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Preparation of alginate hydrogel microparticles using droplet-based microfluidics: a review of methods

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Abstract

This review examines the preparation of alginate hydrogel microparticles. We focus on droplet-based microfluidics, a technique widely employed for its ease of use and excellent control of physicochemical properties, with narrow size distribution. Alginate hydrogel microparticles are obtained by conducting “on-chip” and/or “off-chip” gelation. Various strategies are described and compared. Microparticle properties such as size, shape, concentration, stability and mechanical properties are discussed. Finally, we consider future perspectives for the preparation of hydrogel microparticles and their potential applications.

Key words alginate, hydrogel, microparticle, crosslinking, droplet-based microfluidics

Introduction

Hydrogel microparticles are widely used today, especially in biological and pharmaceutical applications. They are usually used as a matrix to encapsulate bioactive agents such as drugs, proteins, cells, etc. (Ahmed, 2015; Enck et al., 2020; Maitra and Shukla, 2014) in applications like drug delivery (Agüero et al., 2017), cell culture and tissue engineering. (Utech et al., 2015) In addition, fluorescence-encoded hydrogel microparticles are extensively employed in multiplex bioassays. (Shikha et al., 2018; Zhang et al., 2019; Zhao et al., 2020) Another important use is as cell-mimicking microparticles with similar size, shape, deformability and mechanical properties. (Haghgoie et al., 2010; Merkel et al., 2011; Zhang et al., 2020) Hydrogels can be made of various biopolymers such as gelatine, agarose, alginate, pectin, etc. Alginate stands out because of its low cost, non-toxicity and ease of crosslinking (Lee and Mooney, 2012).

With the increasing interest in alginate hydrogel microparticles, various preparation methods have been reported in the literature, including conventional emulsification (Chan et al., 2002), spray-drying (Santa-Maria et al., 2012), extrusion dripping (Barbieri et al., 2017; Lee et al., 2013), microfluidics (Rondeau and Cooper-White, 2008; Yu et al., 2019; Zhang et al., 2006) and soft lithography (Qiu et al., 2007). The huge diversity of techniques and strategies can make it confusing to choose the right method. The present review focuses on a microparticle-producing technique widely used for its efficacy in controlling physicochemical properties: droplet-based microfluidics. The various strategies applied within this technique and the properties of the microparticles obtained are described in this review.

I. Alginate hydrogel microparticles

Hydrogels are described as hydrophilic polymeric networks which can absorb and retain large amounts of water within the structure. The hydrogel network is formed by polymer crosslinking. When crosslinking is realized by molecular entanglement, ionic, H-bonding or hydrophobic forces, hydrogels are called physical or reversible gels. Otherwise, when covalent forces intervene, they are called chemical or permanent gels. (Caccavo et al., 2018; Hoffman, 2012)

I.1. Alginate

Alginate is a natural polysaccharide. Although it can also be synthesized by several bacteria, all the commercially available alginate is produced from the extraction of brown algae (Draget, 2009). Alginate is widely used in the biomedical field because it is biocompatible and non-toxic (Lee and Mooney, 2012).

Sodium alginate (Na-alginate) is the most widely used alginate salt. It dissolves in water to a viscous solution. Alginate is a linear copolymer containing β -D-mannuronate (M) and α -L-guluronate (G) residues (Figure 1).

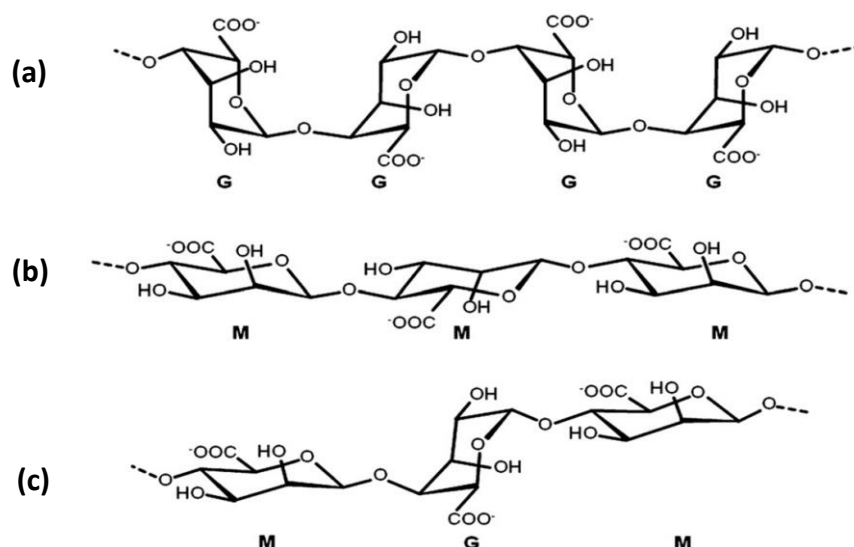


Figure 1. Chemical structures of (a) G-block, (b) M-block and (c) alternating G and M-blocks in alginate. Figure reprinted with permission from Reference (Lee and Mooney, 2012)

I.2. Gelation of alginate

Alginate hydrogel is produced by gelation which is caused by covalent (Fundueanu et al., 1999) or ionic crosslinking. (Gacesa, 1988; Velings and Mestdagh, 1995) Ionic crosslinking is more commonly used because of its simplicity and mild conditions. It can be carried out at room temperature or up to 100°C , usually with divalent cations as crosslinking agents. Calcium chloride is the most widely used (Lee and Mooney, 2012), due to its non-toxicity (Agüero et al., 2017) and availability.

Only G-blocks (Figure 1a) made of consecutive G residues can participate in ionic crosslinking because of their favourable spatial structure (Gacesa, 1988; Lee and Mooney, 2012). Ionic crosslinking of alginate is described by the “egg-box” model (Grant et al., 1973) (Figure 2).

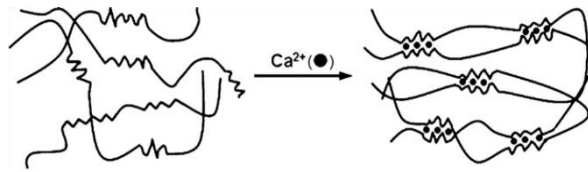


Figure 2. Schematic illustration of the “egg-box” model describing the ionic crosslinking of alginate by calcium cations. Figure reprinted with permission from Reference (Lee and Yuk, 2007)

In this paper, we present various two-step methods of producing alginate hydrogel microparticles. First, sodium alginate droplets are generated using droplet-based microfluidics. Second, internal or external gelation transforms droplets into alginate hydrogel microparticles via different strategies. In internal gelation, the crosslinking agents are inside the alginate droplet whereas in external gelation, the crosslinking agents come from outside the alginate droplet.

II. Droplet-based microfluidics

II.1. Principle of droplet generation

Microfluidics is a technique used to manipulate fluids in channels of micrometric dimensions. Fluids are mixed by adding junctions that connect the channels. When immiscible or partially miscible fluids are mixed in the junction, microdroplets can be generated: this is called droplet-based microfluidics.

The principle is similar to that of conventional emulsification, which consists of blending two immiscible liquids. The advantage of droplet-based microfluidics is monodispersity and repeatability of droplets due to precise control over experimental conditions such as channel geometry, flow rates and viscosities of fluids, etc. (Seemann et al., 2011; Teh et al., 2008) Furthermore, monodisperse droplets can be generated without using surfactant (Liu et al., 2006; Trivedi et al., 2009; Zhang et al., 2020), which is impossible with conventional emulsification.

The droplets generated in droplet-based microfluidics can serve as microreactors to carry out physical, chemical or biological reactions. (Zhu and Wang, 2017) Being small (nL to μ L volume), they require a small quantity of reactants. As droplet composition can be made identical, numerous identical experiments can be performed, enabling a reliable statistical approach to data.

II.2. Flow properties

In droplet-based microfluidics, a continuous fluid and a dispersed fluid are injected separately and then mixed in a junction. Fluids are Newtonian and droplets of the dispersed fluid (**D**) are generated in the flow of the continuous fluid (**C**). The physicochemical properties influencing droplet formation are density, dynamic viscosity, surface tension between the continuous and the dispersed fluids, velocity of the flows and characteristic dimensions of the microfluidic system, such as the diameter of channels (**w**) for cylindrical microfluidic systems. Based on these properties, fluid dynamics is characterized as follows:

- 1- Inertial forces and viscous forces are compared through the Reynolds number, calculated using the continuous fluid properties: density (ρ_c), dynamic viscosity (μ_c) and flow velocity (v_c).

$$Re = \frac{\rho_C \times v_C \times w}{\mu_C} \quad (1)$$

Typically for microfluidics, values of Re are lower than 1: the flow is laminar and the effect of inertia can be ignored. Thus, the average velocity v of a flow is evaluated from its volumetric flow rate Q and w as follows:

$$v = \frac{Q}{\pi(w/2)^2} \quad (2)$$

- 2- The generation of droplets in a microfluidic junction creates a free interface between the two fluids, characterized by the interfacial energy γ_{CD} . The corresponding capillary effects are in competition with gravity effects. The length above which gravity effects dominate capillary effects is the capillary length l_c :

$$l_c = \sqrt{\frac{\gamma_{CD}}{\Delta\rho \times g}} \quad (3)$$

with g the gravity acceleration and $\Delta\rho$ the difference in density between the two fluids. For instance, with fluorinated oil FC70 as the continuous fluid and ethanol as the dispersed fluid, l_c is equal to 2.4 mm (Zhang et al., 2015). Hence gravity does not influence the deformation of the interfaces in millimetric or sub-millimetric channels.

- 3- Shear stress and interfacial energy are compared through the capillary number Ca . When generating droplets of a dispersed fluid in a continuous fluid, Ca is usually calculated using v_C and μ_C of the continuous fluid, and γ_{CD} of the interface between the continuous and the dispersed fluid:

$$Ca = \frac{\mu_C \times v_C}{\gamma_{CD}} \quad (4)$$

II.3. Microfluidic geometry

Microfluidic devices can be in the form of either chips with microchannels and junctions produced by soft lithography, or an assembly of capillaries and junctions (Ren et al., 2013). In terms of materials, polydimethylsiloxane (PDMS) is the most commonly used for microfluidic chips (Liu et al., 2006; Zhang et al., 2006). For capillaries, both glass (Baroud et al., 2010; Hu et al., 2012) and fluoropolymer can be used. (Trivedi et al., 2009; Zhang et al., 2020)

The channel geometry of a microfluidic device influences droplet generation. Three frequently used geometries are “cross-flow”, “co-flow” and “flow-focusing” (Figure 3).

II.3.1. Cross-flowing

For cross-flowing geometry, continuous fluid and dispersed fluid mix with an angle ϑ ($0^\circ < \vartheta \leq 180^\circ$) at the junction (Figure 3 (a)). Where the two fluids meet, first an interface is formed due to the immiscibility of the two fluids. Shear force then pushes the head of the dispersed fluid into the continuous fluid until a part breaks off: the droplet is formed. Then it circulates in the channel of the continuous fluid. (Teh et al., 2008)

Cross-flowing geometry is often called T-junction geometry, where two fluids flow orthogonally (Figure 3 (a) i). However, other shapes of junctions can also be used, such as a junction with an arbitrary angle ϑ (Figure 3 (a) ii), or a Y-shaped junction (Figure 3 (a) iv). For two fluids facing each other ($\vartheta = 180^\circ$, Figure 3 (a) iii), the geometry is called “head-on”. A combination of two junctions (Figure 3 (a) v, vi) can also be used to introduce two different

dispersed fluids and one continuous fluid. Cross-flowing geometry is widely used due to its ease of assembly and handling. (Seemann et al., 2011; Zhu and Wang, 2017)

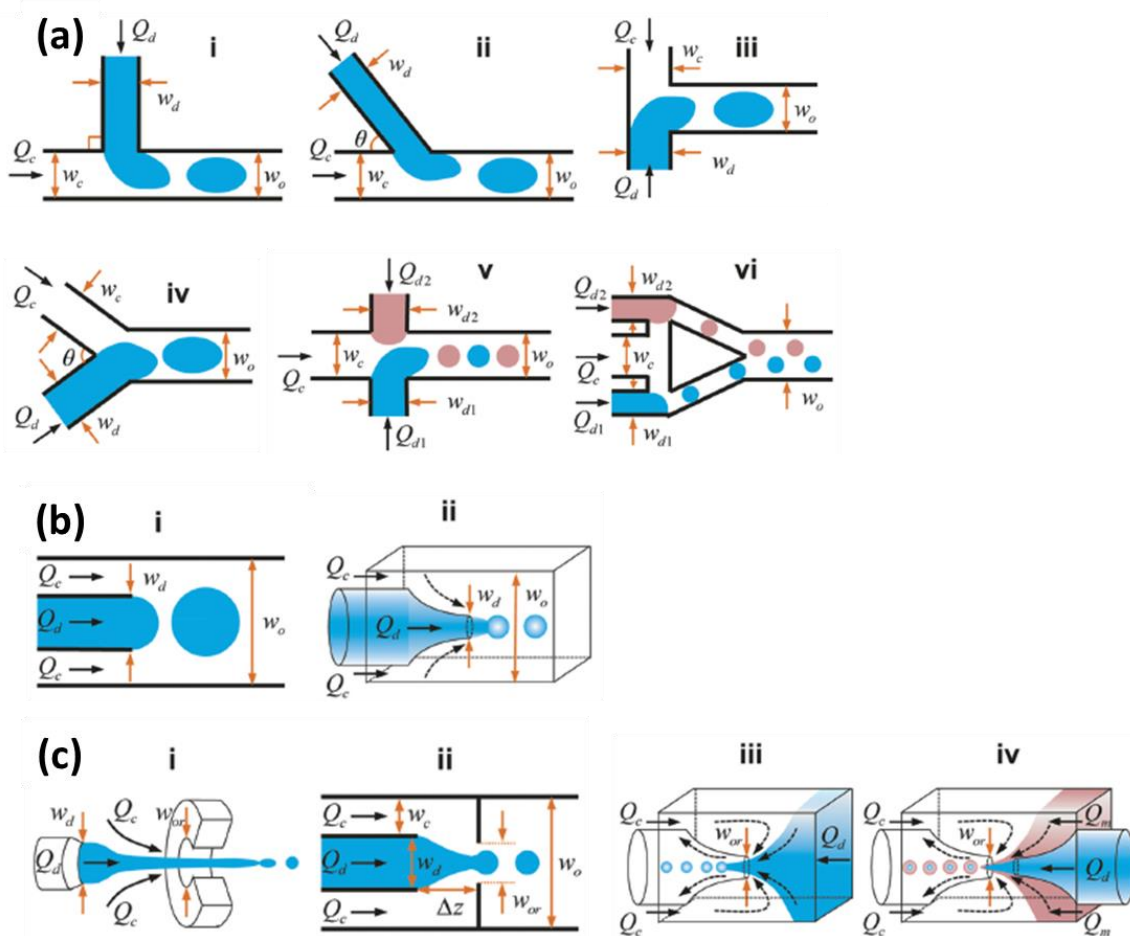


Figure 3. Schematic illustrations (a) “cross-flowing”, (b) “co-flow” and (c) “flow-focusing” geometries for a microfluidic device. Q and w denote respectively flow rate and channel width. Subscripts d , c , o and or denote respectively dispersed fluid, continuous fluid, outlet channel and orifice. Figure reprinted with permission from Reference (Zhu and Wang, 2017)

II.3.2. Flow-focusing

For co-flow geometry, two immiscible fluids flow in two concentric channels (Figure 3 (b)). Droplets are formed at the outlet of the inner channel. Flow-focusing geometry is actually similar to co-flow geometry. The distinction presented in the literature is somewhat ambiguous (Zhu and Wang, 2017), leading some to consider flow-focusing as a special co-flow geometry (Seemann et al., 2011). For flow-focusing geometry (Figure 3 (c)), two immiscible fluids are focused through an orifice, which allows smaller droplets to be generated than with co-flow geometry.

