

### Neglected viral diseases in freshwater fish farming

Olga Haenen, L. Bigarré, Takafumi Ito, Jean-Christophe Avarre, Niccoló Vendramin

#### ▶ To cite this version:

Olga Haenen, L. Bigarré, Takafumi Ito, Jean-Christophe Avarre, Niccoló Vendramin. Neglected viral diseases in freshwater fish farming. Bulletin of the European Association of Fish Pathologists, 2018, 38 (3), pp.105-115. hal-02922463

HAL Id: hal-02922463

https://hal.science/hal-02922463

Submitted on 26 Aug 2020

**HAL** is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

# EAFP WORKSHOP ON NEGLECTED VIRAL DISEASES IN FRESHWATER FISH FARMING Open workshop at the 18<sup>th</sup> EAFP Conference, Belfast, UK, 4<sup>th</sup> September 2017

O. Haenen\*<sup>1</sup>, L. Bigarré<sup>2</sup>, T. Ito<sup>3</sup>, J.-C. Avarre<sup>4</sup>, N. Vendramin<sup>5</sup>

#### **INTRODUCTION**

Worldwide, viral diseases pose a serious challenge to the fish farming industry and wild fish stocks. In order to prevent the spread of serious fish diseases and to contain and control fish disease outbreaks at a global scale, the World Organisation for Animal Health (OIE) has published relevant data on these fish diseases, both in the Aquatic Animal Health Code (OIE, 2017a), including the OIE listed fish diseases, and in the Manual of Diagnostic Tests for Aquatic Animals (OIE, 2017b), including recommended diagnostic methods. The list of viral OIE notifiable fish diseases currently includes: Epizootic haematopoietic necrosis disease (EHN), Infection with HPR-deleted or HPRO infectious salmon anaemia virus (ISA), Infection with salmonid alphavirus, causing Pancreas disease (PD) and Sleeping disease (SD), Infectious haematopoietic necrosis (IHN), Koi herpesvirus disease (KHVD), Red sea bream iridoviral disease (RSIVD), Spring viraemia of carp (SVC), and Viral haemorrhagic septicaemia (VHS). At European Union level, Commission Decision 2006/88/EC and Council Implementing Decision 2015/1554/EC provide specific regulation for surveillance and control of listed infectious aquatic diseases in Europe which include the fish viral diseases VHS, IHN, EHN, ISA, and KHVD (European Commission 2006, 2015).

Due to the frequent emergence of new serious fish viral diseases, the OIE list of fish viral diseases is regularly being extended. However, apart from these well studied viral diseases, other non-notifiable serious fish viral diseases occur in freshwater fish farming. To focus on these new viral threats for freshwater fish farming, an open workshop was organized at the EAFP Conference at Belfast, 4<sup>th</sup> September 2017. The workshop consisted of five short lectures and a discussion, involving an audience of 69 international experts, originating from 25 countries, of which 6 outside Europe.

The main topics presented included issues related to:

i) the difficulty of preventing the global spread of cyprinid herpesvirus 2 (CyHV-2), ii) perhabdoviruses as a threat for percid farming, iii) pathogenesis and diagnostics of piscine orthoreoviruses in farmed rainbow trout, iv) Carp Edema Virus (CEV) in Europe, and v) the potential role of fish endogenous retroviruses in disease emergence. The general aim of the workshop was to identify potential collaborative approaches to carry out multidisciplinary studies aiming to define risks, diagnostic methods and suggest adequate prophylactic measures.

<sup>&</sup>lt;sup>1#</sup>Corresponding author, Leading organizer, Wageningen Bioveterinary Research, NRL for Fish, Crustacean and Shellfish Diseases, Lelystad, The Netherlands, <u>olga.haenen@wur.nl;</u> <sup>2\*</sup>ANSES, Plouzané, France; <sup>3\*</sup>Tamaki Laboratory, Aquatic Animal Health Division, National Research Institute of Aquaculture (NRIA), Fisheries Research Agency, Mie 519-0423 Japan; <sup>4\*</sup>ISEM, IRD, CNRS, EPHE, University of Montpellier, Montpellier, France: <sup>5\*</sup>Technical University of Denmark, NVI, Fish Diseases Unit (EURL), Lyngby, Denmark.

