

# Identification of compounds active against quiescent artemisinin-resistant Plasmodium falciparum parasites via the quiescent-stage survival assay (QSA)

Thibaud Reyser, Lucie Paloque, Manel Ouji, Michel Nguyen, Sandie Ménard, Benoit Witkowski, Jean-Michel Augereau, Françoise Benoit-Vical

# ▶ To cite this version:

Thibaud Reyser, Lucie Paloque, Manel Ouji, Michel Nguyen, Sandie Ménard, et al.. Identification of compounds active against quiescent artemisinin-resistant Plasmodium falciparum parasites via the quiescent-stage survival assay (QSA). Journal of Antimicrobial Chemotherapy, 2020, 75 (10), pp.2826-2834. 10.1093/jac/dkaa250. hal-02948398

# HAL Id: hal-02948398 https://hal.science/hal-02948398v1

Submitted on 6 Oct 2021

**HAL** is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

1	Identification of compounds active against quiescent artemisinin-resistant
2	Plasmodium falciparum parasites via the Quiescent-stage Survival Assay
3	
4	Thibaud REYSER <sup>1,2,\$</sup> , Lucie PALOQUE <sup>1,2,\$,‡</sup> , Manel OUJI <sup>1,2</sup> , Michel NGUYEN <sup>1,2</sup> ,
5	Sandie MENARD <sup>3</sup> , Benoit WITKOWSKI <sup>4</sup> , Jean-Michel AUGEREAU <sup>1,2</sup> ,
6	Françoise BENOIT-VICAL <sup>1,2,5,‡*</sup>
7	
8	<sup>1</sup> LCC-CNRS, Laboratoire de Chimie de Coordination, Université de Toulouse, CNRS,
9	Toulouse, France.
LO	<sup>2</sup> Institut de Pharmacologie et de Biologie Structurale, IPBS, Université de Toulouse, CNRS
l1	UPS, France.
L2	<sup>3</sup> Present adress: CPTP, Centre de Physiopathologie de Toulouse Purpan, INSERM U1043,
L3	CNRS UMR5282, Université de Toulouse, Toulouse, France
L4	<sup>4</sup> Malaria Molecular Epidemiology Unit, Pasteur Institute in Cambodia, Phnom Penh,
L5	Cambodia.
L6	<sup>5</sup> INSERM, Institut National de la Santé et de la Recherche Scientifique, France
L7	
L8	Running title: Compounds active against quiescent artemisinin-resistant malaria parasites
L9	
20	

CNRS, LCC (Laboratoire de Chimie de Coordination), 205 route de Narbonne, BP 44099, F-31077 Toulouse Cedex 4, France. Phone: +00 33 561 175 823

 $<sup>\</sup>hbox{* Author for correspondence: } \underline{Francoise.Vical@inserm.fr}$ 

<sup>\$</sup> These authors contributed equally to this work. \*LP and FBV share senior authorship.

22

#### **ABSTRACT**

23

**Background**: Quiescence is an unconventional mechanism of *Plasmodium* survival mediating 24 artemisinin resistance. This phenomenon increases the risks of clinical failures following 25 Artemisinin-based Combination Therapies (ACTs) by slowing parasite clearance and allowing 26 the selection of parasites resistant to partner drugs. 27 **Objectives**: To thwart this multi-resistance, the quiescent state of artemisinin-resistant parasites 28 must be taken into consideration from the very early stages of the drug discovery process. 29 30 Methods: We designed a novel phenotypic assay named Quiescent-stage Survival Assay (QSA) to assess the antiplasmodial activity of drugs on quiescent parasites. This assay was first 31 validated on quiescent forms from different artemisinin-resistant parasite lines (laboratory 32 33 strain and field isolates), using two reference drugs with different mechanisms of action: chloroquine and atovaquone. Furthermore, the efficacies of different partner drugs of 34 artemisinins used in ACTs were investigated against both laboratory strains and field isolates 35 from Cambodia. 36 **Results**: Our results highlight that because of quiescence-based mechanism and the respective 37 38 pharmacological targets of drugs, their efficacies on artemisinin-resistant parasites may be different between quiescent parasites and their proliferating forms. 39 **Conclusions**: These data confirm the high relevance of adding the chemosensitivity evaluation 40 41 of quiescent parasites by the specific in vitro QSA to the antiplasmodial drug development

process in the current worrisome context of artemisinin resistance.

43

42

44

48

49

50

51

52

53

54

55

56

57

58

59

60

61

62

63

64

65

66

67

68

69

70

#### INTRODUCTION

Artemisinin-based combination therapies (ACTs), using artemisinin or its derivatives (ARTs) combined with other antiplasmodial drugs, are recommended for the treatment of *Plasmodium* falciparum malaria. ARTs produce a fast reduction in the parasite numbers, leaving the remaining parasites exposed to the long-lasting partner drug. <sup>1</sup> The widespread use of ACTs has significantly contributed to the historical decrease of malaria mortality observed over the last 15 years in all endemic countries. <sup>2</sup> However, *Plasmodium* resistance to ARTs emerged in 2006 and could now jeopardize malaria elimination programs. 3-7 ARTs-resistance is linked to a mutation on the kelch13 gene, 8 which encodes for a protein involved in different mechanisms, among which hemoglobin uptake. 9 ARTs-resistance is mediated by a quiescent state upon ARTs exposure, <sup>10</sup> in a mechanism that dramatically differs from other antimalarial drug resistance mechanisms. This parasite cell cycle arrest is characterized by a drastic downregulation of the overall metabolism to allow parasites to face ARTs-induced cellular damage, 11,12 and subsequent up-regulation to enable parasite recrudescence after drug elimination. <sup>10,13</sup> In the field, ACT-resistance is always associated with ARTs-resistance. Either ARTs-resistance facilitates emergence of ACT-resistance by allowing partner drug resistance selection in artemisinin resistant parasites or ACT-resistance is acquired after recombination between ARTs-resistant and partner drug-resistant parasite lineages. 7,14-19 Long-term artemisinin drug pressure also triggers in vitro acquisition of a multidrug tolerance phenotype based on the extended capacity of ARTs-resistant parasites to survive other antiplasmodial drugs through a quiescence mechanism. <sup>20</sup> Extending ACT treatment from 2 to 3 days in order to improve clinical efficacy thus increases artemisinin drug pressure, leading to a higher frequency of P. falciparum ARTs-resistance cases. 21,22 During ACT treatment, due to the respective pharmacokinetics properties of both drugs, parasites are first exposed to ARTs. That is why, in an ARTs-resistance scenario, ring-stage parasites first enter a quiescent state and are then exposed to partner drugs. Therefore, a partner drug that is active against quiescent parasites should be the gold standard for malaria eradication policies and programs. Because standard chemo-sensitivity assays have shown their inability to study artemisinin resistance, <sup>8,10,20</sup> we propose a novel *in vitro* assay named Quiescent-stage Survival Assay (QSA) to assess the efficacy of compounds against ARTs-induced quiescent parasites.