II.4. Droplet generation regime

For each geometry, droplets can be generated following three different break-off mechanisms. The transition from one mechanism to another can be achieved by varying capillary numbers

Ca. (Zhu and Wang, 2017) Figure 4 shows an example of three mechanisms for a cross-flowing geometry.

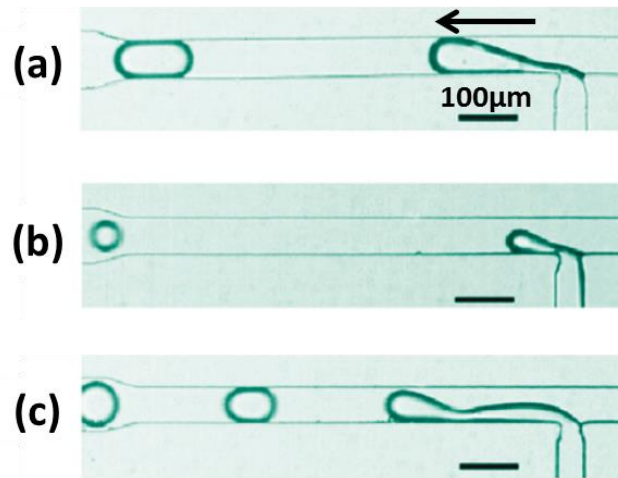


Figure 4. Three break-off mechanisms of droplet generation with a cross-flowing geometry: (a) squeezing, (b) dripping and (c) jetting. The arrow indicates the droplet flow direction. Figure reprinted with permission from Reference (Zagnoni et al., 2010). Copyright 2010 American Chemical Society.

II.4.1. Squeezing

As Figure 4 (a) shows, as it is injected into the principal channel, the dispersed fluid is pushed forward by the continuous fluid. A thin “neck” is thus formed. Because the continuous fluid applies weak shear force, the forming droplet reaches the opposing channel wall without breaking off. The neck becomes thinner until it breaks, so that a plug-shaped droplet confined by channel wall is formed. Squeezing mechanism appears when Ca is low ($Ca \leq 0.01$). (De Menech M. et al., 2008)

II.4.2. Dripping

As Figure 4 (b) shows, the shear force applied is now higher. The forming droplet breaks off before touching the opposing channel wall. A spherical droplet is formed with a diameter smaller than that of the channel. This dripping mechanism appears at a higher Ca ($Ca \geq 0.02$). (De Menech M. et al., 2008)

II.4.3. Jetting

As Figure 4 (c) shows, a liquid jet is emitted from the dispersed fluid channel. It flows and remains attached to the channel wall, due to a strong shear force from the continuous fluid. (Christopher and Anna, 2007) The jet breaks up into droplets at the end because of Rayleigh-Plateau instability. (Zhu and Wang, 2017) Droplets of polydisperse sizes are formed. This jetting mechanism appears at the highest Ca ($Ca \approx 0.2$).

III. Gelation

III.1. Internal gelation

For internal gelation, crosslinking agents are either soluble or insoluble/slightly soluble in water.

III.1.1. Water-soluble crosslinking agents

With water-soluble crosslinking agents such as barium chloride (BaCl_2) and calcium chloride (CaCl_2), alginate is crosslinked directly at the interior of droplets. These agents can be mixed with Na-alginate before or after droplet generation.

III.1.1.1. Mixing crosslinking agents before droplet generation

A first category of strategies is based on mixing water-soluble crosslinking agents with Na-alginate before droplet generation. The crosslinking agents used in these studies are BaCl_2 and CaCl_2 ; these strategies are summarized in Table 1.

Table 1. Internal gelation with water-soluble crosslinking agents: mixing crosslinking agents with Na-alginate before droplet generation

References		Trivedi et al., 2009	Zhang et al., 2006	Rondeau and Cooper-White, 2008	This paper	This paper
Droplet generation	Concentration of Na-alginate	1 wt%	0.5 wt%	0.5 wt%	0.06wt%	0.006wt%
	Continuous fluid	Silicone oil	Mineral oil	DMC*	DMC*	DMC*
	Use of surfactant	NO	Span 80	Not mentioned	NO	NO
	Geometry	Flow-focusing	Flow-focusing	Flow-focusing	Cross-flowing	Cross-flowing
	Microfluidic material	Fluoropolymer tubing and junctions	PDMS chip	PDMS chip	Fluoropolymer tubing and junctions	Fluoropolymer tubing and junctions
Internal gelation by mixing Na-alginate and water-soluble crosslinking agent	Crosslinking agent	BaCl_2 (50 mM)	CaCl_2 (0.1 wt%)	CaCl_2 (0.25 wt%)	CaCl_2 (0.06 wt%)	CaCl_2 (0.002 wt%)
	Geometry	Flow-focusing	Cross-flowing	Flow-focusing	-	Cross-flowing
	Mixing	Before droplet generation	During droplet generation	Before droplet generation	Off-line, before droplet generation	During droplet generation

* Continuous and dispersed fluids partially miscible

Trivedi et al. worked on cell encapsulation by alginate hydrogel microparticles. (Trivedi et al., 2009) For the preparation of microparticles, an aqueous solution of cell-containing Na-alginate (1 %) and a solution of BaCl_2 (50 mM) were injected into the capillary and mixed via a Y-shaped junction (Figure 5 (a)). At the exit from the mixing region, highly viscous silicone oil (10 centistokes) without surfactant was injected by flow-focusing in order to generate droplets. However, the mixing of Na-alginate and barium cations triggered ionic crosslinking,

causing gelation in the mixing region which impacted droplet generation. Finally, instead of generating droplets as expected, a jet of gel was produced with a partially formed droplet head and a long gelatinous tail (Figure 5 (b)).

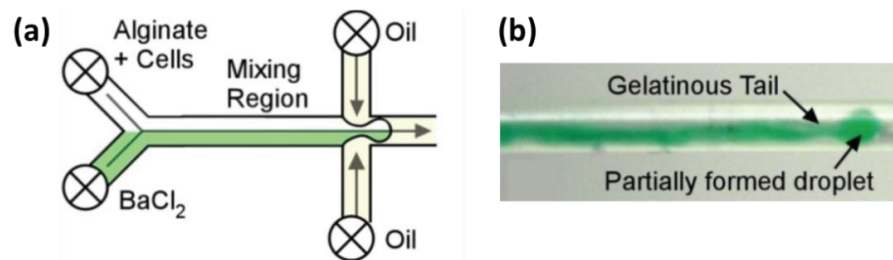


Figure 5. (a) Schematic diagram of mixing Na-alginate/cells and BaCl₂ solutions before droplet generation in a microfluidic device assembled from fluoropolymer capillaries and junctions. (b) Image of the jet of gel produced in oil. Figure reprinted from Reference (Trivedi et al., 2009)

To deal with this issue, the mixing region can be reduced before droplet generation, as Zhang et al. did (Zhang et al., 2006) Using a 5-channel microfluidic device, they mixed Na-alginate fluid (0.5 wt%), CaCl₂ fluids (0.1 wt%) and mineral oil fluids with a surfactant (Span 80, no concentration mentioned) as shown in Figure 6. Droplets were generated by co-flow. However, instead of producing discrete droplets, a line of knots connected with each other was formed. This phenomenon persisted with a wide range of flow rates of oil due to viscosity which increased instantly when Na-alginate and CaCl₂ were mixed, because of rapid gelation. It was therefore impossible to generate droplets at the junction, despite the use of surfactant.

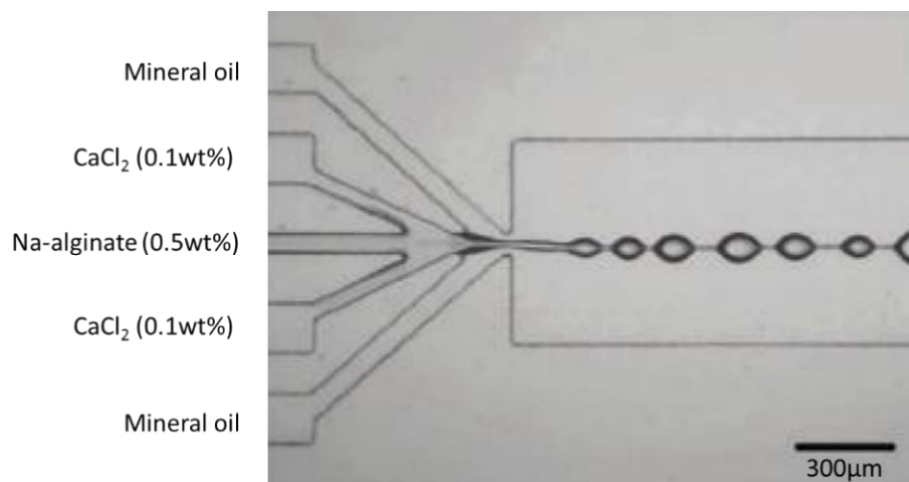


Figure 6. Image of connected knots formed after mixing Na-alginate and CaCl₂ solutions in mineral oil with surfactant, in a PDMS-based microfluidic chip. Figure reprinted with permission from Reference (Zhang et al., 2006). Copyright 2006 American Chemical Society.

The problem can be solved by using low concentrations of Na-alginate and CaCl₂ solutions. In this case, gelation proceeds after droplet generation and is enhanced by using partially miscible fluids. Rondeau and Cooper-White used dimethyl carbonate (DMC) as the continuous fluid (Rondeau and Cooper-White, 2008) (Figure 7). The solubility of water in DMC is about 3 wt% at room temperature. (Stephenson and Stuart, 1986) Aqueous solutions of Na-alginate (0.5 wt%) and CaCl₂ (0.25 wt%) were injected respectively from inlets A and B (Figure 7 (a)). After a short pre-gelation channel, DMC was injected from inlet C. Na-alginate/CaCl₂ droplets

were generated in DMC (no mention of surfactant usage) by flow-focusing. Along the serpentine channel, because of the low solubility of water in DMC, water diffused gradually from droplets into DMC, causing the shrinkage of droplets along the channel. Internal gelation occurred at the same time. Microparticles with a diameter of 20 μm were observed at the outlet of the channel and collected in an aqueous solution of CaCl_2 (2N) to reinforce the gelation (Figure 7 (b)). The diameter of Ca-alginate hydrogel microparticles was influenced by the experimental parameters such as the initial concentration of Na-alginate, flow rates of fluids and channel size. However, DMC is also slightly soluble in water, with a solubility of 12.7 wt% at 20°C. (Stephenson and Stuart, 1986) Thus, during diffusion of water from droplets into DMC, DMC can also diffuse into droplets. This means that, after gelation, DMC can be captured inside alginate hydrogel microparticles. Additional work measuring the amount of DMC residue within microparticles could open the way to further applications.

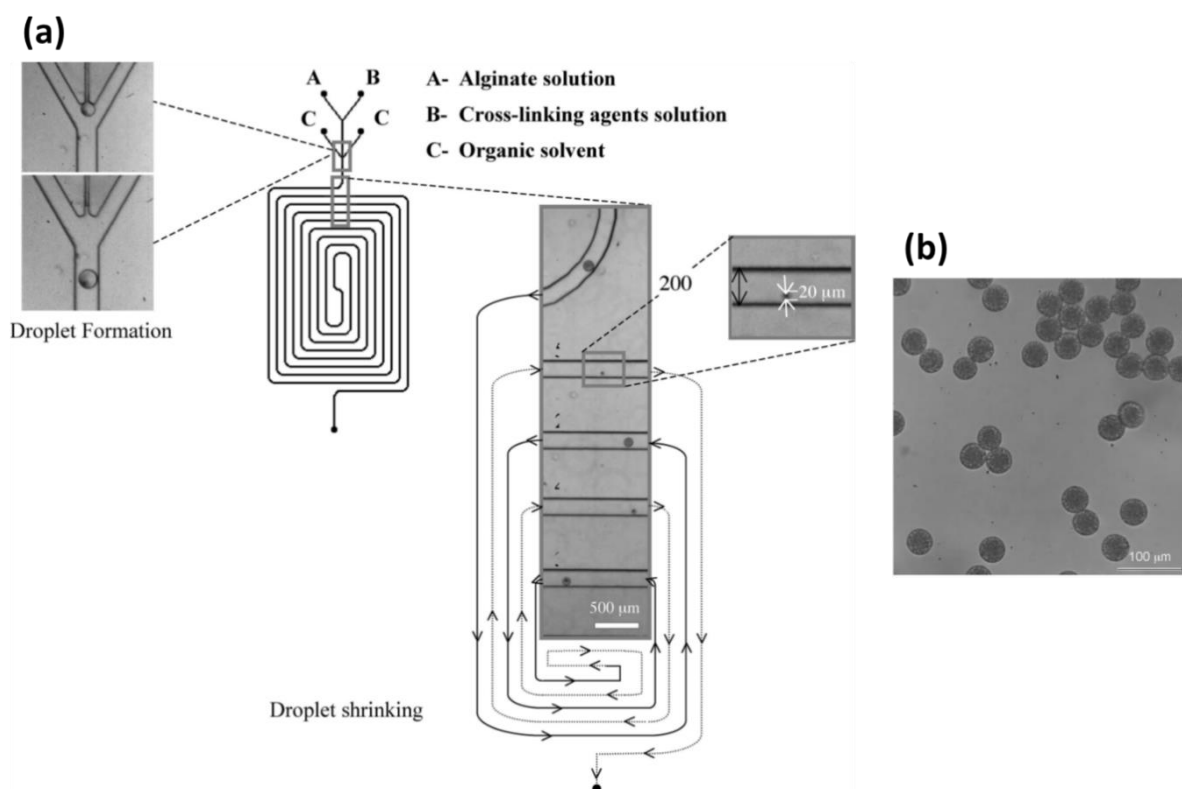


Figure 7. (a) Schematic diagram of a PDMS-based microfluidic device using DMC as the continuous fluid in which water is partially soluble. Droplet shrinkage is observed for an initial concentration of Na-alginate of 0.5 wt%. (b) Micrograph of Ca-alginate hydrogel microparticles collected in an aqueous solution of CaCl_2 . Figure reprinted with permission from Reference (Rondeau and Cooper-White, 2008). Copyright 2008 American Chemical Society.

Following the work of Rondeau and Cooper-White, we tested, in a T-junction (Figure 8), the direct generation of droplets of Ca-alginate in DMC without surfactant from a mixture of more diluted Na-alginate and CaCl_2 solutions (both at 0.06 wt% after mixing). However, this solution was not clear and local gelation was occasionally observed with the naked eye. When these gels entered the channel, droplets were generated in a discontinuous way. This indicates that, even at very low concentrations, thorough mixing of Na-alginate and CaCl_2 solutions leads to gelation, disturbing droplet generation.

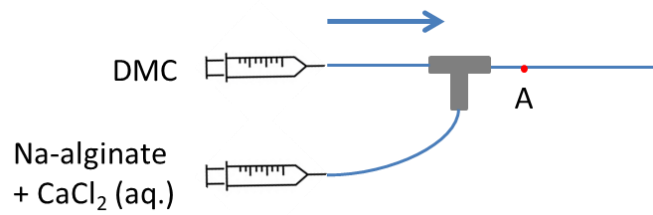


Figure 8. Schematic diagram of the generation of droplets of Ca-alginate with DMC as the continuous fluid and an aqueous mixed solution of Na-alginate and CaCl_2 as the dispersed fluid. The device is composed of polyether ether ketone (PEEK) junctions and Teflon-like tubing (IDEX Health and Science). The arrow indicates the flow direction.