<sup>#</sup>Corresponding Author / Workshop leading organizer: <a href="olga.haenen@wur.nl">olga.haenen@wur.nl</a>

<sup>\*</sup> Workshop co-organizer

#### DIFFICULTY OF PREVENTING SPREAD OF CYPRINID HERPESVIRUS 2 (CyHV-2)

Takafumi Ito and Olga L. M. Haenen

Cyprinid herpesvirus 2 (CyHV-2) is known as the causative agent of herpesviral haematopoietic necrosis (HVHN) in goldfish (*Carassius auratus auratus*) and the occurrence of the virus has also been reported from multiple countries. CyHV-2 has caused death in goldfish regardless of the economic value. Moreover, the virus has also been detected from Prussian carp (*C. gibelio*) and crucian carp (*C. carassius*) from European and Asian countries.

In order to reduce economic losses to the aquaculture industry and impacts on natural resources by CyHV-2 infection, prevention of the spread and any further introductions of the virus are important. However, this disease is difficult to prevent from spreading due to the following characteristics.

- Since CyHV-2-infected fish look healthy for multiple days after initial infection (Fig. 1) (e.g. 7-10 days depending on conditions), it is difficult to identify and therefore isolate and treat diseased fish. In addition, a behavioural change (lethargy) is observed only a few days before death. These characteristics of CyHV-2 infection may be the reason why the infection might spread in markets, wholesalers and retailers (Ito et al., 2013).
- It is suggested that CyHV-2 is spread through deals including the global trade of goldfish (Table 1) (Ito et al., 2017), and by movements of infected wild fish.
- Since CyHV-2 has caused death in goldfish, and wild and cultured Prussian carp, it is difficult to make general prevention measures.

In order to overcome these difficulties, we think that it is important to develop a quick, simple and highly sensitive diagnostic method for detection of diseased fish as quickly as possible, and to provide goldfish lovers with more general information on the characteristics and symptoms of this disease to enable greater awareness of this disease.

#### **Discussion:**

Sven Bergmann asked, if the virus can be passaged in cell cultures. Takafumi Ito replied, that GFF (Goldfish Fin) cells from goldfish were used, in which the virus was sub-cultivated more than 7 passages. The virus supernatant after primary virus isolation was used for experimentally induced infections and vaccination trials with goldfish.

Fig.1 Infected and dead fish in infection experiment by immersion of CyHV-2



This characteristic of CyHV-2 infection may be the reason that goldfish farmers have difficulty in detecting infected fish on-farm.

Table 1. Results of PCR detection of CyHV-2 DNA and of virus isolation from the imported goldfish

Import source	Clinical status	CyHV-2 DNA	Virus isolation
Singapore	Apparently healthy	Positive	Positive
China	Apparently healthy	Negative	Negative
Israel	Apparently healthy	Positive	Negative
Singapore	Apparently healthy	Negative	Negative
Hong Kong	Apparently healthy	Negative	Negative
Singapore	Apparently healthy	Positive	Negative
Singapore	Apparently healthy	Negative	Negative
Israel	Diseased	Positive	Negative

