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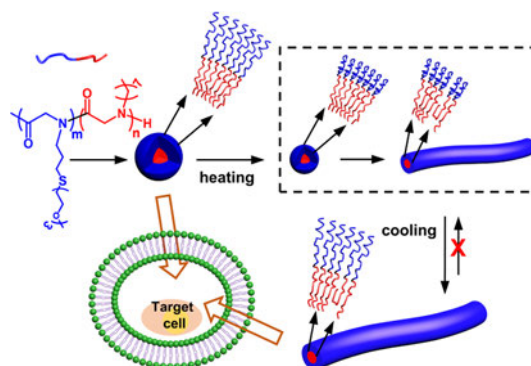
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Thermoinduced Crystallization-Driven Self-Assembly of Bioinspired Block Copolymers in Aqueous Solution

Zhiwei Wang,^{||} Min Lin,^{||} Colin Bonduelle, Rongye Li, Zhekun Shi, Chenhui Zhu, Sébastien Lecommandoux,* Zhibo Li,* and Jing Sun*

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ABSTRACT: Delicate control over architectures via crystallization-driven self-assembly (CDSA) in aqueous solution, particularly combined with external stimuli, is rare and challenging. Here, we report a stepwise CDSA process thermally initiated from amphiphilic poly(*N*-allylglycine)-*b*-poly(*N*-octylglycine) (PNAG-*b*-PNOG) conjugated with thiol-terminated triethylene glycol monomethyl ethers ((PNAG-*g*-EG₃)-*b*-PNOG) in aqueous solution. The diblock copolymers show a reversible thermoresponsive behavior with nearly identical cloud points in both heating and cooling runs. In contrast, the morphology transition of the assemblies is irreversible upon a heating–cooling cycle because of the presence of a confined domain arising from crystalline PNOG, which allows for the achievement of different nanostructured assemblies by the same polymer. We demonstrated that the thermoresponsive property of PNAG-*g*-EG₃ initiates assembly kinetically that is subsequently promoted by crystallization of PNOG thermodynamically. The irreversible morphology transition behavior provides a convenient platform for comparing the cellular uptake efficiency of nanostructured assemblies with various morphologies that are otherwise similar.



INTRODUCTION

Biological organisms fabricate elegant architectures with remarkable performance through a bottom-up assembly process.^{1–4} The assembly mechanism underlying such a dynamic process is often complex and remains a challenging question. Typically, several environmental factors are combined in a single biological event. For instance, spider dragline silk production is accompanied by a structural transition regulated by sequential pH variations.⁵ In recent years, block copolymers obtained by a comparatively simpler synthetic approach represent ideal building blocks for the design of bioinspired structures despite less delicacy and control. Particularly, block copolymers containing a crystalline core, such as polyferrocenyl dimethylsilane, polyethylene, and polythiophene can induce crystallization-driven self-assembly (CDSA), a strategy that has offered a higher level tunability on the structure and kinetic process as compared to other self-assemblies.^{6–12} A couple of biocompatible copolymers including poly(2-isopropyl-2-oxazoline) and poly(L-lactide) segments have already been used in the CDSA strategy, which benefits biomedical applications.^{13–19} However, challenges of their controlled fabrication, in particular combined with other assembly processes involving stimuli-responsive features, represent an important limitation. To conceive such a well-defined architecture, motifs with excellent tunability on both chemical structures and functionalities are desired.

Bioinspired peptide-based polymers have been reported as the most explored building blocks for the formation of a hierarchical biomimetic architecture.^{20–22} Their structural isomers, namely, polypeptoids, offer a flexible backbone, simplified molecular interactions, and controllable fabrication over architectures, and have emerged as promising self-assembly motifs for de novo hierarchical structures.^{23–27} In particular, polypeptoids with alkyl groups exhibit a tunable semicrystalline behavior that differs from polypeptides.^{28–33} For example, in a previous report on a block copolypeptoids, the thermoresponsive poly(*N*-(*n*-propyl)glycine) block serves as a crystallizable micellar core with a soluble poly(*N*-methylglycine) shell as the temperature increases above the cloud point temperature.³³ Instead, the hydrophobic poly(*N*-alkylglycine) with longer alkyl groups is known to arrange in a rectangular crystalline lattice.³¹

It is generally accepted that water is unfavorable for the CDSA.³⁴ A couple of solutions have been addressed, for example, an increasing chain length of hydrophilic segments or

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addition of cosolvents.^{14,15,17} However, cosolvent methods employ organic solvents that limit the potential for biomedical application; while increasing the chain length of hydrophilic segments generally disfavors the organization of a crystalline core for further assembly because of the strong repulsion of long hairy shells. Herein, to tackle these limitations, we designed a new type of block copolymer, a poly(*N*-allylglycine)-*b*-poly(*N*-octylglycine) (PNAG-*b*-PNOG) diblock copolypeptoid conjugated with thiol-terminated triethylene glycol monomethyl ethers ((PNAG-*g*-EG₃)-*b*-PNOG). The aim of such a design is to embed long hydrophilic PNAG-*g*-EG₃ segments with a lower critical solution temperature (LCST) behavior in a crystalline block copolymer, by which the thermostimulus triggers the collapse of hydrophilic chains to facilitate the ordered fabrication of the crystalline PNOG block upon heating. The obtained diblock copolymers exhibit a reversible thermoresponsive behavior with nearly the same cloud points (CPs) during the heating and cooling runs in aqueous solution. Instead, the morphology transition of the assemblies is irreversible upon a heating–cooling cycle that results in different morphologies by the same copolymer. By taking this advantage, we performed cellular uptake experiments using three types of cells on both spherical and cylindrical constructs wherein the surface chemistry remained similar. We demonstrate that the copolymers formulated in cylindrical assemblies are initially internalized to a greater extent than those in the spherical ones and eventually reach a comparable value.

EXPERIMENTAL SECTION

Materials and Methods. Tetrahydrofuran (THF), dichloromethane (DCM), and hexane were first purified by purging with dry N₂, followed by passing through a column of activated alumina. Glyoxylic acid monohydrate (98%), allylamine (98%), octylamine (98%), benzylamine (99%), di-*tert*-butyl decarbonate (Boc₂O) (97%), and 2,2-dimethoxy-2-phenylacetophenone (DMPA) were purchased from Aladdin Reagent. All other chemicals were purchased from commercial suppliers and used without further purification unless otherwise noted. ¹H NMR spectra were recorded on a Bruker AV500 FT-NMR spectrometer. Gel permeation chromatography (GPC) analysis was conducted using an SSI pump connected to a Wyatt Optilab DSP with 0.02 M LiBr in DMF as the eluent at a flow rate of 1.0 mL/min at 50 °C. All GPC samples were prepared at concentrations of 5–10 mg/mL. Conventional calibrations were performed using polystyrene (PS) standards. Differential scanning calorimetry (DSC) studies were conducted using a TA DSC Q20 calorimeter under nitrogen. Powder samples enclosed in aluminum pans were heated from –40 to 200 °C at 10 °C/min for three cycles. Cloud point measurements of aqueous solutions of diblock copolymers (1 wt %) were conducted using a HITACHI U-2900 UV–vis spectrophotometer equipped with a Thermo/Neslab RTE-7 refrigerated bath circulator for temperature control. The experimental temperature range was 25–75 °C and all UV–vis absorptions were referenced against distilled water. The temperature at 50% UV–vis transmittance ($\lambda = 500$ nm) is defined as the cloud point. Atomic force microscopy (AFM) studies were conducted using tapping mode AFM (Bruker Multimode 8 AFM/SPM system) in ambient air with Nanoscope software with probes acquired from Bruker probes. A volume of the polymer solution (~10 μ L) was drop-deposited and dried on freshly cleaved mica under ambient conditions for 24 h before AFM imaging. Minimal processing of the images was done using NanoScope Analysis software from Bruker. Transmission electron microscopy (TEM) experiments were conducted on a FEI TECNAI 20, with a Gatan digital camera and analyzed using Gatan Digital Micrograph software. Samples were prepared on holey carbon-coated 200 mesh copper grids. The polymer solution (6 μ L) was

pipetted onto the carbon-coated side. The grids were then blotted to remove the excess solution and then negatively stained with 0.5 wt % uranyl acetate. The solvent was evaporated under ambient conditions. Confocal laser scanning microscopy (CLSM) images were recorded on a Nikon C2 plus (Nikon Corporation, Japan). Solution small-angle X-ray scattering (SAXS) experiments were performed at the Center de Recherche Paul Pascal (CRPP) in Bordeaux using a high-resolution Xeuss 2.0 X-ray spectrometer from Xenox operating at 8.04 keV. Two-dimensional (2D) scattering patterns were collected using a Pilatus 300K Dectris detector and the beam center position and the angular range were calibrated using a silver behenate standard sample. Finally, the two-dimensional SAXS patterns were radially averaged to obtain the one-dimensional scattering profiles.

Synthesis of Poly(*N*-allylglycine)-*b*-poly(*N*-octylglycine) Diblock Copolymers. The *N*-allyl *N*-carboxyanhydride (Allyl-NCA, M₁) and *N*-octyl *N*-carboxyanhydride (Oct-NCA, M₂) monomers were synthesized following a published procedure.³⁵ All of the diblock copolypeptoids were synthesized by benzylamine-initiated ring-opening polymerization of the corresponding *N*-substituted *N*-carboxyanhydrides in a sequential manner. In a representative procedure, allyl-NCA (508.1 mg, 3.6 mmol, 0.4 M) was dissolved in anhydrous THF in a glovebox. A stock solution of benzylamine (255.0 μ L, 24.0 μ mol, 94.2 mM, [M₁]₀/[–NH₂]₀ = 150:1) was added. The polymerization was performed under a N₂ atmosphere at 50 °C for 24 h and monitored by FTIR until the characteristic peaks of allyl-NCA disappeared. A solution of Oct-NCA in THF (1.8 mL, 0.7 mmol, 0.4 M, [M₁]₀/[M₂]₀/[–NH₂]₀ = 150:30:1) was then added to the above mixture and allowed to stir at room temperature for another 24 h to reach full conversion. The sample solution was precipitated into excess ether. The polymer was collected by filtration and washed with ample hexane followed by drying under vacuum (340.0 mg, 72.0% yield). The composition of the polymer was determined using end-group analysis by ¹H NMR spectroscopy. The number-average degree of polymerization (DP) of each block was calculated based on the integration ratios of characteristic proton peaks of individual blocks.

