

# Neural encoding of time in the animal brain

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1	Neuroscience and Biobehavioral Reviews
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3	<u>Neural encoding of time in the animal brain</u>
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#### 13 Abstract

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The processing of temporal intervals is essential to create causal maps and to predict 15 future events so as to best adapt one's behavior. In this review, we explore the different 16 brain activity patterns associated with processing durations and expressing temporally-17 adapted behavior in animals. We begin by describing succinctly some of the current 18 models of the internal clock that can orient us in what to look for in brain activity. We 19 then outline how durations can be decoded by single cell activity and which activity 20 21 patterns could be associated with interval timing. We further point to similar patterns that have been observed at a more global level within brain areas (e.g. local field potentials) 22 or, even, between these areas, that could represent another way of encoding duration or 23 24 could constitute a necessary part for more complex temporal processing. Finally, we discuss to what extent neural data fit with internal clock models, and highlight 25 improvements for experiments to obtain a more in-depth understanding of the brain's 26 temporal encoding and processing. 27

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*Keywords (3-12)*: interval timing; electrophysiology; local field potentials; oscillations;
time cells; single unit recordings; multi-unit recordings; animal models

#### 31 Highlights

- Durations can be encoded in various patterns of single unit activity 32 • Population activity and communication between brain areas can also encode 33 • durations 34 Combination of unit and population activity may be the basis of our internal 35 clock 36 37 • Experiments designed to study the neural basis of interval timing are needed 38
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## 1. <u>Introduction</u>

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#### 1.1 Time and the internal clock

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A sense of time exists in most species, from drosophila to fish, pigeons, rats, 44 humans (for review, see Buhusi and Meck, 2005) and even honeybees (Craig et al., 2014). 45 It is essential for survival as it allows individuals to extract the predictability of events 46 47 and to adapt their behavior to respond at the appropriate time. We use our implicit sense of time to be able to cross the street while staying safe by estimating the speed and 48 49 time it takes for a car to reach us, or to know when to give up on a presumed broken 50 traffic light. Although we may be aware of how much time is passing by, we may not pay attention to the absolute duration of events. We can, however, be taught to explicitly pay 51 52 attention to how many seconds are passing between lightning and the associated thunder 53 clap to determine if we should hide or not. Both implicit and explicit timing are used in our daily life. Although crucial for complex cognitive functions, understanding how the 54 55 brain encodes durations remains an unsolved problem, especially because brain processing works at a millisecond range (e.g. one action potential lasts for around 5ms), 56 thus providing high precision in short timescales but creating a challenge to timing multi-57 58 seconds to minutes long durations.

59 While circadian rhythm (involved in hunger and sleep) - built around a 24h cycle 60 - is poorly adjustable (as exemplified by jetlag), interval timing - in the seconds to minutes 61 range - is flexible, learned, and covers a larger range of durations to allow for a rapid 62 adaptation to changes in the environment. In contrast to the well-described dependence 63 of circadian rhythm on the suprachiasmatic nucleus, many brain structures have been 64 linked to interval timing. On the other end of the timescale (in the range of a few hundred 65 milliseconds), there is motor timing, which is involved in the automatic synchronization and control of movements (Rammsayer, 1999). The cerebellum is often considered to be
the seat of motor timing because of its role in the processing and integration of multiple
sensory and somatosensory inputs, as well as the optimization of motor output (for a
review, see Spencer and Ivry, 2013). We will focus the present review on interval timing,
which enables organisms to create temporal maps and manage predictions about the
outcome of situations, and has therefore a strong cognitive component (Buhusi and Meck,
2005).

Most species, from insects to primates, process temporal information as if they 73 were using a stopwatch, suggesting the existence of a conserved function for an internal 74 75 clock across evolution (Buhusi and Meck, 2005; Church, 1984, 1978; Matell and Meck, 2000). Animals' internal clock seems to encode time in a linear fashion (Church, 1984; 76 77 Gibbon and Church, 1981) and can be used to time signals from different modalities in a 78 sequential or a simultaneous manner (Gibbon et al., 1984; Olton et al., 1988). The internal clock can also be stopped and reset, as shown in gap paradigms (where the insertion of a 79 80 "pause" in the timed stimulus induces a shift of the temporal behavior dependent on the duration of the "pause"), whether in an instrumental or Pavlovian, appetitive or aversive 81 condition (e.g. Aum et al., 2004; Church, 1984, 1978; Roberts and Church, 1978; Tallot 82 et al., 2016). 83