In a microfluidic device (Figure 9) of similar design to Zhang et al., we were able to generate discrete droplets by using extremely diluted solutions of Na-alginate (0.006 wt%) and CaCl_2 (0.002 wt%). The continuous fluid was DMC without surfactant. Droplets were observed after the cross-junction (point A in Figure 9 (a)). Since they were relatively close to each other in the channel, causing coalescence at the outlet (point B in Figure 9 (a)), a second flow of DMC was introduced as a spacer using a T-junction. When the second DMC flow rate was relatively low, the generation of droplets upstream was not disturbed, so that droplets were uniform (Figure 9 (b)). However, the coalescence at the outlet persisted. Thus, high second DMC flow rates were applied to sufficiently increase the distance between droplets. Nevertheless, this quickly disturbed the generation of droplets upstream, as indicated by heterogeneities in droplet size and frequency (Figure 9 (c)). Using surfactant would prevent droplet coalescence.

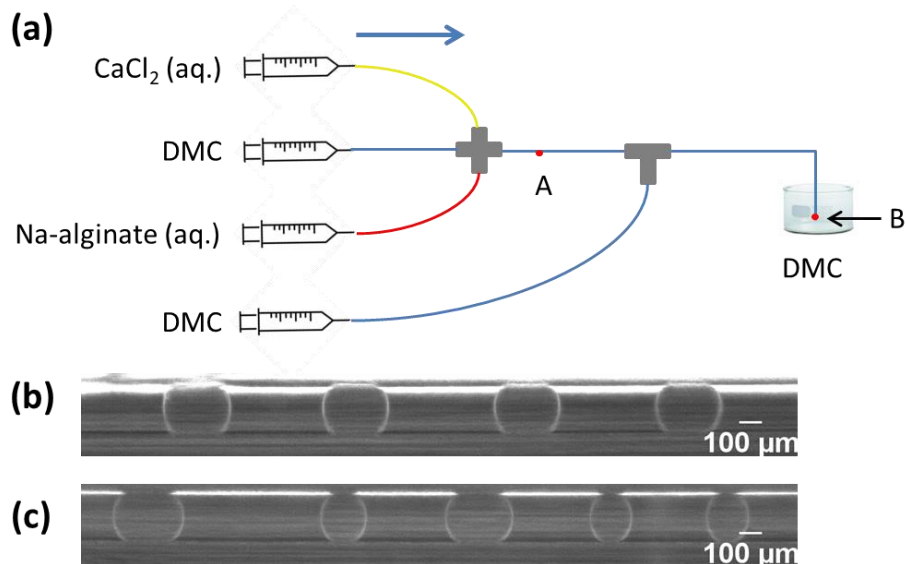


Figure 9. (a) Schematic diagram of the generation of Ca-alginate droplets in dimethyl carbonate (DMC) using a cross-junction. The T-junction served to introduce DMC as a spacer to increase the distance between droplets. The device is composed of polyether ether ketone (PEEK) junctions and Teflon-like tubing (IDEX Health and Science). Micrograph of droplets observed at point A when (b) droplet generation was not disturbed by introducing the spacer and (c) when it was disturbed.

To summarize (Table 1), authors mixed highly concentrated solutions of Na-alginate and water-soluble crosslinking agents before droplet generation to make them gelate. However,

droplet generation was hindered by rapid gelation and microparticles were difficult to obtain. To delay gelation, less concentrated solutions of Na-alginate and water-soluble crosslinking agents were mixed before droplet generation, in the microfluidic device or off-line. Then, concentrations were increased after droplet generation by diffusion of water from the droplets to the continuous fluid, due to their partial miscibility with water. Hence, gelation proceeded slowly with droplet shrinkage. However, mixing crosslinking agents with Na-alginate before droplet generation led to heterogeneities in droplet size and frequency.

III.1.1.2. Mixing crosslinking agents after droplet generation

To delay gelation, water-soluble crosslinking agents need to be mixed with Na-alginate only after droplet generation. Studies doing so, and which also use BaCl₂ and CaCl₂ as crosslinking agents, are summarized in Table 2.

Table 2. Internal gelation with water-soluble crosslinking agents: mixing crosslinking agents with Na-alginate after droplet generation

Reference		Xu et al., 2008	Liu et al., 2006	Trivedi et al., 2010
Droplet generation	Concentration of Na-alginate	2 wt%	2 wt%	1 wt%
	Continuous fluid	octyl alcohol oil	soybean oil	silicone oil
	Use of surfactant	Not mentioned	NO	NO
	Geometry	Flow-focusing	Flow-focusing	Flow-focusing
	Microfluidic material	PMMA chip	PDMS chip	Fluoropolymer tubing and junctions
Internal gelation by mixing Na-alginate and water-soluble crosslinking agent	Crosslinking agents	CaCl ₂ (2 wt%)	CaCl ₂ (2 wt%)	BaCl ₂ (50 mM)
	Geometry	Cross-flowing	Expansion chamber	Cross-flowing
	Mixing	Coalescence of flows	Coalescence of droplets	Coalescence of droplets with flow

Xu et al. prevented rapid gelation by delaying the direct contact between Na-alginate and calcium cations. (Xu et al., 2008) In a first cross-junction, two face-to-face channels were used to introduce CaCl₂ (2 wt%) and Na-alginate (2 wt%) solutions (Figure 10 (a)) perpendicularly to a flow of water. Thus, after the first cross-junction, a flow of water (acting as a buffer) separates the flows of Na-alginate and CaCl₂. Then octyl alcohol oil (no mention of surfactant) was injected at a second cross-junction. Droplets of Na-alginate/CaCl₂ were generated by flow-focusing. In the “synthesizing channel” (Figure 10 (a)), within each droplet, mixing Na-alginate and CaCl₂ induced internal gelation. In this way droplets were transformed into Ca-

alginate hydrogel microparticles (Figure 10 (b)). For this strategy, the size of Ca-alginate hydrogel microparticles is entirely dependent on the experimental conditions, such as flow rates of fluids and channel size. Manipulation of microparticles is difficult if their diameter is smaller than $10\mu\text{m}$.

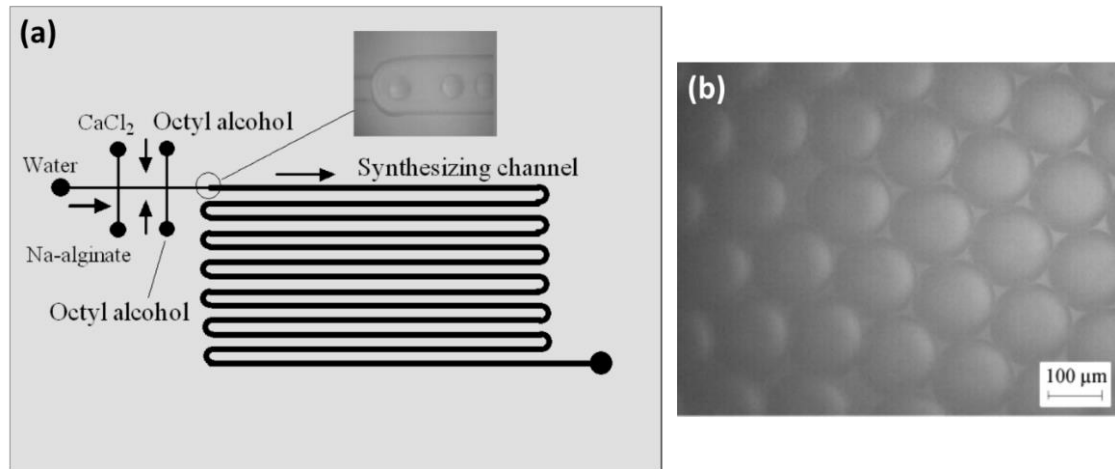


Figure 10. (a) Schematic diagram of Ca-alginate hydrogel microparticles prepared in a PMMA based microfluidic device. (b) Micrographs of Ca-alginate hydrogel microparticles. Figure reprinted with permission from Reference (Xu et al., 2008)

Another strategy to delay gelation was carried out by Liu et al. (Liu et al., 2006) involving coalescence of Na-alginate droplets with CaCl_2 droplets generated separately. First, on a microfluidic chip (Figure 11 (a)), Na-alginate (2 wt%) droplets (Figure 11 (b)) and CaCl_2 (2 wt%) droplets (Figure 11 (c)) were generated in soybean oil without surfactant by flow-focusing using two independent cross-junctions. Then droplets converged via a T-junction (Figure 11 (d)) followed by two successive circular expansion chambers (Figure 11 (d, e)). Thus, droplets could collide either at the T-junction or in circular chambers. Within the coalesced droplets, Na-alginate was crosslinked by calcium cations forming Ca-alginate hydrogel microparticles. With different flow rates and channel geometries, various shapes and sizes of microparticles could be produced (Figure 11 (f)). Nevertheless, the design of circular expansion chambers gives rise to local changes in flow velocity. Droplet circulation can be disturbed, thereby affecting homogeneity in droplet shape, size and frequency.

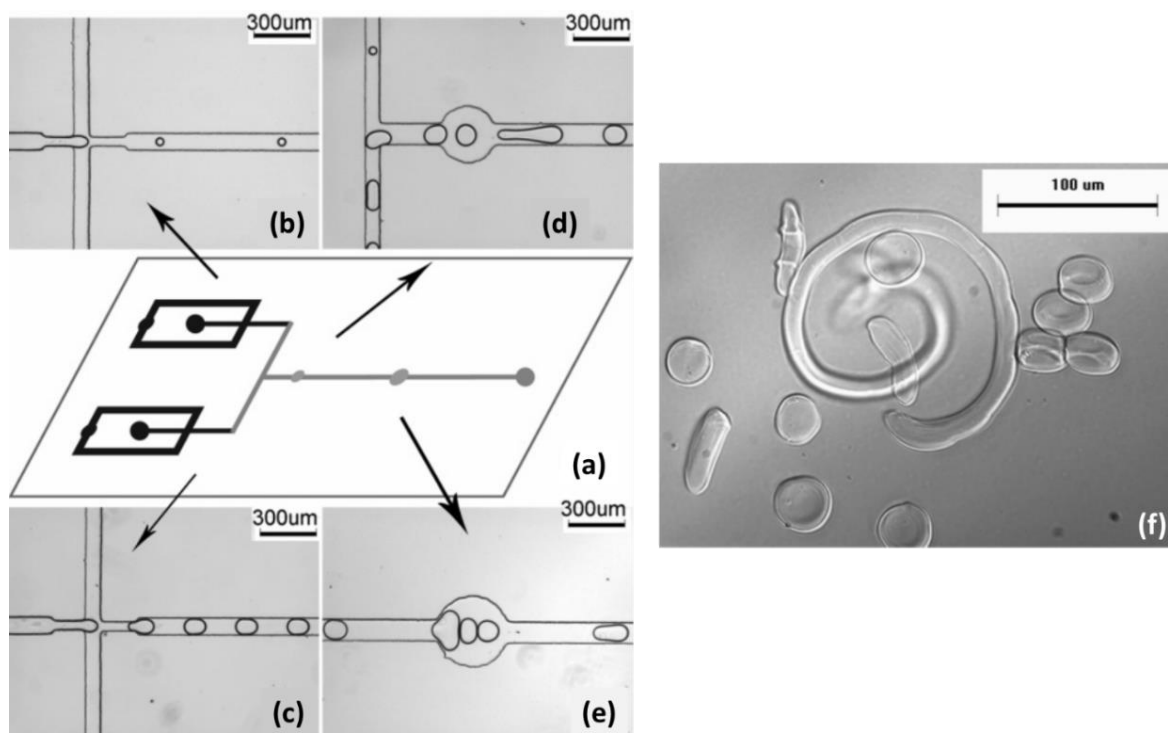


Figure 11. (a) Schematic diagram of the PDMS-based microfluidic device. (b) Flow-focusing channel to generate alginate droplets. (c) Flow-focusing channel to generate CaCl_2 droplets. (d) T-junction followed by a first circular expansion chamber. (e) A second circular expansion chamber. (f) Ca-alginate hydrogel microparticles of different shapes and sizes. Figure reprinted with permission from Reference (Liu et al., 2006). Copyright 2006 American Chemical Society.

Droplets could also be coalesced by exploiting physicochemical parameters between the continuous fluid and the dispersed fluid. (Trivedi et al., 2010; Trivedi et al., 2009) As shown in Figure 12 (a), droplets of Na-alginate (1 wt%) containing cells were generated upstream in a highly viscous silicone oil (10 centistoke) by flow-focusing without surfactant. An aqueous solution of BaCl_2 (50 mM) was injected downstream by a T-junction. With the help of dye, observations at the T-junction indicated that BaCl_2 fluid merged spontaneously with Na-alginate/cells droplets (Figure 12 (b)), instead of forming independent BaCl_2 droplets. However, this strategy lacks flexibility. The expected coalescence happens only when appropriate solvents are used. For instance, when using low-viscosity and low-interfacial energy γ_{CD} soybean oil, independent droplets of BaCl_2 were observed (Figure 12 (c)). They coalesced downstream with Na-alginate/cells droplets. This implied that successful coalescence of droplets could only take place with appropriate interfacial energy and viscosity (Trivedi et al., 2009).

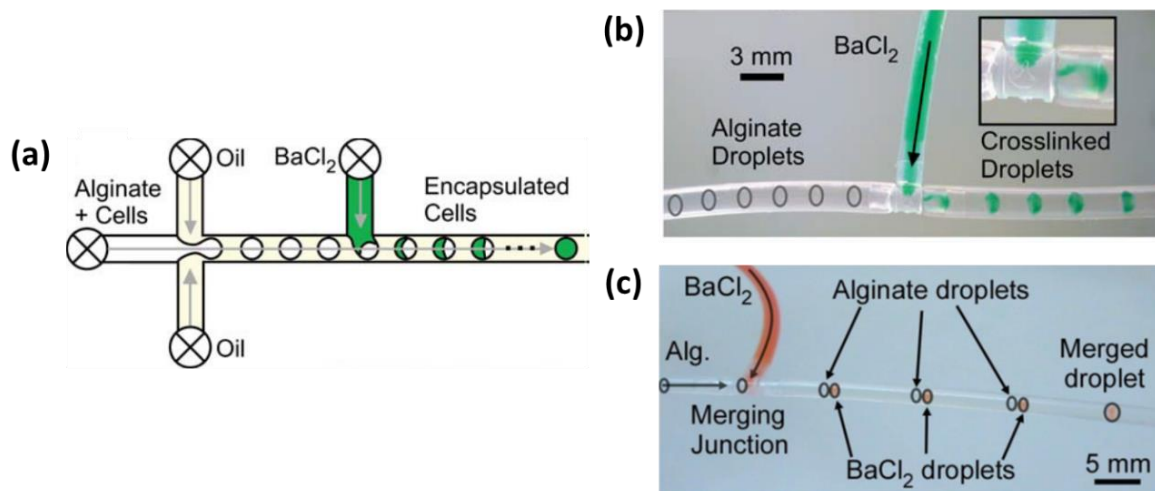


Figure 12. (a) Schematic diagram of Ba-alginate microparticle preparation for cell encapsulation in a microfluidic device assembled from fluoropolymer capillaries and junctions. Droplets observed at the T-junction when (b) highly viscous silicone oil (10 centistoke) and (c) low-viscosity soybean oil is used as the continuous fluid. Figure reprinted from Reference (Trivedi et al., 2010).

III.1.2. Water-insoluble or weakly soluble crosslinking agents

To delay gelation, Na-alginate can be mixed with water-insoluble or weakly soluble crosslinking agents, in water. This will not lead to instant gelation since there are no available cations in water. In the case of crosslinking agents which are pH-sensitive, such as calcium carbonate (CaCO_3) and calcium-ethylenediaminetetraacetic acid (Ca-EDTA) complex, an acid is used in the continuous fluid to release the cations from inert crosslinking agents. Therefore, gelation by the available cations happens after droplet generation. These strategies are summarized in

Table 3.