#### MATERIALS AND METHODS

#### Parasite culture

F32-ART5 (artemisinin-resistant), F32-TEM (artemisinin-sensitive) *Plasmodium falciparum* laboratory strains were cultivated at 37°C in RPMI-1640 medium (Biowest) at 2 % hematocrit in human red blood cells and 5 % AB human serum (EFS, French Blood Bank) under 5 % CO<sub>2</sub> in a humid atmosphere at 37°C. <sup>10,20,23,24</sup> Artemisinin-resistant field isolates (from Pasteur Institute of Cambodia) IPC8262, IPC8317, IPC6388 and IPC6938 were cultivated under 5 % O<sub>2</sub>, 5 % CO<sub>2</sub>, and 90 % N<sub>2</sub> (FIVAL, Air Liquide) at 2 % hematocrit in RMPI-1640 medium supplemented with 5 % human serum (EFS, French Blood Bank) and 0.55 % (w/w) albumax II (Thermo Fisher). <sup>25</sup> The characteristics of the different strains used in this study are reported in Table 1. Genotyping of field isolates were given by Pasteur Institute of Cambodia.

# Drugs

Chloroquine diphosphate, lumefantrine and mefloquine hydrochloride were purchased from Sigma-Aldrich (France), artemisinin from TCI Europe, atovaquone from GlaxoSmithKline, dihydroartemisinin, *N*-desethyl amodiaquine and piperaquine tetraphosphate tetrahydrate were provided by the WWARN. Drugs were used *in vitro* in combination assays at pharmacologically relevant concentrations (700 nM dihydroartemisinin, 3 µM artemisinin,

200 nM chloroquine, 7 μM atovaquone, 200 nM piperaquine, 964 nM mefloquine, 800 nM *N*-desethyl amodiaquine, 5 μM lumefantrine), equal or close to their plasma peak concentrations in patients. <sup>26</sup> Stock solutions were obtained by dissolving piperaquine in 0.5 % lactic acid (Sigma-Aldrich), *N*-desethyl amodiaquine and chloroquine in RPMI-1640, while the other compounds were dissolved in DMSO (Sigma-Aldrich).

# Standard chemosensitivity assay

The chemosensitivity assay was carried out in 96-well culture plates on D-sorbitol (Sigma-Aldrich) ring-stage synchronized parasites. <sup>27</sup> The antiplasmodial activities were evaluated using the SYBR Green I method. <sup>28</sup> Each drug concentration was tested in triplicates. The parasites were incubated with the drugs for 48 h. Parasite pellets were then washed in 1X PBS prior to lysing red blood cells at -20 °C overnight. Then, the plates were thawed and 100  $\mu$ L of each well were transferred into a black 96-well plate. 100  $\mu$ L of SYBR Green I (Thermo Fisher), diluted at a final concentration of 2X in lysis buffer, were added to each well and incubated for 1 h at room temperature prior to reading the plates on BioTek FLx800 Microplate Fluorescence Reader ( $\lambda$ excitation = 485 nm,  $\lambda$ emission = 528 nm). IC<sub>50</sub> values were determined using GraphPad Prism software.

## **Quiescent-stage Survival Assay (QSA)**

To evaluate drug activity in quiescent artemisinin-resistant parasites, the Quiescent-stage Survival Assay (QSA) was first developed using the F32-ART5 strain. D-sorbitol synchronized ring-form cultures at 3 % parasitemia and 2 % hematocrit (5 mL of culture per well in 6-well plate) were first exposed to dihydroartemisinin at 700 nM for 6 h to induce quiescence of artemisinin-resistant parasites. Then parasite cultures were exposed to the combination, dihydroartemisinin + compound to be tested, for 48 h (Fig. 1A, condition a). Keeping parasites

under dihydroartemisinin treatment during exposure to the compound to be tested ensured quiescence maintenance. In parallel, other combinations were tested as control conditions (Fig. 1A, conditions b-d). The compounds were also evaluated on proliferating artemisinin-resistant parasites i.e. without any pre-treatment (Fig. 1A, condition e). Between each drug treatment, parasite cultures were washed with 40 mL of RPMI-1640 medium and put back in culture in new wells for the next treatment. After drug exposure, parasite cultures were washed with 40 mL of RPMI-1640 medium and cultured in drug-free conditions supplemented with 10 % human serum in new wells. The parasitemia for each condition was then microscopically monitored every 2 days to determine the time required to reach the initial parasitemia (3 %), this time point being referred to as recrudescence day. Culture media were renewed twice a week and 25 µL of fresh red blood cells were added once a week. If no recrudescence occurred after 30 days, the experiment was stopped. <sup>10,20</sup> Delay in recrudescence time is the difference between the median recrudescence time from different independent experiments obtained with the drug combination (dihydroartemisinin 6 h / (dihydroartemisinin + drug) 48 h) (Fig. 1A, condition a) and the median recrudescence time obtained with dihydroartemisinin alone (dihydroartemisinin 6 h / dihydroartemisinin 48 h) (Fig. 1A, condition b). This delay is representative of the capacity of the tested compound to be active against quiescent parasites. Concentrations and treatment durations were chosen for their pharmacological relevance. During the QSA development, several conditions were tested to optimize the assay: i) 24-h or 48-h incubation of the compounds, ii) use of a physiological dose of 3 µM of ART or 700 nM of DHA to induce quiescence, iii) repetition of the drug combination treatments to mimic ACT pharmacology, and iv) 2 sorbitol treatments, at 30 h and 54 h during drug exposure to be sure that parasites were in a quiescent state.