Treisman (1963) was one of the first to propose a model for this internal clock. He described three components: a pacemaker, an accumulator, and a memory stage. The beginning of a stimulus closes a switch between the pacemaker and accumulator which starts the accumulation of 'ticks' (i.e. temporal units) for the duration of the stimulus. At the end of the stimulus, the number of ticks accumulated is saved in the memory stage to be retrieved and compared with future durations. In most cases, interval timing follows the scalar property (i.e., Weber's law applied to time), that is, the precision with which a

given duration is estimated decreases in an inversely proportional manner to the length of
that duration. However, not all temporal behaviors follow the scalar property
whether in humans (Wearden and Lejeune, 2008) or in animals (Lejeune and
Wearden, 2006).

To account for the scalar property of time, storage and retrieval from memory are assumed imperfect. The scalar expectancy theory - the most influential timing model up to now - was developed by Gibbon in (1977), and further improved by Church in 1984. It expands on the memory stage of Treisman's internal clock by incorporating: a working memory component, a multiplicative factor for storage in a reference memory, and a decision rule to determine if 'yes' or 'no' the duration being measured is similar to previously encoded durations.

Our main question - "What are the potential neurobiological correlates of time?" - is therefore multifaceted, as we are looking for the different modules of our timing system, from the attention to duration to the retrieval of specific intervals, including the formation in memory of temporal associations between events. The literature we review here is not intended to be exhaustive, but rather to cover a large range of behavioral tasks and highlight various examples from the literature in order to bring together different angles of view on potential neural correlates of time processing.

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## 111 1.2 <u>Neurobiological basis of the internal clock</u>

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Although the scalar expectancy theory explains most behavioral results, it has not yet been supported at the neurobiological level. Pharmacological, lesion and human imaging studies have highlighted the importance of several brain areas in interval timing,

results that have been discussed many times before, and will not be part of this review. 116 Briefly, a few structures have been detected as playing a role in timing across a lot of 117 studies: the supplementary motor area (SMA), the pre-SMA, the prefrontal cortex (PFC), 118 the striatum, the substantia nigra, the inferior parietal cortex and the cerebellum (e.g. 119 Brannon et al., 2008; Buhusi and Meck, 2005; Coull et al., 2011; Harrington et al., 2010, 120 2004; Lewis and Miall, 2006; Wiener et al., 2010). For example, in Huntington's disease, 121 122 in which the striatum is degenerating, temporal deficits have been reported (Garces et al., 2018; Höhn et al., 2011; Paulsen et al., 2004; Rowe et al., 2010; Zimbelman et al., 2007). 123 Patients with Parkinson's disease show impaired timing in motor and sensory tasks, 124 125 linked with the degeneration of their dopaminergic neurons (e.g., Pastor et al., 1992). Pharmacological studies in animals confirm a role of dopamine in timing (Drew et al., 126 2003; Kim et al., 2016; Meck, 1996), although its function is complex (in part related to 127 128 its multi-structures targets), and interacts with motivational states (for a recent review, see Balc1, 2014). Meanwhile, the prefrontal cortex is critically involved in simultaneous 129 temporal processing, as shown in lesion studies in rodents (Meck and MacDonald, 2007; 130 Olton et al., 1988; Pang et al., 2001). 131

One of the core debates on the neurological basis of timing is whether it is dependent 132 133 on one central timing center, or whether timing is present all over the brain in separate clusters. One argument for a central clock is the fact that, in different tasks, there is a 134 similar temporal variance, at least for intervals larger than hundreds of milliseconds 135 (Gibbon et al., 1997). There is also a strong correlation between performance in self-136 paced timing tasks and duration discrimination, implying, again, the use of a common 137 timing mechanism (Keele et al., 1985). However there are also studies showing that brain 138 slices can encode durations (up to 2s) which implies that a very restricted amount of 139

connected neurons is sufficient for simple temporal encoding (Chubykin et al., 2013; Goeland Buonomano, 2016; Johnson et al., 2010).