Synthesis of Poly(*N*-allylglycine)-*b*-poly(*N*-octylglycine) Modified with Thiol-Terminated Triethylene Glycol Monomethyl Ether. Thiol-terminated diethylene glycol monomethyl ethers (OEG₃-SH) were synthesized using relative oligo(ethylene glycol) monomethyl ethers according to literature procedures.³⁶ Typically, PNAG₁₄₂-*b*-PNOG₂₃ (300.0 mg) and DMPA (16.0 mg) were dissolved in DMF. Then OEG₃-SH (1.4 mL, [–SH]/[–C=C] = 5) was added. The reaction mixture was degassed through three freeze/thaw/pump cycles before being irradiated with UV light at room temperature for 4 h. The mixture was then dialyzed against Millipore water for 3 days and lyophilized to yield (PNAG-*g*-EG₃)₁₄₂-*b*-PNOG₂₃ as a white powder (473.0 mg, 68.0% yield). All of the other copolymers were modified in a similar way.

Preparation of Polymer Solutions. The block polymer was dispersed in water at a desired concentration in a clean vial. The solution was stirred at room temperature for 5 days, after which the solution was heated to 60 °C and held at the temperature for 2 h. Then the solution was cooled to room temperature naturally.

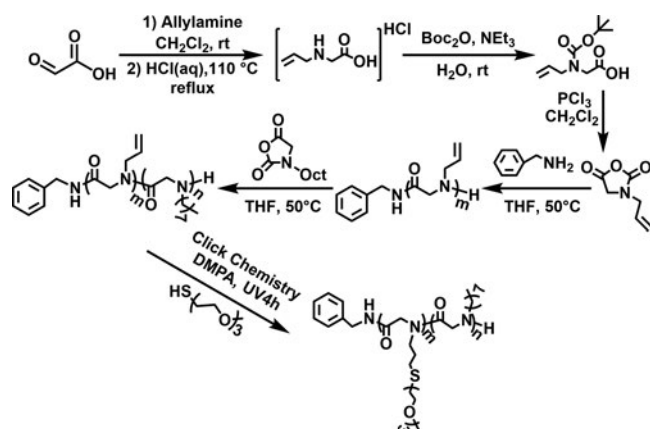
Cellular Uptake. The fluorescein isothiocyanate (FITC)-labeled polymer was prepared by the following procedure. (PNAG-*g*-EG₃)₁₄₂-*b*-PNOG₂₃ (20.0 mg), diisopropylethylamine (10.0 μ L, 0.06 mmol), and FITC (1.0 mg) were dissolved in 2 mL of DMF, followed by stirring at room temperature in the dark overnight. The product was dialyzed with DMSO and then deionized water for a few days to remove the unreacted fluorescent dye and lyophilized. Cells were seeded in a 96-well plate with a density of 5000 cells per well. The plate was then incubated in a humidified 5% CO₂ in air incubator at 37 °C for 24 h. Then, 100 μ L of fresh culture medium replaced the original medium, and the aqueous solution of the assembly was added into the plate with the final concentration of 0.5 mg/mL. The plate was then incubated in a humidified 5% CO₂ in air incubator at 37 °C for different times. The cells were washed with 100 μ L of phosphate buffer saline (PBS) three times. Finally, the total volume of 100 μ L was added into the plate. The cells were imaged by a confocal laser

scanning microscope. All parameters of the microscope remained constant in the entire testing process. The fluorescence intensity of each cell was measured by CLSM. The mean fluorescence intensity was defined as the total fluorescence intensity divided by the total cell number.^{37,38}

RESULTS AND DISCUSSION

The PNAG-*b*-PNOG diblock copolypeptoids were synthesized by sequential ring-opening polymerization of Oct-NCA using PNAG as the macroinitiator (Scheme 1). By varying the

Scheme 1. Synthetic Route to the Diblock Copolypeptoids



monomer/initiator ratio, a series of diblocks with different DPs were obtained. The chemical structures and DPs of the blocks were investigated by ¹H NMR spectroscopy (Figure 1a). All

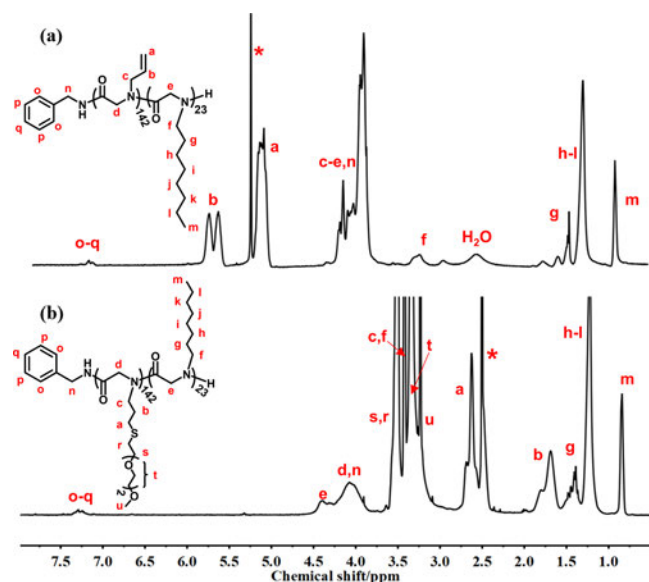


Figure 1. (a) ¹H NMR spectra of PNAG₁₄₂-*b*-PNOG₂₃ in CD₂Cl₂ (* indicates CD₂Cl₂). (b) ¹H NMR spectra of (PNAG-*g*-EG₃)₁₄₂-*b*-PNOG₂₃ in DMSO-*d*₆ (* indicates DMSO-*d*₆). The molecular characteristics are shown in Table S1.

peaks of the diblock copolymers are well assigned in the ¹H NMR spectra, confirming the diblock chemical structures. Table S1 summarizes the molar mass and dispersity (*D*) of all of the samples, where the subscripts correspond to the DPs, respectively. The GPC traces show narrow weight distributions with *D* ≤ 1.24. The polypeptoids were then conjugated with

OEG₃-SH to yield (PNAG-*g*-EG₃)-*b*-PNOG. In a typical ¹H NMR spectrum, the protons of the OEG moiety at 3.5 ppm are visible, while the protons of the allyl group completely disappear, suggesting the quantitative conversion of the alkenyl group (Figure 1b). GPC traces of (PNAG-*g*-EG₃)-*b*-PNOG reveal a similar dispersity with a noticeable shift to a higher molar mass region, indicative of insignificant side reactions in the process of postmodification (Figure S1). The minor shoulder can be attributed to the byproducts during thiol-ene click chemistry,³⁹ which is typically seen in previous reports. Similarly, a series of PNAG-*g*-EG₃ with different DPs were obtained. The molecular characteristics of (PNAG-*g*-EG₃)-*b*-PNOG along with their abbreviations are shown in Table S1.

The thermal properties of (PNAG-*g*-EG₃)-*b*-PNOG were first investigated by DSC measurements as shown in Figure S2 and summarized in Table S2. The PNOG₂₀ homopolymer is crystallizable with two melting transitions at around 51 and 163 °C as reported, which are associated with different chain packings of both side chains and backbones.³¹ In marked contrast, the lack of melting peaks and crystallization exotherms suggests that the PNAG-*g*-EG₃ segment is non-crystalline. With a high molar fraction of PNOG, the DSC endotherms of diblock copolymers display two peaks with similar melting transitions to PNOG homopolymers. As the molar fraction of PNOG is decreased to 0.27 or lower, merely one broad melting transition with centered peaks in the range of 53–61 °C is present, suggesting that crystallization of PNOG is largely suppressed by the presence of PNAG-*g*-EG₃. This is further confirmed by the decreased melting enthalpy and melting temperature with increasing molar fraction of PNAG-*g*-EG₃.

Pegylated poly(*N*-propargylglycine) has been reported to show an LCST behavior in aqueous solution.³⁶ These polymers can readily dissolve in water at room temperature and collapse at elevated temperatures. Further cooling can result in a reversible phase transition. As expected, PNAG-*g*-EG₃ also exhibits a similar reversible LCST behavior, as shown in Figure S3. A representative plot of PNAG₉₅-*g*-EG₃ demonstrates a CP of around 49.4 °C with 50% transmittance during heating. The CP_{cooling} of 48.8 °C is shown with 100% recovered transmittance during the cooling ramp. This variation in CPs could be explained by the overcooling during rehydration of OEG units as reported.⁴⁰ As the DP of the polymer decreases from 95 to 59, the CP increases from 49.4 to 51.2 °C. We attribute this to the increased solubility of shorter polymer chains that results in enhanced CPs.⁴¹ The incorporation of the PNOG block significantly decreases the CPs of the systems by 10–15 °C, which is associated with the decreased solubility (Figure 2). Further, as compared to the homopolymer PNAG₉₅-*g*-EG₃ that shows high dependence of CP on concentration, the CP of the block copolymer (PNAG₉₄-*g*-EG₃)-*b*-PNOG₁₅ with almost the same DP of pegylated PNAG varies slightly with concentration change. We assume that this is because the formation of the aggregates reduces the intermolecular interaction at high concentrations. We will address this shortly. More interestingly, the CPs during the heating and cooling runs remain nearly the same, which indicates the rapid rehydration of OEG units. This is possibly because the OEG moieties are aggregated, which increases the hydrogen-bonding interaction with water.⁴² Furthermore, with a fixed DP of the PNOG block, increasing the DP of PNAG-*g*-EG₃ reduces the CP, consistent with the homopolypeptoids. In addition, increasing the DP of PNOG can also decrease the CP

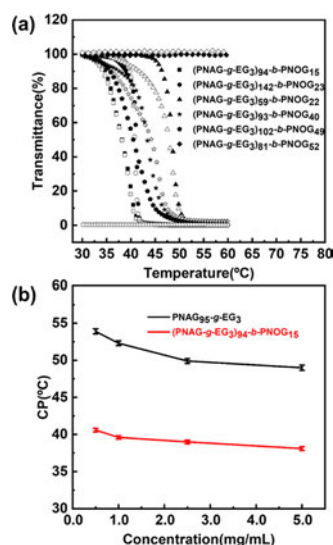


Figure 2. (a) Plots of transmittance versus temperature for aqueous solutions of (PNAG-g-EG₃)-b-PNOG at a concentration of 5 mg/mL. Filled symbol: heating ramp and open symbol: cooling ramp. (b) Plots of CP versus concentration for aqueous solutions of (PNAG-g-EG₃)₉₄-b-PNOG₁₅ and PNAG₉₅-g-EG₃ at different concentrations.

with a fixed DP of PNAG-g-EG₃ due to the increased hydrophobicity of PNOG. Note that as the DP of the PNOG increases to 49 and the molar fraction of PNOG is 0.32, the sample shows an irreversible LCST behavior. Further increasing the DP of PNOG to 52 and the molar fraction to 0.39 results in the absence of the LCST property. In most cases, the phase transition of both the homopolymer and block copolymer is recovered after 8 heating and cooling cycles between 30 and 60 °C, suggesting good stability (Figure S4a). Further, the CPs remain constant with addition of salt. This suggests that both systems are stable to brine, which provides great potential for biomedical applications (Figure S4b).

