was approved in 2012-2013 by local 121 Ethics Committees and health authorities in Switzerland and France.

122 Maternal blood samples and breast milk samples were obtained from pregnant and 123 breastfeeding women treated either with SCIT (n=21) or racemic citalopram (n=12). Maternal 124 blood samples collected around the moment of delivery were available for 28 patients (1 sample

per patient; 14 during labor, 14 right after delivery). Paired blood and breast milk samples during 125 126 the first week postpartum and at 1 month after delivery were collected in 29 and 23 patients, respectively (1 blood and 2 milk samples (fore- and hindmilk) per patient at each occasion). The 127 128 analysis included therefore drug concentrations from a total of 80 blood samples and 104 breast 129 milk samples. Median SCIT and citalopram doses were 10 mg/day (range 5-20 mg/day) and 15 mg/day (range 5-60 mg/day), respectively. Sampling occurred under steady-state conditions at 130 131 a median time-lapse of 11.17 h (range 0.02-34.00 h) and 10.25 h (range 0.25-52.25 h) after the last SCIT or citalopram dose intake, respectively. 132

#### 133 2.2. Analytical methods

134 Venous blood samples (5 ml) were drawn on potassium-EDTA tubes and stored after centrifugation as plasma samples in polypropylene tubes at -20°C. Breast milk was collected in 135 136 lactating mothers using an automatic or manual pump, and stored at -20°C or -80°C in 137 polystyrene tubes until analysis. Subsequent quantification of SCIT and SDCIT in plasma and breast milk was performed using a validated stereoselective HPLC-MS/MS method (Acquity® 138 139 UPLC I-Class, Waters, Milford, USA) described in details previously [15]. Composition of breast milk was determined using the Human Milk Analyzer® (Miris, Uppsala, Sweden) which provided 140 information on fat, protein, carbohydrate and calorie content. Plasma concentrations of alpha-1-141 acid-glycoprotein (AAG) were measured by immunoturbidimetry using the Cobas Integra® 400 142 plus analyzer (Roche Diagnostics, Rotkreuz, Switzerland). 143

# 144 2.3. Genotyping

Genomic DNA was first extracted from potassium-EDTA blood samples collected at delivery or later. Most single nucleotide polymorphisms (SNP) were analyzed by TaqMan<sup>®</sup>-Assay based real-time polymerase chain reaction (Applied Biosystems, Rotkreuz, Switzerland): *CYP2C19\*2*, *CYP2C19\*3*, *CYP219\*17*, *CYP2D6\*3*, *CYP2D6\*4*, *CYP2D6\*6*, *CYP3A4\*22*, *CYP3A5\*3* and

POR\*28. Duplication/multiplication of CYP2D6\*xN and CYP2D6\*5 gene deletion were detected 149 by TaqMan<sup>®</sup> copy number assay. Regarding CYP2C19, patients were classified into 4 predicted 150 phenotypes: poor metabolizer (PM) if they were homozygous for the \*2/\*2 allele, intermediate 151 152 metabolizer (IM) if they carried any of \*1/\*2 or \*17/\*2 allele, extensive metabolizer (EM) if they 153 were homozygous for the wild-type allele  $\frac{1}{1}$  and ultrarapid metabolizer (UM) if they carried either \*1/\*17 or \*17/\*17 genotype. Patients were also divided in 4 different predicted phenotypes 154 155 according to CYP2D6 genotype: poor metabolizer (PM; carriers of 2 non-functional alleles, e.g. \*4/\*4, \*4/\*5, \*4/\*6), intermediate metabolizer (IM; carriers of 1 functional and 1 non-functional 156 allele, e.g. \*1/\*3, \*1/\*4, \*1/\*5), extensive metabolizer (EM; carriers of two functional alleles, 157 158 (1/1) and ultrarapid metabolizer (UM; carriers of 1 functional allele and an increased number of gene copies, \*1/\*xN, or carriers of an increased number of gene copies, \*xN/\*xN. For control 159 160 purposes, each analyzed batch of samples contained also DNAs with known genotypes.