144

145

143

121

122

123

124

125

126

127

128

129

130

131

132

133

134

135

136

137

138

139

140

141

142

#### Recrudescence assay

Recrudescence assay was carried out as previously reported to evaluate the ability of the F32-ART5 parasite line to survive drug exposure comparatively to the artemisinin-sensitive F32-TEM parasite line. <sup>10,20</sup> After 48-h drug exposure, synchronized ring-stage parasites were washed with RPMI-1640 medium and placed in normal drug-free culture conditions. The time to parasite recrudescence was determined as the time required to reach initial parasitemia. The experiment was stopped if no recrudescence was microscopically observed within 30 days. <sup>10,20</sup>

#### **Ring-stage Survival Assay (RSA)**

The Ring-stage Survival Assay (RSA) was carried out as previously reported. <sup>29</sup> Briefly, 0-3 hours old ring-stage parasites (obtained after a Percoll® (Sigma-Aldrich)-sorbitol treatment) were exposed to 700 nM DHA or 0.1 % DMSO (control) for 6 hours in duplicate, then washed in RPMI-1640 and replaced in normal culture conditions for the next 72 h. Survival rates were measured microscopically on Diff-Quick® (RAL Diagnostics)-stained thin blood smears. The parasitemia was calculated by counting 10,000 red blood cells per assay by two independent microscopists. A parasite growth rate  $\geq$  2 in non-exposed culture was deemed necessary to validate the assay. The strains with a survival rate of dihydroartemisinin-treated parasites relative to control above 1 % in RSA were referred to as artemisinin-resistant. <sup>29</sup>

## Piperaquine survival assay (PSA)

The piperaquine survival assay (PSA) was performed as previously described to evaluate the ability of ring-stage parasites to survive piperaquine exposure. <sup>30</sup> Briefly, 0-3 h old ring-stage parasites (obtained after a percoll-sorbitol treatment) were exposed to 200 nM of piperaquine or culture medium alone (control) for 48 h in duplicate. After drug exposure, parasites were washed with RPMI-1640 medium and then cultivated for the next 24 h without drug. Survival rates were measured similarly to the RSA (ratio of viable parasites in piperaquine-exposed

relative to DMSO-exposed samples). The strains with a survival rate above 10 % in PSA were referred to as piperaquine-resistant. <sup>30</sup>

#### **Data Analysis**

Where relevant, a statistical analysis (log-rank test) was carried out on data from recrudescence assay and QSA by performing Kaplan-Meier survival analyses considering censored data. The outcome was defined as the time necessary for cultures to reach initial parasitemia. Observations were considered as censored if no recrudescence was observed within the experiment timeframe (30 days).

#### **RESULTS AND DISCUSSION**

The QSA (quiescent-stage survival assay), a relevant method to specifically evaluate

# chemosensitivity of quiescent parasites

Quiescence-based artemisinin resistance is a new challenge to consider when proposing relevant antiplasmodial treatments. In this context, we designed a novel assay, the QSA, to evaluate drug activity against artemisinin-resistant parasites in a quiescence state. The QSA interpretation is based on a delay in recrudescence time between cultures exposed to (dihydroartemisinin 6 h / (dihydroartemisinin + molecule) 48 h) and cultures exposed to (dihydroartemisinin 6 h / dihydroartemisinin 48 h) (Fig. 1A). A delayed recrudescence (Fig. 1B) or a lack thereof indicates that most or all quiescent parasites are killed by the partner drug tested. Therefore, it is suggested that the more potent the drug against quiescent parasites, the longer the delay because the fewer quiescent parasites, the slower the culture will grow back to initial parasitemia. According to the different results obtained and in order to select relevant compounds, we considered a molecule to be active against quiescent parasites if its recrudescence delay was above a 6-day threshold (Fig. 1B). Below this threshold, the difference

could be statistically significant but not necessarily biologically relevant since this could be due to variations in culture parameters independent of the number of surviving parasites. Therefore, from one experiment to the other, there can be a two to three-day variation in recrudescence days, which is why, by setting a 6-day confidence interval, we made sure that the differences were meaningful. To develop the QSA model, chloroquine and atovaquone were used because of their different and well-documented mechanisms of action. The highly artemisinin-resistant *P. falciparum* strain F32-ART5 harboring none of the genotypic markers associated with chloroquine, mefloquine or atovaquone resistance was used (Table 1). <sup>10,20</sup> Results were then confirmed on artemisinin-resistant isolates from the field.

# Chloroquine cannot kill quiescent parasites

In the absence of dihydroartemisinin pre-treatment, F32-ART5 proliferative parasites appeared sensitive to a single dose of 200 nM chloroquine for 48 h leading to parasite elimination with no recrudescence at day 30 (Fig. 2, Table S1). This chloroquine sensitivity of F32-ART5 is in accordance with the obtained  $IC_{50}$  values below the chloroquine-resistance cut-off of 100 nM (Table S2). The combination dihydroartemisinin 6 h / chloroquine 48 h delays recrudescence time of resistant parasites by 7 days compared to dihydroartemisinin 6 h (Table S1). This delayed recrudescence time could be interpreted as the ability of chloroquine to either be active against quiescent parasites or to repress cell cycle recovery of quiescent parasites. To ensure the targeting of quiescent parasites, the quiescent state was maintained with dihydroartemisinin all over the chloroquine treatment. Surprisingly, the 2 conditions (dihydroartemisinin 6 h / (dihydroartemisinin + chloroquine) 48 h) and (dihydroartemisinin 6 h / dihydroartemisinin 48 h) showed similar recrudescence profiles (8 and 6 days, respectively) (Fig. 2, Table S1), which is interpreted as a lack of chloroquine activity against quiescent parasites. The mechanism of action of chloroquine, through the inhibition of hemozoin formation, could

explain this result since such a mechanism is absent at the quiescent ring-stage. <sup>31</sup> A recent study showed that chloroquine is active against both early ring stages and trophozoites, suggesting that hemoglobin digestion starts early on in the asexual cycle. 32 Hence, this showcases that the efficacy of a drug against quiescent parasites cannot be deduced from its efficacy against proliferating parasites. The QSA condition (dihydroartemisinin 6 h / (dihydroartemisinin + drug to be tested) 48 h) appears thus highly selective for identification of compounds genuinely active against quiescent forms. These results evidenced that drug activity differs according to the proliferating or quiescent state of parasites. Therefore, QSA appears as a global chemosensitivity assay that is able to evaluate drug activity against both artemisininresistant proliferating and quiescent parasites. In order to better mimic the antimalarial drug regimens, the impact of repetitive chloroquine treatments was tested on quiescent parasites. Because a repeated (dihydroartemisinin 6 h / dihydroartemisinin 48 h) treatment did not yield any recrudescence (data not shown), the 48 hexposure was thus reduced to 24 h in each cycle. In these conditions, two successive treatments (dihydroartemisinin 6 h / (dihydroartemisinin + chloroquine ) 24 h / dihydroartemisinin 6 h / (dihydroartemisinin + chloroquine) 24 h) led to the same recrudescence time of 12 days as a single 60 h exposure to dihydroartemisinin alone (Table S3). This result was consistent with the total absence of chloroquine activity against quiescent parasites, independent of the treatment duration.

