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## Post-lockdown detection of SARS-CoV-2 RNA in the wastewater of Montpellier, France

Julie Trottier<sup>1,\*</sup>, Regis Darques<sup>1</sup>, Nassim Ait Mouheb<sup>2,3</sup>, Emma Partiot<sup>2,4</sup>, William Bakhache<sup>4</sup>, Maïka S. Deffieu<sup>2,4</sup>, Raphael Gaudin<sup>2,4,\*</sup>.

### Affiliations:

<sup>1</sup> CNRS, ART-Dev, Site Saint-Charles, rue du Professeur Henri Serre, 34090 Montpellier, France

<sup>2</sup> Université de Montpellier, 163 rue Auguste Broussonnet – 34 090 Montpellier, France

<sup>3</sup> INRAE, UMR G-eau, 361 rue Jean-François Breton, 34196 Montpellier cedex 5, France

<sup>4</sup> CNRS, Institut de Recherche en Infectiologie de Montpellier (IRIM), 1919 route de Mende, 34293 Montpellier, France

\* Co-corresponding authors:

Julie Trottier: [Julie.trottier@cnrs.fr](mailto:Julie.trottier@cnrs.fr) and Raphael Gaudin [Raphael.gaudin@irim.cnrs.fr](mailto:Raphael.gaudin@irim.cnrs.fr)

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### Abstract

The evolution of the COVID-19 pandemic can be monitored through the detection of SARS-CoV-2 RNA in sewage. Here, we measured the amount of SARS-CoV-2 RNA at the inflow point of the main waste water treatment plant (WWTP) of Montpellier, France. We collected samples 4 days before the end of lockdown and up to 70 days post-lockdown. We detected increased amounts of SARS-CoV-2 RNA at the WWTP from mid-June on, whereas the number of new COVID-19 cases in the area started increasing a couple of weeks later. Future epidemiologic investigations shall explain such asynchronous finding.

## Main text

SARS-CoV-2 is the etiologic agent responsible for the current coronavirus disease 2019 (COVID-19) pandemic. Wastewater-based epidemiology represents an attractive strategy to surveil the evolution of virus circulation in populations (Carducci et al., 2020, O'Brien & Xagorarakis, 2019), contributing to cost-effective virus control without infringing on individual liberties. About half of symptomatic patients persistently shed SARS-CoV-2 RNA in their feces at levels going up to  $10^8$  RNA copies per stool sample (Chen et al., 2020, Wang et al., 2020, Wolfel et al., 2020, Wu et al., 2020, Xiao et al., 2020, Xu et al., 2020), suggesting that a single patient can shed billions of SARS-CoV-2 RNA copies in the wastewater at a time. Moreover, an asymptomatic child was recently reported as negative for SARS-CoV-2 RNA based on throat swab specimen, while his stools were positive (Tang et al., 2020), which means that symptomatic and asymptomatic persons are likely to release SARS-CoV-2 RNA in city sewerages. Of note, subgenomic viral RNA was not detected in stools (Wolfel et al., 2020), e.g. no infectious virus is shed (although this finding remains to be further investigated).

Several reports indicate that SARS-CoV-2 RNA was readily detected in wastewater, and it is proposed that such approach could anticipate the occurrence of novel COVID-19 outbreaks in low prevalence regions (Ahmed et al., 2020, La Rosa et al., 2020, Medema et al., 2020, Orive et al., 2020, Randazzo et al., 2020, Wurtzer et al., 2020). The end of the stringent lockdown (that occurred in France as of May 11<sup>th</sup>) is therefore an adequate time to measure the re-emergence of the virus through the monitoring of wastewater. Here, we collected effluent composite samples (using a 24-hour automatic sampler) in wastewater upstream of the main waste water treatment plant (WWTP) of Montpellier metropolitan area located in Lattes, France, which receives the wastewater from  $\approx$  470 000 inhabitants. The sampling dates were May 7<sup>th</sup>, 18<sup>th</sup>, 26<sup>th</sup>, June 4<sup>th</sup>, 15<sup>th</sup>, 25<sup>th</sup>, and July 20<sup>th</sup> to monitor SARS-CoV-2 RNA expression levels during lockdown and up to 70 days after its end. After the period high was reached at the beginning of April, the number of inpatients was decreasing gradually. However, the virus was still circulating in the area, and even more importantly with the upsurge of seasonal tourist movements.

