

Neuroprotective role of polyphenols against oxidative stress-mediated neurodegeneration

Md. Sahab Uddin, Abdullah Al Mamun, Md. Tanvir Kabir, Jamil Ahmad, Philippe Jeandet, Md. Shahid Sarwar, Ghulam Md Ashraf, Lotfi Aleya

To cite this version:

Md. Sahab Uddin, Abdullah Al Mamun, Md. Tanvir Kabir, Jamil Ahmad, Philippe Jeandet, et al.. Neuroprotective role of polyphenols against oxidative stress-mediated neurodegeneration. European Journal of Pharmacology, 2020, 886, pp.173412. 10.1016/j.ejphar.2020.173412. hal-02916840

HAL Id: hal-02916840 <https://hal.science/hal-02916840v1>

Submitted on 22 Aug 2022

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

[Distributed under a Creative Commons Attribution - NonCommercial 4.0 International License](http://creativecommons.org/licenses/by-nc/4.0/)

Version of Record:<https://www.sciencedirect.com/science/article/pii/S0014299920305045> Manuscript_0e27a85fe6baad17f2bb6a64c80f554d

- 29
- 30

31 **Abstract**

32 Neurodegenerative diseases (NDs) are characterized by disorders with progressive deterioration 33 of the structure and/or function of neurons. Genetic mutations can lead to many NDs. 34 Nevertheless, neurodegeneration can also take place due to several biological processes. The 35 pathogenesis of several NDs including Alzheimer's (AD), Parkinson's (PD), and Huntington's 36 (HD) diseases are associated with oxidative stress (OS). In order to maintain the normal 37 functions of neurons, lower levels of reactive oxygen species (ROS) and reactive nitrogen 38 species (RNS) are important, since their increased levels can cause neuronal cell death. It has 39 been found that OS-mediated neurodegeneration involves a number of events including 40 mitochondrial dysfunction, Ca^{2+} overload, and excitotoxicity. A growing number of studies are 41 suggesting the benefit of using polyphenols for the treatment of neurodegenerative disorders. 42 Indeed, in order to treat most of the NDs, synthetic drugs are extensively used which are found to 43 exert side effects in the course of the treatment. There is mounting evidence that researchers have 44 identified several naturally-occurring chemical compounds in plants, which are used for the 45 management of NDs. Overall, polyphenolic phytochemicals are safer in nature and have 46 negligible side effects. In this article, we have focused on the potential efficacy of polyphenols 47 such as epigallocatechin-3-gallate, curcumin, resveratrol, quercetin and methylated polyphenols 48 berberine against the most common neurodegenerative disorders. 49 **Keywords:** Neuroprotection, Polyphenols, Oxidative stress, Neurodegeneration, Neurotrophins

50

- 51
- 52
- 53
- 54
- 55
- 56
- 57

58 **1. Introduction**

59 Over time, neurodegenerative diseases (NDs) have become a global threat. Presently, there is no 60 available treatment that can cure these NDs. It has been found that NDs are most common 61 among elderly people (Uddin et al., 2018). Globally, since there is a rise in the elderly 62 population, therefore the number of people who are suffering from NDs is also increasing. Older 63 people often develop NDs which ultimately can lead to death. Parkinson's disease (PD), 64 Alzheimer's disease (AD), Huntington's disease (HD), and amyotrophic lateral sclerosis (ALS) 65 are the commonly seen NDs among elderly people (Islam et al., 2018; Sahab Uddin and Shah 66 Amran, 2018; Uddin et al., 2017, 2019c). Indeed, movement disorders are linked with PD 67 patients (Kaur et al., 2020), whereas dementia is most common among patients with AD (Uddin 68 et al., 2020g). It is well-known that neurons of the brain are susceptible to oxidative stress (OS) 69 (Satoh et al., 1998) owing to high oxygen demand, low levels of glutathione, abundantly of 70 redox-active metals (iron, copper), and high levels of polyunsaturated fatty acids (Geon Ha Kim 71 et al., 2015). Furthermore, OS has the ability to disrupt the overall functions of the brain (Uddin 72 and Kabir, 2019). At molecular and cellular levels, a number of processes exist that provide 73 protection against oxidative toxicity. In the case of NDs, it has been found that disruptions take 74 place in the defensive mechanisms which can trigger neurological impairments. It was suggested 75 by various studies that OS may trigger cellular damage (Liguori et al., 2018), mitochondrial 76 dysfunction (Van Houten et al., 2018) and impairment of the deoxyribonucleic acid repair 77 system (Van Houten et al., 2018), all of these are crucial factors associated with multiple NDs 78 including AD, PD, and HD (Andersen, 2004; Uddin and Upaganlawar, 2019). Increased 79 generation of reactive oxygen species (ROS) inside mitochondria could lead to oxidative injury 80 against mitochondrial membranes, mitochondrial DNA, and mitochondrial proteins, ultimately 81 causing mitochondrial damage (Higgins et al., 2010). ROS affect the heme-comprising 82 cytochrome c oxidase I (complex IV) of the respiratory chain and trigger further injury to the 83 components of complexes I, II, and III (Lassmann, 2011). Oxidative injury in mitochondria plays 84 a crucial role in the release of cytochrome c inside the cytosol causing apoptosis. Furthermore, 85 the internal mitochondrial membrane displays higher penetrability to small molecules (Chen et 86 al., 2012). It has indeed been found that several events such as the overproduction of ROS, 87 mitochondrial dysfunction, as well as oxidative injury are accountable for neurodegeneration 88 (Hroudová et al., 2014).

90 An association between aging and buildup of OS has also been confirmed (Castelli et al., 2019; 91 Uddin et al., 2019a). Therefore, it is essential to elucidate the complete relationship between 92 cellular damage and OS in the case of the aging process and NDs (Harman, 2006). Indeed, it is 93 established that different cellular events such as mitochondrial dysfunction, an overload of Ca^{2+} , 94 excitotoxicity, and the induction of intracellular signaling mechanisms have significant 95 contributions in the molecular and cellular processes underlying OS-stimulated cell death 96 (Bossy-Wetzel et al., 2004; Liu et al., 2017). It has been found that the survival of neurons under 97 OS is promoted by several polyphenols (Numakawa et al., 2011; Potì et al., 2019; Uddin et al., 98 2020b). Polyphenols are involved with a wide range of neuroprotective activities such as 99 inhibiting neuroinflammation, providing protection to neurons against neurotoxin-induced injury, 100 and improving cognitive function, learning, and memory (Choi et al., 2012; Uttara et al., 2009; 101 Vauzour, 2012). Furthermore, phytosterols, for example, have important actions on the redox 102 state. A study reported that β-sitosterol reversed the impaired ratio of glutathione/oxidized 103 glutathione mediated via phorbol esters, in RAW 264.7 macrophage cultures. The findings of 104 this study can be linked with the reduction in the activity of catalase and the rise in the activities

106 Currently available drugs only provide symptomatic treatment to the individuals with NDs, 107 rather than preventing or curing neurodegeneration (Hossain et al., 2019; Kabir et al., 2019b, 108 2019a). Therefore, effective therapeutic agents are still lacking to treat NDs (Mizuno, 2014). In 109 this regard, it is essential to develop novel and more therapeutic approaches to fight against these 110 NDs. In recent times, there is growing popularity in the use of polyphenols to treat against 111 chronic diseases. Interestingly, polyphenols can also be used in association with other 112 phytochemicals leading to both additive and synergistic actions as shown, for example, in the 113 case of the combined effects of the polyphenol resveratrol, the monophenol tyrosol and the 114 phytosterol β-sitosterol on oxidative stress in RAW 264.7 macrophages (Vivancos and Moreno, 115 2008). Furthermore, a number of studies have been published regarding the use of dietary plant 116 constituents in the treatment of NDs (Debnath et al., 2017; Hossain et al., 2017; Makkar et al.,

105 of glutathione peroxidase and manganese superoxide dismutase (Vivancos and Moreno, 2005).

117 2020; Rahman et al., 2017; Venkatesan et al., 2015). In this article, we have focused on the

118 mechanism of actions and the significance of some dietary polyphenols in neurodegenerative 119 diseases.

120

121 **2. Oxidative stress in neurodegenerative diseases**

122 In terms of cellular functions, low levels of RNS and ROS have a physiological role including 123 neuronal plasticity (Kishida and Klann, 2007). Nevertheless, excessive RNS/ROS can cause 124 nitrosylation/oxidation of nucleic acids, proteins, and lipids, which can ultimately lead to 125 neuronal cell death (Meo et al., 2016). It has been observed that such damage can take place 126 either due to the decreased activity of non-enzymatic and enzymatic antioxidants or owing to the 127 excessive production of RNS/ROS. Therefore, an intricate balance is required between pro- and 128 antioxidant reactions so as to maintain normal neuronal functions.

129

130 It has been found that OS-mediated toxicity might be closely associated with the pathogenesis of

131 NDs including HD, PD, AD, and ALS (Andersen, 2004; Uddin et al., 2020e) as shown in **Fig. 1**.

132 In this regard, for instance, markers for DNA oxidation (8-hydroxy-2-deoxyoguanine [8-

133 OHdG]), lipid oxidation (4-hydroxy-2-nonenal [4-HNE]), and protein oxidation (protein

134 carbonyls and 3-nitro-tyrosine [3-NT]) are found to be increased in AD brains (Butterfield et al.,

135 2007). Amyloid beta (Aβ) buildup is regarded as an AD hallmark, which can trigger the

136 production of ROS including hydrogen peroxide (H_2O_2) in the presence of Cu^{2+} or Fe³⁺ (Huang

137 et al., 1999; Jiang et al., 2007; Xudong Huang et al., 1999). Unfortunately, a progressive and

138 selective loss of dopamine (DA) neurons takes place in the substantia nigra pars compacta in the

139 brains of individuals with PD. In addition to this, the levels of 8-OHdG, 3-NT, protein carbonyls,

140 and 4-HNE are found to be elevated, while the level of glutathione (GSH, a major intracellular

141 antioxidant) is reduced (Zhou et al., 2008). It has been observed that 4-HNE has the ability to

142 covalently bind with the central protein in PD pathogenesis, α -synuclein (α -Syn), which

143 eventually leads to neurotoxic actions on GABAergic and DAergic neuronal cultures (Qin et al.,

144 2007). On the other hand, increased levels of 8-OHdG, malondialdehyde (a marker for lipid

145 oxidation), lipofuscin (a product of unsaturated fatty acid peroxidation), and 3-NT have been

146 observed in case of HD brains (Stack et al., 2008). Decreased GSH level has been observed in

147 cultured neurons obtained from mice that express mutant huntingtin protein (Htt140Q/140Q) (Li 148 et al., 2010).

149

150 **- Insert Figure 1-**

151

152 Oxidative toxicity has also been found to play a pivotal role in reperfusion injury/cerebral 153 ischemia (Uddin et al., 2019b; Zaplatic et al., 2019). There are limited areas of brain and types of 154 neurons that are susceptible to ischemia. Cerebral blood flow is regarded as the primary reason 155 behind the aforesaid phenomenon since this flow is highly spatiotemporally regulated (Cremer 156 and Seville, 1983). The phenomena outlined before may also reveal why certain neuron types of 157 different brain areas are affected in each ND. It has been suggested by copious evidence that 158 aggregation of OS-dependent damage can take place in the course of normal aging, which can 159 further result in substantial impairment in cognitive function (Bishop et al., 2010; Harman, 2006; 160 Jakaria et al., 2019).

161

162 In many NDs, cell death primarily takes place due to apoptosis (a form of programmed cell

163 death). A number of processes usually contribute to apoptosis, such as OS, endoplasmic

164 reticulum (ER) stress, mitochondrial dysfunction, Ca²⁺ overload, and excitotoxicity (Bossy-

165 Wetzel et al., 2004) as given in **Fig. 2**. It is known that via the electron transport chain (ETC),

166 mitochondria generate a lower level of ROS in cellular respiration. There are five protein

167 complexes (I–V) that comprise the ETC. It has been found that excess ROS production can occur

168 due to the disruption of ETC (Nicholls, 2008). Various studies have indeed revealed that

169 mitochondrial dysfunction (including the disrupted activity of the ETC) may be involved in AD

170 animal models and patients with HD (Gil and Rego, 2008), PD (Henchcliffe and Beal, 2008),

171 and AD (Adam-Vizi and Starkov, 2010).

172

173 Furthermore, mitochondria affect the homeostasis of Ca^{2+} by sequestering superfluous cytosolic

 Ca^{2+} within their matrix. However, neurodegeneration may occur through an uncontrolled Ca^{2+}

175 loading. According to the study of Oliveira et al. (Oliveira et al., 2007), disrupted Ca^{2+}

176 homeostasis was observed in striatal mitochondria of Hdh150 knock-in HD mice. It has been

177 found that a lack of phosphatase and tensin homolog deleted on chromosome 10-mediated

- 178 putative kinase 1, causes mitochondrial aggregation of Ca^{2+} in cultured neurons (Gandhi et al.,
- 179 2009). Moreover, ER controls intracellular levels of Ca^{2+} via ryanodine receptors as well as
- 180 inositol-1,4,5-triphosphate receptors (InsP3Rs). Remarkably, presenilin (*PSEN*) 1 and 2 play a
- 181 pivotal role in AD pathology, act as a passive ER Ca^{2+} channel to keep up the equilibrium state
- of Ca2+ 182 levels that disrupts by mutant *PSEN2*-N141I and *PSEN1*-M146V (Tu et al., 2006; Zhang
- 183 et al., 2010c). These *PSEN* mutants increased the gating of InsP3Rs activity, resulting in the
- 184 aggregation of Aβ (Cheung et al., 2008). It has also been observed that Aβ-comprising senile
- 185 plaques lead to Ca^{2+} overload (Kuchibhotla et al., 2008). Collectively, it is assumed that mutant
- *PSEN* as well as A β lead to the disturbance of Ca^{2+} homeostasis that might result in
- 187 mitochondrial dysfunction causing neurodegeneration (Adam-Vizi and Starkov, 2010).
- 188

189 **- Insert Figure 2-**

190

191 **3. Polyphenols as antioxidative factors**

192 Since there is an involvement of OS in the pathogenesis of NDs, thus a crucial therapeutic 193 approach can delay or block the OS through elevating the activity of endogenous antioxidants 194 and/or inhibiting the generation of ROS. However, it is still elusive whether OS is a consequence 195 or cause of ND (Andersen, 2004). It has been suggested by evidence that OS directly starts and 196 progresses to neuronal cell death. Nevertheless, there is a chance that OS accumulation is easily 197 stimulated in neurons that are damaged by other insults. Death signaling pathways can be 198 activated in the apoptotic process through a number of cellular processes such as excitotoxicity, 199 $Ca²⁺$ overload, and mitochondrial dysfunction. In fact, cell viability may get affected by such 200 negative feedback loops. The aforesaid processes can take place in parallel and have an additive 201 action in case of stimulation of cell death. Henceforth, along with the blockage of OS 202 accumulation, activating survival signaling and suppressing death-signaling cascades may also 203 prove effective.