Table 3. Internal gelation with water-insoluble or weakly soluble crosslinking agents which are pH sensitive.

References		Zhang et al., 2007	Akbari and Pirbodaghi, 2014	Yu et al., 2019	Utech et al., 2015
Droplet generation	Concentration of Na-alginate	2 wt%	1,5 wt%	2 w/v%	2 wt%
	Continuous fluid	Soybean oil	Fluorocarbon oil	Mineral oil	Fluorinated carbon oil
	Use of surfactant	Span 80	Fluorinated surfactant	Span 80	Biocompatible surfactant
	Geometry	Flow-focusing	Flow-focusing	Flow-focusing	Flow-focusing
	Microfluidic material	PDMS chip	PDMS chip	PDMS chip	PDMS chip
Internal gelation by mixing Na-alginate and water-insoluble or weakly soluble crosslinking agents	Crosslinking agents	CaCO ₃ (0,1 wt%)	CaCO ₃ (35 mM)	CaCO ₃ (200 mM)	Ca-EDTA (50 mM)
	Mixing	Off-line, before droplet generation	Before droplet generation	Off-line, before droplet generation	Off-line, before droplet generation
	Gelation by acid addition	In continuous fluid	In continuous fluid	In collecting fluid	In continuous fluid

In the work of Zhang et al. (Zhang et al., 2007), fine particles of CaCO₃ (0.1 wt%) were dispersed in an aqueous solution of Na-alginate (2 wt%). Soybean oil with a surfactant (Span 80, 3 wt%) and containing acetic acid (5 wt%) was used as the continuous fluid (Figure 13 (a)). Droplets of Na-alginate/CaCO₃ were generated by flow-focusing in soybean oil/acetic acid (Figure 13 (b)). Droplets pH decreased because of the acetic acid in the oil. As a result, calcium cations were released from CaCO₃, causing internal gelation of the alginate. Finally, Ca-alginate hydrogel microparticles were collected in oil (Figure 13 (c)). However, when collected on a substrate, they had a “pancake” shape and were soluble in aqueous solution owing to insufficient gelation. No improvement was observed from increasing the concentration of acetic acid or that of CaCO₃. Moreover, a higher concentration of CaCO₃ particles would give rise to their aggregation in the channel (Zhang et al., 2007). The mechanical properties of the microparticles could not therefore be improved.

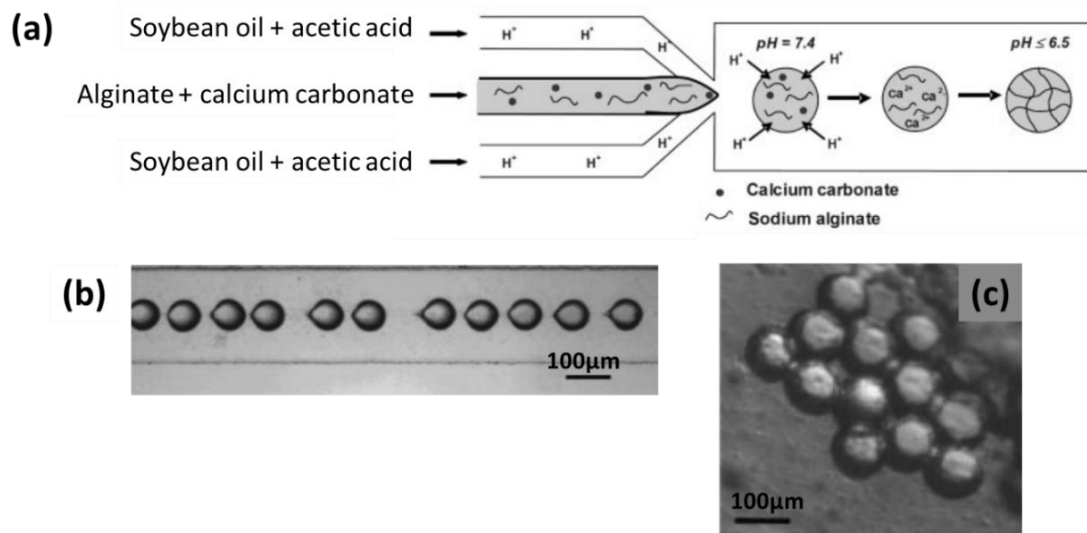


Figure 13. (a) Schematic diagram of the preparation of Ca-alginate hydrogel microparticles by using CaCO_3 to perform internal gelation of alginate in a PDMS-based microfluidic device. Micrograph of (b) droplets generated in the channel and (c) Ca-alginate hydrogel microparticles collected in oil. Figure reprinted with permission from Reference (Zhang et al., 2007)

The same principle was also applied by Akbari and Pirbodaghi to prepare cell-encapsulating microparticles (Figure 14). (Akbari and Pirbodaghi, 2014) At a first T-junction, a fluid of Na-alginate (1.5 wt%) containing cells flowed into the middle channel (Figure 14 (a)), while the Na-alginate fluid (1.5 wt%) containing CaCO_3 nanoparticles (35 mM) was introduced by two side channels (Figure 14 (b)). This geometry was used to create a coaxial stream while avoiding direct mechanical contact between cells and the potentially damaging CaCO_3 particles. At a second T-junction, fluorocarbon oil with surfactant (fluorinated surfactant, 1 wt%) was injected. Droplets of Na-alginate/cells/ CaCO_3 were then generated by flow-focusing. After droplet collection, acetic acid (0.1 vol%) dissolved in oil was added to release calcium cations within droplets, causing gelation of alginate. Droplets were thus transformed into Ca-alginate hydrogel microparticles, some with cells encapsulated (Figure 14 (c)). However, the mixture of CaCO_3 and Na-alginate was not homogeneous, which can be seen from Figure 14 (b). Thus, the varying amounts of CaCO_3 influenced the degree of gelation in each droplet, yielding Ca-alginate hydrogel microparticles with different mechanical properties. This issue is not discussed by Akbari and Pirbodaghi. (Akbari and Pirbodaghi, 2014) Furthermore, not all microparticles encapsulated cells, for reasons not explored in the article. Sorting is therefore required after the preparation of microparticles, which complicates the procedure.

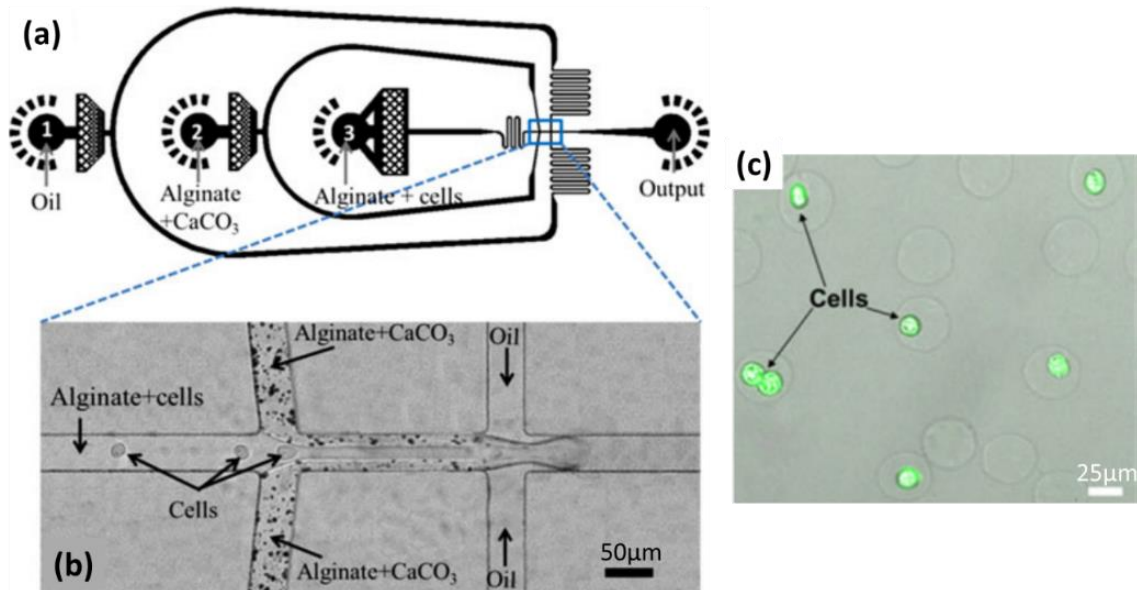


Figure 14. (a) Schematic diagram of a PDMS-based microfluidic device for the generation of droplets. (b) Micrograph of the two cross-junctions in the microfluidic device. (c) Confocal microscopic image of Ca-alginate hydrogel microparticles, some with cells encapsulated (Green fluorescence represents live cells stained by calcein AM). Figure reprinted with permission from Reference (Akbari and Pirbodaghi, 2014)

Combining the strategy of Zhang et al. and Akbari and Pirbodaghi to conduct gelation both in the microfluidic device and in the collection bath, Yu et al. (Yu et al., 2019) produced Ca-alginate hydrogel microparticles for protein encapsulation. First, from inlet 4 (Figure 15 (a)), an aqueous solution of antigen or protein was injected. It co-flowed with another aqueous solution of alginate (2 w/v%) mixed with CaCO_3 particles (200 mM) and injected from inlet 3. Mineral oil with Span 80 added was injected from inlet 2 as a continuous fluid. In the flow-focusing channel, droplets containing alginate, CaCO_3 and protein were formed. From inlet 1, another continuous fluid, mineral oil containing Span 80 and acetate acid, was introduced. When the acetate acid diffused into droplets, calcium cations were released. The alginate was then crosslinked, leading to preliminary gelation. The droplets were collected in an aqueous solution of CaCl_2 (0.27 M) to enhance gelation. In the end, spherical hydrogel microparticles were formed, with protein encapsulated (Figure 15 (b-c)). According to the authors, the preliminary gelation in the microchannel prevented the deformation that occurs when droplets are collected directly in an aqueous solution of CaCl_2 . As mentioned above, however, since CaCO_3 is not soluble in water, a high concentration of CaCO_3 will clog the microchannel. Thus, the scope for preliminary gelation is limited. Moreover, it takes time (in this case, overnight) to obtain a mixture where CaCO_3 particles are well dispersed.

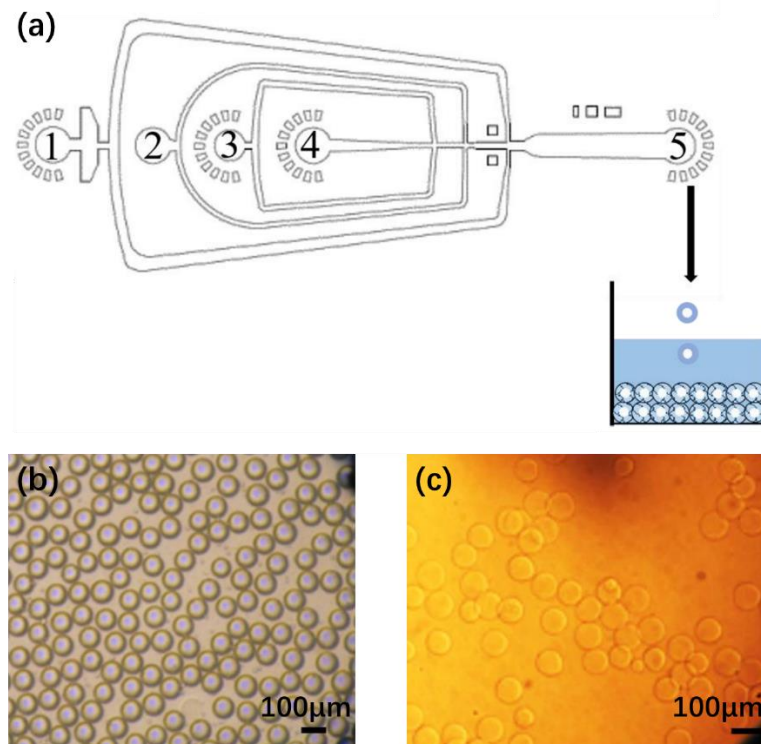


Figure 15. (a) Schematic diagram of a PDMS-based microfluidic device for the preparation of antigen-core alginate-shell microparticles. Inlet 1: Mineral oil with 3 wt% Span 80 and 0.2 vol% acetic acid; Inlet 2: Mineral oil with 3 wt% Span 80; Inlet 3: 2 w/v% alginate solution containing 200 mM CaCO_3 ; Inlet 4: an antigen or protein aqueous solution. Micrographs of Ca-alginate hydrogel microparticles in (b) oil and (c) water. Figure reprinted with permission from Reference (Yu et al., 2019)

In order to obtain a homogeneous internal structure of hydrogel microparticles, Utech et al. used a slightly water-soluble calcium-ethylenediaminetetraacetic acid (Ca-EDTA) complex as the crosslinking agent (Utech et al., 2015). An aqueous solution of Na-alginate (2 wt%) mixed with Ca-EDTA (50 mM) was first prepared. This homogeneous mixture was used as the dispersed fluid for the microfluidic system. The continuous fluid was a fluorinated carbon oil with a biocompatible surfactant (1 wt%) containing acetic acid (0.05 vol%). Droplets of Na-alginate/Ca-EDTA were generated in oil/acetic acid by flow-focusing (Figure 16 (a)). Due to the use of acetic acid, calcium cations were released from Ca-EDTA in each droplet (Figure 16 (b)), causing internal gelation of the alginate. The Ca-alginate hydrogel microparticles formed (Figure 16 (c)) had a homogeneous internal structure and were stable in an aqueous medium without dissolution. It should be noted that, the solubility of Ca-EDTA in water being low (0.26 M at 20°C), the concentration of Ca-EDTA in the Na-alginate solution was limited in order to keep the solution homogeneous. Thus, this strategy is not appropriate when microparticles need to be highly crosslinked. Furthermore, care should be taken with Ca-EDTA, as EDTA is used to dissolve alginate hydrogel microparticles in the literature. (Liu et al., 2019; Yu et al., 2019)

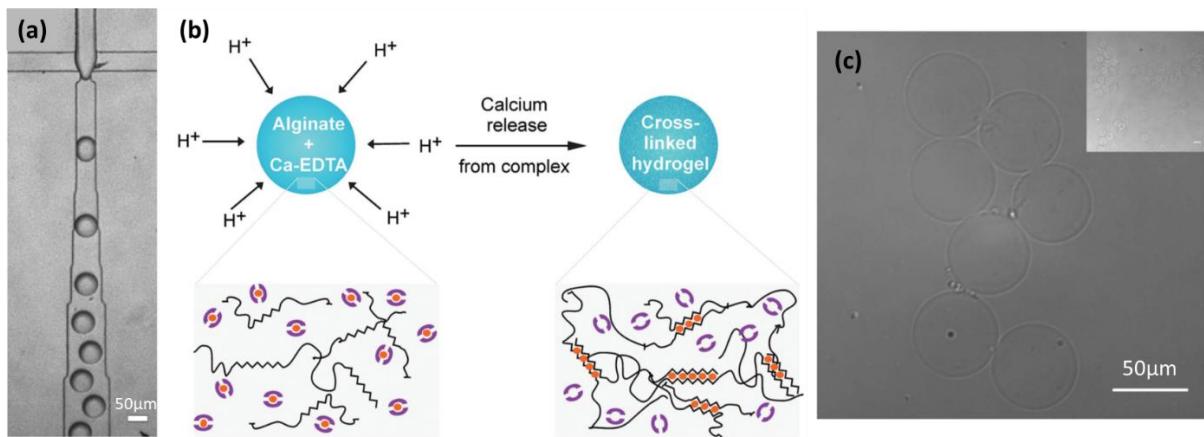


Figure 16. (a) Micrograph of the T-junction in a microfluidic device, where droplets of Na-alginate/Ca-EDTA were generated in oil/acetic acid. (b) Schematic illustration of the crosslinking process in each droplet. (c) Micrograph of Ca-alginate hydrogel microparticles in an aqueous medium. Figure reprinted with permission from Reference (Utech et al., 2015)

In conclusion, internal gelation of alginate can be realized by using crosslinking agents that are soluble or insoluble/slightly soluble in water. When water-soluble crosslinking agents are used, the instant gelation disturbs droplet generation. The problem can be solved by using partially miscible fluids with limited mixing prior to droplet generation, and/or by using extremely diluted solutions and surfactant (Table 1). Mixing crosslinking agents and Na-alginate after droplet generation involves merging droplets or flows of Na-alginate and water-soluble crosslinking agents (Table 2). The resulting droplets are dependent on physicochemical properties like viscosity and interfacial energy. If water-insoluble/slightly soluble crosslinking agents are used, they are mixed with alginate before droplet generation. For pH-sensitive crosslinking agents, acid is then needed to release cations, after which internal gelation takes place (

Table 3). A homogeneous microparticle internal structure can be achieved by choosing appropriate crosslinking agents. However, because of low solubility in water, it is important to limit the concentration of crosslinking agents to avoid precipitates in the channel.