Collected wastewater was processed as follows: on the day of water collection, samples were maintained at 4°C for transport and immediately cleared by centrifugation at 4500 g for 30 min at 4°C. The supernatant was passed through a 40  $\mu$ m cell strainer (Corning) to remove large floating components. At this stage, the samples were frozen at -20°C for later analyses with samples collected at other timepoints. Upon thawing, RNAs were concentrated on a Vivaspin 50 kDa MWCO filter membrane (Sartorius). Starting from 50 ml of water, the sample was concentrated down a hundred times to an adjusted volume of 500  $\mu$ l. RNA extraction was performed using the NucleoSpin RNA Virus kit (Macherey-Nagel), including harsh lysis conditions (lysis buffer enriched in guanidinium isothiocyanate heat at 70°C for 5 min). RT-qPCR was performed on 10  $\mu$ l of purified RNA using the highly sensitive TaqPath One-Step RT-qPCR, CG master mix (ThermoFisher Scientific). The N1 and N3 primer/probe sets designed by the center for disease control (CDC) were used to detect SARS-CoV-2 RNA and a standard curve was run in parallel using a positive control plasmid (Integrated DNA Technologies) coding for the nucleoprotein (N) of SARS-CoV-2. Using RNA extracted from Vero E6 cells either non-infected or infected with SARS-CoV-2 *in vitro*, we showed that the N1 and N3 primer/probe sets recognized solely the RNA from infected cells (Table 1). A recent study used a Dengue virus (DENV) sequence surrogate to determine PCR efficiency (Medema et al., 2020) but in Montpellier area, this approach would be risky, as autochthonous DENV infections have been repeatedly reported in the south of France (European Centre for Disease Prevention and Control). In order to determine the RT-qPCR efficiency intrinsic to each sample, we used a sensitive primer/probe set previously described (Ro et al., 2017) that target the VP40-encoding RNA of Ebola

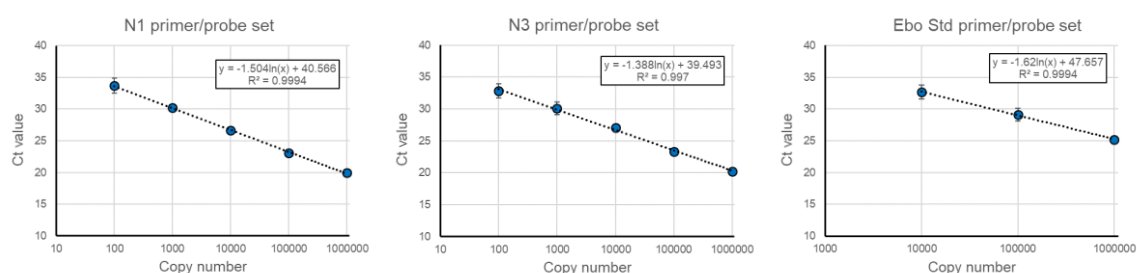
virus (Zaire strain). First, we showed that the Ebola standard (Ebo Std) primer/probe set was not detecting RNA from SARS-CoV-2-infected Vero E6 cells (Table 1). Using water samples from upstream WWTP of the Montpellier metropolitan area collected on June 15<sup>th</sup>, we found that the Ebo Std primer/probe set gave no signal while the primer/probes N1, N3, and RLP27 (targeting the human *rlp27* gene) returned positive signals (Table 1). In comparison, the nearby WWTP of Murviel-lès-Montpellier, treating the wastewater from 2 000 inhabitants, and a Montpellieran household in which no one was sick nor had symptoms, were negative for SARS-CoV-2 RNA on May 29, 2020.

**Table 1. Specificity of the primer/probe sets from *in vitro* SARS-CoV-2 infected cells.** The Table shows the cycle threshold (Ct) of individual RT-qPCR reaction. The data are representative of experiments performed at least in duplicates. “No Ct” indicates that no signal was detected over 40 cycles. nt: not tested.