#### 161 2.4. Population pharmacokinetic analysis

All popPK analyses were conducted using the non-linear mixed effects modeling tool 162 NONMEM® (version 7.3.0, ICON Development Solutions, Ellicott City, USA), which enables the 163 estimation of average population parameters as well as between-subject and residual variability. 164 Due to the use of a stereoselective analytical method (see section 2.2), concentration data from 165 166 both racemic citalopram and pure escitalopram could be pooled assuming a negligible effect of the R-(-) enantiomer on the SCIT pharmacokinetic profile. This assumption was confirmed by 167 168 testing the influence of maternal treatment on SCIT profile (citalopram vs. pure escitalopram as 169 a dichotomous variable), which demonstrated to be non-significant.

170 2.4.1 Structural and statistical models

A stepwise approach was chosen in order to obtain a combined model with the best fit for both parent compound and its metabolite. First, one and two compartment models with linear drug

elimination were evaluated for SCIT plasma concentrations alone. Then, one additional
compartment was used to describe SDCIT plasma concentrations assuming linear metabolism.
Finally, parent drug and metabolite transfers into breast milk were characterized by MPR as a
scaling factor between concentration profiles in plasma and milk, as previously described [16].
The final structural model is illustrated in Figure 1 and described by the following differential
equations:

$$\frac{dSCIT_{dose}}{dt} = -k_{12} \times SCIT_{dose} \tag{1}$$

$$\frac{d(SCIT_{plasma} \times V_{SCIT})}{dt} = k_{12} \times SCIT_{dose} - k_{20} \times SCIT_{plasma} - k_{23} \times SCIT_{plasma}$$
(2)

$$\frac{d(SDCIT_{plasma} \times V_{SDCIT})}{dt} = k_{23} \times SCIT_{plasma} - k_{30} \times SDCIT_{plasma}$$
(3)

$$SCIT_{milk} = MPRD \times SCIT_{plasma}$$
 (4)

$$SCDIT_{milk} = MPRM \times SDCIT_{plasma}$$
 (5)

179 where SCIT<sub>dose</sub> is the SCIT amount in the dose compartment, k<sub>12</sub> the absorption rate constant, 180 k<sub>20</sub> the elimination rate constant from the parent drug compartment, k<sub>23</sub> the metabolic rate 181 constant from SCIT to SDCIT, k<sub>30</sub> the elimination rate constant from the metabolite 182 compartment, V<sub>SCIT</sub> the volume of distribution of SCIT, SCIT<sub>plasma</sub> and SDCIT<sub>plasma</sub> the parent drug and metabolite concentrations in plasma, respectively, SCIT<sub>milk</sub> and SDCIT<sub>milk</sub> the parent 183 drug and metabolite concentration in breast milk, MPRD and MPRM the milk-to-plasma ratio for 184 185 the parent drug and the metabolite, respectively. Due to identifiability issues, the volume of distribution of SDCIT (V<sub>SDCIT</sub>) could not be estimated. We assumed that  $k_{23} = k_{30}$  and estimated 186 V<sub>SDCIT</sub> through the ratio of metabolite to parent drug volume of distribution (with VMR being the 187 188 volume metabolite ratio,  $V_{SDCIT} = V_{SCIT} \times VMR$ ).

Between-subject variability (BSV) of each pharmacokinetic parameter was described using an
 exponential error model. A combined proportional and additive error model was first tested to

describe residual variations in plasma for SCIT and then simplified to a proportional error model,which was retained for both drug and metabolite.

193 2.4.2 Covariate models

194 The effect of genetic and environmental covariates on the final structural model was assessed by a forward insertion/backward deletion approach. Maternal age (AGE), bodyweight (BW), 195 alpha-1-acid-glycoprotein concentration (AAG) and important CYP polymorphisms (CYP2C19, 196 197 CYP2D6, CYP3A4, CYP3A5) were tested as covariates. Genotypes for both CYP2C19 and 2D6 198 isoenzymes were classified into 4 distinct groups according to their predicted phenotype, while the actual genotype was used for CYP3A4 (\*1/\*1, \*1/\*22, \*22/\*22), 3A5 (\*1/\*1, \*1/\*3, \*3/\*3) and 199 200 POR (\*1/\*1, 1\*/\*28, \*28/\*28). Regarding drug transfer into breast milk, feeding occasion (FEED; fore- versus hindmilk), fat (FAT), protein (PROT), carbohydrate (CARBO) and calorie content 201 202 (ENERGY) were evaluated as potentially important covariates. The moment of blood or milk sampling (MOM; 4 categories: during labor, after delivery, 1<sup>st</sup> week postpartum or 1 month after 203 204 delivery) was also tested on various parameter estimates as drug concentrations during the 205 perinatal period may vary significantly due to physiological changes.