III.2. External gelation

In external gelation, crosslinking agents come from outside the alginate droplets and are diffused into the alginate droplets or the microparticles formed, inducing crosslinking. Unlike internal gelation, in which crosslinking agents are always introduced “on-chip” (in the microfluidic device), in external gelation, crosslinking agents can be introduced both “on-chip” and/or “off-chip” (outside the microfluidic device).

III.2.1. On-chip introduction of crosslinking agents

For external gelation, several authors introduced crosslinking agents “on-chip”. They used calcium acetate ($\text{Ca}(\text{CH}_3\text{COO})_2$) or CaCl_2 as crosslinking agents, as summarized in

Table 4.

Table 4. External gelation with on-chip introduction of crosslinking agents

Reference		Zhang et al., 2007	Liu et al., 2019	Sugaya et al., 2011	Pittermannová et al., 2016
Droplet generation	Concentration of Na-alginate	2 wt%	3 wt%	0.025-0.15 wt%	1 w/v%
	Continuous fluid	Soybean oil	Corn oil	Methyl acetate	1-undecanol
	Use of surfactant	Span 80	SY-Glyster CRS-75	Not mentioned	Abil Em 90
	Geometry	Flow-focusing	Flow-focusing	Flow-focusing	Flow-focusing
	Microfluidic material	PDMS chip	Glass chip	PDMS chip	PDMS chip
External gelation by introducing Na-alginate and crosslinking agents on chip before collection	Crosslinking agents	Ca(CH ₃ COO) ₂ (2 wt%) in the continuous fluid	Emulsion of CaCl ₂ in the continuous fluid	CaCl ₂ (1 M) in the dispersed fluid	Emulsion of CaCl ₂ (2 wt%) droplets in the continuous fluid
	Geometry	-	Flow-focusing	Cross-flowing	Flow-focusing
	Mixing	During droplet generation	After droplet generation	After droplet generation	After droplet generation

Crosslinking agents can be contained in the continuous fluid, as described in Zhang et al. (Zhang et al., 2007) Ca(CH₃COO)₂ (2 wt%) was dissolved in soybean oil, the continuous fluid. In the microfluidic device detailed previously (III.1.2.), Na-alginate (2 wt%) droplets were generated by flow-focusing (Figure 17 (a)) in oil/Ca(CH₃COO)₂, with surfactant (Span 80, 3 wt%). Ca(CH₃COO)₂ diffused and dissolved in Na-alginate droplets along the channel (Figure 17 (b)), causing external gelation on-chip. Finally, Ca-alginate hydrogel microparticles were collected in oil (Figure 17 (c)). They showed better stability in an aqueous medium and had a higher Young's modulus compared with those produced by internal gelation (III.1.2.). Consequently, stronger gelation was achieved by external gelation. However, increasing the concentration of Ca(CH₃COO)₂ in soybean oil caused clogging in the microchannel. (Zhang et al., 2007) Thus, it is difficult to vary the rate of gelation of microparticles.

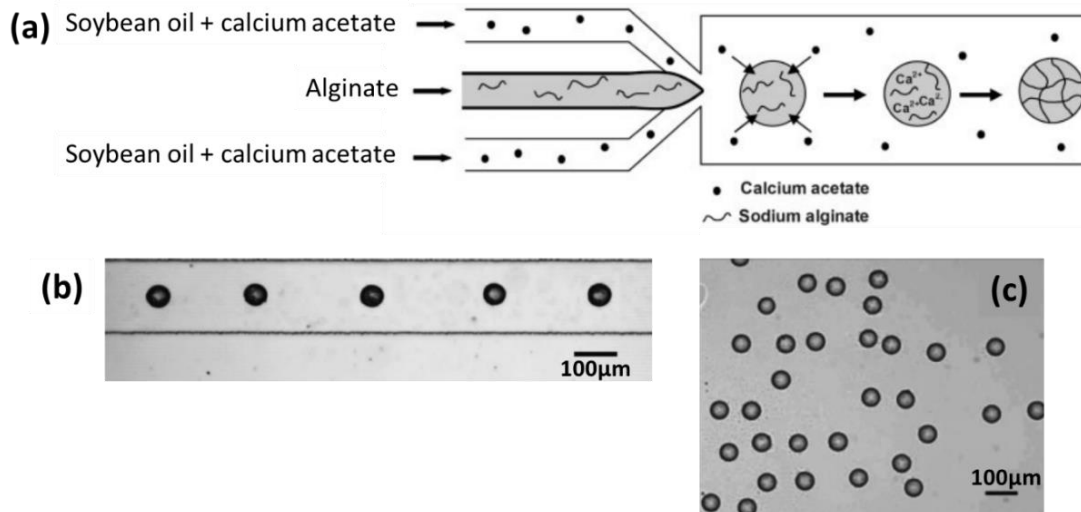


Figure 17. (a) Schematic diagram of the preparation of Ca-alginate hydrogel microparticles via on-chip external gelation in a PDMS-based microfluidic device. Micrographs of Ca-alginate hydrogel microparticles (b) in the downstream channel and (c) in the collecting container in soybean oil. Figure reprinted with permission from Reference (Zhang et al., 2007)

One way to limit channel clogging is to make the crosslinking agents diffuse slowly in Na-alginate droplets. Thus, Liu et al. used emulsion fluids to introduce crosslinking agents. (Liu et al., 2019) A glass-based microfluidic device was used, with channels modified so as to be hydrophobic. Droplets of Na-alginate (3 wt%) were first generated in corn oil at the first flow-focusing channel (Figure 18 (a)). The emulsion of CaCl_2 , containing CaCl_2 droplets in corn oil (with surfactant SY-Glyster CRS-75), was injected downstream. The contact between CaCl_2 and Na-alginate droplets caused ionic crosslinking, leading to gelation. Ca-alginate hydrogel microparticles were obtained. However, it was found that the microparticles could easily be deformed (Figure 18 (b-A, b-C)) by several parameters, such as the mass fraction of the aqueous CaCl_2 solution in emulsion (W). Deformation occurred when the value of W was too high or too low, so that an optimal value of W was required for homogeneous spherical microparticles (Figure 18 (b-B)). The morphology and homogeneity of microparticles also varied with flow rates and surfactant concentrations. Lacking flexibility, this strategy is thus not appropriate for producing spherical hydrogel microparticles. Moreover, generating small particles requires reducing the channel size, involving a risk of droplet coalescence before gelation in CaCl_2 emulsion.

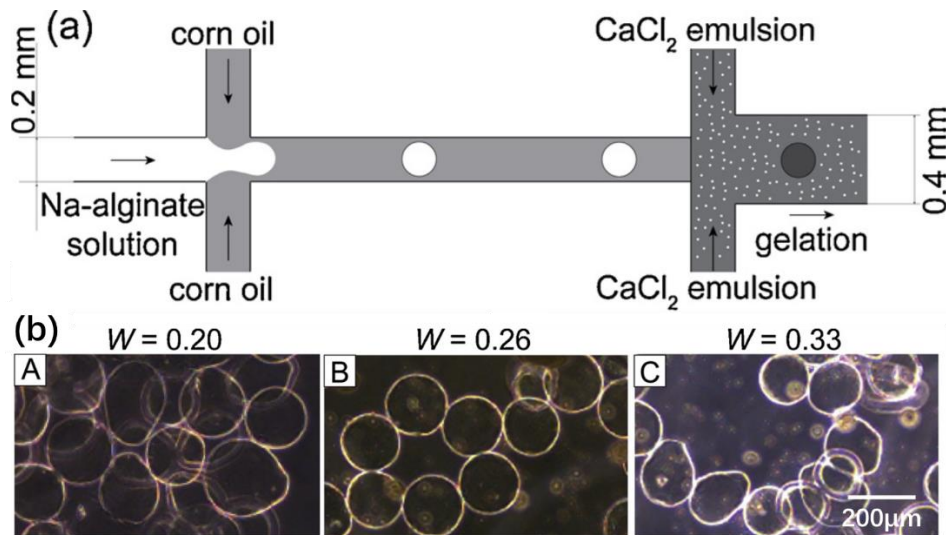


Figure 18. (a) Schematic diagram of a glass-based microfluidic device for the preparation of Ca-alginate hydrogel microparticles, with channels modified so as to be hydrophobic. (b) Micrographs of hydrogel microparticles obtained with different mass fractions of the aqueous CaCl_2 solution in emulsion (W). Figure reprinted with permission from Reference (Liu et al., 2019).

To avoid reducing the channel size, partially miscible fluids can be used as the dispersed and continuous fluids. Sugaya et al. used methyl acetate as the continuous fluid (Sugaya et al., 2011). Na-alginate (0.025-0.15 wt%) droplets were generated in methyl acetate (no mention of surfactant usage) by flow-focusing (Figure 19 (a)). In the following channel, because of the solubility of water in methyl acetate (8 wt%), water dissolved gradually from the droplets into methyl acetate. Thus, the droplets shrank and became more concentrated downstream. CaCl_2 solution (1M) was then injected by side channels and flowed with the droplets by co-flow. Calcium cations diffused into the droplets, inducing on-chip external gelation of alginate. Finally, spherical Ca-alginate hydrogel microparticles with a diameter of less than $20\mu\text{m}$ were obtained (Figure 19 (b)). In this strategy, after CaCl_2 fluids were introduced, two competing processes occurred simultaneously in each droplet: gelation and shrinkage of droplets. The competition between gelation and shrinkage is not discussed in this article. However, the results indicate that extremely small droplets tend to approach the channel wall after shrinkage (Figure 19 (b)). With the CaCl_2 fluid, after gelation, Ca-alginate hydrogel microparticles adhere to the channel wall.

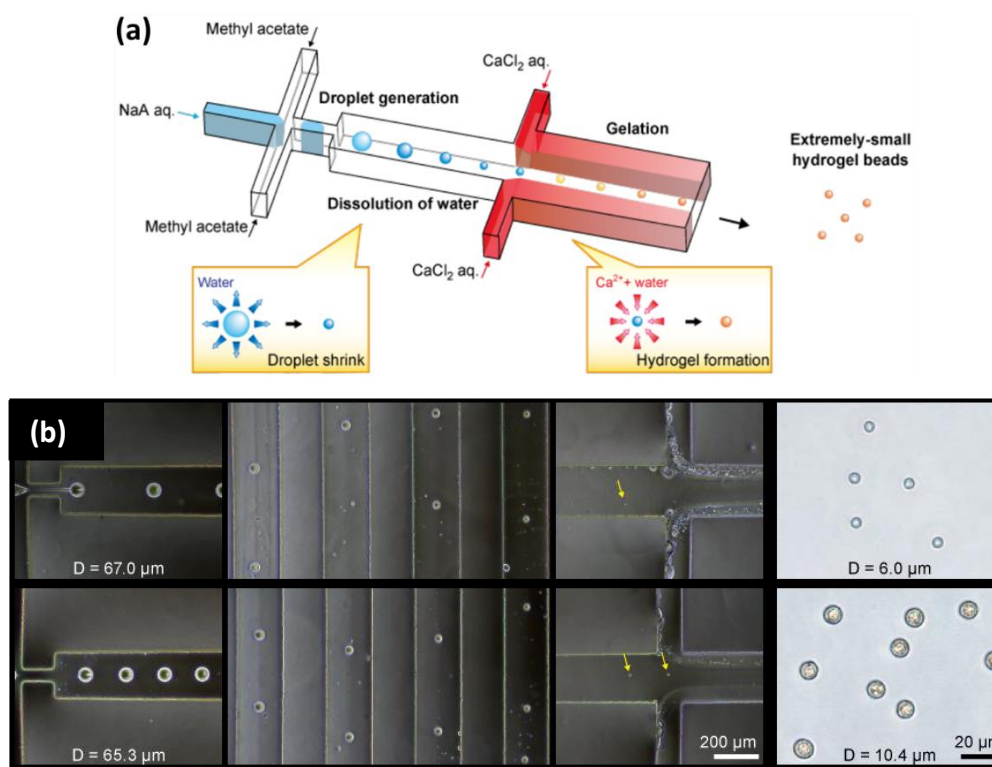


Figure 19. (a) Schematic diagram of a PDMS-based microfluidic device for the preparation of Ca-alginate hydrogel microparticles using methyl acetate as the continuous fluid, in which water is partially soluble. (b) Micrograph of Ca-alginate hydrogel microparticles collected in water, Na-alginate concentrations were 0.025 wt% (top) and 0.15 wt% (bottom), respectively. Figure reprinted from Reference (Sugaya et al., 2011)

Adhesion to the channel and coalescence of microparticles can be avoided thanks to progressive addition of the crosslinking agent. Pittermannová et al. used as continuous fluid 1-undecanol, whose water-solubility is 2.7 vol%. (Pittermannová et al., 2016) The experiment was carried out in a PDMS-based microfluidic device (Figure 20 (a)). An aqueous alginate solution (1 wt%) was first injected. After 1-undecanol (with 5 wt% surfactant Abil Em 90), shown as “oil” in Figure 20 (a), was injected into the flow-focusing channel, droplets of alginate were formed. CaCl_2 (2 wt%) was dispersed in another fluid, 1-undecanol with 5 wt% surfactant Abil Em 90, yielding an emulsion. This emulsion was injected after droplet generation, through successive channels (Figure 20 (a)). Hence, the droplets were increasingly separated from each other, avoiding coalescence. Moreover, they were surrounded by more and more CaCl_2 , increasing gelation, and by more and more 1-undecanol, increasing diffusion of water. Thus, gelation and shrinkage of droplets occurred gradually and simultaneously. According to the authors, this procedure avoids the droplet generation instability caused by pre-gelation. However, spherical hydrogel microparticles (Figure 20 (b)) were only obtained using certain flow rates and calcium concentrations. Otherwise, the microparticles were slightly deformed (Figure 20 (c)) or collapsed (Figure 20 (d)), which was explained using a core-shell model. (Pittermannová et al., 2016) Unfortunately, this explanation does not take into account the change in alginate concentration in the droplets due to water extraction, a factor which is bound to impact deformation. Simply prolonging the water extraction process before

introducing crosslinking agents, as done by Sugaya et al., could avoid deformation. (Sugaya et al., 2011)

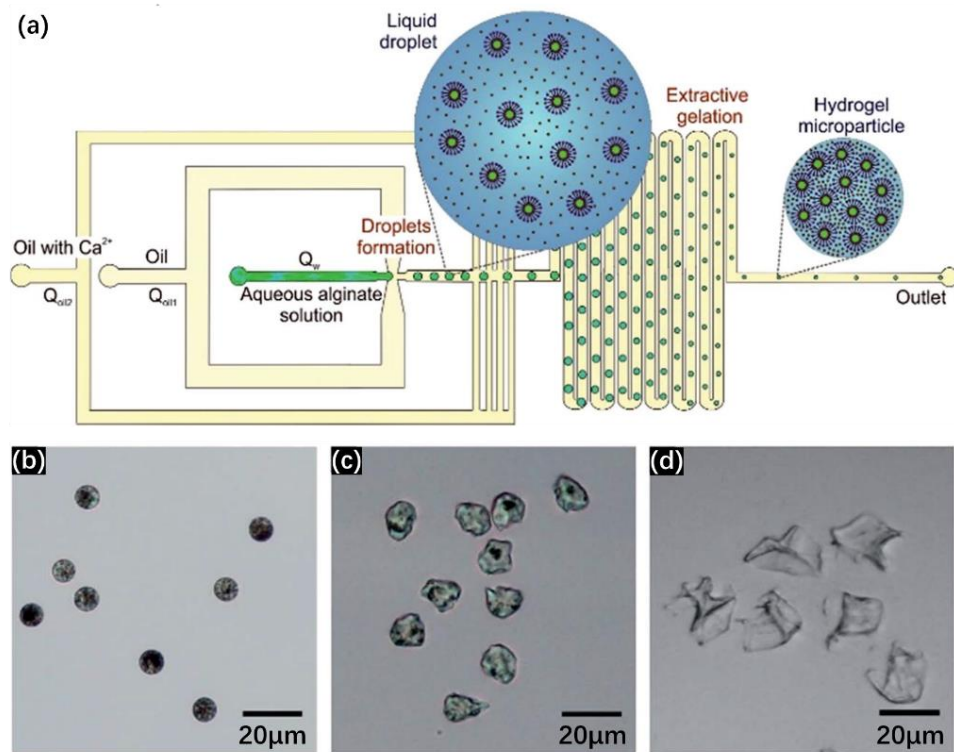


Figure 20. (a) Schematic diagram of a PDMS-based microfluidic device for the preparation of Ca-alginate hydrogel microparticles. Micrographs of microparticles with a (b) spherical, (c) slightly deformed and (d) collapsed morphology obtained using different experimental flow rates and calcium concentrations. Figure reprinted from Reference (Pittermannová et al., 2016).