204

205 Some studies revealed that one or more mechanisms related to adaptive cellular stress responses

206 might get activated by lower amounts of dietary polyphenols (Mandel et al., 2008; Schroeter et

- 207 al., 2001, 2007; Vauzour et al., 2007). Activation of such hormetic signaling pathways in
- 208 neurons can lead to the generation of various types of cytoprotective proteins such as anti-

209 apoptotic proteins, antioxidant and phase II enzymes, protein chaperones, and neurotrophic 210 factors (Calabrese, 2008; Mattson and Cheng, 2006). There is a growing interest in the hermetic 211 actions in neurons that involve nuclear factor erythroid 2-related factor 2 (Nrf2). It has indeed 212 been reported that Nrf2 has the ability to bind with antioxidant-responsive element (ARE) with 213 high affinity and has a significant contribution in the up-regulation of genes that are involved in 214 the modulation of the cellular redox status and the protection of the cells from OS (Kraft et al., 215 2004; Lee and Johnson, 2004; Uddin et al., 2020h). Furthermore, Nrf2 can also interact with a 216 cytosolic repressor protein Keap1 (Kelch ECH associating protein) limiting Nrf2-induced gene 217 expression, under basal conditions (Itoh et al., 1999). Upon exposure to oxidative damage, Nrf2 218 is released in cells from Keap1 and translocates to the nucleus, where it causes activation of 219 ARE-dependent transcription of phase II and antioxidant defense enzymes including heme 220 oxygenase-1 (HO-1), glutathione peroxidase (GPx), and glutathione-S-transferase (GST)

221 (Nguyen et al., 2003).

222 A study by Calabrese et al. (Calabrese, 2008) demonstrated that most polyphenols responded in a 223 bell-shaped dose-response mode, showed cellular toxicity at high concentrations whereas 224 triggering light chemical stress at low doses with the initiation of physiological hormesis in cells, 225 causing increased expression of protective genes including those initiated by Nrf2. Some studies 226 revealed that resveratrol protected PC12 cells from H_2O_2 -induced oxidative stress (Chen et al., 227 2005b) and attenuated cerebral ischemic injury in an animal model (Ren et al., 2011) through the 228 initiation of Nfr2 as well as the upregulation of HO-1. Numerous upstream signaling pathways 229 might activate Nrf2 either independently, or in a combined way. These have specific functions 230 on several lipid and protein kinase signaling pathways including the mitogen-activated protein 231 kinase (MAPK) and phosphoinositide 3-kinase (PI3k)/protein kinase B (Akt) cascades that 232 control prosurvival transcription factors as well as the expression of the gene (Williams et al., 233 2004). Furthermore, activation of the extracellular signal-regulated kinase (ERK) plays an 234 essential role in the initiation of cyclic adenosine monophosphate (cAMP)-response element-235 binding protein (CREB), a transcription factor that leads to the initiation of long-term alterations 236 in memory and synaptic plasticity (Bourtchuladze et al., 1994; Impey et al., 1998). The 237 activation of CREB also controls the expression of many essential genes, such as brain-derived 238 neurotrophic factor (BDNF), consequently playing an important role in regulating synaptic 239 activities and neuronal survival in the adult central nervous system (CNS) (Finkbeiner, 2000;

240 Tully et al., 2003). Besides, another polyphenol, fisetin, is commonly observed in strawberries,

- 241 which has been revealed to enhance long-term potentiation as well as to improve object
- 242 recognition in an animal model by the initiation of CREB and ERK (Kong et al., 2000).
- 243

244 **4. Neuroprotective potential of polyphenols against neurodegenerative events**

245 NDs including multiple sclerosis (MS), HD, PD, and AD fall under the category of NDs along 246 with neurotraumatic disorders (Helman and Murphy, 2016; Winner et al., 2011). Environmental 247 and genetic factors also have a significant contribution to the progression of NDs.

248 Neuroinflammation, OS, and buildup of certain aggregated proteins are the common pathological

249 characteristics of NDs (Farooqui, 2012; Uddin et al., 2020d, 2020f). These stimuli are embedded

250 in the plasma membrane via extracellular receptors. Chronic inflammation is directly related to

251 oxidative/nitrosative stress (Mitjavila and Moreno, 2012). Lipoxygenase (LOX), as well as

252 cyclooxygenase (COX) cascades, are upregulated in chronic and age-related brain pathologies

- 253 (Bishnoi et al., 2005; Vivancos and Moreno, 2002). The metabolism of polyunsaturated fatty
- 254 acids (PUFAs) are impaired by COX. Furthermore, COX-1, COX-2 are accountable for the

255 generation of many eicosanoids, as well as by LOXs that catalyze the addition of oxygen to

256 docosahexaenoic linolenic, and arachidonic acids and other PUFA playing a crucial role in the

257 generation of bioactive lipids, which considerably affects the event of neurodegeneration. Amid

258 numerous isoforms, 12/15-LOX and 5-LOX are particularly significant in neuroinflammation or

259 neurodegeneration (Czapski et al., 2016).

260 In brain, increased levels of PUFAs including arachidonic acid and docosahexaenoic acid have

261 been observed as compared to other organs. Since these fatty acids are extremely unsaturated,

262 they are thus prone to OS-mediated lipid peroxidation, which has been found to be one of the

263 major consequences of free radical-induced injury (Montine et al., 2002). Breakdown of

264 neuronal membrane-associated PUFAs can lead to the release of increased concentrations of

265 reactive electrophilic aldehydes, which can further covalently bind proteins to generate multiple

266 adducts (Tramutola et al., 2017) including 4-HNE. Various studies have confirmed elevated

- 267 levels of nitrosative stress and lipid peroxidation in the brains of individuals with AD and PD
- 268 (Butterfield et al., 2006; Hardas et al., 2013; Yoritaka et al., 1996). Numerous studies have
- 269 revealed an association between OS and protein homeostasis. Proteostasis network is essential

270 for the degradation of proteins that are irreversibly oxidized (Jung et al., 2009; Sahab Uddin and 271 Ashraf, 2020; Tramutola et al., 2017). During aging, the ability of the cells to preserve 272 proteostasis is reduced, which can make the organism vulnerable towards neurodegeneration 273 (Uddin et al., 2020c). Furthermore, the main outcome of protein oxidation is the generation of 274 large protein aggregates, whose buildup develops toxicity in cells. Insoluble aggregates can be 275 generated due to covalent cross-links among peptide chains, as in the case of the mutant 276 superoxide dismutase 1 in ALS, α-synuclein in PD, and neurofibrillary tangles (NFTs) and \overrightarrow{AB} in 277 AD (Kabir et al., 2020; Uddin et al., 2020a; Uddin and Kabir, 2019).

278

279 Furthermore, neurotrophins (NTs) can cause cellular stress and stimulate release and generation 280 of the BDNF, and activation of the tropomyosin-related kinase B (TrkB) receptor family and 281 other downstream protein kinases (**Fig. 3**) (Barbacid, 1994; Marini et al., 2004). In the brain, 282 NTs play vital roles in the regeneration, survival, and maintenance of specific neuronal 283 populations (Venkatesan et al., 2015). Moreover, the expression of NT has a strong association 284 with the occurrence of ND (Cho et al., 2013). On the other hand, mammalian NTs including NT-285 3, BDNF, nerve growth factor (NGF), and NT-4/5 play a significant role in the maintenance of 286 neurons of both central and peripheral nervous systems (Reichardt, 2006). Indeed, NTs have 287 been identified as promoters of neuron survival. It was also suggested that the levels of NT drop 288 drastically in the case of NDs (Dawbarn and Allen, 2003). Henceforth, NTs are regarded as vital 289 targets for polyphenols that exert effects against ND. Through the binding and activation of Trk 290 receptors (located in the plasma membrane), NTs avert neuronal degeneration as depicted in **Fig.** 291 **3**.

292 **- Insert Figure 3-**

293 In addition, the aforesaid binding of NTs with Trk receptors ultimately helps in the establishment 294 of a growth-encouraging microenvironment for neurons (Huang and Reichardt, 2001). Following

295 this, several intracellular signaling processes occur including the ERK pathway and PI3k/Akt

296 cascade which stimulate cell survival and help in protecting the neurons against

297 neurodegeneration. Besides activation of the signaling pathway, expression of B-cell lymphoma

298 2 (Bcl-2) genes (an inhibitor of intracellular apoptosis) can also be promoted by NT. On the

299 other hand, gradual degeneration of neurons has been observed in the absence of binding of NTs

300 to the tropomyosin-related kinase (Trk) receptors. Therefore, the administration of NTs might be 301 an effective way to combat neurodegeneration and ultimately overcome this disorder. 302 Nevertheless, crossing the blood-brain barrier (BBB) is still a big challenge to treat CNS 303 disorders. In order to reverse NT loss, an enhancer or modulator (i.e. therapeutic agent) 304 particularly targeting the Trk receptor might be a good candidate (Reichardt, 2006). Moreover, 305 due to the promising findings of several studies, there is a rise in the intake of certain plant 306 ingredients for the treatment of NDs (Essa et al., 2012; Howes and Houghton, 2012). 307 Polyphenols are most commonly found in vegetables, fruits, and natural products. Furthermore, 308 polyphenols contain several hydroxyl groups on aromatic rings (Tresserra-Rimbau et al., 2018; 309 Vauzour, 2012). Interestingly, *in vitro* studies have revealed that polyphenols exert their 310 beneficial effects by directly scavenging free radicals and nitrogen species (Halliwell, 2006; 311 Pannala et al., 1997; Russo et al., 2000; Visioli et al., 1998). Nonetheless, polyphenols are 312 limitedly bioavailable and get extensively metabolized. These factors attenuate their antioxidant 313 properties making it also difficult to estimate the *in vivo* concentrations at which they will exert 314 them (Williams et al., 2004).

315

316 **4.1. Epigallocatechin-3-gallate**

317 EGCG (**Fig. 4**) is the most common polyphenol found in tea. Because of its potential to delay 318 neurodegeneration, this plant compound has drawn a lot of attention in the last few decades. 319 Interestingly, it has been observed that the occurrence of NDs is inversely connected with the 320 consumption of tea (Pervin et al., 2018). In AD models, several experiments have been carried 321 out where EGCG was provided. Many studies have revealed that the administration of a drug 322 named D-gal in AD models played a significant role in the reduction of Aβ (Chan et al., 2016; 323 Walker et al., 2015; Wobst et al., 2015). A study by Liu et al., showed that the administration of 324 EGCG decreased the level of β- and γ-secretases [i.e. amyloid precursor protein (APP) cleaving 325 enzyme)] via suppression of the activity of nuclear factor kappa B (NF-κβ) and ERK, which 326 ultimately leads to the prevention of neuronal cell death (Liu et al., 2014b). It was also found that 327 EGCG administration resulted in a reduced level of A β due to the α -secretase-mediated APP 328 cleavage (Smith et al., 2010). On the other hand, Choi et al., (Choi et al., 2002) recommended

329 that consumption of three or more cups of tea containing high levels of EGCG led to a reduced 330 risk of PD development.

331 It has been observed that there is a substantial loss of tyrosine hydroxylase (TH) positive cells in

332 PD. Koh et al., (Koh et al., 2003) demonstrated that the concurrent consumption of EGCG and

333 tea could help to prevent the loss of these cells in the substantia nigra. Collectively, these

334 findings denote that the use of EGCG can be useful in the treatment of ND. Furthermore, by

335 preventing the uptake of DA and by displaying neuroprotective effects, EGCG can inhibit

336 neuronal cell death. EGCG exerts this aforesaid function through its activity on the catechol-*O*-

337 methyltransferase (COMT) enzyme that blocks DA metabolism. α-Syn accumulation is also

338 observed in ND. It is supposed that EGCG regulates the proteolytic cleavage of α-Syn converting

339 it into a less toxic form (Koh et al., 2003; Liu et al., 2014b).

340 EGCG exerts neuroprotective effects on oxidative-stress-mediated apoptosis in neuronal-

341 differentiated PC12 cells by downstream signaling via mitochondrial damage, poly(ADP-ribose)

342 polymerase, and caspase (CASP)-3 as well as upstream signaling via the PI3K/Akt, and GSK-3

343 cascades (Koh et al., 2003). Interestingly, EGCG also counteracts radicals and exerts antioxidant

344 effects, since EGCG has a 3′, 4′, 5-trihydroxy B ring containing a member of the catechol family

345 (Kalaiselvi et al., 2013). The Nrf2/ARE pathway outlined above plays an important role in

346 mediating the antioxidant effects of EGCG (Han et al., 2014). In addition, catalase and

347 superoxide dismutase (SOD) are the two radical scavenging enzymes that are typically activated

348 by EGCG (Kalaiselvi et al., 2013; Srividhya et al., 2008).

349 Several properties of EGCG such as its BBB penetrating and metal chelating features make this

350 plant compound a promising solution for neurodegenerative events. Numerous studies performed

351 in animal models demonstrated the BBB permeability of EGCG and showed that it can prevent

352 cognitive dysfunctions (Pervin et al., 2017; Wei et al., 2019) (**Table 1**). It has already been

353 observed that EGCG restores DA neurons by preventing the aggregation of iron-dependent α-

354 Syn (Bieschke et al., 2010). A number of clinical studies have confirmed that EGCG suppresses

355 the activity of cytochrome p450 and that this plant compound is also accountable for lipid

- 356 peroxidation *in vitro* as well as *in vivo* (Mangialasche et al., 2010; Schneider et al., 2014). In
- 357 their work, Castellano-González et al., (Castellano-González et al., 2016) observed that EGCG
- 358 mediates oxidative phosphorylation via activating cytochrome c oxidase (CcO) in human

359 cultured astrocytes and neurons. Furthermore, they also noticed that there is an increased activity 360 of CcO in 10 μM EGCG-treated neurons as compared to the controls. However, the increased 361 activity of CcO recorded in astrocytes was not statistically significant. EGCG treatments up to 50 362 μM did not elevate the generation of ROS in both human cultured neurons and astrocytes, while 363 EGCG at 100 μM elevated the generation of ROS in neurons. Over generation of ROS is found 364 to be involved in the stimulation of apoptosis. The presence of CcO dysfunction has also been 365 detected in individuals with NDs including AD (Castellano-González et al., 2016). Thus it is 366 likely that treatment with EGCG may restore the functions of mitochondria and avert the 367 following synaptic functional loss.

368 EGCG exerts neuroprotective effects in SH-SY5Y cells by playing a significant role in the 369 intracellular signaling mechanism of protein kinase C (PKC). It has been observed that inhibitors 370 of PKC also take part in the neuroprotective mechanisms of EGCG. On the other hand, the 371 neurotoxin, 6-hydroxydopamine (6-OHDA), was reported to be inhibited by EGCG through its 372 activation of PKC phosphorylation (Koh et al., 2003). It has been found that EGCG not only 373 activates the PKC signaling pathway, but also other pathways including the MAPK, PI3k/Akt, 374 and pathways regulating calcium influxes (Chen et al., 2017; Ortiz-López et al., 2016). 375 According to the study by Levites et al., (Levites et al., 2002) EGCG displayed a neuroprotective 376 effect by restoring the reduced ERK1/2 and PKC phosphorylation levels induced by 6-OHDA, 377 and by modulating of antiapoptotic-related genes. Furthermore, the study of Ding et al., (Ding et 378 al., 2017) has demonstrated that EGCG was capable to efficiently suppress sevoflurane-induced 379 neurodegeneration as well as enhance memory and learning retention of mice through the 380 activation of CREB/BDNF/TrkB-PI3K/Akt signaling (**Table 2**).