Differences in neural encoding may relate to differences in the timing process 142 involved, e.g. whether it is of implicit or explicit nature. In implicit tasks, a subject's 143 knowledge of the durations experienced is not required for its performance, that is, the 144 subject does not have to time during the task but may do so nonetheless (like during 145 146 Pavlovian conditioning, working memory tasks and entrainment). In these tasks, accurate timing may facilitate detection of a stimulus or allow for a better regulation of behavior, 147 but is not necessary to perform. In explicit tasks, durations have to be processed for the 148 149 subject to respond accurately and learning of the duration is necessary (like in temporal 150 discrimination, temporal production and reproduction tasks). To what extent these two types of timing task rely on distinct or overlapping neural networks is still debated. 151 152 However, in humans at least, explicit timing seems to involve a fronto-striatal network (SMA, right inferior frontal cortex and basal ganglia) (Coull et al., 2013; Wiener et al., 153 154 2010) whereas implicit timing activates the left inferior parietal cortex and the right prefrontal cortex (Coull et al., 2000; Coull and Nobre, 1998; Vallesi et al., 2009). 155

To address the issue of the origin of the internal clock, assuming there is one, we 156 157 must look at neuronal activity from individual neurons to groups of neurons in a large range of brain areas and in different types of timing tasks. In vivo deep brain recording in 158 the awake behaving animal gives access to single cell and population neuronal activity 159 160 with high temporal resolution in any brain area during behavioral tasks with a large range of cues' duration. Patterns of single cell firing activity and synchronous spike activity of 161 neural ensembles could reflect a local processing of time. This synchronous cell activity 162 can generate depolarization/hyperpolarization oscillatory rhythms either locally (through 163 recurrent networks) or in distant brain areas, which can be recorded with local field 164

potentials (LFP). Neural oscillations also give access to subthreshold depolarization, 165 166 which may not be translated into spikes by the integrating neurons. As such, recordings of spike activity and oscillations provide complementary, non-fully-overlapping, 167 information. Neural oscillations are an ubiquitous property of brain function and have 168 important roles in learning, memory and cognitive processes such as those involved in 169 170 timing and time perception (for reviews, Buzsáki et al., 2013; Buzsáki and Draguhn, 2004; Hanslmayr and Staudigl, 2014; Matell and Meck, 2004). Slow (<50Hz) oscillations 171 are associated with large fluctuations of neurons' membrane potential and are considered 172 to cover large brain areas, whereas fast oscillations result from smaller fluctuations in 173 174 membrane potential and should be restricted to smaller neural volumes. Changes in oscillatory rhythms' frequency or power may represent timing function at a network level. 175 Here we have selected studies in the animal literature - not only in tasks designed to 176 177 study timing, but also in tasks studying other aspects of learning in which time was involved or manipulated - to see whether general principles emerge on how duration can 178 179 be encoded/decoded in the brain. We classify the studies between explicit and implicit timing tasks as a way to determine if they may rely on different neural systems or if they 180 belong to a common network that could represent 'pure' timing. To help the reader for 181 having a global view of the literature reviewed here, we summarize it in table 182 formats, organized around the type of tasks and neurophysiological correlates. 183

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187 Since the pioneering study by Fuster & Alexander (1971) which pointed to cells in 188 the prefrontal cortex modulating their firing during a delay in monkeys performing a 189 delayed response task, many different studies have been interested in determining how

2. Can time be encoded by a single neuron activity?

single neuron activity (i.e. spikes) can encode durations; and it is a growing field of 190 191 research, as about 30% of these studies have been published within the last five years. Indeed, how can an event of less than ten milliseconds encode durations of several 192 seconds to minutes? We are compiling here more than 80 studies that have in some way 193 looked at different patterns of single neuron firing that can represent time in explicit 194 (Table 1) or implicit (Table 2) temporal tasks. We have organized these studies according 195 196 to the type of task used, as well as the brain area where timing-related activity was reported. The studied species, as well as the range of durations used, are also mentioned. 197 We have categorized the modification of neurons' firing patterns, as compared to baseline 198 199 activity, in four types (see Figure 1 for a schematic representation of the different activity patterns): (1) sustained change, (2) phasic change in activity at the stimulus onset or 200 offset, whose amplitude and/or duration is proportional to the duration of the event, (3) 201 202 peak modulation at a specific time point ('event time' cells), and (4) ramping activity. The changes reported are in majority in the direction of an increased cell firing, but several 203 204 studies have also reported a decrease in cell firing, in particular when baseline levels of activity are high (e.g. Fuster and Alexander, 1973; Oshio et al., 2006). 205