III.2.2. Off-chip introduction of crosslinking agents

Other strategies of external gelation introduce crosslinking agents “off-chip”, i.e., during droplet collection. $\text{Ca}(\text{CH}_3\text{COO})_2$ or CaCl_2 are used as crosslinking agents and the collection bath procedure depends on the strategy, as summarized in

Table 5.

Table 5. External gelation with off-chip introduction of crosslinking agents

References		Hu et al., 2012		Zhang et al., 2020	This paper	
Droplet generation	Na-alginate	1.5 wt%		0.006-1 wt%	0.006-1 wt%	
	Continuous fluid	N-decanol		DMC*	DMC*	
	Use of surfactant	Span 80		NO	NO	
	Geometry	Flow-focusing		Cross-flowing	Cross-flowing	
	Microfluidic material	Glass chip		Fluoropolymer tubing and junctions	Fluoropolymer tubing and junctions	
External gelation by introducing Na-alginate and crosslinking agents off-chip during collection	Number of phases or steps	2 phases - 1 collection bath		1 phase - 1 collection bath	2 phases - 2 successive collection baths	
	Description of phases or steps	Phase 1	CaCl ₂ (15 wt%) in n-decanol	CaCl ₂ (0.1-1 wt%)	Step 1	DMC and evaporation of DMC
		Phase 2	Ba(CH ₃ COO) ₂ (15 wt%) in water and glycerol		Step 2	CaCl ₂ (0.5-10 wt%) in water
	Surfactant	Span 80		NO	NO	

* Continuous and dispersed fluids partially miscible

Hu et al. (Hu et al., 2012) studied the influence of external gelation conditions on the shape of microparticles. Na-alginate (1.5 wt%) droplets were first generated in n-decanol with surfactant (Span 80, 5 wt%), using concentric glass capillaries in co-flow geometry (Figure 21 (A)). For off-chip external gelation, droplets were collected in a two-phase gelation bath: the upper phase of n-decanol with surfactant (Span 80, 5 wt%) containing CaCl₂ (15 wt%) allowed for pre-gelation of alginate; the bottom phase, an aqueous solution of barium acetate (15 wt%), strengthened the gelation. Glycerol (0-70 wt%) was added to the bottom phase to regulate viscosity. Ca-alginate hydrogel particles of different shapes (Figure 21 (B)) were obtained by varying gelation conditions such as the interfacial energy γ_{CD} , the concentration and type of surfactant, the height h between the end of the capillary and the surface of the gelation bath, and the viscosity of the bottom phase in the gelation bath. The shape of microparticles was shown to depend on forces applied to the surface of droplets when they passed through the interface in the gelation bath. The force from γ_{CD} maintains the spherical

with the loss of water, the alginate concentration in droplets increased. Then, the channel outlet (point B in Figure 22 (a)) was immersed in an aqueous solution of CaCl_2 (0.1-1 wt%). An interface was created at the channel outlet (Figure 22 (b)) because of the non-total miscibility between DMC and water. Na-alginate droplets passed through the interface and entered the CaCl_2 solution, leading to off-chip external gelation. After gelation, Ca-alginate hydrogel microparticles were droplet-shaped (Figure 22 (c)) and tadpole-shaped (Figure 22 (d)), as in Figure 21 (B, b-c). The shape of the microparticles varied with the flow rates, the concentration of Na-alginate and that of CaCl_2 . It is likely that the deformation mechanism involved the forces applied to droplets at the interface, as explained by Hu et al. (Hu et al., 2012)

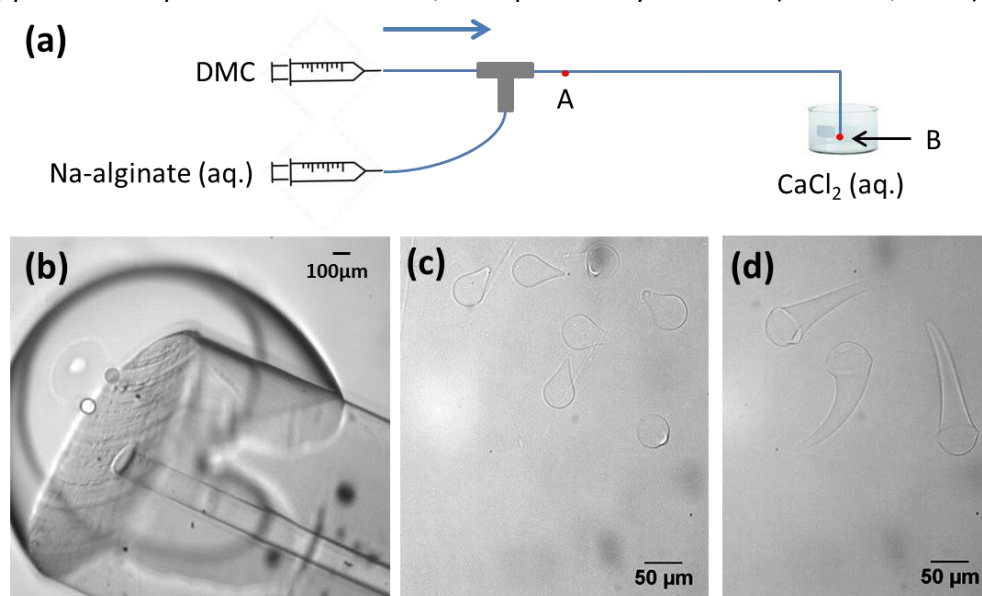


Figure 22. (a) Schematic diagram of the preparation of Ca-alginate hydrogel microparticles by off-chip external gelation without pre-gelation. Droplets were generated using a microfluidic device assembled from fluoropolymer capillaries and a T-junction. Micrographs of (b) the channel outlet immersed in an aqueous solution of CaCl_2 ; Ca-alginate hydrogel microparticles prepared by collecting droplets in an aqueous solution of CaCl_2 at concentrations of (c) 1 wt% and (d) 0.1 wt%.

To improve the spherical shape of microparticles, we collected droplets in a bath of the continuous fluid, i.e., DMC. Hence, the droplets continued to shrink and were finally transformed into spherical condensed Na-alginate microparticles, not yet gelated. For the gelation of the microparticles, the bath of DMC was first evaporated. Then, an aqueous solution of CaCl_2 (0.5-10 wt%) was added to the dried Na-alginate microparticles, inducing off-chip external gelation. Observations showed that this process was accompanied by the swelling of the microparticles without deformation (Figure 23 (b-c)). In the end, spherical Ca-alginate hydrogel microparticles were obtained. They were insoluble in water, indicating efficient gelation. Moreover, the concentration of CaCl_2 had no significant effect on the size of the Ca-alginate microparticles. Since no surfactant is used in this method, no surfactant-removing step is needed, which simplifies the process. However, the quantity of microparticles produced is limited by the need to avoid droplet coalescence.

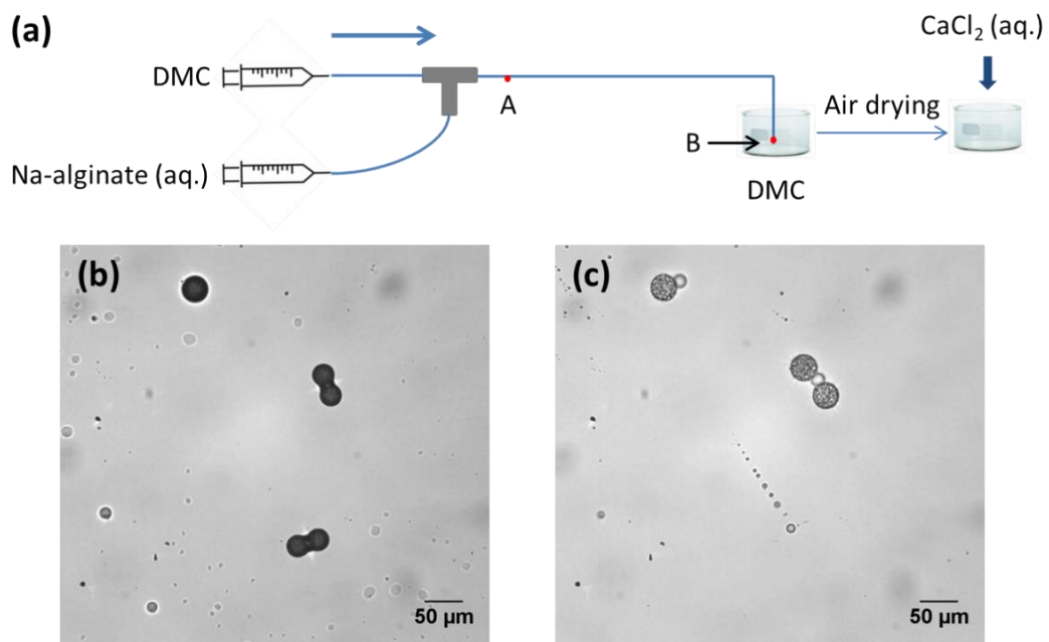


Figure 23. (a) Schematic diagram of a two-step preparation of Ca-alginate hydrogel microparticles using a microfluidic device constructed from fluoropolymer capillaries and junctions. Micrographs of (b) dried Na-alginate microparticles in air and (c) corresponding Ca-alginate microparticles after gelation in CaCl₂ solution. Figure reprinted and adapted with permission from Reference (Zhang et al., 2020)

In conclusion, external gelation of alginate can be performed both on-chip and off-chip. For on-chip external gelation (

Table 4), crosslinking agents can be added to the continuous fluid, i.e., the oil. However, only limited concentrations can be used, since most are slightly soluble in oil. Therefore, introducing crosslinking agents in emulsion form increases the quantity of alginate available for gelation. However, the particles are large and deformed. On the other hand, if partially miscible fluids are used, an aqueous solution of crosslinking agents can be injected after droplet shrinkage. Particle size is reduced but the gelation is too rapid. Things can be improved by dissolving crosslinking agents in an oil-based emulsion, introduced in small quantities but repeatedly.

For off-chip external gelation (

Table 5), crosslinking agents are introduced into the collection bath. A two-phase collection bath permits pre-gelation of Na-alginate droplets before gelation. However, the particles are large. Droplet size can be reduced to below the channel diameter by using partially miscible fluids for droplet generation, and the droplets can then be collected directly in the dispersed phase containing the crosslinking agent. However, the microparticles are deformed. To further reduce particle size and improve gelation, our solution is to perform two-step collection. Thus, off-chip external gelation can be used to produce shape-controlled and size-controlled microparticles.

IV. Properties of alginate hydrogel microparticles

After preparation, alginate hydrogel microparticles should be characterized to obtain better knowledge of their properties, which will determine their further applications. This section discusses characterization approaches and factors influencing particle properties.

IV.1. Size

Size is one of the most important properties of alginate hydrogel microparticles. For example, in drug delivery, microparticle size and size distribution affect drug release kinetics. (Uyen et al., 2020) Size can be measured by optical or light-scattering (sub-micrometer range) microscopy (Joye and McClements, 2014), or using microgrippers. (Zhang, 2020)

Droplet-based microfluidics allows monodisperse microparticles to be produced with accurate control of size and size distribution. Table 6 shows the average size attained under droplet-based microfluidics using different gelation methods.

Table 6. Average size of alginate microparticles prepared using droplet-based microfluidics with different gelation methods.

Average size of microparticles	Gelation method	Reference
1-50 μm 10-300 nm	Internal gelation with water-soluble crosslinking agents mixed with Na-alginate before droplet generation	Rondeau and Cooper-White 2008*
50-300 μm	Internal gelation with water-soluble crosslinking agents mixed with Na-alginate after droplet generation	Xu et al. 2008
20-50 μm		Liu et al. 2006
22-42 μm		Trivedi et al. 2009
60-100 μm	Internal gelation with water-insoluble crosslinking agents mixed with Na-alginate after droplet generation	Zhang et al. 2007
26 μm		Akbari and Pirbodaghi 2014
50-100 μm		Yu et al. 2019
10-50 μm	Internal gelation with slightly water-soluble crosslinking agents mixed with Na-alginate after droplet generation	Utech et al. 2015
50-70 μm	External gelation with on-chip introduction of crosslinking agents	Zhang et al. 2007
147-176 μm		Liu et al. 2019
5-10 μm		Pittermannová et al. 2016*
6-10 μm		Sugaya et al. 2011*

100-200 μm	External gelation with off-chip introduction of crosslinking agents	Hu et al. 2012
7-40 μm		Zhang et al. 2020*

* Partially miscible fluids were used.