240

241

242

243

244

245

221

222

223

224

225

226

227

228

229

230

231

232

233

234

235

236

237

238

239

## Atovaquone targets both quiescent and proliferating parasites

The delayed parasite recrudescence time of 12 days for the combination (dihydroartemisinin 6 h / (dihydroartemisinin + atovaquone) 48 h) compared to dihydroartemisinin alone (Fig. 2, Table 2 and Table S4) evidenced that atovaquone is not only highly active against quiescent parasites but also against proliferating artemisinin-resistant parasites (Fig. 1, control condition

e). This is in accordance with previously published atovaquone activity towards the mitochondrial electron transport chain, <sup>33,34</sup> whose activity seems maintained during quiescence. <sup>10,12,35</sup>

#### The newly-designed QSA specifically targets quiescent parasites

The induction of quiescence by ARTs had already been demonstrated. <sup>10</sup> Here, two sorbitol treatments were performed during the QSA to eliminate any potential non ring-stage and thus non-quiescent parasites or awaking parasites (Fig. 3A). Regardless of the treatment conditions and the drugs used, sorbitol treatment did not lead to any delay in the recrudescence times compared to the no-sorbitol condition (Fig. 3B, Table S5). These data confirm that the QSA allows the specifically evaluation of the activities of drugs on *P. falciparum* quiescent parasites.

#### QSA applied to artemisinin-resistant field isolates confirms its stage specificity

The QSA was also performed on artemisinin-resistant isolates from the field to confirm its potential as a new screening tool for antimalarial development. Chloroquine and atovaquone were tested on four *P. falciparum* Cambodian isolates all harboring the C580Y mutation on the *kelch13* gene and having survival rates above 1 % in RSA, confirming their artemisinin resistance (Table 1). Notably, all the isolates used harbored a chloroquine-resistant phenotype with IC<sub>50</sub> values superior to 100 nM (Table S2). This phenotype was confirmed by the fast parasite recrudescence after 48-h chloroquine treatment (2-day recrudescence time for IPC8262, IPC6938, IPC6388, and 6-day recrudescence time for IPC8317, compared to no recrudescence of the chloroquine-sensitive F32-ART strain) (Fig. 2, Table S1). Independent of the isolate used, the QSA confirmed the lack of chloroquine activity towards quiescent parasites with no significant delay between drug combination and the dihydroartemisinin control (Table 2, Table S1).

The efficacy of atovaquone was confirmed regardless of the field isolate and the combination 271 272 tested, with delays in recrudescence times all superior (at least 6 days) to the control condition (dihydroartemisinin 54 h) supporting atovaquone activity against both quiescent (conditions a-273 b) and artemisinin-resistant proliferating parasites (condition e) from the field (Fig. 2, Table 2, 274 275 Table S4). By validating the QSA protocol on both laboratory and field-isolated artemisinin-resistant 276 strains, we evidenced that this newly designed protocol was able to evaluate the activity of 277 compounds against artemisinin-resistant parasites both in the specific artemisinin-induced 278 quiescent state and in proliferating forms. 279 280 ACT partner drugs display different activities against artemisinin-resistant quiescent 281 parasites 282 283 After standardizing the QSA protocol, the activity of WHO-recommended ACT partner drugs, namely N-desethyl amodiaguine (the active metabolite of amodiaguine), piperaguine, 284 lumefantrine, and mefloquine were evaluated on quiescent parasites. 285 Although active towards proliferative parasites (Table S6, Fig. 1A condition e), N-desethyl 286 287 amodiaquine showed a very weak activity against quiescent parasites from the laboratory strain 288 F32-ART5 and the field isolate IPC8317 with a delay of 3-4 days (Fig. 2, Table S6), which is below the 6-day delay threshold. This lack of efficacy towards quiescent parasites could be 289 explained by the mode of action of the molecule which, like chloroquine, <sup>36,37</sup> targets the 290 polymerization of heme into hemozoin, a mechanism absent during quiescence. This is further 291 corroborated by the stage specificity of N-desethyl amodiaquine, which is mostly active against 292 trophozoites. 38 293 In a similar fashion, a weak piperaquine activity was observed in QSA against quiescent 294 parasites with a delay in recrudescence time below the 6-day threshold for F32-ART5, 295

IPC8317, IPC8262 and IPC6388, (Fig. 2, Table 2, Table S7). These strains harbor a single copy of the pm2 gene (except for IPC8262 with 1.58 pm2 copies) and a PSA survival rate below 10 % (Table 1). Firstly, the absence of delay between F32-ART and F32-TEM in piperaquine recrudescence assay and PSA data confirm that the partial lack of piperaquine activity against quiescent parasites is not linked with any piperaquine resistance or cross-resistance with artemisinin (Table S8). Secondly, these results are in accordance with the purported mode of action of piperaquine which is thought to interfere with heme detoxification in a similar fashion and with similar stage specificity than chloroquine. <sup>32,39</sup> The field isolate IPC6938 showed no recrudescence after treatment with piperaquine alone whereas this line harbors a PSA survival rate of 10.7 % and 3 copies of the pm2 gene consistent with piperaquine resistance (Table 1). These conflicting results could be explained by the age of ring-stage parasites during drug exposure in the different assays. Indeed, when PSA was performed on ring-stage parasites older than 6 hours (condition similar to the QSA), the survival rate dropped to 0.16 %. Unlike the other parasite lines, preliminary data obtained with this isolate showed a delayed recrudescence with piperaquine above 6 days in QSA (Table 2, Table S7). Further experiments should be conducted in order to clarify the impact of piperaquine resistance on quiescent parasites. In contrast, lumefantrine appeared active against quiescent parasites both on the laboratory strain F32-ART5 and on the parasite isolate from the field IPC8317 with a recrudescence time delay above 9 days (Fig. 2, Table 2, Table S9). Recently, lumefantrine has been shown to be active against all but the schizont stage. 32 These data, in addition to our results which also showed its activity against quiescent parasites, seem to indicate that lumefantrine targets other pathway(s) apart from the inhibition of hemozoin formation. <sup>40</sup> Mefloquine also showed an activity against quiescent parasites on all the tested strains with very high recrudescence delays (Fig. 2, Table 2, Table S10). One field isolate, IPC6388, showed