All of the block copolymers can be readily dispersed in water at room temperature, possibly due to the significantly long chain length of the PNAG-g-EG₃ block with good water solubility. Considering the amphiphilicity of the system, the copolymers are expected to form assemblies. We performed TEM characterization to determine the shape of nanostructures. Upon negative staining, TEM images of the (PNAG-g-EG₃)₁₄₂-b-PNOG₂₃ copolymer with the lowest molar fraction of PNOG of 0.14 reveal the presence of a spherical-shaped-structure with an average diameter of 11.5 nm when prepared at room temperature after 1 day of stirring (Figure 3a). AFM characterization confirms the spherical structures with a similar diameter (~11.2 nm) and comparable height (~4.1 nm), consistent with TEM results (Figure 3b). It is conceivable that the hydrophilic PNAG-g-EG₃ forms the shell and the hydrophobic PNOG constitutes the core of the spheres, thus leading to the micellar core-shell structure. Similarly, the diblock copolymer (PNAG-g-EG₃)₉₄-b-PNOG₁₅ with the same molar fraction of PNOG but a smaller molar mass shows a majority of sphere-like morphology with an average diameter of 9.8 nm, as observed by TEM (Figure 3c). A few cylindrical aggregates with a diameter similar to that of the spheres are occasionally present. The presence of cylinders probably result from a fusion process of the spheres, as evidenced by AFM (Figure 3d).^{43,44} As the molar fraction of PNOG increases, the predominant morphology is observed to be cylindrical,

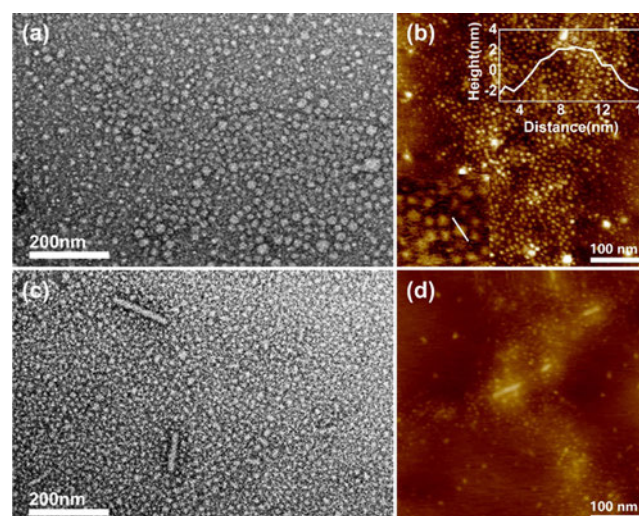


Figure 3. TEM images of (PNAG-g-EG₃)₁₄₂-b-PNOG₂₃ (a) and (PNAG-g-EG₃)₉₄-b-PNOG₁₅ (c) stirred for 24 h at room temperature in aqueous solution at a concentration of 1 mg/mL. AFM images of (PNAG-g-EG₃)₁₄₂-b-PNOG₂₃ (b) and (PNAG-g-EG₃)₉₄-b-PNOG₁₅ (d) stirred for 24 h at room temperature in aqueous solution at a concentration of 1 mg/mL.

irrespective of the molar mass of the block copolymer (Figure S5).

As expected, the self-assemblies exhibit a thermoresponsive behavior upon heating. With the (PNAG-g-EG₃)₁₄₂-b-PNOG₂₃ solution heated up to 60 °C (>CP of ~41.1 °C), a sphere-to-cylinder transition is explicitly shown. Figure 4a shows that the length of the cylinders is up to ~560 nm and the diameter is ~11.3 nm, nearly identical to the diameter of the spheres. The PNAG-g-EG₃ collapses with the temperature increasing above the CP, which enables protruding sticky ends of PNOG to fuse with the adjacent one to grow into cylinders. Note that the amide bonds on the backbone of the copolymers can form hydrogen bonding with water, which stabilizes the system. It is known that polysarcosine with one methyl group on the side chain shows excellent solubility in water.²⁷ A type of semi-bald micelle has been reported by both simulations and experiments for the thermogelling mechanism, which is likely to occur.⁴⁵ In most previous reports, the assemblies from thermoresponsive diblock copolymers exhibit reversible morphological evolution with temperature.^{46–50} Remarkably, Figure 4b shows that the cylindrical nanostructure persists with similar dimensions as the solution cools back to the room temperature. We assume that this irreversible transition is due to the presence of crystalline PNOG within the core that restricts the disassembly of the system. To probe the detailed molecular packing and assembly mechanism, grazing incidence wide-angle X-ray scattering (GIWAXS) was performed (Figure 5a). The scattering peak at the $q = q^* = 3.0 \text{ nm}^{-1}$ in plane is due to the Bragg reflections of PNOG crystals, associated with the side-chain packing as reported.³⁰ The spacing is given by $d = 2\pi/q = 2.1 \text{ nm}$. An additional broad peak at around 14.1 nm^{-1} indicates the disordered packing, confirming that the crystallization of PNOG is suppressed. It is shown that as the solution is heated to 60 °C additional higher-order peaks at $2q^*$ and $3q^*$ are visible, indicative of the presence of the lamellae. The variations suggest that the cylinders contain higher-ordered packing of PNOG domains (Scheme 2). Note that the (PNAG-g-EG₃)₁₄₂-b-PNOG₂₃ bulk sample melts over

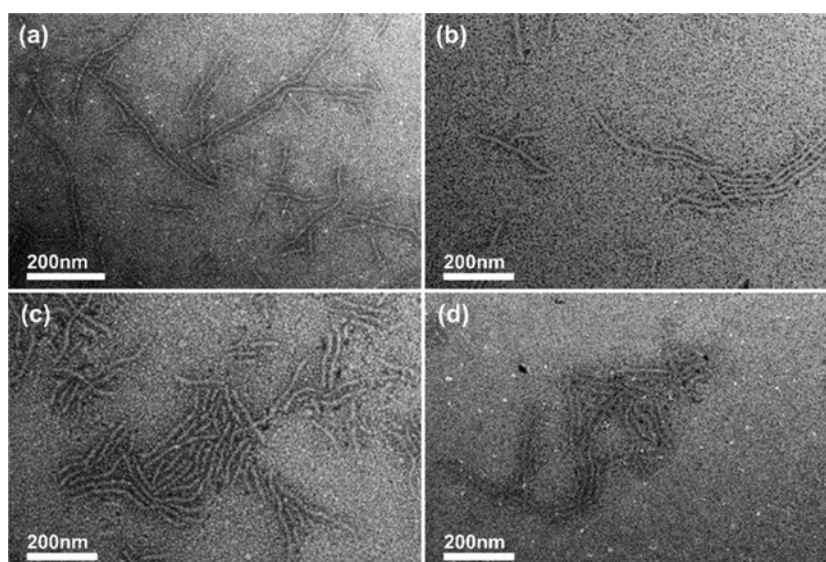


Figure 4. TEM images of (PNAG-*g*-EG₃)₁₄₂-*b*-PNOG₂₃ (a) and (PNAG-*g*-EG₃)₉₄-*b*-PNOG₁₅ (c) incubated at 60 °C for 2 h after stirring for 24 h at room temperature in aqueous solution at a concentration of 1 mg/mL (grid prepared at 60 °C). TEM images of (PNAG-*g*-EG₃)₁₄₂-*b*-PNOG₂₃ (b) and (PNAG-*g*-EG₃)₉₄-*b*-PNOG₁₅ (d) after cooling the solutions in (a) and (c) to the room temperature, respectively.

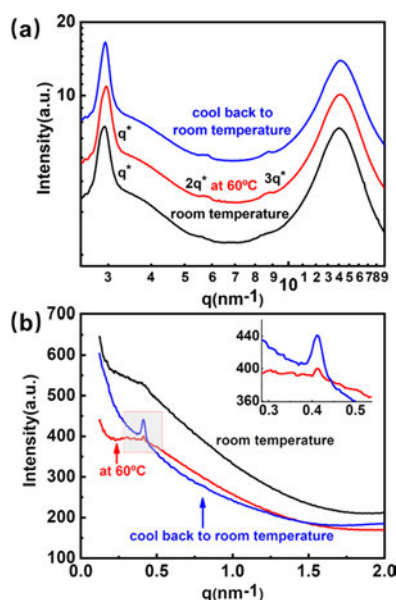
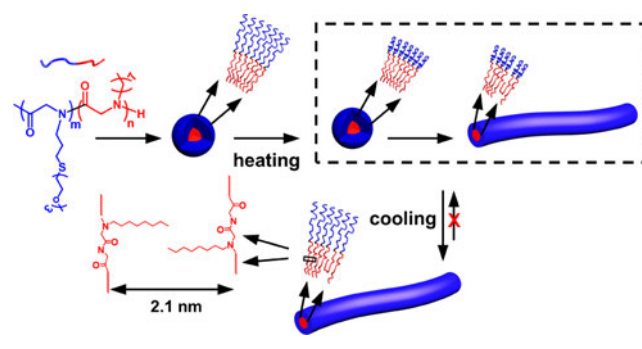