Figure 24 shows an example of the narrow size distribution of Na-alginate and Ca-alginate microparticles produced by droplet-based microfluidics (Zhang et al., 2020) (part III.2.2.), indicating the monodispersity of the particle size. This is an advantage compared to conventional emulsification, which yields a broader size distribution. (Xu et al., 2009)

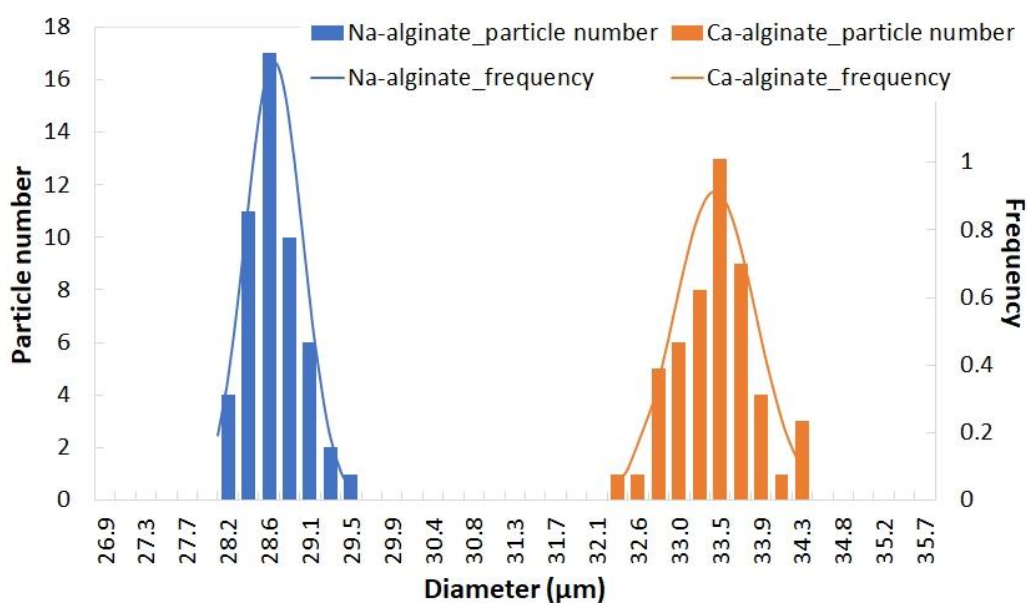


Figure 24. Size distribution of microparticles of Na-alginate (blue) and Ca-alginate (orange) produced using droplet-based microfluidics. The curves show Gaussian fitting. Figure reprinted and adapted with permission from Reference (Zhang et al., 2020)

With droplet-based microfluidics, the size of alginate hydrogel microparticles is influenced by several factors linked to the fluids used to generate them. When immiscible fluids are used for the dispersed and the continuous fluids, the size of alginate hydrogel microparticles is completely dependent on the size of the droplets first generated. Droplet size is influenced by channel size, and smaller droplets can be generated by using narrower channels. Other important factors are flow rates, alginate concentration (Rondeau and Cooper-White, 2008; Zhang et al., 2020), and fluid viscosities. (Seemann et al., 2011; Teh et al., 2008) However, reducing channel size increases hydraulic resistance, as well as the pressure required to generate droplets. Moreover, it should be noted that in most cases, the Na-alginate solution used is relatively viscous. Therefore, high pressure may cause leakage or even destruction of the microfluidic device. (Akbari and Pirbodaghi, 2014; Utech et al., 2015) Thus, even when channel diameter is decreased and/or the flow rate of the continuous fluid is increased, producing droplets of a diameter below 10 μm remains challenging.

Droplets of this size can be obtained without applying high pressure (Table 6), by using partially miscible fluids with low solubility in each other. (Rondeau and Cooper-White, 2008;

Sugaya et al., 2011; Zhang et al., 2020) The dispersed fluid is an aqueous solution containing Na-alginate; the continuous fluid is an organic solvent that is partially miscible with water and in which water has low solubility. The partial miscibility between the continuous and the dispersed fluids should be slight enough so that interfacial energy γ_{CD} still allows the generation of droplets. The low solubility of water in the continuous fluid allows water diffusion from droplets into it, causing the droplets to shrink. As a result, the initially obtained diluted large droplets are transformed into concentrated small droplets or microparticles. Thus, their size is no longer dependent on the size of droplets initially generated but varies with the interaction between water and the continuous fluid.

IV.2. Shape

The shape of microparticles is another important property. A specific shape is sometimes needed; for example, red blood cell-mimicking microparticles are often required in a biconcave shape. (Merkel et al., 2011) In drug delivery, the shape of microparticles has an impact on the drug-release profile. (Freiberg and Zhu, 2004) The overall shape of microparticles can be observed by using optical microscopy. Confocal microscopy of fluorescent samples can be used to form a spatial 3D image. (Joye and McClements, 2014) Better resolution can be obtained by using atomic force microscopy (AFM) or scanning electron microscopy (SEM). (Zhang et al., 2020)

With droplet-based microfluidics, the spherical droplets initially generated can be transformed into spherical alginate hydrogel microparticles after gelation. Non-spherical microparticles can also be obtained. For example, as presented previously, Liu et al. first generated droplets of Na-alginate and CaCl_2 separately. (Liu et al., 2006) Then the droplets were fused in a specifically designed microfluidic device, leading to gelation. By varying the channel geometry and controlling the flow rates of fluids, Ca-alginate microparticles of different shapes were obtained (Figure 11 (f)). A different method was presented by Hu et al. (Hu et al., 2012) Na-alginate droplets were first generated and then collected in a two-phase gelation bath. Spherical droplets were deformed via interfacial energy derived from surfactant and viscous force. Thus, different shapes were produced (Figure 21 (B)) by controlling the surfactant used and the viscosity of the gelation bath.

IV.3. Concentration

After preparation, the concentration of alginate in the microparticles can be calculated approximately. For instance, Zhang et al. used partially miscible fluids. (Zhang et al., 2020) An aqueous solution of Na-alginate was prepared with a known concentration. After droplet generation, droplet shrinkage occurred during passage through the channel due to water diffusion into the continuous fluid. Droplets were hence transformed into microparticles. As the diffusion of Na-alginate into the continuous fluid is negligible (Rondeau and Cooper-White, 2008), the quantity of Na-alginate is constant. It can be calculated by multiplying the droplet volume and initial concentration. Finally, by measuring microparticle size, the concentration of Na-alginate can be calculated. The final concentration of Na-alginate varies from 20 to 100 wt%, depending on the initial concentration and diameter of the droplets generated. (Zhang et al., 2020) Furthermore, as presented in Utech et al., the homogeneity of composition of microparticles can be determined with the help of fluorescence technology. (Utech et al., 2015)

IV.4. Stability

In most cases, surfactant is added in the continuous fluid (Trivedi et al., 2009; Utech et al., 2015; Zhang et al., 2007) to lower interfacial energy between the continuous and the dispersed fluids γ_{CD} . Note that for each of the above studies, use or non-use of surfactant is mentioned when indicated by the authors (Table 1-5). Surfactant facilitates the creation of a new interface, and thus the formation of droplets. It also stabilizes the formed droplets by preventing their coalescence. (Seemann et al., 2011) Before the application of microparticles, the surfactant should be dissolved (Akbari and Pirbodaghi, 2014), except for biocompatible surfactant (Utech et al., 2015), although protocols for removing surfactant are rarely reported in the literature. To remove the oil used during the preparation, microparticles should be washed several times with an aqueous solution, followed by centrifugation. (Zhang et al., 2007)

However, despite its advantages, the use of surfactant may be undesirable. Surfactant has been shown to impact the surface properties of microparticles, such as morphology (Sundberg et al., 1990) and surface hydrophobicity. (Kidane et al., 2002) Additionally, if rinsing is insufficient, the traces of surfactant on microparticles can damage the devices during application. In this situation, microparticles should be prepared without surfactant, which is possible with droplet-based microfluidics. In the microchannel, the coalescence of droplets can be avoided by enlarging the distance between droplets, which can be achieved by regulating flow rates. Furthermore, gelation, either on-chip or off-chip, solidifies droplets and thus helps to avoid coalescence as well. Another strategy consists of using partially miscible fluids. This means that the droplets shrink and become more and more condensed during passage through the channel. At the outlet, either gelation (Rondeau and Cooper-White, 2008) or a final shrinkage (Zhang et al., 2020) can help avoid coalescence.

Moreover, for their stability, alginate hydrogel microparticles should be insoluble in water. This can be achieved by adopting proper gelation methods using a sufficient quantity of crosslinking agents for effective gelation.

IV.5. Mechanical properties

Mechanical properties of alginate hydrogel microparticles are usually characterized by measuring the Young's modulus, which varies with several factors. According to the type of bond between alginate and crosslinking agents, covalent crosslinking results in a higher Young's modulus than ionic crosslinking in microparticles. (Caccavo et al., 2018) For ionic crosslinking, different cations present different affinities, i.e. different forces with alginate, thus different Young's moduli. (Mørch et al., 2006) In addition, the Young's modulus increases with the concentration of alginate. (Markert et al., 2013) To measure the Young's modulus of a microparticle, it needs to be deformed under a known force, which can be either compressive or tensile. (Guevorkian and Maître, 2017) The techniques used in the literature include micropipette aspiration (Kleinberger et al., 2013), compression (Carin et al., 2003; Wang et al., 2005) or Atomic Force Microscopy (AFM). (Zhang et al., 2020; Zhang et al., 2007)

IV.5.1. Micropipette aspiration technique

In the micropipette aspiration technique, controlled pressure is used to pull on the sample surface. When this pressure is high enough, the sample behaves like a viscoelastic fluid flowing inside the micropipette. (Guevorkian and Maître, 2017) With a known pressure applied, the Young's modulus is calculated by applying the homogenous half-space model as described by Kleinberger et al. (Kleinberger et al., 2013)

IV.5.2. Compression technique

This technique consists in compressing a microparticle between two parallel plates (Carin et al., 2003) or between the flat end of a glass fiber and a glass surface. (Wang et al., 2005) A force transducer is connected to the equipment to measure the force applied. By varying the force, microparticle deformation can be recorded. Finally, according to the force-deformation curve and equations based on theoretical models, the Young's modulus is calculated.

However, both the micropipette aspiration technique and the compression technique are unsuitable for microparticles with high resistance to deformation, like the Na- and Ca-alginate microparticles generated by Zhang et al. (Zhang et al., 2020) In this case, Atomic Force Microscopy was used to measure resistance to deformation.

IV.5.3. Atomic Force Microscopy

Measurement of the Young's modulus of a microparticle with Atomic Force Microscopy (AFM) involves indenting it. The indentation depth (order of 100 nm) is generally about 100 times less than the diameter of the microparticle (order of 10 μm). Hence the Young's modulus represents the local mechanical property on the surface, depending on the measuring point, as in Zhang et al. (Zhang et al., 2020) As the surface of their microparticles was smooth, variation in the local mechanical property was explained by the porous inner structure observed by Scanning Electron Microscopy (SEM) (Figure 25).

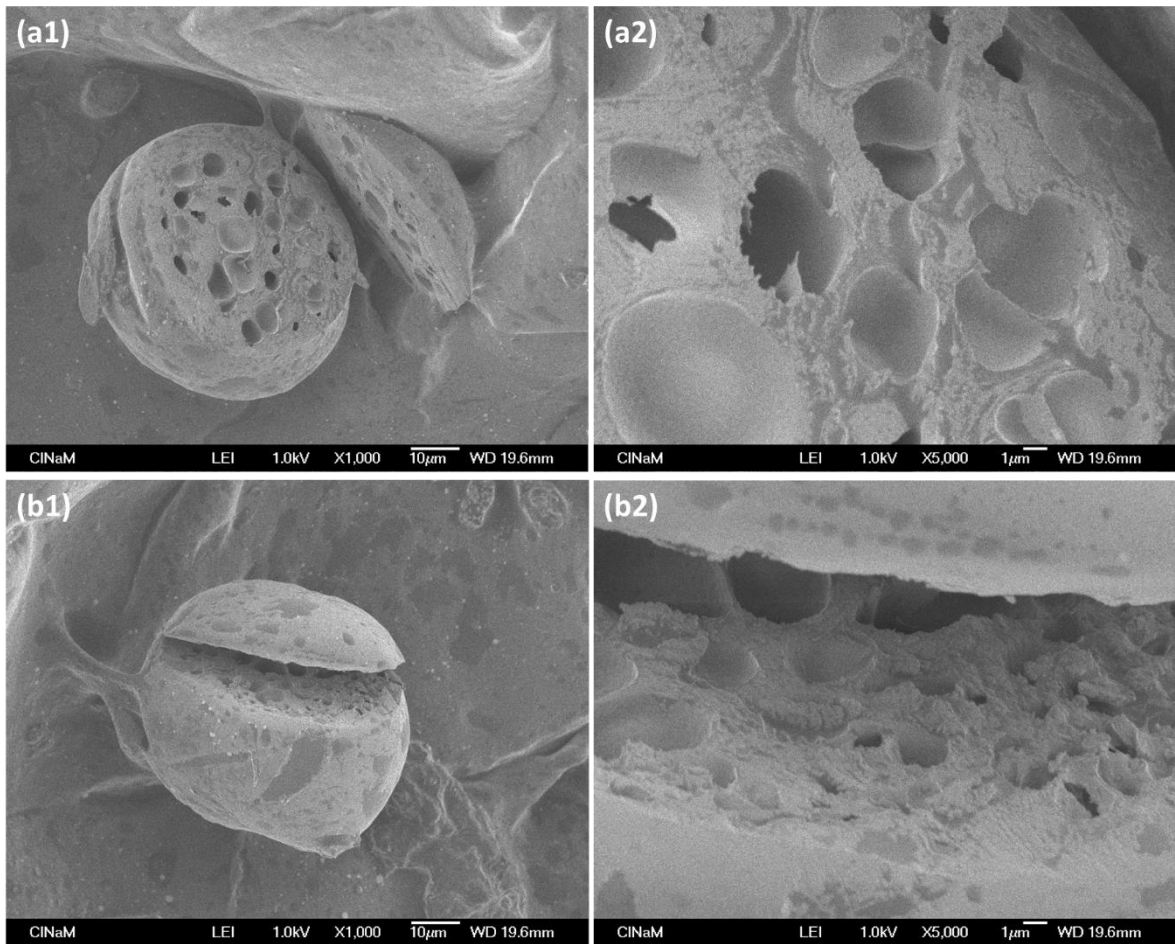


Figure 25. SEM photographs of 2 Na-alginate microparticles (a) and (b), magnified 1000x (a1 and b1) and 5000x (a2 and b2). Na-alginate microparticles were prepared following the method mentioned in the publication (Zhang et al. 2020). Figure reprinted and adapted with permission from Reference (Zhang et al., 2020)

V. Conclusion

This review focuses on the preparation of alginate hydrogel microparticles via droplet-based microfluidics. Various strategies are presented and classified within categories that represent the full range of methods used in the literature. Thus, readers will find that any strategy encountered can fit into one of the categories we present here.

To summarize, gelation is indispensable to transform alginate microdroplets into alginate hydrogel microparticles. It is realized by crosslinking, which requires crosslinking agents to be introduced either inside or outside the microdroplets, causing respectively internal gelation or external gelation. For internal gelation, crosslinking agents are introduced “on-chip” (in the microfluidic device). For external gelation, crosslinking agents can be introduced both “on-chip” and/or “off-chip” (outside the microfluidic device). The review describes the various strategies applied under the microfluidic technique, and the size, shape, concentration, stability and mechanical properties of the alginate hydrogel microparticles obtained.

Lastly, we wish to stress the ease of constructing a microfluidic device with Fluoropolymer tubing and junctions compared to a chip, whatever its composition: PDMS, PMMA or glass. A microfluidic device’s geometry can be tuned flexibly, whereas in a chip, the geometry of channels is fixed. Chip fabrication is laborious, time-consuming and expensive. (Enck et al., 2020) Clean room facilities are indispensable and the products used for the photolithography are toxic.

Thus, in terms of flexibility, cost and efficiency, we highly recommend using a microfluidic device with Fluoropolymer tubing and junctions, especially for proof-of-concept demonstrations.

VI. Outlook

There is scope for several future improvements in the preparation of alginate hydrogel microparticles via droplet-based microfluidics.

1. When preparing microparticles using droplet-based microfluidics, surfactant is usually used. Unfortunately, surfactant has been shown to affect particle morphology (Pittermannová et al., 2016). Moreover, surfactant can damage the equipment involved in subsequent applications. (Zhang et al., 2020) It obviously needs to be removed; however, the literature contains little information on how to remove surfactant. Moreover, despite its importance, the microparticle purifying process is barely touched on. (Pittermannová et al., 2016) Future work could therefore usefully provide more details of the full preparation process, including surfactant removal.
2. In practice, a large number of microparticles usually needs to be produced. However, the throughput of microparticles fabricated by microfluidics is still limited. Exploring ways to scale up microparticle production would therefore be a welcome contribution.
3. Currently, most publications concern proof-of-concepts or preliminary demonstrations. Little work is available on microparticles in real biological or biomedical applications. (Wang and Zhou, 2020) Apart from biological barriers (Poon et al., 2020), the physiological environment is quite complex in terms of composition and rheological characteristics, which makes it extremely unlikely that microparticles can provide the functions required. The feasibility of using alginate hydrogel microparticles in real-life applications needs to be assessed.