381

382 **- Insert Figure 4-**

383

384 **4.2. Berberine**

385 Berberine (**Fig. 4**), a naturally-occurring isoquinoline alkaloid, can be isolated from the stems,

- 386 rhizome, roots, and bark of plants of the genus *Berberis* and also from plants including *Hydrastis*
- 387 *canadensis* and *Coptis chinensis* (Koppen et al., 2017). Berberine also crosses BBB, exerts a
- 388 direct action on neurons and accumulates in the hippocampus in several brain diseases (Wang et

389 al., 2005). According to the study of Zhang et al., (Zhang et al., 2012d) berberine improves BBB 390 permeability and displays its neuroprotective effects through the activation of Akt/glycogen 391 synthase kinase (GSK) signaling, downregulation of NF-κB nuclear transposition, and 392 upregulation of phosphorylated cyclic AMP response element binding protein (pCREB) (**Table** 393 **1**). On the other hand, berberine has the ability to stop the activity of a number of enzymes that 394 are associated with AD progression (Hsu et al., 2013a). According to the study of Hsu et al., 395 (Hsu et al., 2013a) berberine demonstrates neuroprotective effects by activating Akt/GSK-396 3β/Nrf2-mediated regulation, triggers NGF and BDNF secretion and suppresses tumor necrosis 397 factor alpha (TNF-α), cyclooxygenase-2 (COX-2), interleukin 1β (IL-1β), and nuclear factor-398 kappa B (NF- κ B) levels (**Table 2**). A number of *in vitro* studies have revealed that H₂O₂ and 399 glutamate can induce neurotoxicity. In addition, it was found that cobalt chloride can facilitate a 400 decreased oxygen environment that connected with neurotoxicity, as well. It was also established 401 in these *in vitro* studies that berberine has the ability to protect neuronal cells from H_2O_2 , 402 glutamate and cobalt chloride-induced neurotoxicity (Cui et al., 2009; Hsu et al., 2013a; Simões 403 Pires et al., 2014). Furthermore, berberine is useful in the treatment of cobalt chloride-induced 404 hypoxia. In this regard, berberine plays a key role as a scavenger of ROS (i.e. which can inhibit 405 multiple apoptosis-promoting agents) produced upon hypoxic conditions, therefore providing 406 neuroprotective effects (Zhang et al., 2012c). Nevertheless, a study by Kwon et al. (Kwon et al., 407 2010) showed that berberine could exert substantial side effects including a fall in the level of 408 DA neurons as a result of the cytotoxicity of 6-hydroxydopamine. 409 Berberine exerts its neuroprotective properties through activating the PI3K/Akt/Nrf2 pathway by 410 scavenging radicals. Additionally, berberine exerts anti-apoptotic effects by decreasing the

411 expression of CASP-1 and -3, Bax along with Bcl-2 upregulation (Asai et al., 2007). Another

412 study by Hsu et al., (Hsu et al., 2012) revealed that the administration of berberine can prevent

413 H2O2-stimulated neurotoxicity by decreasing the expression of cyclin D1, CASP, p53 and

414 elevating the expression of Bcl-2 associated agonist of cell death (BAD). It has been observed

415 that at a nanomolar concentration, berberine reduced OS and promoted cell survival via the

416 downregulation of multiple factors including CASP, Bax, and cytochrome c (Maleki et al.,

417 2018).

- 418 Zhang et al. (Zhang et al., 2009) demonstrated that the administration of berberine reduced the
- 419 decrease of choline acetyltransferase and SOD activities in an aluminum-induced rat model of
- 420 NDs. Interestingly, it has been observed that berberine provides protection against ischemic
- 421 stroke by decreasing OS (Kim et al., 2014). Berberine administration can restore memory
- 422 function and BDNF levels maintains cholinergic enzyme activity and reduces the expression of
- 423 inflammatory molecules (Durairajan et al., 2012).
- 424 On the other hand, numerous studies have revealed that chronic administration of berberine can 425 lead to loss of dopaminergic substantia nigra neurons (Kim et al., 2014; Kwon et al., 2010; Shin 426 et al., 2013). Individuals receiving chronic therapy with levodopa might require the use of 427 neurotoxic isoquinoline agents along with berberine, though they should be monitored for 428 adverse reactions (Shin et al., 2013). Berberine exerts anti-apoptotic effects by reducing the 429 hypoxia-inducible factor 1α and p53 expression, which are both regarded as the major processes 430 in order to avert apoptosis via berberine administration (Zhang et al., 2012c). Moreover, 431 berberine is also believed to reduce ischemic stroke-caused damages via reducing the activity of 432 gelatinase and by decreasing the expression of the matrix metalloproteinase-9, laminin, and that 433 of neuronal nuclear protein (Hong et al., 2012). As berberine was reported to solubilize Aβ 434 plaques in mouse models, this compound can thus be useful in the AD treatment (Asai et al., 435 2007). It has also been suggested that berberine binds with the N-methyl-D-aspartate (NMDA) 436 receptor in order to exert its neuroprotective effects. In a caspase-independent manner, berberine 437 exceeding 1 μM decreased the viability of neurons through early changes of mitochondrial 438 morphology and function. According to the study of Kysenius et al., (Kysenius et al., 2014) 439 MK-801 and memantine played a significant role in the inhibition of NMDA receptors, which
- 440 totally blocked berberine-mediated neurotoxicity.
- 441 There are still some concerns related to the safety profile of berberine, since some studies 442 reported that bradycardia and exacerbation of neurodegeneration can take place owing to the use 443 of berberine. In the study of Kysenius et al. (Kysenius et al., 2014), it was revealed that at 444 micromolar level, berberine can cause rapid mitochondria-dependent toxicity in primary neurons 445 and this toxicity was characterized by elevated OS, mitochondrial swelling, reduced 446 mitochondrial membrane potential and depleted ATP content. Activation of caspase-3 is not 447 induced by berberine and the resultant neurotoxicity stays unchanged due to the treatment with

448 pan-caspase inhibitor. MK-801 and memantine inhibited the NMDA receptors, which eventually 449 leads to the complete blockage of berberine-mediated neurotoxicity. In addition, subtoxic 450 nanomolar berberine concentrations were sufficient to cause sensitization of neurons to rotenone 451 injury and glutamate excitotoxicity. Nevertheless, more studies are required on the safety profile

452 of berberine, mainly because of its propensity to gather in the CNS and due to the potential risk

453 of neurotoxicity linked to its increasing bioavailability.

454 **-Insert Table 1-**

455 **-Insert Table 2-**

456

457 **4.3. Curcumin**

458 One of the significant constituents of turmeric is curcumin (**Fig. 4**). A proposed mechanism of 459 action for curcumin in AD is that it can bind to Aβ by suppressing NF-κβ, conferring this 460 polyphenol the potential to reduce AD pathogenesis (Nam et al., 2014). A study by Kim et al., 461 (Kim et al., 2005) reported that out of about 214 compounds possessing anti-oxidant effects, 462 curcumin exhibited the highest affinity towards Aβ. In aged mice, upon administration of 463 curcumin, cognitive deficits were ameliorated because of the rise in the levels of BDNF and 464 CREB (Nam et al., 2014). Furthermore, in a PD model, it has been observed that curcumin can 465 aid in the regeneration of neurons via activating Trk/PI3K signaling pathways which eventually 466 increase the levels of BDNF (**Table 2**) (Yang et al., 2014). A study by Hoppe et al., (Hoppe et 467 al., 2013) exposed that nanoparticles of curcumin can improve cognitive deficit through 468 recovering the levels of BDNF by activating Akt/GSK-3β signaling pathways. Indeed, curcumin 469 plays a crucial role in several aspects of NDs. Therefore, curcumin can be used as a cheap and 470 safe preventive measure in NDs.

471 Current studies are mainly focusing on the discovery of curcumin derivatives which possess 472 more lipophilic properties helping them to penetrate BBB more easily and thus providing them 473 more affinity towards Aβ (Mourtas et al., 2011). A study of Garcia‐Alloza et al., (Garcia-Alloza 474 et al., 2007) demonstrated that curcumin could permeate the BBB and labeled senile plaques and 475 ultimately ameliorates existing plaques in AD (**Table 1**). Present studies carried out in cell 476 cultures and *in vivo* have also shown that the formation of Aβ plaques can be reduced by 477 curcumin (Wang et al., 2018). Zhang et al. reported that the activity of curcumin is not only

478 limited to disruption of Aβ levels but also averts APP maturation in mouse neurons (Zhang et al.,

479 2010a). Indeed, NF- κ B inhibition is the main mechanism by which curcumin protects from A β

480 plaques. Curcumin can provide protection against the PD through α -Syn destabilization, as well

481 (Liu et al., 2014a).

482 Studies have revealed that Aβ plaques in AD individuals possess iron and the presence of this 483 metal is also accountable for the production of free radicals, lipid peroxidation, advanced 484 glycation end-products (AGEs), oxidation of DNA and proteins. Interestingly, *in vitro* studies 485 have revealed that curcumin exhibits an affinity for iron (Liu et al., 2014a). Many works have 486 reported that 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine (MPTP) and 6-OHDA can be used to 487 generate PD models and curcumin was shown to exert neuroprotective effects in these models 488 through ROS inhibition (Yang et al., 2014; Zbarsky et al., 2005). According to the study of Yu et 489 al., curcumin suppresses the c-Jun N-terminal kinase (JNK) pathway in MPTP- and 1-methyl-4- 490 phenylpyridinium (MPP)-induced neurotoxicity in mice models (Yu et al., 2010). It has also 491 been found that curcumin ameliorates locomotion and lifespan in different fruit fly AD models 492 (Caesar et al., 2012).

493 In mammals, prepared Aβ-expressing mice were administered with curcumin (Lim et al., 2001), 494 resulting in curcumin-improved NDs by scavenging ROS, disrupting Aβ plaques, and showing 495 anti-apoptotic and anti-inflammatory properties (Hoppe et al., 2013). In a recent animal study, 496 administration of curcumin at a dose of 500 ppm inhibited the aggregation of Huntingtin protein 497 (Frautschy et al.). Curcumin has also shown various beneficial effects *in vitro*, for example 498 enhancing the lifespan in *Caenorhabditis elegans* and Drosophila, though no improvement in 499 mice was observed (Liao et al., 2011; Strong et al., 2013). Poor bioavailability of curcumin is 500 one of the main issues associated with it, however, it can be managed by conjugating curcumin 501 with lipophilic compounds or by changing its chemical structure (Purpura et al., 2018). 502

503 **4.4. Resveratrol**

504 Resveratrol (**Fig. 4**) is a type of polyphenol found in peanuts, grapes, wine, and tea displaying 505 numerous biological activities (Jeandet et al., 2018, 2019; Storniolo and Moreno, 2019). A lot of 506 *in vitro* as well as *in vivo* investigations have shown the potential neuroprotective properties of 507 resveratrol. For example, copious studies have revealed that resveratrol provides cytoprotective

508 activity in cells exposed to Aβ and/or Aβ -metal complex through sirtuin-3 (SIRT3)-mediated 509 mechanisms (Granzotto and Zatta, 2011; Yan et al., 2018). In a study of mice with cerebral Aβ 510 deposition, oral administration of resveratrol decreased the activation of microglia associated 511 with the formation of cortical Aβ plaques (Capiralla et al., 2012). Moreover, long-term 512 consumption of dietary resveratrol has shown a neuroprotective role of this compound together 513 with reduced cognitive impairment, tau hyperphosphorylation, and amyloid burden in age-related 514 AD (SAMP8) mice (Porquet et al., 2013). In addition, increasing scientific evidence suggests 515 that resveratrol can provide beneficial effects in cells or animals representing PD. For example, 516 in primary midbrain neuron-glia cultures of rat, resveratrol has been shown to protect against 517 lipopolysaccharide-induced neurotoxic dopaminergic neurons in a time- and dose-dependent 518 way, by inhibiting the activation of microglia and the resultant reduction in the release of pro-519 inflammatory factors (Zhang et al., 2010b). In a clinical study, resveratrol also improved 520 pathological changes and motor deficits in MPTP-treated mice by activating Sirtuin-1 (SIRT1) 521 and the resultant light chain 3 (LC3) deacetylation-mediated autophagic degradation of α-Syn 522 (Guo et al., 2016). All of the above investigations have proposed that resveratrol is likely to be a 523 possible therapeutic or prophylactic solution for NDs.

524

525 The anti-oxidative activities of resveratrol have comprehensively been reported since the past 526 years (Chen et al., 2005a; de Sá Coutinho et al., 2018; Wang et al., 2001; Zhang et al., 2008), but 527 the recent popular discovery of the activation of vitagenes and SIRT1 by resveratrol, has caught 528 fire (Yazir et al., 2015). Timmers et al., (Timmers et al., 2011) demonstrated that both resveratrol 529 and energy restriction work in a similar way in increasing life expectancy. The possible 530 mechanism of resveratrol includes the activation of peroxisome proliferator-activated receptor 531 (PPAR) and the deacetylation of PGC-1α (PPARγ coactivator-1) (Zhang et al., 2012a). PPAR 532 activation can consecutively activate the catalase gene by PI3K/Nrf/Keap pathway (Rubiolo et 533 al., 2008). Moreover, resveratrol provides neuroprotection by encouraging mitochondrial 534 biogenesis through adenosine monophosphate (AMP)-activated protein kinase (AMPK) 535 activation (Wu et al., 2011). The study by Anastácio et al., (Anastácio et al., 2014) reported that 536 resveratrol exhibits neuroprotective effects by activating ERK-induced CREB regulation, 537 triggers BDNF, NGF, and glial cell-derived neurotrophic factor (GDNF) secretion, and 538 suppresses IL10, IL-1β, and NF-κB levels (**Table 2**). One of the main reasons for

539 neurodegeneration is ROS, which is caused by mitochondrial dysfunction and the nicotinamide

540 adenine dinucleotide phosphate (NADPH) oxidases (Zhang et al., 2010b). All of the above

- 541 studies afford evidence that resveratrol provides a favorable solution for neurodegenerative
- 542 diseases. Resveratrol is also effective in preventing BBB impairment and inhibiting Aβ1–42
- 543 from crossing the BBB and accumulating in the hippocampus (Omar, 2017; Zhao et al., 2015).
- 544 According to the study of Wang et al., (Wang et al., 2002) resveratrol could cross the BBB and
- 545 demonstrate protective effects against cerebral ischemic injury (**Table 1**).

546

- 547
- 548

549 **4.5. Quercetin**

550 Flavonoids are widely known for their anti-oxidant activities and quercetin, one member of the 551 flavonoid group (**Fig. 4**), has been extensively investigated for its anti-proliferative properties 552 (Lesjak et al., 2018). Heo and Lee (Heo and Lee, 2004) reported the protective activities of 553 quercetin on H_2O_2 -mediated neurodegeneration in pheochromocytoma cells (PC12) cells. 554 Findings also revealed that quercetin treatment convincingly ameliorates cell viability. In 555 addition to this, quercetin exhibited an increased antioxidant protective activity as compared to 556 vitamin C. Indeed, 10 µM quercetin exerted protective activities in neurons and also exhibited a 557 superior cell viability effect over 30 µM (Heo and Lee, 2004). It has been shown to cross the 558 BBB and prevents H_2O_2 -induced cytotoxicity, as well (Heo and Lee, 2004). According to the 559 study of Ishisaka et al., (Ishisaka et al., 2011) quercetin could permeate the BBB and protect it 560 from disorders connected with oxidative stress in an animal model (**Table 1**).