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Sustained change in activity is often described in working memory tasks, and may represent the temporary maintenance in memory of a stimulus until a response has to be produced (e.g., Hampson and Deadwyler, 2003; Hikosaka et al., 1989; Narayanan and Laubach, 2009; Ohmae et al., 2008; Soltysik et al., 1975; Tremblay et al., 1998). Sustained increase or decrease in the number of spikes for the whole duration of a stimulus has been observed in 21 studies, in both implicit and explicit tasks. This pattern

2.1 Sustained change in cell's firing

has mostly been observed in the cortex, except for the four following studies. Pendyam 215 216 et al. (2013) reported sustained activity in the basal amygdala of rats in a Pavlovian aversive conditioning between the onset of a tone used as a conditioned stimulus (CS) 217 218 and the arrival of the footshock (used as an unconditioned stimulus, US). In Pavlovian associative tasks, animals not only learn the association but also memorize the 219 interval between the CS and US, whether they co-terminate, as in delay 220 221 conditioning, or are separated by a gap, as in trace conditioning (for a review, Balsam et al., 2010), and it has been suggested that the amygdala plays a role in 222 processing the CS-US interval (Díaz-Mataix et al., 2014). Soltysic et al. (1975) and 223 224 Hikosaka et al. (1989) observed this pattern in the basal ganglia of monkeys engaged in a working memory task, and Hampson and Deadwyler (2003) saw it in the subiculum of 225 226 rats in a similar task.

227 However, when the stimulus is present during the whole duration to be timed, it is difficult to differentiate a sustained change in activity due to the presence of the 228 229 stimulus, from one representing the online processing of duration. Circumventing this issue, Namboodiri et al (2015) have shown recently that the primary visual cortex can 230 encode time with sustained modulation in the absence of a continuous stimulus, allowing 231 232 us to better sort out sensory from temporal encoding. They used a task somewhat related to a DRL-LH (differential reinforcement of low rates with limited hold) task, in which a 233 brief (100 ms) visual stimulus delivered through goggles indicates the availability of a 234 reward for a fixed amount of time (1.5 s), but with increasing reward value (i.e. quantity 235 of liquid) as time passes. Thus, the rat has to perform in a temporally precise manner to 236 optimize the amount of reward. After a substantial amount of training, the rats developed 237 a stereotyped behavior with an optimal time of responding very close to the programmed 238 optimum (~1.1 s). The authors determined that 10% of the cells they recorded in the 239

primary visual cortex were timing units, as their sustained change in activity was highly correlated with the behavioral timing, but only on trials on which the action was correctly timed. Furthermore, the temporal information encoded by these neurons before the start of the action was predictive of timing behavior. Optogenetic modulation of primary visual cortical neurons' activity induced a shift in behavior to earlier responding, showing that the primary visual cortex participates in the temporal control of this highly stereotyped action.

Sustained activity does not seem sufficient to encode duration by itself, but could be 247 used as a pacemaker in an internal clock model with each spike representing a 'tick'. The 248 249 activity from the 'sustained' cells would need to be accumulated and transformed to be memorized, and then retrieved for comparison to previous durations. Therefore, it does 250 251 require other forms of activity, either from the same brain area or from others, to form a 252 complete internal clock. It may thus not be surprising that, most of the time, this type of activity was observed at the same time as other patterns of activity in the same brain area 253 254 (see Table 1 and 2).

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## 256 2.2 <u>Phasic response at onset or offset of the stimulus</u>

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Neuronal activity at either the onset or the offset of a stimulus may encode its duration through changes in the firing rate at its onset for representing its expected duration or at its offset for representing its passed duration. Such type of neural encoding has been observed for a large range of durations (from 1 to tens of seconds), and often when several durations are presented within the task, suggesting that it may have a role in differentiating durations (Chiba et al., 2015, 2008; Fiorillo et al., 2008; Jaramillo and Zador, 2011; Ohmae et al., 2008; Roux et al., 2003; Sakurai et al., 2004; Yumoto et al.,

2011). For example, Fiorillo et al. (2008) used a Pavlovian appetitive task where each of 265 266 four CSs predicted different intervals (from 1 to 8 seconds) between the CS onset and the US. The authors showed that dopaminergic neurons from the substantia nigra and ventral 267 tegmental area of trained monkeys respond less to the CS onset and more to the US for 268 longer CS-US intervals, and that the response to the US was also modulated when it was 269 delivered earlier or later than expected. The lawful relationship between US-evoked 270 271 responses and time is indicative of some underlying mechanisms of temporal encoding, and may be the expression of it, but is not the neural encoding of time per se. The 272 modulation of neural responses to the CS onset may, on the contrary, be decoded as the 273 274 length of time that the animal expects to wait before receiving the US, since one of the elements recalled when a CS is presented is the CS-US interval. The issue, though, is that 275 expectation of the reward includes also its salience, a reward closer in time being more 276 277 rewarding.