Primer/probe set	N1	N3	Ebo Std	RLP27
RNA from non-infected cells	No Ct	No Ct	No Ct	nt
RNA from SARS-CoV-2 infected cells	19	17.4	No Ct	nt
Water samples collected upstream of the Montpellier WWTP	36.4	37.6	No Ct	30.9
	No Ct	37.3	No Ct	30.2
Water samples collected upstream of the Murviel-lès-Montpellier WWTP	No Ct	nt	nt	39.0
	No Ct	nt	nt	34.5
Sample collected in Montpellier household wastewater	No Ct	nt	nt	24.6
	No Ct	nt	nt	27.5

A synthetic RNA sequence of 91 nucleotides (gcagaucgaauccacucaggccaaucauggugaauugucauacggggcccaagugcuaaagaaguuuggcuuccucaggugucggcag) derived from the VP40-coding gene of Ebola virus and recognized by the above-mentioned Ebo Std primer/probe set was purchased (Integrated DNA Technologies) to be used as an internal normalization standard. Standard curves were run using the N control plasmid (N1 and N3 primer/probe sets) or the Ebo Std RNA (Ebo Std primer/probe set) to estimate copy numbers (Figure 1). Of note, the Ebo Std primer/probe set was less sensitive than N1 and N3, probably because the Ebo Std synthetic template RNA is relatively fragile and short (91 nucleotides).

**Figure 1.** Standard curves of the N1 (left), N3 (middle) and Ebo Std (right) primer/probe sets showing the cycle threshold (Ct) value at indicated template copy number. The mean +/- SD of duplicates from a representative experiment is shown.

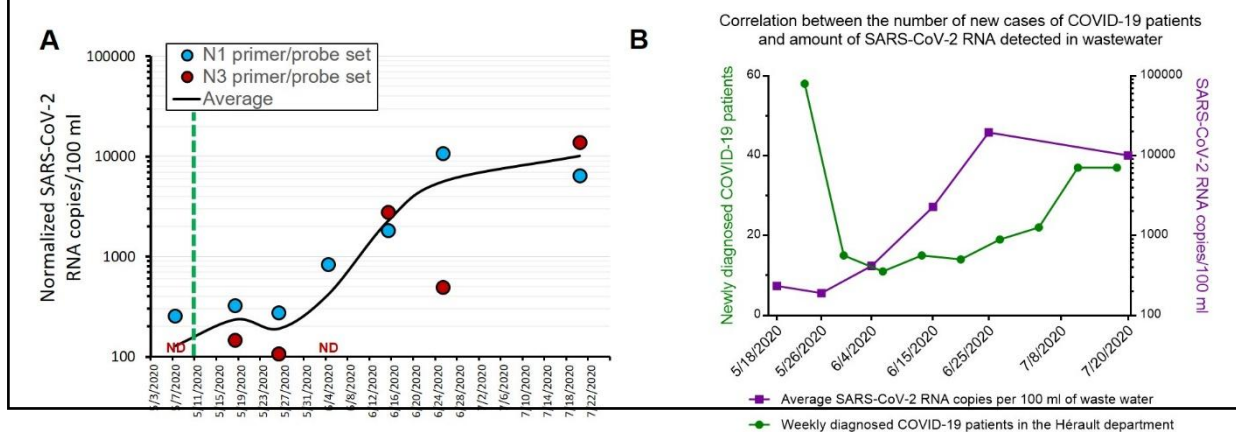


Next, we measured the SARS-CoV-2 RNA levels using N1 and N3 primer/probe sets in wastewater collected upstream of the main WWTP of the Montpellier metropolitan area on May 7<sup>th</sup>, 18<sup>th</sup>, 26<sup>th</sup>, June 4<sup>th</sup>, 15<sup>th</sup>, 25<sup>th</sup>, and July 20<sup>th</sup> corrected to PCR efficiency level calculated for each sample (Figure 2A). Our data highlights that the wastewater from the three later dates (June 15<sup>th</sup>, 25<sup>th</sup>, and July 20<sup>th</sup>) contained about fifty-fold more SARS-CoV-2 RNA than from previous dates. Of note, the Montpellier wastewater network is partially unitary (part of it channels runoff water together with sewage) and thus, we checked that rain precipitation was negligible (between 0 and 2 mm of rain on the 24 h preceding sample collection. Similarly, inlet water flowrate entering the WWTP was varying by less than 25% from sample to sample and thus, could not explain the magnitude of the SARS-CoV-2 RNA increase we observed (Figure 2A).