561 Quercetin regulates the activity of kinases that consecutively modulate gene expression and

562 cellular function. The arrangement of the functional groups also plays an important role in the

563 properties of quercetin (Barreca et al., 2016). The activity of quercetin is influenced by some

564 factors including time exposure and cell type, and at higher doses $(2100 \,\mu\text{M})$ it shows cytotoxic,

565 apoptotic, genotoxic, and anti-proliferative properties. Dajas et al., (Dajas et al., 2015)

- 566 demonstrated that quercetin regulated NF-κB which could consecutively improve inflammatory
- 567 processes associated with neurodegenerative diseases. Numerous studies (Liu et al., 2013, 2015;
- 568 Suganthy et al., 2016) have reported that quercetin demonstrates neuroprotective activity by

569 regulating synaptic plasticity, learning, and neuronal adaptation through signaling pathways of

- 570 the PI3K/Akt/Nrf2, ERK/CREB/BDNF, PGC-1α/fibronectin type III domain containing 5
- 571 (FNDC5)/BDNF, and activation of SIRT1 (**Table 2**).

572 Quercetin has also been found to modulate protein kinases, lipid kinases including the PI3k, 573 tyrosine kinases, Akt/PKB, MAP kinases, and PKC (Dajas et al., 2015). This is useful as it 574 modifies the state of phosphorylation of the target molecules hence deciding the cellular fate. 575 Moreover, quercetin has been shown to activate pro-apoptotic as well as anti-apoptotic pathways 576 (Bournival et al., 2009). Quercetin also plays an important role in increasing the biogenesis of 577 mitochondria and this is an essential feature as mitochondrial dysfunction results in degenerating 578 neurons by depleting adenosine triphosphate levels and generating ROS. Therefore, quercetin 579 has the potential to protect from NDs through mitochondria-targeted activities (Dajas et al., 580 2015). Nonetheless, before the clinical trials of quercetin, there is a need for more evidence about 581 the absorption and metabolism of quercetin after its oral administration (Moreno et al., 2017).

582

583 **5. Conclusion**

584 At the molecular and cellular levels, an increased level of neuronal damage might be associated 585 with the pathogenesis of NDs. Possibly, OS can cause neuronal cell death by elevating the 586 concentration of intracellular Ca^{2+} , glutamate-mediated excitotoxicity, reducing overall survival 587 signaling, activation of death-signaling cascades, and mitochondrial dysfunction. Polyphenols 588 indeed provide protection against neuronal damage and this article elucidates the multiple 589 mechanisms by which they can act against NDs. Although a number of studies have revealed 590 their potential benefits, more extensive studies are required to establish the efficacy and long-591 term effects of using polyphenols as therapeutics for NDs.

592 593

594 **Author contributions**

595 MSU conceived the original idea and designed the outlines of the study. MSU, AAM, MTK, and

596 JA wrote the draft of the manuscript. MSU and AAM prepared the figures for the manuscript. PJ

- 597 edited the whole manuscript. MSS, GMA, and LA performed the literature review and aided in
- 598 revising the manuscript. All authors have read and approved the final manuscript.

- 626 Barreca, D., Bellocco, E., D'Onofrio, G., Nabavi, S. F., Daglia, M., Rastrelli, L., et al. (2016). 627 Neuroprotective Effects of Quercetin: From Chemistry to Medicine. *CNS Neurol. Disord.* 628 *Drug Targets* 15, 964–975.
- 629 Bieschke, J., Russ, J., Friedrich, R. P., Ehrnhoefer, D. E., Wobst, H., Neugebauer, K., et al.
- 630 (2010). EGCG remodels mature α-synuclein and amyloid-β fibrils and reduces cellular
- 631 toxicity. *Proc. Natl. Acad. Sci.* 107, 7710–7715. doi:10.1073/pnas.0910723107.
- 632 Bishnoi, M., Patii, C. S., Kumar, A., and Kulkarni, S. K. (2005). Relative role of
- 633 cyclooxygenase-2 (cox-2) inhibitors and lipoxygenase (lox) inhibitors in aging induced
- 634 dementia and oxidative damage. *Ann. Neurosci.* 12, 6–11.
- 635 doi:10.5214/ans.0972.7531.2005.120202.
- 636 Bishop, N. A., Lu, T., and Yankner, B. A. (2010). Neural mechanisms of ageing and cognitive 637 decline. *Nature* 464, 529–535. doi:10.1038/nature08983.
- 638 Bossy-Wetzel, E., Schwarzenbacher, R., and Lipton, S. A. (2004). Molecular pathways to 639 neurodegeneration. *Nat. Med.* 10, S2–S9. doi:10.1038/nm1067.
- 640 Bournival, J., Quessy, P., and Martinoli, M.-G. (2009). Protective Effects of Resveratrol and
- 641 Quercetin Against MPP+ -Induced Oxidative Stress Act by Modulating Markers of
- 642 Apoptotic Death in Dopaminergic Neurons. *Cell. Mol. Neurobiol.* 29, 1169–1180.
- 643 doi:10.1007/s10571-009-9411-5.
- 644 Bourtchuladze, R., Frenguelli, B., Blendy, J., Cioffi, D., Schutz, G., and Silva, A. J. (1994). 645 Deficient long-term memory in mice with a targeted mutation of the cAMP-responsive 646 element-binding protein. *Cell* 79, 59–68. doi:10.1016/0092-8674(94)90400-6.
- 647 Butterfield, D. A., Reed, T., Newman, S. F., and Sultana, R. (2007). Roles of amyloid β-peptide-
- 648 associated oxidative stress and brain protein modifications in the pathogenesis of
- 649 Alzheimer's disease and mild cognitive impairment. *Free Radic. Biol. Med.* 43, 658–677.
- 650 doi:10.1016/J.FREERADBIOMED.2007.05.037.
- 651 Butterfield, D. A., Reed, T., Perluigi, M., De Marco, C., Coccia, R., Cini, C., et al. (2006).
- 652 Elevated protein-bound levels of the lipid peroxidation product, 4-hydroxy-2-nonenal, in
- 653 brain from persons with mild cognitive impairment. *Neurosci. Lett.* 397, 170–173. 654 doi:10.1016/j.neulet.2005.12.017.
- 655 Caesar, I., Jonson, M., Nilsson, K. P. R., Thor, S., and Hammarström, P. (2012). Curcumin 656 Promotes A-beta Fibrillation and Reduces Neurotoxicity in Transgenic Drosophila. *PLoS* 657 *One* 7, e31424. doi:10.1371/journal.pone.0031424.
- 658 Calabrese, E. J. (2008). Neuroscience and hormesis: Overview and general findings. *Crit. Rev.* 659 *Toxicol.* 38, 249–252. doi:10.1080/10408440801981957.
- 660 Capiralla, H., Vingtdeux, V., Zhao, H., Sankowski, R., Al-Abed, Y., Davies, P., et al. (2012).
- 661 Resveratrol mitigates lipopolysaccharide- and Aβ-mediated microglial inflammation by 662 inhibiting the TLR4/NF-κB/STAT signaling cascade. *J. Neurochem.* 120, 461–472.
- 663 doi:10.1111/j.1471-4159.2011.07594.x.
- 664 Castellano-González, G., Pichaud, N., Ballard, J. W. O., Bessede, A., Marcal, H., and Guillemin, 665 G. J. (2016). Epigallocatechin-3-gallate induces oxidative phosphorylation by activating 666 cytochrome c oxidase in human cultured neurons and astrocytes. *Oncotarget* 7, 7426–40. 667 doi:10.18632/oncotarget.6863.
- 668 Castelli, V., Benedetti, E., Antonosante, A., Catanesi, M., Pitari, G., Ippoliti, R., et al. (2019).
- 669 Neuronal cells rearrangement during aging and neurodegenerative disease: Metabolism,
- 670 oxidative stress and organelles dynamic. *Front. Mol. Neurosci.* 12.
- 671 doi:10.3389/fnmol.2019.00132.
- 672 Chan, S., Kantham, S., Rao, V. M., Palanivelu, M. K., Pham, H. L., Shaw, P. N., et al. (2016).
- 673 Metal chelation, radical scavenging and inhibition of Aβ42 fibrillation by food constituents
- 674 in relation to Alzheimer's disease. *Food Chem.* 199, 185–194.
- 675 doi:10.1016/j.foodchem.2015.11.118.
- 676 Chen, C.-Y., Jang, J.-H., Li, M.-H., and Surh, Y.-J. (2005a). Resveratrol upregulates heme
- 677 oxygenase-1 expression via activation of NF-E2-related factor 2 in PC12 cells. *Biochem.*
- 678 *Biophys. Res. Commun.* 331, 993–1000. doi:10.1016/j.bbrc.2005.03.237.
- 679 Chen, C. Y., Jang, J. H., Li, M. H., and Surh, Y. J. (2005b). Resveratrol upregulates heme

- 707 Czapski, G. A., Czubowicz, K., Strosznajder, J. B., and Strosznajder, R. P. (2016). The 708 lipoxygenases: Their regulation and implication in alzheimer's disease. *Neurochem. Res.*
- 709 41, 243–257. doi:10.1007/s11064-015-1776-x.
- 710 Dajas, F., Abin-Carriquiry, J. A., Arredondo, F., Blasina, F., Echeverry, C., Martínez, M., et al.
- 711 (2015). Quercetin in brain diseases: Potential and limits. *Neurochem. Int.* 89, 140–148. 712 doi:10.1016/j.neuint.2015.07.002.
- 713 Dawbarn, D., and Allen, S. J. (2003). Neurotrophins and neurodegeneration. *Neuropathol. Appl.* 714 *Neurobiol.* 29, 211–230. doi:10.1046/j.1365-2990.2003.00487.x.
- 715 de Sá Coutinho, D., Pacheco, M., Frozza, R., and Bernardi, A. (2018). Anti-Inflammatory
- 716 Effects of Resveratrol: Mechanistic Insights. *Int. J. Mol. Sci.* 19, 1812.
- 717 doi:10.3390/ijms19061812.
- 718 Debnath, K. C., Uddin, M. S., Goswami, S., Herrera-Ca, O., Kabir, M. T., Hasan, M. F., et al. 719 (2017). Impact of Tobacco Smoking, Betel Quid Chewing and Alcohol Consumption Habits 720 in Patients with Oral Cavity Cancer in Bangladesh. *J. Med. Sci.* 17, 46–52.
- 721 doi:10.3923/jms.2017.46.52.
- 722 Deng, Z., Peng, Y., and Gao, W. (2018). Berberine protects against chronic social defeat stress-723 induced depressive-like behaviors with upregulation of neuronal PAS domain protein 724 4/brain-derived neurotrophic factor signaling pathway. *Pharmacogn. Mag.* 14, 501. 725 doi:10.4103/pm.pm 268 18.
- 726 Ding, M. L., Ma, H., Man, Y. G., and Lv, H. Y. (2017). Protective effects of a green tea 727 polyphenol, epigallocatechin- 3-gallate, against sevoflurane-induced neuronal apoptosis 728 involve regulation of CREB/BDNF/TrkB and PI3K/Akt/mTOR signalling pathways in
- 729 neonatal mice. *Can. J. Physiol. Pharmacol.* 95, 1396–1405. doi:10.1139/cjpp-2016-0333.
- 730 Durairajan, S. S. K., Liu, L.-F., Lu, J.-H., Chen, L.-L., Yuan, Q., Chung, S. K., et al. (2012).
- 731 Berberine ameliorates β-amyloid pathology, gliosis, and cognitive impairment in an
- 732 Alzheimer's disease transgenic mouse model. *Neurobiol. Aging* 33, 2903–2919.
- 733 doi:10.1016/j.neurobiolaging.2012.02.016.
- 734 Essa, M. M., Vijayan, R. K., Castellano-Gonzalez, G., Memon, M. A., Braidy, N., and
- 735 Guillemin, G. J. (2012). Neuroprotective Effect of Natural Products Against Alzheimer's 736 Disease. *Neurochem. Res.* 37, 1829–1842. doi:10.1007/s11064-012-0799-9.
- 737 Farooqui, A. A. (2012). *Phytochemicals, Signal Transduction, and Neurological Disorders*. New
- 738 York, NY: Springer New York doi:10.1007/978-1-4614-3804-5.
- 739 Finkbeiner, S. (2000). CREB couples neurotrophin signals to survival messages. *Neuron* 25, 11– 740 14. doi:10.1016/S0896-6273(00)80866-1.
- 741 Frautschy, S. A., Hu, W., Kim, P., Miller, S. A., Chu, T., Harris-White, M. E., et al. Phenolic

742 anti-inflammatory antioxidant reversal of Abeta-induced cognitive deficits and

- 743 neuropathology. *Neurobiol. Aging* 22, 993–1005. doi:10.1016/s0197-4580(01)00300-1.
- 744 Gandhi, S., Wood-Kaczmar, A., Yao, Z., Plun-Favreau, H., Deas, E., Klupsch, K., et al. (2009). 745 PINK1-Associated Parkinson's Disease Is Caused by Neuronal Vulnerability to Calcium-746 Induced Cell Death. *Mol. Cell* 33, 627–638. doi:10.1016/j.molcel.2009.02.013.
- 747 Garcia-Alloza, M., Borrelli, L. A., Rozkalne, A., Hyman, B. T., and Bacskai, B. J. (2007).
- 748 Curcumin labels amyloid pathology in vivo, disrupts existing plaques, and partially restores
- 749 distorted neurites in an Alzheimer mouse model . *J. Neurochem.* 102, 1095–1104.
- 750 doi:10.1111/j.1471-4159.2007.04613.x.
- 751 Geon Ha Kim, Jieun E. Kim, Sandy Jeong Rhie, and Sujung Yoon (2015). The role for oxidative 752 stress in neurodegenerative diseases. *Exp Neurobiol* 24, 325–340. 753 doi:10.5607/en.2015.24.4.325.
- 754 Ghosh, A., Sarkar, S., Mandal, A. K., and Das, N. (2013). Neuroprotective Role of
- 755 Nanoencapsulated Quercetin in Combating Ischemia-Reperfusion Induced Neuronal
- 756 Damage in Young and Aged Rats. *PLoS One* 8, e57735. doi:10.1371/journal.pone.0057735.
- 757 Gil, J. M., and Rego, A. C. (2008). Mechanisms of neurodegeneration in Huntington's disease. 758 *Eur. J. Neurosci.* 27, 2803–2820. doi:10.1111/j.1460-9568.2008.06310.x.
- 759 Granzotto, A., and Zatta, P. (2011). Resveratrol Acts Not through Anti-Aggregative Pathways 760 but Mainly via Its Scavenging Properties against Aβ and Aβ-Metal Complexes Toxicity.
- 761 *PLoS One* 6, e21565. doi:10.1371/journal.pone.0021565.
- 762 Guo, Y.-J., Dong, S.-Y., Cui, X.-X., Feng, Y., Liu, T., Yin, M., et al. (2016). Resveratrol