296

297

298

299

300

301

302

303

304

305

306

307

308

309

310

311

312

313

314

315

316

317

318

319

a faster recrudescence after treatment with mefloquine alone compared to the other strains, correlating with a higher IC<sub>50</sub> value. Consistently, this isolate harbors 3 copies of the *mdr1* gene which is associated with mefloquine resistance (Table 1). Interestingly, its recrudescence delay was of 6 days in QSA. Therefore, these results suggest that although mefloquine-resistant parasites in quiescent state are less sensitive to the drug than mefloquine-sensitive parasites, they can still be killed by mefloquine treatment. Its activity against quiescent parasites is surprising considering its specificity towards trophozoites. <sup>32</sup> It cannot be explained by its ability to impair hemoglobin uptake, <sup>41</sup> hemozoin biosynthesis, <sup>42</sup> or even inhibit protein synthesis by targeting the 80S ribosome, <sup>43</sup> since these different pathways are supposed to be inactive during quiescence. Although the effect of dihydroartemisinin treatment on metacaspase transcripts remains to be determined, <sup>44</sup> the mefloquine activity against quiescent parasites could be due to its capacity to trigger an apoptotic process by *Plasmodium* metacaspase activation leading to parasite death. <sup>45,46</sup>

#### Targeting quiescent parasites can be achieved with different treatment schedules

In order to optimize the QSA protocol, different quiescence-inducing drugs and different drug schedules were tested against F32-ART5 in combination with 4 antimalarials namely chloroquine, atovaquone, piperaquine and mefloquine. Similar activities were obtained regardless of the artemisinin derivative used (3 µM of artemisinin or 700 nM of dihydroartemisinin) or the treatment duration (24 h or 48 h; Table S11) except when treating quiescent parasites with piperaquine. Indeed, a 24-h exposure to piperaquine has no effect on quiescent parasites as shown by the lack of delay between the combination and the control regardless of the drug used to induce quiescence (Table S11), whereas a 48-h exposure to piperaquine led to a 5-day delay (Table 2). Because ACT partner drugs have half-lives of several days, we reasoned that lengthening drug exposure to 48 hours (corresponding to a

hypothetical complete erythrocytic cycle) allowed better approximation of physiological conditions.

Tests of molecules against quiescent parasites had already been reported. <sup>11,47–49</sup> However, our results underlined the importance of i) pre-treatment by ARTs to induce quiescence, ii) maintaining quiescence throughout the whole drug evaluation, and iii) comparing the combination dihydroartemisinin / (dihydroartemisinin + drug to be tested) to dihydroartemisinin / dihydroartemisinin in order to be able to conclude on the genuine efficacy of the drug towards quiescent parasites.

# Utility and limitations of the QSA

Standard chemosensitivity assays are proliferation-based and hence, cannot allow the evaluation of the missing variable of partner drug activity against quiescent parasites. Because of its experimental design, the QSA presented herein can overcome these limitations. This assay appears as a highly relevant tool to study artemisinin-induced quiescence in *P. falciparum* parasites. It also constitutes an *in vitro* model that tries to grasp the pharmacological complexity of ACTs by triggering quiescence before exposing parasites to the partner drug. Compounds tested were used at high concentrations over 48 hours to approximate their physiological conditions since ACT partner drugs have half-lives of several days and high plasma peak concentrations. <sup>26</sup> Therefore, these parameters could be fine-tuned when evaluating the activity of other compounds. The assay was not designed to compare the activity of different drugs on quiescent parasites but rather to determine whether a drug efficiently kills quiescent parasites. Moreover, it does not allow direct assessment of parasite death for obvious quantitation limitations. The outcome of the assay is thereby indirect as it relies on a delayed parasite recrudescence, correlated with the ability of a drug to kill quiescent parasites.

In conclusion, thanks to the QSA, we highlighted that i) drug activity against quiescent parasites depends on the metabolic status of its target throughout quiescence, regardless of the drug's stage specificity in asexual replication, ii) because of quiescence-based mechanism, drug efficacy against artemisinin-induced quiescent forms of the *falciparum* malaria parasite is unrelated to its activity against proliferating parasites, and iii) some molecules active against ARTs-induced quiescent parasites can be identified and can shed light on pathways that remain active within this dormant state, potentially guiding the search for new antimalarials. Custom ACTs through the addition of a third compound are already evaluated in clinical trials. <sup>50</sup> However the added value of QSA-based drug design would be to enable the design of novel ACTs that effectively target dormant parasites. This strategy could greatly enhance ACT clinical efficacy and drastically limit the emergence of resistant parasites. Ultimately, this approach could also confer an extended lifespan to ACTs even in a drug resistance context.

#### Acknowledgements

We would like to thank Chinedu O. Egwu for proofreading the article and we would also like to acknowledge the WorldWide Antimalarial Resistance Network (WWARN) Reference Standards Program for reference drug providing.

# **Funding**

This study was supported in part by the Centre National de la Recherche Scientifique and the French Agence Nationale de la Recherche (ANR-13-BSV3-0018 & ANR-13-CE35-0003). T. R. was funded by a Graduate Fellowship Program from the University Paul Sabatier of Toulouse. The funding sources of the study had no role in the study design, data collection, data analysis, data interpretation or writing of the report.

396 Transparency declarations

None to declare.