Figure 5. GIWAXS in-plane (a) and SAXS (b) measurements of the sphere and cylinder from (PNAG-*g*-EG₃)₁₄₂-*b*-PNOG₂₃ in aqueous solution at a concentration of 1 mg/mL. The black line represents the (PNAG-*g*-EG₃)₁₄₂-*b*-PNOG₂₃ stirred for 24 h at room temperature in aqueous solution at a concentration of 1 mg/mL. The red line represents the (PNAG-*g*-EG₃)₁₄₂-*b*-PNOG₂₃ incubated at 60 °C for 1 h after stirring for 24 h at room temperature in aqueous solution at a concentration of 1 mg/mL. The blue line represents the (PNAG-*g*-EG₃)₁₄₂-*b*-PNOG₂₃ cooled to the room temperature after stirring for 24 h at room temperature and incubating at 60 °C for 1 h in aqueous solution at a concentration of 1 mg/mL.

a wide range of ~40–70 °C. Thus, the PNOG may partially melt and the increased mobility enables the long-range order of polymer chains for the thermodynamically favorable transition. As the solution cools to room temperature, a sharpened primary peak and higher-order peaks suggest more ordered chain packing. It is thus conceivable that the restrained PNOG chains prevent the reversible morphological evolution from

Scheme 2. Sphere and Cylinder Structure of the Crystalline Diblock Copolyptoids



cylinders to spheres. We therefore propose that the LCST behavior of the system initiates the assembly process by enabling collapsed PNAG-*g*-EG₃ chains and exposure of PNOG sites with the temperature increasing above the CP. Subsequent crystallization of PNOG promotes the formation of the cylinder and further stabilizes the architecture with confined crystalline domains. Note that despite the thermoresponsive behavior being recovered after 8 times of heating and cooling runs, the cylindrical assembly is significantly stable after the first cooling process (Figure S4a). This suggests that the “locked” morphology is independent of the environmental conditions once formed.

Aqueous solution of the assemblies was further analyzed by SAXS. A clear Bragg-like scattering peak at 0.41 nm^{−1} in a low-*q* range is observed in the heating and recooling solution (Figure 5b). Note that the peak intensity in the recooling process is more pronounced than that in the heating process. The spacing is ca. 15.2 nm, which can be attributed to the repeating units of a cluster of laterally arrayed filaments, consistent with the TEM images in the dry state. We have previously observed that the fibers assembled from poly(ethylene glycol)-*b*-poly(*N*-(2-phenylethyl)glycine) align laterally in water and further fuse into platelets.⁵¹ In this case, the absence of nanosheets is perhaps due to the long hydrophilic

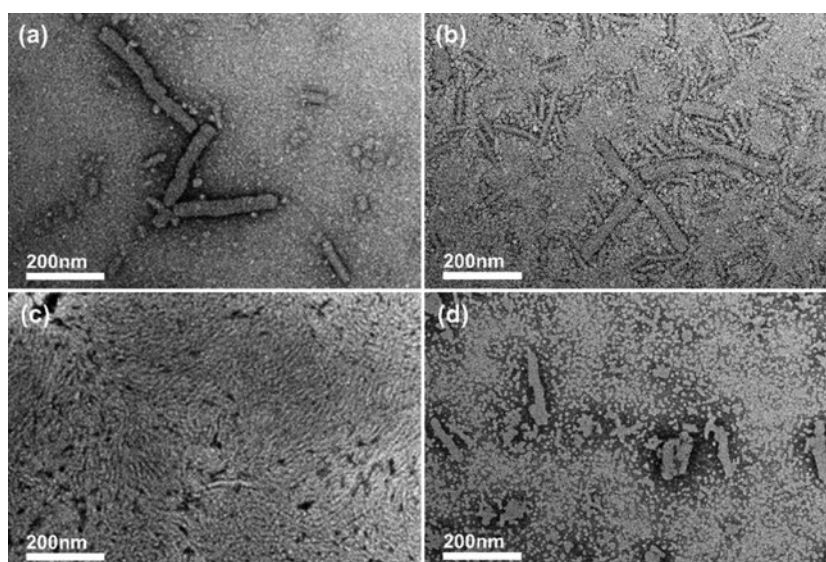


Figure 6. TEM images of (PNAG-*g*-EG₃)₁₀₂-*b*-PNOG₄₉ (a) and (PNAG-*g*-EG₃)₈₁-*b*-PNOG₅₂ (c) incubated at 60 °C for 2 h after stirring for 24 h at room temperature in aqueous solution at a concentration of 1 mg/mL (grid prepared at 60 °C). TEM images of (PNAG-*g*-EG₃)₁₀₂-*b*-PNOG₄₉ (b) and (PNAG-*g*-EG₃)₈₁-*b*-PNOG₅₂ (d) after cooling the solutions in (a) and (c) to the room temperature, respectively.

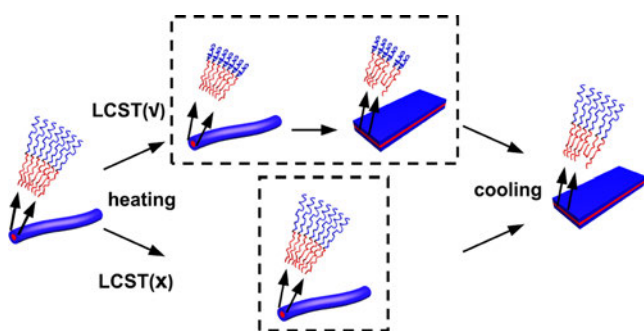
PNAG-*g*-EG₃ segments that prevent the lateral cohesion of the fibers. Note that the diameter from TEM is smaller than that from SAXS, which is possibly because of the volume shrinkage of the assemblies during the drying process and an invisible corona with a low electron density by TEM.⁵² This result confirms that the fibers assemble at an elevated temperature in aqueous solution and remain stable upon recooling, which agrees well with the proposed mechanisms.

Note that simply increasing the stirring time to 5 days and the concentration to 5 mg/mL at room temperature results in exclusively spheres (Figure S6). Increasing the incubation time at elevated temperatures results in consistent morphologies as well (Figure S7). We thus assume that the temperature is significantly crucial for the cylinder formation. To further study the detailed assembly kinetics, we performed the experiments at various temperatures (Figure S8). As expected, it is shown that merely spheres are visible as the (PNAG-*g*-EG₃)₁₄₂-*b*-PNOG₂₃ is heated to 40 °C, slightly lower than the CP. Heating the sample up to 50 °C (>CP) results in the coexistence of the spheres and cylinders with a similar diameter of ~11.3 nm. Since the (PNAG-*g*-EG₃)₁₄₂-*b*-PNOG₂₃ bulk sample melts over a wide range centered at around 60 °C, it is likely that the mobility of PNOG is insufficient to organize into dense-packed crystals at 50 °C, which leads to the reversible transition to spheres at room temperature. Further increasing the temperature up to 80 °C results in the formation of cylindrical structures. However, the length of the cylinders (~150 nm) is typically shorter than that prepared at 60 °C. We assume that this is possibly because the PNOG completely melts at 80 °C, and the assemblies are mainly driven by hydrophobicity and the lack of order. Further cooling down to below the CP results in formation of a long hairy shell that disfavors the long-range ordered packing.

Similarly, Figure 4c,d shows that (PNAG-*g*-EG₃)₉₄-*b*-PNOG₁₅ with the same molar fraction of PNOG exhibit an irreversible sphere-to-cylinder transition. As the molar fraction of PNOG is increased to 0.27 and 0.30, both (PNAG-*g*-EG₃)₅₉-*b*-PNOG₂₂ and (PNAG-*g*-EG₃)₉₃-*b*-PNOG₄₀ remain as the cylindrical structures during the heating–cooling cycle

despite the different molar masses (Figure S9). In both cases, the lengths of cylinders significantly grow from ~143 to 386 nm and ~156 to 403 nm upon heating, respectively. Both lengths persist after cooling back to the room temperature. This short-to-long cylinder transition is very consistent with the sphere-to-cylinder transition. Further increasing the molar fraction of PNOG results in the formation of a two-dimensional nanosheet of (PNAG-*g*-EG₃)₁₀₂-*b*-PNOG₄₉ and (PNAG-*g*-EG₃)₈₁-*b*-PNOG₅₂, as shown in Figure 6. In both cases, two melting transitions are observed by DSC. The AFM images show sheetlike structures with thicknesses of ~4.2 and 4.5 nm, respectively, which are comparable to the spheres (Figure S10). Interestingly, it is observed that both samples behave fairly distinctly at elevated temperatures. In the case of (PNAG-*g*-EG₃)₁₀₂-*b*-PNOG₄₉, the nanosheet forms upon heating and remains during the cooling process. As the temperature increases to the first melting transition temperature, the PNAG-*g*-EG₃ chains collapse and simultaneously PNOG chains transition into a mesophase, both of which enable the higher-ordered structures. However, (PNAG-*g*-EG₃)₈₁-*b*-PNOG₅₂ assembles into nanosheets merely after the solution is cooled back to room temperature. The cylinders are exclusively observed at elevated temperatures, which suggests that the assembly of nanosheets is initiated upon cooling. Note that (PNAG-*g*-EG₃)₈₁-*b*-PNOG₅₂ excludes the thermoresponsive property over the entire experimental range. Although largely the chain mobility increases in the mesophase, the formation of nanosheets is inhibited by the long hairy PNAG-*g*-EG₃ block during heating (Scheme 3). In this specific case, the formation of nanosheets is entirely driven by crystallization of PNOG upon cooling. This confirms the proposed formation mechanism, where the collapsed PNAG-*g*-EG₃ chains first facilitate the packing of exposed PNOG, followed by promotion of crystallization. Note that the nanosheets assembled from (PNAG-*g*-EG₃)₈₁-*b*-PNOG₅₂ appear to be more irregular than those from (PNAG-*g*-EG₃)₁₀₂-*b*-PNOG₄₉. This suggests that well-controlled structures can be obtained by incorporating hydrophilic segments with the thermores-

Scheme 3. Cylinder and Nanosheet Structure of the Crystalline Diblock Copolypeptoids



ponsive property, which offers great potential for biomedical applications.