We then put in perspective the amount of SARS-CoV-2 RNA in Montpellier wastewater with the number of new COVID-19 cases weekly recorded in the Hérault department (> 40% inhabitant living in the Montpellier metropolitan area). Interestingly, a mild increase in the number of newly tested COVID-19 patients was observed in early July (a figure up to three times higher than in May). This surge occurred roughly 2-3 weeks after the increase of SARS-CoV-2 RNA levels in wastewater (Figure 2B). Our data are reminiscent of a recent Spanish study, in which the authors could detect SARS-CoV-2 RNA in wastewater several weeks before the first COVID-19 cases were reported (Randazzo et al., 2020). However, they could not see a correlation between SARS-CoV-2 RNA levels in wastewater and the number of newly diagnosed COVID-19 patients. On the same line, Medema & colleagues showed a correlation between the cumulative cases of COVID-19 and SARS-CoV-2 RNA although the data were not correlated as a function of time (Medema et al., 2020). Other work in Paris, France, is ongoing to more finely determine the temporal correlation between wastewater SARS-CoV-2 RNA levels and COVID-19 epidemiological features (Wurtzer et al., 2020). Moreover, it should be noted that only 1000-1500 people are tested daily in the department (regular working days), which accounts for 0.15% of local population (1.1 M, without considering tourist flows).

In conclusion, we report effective detection of SARS-CoV-2 RNA in the wastewater of Montpellier area upstream of the treatment plant and identified an increase of the amount of detected viral RNA mid-June, associated with a rising number of newly identified COVID-19 cases in the department. Although a delayed correlation may exist, further investigations are required to better characterize the intricate relationship between these two variables. Indeed, we are unable at this stage to determine whether this increase announces an upcoming increase of hospital admissions in the area or relates to intrinsic SARS-CoV-2 RNA variations associated with uneven virus shedding (from patient-to-patient and depending on the stage of the disease for a given patient). Moreover, various other parameters might also impact these results, such as people from distant clusters moving to second homes and tourist accommodation, the chronic underestimation of prevalence rates, or local variability in the geographical pattern of virus spread. These hypotheses are non-exclusive and future multiparametric investigations are required to routinely use wastewater surveillance as a powerful predictive tool for ongoing and future epidemic outbreaks.

**Figure 2. SARS-CoV-2 RNA detection in the Montpellier wastewater and number of COVID-19 cases.** (A) The number of SARS-CoV-2 RNA copies was measured using either the N1 or N3 primer/probe sets. Each dot corresponds to the concentration of SARS-CoV-2 RNA in wastewater measured from the sum of the RNA copy number calculated from two RNA extractions and four RT-qPCR reactions performed in two individual runs. The first five samples were treated in parallel and therefore, differences in these samples cannot be attributed to experimental variation. Sterile water wells were included in duplicates for each primer/probe set in each plate as negative control and returned “No Ct” (not shown on the graph). Each datapoint was normalized to consider variability of PCR efficiency. However, precipitation and flowrates were negligible and not considered in the calculation. The green dotted line indicates the end of the strict lockdown in France. The black line corresponds to the smoothed average of the N1 and N3 primer/probe sets. ND: not detected. (B) The graph shows the a number of new positive patients tested in the Hérault department over a 7-day period (green; left axis) as a function of time (source: SI-DEP, Santé Publique France) and on the right Y axis, the average SARS-CoV-2 RNA levels measured with the N1 and N3 primer/probe sets (purple). Of note, the outlier number of cases on week 17-23 May is surprisingly high, which could be linked to readjustments following the creation of a new virological testing database as of May 13. This figure is contradicted by the stable relatively low number of positive tests recorded in the first statistical database ending May 29.



**Authors' contributions:** J.T established our consortium and initiated the project. J.T and R.G designed the study. R.D provided epidemiological datasets and geomatics expertise. N.A.M provided the rain precipitation dataset and hydrology expertise. E.P, W.B, M.D and R.G performed and optimized the technical experimentations. R.G wrote the manuscript and J.T, R.D, N.A.M and W.B edited and commented on the manuscript. All authors agree on the final version of the manuscript.