#### PERHABDOVIRUSES, A THREAT FOR PERCID FARMING

L. Bigarré, F. Pozet, Y. Ledoré, P. Fontaine, and F. Lieffrig

Since their first discovery in 1980 and the development of percid farming since two decades, viruses from the family Rhabdoviridae have been responsible of high losses in percid farms and experimental facilities in Europe (Belgium, France, Ireland, Norway, Switzerland, etc.) (Dannevig et al., 2001; Dorson et al., 1984; Nougayrède et al., 1992; Rodger and Girons, 2008; Ruane et al., 2014; Wahli et al., 2015). To date, reports described mostly losses on perch, but several outbreaks have impacted pike-perch production in 2016. The genetic diversity of these viruses (genus perhabdovirus) is high. At least two species are described: perch rhabdovirus (PRhV) and sea trout rhabdovirus (STRV), each species regrouping a number of variants known and probably a larger number yet to be discovered (Talbi et al., 2011). These viruses may affect non-percid fish species which are therefore potential reservoirs in the wild. The transmissions of these viruses are horizontal (fish to fish) and vertical (genitor to fry). Consequently, the control of the disease is based on disinfection of the eggs and prevention of viral introduction in farms. However, both strategies are insufficiently used in Europe. Diagnostics tools are still poorly developed and rely on complementary methods in specialized laboratories: in vitro culture of the virus and PCR. These perhabdoviruses have been considerably neglected these last years, although they are a main infectious threat that should be actively controlled in the future if the percid farming is to be developed.

<u>Discussion</u>: Torsten Boutrup asked, if it is possible to access samples from the genitors in the Czech Republic. Laurent Bigarré replied, this is not possible. Anne Nichols from Ghent University asked, how long fish should be kept in quarantine. Laurent answered, two to three days for larvae at the right temperature. If there are no symptoms and no mortality, fish may be transferred. Anne asked then about the quarantine period for larger fish. Laurent replied that it is rare to observe symptoms, but it might be needed to quarantine for a longer time (several weeks). Tom Murphy from Dublin asked, how to secure genitors coming from the wild. Laurent replied, that from these fish the virus might

emerge. There is currently a need for a diagnostic tool. Tomas Vesély from Czech Republic asked about sensitive cell lines and temperature. Laurent replied, EPC and BF-2, and depending on the laboratory, incubate these at 14 or 21°C.



Mortality of pike perch larvae caused by disease through a perhadbovirus (picture Y. Ledoré)

### PISCINE ORTHOREOVIRUSES IN FARMED RAINBOW TROUT: PATHOGENESIS UNDER EXPERIMENTAL CONDITIONS AND DIAGNOSTICS

Niccolò Vendramin, Helena Hauge, Anne Berit Olsen, Torunn Taskdal, Øystein Wessel, Anna Luiza Farias Alencar, Maria K Dahle, and Niels Jørgen Olesen

A new disease in farmed rainbow trout *(Onchorhyncus mykiss)* was described in Norway in 2013. Farmed rainbow trout suffered unexplained increased mortality, the diagnostic process ruled out known pathogens and described pathology resembling heart and skeletal muscle inflammation (HSMI) in Atlantic salmon *(Salmo salar)*. HSMI is caused by Piscine orthoreovirus (PRV).

Further investigation led to detection of a sequence with high similarity to PRV in diseased rainbow trout. This finding called for a targeted effort to assess the risk the new PRV-variant pose on farmed rainbow trout by studying transmissibility and disease pathogenesis, and provide information for conducting diagnostic. A cohabitation trial performed in rainbow trout confirmed viral replication in this host, that the virus levels peaked in blood and heart of cohabitants after 6 weeks post challenge. Heart inflammation was diagnosed in all examined rainbow trout cohabitants 8 weeks after challenge.

In conclusion the novel PRV infects and cause heart pathology in rainbow trout. The infection under experimental conditions develops as acute viral infection, characterized by viral peak followed by clearance.

Information on diagnostic tools available and knowledge on viral epidemiology will be provided during the presentation.

<u>Discussion</u>: Mikolaj Adamek informed, they have PRV3 and PRV1 in Germany from an outbreak in rainbow trout. Niccoló requested him to send the isolates to the EURL for Fish Diseases. Mikolaj asked, if you see also mortality in small fish. Niccoló replied, he did not know, but that this disease is normally stress related, like with transport, sea transfer, etc.. Sven Bergmann asked, what is the

relation of PRV to eel, carp, koi, and grass carp reovirus that would grow in cell culture. Niccoló replied, that the viral species mentioned by Sven belong to the group of aquareoviruses, and not to ortho-reoviruses. Currently it is not possible to cultivate piscineorthorevirus in cells culture. Sven inquired about the detection of PRV outside Europe. Niccoló replied that HSMI is described in Canada and Chile, and that Kyle Garver reported PRV in British Columbia, and one case of HSMI in the east coast of Canada. A Canadian colleague mentioned that PRV in British Columbia had been one time HSMI too, and that PRV has been recorded for a long time. A Canadian colleague added, that PRV detection on the East Coast of Canada is not tightly linked to HSMI.