In nature, a number of viruses that infect animals exhibit a cylindrical morphology, indicative of possible morphological advantages.^{53–55} Compared to a large number of studies of size and surface chemistry effects on cellular uptake behaviors, merely a few studies have been conducted on the influence of morphology.^{15,17,56–59} In this study, we addressed this issue by comparing cellular uptakes of spherical and cylindrical structures in three types of cells by CLSM. The diblock copolymer (PNAG-*g*-EG₃)₁₄₂-*b*-PNOG₂₃ was first labeled with FITC quantitatively (Figure S11). The cellular uptakes were assessed in HeLa cells for potential application in tumor therapy. The CLSM images of HeLa cells after incubation with both FITC-labeled assemblies for 2 h clearly show that both assemblies are internalized by the cells (Figure 7). Further, the

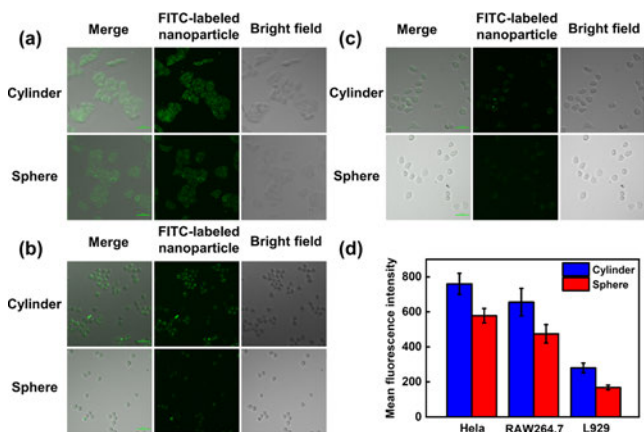


Figure 7. Confocal laser scanning microscopy images of HeLa cells after 2 h incubation (a), RAW264.7 after 2 h incubation (b), and L929 cells after 4 h incubation (c) with FITC-labeled assemblies at a concentration of 0.5 mg/ml at 37 °C. Scale bars = 50 μ m. (d) Mean fluorescence intensity in HeLa cells, RAW264.7, and L929 cells after 2 h incubation with FITC-labeled nanoparticle assemblies at a concentration of 0.5 mg/ml at 37 °C.

fluorescence intensity from the cylindrical assembly is significantly higher than that from the spheres, quantitatively confirmed by mean fluorescence intensity determination (Figure 7d). The time-dependent experiment further shows that the copolymers formulated in cylindrical assemblies are initially internalized to a higher extent than those in spherical ones, and eventually reach a comparable value (Figure S12). Note that nearly all of the cells spread very well after 16 h, indicating the low cytotoxicity of the biocompatible poly-

peptoids as previously reported.⁶⁰ Similar morphology-dependent cellular uptake behaviors were also observed in RAW264.7 and L929 cells by CLSM. In L929 cells, the fluorescence intensity is lower than that in the other two cells, indicating the lower cellular uptake. Note that the cylinders are much larger in volume compared with the spheres. Thus, it is hypothesized that the long and flexible structure may span across a larger contact area for stronger and faster binding with cell membranes.^{56,58}

CONCLUSIONS

In summary, we have designed and synthesized a series of diblock copolypeptoids (PNAG-*g*-EG₃)-*b*-PNOG that exhibit a reversible thermoresponsive behavior with nearly the same CPs during the heating and cooling runs in aqueous solution. Remarkably, the morphology transition of the assemblies observed upon heating becomes irreversible upon cooling because of the presence of crystalline PNOG. This PNOG behaves as a chemical leverage to “freeze” cylindrical morphologies obtained above the CP at temperatures below the same CP. We have therefore demonstrated a CDSA process combined with nanoprecipitation initiated by thermo-stimulus. Specifically, the thermoresponsive property of the system initiates the assembly process, promoted by subsequent crystallization of PNOG and physically locks the morphology eventually. Both thermoresponsive and crystalline features coordinately facilitate the architecture and assembly kinetics. Such a process allows the formation of different nano-structured assemblies from the same block copolymers. We have further studied the influence of nanoparticles’ shape on cell internalization while keeping all molecular parameters constant. Cellular uptake experiments using three types of cells have shown that the copolymers formulated in cylindrical assemblies are initially internalized to a greater extent than those in spherical ones of identical surface chemistry and eventually reach a comparable value. This combined self-assembly strategy offers a new opportunity to construct new generations of bioinspired nanomaterials with bioactive hierarchical nanostructures for biomedical applications.

ASSOCIATED CONTENT

Supporting Information

The Supporting Information is available free of charge at <https://pubs.acs.org/doi/10.1021/acs.biomac.0c00844>.

Additional GPC data, DSC data, NMR data, GIWAXS results, TEM images, and AFM images (PDF)

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Notes

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Supporting Information

Thermoinduced crystallization-driven self-assembly of bioinspired block copolymers in aqueous solution

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Results and Discussion

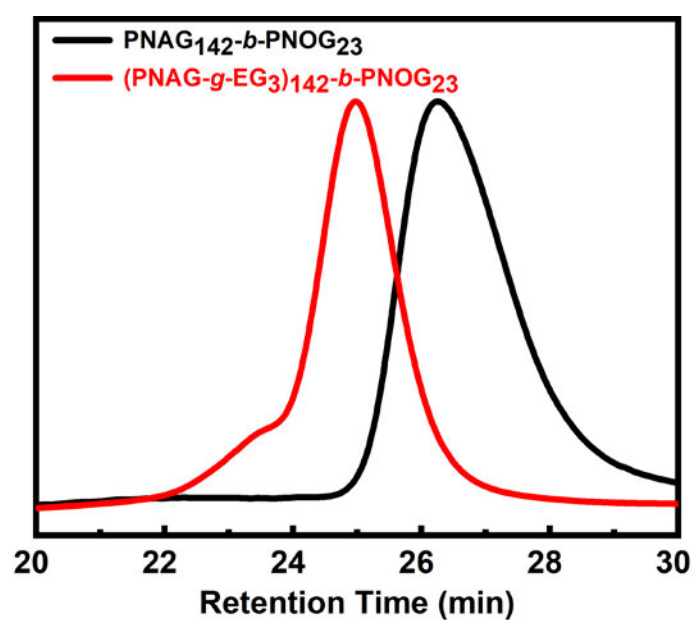
Table S1. Molecular parameters of all the polymers.

Samples	Feed ratio ^[a]	m/n ^[b]	Mn ^[c] (kDa)	Mn ^[d] (kDa)	Dispersity (<i>D</i>) ^[d]	X _{PNOG} ^[e]
PNOG ₂₀	20	20	3.4	2.7	1.07	-
PNAG ₉₅ -g-EG ₃	95	95	18.0	14.9	1.09	-
PNAG ₇₈ -g-EG ₃	80	78	14.7	11.1	1.09	-
PNAG ₅₉ -g-EG ₃	60	59	11.2	8.8	1.06	-
PNAG ₈₁ - <i>b</i> -PNOG ₅₂	95/60	81/52	16.6	12.4	1.16	0.39
(PNAG-g-EG ₃) ₈₁ - <i>b</i> -PNOG ₅₂	-	81/52	24.1	18.5	1.18	0.39
PNAG ₁₀₂ - <i>b</i> -PNOG ₄₉	100/50	102/49	18.1	15.6	1.06	0.32
(PNAG-g-EG ₃) ₁₀₂ - <i>b</i> -PNOG ₄₉	-	102/49	27.6	19.7	1.10	0.32
PNAG ₉₃ - <i>b</i> -PNOG ₄₀	95/30	93/40	15.8	12.3	1.12	0.30
(PNAG-g-EG ₃) ₉₃ - <i>b</i> -PNOG ₄₀		93/40	24.3	18.6	1.15	0.30
PNAG ₅₉ - <i>b</i> -PNOG ₂₂	60/30	59/22	9.4	8.6	1.18	0.27
(PNAG-g-EG ₃) ₅₉ - <i>b</i> -PNOG ₂₂	-	59/22	14.9	11.8	1.24	0.27
PNAG ₉₄ - <i>b</i> -PNOG ₁₅	95/20	94/15	11.6	9.7	1.11	0.14
(PNAG-g-EG ₃) ₉₄ - <i>b</i> -PNOG ₁₅	-	94/15	20.3	17.3	1.13	0.14
PNAG ₁₄₂ - <i>b</i> -PNOG ₂₃	150/30	142/23	17.6	15.2	1.07	0.14
(PNAG-g-EG ₃) ₁₄₂ - <i>b</i> -PNOG ₂₃	-	142/23	30.7	21.2	1.09	0.14

[a] Feed molar ratio of PEG / Oct-NCA; [b] Calculated from ¹HNMR spectra; [c] Calculated from ¹HNMR spectra; [d] Determined from GPC, [e] Mole fraction of PNOG.

Table S2. DSC data for all the polymers.

Samples	ΔH_1 (J/g)	ΔH_2 (J/g)	T_1 (°C)	T_2 (°C)	T_g (°C)
PNAG ₉₅ - <i>g</i> -EG ₃	-	-	-	-	53.0
PNOG ₂₀	12.3	25.2	50.8	163.5	-
(PNAG- <i>g</i> -EG ₃) ₈₁ - <i>b</i> -PNOG ₅₂	7.4	9.2	46.7	150.8	-
(PNAG- <i>g</i> -EG ₃) ₁₀₂ - <i>b</i> -PNOG ₄₉	3.0	2.7	46.8	147.6	-
(PNAG- <i>g</i> -EG ₃) ₉₃ - <i>b</i> -PNOG ₄₀	2.6	1.7	56.0	104.3	-
(PNAG- <i>g</i> -EG ₃) ₅₉ - <i>b</i> -PNOG ₂₂	5.5	-	60.5	-	-
(PNAG- <i>g</i> -EG ₃) ₁₄₂ - <i>b</i> -PNOG ₂₃	4.2	-	60.9	-	-
(PNAG- <i>g</i> -EG ₃) ₉₄ - <i>b</i> -PNOG ₁₅	2.3	-	53.3	-	-