763 alleviates MPTP-induced motor impairments and pathological changes by autophagic

764 degradation of α-synuclein via SIRT1-deacetylated LC3. *Mol. Nutr. Food Res.* 60, 2161–

- 765 2175. doi:10.1002/mnfr.201600111.
- 766 Halliwell, B. (2006). Oxidative stress and neurodegeneration: Where are we now? *J. Neurochem.* 767 97, 1634–1658. doi:10.1111/j.1471-4159.2006.03907.x.
- 768 Han, J., Wang, M., Jing, X., Shi, H., Ren, M., and Lou, H. (2014). (−)-Epigallocatechin Gallate 769 Protects Against Cerebral Ischemia-Induced Oxidative Stress via Nrf2/ARE Signaling. 770 *Neurochem. Res.* 39, 1292–1299. doi:10.1007/s11064-014-1311-5.
- 771 Hardas, S. S., Sultana, R., Clark, A. M., Beckett, T. L., Szweda, L. I., Paul Murphy, M., et al. 772 (2013). Oxidative modification of lipoic acid by HNE in alzheimer disease brain. *Redox* 773 *Biol.* 1, 80–85. doi:10.1016/j.redox.2013.01.002.
- 774 Harman, D. (2006). Free Radical Theory of Aging: An Update: Increasing the Functional Life 775 Span. *Ann. N. Y. Acad. Sci.* 1067, 10–21. doi:10.1196/annals.1354.003.
- 776 He, Y., Tan, D., Mi, Y., Zhou, Q., and Ji, S. (2017). Epigallocatechin-3-gallate attenuates 777 cerebral cortex damage and promotes brain regeneration in acrylamide-treated rats. *Food* 778 *Funct.* 8, 2275–2282. doi:10.1039/c6fo01823h.
- 779 Helman, A. M., and Murphy, M. P. (2016). Vascular cognitive impairment: Modeling a critical 780 neurologic disease in vitro and in vivo. *Biochim. Biophys. Acta - Mol. Basis Dis.* 1862, 975– 781 982. doi:10.1016/j.bbadis.2015.12.009.
- 782 Henchcliffe, C., and Beal, M. F. (2008). Mitochondrial biology and oxidative stress in Parkinson 783 disease pathogenesis. *Nat. Clin. Pract. Neurol.* 4, 600–609. doi:10.1038/ncpneuro0924.
- 784 Heo, H. J., and Lee, C. Y. (2004). Protective Effects of Quercetin and Vitamin C against
- 785 Oxidative Stress-Induced Neurodegeneration. *J. Agric. Food Chem.* 52, 7514–7517. 786 doi:10.1021/jf049243r.

804 podophyllum l.: an incredible therapeutic plant. *Biomed. Pharmacol. J.* 10. 805 doi:10.13005/bpj/1229.

806 Howes, M.-J. R., and Houghton, P. J. (2012). Ethnobotanical treatment strategies against 807 Alzheimer's disease. *Curr. Alzheimer Res.* 9, 67–85.

808 Hroudová, J., Singh, N., and Fišar, Z. (2014). Mitochondrial Dysfunctions in Neurodegenerative 809 Diseases: Relevance to Alzheimer's Disease. *Biomed Res. Int.* 2014. 810 doi:10.1155/2014/175062.

-
- 811 Hsu, Y.-Y., Chen, C.-S., Wu, S.-N., Jong, Y.-J., and Lo, Y.-C. (2012). Berberine activates Nrf2
- 812 nuclear translocation and protects against oxidative damage via a phosphatidylinositol 3-
- 813 kinase/Akt-dependent mechanism in NSC34 motor neuron-like cells. *Eur. J. Pharm. Sci.*
- 814 46, 415–425. doi:10.1016/j.ejps.2012.03.004.
- 815 Hsu, Y.-Y., Tseng, Y.-T., and Lo, Y.-C. (2013a). Berberine, a natural antidiabetes drug,
- 816 attenuates glucose neurotoxicity and promotes Nrf2-related neurite outgrowth. *Toxicol.* 817 *Appl. Pharmacol.* 272, 787–796. doi:10.1016/j.taap.2013.08.008.
- 818 Hsu, Y. Y., Tseng, Y. T., and Lo, Y. C. (2013b). Berberine, a natural antidiabetes drug,
- 819 attenuates glucose neurotoxicity and promotes Nrf2-related neurite outgrowth. *Toxicol.*
- 820 *Appl. Pharmacol.* 272, 787–796. doi:10.1016/j.taap.2013.08.008.
- 821 Huang, E. J., and Reichardt, L. F. (2001). Neurotrophins: Roles in Neuronal Development and 822 Function. *Annu. Rev. Neurosci.* 24, 677–736. doi:10.1146/annurev.neuro.24.1.677.
- 823 Huang, X., Cuajungco, M. P., Atwood, C. S., Hartshorn, M. A., Tyndall, J. D., Hanson, G. R., et
- 824 al. (1999). Cu(II) potentiation of alzheimer abeta neurotoxicity. Correlation with cell-free
- 825 hydrogen peroxide production and metal reduction. *J. Biol. Chem.* 274, 37111–6. 826 doi:10.1074/jbc.274.52.37111.
- 827 Impey, S., Smith, D. M., Obrietan, K., Donahue, R., Wade, C., and Storm, D. R. (1998). 828 Stimulation of cAMP response element (CRE)-mediated transcription during contextual 829 learning. *Nat. Neurosci.* 1, 595–601. doi:10.1038/2830.
- 830 Ishisaka, A., Ichikawa, S., Sakakibara, H., Piskula, M. K., Nakamura, T., Kato, Y., et al. (2011).
- 831 Accumulation of orally administered quercetin in brain tissue and its antioxidative effects in 832 rats. *Free Radic. Biol. Med.* 51, 1329–1336. doi:10.1016/j.freeradbiomed.2011.06.017.
- 833 Islam, M. S., Kanak, F., Iqbal, M. A., Faheema Islam, K., Al-Mamun, A., and Uddin, M. S.
- 834 (2018). Analyzing the Status of the Autism Spectrum Disorder Amid Children with

835 Intellectual Disabilities in Bangladesh. *Biomed. Pharmacol. J.* 11, 689–701.

- 836 doi:10.13005/bpj/1422.
- 837 Itoh, K., Wakabayashi, N., Katoh, Y., Ishii, T., Igarashi, K., Engel, J. D., et al. (1999). Keap1
- 838 represses nuclear activation of antioxidant responsive elements by Nrf2 through binding to
- 839 the amino-terminal Neh2 domain. *Genes Dev.* 13, 76–86. doi:10.1101/gad.13.1.76.
- 840 Itoh, T., Imano, M., Nishida, S., Tsubaki, M., Mizuguchi, N., Hashimoto, S., et al. (2012). (−)-
- 841 Epigallocatechin-3-gallate increases the number of neural stem cells around the damaged
- 842 area after rat traumatic brain injury. *J. Neural Transm.* 119, 877–890. doi:10.1007/s00702- 843 011-0764-9.
- 844 Jakaria, M., Azam, S., Haque, M. E., Jo, S.-H., Uddin, M. S., Kim, I.-S., et al. (2019). Taurine 845 and its analogs in neurological disorders: Focus on therapeutic potential and molecular 846 mechanisms. *Redox Biol.* 24, 101223. doi:10.1016/J.REDOX.2019.101223.
- 847 Jeandet, P., Sobarzo-Sánchez, E., Clément, C., Nabavi, S. F., Habtemariam, S., Nabavi, S. M., et 848 al. (2018). Engineering stilbene metabolic pathways in microbial cells. *Biotechnol. Adv.* 36, 849 2264–2283. doi:10.1016/j.biotechadv.2018.11.002.
- 850 Jeandet, P., Sobarzo-Sánchez, E., Silva, A. S., Clément, C., Nabavi, S. F., Battino, M., et al.
- 851 (2019). Whole-cell biocatalytic, enzymatic and green chemistry methods for the production
- 852 of resveratrol and its derivatives. *Biotechnol. Adv.*, 107461.
- 853 doi:10.1016/j.biotechadv.2019.107461.
- 854 Jiang, D., Men, L., Wang, J., Zhang, Y., Chickenyen, S., Wang, Y., et al. (2007). Redox 855 Reactions of Copper Complexes Formed with Different β-amyloid Peptides and Their 856 Neuropathalogical Relevance. *Biochemistry* 46, 9270–82. doi:10.1021/bi700508n.
- 857 Jin, Z., Ke, J., Guo, P., Wang, Y., and Wu, H. (2019). Quercetin improves blood-brain barrier 858 dysfunction in rats with cerebral ischemia reperfusion via Wnt signaling pathway. *Am. J.* 859 *Transl. Res.* 11, 4683–4695.
- 860 Jung, T., Catalgol, B., and Grune, T. (2009). The proteasomal system. *Mol. Aspects Med.* 30, 861 191–296. doi:10.1016/j.mam.2009.04.001.
- 862 Kabir, M. T., Abu Sufian, M., Uddin, M. S., Begum, M. M., Akhter, S., Islam, A., et al. (2019a). 863 NMDA Receptor Antagonists: Repositioning of Memantine as Multitargeting Agent for
- 864 Alzheimer's Therapy. *Curr. Pharm. Des.* 25, 3506–3518.
- 865 doi:10.2174/1381612825666191011102444.
- 866 Kabir, M. T., Uddin, M. S., Abdeen, A., Ashraf, G. M., Perveen, A., Hafeez, A., et al. (2020). 867 Evidence Linking Protein Misfolding to Quality Control in Progressive Neurodegenerative 868 Diseases. *Curr. Top. Med. Chem.* 20. doi:10.2174/1568026620666200618114924.

869 Kabir, M. T., Uddin, M. S., Begum, M. M., Thangapandiyan, S., Rahman, M. S., Aleya, L., et al.

- 870 (2019b). Cholinesterase Inhibitors for Alzheimer's Disease: Multitargeting Strategy based
- 871 on Anti-Alzheimer's Drugs Repositioning. *Curr. Pharm. Des.* 25, 3519–3535.
- 872 doi:10.2174/1381612825666191008103141.
- 873 Kalaiselvi, P., Rajashree, K., Bharathi Priya, L., and Padma, V. V. (2013). Cytoprotective effect
- 874 of epigallocatechin-3-gallate against deoxynivalenol-induced toxicity through anti-oxidative 875 and anti-inflammatory mechanisms in HT-29 cells. *Food Chem. Toxicol.* 56, 110–118. 876 doi:10.1016/j.fct.2013.01.042.
- 877 Kaur, G., Behl, T., Bungau, S., Kumar, A., Uddin, M. S., Mehta, V., et al. (2020). Dysregulation
- 878 of the Gut-Brain Axis, Dysbiosis and Influence of numerous factors on Gut Microbiota
- 879 associated Parkinson's Disease. *Curr. Neuropharmacol.* 18.
- 880 doi:10.2174/1570159x18666200606233050.
- 881 Kim, H., Park, B.-S., Lee, K.-G., Choi, C. Y., Jang, S. S., Kim, Y.-H., et al. (2005). Effects of 882 Naturally Occurring Compounds on Fibril Formation and Oxidative Stress of β-Amyloid. *J.* 883 *Agric. Food Chem.* 53, 8537–8541. doi:10.1021/jf051985c.
- 884 Kim, M., Cho, K.-H., Shin, M.-S., Lee, J.-M., Cho, H.-S., Kim, C.-J., et al. (2014). Berberine 885 prevents nigrostriatal dopaminergic neuronal loss and suppresses hippocampal apoptosis in 886 mice with Parkinson's disease. *Int. J. Mol. Med.* 33, 870–878. doi:10.3892/ijmm.2014.1656.
- 887 Kim, M., Shin, M. S., Lee, J. M., Cho, H. S., Kim, C. J., Kim, Y. J., et al. (2014). Inhibitory
- 888 Effects of Isoquinoline Alkaloid Berberine on Ischemia-Induced Apoptosis via Activation 889 of Phosphoinositide 3-Kinase/Protein Kinase B Signaling Pathway. *Int. Neurourol. J.* 18, 890 115. doi:10.5213/inj.2014.18.3.115.
- 891 Kishida, K. T., and Klann, E. (2007). Sources and Targets of Reactive Oxygen Species in 892 Synaptic Plasticity and Memory. *Antioxid. Redox Signal.* 9, 233–244. 893 doi:10.1089/ars.2007.9.233.
- 894 Koh, S.-H., Kim, S. H., Kwon, H., Park, Y., Kim, K. S., Song, C. W., et al. (2003).
- 895 Epigallocatechin gallate protects nerve growth factor differentiated PC12 cells from 896 oxidative-radical-stress-induced apoptosis through its effect on phosphoinositide 3-
- 897 kinase/Akt and glycogen synthase kinase-3. *Mol. Brain Res.* 118, 72–81. 898 doi:10.1016/j.molbrainres.2003.07.003.
- 899 Kong, A. N. T., Yu, R., Chen, C., Mandlekar, S., and Primiano, T. (2000). Signal Transduction 900 Events Elicited by Natural Products: Role of MAPK and Caspase Pathways in Homeostatic 901 Response and Induction of Apoptosis. *Arch. Pharm. Res.* 23, 1–16.
- 902 doi:10.1007/BF02976458.
- 903 Koppen, L. M., Whitaker, A., Rosene, A., and Beckett, R. D. (2017). Efficacy of Berberine 904 Alone and in Combination for the Treatment of Hyperlipidemia: A Systematic Review. *J.* 905 *Evid. Based. Complementary Altern. Med.* 22, 956–968. doi:10.1177/2156587216687695.
- 906 Kraft, A. D., Johnson, D. A., and Johnson, J. A. (2004). Nuclear Factor E2-Related Factor 2- 907 Dependent Antioxidant Response Element Activation by tert-Butylhydroquinone and 908 Sulforaphane Occurring Preferentially in Astrocytes Conditions Neurons against Oxidative 909 Insult. *J. Neurosci.* 24, 1101–1112. doi:10.1523/JNEUROSCI.3817-03.2004.
- 910 Kuchibhotla, K. V., Goldman, S. T., Lattarulo, C. R., Wu, H. Y., Hyman, B. T., and Bacskai, B.
- 911 J. (2008). Aβ Plaques Lead to Aberrant Regulation of Calcium Homeostasis In Vivo
- 912 Resulting in Structural and Functional Disruption of Neuronal Networks. *Neuron* 59, 214–
- 913 225. doi:10.1016/j.neuron.2008.06.008.
- 914 Kwon, I. H., Choi, H. S., Shin, K. S., Lee, B. K., Lee, C. K., Hwang, B. Y., et al. (2010). Effects 915 of berberine on 6-hydroxydopamine-induced neurotoxicity in PC12 cells and a rat model of 916 Parkinson's disease. *Neurosci. Lett.* 486, 29–33. doi:10.1016/j.neulet.2010.09.038.
- 917 Kysenius, K., Brunello, C. A., and Huttunen, H. J. (2014). Mitochondria and NMDA Receptor-918 Dependent Toxicity of Berberine Sensitizes Neurons to Glutamate and Rotenone Injury. 919 *PLoS One* 9, e107129. doi:10.1371/journal.pone.0107129.
- 920 Lassmann, H. (2011). Mechanisms of neurodegeneration shared between multiple sclerosis and 921 Alzheimer's disease. *J. Neural Transm.* 118, 747–752. doi:10.1007/s00702-011-0607-8.
- 922 Lee, J. M., and Johnson, J. A. (2004). An important role of Nrf2-ARE pathway in the cellular 923 defense mechanism. *J. Biochem. Mol. Biol.* 37, 139–143.
- 924 doi:10.5483/bmbrep.2004.37.2.139.
- 925 Lesjak, M., Beara, I., Simin, N., Pintać, D., Majkić, T., Bekvalac, K., et al. (2018). Antioxidant 926 and anti-inflammatory activities of quercetin and its derivatives. *J. Funct. Foods* 40, 68–75. 927 doi:10.1016/J.JFF.2017.10.047.
- 928 Levites, Y., Amit, T., Youdim, M. B. H., and Mandel, S. (2002). Involvement of Protein Kinase 929 C Activation and Cell Survival/ Cell Cycle Genes in Green Tea Polyphenol (−)- 930 Epigallocatechin 3-Gallate Neuroprotective Action. *J. Biol. Chem.* 277, 30574–30580. 931 doi:10.1074/jbc.M202832200.
- 932 Li, X., Valencia, A., Sapp, E., Masso, N., Alexander, J., Reeves, P., et al. (2010). Aberrant
- 933 Rab11-Dependent Trafficking of the Neuronal Glutamate Transporter EAAC1 Causes
- 934 Oxidative Stress and Cell Death in Huntington's Disease. *J. Neurosci.* 30, 4552–4561. 935 doi:10.1523/JNEUROSCI.5865-09.2010.
-
- 936 Liao, V. H.-C., Yu, C.-W., Chu, Y.-J., Li, W.-H., Hsieh, Y.-C., and Wang, T.-T. (2011). 937 Curcumin-mediated lifespan extension in Caenorhabditis elegans. *Mech. Ageing Dev.* 132, 938 480–487. doi:10.1016/j.mad.2011.07.008.
- 939 Liguori, I., Russo, G., Curcio, F., Bulli, G., Aran, L., Della-Morte, D., et al. (2018). Oxidative 940 stress, aging, and diseases. *Clin. Interv. Aging* 13, 757–772. doi:10.2147/CIA.S158513.
- 941 Lim, G. P., Chu, T., Yang, F., Beech, W., Frautschy, S. A., and Cole, G. M. (2001). The curry 942 spice curcumin reduces oxidative damage and amyloid pathology in an Alzheimer 943 transgenic mouse. *J. Neurosci.* 21, 8370–7.
- 944 Liu, D., Wang, Z., Gao, Z., Xie, K., Zhang, Q., Jiang, H., et al. (2014a). Effects of curcumin on 945 learning and memory deficits, BDNF, and ERK protein expression in rats exposed to 946 chronic unpredictable stress. *Behav. Brain Res.* 271, 116–121.
- 947 doi:10.1016/j.bbr.2014.05.068.
- 948 Liu, M., Chen, F., Sha, L., Wang, S., Tao, L., Yao, L., et al. (2014b). (−)-Epigallocatechin-3- 949 Gallate Ameliorates Learning and Memory Deficits by Adjusting the Balance of 950 TrkA/p75NTR Signaling in APP/PS1 Transgenic Mice. *Mol. Neurobiol.* 49, 1350–1363.