#### CARP EDEMA VIRUS DISEASE (CEVD) IN EUROPE

Olga Haenen

An outline of the current status of CEV detections in carp and koi (Cyprinus carpio) in Europe will be presented with some research results, based on the alert paper in Dis. Aquat. Org. 2017, written by experts from 15 institutions in 11 European countries and USA: Way K., O. Haenen, D. Stone, M. Adamek, S.M. Bergmann, L. Bigarré, N. Diserens, M. El-Matbouli, M.C. Gjessing, V. Jung-Schroers, E. Leguay, M. Matras, N.J. Olesen, V. Panzarin, V. Piačková, A. Toffan, N. Vendramin, T. Veselý, T. Waltzek: The emergence of carp edema virus (CEV) and its significance to European common carp and koi, Cyprinus carpio.

Carp edema virus (CEV) disease, also known as koi sleepy disease, is caused by a poxvirus associated with outbreaks of clinical disease in koi and common carp. It was originally characterised in Japan in the 1970's. International trade in koi has led to the spread of CEV. In 1996, CEVD was reported in the USA. In 2009, the disease was first recognised in Europe. Since its first detection in Europe, detection and diagnostic methods improved, and more EU member states reported CEV associated with disease outbreaks. So far, in Europe, Austria, Czech Republic, France, Germany, Italy, Poland, Switzerland, The Netherlands, and the UK have detected CEV.

The genome of the virus has been partly sequenced, and suggested the existence of distinct geographical populations of CEV infecting both koi and common carp. New qPCR primers successfully detected CEV DNA in formalin-fixed, paraffin-embedded archive material from investigations of unexplained carp mortalities of over 15 years ago.

Disease management and control methods, and biosecurity, good health management and disease surveillance, applied to koi herpesvirus disease, can be equally applied to CEVD. There is little chance, CEVD would be considered for notifiable disease listing. However, governments might take instigating disease control measures. Further research is needed to develop susceptible cells to isolate CEV, and to study the disease pathogenesis and epidemiology to estimate the likely impact of CEVD on European koi and common carp aquaculture and on wild carp stocks.

There is an active CEV network which organized two workshops (EURL, 2015a, 2015b). Please contact Olga Haenen to be added to the mailing list.

<u>Discussion</u>: Mikolaj Adamek invited the audience to attend his presentation on CEV in gill disease at this conference. Sven Bergmann worked on the diagnostics, based on the old Oyamatsu PCR: In India and China there are varieties and differences in strains found. It would be good to investigate different strains circulating in warm water in Europe. Bartolomeo Gorgoglione inquired about the status of the Full genome sequencing of CEV. Mikolaj Adamek replied, they are nearly ready. Maria

Dahle told, her laboratory at Norway works on SGPDV (Salmon Gill Pox Disease Virus) and that they are struggling finding cell cultures and similar challenges. She invited the audience to cooperate. They have finished transcriptional analysis, and have interesting findings. This was warmly welcomed by the audience.

### THE POTENTIAL ROLE OF FISH ENDOGENOUS RETROVIRUSES IN DISEASE EMERGENCE Jean-Christophe Avarre

This presentation raised an interesting question that is often overlooked in the field of viral (re)emergence: the potential role of endogenous retroviruses in fish diseases. While several exogenous retroviruses have been unambiguously identified as etiological agents of some fish diseases (Quackenbush, 2016), no clear association between endogenous retroviruses and diseases has been established in fish so far, perhaps with the exceptions of the salmon leukemia virus, which may have an endogenous origin (Eaton et al, 1994), and the zebrafish endogenous retrovirus, which may be involved in the zebrafish T-cell leukemia (Frazer et al, 2012) (Table 1). The advent of next-generation sequencing has contributed to the accumulation of large fish genomic datasets. Scrutiny of these latter has started to uncover new types of endogenous retroviruses, some of which with a potential for retained infectivity or re-emergence, as is the case in other animal groups (Naville and Volff, 2016). These new findings were presented and discussed.