206 Continuous covariates (normalized and centered on their median population value, missing data 207 were replaced by the median value) and dichotomous covariates (coded as 0/1) were tested 208 using a linear function. Discrete variables including more than 2 groups were described as 209 categorical covariates assigning a fixed effect to each category and reduced models combining 210 different groups were tested as well for statistical significance.

211

212 2.4.3 Parameter estimation and model selection

Data were fitted using the first-order conditional estimation method with interaction (FOCE-I) in 213 NONMEM® (reference for NONMEM: Beal SL SL, Boeckmann A, Bauer RJ. NONMEM user's 214 quide (1989–2009) Ellicott City: Icon Development Solutions. 2009). The log-likelihood ratio test 215 216 was used to discriminate between two nested models (difference in the objective function value ( $\triangle OFV$ ) approximates a  $\chi^2$  distribution). A  $\triangle OFV < -3.84$  ( $\chi^2$ , p<0.05, df =1) and >6.63 217  $(\chi^2, p<0.01, df = 1)$  were considered statistically significant for one additional parameter during 218 219 forward insertion and backward deletion steps, respectively. Goodness-of-fit plots, pharmacokinetic parameter precision and decrease in between-subject variability were also 220 considered for model quality assessment. 221

### 222 2.4.4 Model validation

223 The precision of parameter estimates and stability of the final model were assessed using the 224 non-parametric bootstrap method. Briefly, two thousand datasets were created through random resampling of original data (1000 stratified on CYP2C19 and 1000 on CYP2D6 genotypes, 225 respectively). Median parameter values and percentile intervals (90% PI) were generated for 226 227 each pharmacokinetic parameter based on all replicated datasets and compared to estimations from the original model. Prediction-corrected visual predictive checks (pcVPC) were also carried 228 out by simulating 1000 individuals based on the final covariate model estimates using the PsN 229 toolkit [17]. 230

231

### 232 2.4.5 Simulation of drug concentrations and prediction of infant drug exposure

Plasma and breast milk concentrations of SCIT and SDCIT at steady-state were simulated according to different dose regimens in 5'000 mothers based on the final structural model (without any covariates) as well as according to relevant covariates in 10'000 mothers based on intermediate covariate models (one single covariate added alternatively to the final structural

model). For this purpose, virtual datasets of breastfeeding women were generated with R<sup>®</sup> (R 237 238 Development Core Team, Foundation for Statistical Computing, Vienna, Austria) by assigning random breastfeeding frequencies and intervals to each woman after the last dose intake. In 239 240 order to best reflect usual breastfeeding frequencies, the mean frequency was set to 11±3 241 feedings per day with a range between 6 and 18 times a day [18]. Finally, the daily infant dosage that is equivalent to the drug dose ingested by an exclusively breastfed infant through 242 243 breast milk was calculated for each mother-infant pair and expressed in mg/kg/day, using the 244 following equation:

Daily infant dosage = 
$$\sum_{i=1}^{n} C_{milk}^{i} * V_{milk}$$

where  $C_{milk}^{i}$  is the simulated drug concentration in milk at the i<sup>th</sup> feeding time, n is the daily feeding frequency and  $V_{milk}$  is the ingested milk volume by a suckling child during a feeding occasion. For simplicity, typical  $V_{milk}$  was derived from literature by dividing the usual weightadjusted daily milk intake of 150 ml/kg/day by the feeding frequency [19].

249

In order to better evaluate the clinical relevance of the daily infant dosage, two additional exposure estimates were determined, namely the relative infant dose (RID) and the adult dose equivalent (ADE). RID represents the ratio between the daily infant dosage and the weightadjusted maternal dose (based on a standard adult weight of 70 kg) and is expressed as a percentage [19]. For example, a RID of 10% indicates that a suckling child would daily ingest via breast milk 10% of the weight-adjusted maternal SCIT dose.