951 doi:10.1007/s12035-013-8608-2.

- 952 Liu, P., Zou, D., Yi, L., Chen, M., Gao, Y., Zhou, R., et al. (2015). Quercetin ameliorates 953 hypobaric hypoxia-induced memory impairment through mitochondrial and neuron function
- 954 adaptation via the PGC-1α pathway. *Restor. Neurol. Neurosci.* 33, 143–157.
- 955 doi:10.3233/RNN-140446.
- 956 Liu, R., Zhang, T. T., Zhou, D., Bai, X. Y., Zhou, W. L., Huang, C., et al. (2013). Quercetin 957 protects against the Aβ25-35-induced amnesic injury: Involvement of inactivation of 958 RAGE-mediated pathway and conservation of the NVU. *Neuropharmacology* 67, 419–431. 959 doi:10.1016/j.neuropharm.2012.11.018.
- 960 Liu, Z., Zhou, T., Ziegler, A. C., Dimitrion, P., and Zuo, L. (2017). Oxidative Stress in 961 Neurodegenerative Diseases: From Molecular Mechanisms to Clinical Applications. *Oxid.* 962 *Med. Cell. Longev.* 2017. doi:10.1155/2017/2525967.
- 963 Lu, C., Guo, Y., Yan, J., Luo, Z., Luo, H. Bin, Yan, M., et al. (2013). Design, synthesis, and 964 evaluation of multitarget-directed resveratrol derivatives for the treatment of Alzheimer's 965 disease. *J. Med. Chem.* 56, 5843–5859. doi:10.1021/jm400567s.
- 966 Makkar, R., Behl, T., Bungau, S., Zengin, G., Mehta, V., Kumar, A., et al. (2020). Nutraceuticals 967 in Neurological Disorders. *Int. J. Mol. Sci.* 21. doi:10.3390/ijms21124424.
- 968 Maleki, S. N., Aboutaleb, N., and Souri, F. (2018). Berberine confers neuroprotection in coping 969 with focal cerebral ischemia by targeting inflammatory cytokines. *J. Chem. Neuroanat.* 87, 970 54–59. doi:10.1016/j.jchemneu.2017.04.008.
- 971 Mandel, S. A., Amit, T., Kalfon, L., Reznichenko, L., Weinreb, O., and Youdim, M. B. H.
- 972 (2008). Cell signaling pathways and iron chelation in the neurorestorative activity of green
- 973 tea polyphenols: special reference to epigallocatechin gallate (EGCG). *J. Alzheimers. Dis.* 974 15, 211–22.
- 975 Mangialasche, F., Solomon, A., Winblad, B., Mecocci, P., and Kivipelto, M. (2010).
- 976 Alzheimer's disease: clinical trials and drug development. *Lancet Neurol.* 9, 702–716.
- 977 doi:10.1016/S1474-4422(10)70119-8.
- 978 Marini, A. M., Jiang, X., Wu, X., Tian, F., Zhu, D., Okagaki, P., et al. (2004). Role of brain-979 derived neurotrophic factor and NF-kappaB in neuronal plasticity and survival: From genes 980 to phenotype. *Restor. Neurol. Neurosci.* 22, 121–30.
- 981 Mattson, M. P., and Cheng, A. (2006). Neurohormetic phytochemicals: low-dose toxins that 982 induce adaptive neuronal stress responses. *Trends Neurosci.* 29, 632–639.
- 983 doi:10.1016/j.tins.2006.09.001.
- 984 Md. Sahab Uddin;, and Aman B. Upaganlawar (2019). *Oxidative Stress and Antioxidant Defense* 985 *Biomedical Value in Health and Diseases*. USA: Nova Science Publishers.
- 986 Meo, S. Di, Reed, T. T., Venditti, P., and Victor, V. M. (2016). Role of ROS and RNS Sources 987 in Physiological and Pathological Conditions. *Oxid. Med. Cell. Longev.* 2016.
- 988 doi:10.1155/2016/1245049.
- 989 Mitjavila, M. T., and Moreno, J. J. (2012). The effects of polyphenols on oxidative stress and the 990 arachidonic acid cascade. Implications for the prevention/treatment of high prevalence 991 diseases. *Biochem. Pharmacol.* 84, 1113–1122. doi:10.1016/j.bcp.2012.07.017.
- 992 Mizuno, Y. (2014). Recent Research Progress in and Future Perspective on Treatment of 993 Parkinson's Disease. *Integr. Med. Int.* 1, 67–79. doi:10.1159/000365571.
- 994 Mokni, M., Elkahoui, S., Limam, F., Amri, M., and Aouani, E. (2007). Effect of resveratrol on 995 antioxidant enzyme activities in the brain of healthy rat. *Neurochem. Res.* 32, 981–987. 996 doi:10.1007/s11064-006-9255-z.
- 997 Montine, T. J., Neely, M. D., Quinn, J. F., Beal, M. F., Markesbery, W. R., Roberts, L. J., et al. 998 (2002). Lipid peroxidation in aging brain and Alzheimer's disease. *Free Radic. Biol. Med.* 999 33, 620–626. doi:10.1016/S0891-5849(02)00807-9.
- 1000 Moreno, L. C. G. e I., Puerta, E., Suárez-Santiago, J. E., Santos-Magalhães, N. S., Ramirez, M.
- 1001 J., and Irache, J. M. (2017). Effect of the oral administration of nanoencapsulated quercetin
- 1002 on a mouse model of Alzheimer's disease. *Int. J. Pharm.* 517, 50–57.
- 1003 doi:10.1016/j.ijpharm.2016.11.061.
- 1004 Mourtas, S., Canovi, M., Zona, C., Aurilia, D., Niarakis, A., La Ferla, B., et al. (2011).
- 1005 Curcumin-decorated nanoliposomes with very high affinity for amyloid-β1-42 peptide. 1006 *Biomaterials* 32, 1635–1645. doi:10.1016/j.biomaterials.2010.10.027.
- 1007 Nam, S. M., Choi, J. H., Yoo, D. Y., Kim, W., Jung, H. Y., Kim, J. W., et al. (2014). Effects of 1008 curcumin (Curcuma longa) on learning and spatial memory as well as cell proliferation and 1009 neuroblast differentiation in adult and aged mice by upregulating brain-derived neurotrophic 1010 factor and CREB signaling. *J. Med. Food* 17, 641–649. doi:10.1089/jmf.2013.2965.
- 1011 Nguyen, T., Sherratt, P. J., and Pickett, C. B. (2003). Regulatory mechanisms controlling gene 1012 expression mediated by the antioxidant response element. *Annu. Rev. Pharmacol. Toxicol.* 1013 43, 233–260. doi:10.1146/annurev.pharmtox.43.100901.140229.
- 1014 Nicholls, D. G. (2008). Oxidative Stress and Energy Crises in Neuronal Dysfunction. *Ann. N. Y.* 1015 *Acad. Sci.* 1147, 53–60. doi:10.1196/annals.1427.002.
- 1016 Numakawa, T., Matsumoto, T., Numakawa, Y., Richards, M., Yamawaki, S., and Kunugi, H. 1017 (2011). Protective Action of Neurotrophic Factors and Estrogen against Oxidative Stress-1018 Mediated Neurodegeneration. *J. Toxicol.* 2011, 1–12. doi:10.1155/2011/405194.
- 1019 Ola, M. S., Ahmed, M. M., Shams, S., and Al-Rejaie, S. S. (2017). Neuroprotective effects of 1020 quercetin in diabetic rat retina. *Saudi J. Biol. Sci.* 24, 1186–1194.
- 1021 doi:10.1016/j.sjbs.2016.11.017.
- 1022 Oliveira, J. M. A., Jekabsons, M. B., Chen, S., Lin, A., Rego, A. C., Gonçalves, J., et al. (2007). 1023 Mitochondrial dysfunction in Huntington's disease: The bioenergetics of isolated and in situ 1024 mitochondria from transgenic mice. *J. Neurochem.* 101, 241–249. doi:10.1111/j.1471- 1025 4159.2006.04361.x.
- 1026 Omar, S. H. (2017). Biophenols pharmacology against the amyloidogenic activity in Alzheimer's 1027 disease. *Biomed. Pharmacother.* 89, 396–413. doi:10.1016/j.biopha.2017.02.051.
- 1028 Ortiz-López, L., Márquez-Valadez, B., Gómez-Sánchez, A., Silva-Lucero, M. D. C., Torres-
- 1029 Pérez, M., Téllez-Ballesteros, R. I., et al. (2016). Green tea compound epigallo-catechin-3-
- 1030 gallate (EGCG) increases neuronal survival in adult hippocampal neurogenesis in vivo and
- 1031 in vitro. *Neuroscience* 322, 208–220. doi:10.1016/j.neuroscience.2016.02.040.

- 1049 Purpura, M., Lowery, R. P., Wilson, J. M., Mannan, H., Münch, G., and Razmovski-Naumovski, 1050 V. (2018). Analysis of different innovative formulations of curcumin for improved relative 1051 oral bioavailability in human subjects. *Eur. J. Nutr.* 57, 929–938. doi:10.1007/s00394-016- 1052 1376-9.
- 1053 Qin, Z., Hu, D., Han, S., Reaney, S. H., Di Monte, D. A., and Fink, A. L. (2007). Effect of 4- 1054 hydroxy-2-nonenal modification on alpha-synuclein aggregation. *J. Biol. Chem.* 282, 5862– 1055 70. doi:10.1074/jbc.M608126200.
- 1056 Rahman, A., Haque, A., Uddin, M. S., Mian, M. M., Sufian, M. A., Rahman, M. M., et al.
- 1057 (2017). In vitro screening for antioxidant and anticholinesterase effects of Uvaria littoralis
- 1058 Blume.: A nootropic phytotherapeutic remedy. *J. Intellect. Disabil. Diagnosis Treat.* 5.
- 1059 Reichardt, L. F. (2006). Neurotrophin-regulated signalling pathways. *Philos. Trans. R. Soc. B* 1060 *Biol. Sci.* 361, 1545–1564. doi:10.1098/rstb.2006.1894.
- 1061 Ren, J., Fan, C., Chen, N., Huang, J., and Yang, Q. (2011). Resveratrol pretreatment attenuates 1062 cerebral ischemic injury by upregulating expression of transcription factor Nrf2 and HO-1 1063 in Rats. *Neurochem. Res.* 36, 2352–2362. doi:10.1007/s11064-011-0561-8.
- 1064 Rubiolo, J. A., Mithieux, G., and Vega, F. V. (2008). Resveratrol protects primary rat 1065 hepatocytes against oxidative stress damage: activation of the Nrf2 transcription factor and 1066 augmented activities of antioxidant enzymes. *Eur. J. Pharmacol.* 591, 66–72. 1067 doi:10.1016/j.ejphar.2008.06.067.
- 1068 Russo, A., Acquaviva, R., Campisi, A., Sorrenti, V., Di Giacomo, C., Virgata, G., et al. (2000). 1069 Bioflavonoids as antiradicals, antioxidants and DNA cleavage protectors. *Cell Biol. Toxicol.* 1070 16, 91–98. doi:10.1023/A:1007685909018.
- 1071 Sahab Uddin, M., and Ashraf, G. M. (2020). *Quality control of cellular protein in* 1072 *neurodegenerative disorders*. Hershey: IGI Global doi:10.4018/978-1-7998-1317-0.
- 1073 Sahab Uddin, M., and Shah Amran, M. (2018). *Handbook of research on critical examinations* 1074 *of neurodegenerative disorders*. First. IGI Global: USA.
- 1075 Satoh, T., Enokido, Y., Kubo, T., Yamada, M., and Hatanaka, H. (1998). Gram Positive Bacteria 1076 Induce IL-6 and IL-8 Production in Human Alveolar Macrophages and Epithelial Cells. 1077 *Cell. Mol. Neurobiol.* 18, 649–666. doi:10.1023/A:1020269802315.
- 1078 Schneider, L. S., Mangialasche, F., Andreasen, N., Feldman, H., Giacobini, E., Jones, R., et al. 1079 (2014). Clinical trials and late-stage drug development for Alzheimer's disease: an appraisal 1080 from 1984 to 2014. *J. Intern. Med.* 275, 251–283. doi:10.1111/joim.12191.
-
- 1081 Schroeter, H., Bahia, P., Spencer, J. P. E., Sheppard, O., Rattray, M., Cadenas, E., et al. (2007).
- 1082 (-)Epicatechin stimulates ERK-dependent cyclic AMP response element activity and up-
- 1083 regulates GluR2 in cortical neurons. *J. Neurochem.* 101, 1596–1606. doi:10.1111/j.1471- 1084 4159.2006.04434.x.
- 1085 Schroeter, H., Spencer, J. P. E., Rice-Evans, C., and Williams, R. J. (2001). Flavonoids protect