#### 399 References

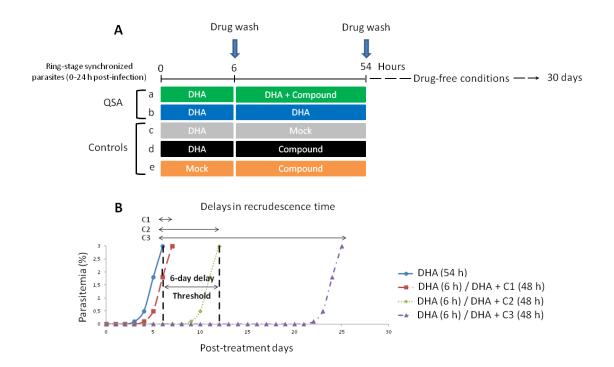
- 400 1. White N. Antimalarial drug resistance and combination chemotherapy. *Philos Trans R Soc B Biol*
- 401 *Sci* 1999; **354**: 739–49.
- 402 2. World Health Organization. World Malaria Report 2019. 2019.
- 403 https://www.who.int/publications-detail/world-malaria-report-2019
- 3. Noedl H, Se Y, Schaecher K et al. Evidence of artemisinin-resistant malaria in Western Cambodia.
- 405 N Engl J Med 2008; **359**: 2619–20.
- 4. Dondorp AM, Nosten F, Yi P et al. Artemisinin Resistance in Plasmodium falciparum Malaria. N
- 407 *Engl J Med* 2009; **361**: 455–67.
- 408 5. Ashley EA, Dhorda M, Fairhurst RM et al. Spread of Artemisinin Resistance in Plasmodium
- 409 *falciparum* Malaria. *N Engl J Med* 2014; **371**: 411–23.
- 410 6. Ménard D, Khim N, Beghain J et al. A worldwide map of Plasmodium falciparum K13-propeller
- 411 polymorphisms. *N Engl J Med* 2016; **374**: 2453–64.
- 7. World Health Organization. *Status report on Artemisinin and ACT resistance (April 2017)*. 2017.
- 413 http://www.who.int/malaria/publications/atoz/artemisinin-resistance-april2017/en/
- 8. Ariey F, Witkowski B, Amaratunga C et al. A molecular marker of artemisinin-resistant
- 415 *Plasmodium falciparum* malaria. *Nature* 2014; **505**: 50–5.
- 416 9. Birnbaum J, Scharf S, Schmidt S et al. A Kelch13-defined endocytosis pathway mediates
- artemisinin resistance in malaria parasites. *Science* 2020; **367**: 51–59.
- 418 10. Witkowski B, Lelièvre J, Barragán MJL et al. Increased tolerance to artemisinin in Plasmodium
- falciparum is mediated by a quiescence mechanism. Antimicrob Agents Chemother 2010; 54: 1872–7.
- 420 11. Chen N, LaCrue AN, Teuscher F et al. Fatty Acid Synthesis and Pyruvate Metabolism Pathways
- Remain Active in Dihydroartemisinin-Induced Dormant Ring Stages of *Plasmodium falciparum*.
- 422 *Antimicrob Agents Chemother* 2014; **58**: 4773–81.
- 423 12. Paloque L, Ramadani AP, Mercereau-puijalon O et al. Plasmodium falciparum: multifaceted
- 424 resistance to artemisinins. *Malar J* 2016; **15**: 149.
- 425 13. Witkowski B, Khim N, Chim P et al. Reduced artemisinin susceptibility of Plasmodium
- 426 falciparum ring stages in western cambodia. Antimicrob Agents Chemother 2013; 57: 914–23.

- 427 14. Na-Bangchang K, Muhamad P, Ruaengweerayut R et al. Identification of resistance of
- 428 Plasmodium falciparum to artesunate-mefloquine combination in an area along the Thai-Myanmar
- border: integration of clinico-parasitological response, systemic drug exposure, and in vitro parasite
- 430 sensitivity. *Malar J* 2013; **12**: 263.
- 431 15. Saunders DL, Vanachayangkul P, Lon C. Dihydroartemisinin-piperaquine failure in Cambodia. N
- 432 *Engl J Med* 2014; **371**: 484–5.
- 433 16. Spring MD, Lin JT, Manning JE et al. Dihydroartemisinin-piperaquine failure associated with a
- 434 triple mutant including kelch13 C580Y in Cambodia: An observational cohort study. *Lancet Infect Dis*
- 435 2015; **15**: 683–91.
- 436 17. Dondorp AM, Fairhurst RM. Artemisinin-Resistant *Plasmodium falciparum* Malaria. *Microbiol*
- 437 *Spectr* 2016; **4**: 409–29.
- 438 18. Amaratunga C, Lim P, Suon S et al. Dihydroartemisinin-piperaquine resistance in *Plasmodium*
- falciparum malaria in Cambodia: a multisite prospective cohort study. Lancet Infect Dis 2016; 16:
- 440 357–65.
- 19. Amato R, Pearson RD, Almagro-Garcia J et al. Origins of the current outbreak of multidrug-
- resistant malaria in southeast Asia: a retrospective genetic study. *Lancet Infect Dis* 2018; **18**: 337–45.
- 20. Ménard S, Ben Haddou T, Ramadani AP et al. Induction of multidrug tolerance in Plasmodium
- falciparum by extended artemisinin pressure. *Emerg Infect Dis* 2015; **21**: 1733–41.
- 21. Putaporntip C, Kuamsab N, Kosuwin R et al. Natural selection of K13 mutants of Plasmodium
- falciparum in response to artemisinin combination therapies in Thailand. Clin Microbiol Infect 2016;
- **22**: 285.e1-8.
- 448 22. WHO. Status report on artemisinin resistance. 2014.
- https://www.who.int/malaria/publications/atoz/status-rep-artemisinin-resistance-sep2014.pdf
- 450 23. Benoit-Vical F, Lelièvre J, Berry A et al. Trioxaquines are new antimalarial agents active on all
- 451 erythrocytic forms, including gametocytes. Antimicrob Agents Chemother 2007. 51: 1463-72
- 452 24. Wenger RH, Kurtcuoglu V, Scholz CC *et al.* Frequently asked questions in hypoxia research.
- 453 *Hypoxia* (Auckland, NZ) 2015; **3**: 35–43.
- 454 25. Niaré K, Paloque L, Ménard S et al. Multiple phenotypic and genotypic artemisinin sensitivity