<u>Discussion</u>: Sven Bergmann asked if there is a possibility that fish retroviruses become zoonotic in the future. Jean-Christophe Avarre replied that nothing was known about that so far, and it just started to being studied now. In murine, endogenous retroviruses may be transmitted between different species (Stocking and Kozak, 2008). Torsten Boutrup added that there could be a risk for pathogens in warm waters, while it was unlikely in cold waters. He added that walleye sarcoma virus was temperature dependent.

#### Figure(s)?

#### **General discussion**

#### 1- Which is the most important neglected viral disease in freshwater fish farming?

Jason Mewett from CEFAS answered, potentially this would be CEV. It is a real problem, also related to carp angling in the carp sport fisheries branch. Sven Bergmann adds, that losses in eels due to rhabdovirus, reovirus and picornavirus are also important, and need increased attention for the eel sector.

#### 2- Are diagnostic methods up to date, which ones to use?

There are various PCR tests possible to use. The Way et al. paper (2017) gives information on the possible and best diagnostic methods at various laboratories. The development of susceptible cell lines is now crucial to have tools for further development of diagnostic tests, such as serological tests, like ELISA, and for replicating the virus for pathogenicity and pathogenesis studies.

#### 3- Are prevention measures in place, and which specific vaccines are needed?

For development of a vaccine, for CEV we first need susceptible fish cell lines. Sven Bergmann added, that mononuclear cells are needed. Olga asked about the need for vaccines regarding perch viruses. Laurent answered, that there is a need for diagnostic tests, and to improve and increase egg

disinfection. Eggs of perch are sticky and are therefore difficult to disinfect, although egg disinfection is easy for pike perch eggs. Torsten Boutrup advised, we need to have a quality check to secure disinfection protocols. Uwe Fischer from FLI stated, for rhabdoviruses, DNA vaccination is a good option with other rhabdoviruses than from perch. Recombinant vaccines could also be an approach for CEV. Eva Lewisch from Vienna underlined the importance that, for prevention, we need to validate protocols and know how long the virus will persist in the environment.

## 4. What recommendations can be made for effective diagnosis and prevention of neglected viral disease problems in freshwater fish farming?

It is recommended to form a group of experts to sequence various pathogens, and set up an international platform. Jean-Christophe Avarre agrees on such a platform, and emphasized that with the advent of 3rd and 4<sup>th</sup> generations of sequencing, it is important to have a pipeline (see Bayliss et al. (2017) for illustration). Richard Paley from CEFAS will seek UK national funding. Olga Haenen hopes that the current publication provides necessary basic information on these emerging and neglected viruses to raise appropriate funding for future research initiatives.

#### **Overall Conclusions**

It was concluded that this workshop was very useful to address neglected viral diseases of freshwater fish farming, which should be strongly considered in the near future.

#### **Acknowledgements:**

We thank the organizers of the 18th EAFP International Conference, particularly Ivona Mladineo, the EAFP meeting secretary, and José Garcia, the EAFP president, for providing this important opportunity to address this workshop to a large international audience. We also thank all co-authors and colleagues collaborating to any presented work, and all participants for contributing to this successful event.