- 1089 Selvakumar, K., Bavithra, S., Krishnamoorthy, G., and Arunakaran, J. (2019). Impact of 1090 quercetin on tight junctional proteins and BDNF signaling molecules in hippocampus of 1091 PCBs-exposed rats. *Interdiscip. Toxicol.* 11, 294–305. doi:10.2478/intox-2018-0029.
- 1092 Shin, K. S., Choi, H. S., Zhao, T. T., Suh, K. H., Kwon, I. H., Choi, S. O., et al. (2013).
- 1093 Neurotoxic effects of berberine on long-term l-DOPA administration in 6-
- 1094 hydroxydopamine-lesioned rat model of Parkinson's disease. *Arch. Pharm. Res.* 36, 759–
- 1095 767. doi:10.1007/s12272-013-0051-4.
- 1096 Simões Pires, E. N., Frozza, R. L., Hoppe, J. B., Menezes, B. de M., and Salbego, C. G. (2014). 1097 Berberine was neuroprotective against an in vitro model of brain ischemia: Survival and 1098 apoptosis pathways involved. *Brain Res.* 1557, 26–33. doi:10.1016/j.brainres.2014.02.021.
- 1099 Smith, A., Giunta, B., Bickford, P. C., Fountain, M., Tan, J., and Shytle, R. D. (2010).
- 1100 Nanolipidic particles improve the bioavailability and α-secretase inducing ability of
- 1101 epigallocatechin-3-gallate (EGCG) for the treatment of Alzheimer's disease. *Int. J. Pharm.*
- 1102 389, 207–212. doi:10.1016/j.ijpharm.2010.01.012.
- 1103 Srividhya, R., Jyothilakshmi, V., Arulmathi, K., Senthilkumaran, V., and Kalaiselvi, P. (2008).
- 1104 Attenuation of senescence-induced oxidative exacerbations in aged rat brain by (−)-
- 1105 epigallocatechin-3-gallate. *Int. J. Dev. Neurosci.* 26, 217–223.
- 1106 doi:10.1016/j.ijdevneu.2007.12.003.
- 1107 Stack, E. C., Matson, W. R., and Ferrante, R. J. (2008). Evidence of Oxidant Damage in 1108 Huntington's Disease: Translational Strategies Using Antioxidants. *Ann. N. Y. Acad. Sci.* 1109 1147, 79–92. doi:10.1196/annals.1427.008.
- 1110 Storniolo, C. E., and Moreno, J. J. (2019). Resveratrol Analogs with Antioxidant Activity Inhibit
- 1111 Intestinal Epithelial Cancer Caco-2 Cell Growth by Modulating Arachidonic Acid Cascade.
- 1112 *J. Agric. Food Chem.* 67, 819–828. doi:10.1021/acs.jafc.8b05982.

1113 Strong, R., Miller, R. A., Astle, C. M., Baur, J. A., de Cabo, R., Fernandez, E., et al. (2013).

- 1114 Evaluation of Resveratrol, Green Tea Extract, Curcumin, Oxaloacetic Acid, and Medium-
- 1115 Chain Triglyceride Oil on Life Span of Genetically Heterogeneous Mice. *Journals*
- 1116 *Gerontol. Ser. A Biol. Sci. Med. Sci.* 68, 6–16. doi:10.1093/gerona/gls070.
- 1117 Suganthy, N., Devi, K. P., Nabavi, S. F., Braidy, N., and Nabavi, S. M. (2016). Bioactive effects
- 1118 of quercetin in the central nervous system: Focusing on the mechanisms of actions. *Biomed.* 1119 *Pharmacother.* 84, 892–908. doi:10.1016/j.biopha.2016.10.011.

1120 Timmers, S., Konings, E., Bilet, L., Houtkooper, R. H., van de Weijer, T., Goossens, G. H., et al.

- 1121 (2011). Calorie Restriction-like Effects of 30 Days of Resveratrol Supplementation on
- 1122 Energy Metabolism and Metabolic Profile in Obese Humans. *Cell Metab.* 14, 612–622.
- 1123 doi:10.1016/j.cmet.2011.10.002.
- 1124 Tramutola, A., Lanzillotta, C., Perluigi, M., and Butterfield, D. A. (2017). Oxidative stress,
- 1125 protein modification and Alzheimer disease. *Brain Res. Bull.* 133, 88–96.
- 1126 doi:10.1016/j.brainresbull.2016.06.005.

1127 Tresserra-Rimbau, A., Lamuela-Raventos, R. M., and Moreno, J. J. (2018). Polyphenols, food

1128 and pharma. Current knowledge and directions for future research. *Biochem. Pharmacol.*

- 1129 156, 186–195. doi:10.1016/j.bcp.2018.07.050.
- 1130 Tu, H., Nelson, O., Bezprozvanny, A., Wang, Z., Lee, S. F., Hao, Y. H., et al. (2006). Presenilins 1131 Form ER Ca2+ Leak Channels, a Function Disrupted by Familial Alzheimer's Disease-1132 Linked Mutations. *Cell* 126, 981–993. doi:10.1016/j.cell.2006.06.059.
- 1133 Tully, T., Bourtchouladze, R., Scott, R., and Tallman, J. (2003). Targeting the creb pathway for 1134 memory enhancers. *Nat. Rev. Drug Discov.* 2, 267–277. doi:10.1038/nrd1061.
- 1135 Uddin, M. S., Al Mamun, A., Asaduzzaman, M., Hosn, F., Abu Sufian, M., Takeda, S., et al.
- 1136 (2018). Spectrum of Disease and Prescription Pattern for Outpatients with Neurological
- 1137 Disorders: An Empirical Pilot Study in Bangladesh. *Ann. Neurosci.* 25, 25–37.
- 1138 doi:10.1159/000481812.
- 1139 Uddin, M. S., Al Mamun, A., Kabir, M. T., Jakaria, M., Mathew, B., Barreto, G. E., et al.
- 1140 (2019a). Nootropic and Anti-Alzheimer's Actions of Medicinal Plants: Molecular Insight
- 1141 into Therapeutic Potential to Alleviate Alzheimer's Neuropathology. *Mol. Neurobiol.* 56, 1142 4925–4944. doi:10.1007/s12035-018-1420-2.
- 1143 Uddin, M. S., Al Mamun, A., Rahman, M. A., Behl, T., Perveen, A., Hafeez, A., et al. (2020a). 1144 Emerging Proof of Protein Misfolding and Interactions in Multifactorial Alzheimer's 1145 Disease. *Curr. Top. Med. Chem.* 20. doi:10.2174/1568026620666200601161703.
- 1146 Uddin, M. S., Hossain, M. F., Mamun, A. Al, Shah, M. A., Hasana, S., and Bulbul, I. J. (2020b). 1147 Exploring the Multimodal Role of Phytochemicals in the Modulation of Cellular Signaling 1148 Pathways to Combat Age-Related Neurodegeneration. *Sci. Total Environ.* 725, 138313. 1149 doi:10.1016/j.scitotenv.2020.138313.
- 1150 Uddin, M. S., and Kabir, M. T. (2019). "Oxidative Stress in Alzheimer's Disease: Molecular 1151 Hallmarks of Underlying Vulnerability," in *Biological, Diagnostic and Therapeutic* 1152 *Advances in Alzheimer's Disease* (Singapore: Springer Singapore), 91–115. 1153 doi:10.1007/978-981-13-9636-6_5.
- 1154 Uddin, M. S., Kabir, M. T., Jakaria, M., Mamun, A. Al, Niaz, K., Amran, M. S., et al. (2019b). 1155 Endothelial PPARγ Is Crucial for Averting Age-Related Vascular Dysfunction by Stalling 1156 Oxidative Stress and ROCK. *Neurotox. Res.* 36, 583–601. doi:10.1007/s12640-019-00047- 1157 5.
- 1158 Uddin, M. S., Kabir, M. T., Jakaria, M., Sobarzo-Sánchez, E., Barreto, G. E., Perveen, A., et al. 1159 (2020c). Exploring the Potential of Neuroproteomics in Alzheimer's Disease. *Curr. Top.* 1160 *Med. Chem.* 20. doi:10.2174/1568026620666200603112030.
- 1161 Uddin, M. S., Kabir, M. T., Mamun, A. Al, Barreto, G. E., Rashid, M., Perveen, A., et al.
- 1162 (2020d). Pharmacological approaches to mitigate neuroinflammation in Alzheimer's 1163 disease. *Int. Immunopharmacol.* 84, 106479. doi:10.1016/j.intimp.2020.106479.
- 1164 Uddin, M. S., Kabir, M. T., Niaz, K., Jeandet, P., Clément, C., Mathew, B., et al. (2020e).
- 1165 Molecular Insight into the Therapeutic Promise of Flavonoids against Alzheimer's Disease.
- 1166 *Mol.* 25, 1267. doi:10.3390/MOLECULES25061267.
- 1167 Uddin, M. S., Kabir, M. T., Tewari, D., Al Mamun, A., Mathew, B., Aleya, L., et al. (2020f).
- 1168 Revisiting the role of brain and peripheral Aβ in the pathogenesis of Alzheimer's disease. *J.* 1169 *Neurol. Sci.* 416, 116974. doi:10.1016/j.jns.2020.116974.
- 1170 Uddin, M. S., Kabir, M. T., Tewari, D., Mathew, B., and Aleya, L. (2020g). Emerging signal
- 1171 regulating potential of small molecule biflavonoids to combat neuropathological insults of
- 1172 Alzheimer's disease. *Sci. Total Environ.* 700, 134836. doi:10.1016/j.scitotenv.2019.134836.
- 1173 Uddin, M. S., Mamun, A. Al, Jakaria, M., Thangapandiyan, S., Ahmad, J., Rahman, M. A., et al. 1174 (2020h). Emerging promise of sulforaphane-mediated Nrf2 signaling cascade against 1175 neurological disorders. *Sci. Total Environ.* 707, 135624.
- 1176 doi:10.1016/j.scitotenv.2019.135624.
- 1177 Uddin, M. S., Mamun, A. Al, Labu, Z. K., Hidalgo‐Lanussa, O., Barreto, G. E., and Ashraf, G. 1178 M. (2019c). Autophagic dysfunction in Alzheimer's disease: Cellular and molecular 1179 mechanistic approaches to halt Alzheimer's pathogenesis. *J. Cell. Physiol.* 234, 8094–8112. 1180 doi:10.1002/jcp.27588.
- 1181 Uddin, M. S., Uddin, G. M. S., Begum, M. M., Begum, Y., Herrera-Calderon, O., Islam, M. M.,
- 1182 et al. (2017). Inspection of phytochemical content and in vitro antioxidant profile of
- 1183 Gnaphalium luteoalbum L.: An unexplored phytomedicine. *J. Pharm. Nutr. Sci.* 7, 136–146. 1184 doi:10.6000/1927-5951.2017.07.03.10.
- 1185 Uttara, B., Singh, A., Zamboni, P., and Mahajan, R. (2009). Oxidative Stress and
- 1186 Neurodegenerative Diseases: A Review of Upstream and Downstream Antioxidant 1187 Therapeutic Options. *Curr. Neuropharmacol.* 7, 65–74. doi:10.2174/157015909787602823.
- 1188 Van Houten, B., Santa-Gonzalez, G. A., and Camargo, M. (2018). DNA repair after oxidative
- 1189 stress: Current challenges. *Curr. Opin. Toxicol.* 7, 9–16. doi:10.1016/j.cotox.2017.10.009.
- 1190 Vauzour, D. (2012). Dietary Polyphenols as Modulators of Brain Functions: Biological Actions
- 1191 and Molecular Mechanisms Underpinning Their Beneficial Effects. *Oxid. Med. Cell.*
- 1192 *Longev.* 2012, 1–16. doi:10.1155/2012/914273.
- 1193 Vauzour, D., Vafeiadou, K., Rice-Evans, C., Williams, R. J., and Spencer, J. P. E. (2007).
- 1194 Activation of pro-survival Akt and ERK1/2 signalling pathways underlie the anti-apoptotic 1195 effects of flavanones in cortical neurons. *J. Neurochem.* 103, 1355–1367.
- 1196 doi:10.1111/j.1471-4159.2007.04841.x.
- 1197 Venkatesan, R., Ji, E., and Kim, S. Y. (2015). Phytochemicals That Regulate Neurodegenerative 1198 Disease by Targeting Neurotrophins: A Comprehensive Review. *Biomed Res. Int.* 2015, 1–
- 1199 22. doi:10.1155/2015/814068.
- 1200 Visioli, F., Bellomo, G., and Galli, C. (1998). Free radical-scavenging properties of olive oil 1201 polyphenols. *Biochem. Biophys. Res. Commun.* 247, 60–64. doi:10.1006/bbrc.1998.8735.
- 1202 Vivancos, M., and Moreno, J. J. (2002). Role of Ca2+-independent phospholipase A2 and

1203 cyclooxygenase/lipoxygenase pathways in the nitric oxide production by murine