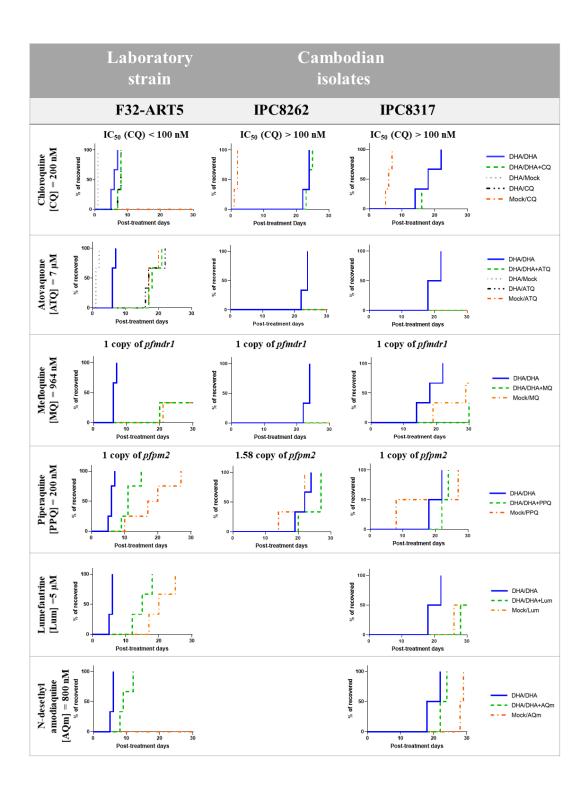
- evaluation of malian *Plasmodium falciparum* isolates. *Am J Trop Med Hyg* 2018; **98**: 1123–31.
- 456 26. Nosten F, White NJ. Artemisinin-based combination treatment of falciparum malaria. *Am J Trop*
- 457 *Med Hyg* 2007; **77**: 181–92.
- 458 27. Lambros C, Vanderberg JP. Synchronization of *Plasmodium falciparum* Erythrocytic Stages in
- 459 Culture. *J Parasitol* 1979; **65**: 418–20.
- 28. Smilkstein M, Sriwilaijaroen N, Kelly JX et al. Simple and Inexpensive Fluorescence-Based
- Technique for High-Throughput Antimalarial Drug Screening. *Antimicrob Agents Chemother* 2004;
- **462 48**: 1803–6.
- 29. Witkowski B, Amaratunga C, Khim N et al. Novel phenotypic assays for the detection of
- artemisinin-resistant *Plasmodium falciparum* malaria in Cambodia: In-vitro and ex-vivo drug-response
- 465 studies. *Lancet Infect Dis* 2013; **13**: 1043–9.
- 30. Duru V, Khim N, Leang R et al. Plasmodium falciparum dihydroartemisinin-piperaquine failures
- in Cambodia are associated with mutant K13 parasites presenting high survival rates in novel
- piperaquine in vitro assays: Retrospective and prospective investigations. *BMC Med* 2015; **13**: 305.
- 31. Fitch CD. Ferriprotoporphyrin IX, phospholipids, and the antimalarial actions of quinoline drugs.
- 470 *Life Sci* 2004; **74**: 1957–72.
- 471 32. Murithi JM, Owen ES, Istvan ES *et al.* Combining Stage Specificity and Metabolomic Profiling to
- 472 Advance Antimalarial Drug Discovery. Cell Chem Biol 2020; 27: 158-171.e3.
- 473 33. Nixon GL, Pidathala C, Shone AE et al. Targeting the mitochondrial electron transport chain of
- 474 Plasmodium falciparum: New strategies towards the development of improved antimalarials for the
- 475 elimination era. *Future Med Chem* 2013; **5**: 1573–91.
- 476 34. Siregar JE, Kurisu G, Kobayashi T et al. Direct evidence for the atovaquone action on the
- 477 *Plasmodium* cytochrome bc1 complex. *Parasitol Int* 2015; **64**: 295–300.
- 478 35. Peatey CL, Chavchich M, Chen N et al. Mitochondrial Membrane Potential in a Small Subset of
- 479 Artemisinin-Induced Dormant *Plasmodium falciparum* Parasites in Vitro. *J Infect Dis* 2015. **212**: 426-
- 480 34
- 481 36. Bray PG, Mungthin M, Ridley RG *et al.* Access to Hematin: The Basis of Chloroquine Resistance.
- 482 *Mol Pharmacol* 1998; **54**: 170–9.

- 483 37. Ginsburg H, Famin O, Zhang J et al. Inhibition of glutathione-dependent degradation of heme by
- chloroquine and amodiaquine as a possible basis for their antimalarial mode of action. *Biochem*
- 485 *Pharmacol* 1998; **56**: 1305–13.
- 486 38. Wilson DW, Langer C, Goodman CD et al. Defining the timing of action of antimalarial drugs
- 487 against Plasmodium falciparum. Antimicrob Agents Chemother 2013; 57: 1455–67.
- 488 39. Sachanonta N, Chotivanich K, Chaisri U et al. Ultrastructural and real-time microscopic changes
- in *P. falciparum*-infected red blood cells following treatment with antimalarial drugs. *Ultrastruct*
- 490 *Pathol* 2011; **35**: 214–25.
- 491 40. Warhurst DC, Adagu IS, Beck H-P et al. Mode of action of artemether/lumefantrine
- 492 (COARTEM): The sole, fixed, oral ADCC and its role in combatting multidrug-resistance. *Southeast*
- 493 *Asian J Trop Med Public Health* 2001; **32**: 4–8.
- 494 41. Famin O, Ginsburg H. Differential effects of 4-aminoquinoline-containing antimalarial drugs on
- hemoglobin digestion in *Plasmodium falciparum*-infected erythrocytes. *Biochem Pharmacol* 2002; **63**:
- 496 393–8.
- 42. Sullivan DJ, Matile H, Ridley RG et al. A common mechanism for blockade of heme
- 498 polymerization by antimalarial quinolines. *J Biol Chem* 1998; **273**: 31103–7.
- 499 43. Wong W, Bai XC, Sleebs BE et al. Mefloquine targets the Plasmodium falciparum 80S ribosome
- to inhibit protein synthesis. *Nat Microbiol* 2017; **2**: 17031
- 501 44. Shaw PJ, Chaotheing S, Kaewprommal P et al. Plasmodium parasites mount an arrest response to
- dihydroartemisinin, as revealed by whole transcriptome shotgun sequencing (RNA-seq) and
- microarray study. *BMC Genomics* 2015; **16**: 830.
- 504 45. Gunjan S, Singh SK, Sharma T et al. Mefloquine induces ROS mediated programmed cell death in
- malaria parasite: *Plasmodium. Apoptosis* 2016; **21**: 955–64.
- 46. Meslin B, Beavogui AH, Fasel N et al. Plasmodium falciparum metacaspase PfMCA-1 triggers a
- 507 z-VAD-fmk inhibitable protease to promote cell death. *PLoS One* 2011; **6**: e23867.
- 508 47. Teuscher F, Gatton ML, Chen N et al. Artemisinin-Induced Dormancy in Plasmodium falciparum:
- 509 Duration, Recovery Rates, and Implications in Treatment Failure. *J Infect Dis* 2010; **202**: 1362–8.
- 48. Dembele L, Gupta DK, Yi-Xiu Lim M et al. Imidazolopiperazines Kill both Rings and Dormant