**Figure S1.** GPC chromatograms of the polymers. The molecular characteristics are shown in Table S1.

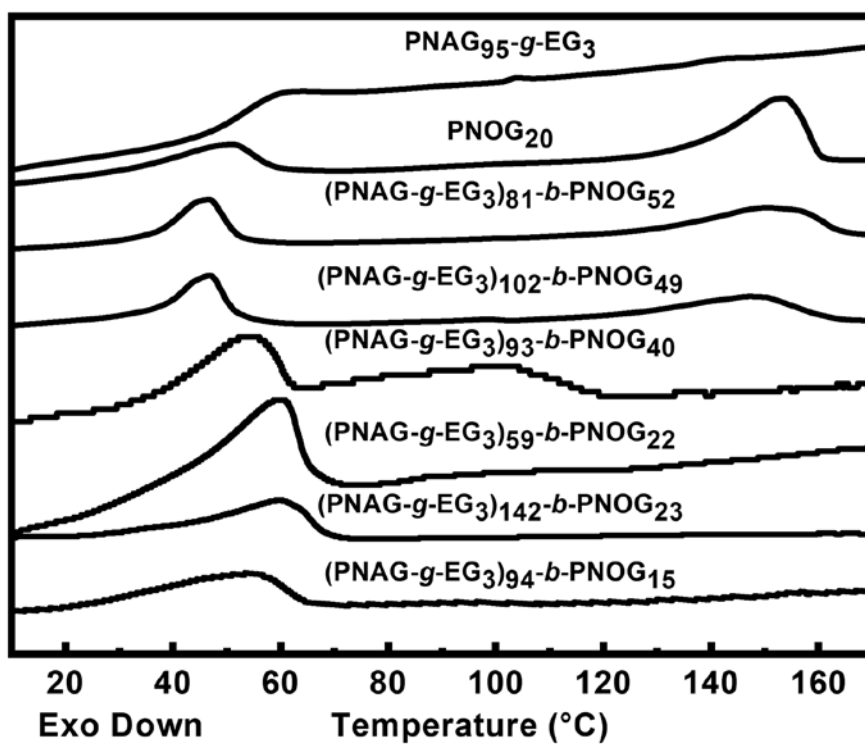


Figure S2. The DSC thermograms of all the polymers. The detailed information is summarized in Supplementary Table 2.

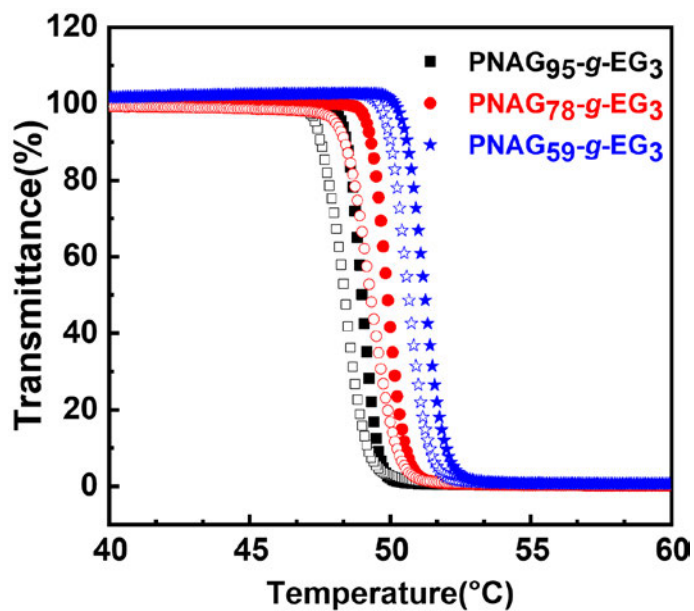


Figure S3. Plots of the transmittance versus temperature for aqueous solutions of PNAG-g-EG₃ at a concentration of 5 mg/mL. Filled symbol: heating ramp, open symbol: cooling ramp.

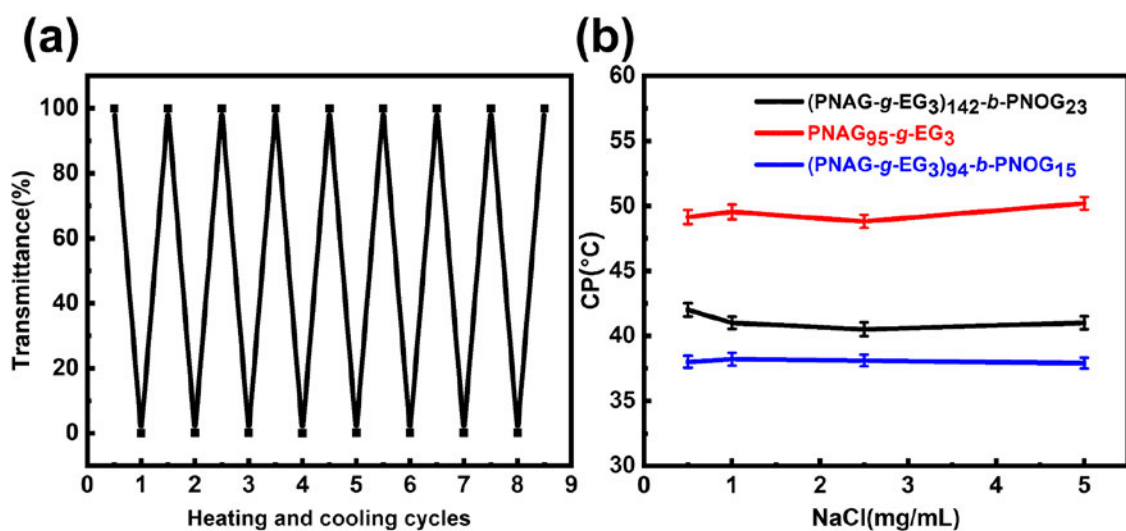


Figure S4. (a) Transmittance of (PNAG-g-EG₃)₁₄₂-b-PNOG₂₃, PNAG₉₅-g-EG₃ and (PNAG-g-EG₃)₉₄-b-PNOG₁₅ aqueous solution at a concentration of 5 mg/mL versus 8 heating and cooling cycles between 30 and 60 °C. (b) Plots of CP as a function of NaCl concentration for (PNAG-g-EG₃)₁₄₂-b-PNOG₂₃, PNAG₉₅-g-EG₃ and (PNAG-g-EG₃)₉₄-b-PNOG₁₅ solution at a concentration of 5 mg/mL.

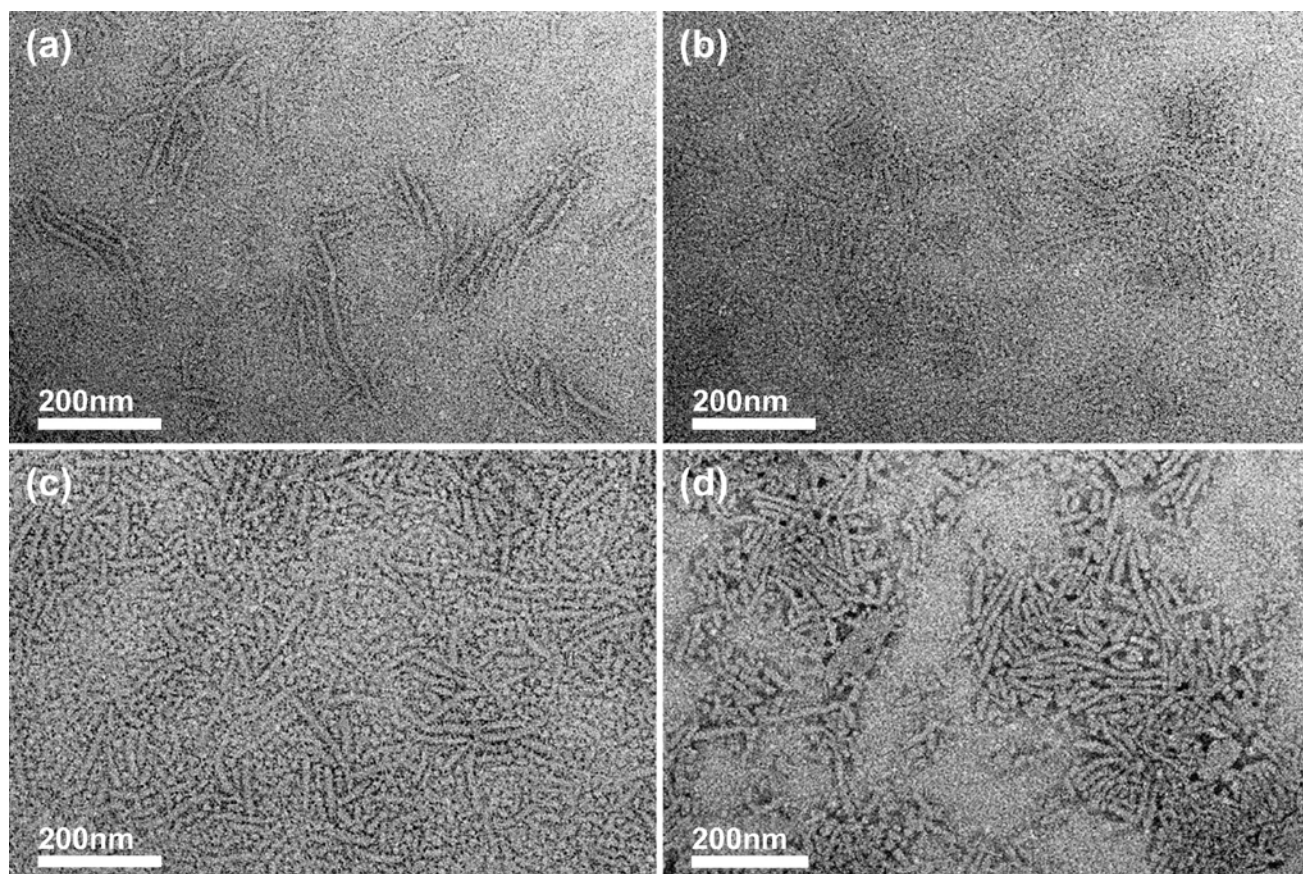


Figure S5. TEM images of (PNAG-g-EG₃)₅₉-b-PNOG₂₂ (a), (PNAG-g-EG₃)₉₃-b-PNOG₄₀ (b), (PNAG-g-EG₃)₁₀₂-b-PNOG₄₉ (c) and (PNAG-g-EG₃)₈₁-b-PNOG₅₂ (d) stirred for 24 h at room temperature in aqueous solution at a concentration of 1 mg/mL.