#### References:

- Adamek M, Jung-Schroers V, Hellmann J, Teitge F, Bergmann SM, Runge M, Kleingeld DW, Way K, Stone DM, Steinhagen D (2016) Concentration of carp edema virus (CEV) DNA in koi tissues affected by koi sleepy disease (KSD). *Dis Aquat Org* 119, 245-251.
- Adamek M, Oschilewski A, Wohlsen P, Jung-Schroers V, Teitge F, Dawson A, Gela D, Piackova V, Kocour M, Adamek J, Bergmann SM, Steinhagen D (2017) Experimental infections of different carp strains with the carp edema virus (CEV) give insights into the infection biology of the virus and indicate possible solutions to problems caused by koi sleepy disease (KSD) in carp aquaculture. Vet Res 48, 12 DOI 10.1186/s 13567-017-0416-7.
- Bayliss SC, Verner-Jeffreys DW, Bartie KL, Aanensen DM, Sheppard SK, Adams A, Feil EJ
  (2017) The Promise of Whole Genome Pathogen Sequencing for the Molecular Epidemiology of
  Emerging Aquaculture Pathogens. Front Microbiol 8, 121.
- Boitard PM, Baud M, Labrut S, de Boisseson C, Jamin M, Bigarré L (2016). First detection of Cyprinid Herpesvirus 2 (CyHV-2) in goldfish (Carassius auratus) in France. Journal of Fish Diseases 39, 673-680.
- Chang PH, Lee SH, Chiang HC and Jong MH (1999). Epizootic of herpes-like virus infection in goldfish, *Carassius auratus* in Taiwan. *Fish Pathology* **34**, 209–210.
- Daněk T, Kalous L, Veselý T, Krásová E, Reschová S, Rylková K, Kulich P, Petrtýl M, Pokorová D and Knytl M (2012). Massive mortality of Prussian carp *Carassius gibelio* in the upper Elbe basin associated with herpesviral hematopoietic necrosis (CyHV-2). *Diseases of Aquatic Organisms* 102, 87-95.

- Dannevig BH, Olesen NJ, Jentoft S, Kvellestad A, Taksdal T, Håstein T (2001). The first isolation
  of a rhabdovirus from perch (*Perca fluviatilis*) in Norway. *Bulletin of the European Association of*Fish Pathologists 21(5), 186-194.
- Di Cicco E, Ferguson HW, Schulze AD, Kaukinen KH, Li S, Vanderstichel R, Wessel Ø, Rimstad E, Gardner IA, Hammell KL, and Miller KM (2017) Heart and skeletal muscle inflammation (HSMI) disease diagnosed on a British Columbia salmon farm through a longitudinal farm study. *PLoS ONE* 12(2), e0171471. doi:10.1371/journal.pone.0171471.
- Dorson M, Torchy C, Chilmonczyk S, de Kinkelin P, Michel C (1984). A rhabdovirus pathogenic for perch, *Perca fluviatilis* L.: isolation and preliminary study. *Journal of Fish Diseases* **7**, 241-245.
- Eaton, W.D., Folkins, B., and Kent, M.L. (1994). Biochemical and histologic evidence of plasmacytoid leukemia and salmon leukemia virus (SLV) in wild-caught chinook salmon Oncorhynchus tshawytscha from british Columbia expressing plasmacytoid leukemia. Diseases of Aquatic Organisms 19, 147-151.
- European Commission (2015). European Commission Implementing Decision (EU) 2015/1554 of 11 September 2015 laying down rules for the application of Directive 2006/88/EC as regards requirements for surveillance and diagnostic methods (notified under document C(2015) 6188) (Text with EEA relevance). Official Journal of the European Union, 23<sup>rd</sup> September 2015, L247, 1-62. Online version: <a href="https://members.wto.org/crnattachments/2015/SPS/EEC/15">https://members.wto.org/crnattachments/2015/SPS/EEC/15</a> 3889 00 e.pdf Accessed 12 Dec 217.
- European Commission (2006). 2006/88/EC. Council Directive 2006/88/EC of 24 October 2006 on animal health requirements for aquaculture animals and products thereof, and on the prevention and control of certain diseases in aquatic animals. Online version: <a href="http://eur-lex.europa.eu/LexUriServ/LexUriServ.do?uri=OJ:L:2006:328:0014:0056:en:PDF">http://eur-lex.europa.eu/LexUriServ/LexUriServ.do?uri=OJ:L:2006:328:0014:0056:en:PDF</a>
- EURL (2015a) Report of Carp Edema Virus CEV Workshop 12th 13th January 2015 13 pp. http://www.eurl-fish.eu/Reports
- EURL (2015b) Report of Carp Edema Virus lunch meeting EAFP 2015. 9th September 2015 4pp. <a href="http://www.eurl-fish.eu/Reports">http://www.eurl-fish.eu/Reports</a>
- Frazer, J.K., Batchelor, L.A., Bradley, D.F., Brown, K.H., Dobrinski, K.P., Lee, C., and Trede, N.S. (2012). Genomic Amplification of an Endogenous Retrovirus in Zebrafish T-Cell Malignancies.
   Advances in Hematology, 12. [627920]. DOI: 10.1155/2012/627920
- Goodwin AE, Khoo L, LaPatra SE, Bonar C, Key DW, Garner M, Lee MV and Hanson L (2006).
   Goldfish hematopoietic necrosis herpesvirus (Cyprinid Herpesvirus 2) in the USA: molecular confirmation of isolates from diseased fish. *Journal of Aquatic Animal Health* 18, 11–18.
- Granzow H, Fichtner D, Schütze H, Lenk M, Dresenkamp B, Nieper H and Mettenleiter TC (2014). Isolation and partial characterization of a novel virus from different carp species suffering gill necrosis ultrastructure and morphogenesis. *Journal of Fish Diseases* 37, 559–569. doi: 10.1111/jfd.12150.
- Groff JM, LaPatra SE, Munn RJ and Zinkl JG (1998). A viral epizootic in cultured populations of juvenile goldfish due to a putative herpesvirus etiology. *Journal of Veterinary Diagnostic Investigation* 10, 375–378.
- Hauge H, Vendramin N, Taksdal T, Olsen AB, Wessel Ø, Mikkelsen SS, Alencar ALF, Olesen NJ, Dahle MK (2017) Infection experiments with novel Piscine orthoreovirus from rainbow trout (Oncorhynchus mykiss) in salmonids. PLoS ONE 12(7): e0180293. https://doi.org/10.1371/journal.pone.0180293