- Rings in Wild-Type and K13 Artemisinin-Resistant *Plasmodium falciparum* in vitro. *Antimicrob*
- 512 *Agents Chemother* 2018; **62**: e02235-17.
- 513 49. Clements RL, Streva V, Dumoulin P et al. A novel antiparasitic compound kills ring-stage
- 514 Plasmodium falciparum and retains activity against artemisinin-resistant parasites. J Infect Dis 2020;
- **221**: 956-62
- 50. Thu AM, Phyo AP, Landier J et al. Combating multidrug-resistant Plasmodium falciparum
- 517 malaria. *FEBS J* 2017; **284**: 2569–78.

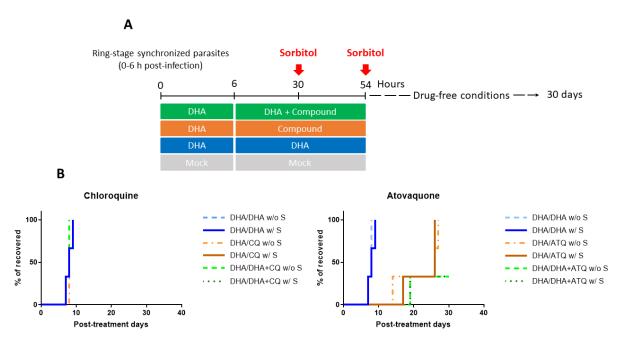


**Figure 1.** QSA (Quiescent-stage Survival Assay) design and interpretation. **A:** Treatment schedule of the different drug combinations studied (DHA: dihydroartemisinin); a-b) conditions to assess activity of the molecule on quiescent parasites; c-e) control conditions. **B:** Schematic curves of recrudescence profiles of artemisinin-resistant parasites to reach the initial 3 % parasitemia obtained *via* the QSA after 54-h treatment with DHA 700 nM alone or in combination with different compounds tested (C1, C2 and C3). C1 appears inactive, C2 is active with a 6-day delay (corresponding to the designated threshold) between **a** and **b** conditions and C3 is highly active against quiescent parasites with a delay largely higher than the 6-day threshold. Double-headed arrows highlight delay in recrudescence times between combination treatments (DHA / DHA + C1, C2 or C3) and DHA (54 h) alone.



**Figure 2.** QSA (Quiescent-stage Survival Assay) results represented by Kaplan-Meier survival curves showing recrudescence days of synchronous ring-stage parasites of the laboratory artemisinin-resistant strain F32-ART5 and two artemisinin-resistant Cambodian isolates after

different treatments. D0 corresponds to the day when all drugs were removed. Raw data are presented in supplementary tables. These 3 strains are artemisinin-resistant (RSA survival rates > 1 %), piperaquine-sensitive (PSA survival rates < 10%) and mefloquine-sensitive.



**Figure 3.** Quiescence specificity of the Quiescent-stage Survival Assay. **A:** Sorbitol treatments schedule applied on parasites submitted to QSA conditions. In this experiment, two consecutive sorbitol treatments were conducted at 24-h interval to make sure that dihydroartemisinintreatment maintained parasites in a quiescent state, i.e. that parasites did not proceed with their cell cycle. Treating parasite cultures with a 5 % sorbitol solution selectively eliminates all possible forms older than 24 h post-invasion preventing any recrudescence. Comparing the effect of sorbitol treatment for a given treatment schedule will thus reveal if the QSA protocol is indeed adequate to target quiescent forms of the parasite.

**B:** Kaplan-Meier survival curves on F32-ART5 showing days needed for cultures to reach initial parasitemia with or without sorbitol treatment in different conditions. Raw data are presented in supplementary Table S5.

w/S (with sorbitol), w/o S (without sorbitol).

**Table 1.** Phenotypic and genotypic traits of the *P. falciparum* strains used in this study

Strain	Description	Genotype				RSA survival	PSA survival
Suam		pfkelch13	pfcrt	pfpm2 copy number	pfmdr1 copy number	rate (%)	rate (%) <sup>b</sup>
<b>F32-ART5</b>	Laboratory	M476I	WT	1 copy	1 copy	10.7 <sup>a</sup>	0.4
F32-TEM	Laboratory	WT	WT	1 copy	1 copy	0.09 a	ND
IPC8262	Field	C580Y	ND	1.58 copy	1 copy	13 <sup>b</sup>	2.7
IPC8317	Field	C580Y	ND	1 copy	1 copy	8.8 °	0
IPC6388	Field	C580Y	ND	1 copy	3 copies	6.8 °	1.9
IPC6938	Field	C580Y	ND	3 copies	1 copy	6.6 °	10.7

558

556

ND: Not Determined; WT: Wild Type

559 a n=3

560 <sup>b</sup> n=2

561 ° n=1

562

563

**Table 2**. Difference in median recrudescence times from independent experiments obtained for the artemisinin-resistant strain F32-ART5 and 4 Cambodian ART-resistant isolates following exposure to different antimalarial drugs in QSA conditions (DHA 6 h / (DHA + compound 48 h)) compared to DHA alone (DHA 6 h / DHA 48 h). Raw data are presented in supplementary tables.

	Delay in recrudescence time (post-treatment days)							
_	CQ	ATQ	AQm	PPQ	LUM	MQ		
F32-ART5	2	12	4	5	10	> 24		
IPC8262	0	> 6		5		> 6		
IPC8317	0	> 10	3	3	> 9	> 12		
IPC6938	3	> 6		> 6		> 6		
IPC6388	0	14		3		6		

CQ (chloroquine), ATQ (atovaquone), AQm (N-desethyl amodiaquine), PPQ (piperaquine), LUM (lumefantrine), MQ (mefloquine).

> means that for at least one of the three replicates, no recrudescence was observed up to 30 days, in which case, the value was censored.