The ADE quantifies the virtual number of tablets that would be ingested by the infant over 6 months of exclusive breastfeeding. This number was obtained by dividing the cumulated daily infant dosage over 6 months by a standard adult daily dose (i.e. 10 mg for escitalopram), assuming a mean child weight of 6 kg.

#### 261 **3. Results**

262 Demographic and clinical characteristics of patients included in the popPk analysis are 263 presented in Table 1.

264 3.1. Structural models

Regarding SCIT, data were best described by a one compartment model with first-order absorption and elimination. The two compartment model did not show any significant improvement of the fit ( $\Delta$ OFV = -1.0, p>0.05). The addition of a BSV on CL, but not on V or k<sub>a</sub> ( $\Delta$ OFV = 0.0, p>0.05), yielded a significantly better fit ( $\Delta$ OFV = -40.9, p<0.001) and was therefore retained (40.1%, CV%).

In the next step, metabolite plasma concentrations (SDCIT) were added to the dataset. Due to identifiability issues, the metabolite's apparent volume of distribution was estimated using a scaling factor VMR (volume metabolite ratio), and resulted in acceptable parameter precision. The assignment of a BSV to VMR slightly improved the fit ( $\Delta$ OFV = -5.2, p=0.02; 20.0%, CV%), while a variability on k<sub>23</sub> did not ( $\Delta$ OFV = -2.9, p>0.05).

Finally, SCIT and SDCIT milk concentrations were integrated into the model using the milk-toplasma ratios (MPRD and MPRM, respectively). No changes in the model fit were observed assigning a BSV to MPRD or MPRM ( $\Delta$ OFV > -0.2, p>0.05. Some individual covariates such as milk fat content were nevertheless tested on these parameters due to a priori knowledge. Pharmacokinetic parameters of the structural model are presented in Table 2.

280

### 281 3.2. Covariate models

282 Univariate analyses showed that MOM, BW and CYP2C19 were significantly associated with 283  $CL_{SCIT}$  ( $\Delta OFV < -14.1$ , p<0.05). No differences could be observed between concentrations

collected during labor, right after delivery and during the 1<sup>st</sup> week postpartum ( $\Delta OFV = 0$ , 284 285 p>0.05), nor between CYP2C19 IM, EM and UM ( $\Delta OFV = 0$ , p>0.05). These categorical covariates could therefore be reduced to dichotomous variables. Maternal age, CYP2D6, 3A4, 286 287 3A5, POR or AAG did not show any significant effect on  $CL_{SCIT}$  ( $\Delta OFV > -3.8$ , p>0.05). Only 288 MOM importantly influenced V<sub>SCIT</sub> ( $\Delta$ OFV = -46.4, p=0.05) but not BW, AGE or AAG ( $\Delta$ OFV >-1.6, p>0.05). VMR was affected by MOM, CYP2C19 and CYP2D6 ( $\Delta$ OFV < -6.9, p<0.05) but 289 290 not by BW, AGE, CYP3A4, 3A5, POR or AAG ( $\triangle OFV > -3.4$ , p>0.05). No differences could be observed between CYP2D6 IM and EM ( $\Delta OFV = 0$ , p>0.05). Regarding univariate analyses 291 conducted on MPRD and MPRM, the covariates MOM, FAT, ENERGY and FEED all 292 significantly influenced the model ( $\triangle OFV < -4.0$ , p=0.05). 293

In multivariate analyses, MOM and CYP2C19 showed an independent impact on CL<sub>SCIT</sub> but not 294 BW ( $\triangle OFV = -3.8$ , p=0.05). SCIT clearance was reduced by 51% in CYP2C19 PM compared to 295 the other genetic polymorphisms. A 10% decrease in CL<sub>SCIT</sub> at 1 month after delivery was 296 297 observed compared to the perinatal period. The influence of both CYP2D6 and CYP2C19 on 298 VMR remained significant in multivariate analysis ( $\Delta OFV < -12.8$ , p<0.05). These results 299 suggest lower plasma concentrations of the active metabolite SDCIT in CYP2C19 PM (-72%) 300 and higher concentrations in UM (+22%) in comparison to IM/EM. In addition, patients carrying 301 CYP2D6 PM genotype had also reduced SDCIT concentrations (-104%) compared to IM/EM.