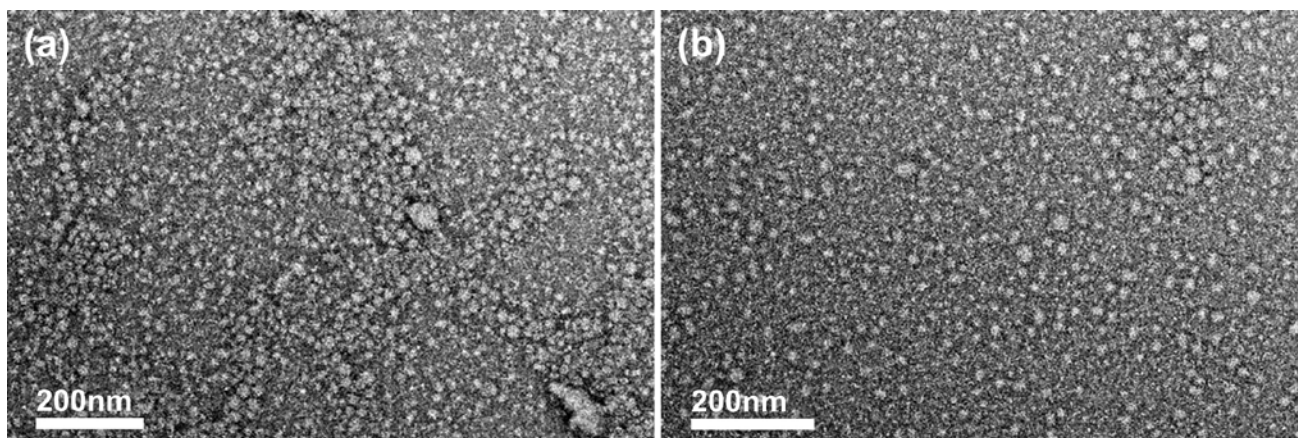


Figure S6. TEM images of (PNAG-*g*-EG₃)₁₄₂-*b*-PNOG₂₃ stirred for 5 days at room temperature in aqueous solution at a concentration of 1 mg/mL (a) and 5 mg/mL (b).

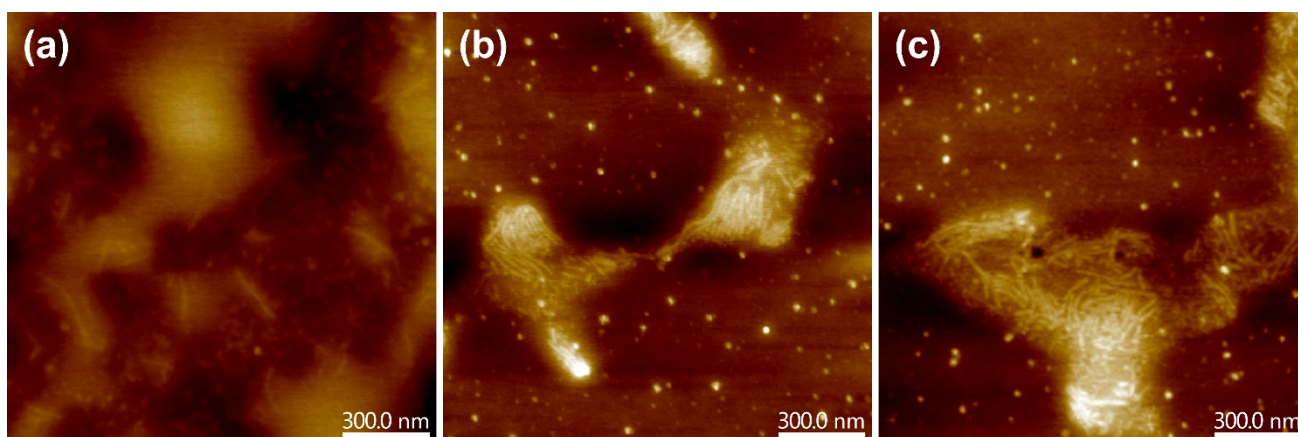


Figure S7. AFM images of (PNAG-*g*-EG₃)₁₄₂-*b*-PNOG₂₃ incubated at 60 °C for 1 h (a), 2 h (b) and 3 h (c) after stirring for 24 h at room temperature in aqueous solution at a concentration of 1 mg/mL.

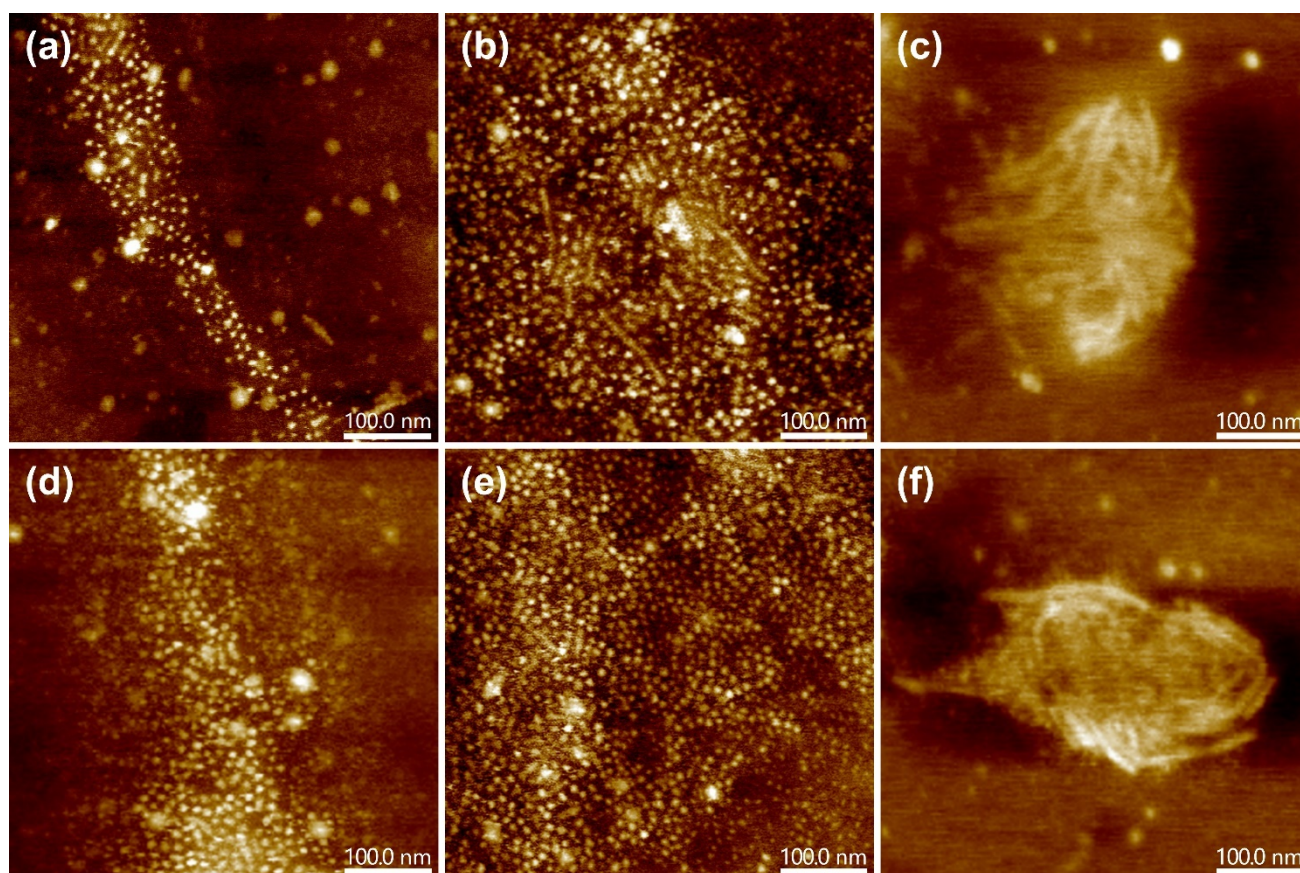


Figure S8. AFM images of (PNAG-*g*-EG₃)₁₄₂-*b*-PNOG₂₃ incubated at 40 °C (a), 50 °C (b) and 80 °C (c) for 2 h after stirring for 24 h at room temperature in aqueous solution at a concentration of 1 mg/mL. AFM images of assemblies (d), (e) and (f) after cooling solution of (a), (b) and (c) to the room temperature, respectively.