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## References

- Ahmed W, Angel N, Edson J, Bibby K, Bivins A, O'Brien JW, Choi PM, Kitajima M, Simpson SL, Li J, Tschärke B, Verhagen R, Smith WJM, Zaugg J, Dierens L, Hugenholz P, Thomas KV, Mueller JF (2020) First confirmed detection of SARS-CoV-2 in untreated wastewater in Australia: A proof of concept for the wastewater surveillance of COVID-19 in the community. *Sci Total Environ* 728: 138764
- Carducci A, Federigi I, Liu D, Thompson JR, Verani M (2020) Making Waves: Coronavirus detection, presence and persistence in the water environment: State of the art and knowledge needs for public health. *Water Res* 179: 115907
- Chen Y, Chen L, Deng Q, Zhang G, Wu K, Ni L, Yang Y, Liu B, Wang W, Wei C, Yang J, Ye G, Cheng Z (2020) The presence of SARS-CoV-2 RNA in the feces of COVID-19 patients. *Journal of medical virology* 92: 833-840
- La Rosa G, Iaconelli M, Mancini P, Bonanno Ferraro G, Veneri C, Bonadonna L, Lucentini L, Suffredini E (2020) First detection of SARS-CoV-2 in untreated wastewaters in Italy. *Sci Total Environ* 736: 139652
- Medema G, Heijnen L, Elsinga G, Italiaander R, Brouwer A (2020) Presence of SARS-Coronavirus-2 RNA in Sewage and Correlation with Reported COVID-19 Prevalence in the Early Stage of the Epidemic in The Netherlands. *Environmental Science & Technology Letters* 0: null
- O'Brien E, Xagoraki I (2019) A water-focused one-health approach for early detection and prevention of viral outbreaks. *One Health* 7: 100094
- Orive G, Lertxundi U, Barcelo D (2020) Early SARS-CoV-2 outbreak detection by sewage-based epidemiology. *Sci Total Environ* 732: 139298
- Randazzo W, Truchado P, Cuevas-Ferrando E, Simon P, Allende A, Sanchez G (2020) SARS-CoV-2 RNA in wastewater anticipated COVID-19 occurrence in a low prevalence area. *Water Res* 181: 115942
- Ro YT, Ticer A, Carrion R, Jr., Patterson JL (2017) Rapid detection and quantification of Ebola Zaire virus by one-step real-time quantitative reverse transcription-polymerase chain reaction. *Microbiol Immunol* 61: 130-137
- Tang A, Tong ZD, Wang HL, Dai YX, Li KF, Liu JN, Wu WJ, Yuan C, Yu ML, Li P, Yan JB (2020) Detection of Novel Coronavirus by RT-PCR in Stool Specimen from Asymptomatic Child, China. *Emerging infectious diseases* 26: 1337-1339
- Wang W, Xu Y, Gao R, Lu R, Han K, Wu G, Tan W (2020) Detection of SARS-CoV-2 in Different Types of Clinical Specimens. *JAMA*
- Wolfel R, Corman VM, Guggemos W, Seilmaier M, Zange S, Müller MA, Niemeyer D, Jones TC, Vollmar P, Rothe C, Hoelscher M, Bleicker T, Brunink S, Schneider J, Ehmann R, Zwirgmaier K, Drosten C, Wendtner C (2020) Virological assessment of hospitalized patients with COVID-2019. *Nature* 581: 465-469
- Wu Y, Guo C, Tang L, Hong Z, Zhou J, Dong X, Yin H, Xiao Q, Tang Y, Qu X, Kuang L, Fang X, Mishra N, Lu J, Shan H, Jiang G, Huang X (2020) Prolonged presence of SARS-CoV-2 viral RNA in faecal samples. *Lancet Gastroenterol Hepatol* 5: 434-435
- Wurtzer S, Marechal V, Mouchel J, Maday Y, Teyssou R, Richard E, Almayrac J, Moulin L (2020) Evaluation of lockdown impact on SARS-CoV-2 dynamics through viral genome quantification in Paris wastewaters. *MedRxiv*
- Xiao F, Sun J, Xu Y, Li F, Huang X, Li H, Zhao J, Huang J, Zhao J (2020) Infectious SARS-CoV-2 in Feces of Patient with Severe COVID-19. *Emerging infectious diseases* 26
- Xu Y, Li X, Zhu B, Liang H, Fang C, Gong Y, Guo Q, Sun X, Zhao D, Shen J, Zhang H, Liu H, Xia H, Tang J, Zhang K, Gong S (2020) Characteristics of pediatric SARS-CoV-2 infection and potential evidence for persistent fecal viral shedding. *Nature medicine* 26: 502-505