The combined addition of MOM and FAT to MPRD improved the description of the data, but ENERGY and FEED did not remain significant in multivariate analyses ( $\Delta OFV > -3.4$ , p>0.05). Only FAT remained statistically significant on MPRM, since it was correlated with ENERGY and FEED ( $\Delta OFV > -3.8$ , p>0.05). The transfer of SCIT and SDCIT into breast milk increased by 28% and 18%, respectively, with a doubling of fat content from 3.1 to 6.2 g/100 ml. The final parameter estimates, influencing covariates and variabilities together with their bootstrap estimations are presented in Table 3 and diagnostic plots are shown in Figure 2 and 3. 310 3.3. Model validation

The pcVPCs shown in Figure 4 supported the good predictive performance of the model. All parameter estimates from the final covariate model lied within the 90% percentile interval (90% PI) calculated from the bootstrap method and were close to the median values (<11%), indicating a reasonably robust model.

315

# 316 3.4. Prediction of infant drug exposure by simulation

317 Based on the final structural model without any covariates, simulation of 5'000 mothers treated with 10 mg escitalopram daily revealed that the daily infant dosage would lie between 0.001 and 318 319 0.016 mg/kg/day (median: 0.005 mg/kg/day). Expressed as the ratio to the weight-adjusted maternal dose, RID values for escitalopram ranged from 0.8% to 11.3% (median: 3.3%; Figure 320 321 5), which is in line with previously reported values [11, 20]. In terms of ADE, an exclusively breastfed infant would ingest over 6 months a cumulated dose of 1.2-17.4 mg (median: 5.0 mg) 322 equivalent to 0.1-1.7 tablets of escitalopram (median: 0.5; Figure 5). RID and ADE would be 323 324 slightly higher in mothers who are PM for CYP2C19, while the influence of other covariates on 325 these exposure estimates remained negligible. Simulations performed on the active metabolite SDCIT showed only a minor impact on overall infant drug exposure (data not shown). 326

327

# 328 4. Discussion

This study described for the first time the population pharmacokinetics of SCIT and its major metabolite SDCIT in depressive patients during the perinatal period, while taking into account genetic, demographic and environmental factors. Estimated pharmacokinetic parameters and

mean MPR of both parent drug and metabolite are in line with available data [7, 8, 21, 22] [12, 13]. These results suggest that this antidepressant drug has a higher affinity for breast milk than for plasma (MPR>1.0), but infant exposure through breast milk remains limited as illustrated by a median RID of 3.3% and an ADE of 0.5 tablets.

336 Model results showed a slight decrease of 10% in clearance of SCIT one month after delivery 337 compared to the perinatal period. A similar finding has been reported previously for racemic 338 citalopram, where concentrations measured at delivery and corrected to a daily dose of 20 mg were 20-30% lower than at 2-4 months postpartum. These small studies observed even higher 339 differences when comparing to the 2<sup>nd</sup> and 3<sup>rd</sup> trimester and attributed them to physiological 340 changes in pregnancy affecting renal and hepatic functions (e.g. increased glomerular filtration 341 rate, induced CYP2D6 and 3A4 activity) [23, 24]. This effect could not be investigated in our 342 343 population due to the lack of drug measurements earlier in pregnancy, but may potentially suggest the need for dose adjustment in pregnant women in order to prevent relapse of 344 345 depressive symptoms. Yet, the clinical relevance of such a measure still requires confirmation 346 from larger studies.

347 The significant influence of CYP2C19 predicted phenotype on SCIT clearance and SDCIT 348 volume metabolite ratio is in agreement with numerous other publications reporting similar 349 effects in the psychiatric population. According to a recent meta-analysis, CYP2C19 PM show a mean increase in (es)citalopram concentrations of 95% compared to EM [25]. In these patients, 350 351 enhanced drug exposure could potentially lead to a lower drug tolerance or even to toxicity (e.g. QT prolongation), reason why authorities and consensus guidelines recommend dose 352 353 adjustments in these situations [26]. Similar considerations are of particular interest for 354 depressive women during the perinatal period as higher maternal drug concentrations will lead to higher exposure of the suckling child through breastmilk. In addition to CYP2C19, 355 356 polymorphisms in genes coding for CYP2D6 had also a significant effect on metabolite plasma 357 concentrations. However, this finding was based on only two CYP2D6 PM patients (and no 16/29

UM), thus no specific conclusions on the real effect size can be drawn. Polymorphisms in CYP3A4, 3A5 or POR did not show any effect on SCIT elimination in exploratory analyses, but study population included only one CYP3A5 \*1/\*1 carrier and no CYP3A4 \*22/\*22 carrier.