- 1204 macrophages stimulated by lipopolysaccharides. *Nitric Oxide Biol. Chem.* 6, 255–262. 1205 doi:10.1006/niox.2001.0410.
- 1206 Vivancos, M., and Moreno, J. J. (2005). β-Sitosterol modulates antioxidant enzyme response in 1207 RAW 264.7 macrophages. *Free Radic. Biol. Med.* 39, 91–97. 1208 doi:10.1016/j.freeradbiomed.2005.02.025.
- 1209 Vivancos, M., and Moreno, J. J. (2008). Effect of resveratrol, tyrosol and β-sitosterol on oxidised 1210 low-density lipoprotein-stimulated oxidative stress, arachidonic acid release and
- 1211 prostaglandin E2 synthesis by RAW 264.7 macrophages. *Br. J. Nutr.* 99, 1199–1207.
- 1212 doi:10.1017/S0007114507876203.
- 1213 Walker, J. M., Klakotskaia, D., Ajit, D., Weisman, G. A., Wood, W. G., Sun, G. Y., et al. (2015). 1214 Beneficial Effects of Dietary EGCG and Voluntary Exercise on Behavior in an Alzheimer's 1215 Disease Mouse Model. *J. Alzheimer's Dis.* 44, 561–572. doi:10.3233/JAD-140981.
- 1216 Wang, J., Xiong, T., Zhou, J., He, H., Wu, D., Du, X., et al. (2018). Enzymatic formation of
- 1217 curcumin in vitro and in vivo. *Nano Res.* 11, 3453–3461. doi:10.1007/s12274-018-1994-z.
- 1218 Wang, K., Chen, Q., Wu, N., Li, Y., Zhang, R., Wang, J., et al. (2019). Berberine Ameliorates 1219 Spatial Learning Memory Impairment and Modulates Cholinergic Anti-Inflammatory 1220 Pathway in Diabetic Rats. *Front. Pharmacol.* 10, 1003. doi:10.3389/fphar.2019.01003.
- 1221 Wang, M. J., Huang, H. M., Hsieh, S. J., Jeng, K. C., and Kuo, J. S. (2001). Resveratrol inhibits 1222 interleukin-6 production in cortical mixed glial cells under hypoxia/hypoglycemia followed 1223 by reoxygenation. *J. Neuroimmunol.* 112, 28–34. doi:10.1016/s0165-5728(00)00374-x.
- 1224 Wang, Q., Xu, J., Rottinghaus, G. E., Simonyi, A., Lubahn, D., Sun, G. Y., et al. (2002).
- 1225 Resveratrol protects against global cerebral ischemic injury in gerbils. *Brain Res.* 958, 439–
- 1226 47. doi:10.1016/s0006-8993(02)03543-6.
- 1227 Wang, R., Li, Y. H., Xu, Y., Li, Y. B., Wu, H. L., Guo, H., et al. (2010). Curcumin produces 1228 neuroprotective effects via activating brain-derived neurotrophic factor/TrkB-dependent 1229 MAPK and PI-3K cascades in rodent cortical neurons. *Prog. Neuro-Psychopharmacology* 1230 *Biol. Psychiatry* 34, 147–153. doi:10.1016/j.pnpbp.2009.10.016.
- 1231 Wang, X., Wang, R., Xing, D., Su, H., Ma, C., Ding, Y., et al. (2005). Kinetic difference of 1232 berberine between hippocampus and plasma in rat after intravenous administration of 1233 Coptidis rhizoma extract. *Life Sci.* 77, 3058–3067. doi:10.1016/j.lfs.2005.02.033.
- 1234 Wei, B. bin, Liu, M. yan, Zhong, X., Yao, W. fan, and Wei, M. jie (2019). Increased BBB 1235 permeability contributes to EGCG-caused cognitive function improvement in natural aging 1236 rats: pharmacokinetic and distribution analyses. *Acta Pharmacol. Sin.* 40, 1490–1500. 1237 doi:10.1038/s41401-019-0243-7.
- 1238 Wiciński, M., Malinowski, B., Węclewicz, M. M., Grześk, E., and Grześk, G. (2017).
- 1239 Resveratrol Increases Serum BDNF Concentrations and Reduces Vascular Smooth Muscle
- 1240 Cells Contractility via a NOS-3-Independent Mechanism. *Biomed Res. Int.* 2017, 9202954. 1241 doi:10.1155/2017/9202954.
- 1242 Williams, R. J., Spencer, J. P. ., and Rice-Evans, C. (2004). Flavonoids: antioxidants or
- 1243 signalling molecules? *Free Radic. Biol. Med.* 36, 838–849.
- 1244 doi:10.1016/j.freeradbiomed.2004.01.001.
- 1245 Winner, B., Kohl, Z., and Gage, F. H. (2011). Neurodegenerative disease and adult neurogenesis. 1246 *Eur. J. Neurosci.* 33, 1139–1151. doi:10.1111/j.1460-9568.2011.07613.x.
- 1247 Wobst, H. J., Sharma, A., Diamond, M. I., Wanker, E. E., and Bieschke, J. (2015). The green tea
- 1248 polyphenol (−)-epigallocatechin gallate prevents the aggregation of tau protein into toxic 1249 oligomers at substoichiometric ratios. *FEBS Lett.* 589, 77–83. 1250 doi:10.1016/j.febslet.2014.11.026.
- 1251 Wu, Y., Li, X., Zhu, J. X., Xie, W., Le, W., Fan, Z., et al. (2011). Resveratrol-Activated 1252 AMPK/SIRT1/Autophagy in Cellular Models of Parkinson's Disease. *Neurosignals* 19, 1253 163–174. doi:10.1159/000328516.
- 1254 Xia, S. F., Xie, Z. X., Qiao, Y., Li, L. R., Cheng, X. R., Tang, X., et al. (2015). Differential 1255 effects of quercetin on hippocampus-dependent learning and memory in mice fed with 1256 different diets related with oxidative stress. *Physiol. Behav.* 138, 325–331.
- 1257 doi:10.1016/j.physbeh.2014.09.008.
- 1258 Xudong Huang, ‡, Craig S. Atwood, ‡, Mariana A. Hartshorn, ‡, Gerd Multhaup, §, Lee E. 1259 Goldstein, ‡, Richard C. Scarpa, ‡, et al. (1999). The Aβ Peptide of Alzheimer's Disease 1260 Directly Produces Hydrogen Peroxide through Metal Ion Reduction. 1261 doi:10.1021/BI990438F.
- 1262 Yan, W.-J., Liu, R.-B., Wang, L.-K., Ma, Y.-B., Ding, S.-L., Deng, F., et al. (2018). Sirt3- 1263 Mediated Autophagy Contributes to Resveratrol-Induced Protection against ER Stress in 1264 HT22 Cells. *Front. Neurosci.* 12. doi:10.3389/fnins.2018.00116.
- 1265 Yang, F., Lim, G. P., Begum, A. N., Ubeda, O. J., Simmons, M. R., Ambegaokar, S. S., et al. 1266 (2005). Curcumin inhibits formation of amyloid β oligomers and fibrils, binds plaques, and 1267 reduces amyloid in vivo. *J. Biol. Chem.* 280, 5892–5901. doi:10.1074/jbc.M404751200.
- 1268 Yang, J., Song, S., Li, J., and Liang, T. (2014). Neuroprotective effect of curcumin on 1269 hippocampal injury in 6-OHDA-induced Parkinson's disease rat. *Pathol. - Res. Pract.* 210, 1270 357–362. doi:10.1016/j.prp.2014.02.005.
- 1271 Yang, J., Yan, H., Li, S., and Zhang, M. (2018). Berberine Ameliorates MCAO Induced Cerebral 1272 Ischemia/Reperfusion Injury via Activation of the BDNF–TrkB–PI3K/Akt Signaling 1273 Pathway. *Neurochem. Res.* 43, 702–710. doi:10.1007/s11064-018-2472-4.
- 1274 Yazir, Y., Utkan, T., Gacar, N., and Aricioglu, F. (2015). Resveratrol exerts anti-inflammatory
- 1275 and neuroprotective effects to prevent memory deficits in rats exposed to chronic
- 1276 unpredictable mild stress. *Physiol. Behav.* 138, 297–304.
- 1277 doi:10.1016/j.physbeh.2014.10.010.
- 1278 Yoritaka, A., Hattori, N., Uchida, K., Tanaka, M., Stadtman, E. R., and Mizuno, Y. (1996).
- 1279 Immunohistochemical detection of 4-hydroxynonenal protein adducts in Parkinson disease.
- 1280 *Proc. Natl. Acad. Sci. U. S. A.* 93, 2696–2701. doi:10.1073/pnas.93.7.2696.
- 1281 Yu, S., Zheng, W., Xin, N., Chi, Z.-H., Wang, N.-Q., Nie, Y.-X., et al. (2010). Curcumin 1282 Prevents Dopaminergic Neuronal Death Through Inhibition of the c-Jun N-Terminal Kinase 1283 Pathway. *Rejuvenation Res.* 13, 55–64. doi:10.1089/rej.2009.0908.
- 1284 Yuan, J., Liu, W., Zhu, H., Zhang, X., Feng, Y., Chen, Y., et al. (2017). Curcumin attenuates 1285 blood-brain barrier disruption after subarachnoid hemorrhage in mice. *J. Surg. Res.* 207, 1286 85–91. doi:10.1016/j.jss.2016.08.090.
- 1287 Zaplatic, E., Bule, M., Shah, S. Z. A., Uddin, M. S., and Niaz, K. (2019). Molecular mechanisms 1288 underlying protective role of quercetin in attenuating Alzheimer's disease. *Life Sci.* 224, 1289 109–119. doi:10.1016/j.lfs.2019.03.055.
- 1290 Zbarsky, V., Datla, K. P., Parkar, S., Rai, D. K., Aruoma, O. I., and Dexter, D. T. (2005).
- 1291 Neuroprotective properties of the natural phenolic antioxidants curcumin and naringenin but 1292 not quercetin and fisetin in a 6-OHDA model of Parkinson's disease. *Free Radic. Res.* 39,
- 1293 1119–1125. doi:10.1080/10715760500233113.
- 1294 Zhang, C., Browne, A., Child, D., and Tanzi, R. E. (2010a). Curcumin Decreases Amyloid-β 1295 Peptide Levels by Attenuating the Maturation of Amyloid-β Precursor Protein. *J. Biol.* 1296 *Chem.* 285, 28472–28480. doi:10.1074/jbc.M110.133520.
- 1297 Zhang, F., Lu, Y.-F., Wu, Q., Liu, J., and Shi, J.-S. (2012a). Resveratrol promotes neurotrophic 1298 factor release from astroglia. *Exp. Biol. Med.* 237, 943–948. doi:10.1258/ebm.2012.012044.
- 1299 Zhang, F., Lu, Y. F., Wu, Q., Liu, J., and Shi, J. S. (2012b). Resveratrol promotes neurotrophic 1300 factor release from astroglia. *Exp. Biol. Med.* 237, 943–948. doi:10.1258/ebm.2012.012044.
- 1301 Zhang, F., Shi, J.-S., Zhou, H., Wilson, B., Hong, J.-S., and Gao, H.-M. (2010b). Resveratrol

1329

1330 **Figure Legends**

1331 **Figure 1: The role of oxidative stress in the pathogenesis of neurodegenerative diseases.** 1332 Several events like environmental factors, mitochondrial dysfunction as well as cellular 1333 metabolic process are responsible for oxidative stress. Protein misfolding and aggregation are 1334 common hallmarks of several neurodegenerative disorders. Oxidative stress activates PP2A, 1335 p38MAPK, and JNK that lead to increase expression of neurotoxic Aβ and tau (hallmarks of 1336 Alzheimer's disease) that further increase cellular levels of ROS and RNS and results in 1337 neurodegeneration. Furthermore, oxidative stress causes neuronal dysfunction by atypical 1338 misfolding and aggregation of α-synuclein (for Parkinson's disease), mHtt (for Huntington's 1339 disease), and TDP-43 for (amyotrophic lateral sclerosis) by augmenting ROS and RNS levels. 1340 ROS, reactive oxygen species; RNS, reactive nitrogen species; PP2, protein phosphatase 2; 1341 p38MAPK, mitogen-activated protein kinase p38; JNK, c-Jun N-terminal kinase; mHtt, mutant 1342 huntingtin; TDP-43, TAR DNA binding protein 43; NOX, NADPH oxidase; NFTs,

1344

1343 neurofibrillary tangles.

1345 **Figure 2: Mechanism of oxidative stress-mediated neuronal apoptosis and**

1346 **neurodegeneration.** A number of pathogenic intracellular events like endoplasmic reticulum 1347 stress and mitochondrial dysfunction cause oxidative stress by increasing Ca^{2+} influx and 1348 subsequent neuronal excitotoxicity. Excessive production of reactive species causes oxidation of 1349 proteins, lipids, and nucleic acids that lead to neurodegeneration. Oxidative stress causes 1350 activation of the p53 apoptotic pathway leads to the expression of Noxa, Bax, and Puma. The 1351 expression of Puma causes the release of cytosolic p53 that held inactive by binding with Bcl-xL. 1352 P53 and Bcl-xL complex cause Bax oligomerization and mitochondrial translocation. 1353 Accumulation of intracellular p53 serves as a major source of mitochondrial p53 that induces 1354 Bax and Bak oligomerization as well as antagonize the antiapoptotic effect of Bcl-2 and Bcl-xL 1355 in the mitochondria. As a result, mitochondrial cytochrome c is released that stimulates the 1356 activation of caspase pathways that lead to neuronal apoptosis and finally neurodegeneration. 1357 p53, tumor suppressor protein p53; 8-OHG, 8-hydroxyguanosine; 8-OHdG, 8-hydroxy-2′- 1358 deoxyguanosine; 4-HNE, 4-hydroxy-2-nonenal; MDA, malondialdehyde; Noxa, phorbol-12- 1359 myristate-13-acetate-induced protein 1; Puma, p53 upregulated modulator of apoptosis; Bax,

1360 Bcl-2-associated X protein, Bak, Bcl-2 antagonist/killer; Bcl-2, B-cell lymphoma 2; Bcl-xL, B-1361 cell lymphoma-extra-large.

1362

1363 **Figure 3: Role of polyphenols involved with neurotrophins to alleviate the progression of** 1364 **neurodegenerative diseases by activating the TrkB receptor.** Polyphenols or neurotrophins 1365 especially BDNF binds to the TrkB receptor, which activates PI3K/Akt, Ras/ERK 1/2 and PL-Cγ 1366 pathways and eventually phosphorylates CREB protein. Subsequently, CREB increases the 1367 transcription of target genes coding NTFs, Bcl-2 and antioxidant genes that regulate cell survival 1368 and ultimately inhibit neurodegeneration. Polyphenols also trigger Keap-Nrf2-ARE cascade and 1369 increase the expression of antioxidant enzymes and proteins, which play an essential role in 1370 increasing the neuroprotective activity. In addition, polyphenols also inhibit the ROS and GSK3β 1371 that leads to neurodegeneration. Trk, tropomyosin-related kinase; BDNF, brain-derived 1372 neurotrophic factor; PI3K, phosphatidylinositol-3-kinase; MEK, MAP kinase kinase; MAPK, 1373 mitogen-activated protein kinase; ERK, extracellular signal-regulated kinases; PL-Cγ, 1374 phospholipase Cγ; PKC, protein kinase C; CaMKII/IV, Ca²⁺-calmodulin kinase II/IV; Nfr2, 1375 nuclear factor erythroid 2-related factor 2; CREB, cyclic adenosine monophosphate response 1376 element-binding protein; CRE, cyclic adenosine monophosphate response elements; GSK3β, 1377 glucose synthase kinase-3β; ROS, reactive oxygen species; Akt, protein kinase B; Ras, a small 1378 GTPase; Keap1, Kelch-like ECH associated protein; ARE, antioxidant response element; NTFs, 1379 neurotrophic factors; Bcl-2, B-cell lymphoma 2. 1380 1381 **Figure 4: Chemical structures of several neuroprotective polyphenols.** 1382 1383 1384 **Tables** 1385 **Table 1: The blood-brain barrier permeability of different polyphenols for** 1386 **neuroprotection.** 1387 1388 **Table 2: The effects of different polyphenols on neurotrophin factors to combat** 1389 **neurodegeneration.** 1390

Table 1: The blood-brain barrier permeability of different polyphenols for neuroprotection.

Table 2: The effects of different polyphenols on neurotrophin factors to combat neurodegeneration.

pathway