- Hedrick RP, Antonio DB, Munn RJ (1997). Poxvirus like agent associated with epizootic mortality in juvenile koi (*Cyprinus carpio*). FHS Newsletter **25** (3) 1-2.
- Hesami S, Yanong RPE, Shelley J, Goodwin A, Kelley KL, Spears S, Thompson PM, Groff J, Francis-Floyd R, Haenen OLM, Waltzek TB (2015). Carp edema virus disease (CEVD)/ Koi Sleepy Disease (KSD). University of Florida. University of Florida EDIS Publication FA18900. http://edis.ifas.ufl.edu/fa189.
- Ito T, Kurita J, Haenen O, 2017. Importation of CyHV-2 infected goldfish into the Netherlands. *Diseases of Aquatic Organisms* 126: 51–62. https://doi.org/10.3354/dao03157
- Ito T, Kurita J, Ozaki A, Sano M, Fukuda H and Ototake M (2013). Growth of Cyprinid herpesvirus 2 (CyHV-2) in cell culture and experimental infection of goldfish *Carassius auratus*. *Diseases of Aquatic Organisms* 105, 193-202.
- Ito T and Maeno Y (2014a). Susceptibility of Japanese Cyprininae fish species to cyprinid herpesvirus 2 (CyHV-2). *Veterinary Microbiology* **169**, 128–134.
- Ito T and Maeno Y (2014b). Effects of experimentally induced infections of goldfish *Carassius auratus* with cyprinid herpesvirus 2 (CyHV-2) at various water temperatures. *Diseases of Aquatic Organisms* **110**, 193–200.
- Ito T and Maeno Y (2015). Effect of booster shot and investigation of vaccination efficacy period against herpesviral haematopoietic necrosis (HVHN) in goldfish *Carassius auratus*. Veterinary Microbiology 175, 139–144.
- Ito T and Ototake M (2013). Vaccination against cyprinid herpesvirus 2 (CyHV-2) infection in goldfish Carassius auratus. Bulletin of the European Association of Fish Pathologists 33, 158–164.
- Jeffery KR, Bateman K, Bayley A, Feist SW, Hulland J, Longshaw C, Stone D, Woolford G and Way K (2007). Isolation of a cyprinid herpesvirus 2 from goldfish, *Carassius auratus* (L.), in the UK. *Journal of Fish Diseases* **30**, 649–656.
- Matras M, Borzym E, Stone D, Way K, StachniK M, Maj-Paluch J, Palusińska M, Reichert M (2016) Emergence of the carp edema virus (CEV) in Polish aquaculture phylogenetic analysis of the first cases. *Journal of Fish Diseases*, published online: 25 July 2016 | DOI: 10.1111/jfd.12518
- Naville, M., and Volff, J.-N. (2016). Endogenous Retroviruses in Fish Genomes: From Relics of Past Infections to Evolutionary Innovations? Frontiers in Microbiology 7.
- OIE (2017a). OIE-Listed diseases, infections and infestations in force in 2017. Aquatic Animal Health Code. World Organisation for Animal Health (OIE), Paris, <a href="http://www.oie.int/animal-health-in-the-world/oie-listed-diseases-2017/">http://www.oie.int/animal-health-in-the-world/oie-listed-diseases-2017/</a>, accessed 29 Nov 2017.
- OIE (2017b). Manual of Diagnostic Tests for Aquatic Animals. World Organisation for Animal Health (OIE), Paris, <a href="http://www.oie.int/international-standard-setting/aquatic-manual/access-online/">http://www.oie.int/international-standard-setting/aquatic-manual/access-online/</a>, accessed 12 Dec 2017.
- Ono S, Nagai A, Sugai N (1986). A histopathological study on juvenile colorcarp, Cyprinus carpio, showing edema. Fish Pathology 21, 167–175
- Quackenbush, S. (2016). "Retroviruses of Fish" in Aquaculture Virology, eds. F. Kibenge, S.B. & M.G. Godoy. (San Diego: Academic Press), 193-204.
- Rodger, H and Girons A (2008). Diseases and health management in perch farming. *Fish Veterinary Journal* **10**, 9-18.