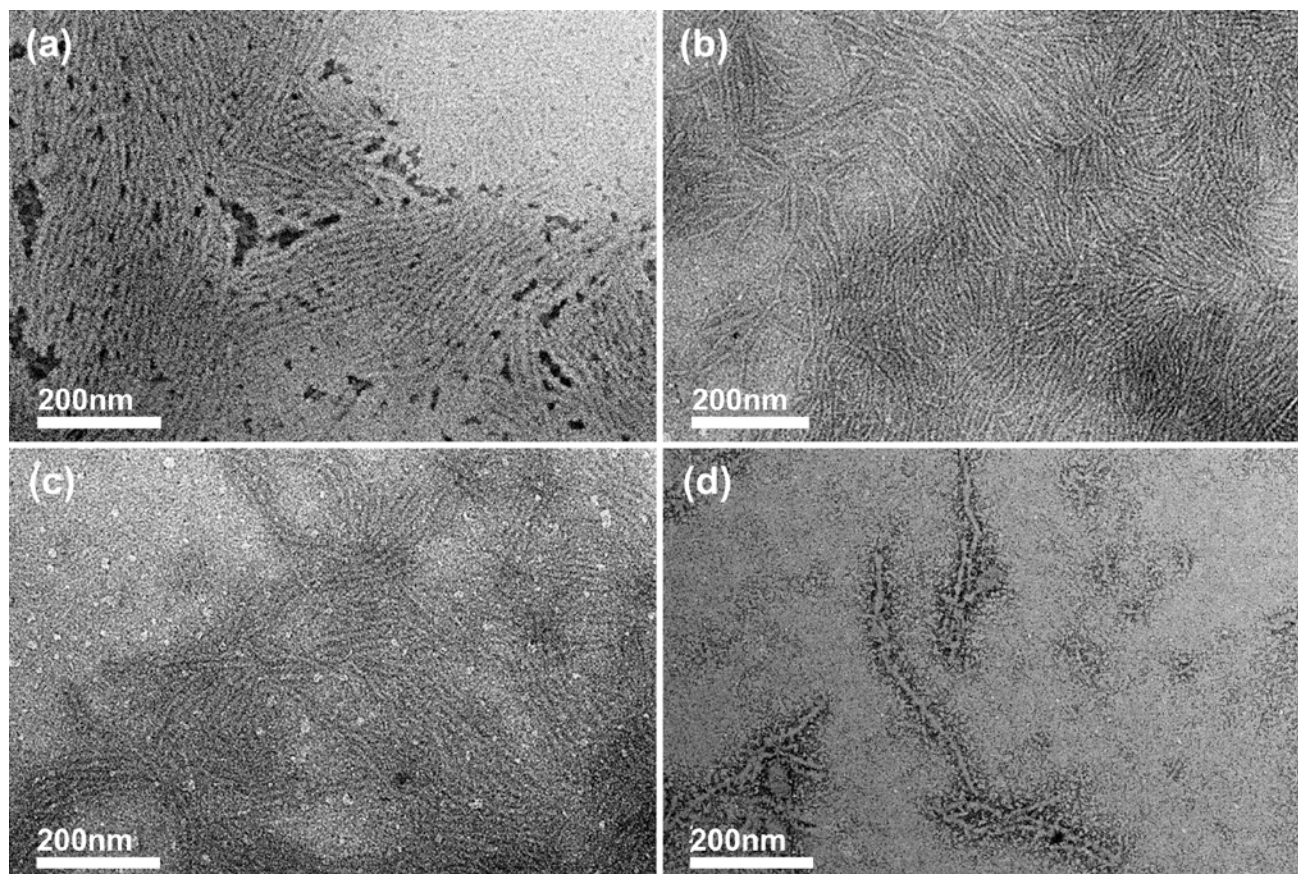


Figure S9. TEM images of (PNAG-*g*-EG₃)₅₉-*b*-PNOG₂₂ (a), (PNAG-*g*-EG₃)₉₃-*b*-PNOG₄₀ (c), incubated at 60 °C for 2 h after stirring for 24 h at room temperature in aqueous solution at a concentration of 1 mg/mL. TEM images of (PNAG-*g*-EG₃)₅₉-*b*-PNOG₂₂ (b), (PNAG-*g*-EG₃)₉₃-*b*-PNOG₄₀ (d) after cooling the solutions in (a) and (c) to the room temperature, respectively.

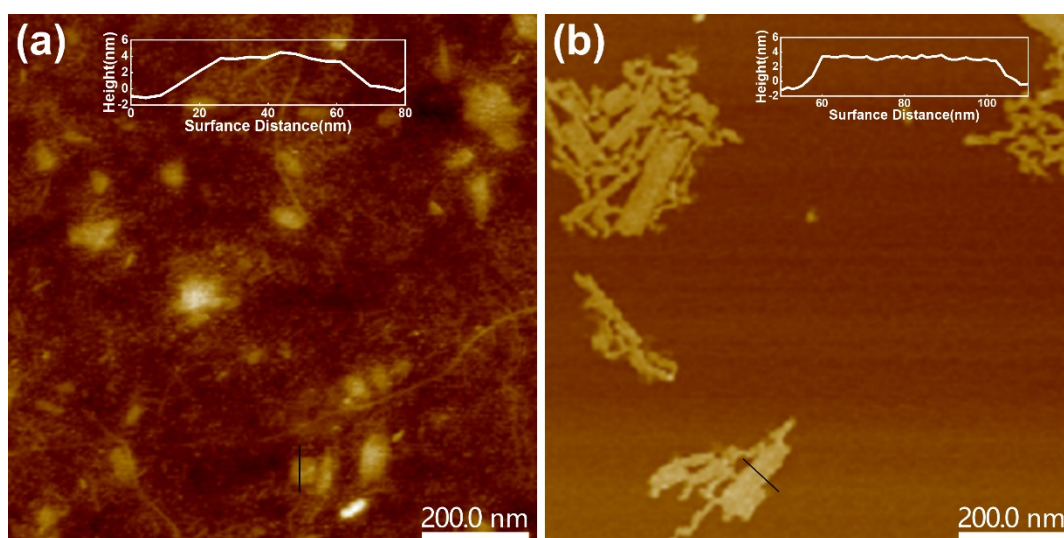


Figure S10. AFM images of (PNAG-*g*-EG₃)₈₁-*b*-PNOG₅₂ (a) and (PNAG-*g*-EG₃)₁₀₂-*b*-PNOG₄₉ (b) incubated at 60 °C for 2 h after stirring for 24 h at room temperature in aqueous solution at a concentration of 1 mg/mL.

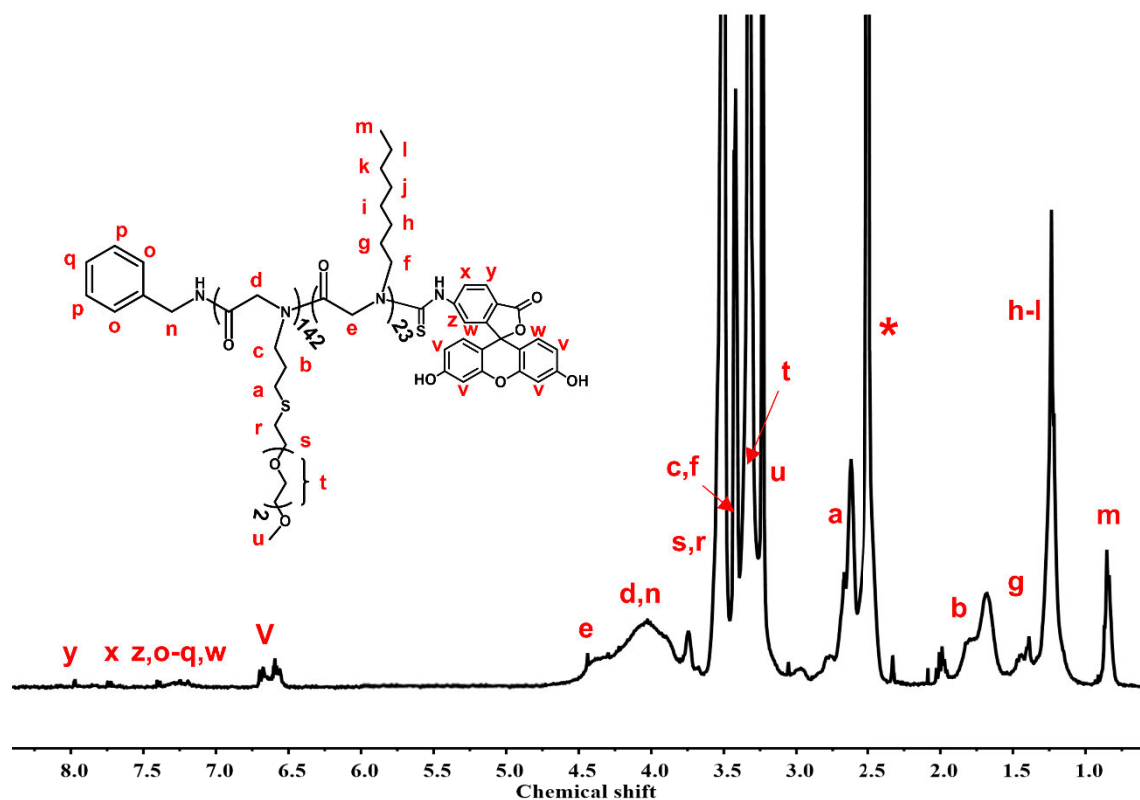


Figure S11. ^1H NMR spectra of $(\text{PNAG-g-EG3})_{142}\text{-b-PNOG}_{23}$ labeled with fluorescein isothiocyanate (FITC) quantitatively in $\text{DMSO-}d_6$ (* indicates $\text{DMSO-}d_6$).

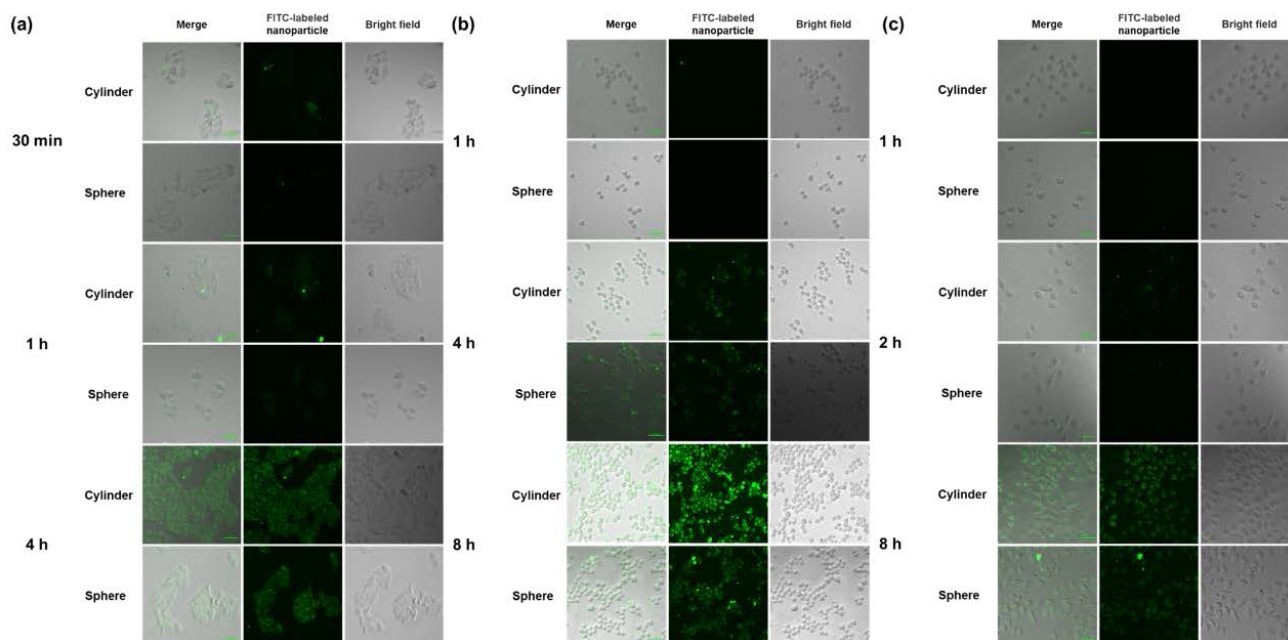


Figure S12. Confocal laser scanning microscope images of HeLa cells (a), RAW264.7 (b) and L929 (c) cells incubated with FITC-labeled assemblies at a concentration of 0.5 mg/ml for different incubation time at 37 °C. Scale bars = 50 μm .