361 SCIT and its major metabolite are excreted into breast milk with a MPR of 1.9. Breast milk fat 362 content, which varies during a feed (i.e. hind milk vs fore milk) and breastfeeding periods (i.e. 363 colostrum vs mature milk), mainly influenced this passage. Similar findings showing a higher affinity of lipophilic drugs for milk richer in lipids have been published previously (Berle et al. 364 2004, PMID: 15367050). In fact, simulations performed in 10'000 mother-infant pairs showed 365 366 that a doubling of fat content from 2.1 g to 4.3 g/100 ml increased median RID from 2.8% to 367 3.5%. Despite this increase, the RID can still be considered as reassuringly low. This clearly outweighs the possible effect of a small additional drug transfer. 368

369 Regarding prediction of infant drug exposure, maternal CYP2C19 phenotype was identified as 370 the most influential covariate in our population. Breastfed infants of poor metabolizing mothers 371 would ingest via milk 5.7% of the weight-adjusted maternal dose in average compared to 3.0% in other phenotypes. Based on average multiethnic frequencies, 2-15% of individuals may be 372 CYP2C19 PM and therefore concerned by elevated drug concentrations [26=Hicks]. However, 373 374 95% of their suckling children would not exceed a RID value of 10% and would receive in total 375 after 6 months of exclusive breastfeeding an ADE of 0.3-2.3 tablets, indicating a low absolute exposure to the drug. Clinical implications of even very low drug exposure for the long term 376 377 development of a breastfed infant remain however largely uncertain. Simulations performed on the active metabolite SDCIT showed only a minor impact on overall infant drug exposure, 378 379 probably due to its low plasma and milk concentrations. The lower potency of SDCIT as an 380 antidepressant compared to SCIT would further limit its potential impact on the suckling child.

381 One limitation of the present study was the limited sample size of 33 patients for the Pk 382 analysis. Large clinical trials are particularly difficult to conduct in pregnant and nursing women

383 as they are considered a vulnerable population. Observed frequencies of CYP2C19 phenotypes 384 were however representative of the general population [26=Hicks] and thereby reinforcing 385 representativeness of the study sample and thus the validity of the model-based simulations. 386 The remaining unexplained between-subject variability observed suggests that information on 387 potentially influencing covariates were not available for analysis as not captured by the study 388 protocol (e.g. compliance, unknown genetic factors). A limited compliance in our study sample 389 would be linked to an underestimated RID. However, the level of compliance is not expected to 390 be below average in our sample of patient with a high level of education (data not shown).

#### 391 **5. Conclusions**

Escitalopram and its metabolite showed moderate between-subject variability in blood concentrations, partially explained by genetic polymorphisms in CYP2C19. Milk concentrations were similarly variable and mainly influenced by the milk fat content. The limited exposure to escitalopram through breast milk, as expected based on previous incomplete data, was confirmed by this population pharmacokinetic model approach. These findings provide reassurance with a good level of certainty for clinicians and patients successfully treated with escitalopram or racemic citalopram in the perinatal period.

399

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- 472

474	<b>Table 1</b> : Demographic and clinical characteristics of the study population
4/4	Table 1. Demographic and clinical characteristics of the study population