- Ruane NM, Rodger HD, McCarthy LJ, Swords D, Dodge M, Kerr RC, Henshilwood K, Stone DM, (2014). Genetic diversity and associated pathology of rhabdovirus infections in farmed and wild perch *Perca fluviatilis* in Ireland. *Diseases of Aquatic Organisms* 112(2), 121-130.
- Stephens FJ, Raidal SR and Jones B (2004). Haematopoietic necrosis in a goldfish (*Carassius auratus*) associated with an agent morphologically similar to herpesvirus. *Australian Veterinary Journal* **82**, 167–169.
- Stocking, C., and Kozak, C.A. (2008). Murine endogenous retroviruses. *Cellular and molecular life sciences* **65**, 3383-3398.
- Wahli T, Bellec L, von Siebenthal B, Cabon J, Schmidt-Posthaus H, Morin T (2015). First isolation
  of a rhabdovirus from perch *Perca fluviatilis* in Switzerland. *Diseases of Aquatic Organisms*116(2), 93-101.
- Wang L, He J, Liang L, Zheng X, Jia P, Shi X, Lan W, Xie J, Liu H and Xu P (2012). Mass mortality caused by Cyprinid Herpesvirus 2 (CyHV-2) in Prussian carp (*Carassius gibelio*) in China. *Bulletin of the European Association of Fish Pathologists* **32**, 164-173.
- Way K., O. Haenen, D. Stone, M. Adamek, S.M. Bergmann, L. Bigarré, N. Diserens, M. El-Matbouli, M.C. Gjessing, V. Jung-Schroers, E. Leguay, M. Matras, N.J. Olesen, V. Panzarin, V. Piačková, A. Toffan, N. Vendramin, T. Veselý, T. Waltzek (2017). The emergence of carp edema virus (CEV) and its significance to European common carp and koi, *Cyprinus carpio. Diseases of Aquatic Organisms* 126, 155–166. https://doi.org/10.3354/dao03164
- Wolf, K (1988). Fish viruses and viral diseases. Cornell University Press, Ithaca, N.Y. ISBN 0-8014-1259-5, 476 pp.