4. The relation between degree of gelation and mechanical properties is rarely discussed in the literature. In fact, the degree of gelation itself is barely mentioned although, from a microscopic point of view, it influences mechanical properties. Thus, we believe it is worth assessing degree of gelation by measuring the concentration of crosslinking agents inside the final hydrogel microparticles. Subsequently, the relation between degree of gelation and mechanical properties should also be addressed.
5. In practical applications, the surface of microparticles usually needs to be modified. For example, one way to detect antigens is to graft antibodies onto the surface of microparticles. (Roh et al., 2019) We note that the surface modification is always an extra step after the preparation of microparticles, thus complicating the process. The feasibility of an “all-in-one” process should be studied: can all the steps be performed within one single microfluidic device, combining microparticle production and surface modification?

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VIII. References

- Agüero, L., Zaldivar-Silva, D., Peña, L., Dias, M.L., 2017. Alginate microparticles as oral colon drug delivery device: A review. *Carbohydrate Polymers* 168, 32-43.
- Ahmed, E.M., 2015. Hydrogel: Preparation, characterization, and applications: A review. *Journal of Advanced Research* 6, 105-121.
- Akbari, S., Pirbodaghi, T., 2014. Microfluidic encapsulation of cells in alginate particles via an improved internal gelation approach. *Microfluidics and Nanofluidics* 16, 773-777.
- Barbieri, M., Cellini, F., Cacciotti, I., Peterson, S.D., Porfiri, M., 2017. In situ temperature sensing with fluorescent chitosan-coated PNIPAAm/alginate beads. *Journal of Materials Science* 52, 12506-12512.
- Baroud, C.N., Gallaire, F., Dangling, R., 2010. Dynamics of microfluidic droplets. *Lab on a Chip* 10, 2032-2045.
- Caccavo, D., Cascone, S., Lamberti, G., Barba, A.A., 2018. Hydrogels: experimental characterization and mathematical modelling of their mechanical and diffusive behaviour. *Chemical Society Reviews* 47, 2357-2373.
- Carin, M., Barthès-Biesel, D., Edwards-Lévy, F., Postel, C., Andrei, D.C., 2003. Compression of biocompatible liquid-filled HSA-alginate capsules: Determination of the membrane mechanical properties. *Biotechnology and Bioengineering* 82, 207-212.
- Chan, L.W., Lee, H.Y., Heng, P.W.S., 2002. Production of alginate microspheres by internal gelation using an emulsification method. *International Journal of Pharmaceutics* 242, 259-262.
- Christopher, G.F., Anna, S.L., 2007. Microfluidic methods for generating continuous droplet streams. *Journal of Physics D: Applied Physics* 40, R319-R336.
- De Menech M., Garstecki P., Jousse F., Stone H. A., 2008. Transition from squeezing to dripping in a microfluidic T-shaped junction. , 595, pp 141-161 *Journal of Fluid Mechanics* 595, 141-161.
- Draget, K.I., 2009. 29 - Alginates, in: Phillips, G.O., Williams, P.A. (Eds.), *Handbook of Hydrocolloids (Second Edition)*. Woodhead Publishing, pp. 807-828.

Enck, K., Rajan, S.P., Aleman, J., Castagno, S., Long, E., Khalil, F., Hall, A.R., Opara, E.C., 2020. Design of an Adhesive Film-Based Microfluidic Device for Alginate Hydrogel-Based Cell Encapsulation. *Annals of Biomedical Engineering* 48, 1103-1111.

Freiberg, S., Zhu, X.X., 2004. Polymer microspheres for controlled drug release. *International Journal of Pharmaceutics* 282, 1-18.

Fundueanu, G., Nastruzzi, C., Carpov, A., Desbrieres, J., Rinaudo, M., 1999. Physico-chemical characterization of Ca-alginate microparticles produced with different methods. *Biomaterials* 20, 1427-1435.

Gacesa, P., 1988. Alginates. *Carbohydrate Polymers* 8, 161-182.

Grant, G.T., Morris, E.R., Rees, D.A., Smith, P.J.C., Thom, D., 1973. Biological interactions between polysaccharides and divalent cations: The egg-box model. *FEBS Letters* 32, 195-198.

Guevorkian, K., Maître, J.L., 2017. Chapter 10 - Micropipette aspiration: A unique tool for exploring cell and tissue mechanics in vivo, in: Lecuit, T. (Ed.), *Methods in Cell Biology*. Academic Press, pp. 187-201.

Haghgooeie, R., Toner, M., Doyle, P.S., 2010. Squishy Non-Spherical Hydrogel Microparticles. *Macromolecular Rapid Communications* 31, 128-134.

Hoffman, A.S., 2012. Hydrogels for biomedical applications. *Advanced Drug Delivery Reviews* 64, 18-23.

Hu, Y., Wang, Q., Wang, J., Zhu, J., Wang, H., Yang, Y., 2012. Shape controllable microgel particles prepared by microfluidic combining external ionic crosslinking. *Biomicrofluidics* 6, 026502.

Joye, I.J., McClements, D.J., 2014. Biopolymer-based nanoparticles and microparticles: Fabrication, characterization, and application. *Current Opinion in Colloid & Interface Science* 19, 417-427.

Kidane, A., Guimond, P., Rob Ju, T.-C., Sanchez, M., Gibson, J., North, A., Hogenesch, H., Bowersock, T.L., 2002. Effects of cellulose derivatives and poly(ethylene oxide)-poly(propylene oxide) tri-block copolymers (Pluronic®surfactants) on the properties of alginate based microspheres and their interactions with phagocytic cells. *Journal of Controlled Release* 85, 181-189.

Kleinberger, R.M., Burke, N.a.D., Dalnoki-Veress, K., Stöver, H.D.H., 2013. Systematic study of alginate-based microcapsules by micropipette aspiration and confocal fluorescence microscopy. *Materials Science and Engineering: C* 33, 4295-4304.

Lee, B.-B., Ravindra, P., Chan, E.-S., 2013. Size and Shape of Calcium Alginate Beads Produced by Extrusion Dripping. *Chemical Engineering & Technology* 36, 1627-1642.

Lee, K.Y., Mooney, D.J., 2012. Alginate: Properties and biomedical applications. *Progress in Polymer Science* 37, 106-126.

Lee, K.Y., Yuk, S.H., 2007. Polymeric protein delivery systems. *Progress in Polymer Science* 32, 669-697.

Liu, K., Ding, H.-J., Liu, J., Chen, Y., Zhao, X.-Z., 2006. Shape-Controlled Production of Biodegradable Calcium Alginate Gel Microparticles Using a Novel Microfluidic Device. *Langmuir* 22, 9453-9457.

Liu, Y., Tottori, N., Nisisako, T., 2019. Microfluidic synthesis of highly spherical calcium alginate hydrogels based on external gelation using an emulsion reactant. *Sensors and Actuators B: Chemical* 283, 802-809.

Maitra, J., Shukla, V., 2014. Cross-linking in hydrogels - a review. *Am J Polym Sci* 4, 25-31.

Markert, C.D., Guo, X., Skardal, A., Wang, Z., Bharadwaj, S., Zhang, Y., Bonin, K., Guthold, M., 2013. Characterizing the micro-scale elastic modulus of hydrogels for use in regenerative medicine. *Journal of the Mechanical Behavior of Biomedical Materials* 27, 115-127.

Merkel, T.J., Jones, S.W., Herlihy, K.P., Kersey, F.R., Shields, A.R., Napier, M., Luft, J.C., Wu, H., Zamboni, W.C., Wang, A.Z., Bear, J.E., Desimone, J.M., 2011. Using mechanobiological mimicry of red blood cells to extend circulation times of hydrogel microparticles. *Proceedings of the National Academy of Sciences* 108, 586.

Mørch, Ý.A., Donati, I., Strand, B.L., Skjåk-Bræk, G., 2006. Effect of Ca²⁺, Ba²⁺, and Sr²⁺ on Alginate Microbeads. *Biomacromolecules* 7, 1471-1480.

Pittermannová, A., Ruberová, Z., Zadražil, A., Bremond, N., Bibette, J., Štěpánek, F., 2016. Microfluidic fabrication of composite hydrogel microparticles in the size range of blood cells. *RSC Advances* 6, 103532-103540.

Poon, W., Kingston, B.R., Ouyang, B., Ngo, W., Chan, W.C.W., 2020. A framework for designing delivery systems. *Nature Nanotechnology* 15, 819-829.

Qiu, C., Chen, M., Yan, H., Wu, H., 2007. Generation of Uniformly Sized Alginate Microparticles for Cell Encapsulation by Using a Soft-Lithography Approach. *Advanced Materials* 19, 1603-1607.

Ren, K., Zhou, J., Wu, H., 2013. Materials for Microfluidic Chip Fabrication. *Accounts of Chemical Research* 46, 2396-2406.

Roh, Y.H., Lee, H.J., Bong, K.W., 2019. Microfluidic Fabrication of Encoded Hydrogel Microparticles for Application in Multiplex Immunoassay. *BioChip Journal* 13, 64-81.

Rondeau, E., Cooper-White, J.J., 2008. Biopolymer Microparticle and Nanoparticle Formation within a Microfluidic Device. *Langmuir* 24, 6937-6945.

Santa-Maria, M., Scher, H., Jeoh, T., 2012. Microencapsulation of bioactives in cross-linked alginate matrices by spray drying. *Journal of Microencapsulation* 29, 286-295.

Seemann, R., Brinkmann, M., Pfohl, T., Herminghaus, S., 2011. Droplet based microfluidics. *Reports on Progress in Physics* 75, 016601.

Shikha, S., Zheng, X., Zhang, Y., 2018. Upconversion Nanoparticles-Encoded Hydrogel Microbeads-Based Multiplexed Protein Detection. *Nano-Micro Letters* 10, 31.

Stephenson, R., Stuart, J., 1986. Mutual binary solubilities: water-alcohols and water-esters. *Journal of Chemical and Engineering Data* 31, 56-70.

Sugaya, S., Yamada, M., Seki, M., 2011. Production of extremely-small hydrogel microspheres by utilizing water-droplet dissolution in a polar solvent, in: Landers, J. (Ed.), 15th International Conference on Miniaturized Systems for Chemistry and Life Sciences. Chemical and Biological Microsystems Society (CBMS), Seattle, Washington, USA, pp. 18-20.

Sundberg, D.C., Casassa, A.P., Pantazopoulos, J., Muscato, M.R., Kronberg, B., Berg, J., 1990. Morphology development of polymeric microparticles in aqueous dispersions. I. Thermodynamic considerations. *Journal of Applied Polymer Science* 41, 1425-1442.

Teh, S.Y., Lin, R., Hung, L.H., Lee, A.P., 2008. Droplet microfluidics. *Lab on a Chip* 8, 198-220.

Trivedi, V., Doshi, A., Kurup, G.K., Ereifej, E., Vandevord, P.J., Basu, A.S., 2010. A modular approach for the generation, storage, mixing, and detection of droplet libraries for high throughput screening. *Lab on a Chip* 10, 2433-2442.

Trivedi, V., Ereifej, E.S., Doshi, A., Sehgal, P., Vandevord, P.J., Basu, A.S., 2009. Microfluidic encapsulation of cells in alginate capsules for high throughput screening, *Proceedings of the 31st Annual International Conference of the IEEE Engineering in Medicine and Biology Society: Engineering the Future of Biomedicine, EMBC 2009*, pp. 7037-7040.

Utech, S., Prodanovic, R., Mao, A.S., Ostafe, R., Mooney, D.J., Weitz, D.A., 2015. Microfluidic Generation of Monodisperse, Structurally Homogeneous Alginate Microgels for Cell Encapsulation and 3D Cell Culture. *Advanced Healthcare Materials* 4, 1628-1633.

Uyen, N.T.T., Hamid, Z.a.A., Tram, N.X.T., Ahmad, N., 2020. Fabrication of alginate microspheres for drug delivery: A review. *International Journal of Biological Macromolecules* 153, 1035-1046.

Velings, N.M., Mestdagh, M.M., 1995. Physico-chemical properties of alginate gel beads. *Polymer Gels and Networks* 3, 311-330.

Wang, C.X., Cowen, C., Zhang, Z., Thomas, C.R., 2005. High-speed compression of single alginate microspheres. *Chemical Engineering Science* 60, 6649-6657.

Wang, W., Zhou, C., 2020. A Journey of Nanomotors for Targeted Cancer Therapy: Principles, Challenges, and a Critical Review of the State-of-the-Art. *Advanced Healthcare Materials*, 2001236.

Xu, J.H., Li, S.W., Tan, J., Luo, G.S., 2008. Controllable Preparation of Monodispersed Calcium Alginate Microbeads in a Novel Microfluidic System. *Chemical Engineering & Technology* 31, 1223-1226.

Xu, Q., Hashimoto, M., Dang, T.T., Hoare, T., Kohane, D.S., Whitesides, G.M., Langer, R., Anderson, D.G., 2009. Preparation of Monodisperse Biodegradable Polymer Microparticles Using a Microfluidic Flow-Focusing Device for Controlled Drug Delivery. *Small* 5, 1575-1581.

Yu, L., Sun, Q., Hui, Y., Seth, A., Petrovsky, N., Zhao, C.-X., 2019. Microfluidic formation of core-shell alginate microparticles for protein encapsulation and controlled release. *Journal of Colloid and Interface Science* 539, 497-503.

Zagnoni, M., Anderson, J., Cooper, J.M., 2010. Hysteresis in Multiphase Microfluidics at a T-Junction. *Langmuir* 26, 9416-9422.

Zhang, C., 2020. Development of a microfluidic method for the preparation of mimetic microparticles of red blood cells with controllable size and mechanical properties. *Université d'Aix-Marseille*.

Zhang, C., Grossier, R., Lacaria, L., Rico, F., Candoni, N., Veessler, S., 2020. A microfluidic method generating monodispersed microparticles with controllable sizes and mechanical properties. *Chemical Engineering Science* 211, 115322.

Zhang, H., Tumarkin, E., Peerani, R., Nie, Z., Sullan, R.M.A., Walker, G.C., Kumacheva, E., 2006. Microfluidic Production of Biopolymer Microcapsules with Controlled Morphology. *Journal of the American Chemical Society* 128, 12205-12210.

Zhang, H., Tumarkin, E., Sullan, R.M.A., Walker, G.C., Kumacheva, E., 2007. Exploring Microfluidic Routes to Microgels of Biological Polymers. *Macromolecular Rapid Communications* 28, 527-538.

Zhang, J., Shikha, S., Mei, Q., Liu, J., Zhang, Y., 2019. Fluorescent microbeads for point-of-care testing: a review. *Microchimica Acta* 186, 361.

Zhang, S., Guivier-Curien, C., Veessler, S., Candoni, N., 2015. Prediction of sizes and frequencies of nanoliter-sized droplets in cylindrical T-junction microfluidics. *Chemical Engineering Science* 138, 128-139.

Zhao, X., Cui, Y., He, Y., Wang, S., Wang, J., 2020. Synthesis of Multi-mode Quantum Dots Encoded Molecularly Imprinted Polymers Microspheres and Application in Quantitative Detection for Dopamine. *Sensors and Actuators B: Chemical* 304, 127265.

Zhu, P., Wang, L., 2017. Passive and active droplet generation with microfluidics: a review. *Lab on a Chip* 17, 34-75.