	Study population (n=33)		
Age (years), median (range)	34.0	(21.0-43.0)	
Bodyweight (kg), median (range)			
Term pregnancy (n=27)	76.0	(60.8-120.0)	
1 <sup>st</sup> week postpartum (n=23)	68.0	(53.0-114.0)	
1 <sup>st</sup> month postpartum (n=19)	68.0	(50.0-87.6)	
Alpha-1-acid-glycoprotein (g/L), median (range)			
Term pregnancy (n=28)	0.42	(0.23-1.16)	
1 <sup>st</sup> week postpartum (n=27)	0.80	(0.25-1.47)	
1 <sup>st</sup> month postpartum (n=23)	0.75	(0.48-1.24)	
CYP2C19 predicted phenotype, n (%)			
Poor metabolizer	4	(12%)	
Intermediate metabolizer	5	(15%)	
Extensive metabolizer	12	(36%)	
Ultrarapid metabolizer	12	(36%)	
CYP2D6 predicted phenotype, n (%)			
Poor metabolizer	2	(6%)	
Intermediate metabolizer	8	(24%)	
Extensive metabolizer	23	(70%)	
Ultrarapid metabolizer	0	(0%)	
CYP3A4 genotype, n (%)			
*1/*1	30	(91%)	
*1/*22	3	(9%)	
*22/*22	0	(0%)	
CYP3A5 genotype, n (%)			
*1/*1	1	(3%)	
*1/*3	6	(18%)	
*3/*3	26	(79%)	
POR genotype, n (%)			
*1/*1	18	(55%)	
*1/*28	10	(30%)	

*28/*28	5	(15%)
Breast milk*, median (range)		
Fat content (g/100 ml; n=29)	3.1	(0.2-9.3)
Protein content (g/100 ml; n=29)	1.4	(0.0-4.3)
Carbohydrate content (g/100 ml; n=29)	6.2	(1.0-7.8)
Calorie content (kcal/100 ml; n=29)	57.0	(18.7-112.0)

475 \* mixed fore- and hindmilk

- 477 **Table 2**: Parameter estimates of intermediate and final structural pharmacokinetic models for
- 478 escitalopram (SCIT) alone or combined with S-desmethylcitalopram (SDCIT) in plasma and/or
- 479 breast milk

	Model for SCIT alone		Model for SCIT+SDCIT		Model for SCIT+SDCIT	
	pla	sma	plasma		plasma + breast milk	
	estimate	RSE (%)	estimate	RSE (%)	estimate	RSE (%)
Structural model						
CL <sub>SCIT</sub> (L/h)	29.6	8	27.7	9	28.7	8
V <sub>SCIT</sub> (L)	900	19	1440	9	1310	24
$k_a / k_{12} (h^{-1})$	0.57	34	0.99	37	0.73	60
k <sub>30</sub> (h⁻¹)			0.74	42	0.54	162
VMR			2.6	8	2.5	8
MPRD					2.1	4
MPRM					1.9	4
Statistical model						
ω <sub>CL</sub> (%)	40.1	16	26.2	24	35.4	18
ω <sub>vmr</sub> (%)			20.0	33	33.9	24
$\sigma$ SCIT <sub>plasma</sub> (%)	33.8	9	41.6	11	35.3	10
$\sigma$ SDCIT <sub>plasma</sub> (%)			20.3	10	21.0	10
σ SCIT <sub>milk</sub> (%)					31.5	12
σ SDCIT <sub>milk</sub> (%)					22.8	9

480 RSE, relative standard error defined as SE/estimate;  $CL_{SCIT}$ , apparent clearance of SCIT;  $V_{SCIT}$ , apparent volume of 481 distribution of SCIT;  $k_a / k_{12}$ , apparent absorption rate constant from dose compartment;  $k_{30}$ , apparent metabolic and 482 elimination rate constant of SDCIT; VMR, scaling factor of  $V_{SDCIT}$ ; MPRD, milk-to-plasma ratio of SCIT; MPRM, milk-483 to-plasma ratio of SDCIT;  $\omega$ , between-subject variability estimate;  $\sigma$ , residual variability expressed as coefficient of 484 variation

485

# 487 **Table 3**: Final covariate population model and bootstrap results for escitalopram and its major

488 metabolite

	Final covariate model		Bootsti	Bootstrap (n=2000)	
	estimate	RSE (%)	median	90% PI	
CL <sub>SCIT</sub> (L/h)	32.3	8	32.0	28.4 – 36.1	
CYP2C19	-0.51	17	-0.50	-0.64 to -0.34	
MOM	-0.10	54	-0.10	-0.17 to -0.01	
V <sub>SCIT</sub> (L)	1590	14	1474	811 – 1856	
$k_a / k_{12} (h^{-1})$	0.87	36	0.83	0.20 – 1.73	
k <sub>30</sub> (h⁻¹)	0.67	53	0.59	0.05 – 1.28	
VMR for CYP2C19 PM	4.1	22	4.1	3.0 - 6.3	
VMR for CYP2C19 IM/EM	2.4	7	2.4	2.2 – 2.8	
VMR for CYP2C19 UM	1.9	7	1.9	1.7 – 2.1	
CYP2D6	1.04	11	1.04	0.82 – 1.26	
MPRD	1.9	4	1.9	1.8 – 2.1	
MOM	0.16	53	0.16	0.03 – 0.30	
FAT	0.28	20	0.28	0.20 – 0.39	
MPRM	1.9	3	1.9	1.8 – 2.0	
FAT	0.18	20	0.17	0.11 – 0.23	
ω <sub>CL</sub> (%)	31.2	12	30.5	23.8 - 36.8	
ω <sub>vmr</sub> (%)	21.0	19	18.9	9.3 – 24.7	
$\sigma$ SCIT <sub>plasma</sub> (%)	33.5	8	33.0	28.7 – 37.4	
$\sigma$ SDCIT <sub>plasma</sub> (%)	20.8	9	20.0	16.7 – 23.2	
$\sigma \operatorname{SCIT}_{milk}$ (%)	24.0	11	23.1	18.7 – 27.7	
$\sigma$ SDCIT <sub>milk</sub> (%)	19.4	9	18.4	14.9 – 21.6	

489 RSE, relative standard error defined as SE/estimate; 90% PI, percentile interval between 5 and 95%; CL<sub>SCIT</sub>, 490 apparent clearance of SCIT; CYP2C19, relative deviation of CL<sub>SCIT</sub> if PM compared to IM/EM/UM; MOM, relative 491 deviation of CL<sub>SCIT</sub> or MPRD if moment of sampling at 1 month after delivery compared to labor/early postpartum; 492 V<sub>SCIT</sub>, apparent volume of distribution of SCIT; k<sub>a</sub> / k<sub>12</sub>, apparent absorption rate constant from dose compartment; 493 k<sub>30</sub>, apparent metabolic and elimination rate constant of SDCIT; VMR, scaling factor of V<sub>SDCIT</sub>; CYP2D6, relative 494 deviation of VMR if PM compared to IM/EM; MPRD, milk-to-plasma ratio of SCIT; FAT, relative deviation of MPRD or 495 MPRM depending on fat content of breast milk; MPRM, milk-to-plasma ratio of SDCIT; ω, between-subject variability 496 estimate;  $\sigma$ , residual variability expressed as coefficient of variation

Figure 1: Compartmental structure of the final model and its main pharmacokinetic parameters
(V<sub>SCIT</sub>: volume of distribution of escitalopram; VMR: volume metabolite ratio; k<sub>xy</sub>:
absorption/metabolic/elimination rate constant; MPRD: milk-to-plasma ratio of escitalopram;
MPRM: milk-to-plasma ratio of S-desmethylcitalopram)



501

503 Figure 2: Goodness-of-fit plots of escitalopram with a) observed concentrations (DV) vs. population predictions (PRED), b) DV vs. individual predictions (IPRED), c) conditional weighted 504 505 residuals (CWRES) vs. PRED and d) CWRES vs. time after dose (TAD) (solid line: unity line)



**Figure 3**: Goodness-of-fit plots of S-desmethylcitalopram with a) observed concentrations (DV) vs. population predictions (PRED), b) DV vs. individual predictions (IPRED), c) conditional weighted residuals (CWRES) vs. PRED and d) CWRES vs. time after dose (TAD) (solid line: unity line)





**Figure 4**: Prediction-corrected visual predictive checks (pcVPC) of the final covariate model for a) escitalopram in plasma, b) S-desmethylcitalopram in plasma, c) escitalopram in breast milk and d) S-desmethylcitalopram in breast milk. (circles: prediction-corrected drug concentrations; continuous line: population median prediction; dashed lines: 2.5<sup>th</sup> and 97.5<sup>th</sup> percentiles; semitransparent grey fields: model-based percentile confidence interval).



519

- 521 Figure 5: Relative infant dose (RID) and adult dose equivalent (ADE) of escitalopram in 5'000-
- 522 10'000 simulated mother-infant pairs under various conditions (median ± range).



523