To isolate time encoding *per se* would necessitate a task without reward. Yumoto and 278 279 colleagues (2011) recorded activity in the PFC (area 9) of monkeys during a temporal reproduction task of a visual stimulus, in which the duration of the stimulus was not linked 280 to the wait for the reward, thus allowing separation between duration and salience. They 281 282 observed that some neurons showed phasic activity after the offset of a visual stimulus of a specific duration (neurons called "duration-recognizing"), whereas other neurons 283 showed sustained activity during the production of a specific duration (neurons called 284 "interval-generating"). Very rarely (less than 10%) a neuron had both roles. The authors 285 further demonstrated a role for area 9 of the PFC in timing by showing that an injection 286 of muscimol (a GABA<sub>A</sub> agonist that induces an inhibition of neuronal activity) resulted 287 in an increase in the temporal error rate as well as a shift to earlier responding. 288

Even though it has been described in fewer studies (only 14) than for other patterns,

duration-related phasic activity has been observed in a wide variety of brain areas, from cortical structures to hippocampus and *subtantia nigra*. Interestingly, this pattern seems to represent a discriminative response, and thus probably makes differentiation of durations more efficient. It appears in both implicit and explicit tasks, strengthening the idea that learning durations and discrimination between durations is important even in implicit tasks. However, this phasic activity represents durations relative to one another and not their absolute time.

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#### 298 2.3 <u>'Event time' cells</u>

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'Event time' activity relates to a transient increase or decrease of the firing rate of a 300 neuron at the end of a learned interval, usually reinforcement time or when the animal 301 302 must respond. One well-known example of such activity, described by Schultz et al. (1992), is the decrease of firing of dopamine neurons at the time of an expected, but 303 304 omitted, reward. To detect an 'event time' activity requires paradigms in which the period post-expected reinforcement can be studied; for example, by using probe trials where the 305 cue is presented for a longer duration and in the absence of reinforcement. Therefore, 306 307 unlike the previous two patterns, it is not dependent on the presence vs. absence of an external stimulus but only on the memorized duration. 308

<sup>309</sup> 'Event time' activity has been observed in the cortex (prefrontal, premotor, motor and <sup>310</sup> visual), striatum and hippocampus, and for a wide range of durations. Most of these <sup>311</sup> structures are part of the SBF and/or have been studied in the context of timing for a long <sup>312</sup> time. 'Event time' activity has also been observed in a peak interval timing task, when <sup>313</sup> unreinforced probe trials are introduced in a fixed-interval task (where the reward is <sup>314</sup> available after a certain amount of time has passed since a stimulus). In such a paradigm,

the animal develops a pattern of responding that is time-appropriate which, on average, 315 316 follows a Gaussian curve with a peak around the expected time of reward in probe trials. Using this task in rats, Matell et al. (2003) have shown that some neurons in the PFC and 317 in the dorsal striatum (the two main brain areas of the SBF model) follow patterns of 318 responding that are very similar to the temporal behavior of the animal, that is, their 319 activity reaches a maximum around the expected time of arrival of the reinforcement. 320 321 'Event time' cells have also been described in implicit timing tasks, such as Pavlovian conditioning and entrainment. In entrainment paradigms, the regular repetition of a 322 stimulus is interrupted and the expectation of the next stimulus can thus be recorded 323 324 without the interference associated to its presentation (Bartolo et al., 2014; Crowe et al., 325 2014; Merchant et al., 2013b, 2011). In a Pavlovian tone-shock conditioning paradigm, Armony et al. (1998) and Quirk et al. (1997) described a late tone-induced response that 326 327 appeared in the auditory cortex of rats after a single session of twenty CS-US trials. An increase in firing was observed late during the CS, just before the arrival of the US, 328 329 suggesting that it reflected the higher expectation of the US near its time of arrival. Although the US was not omitted, this neural activity close to the time of the US arrival 330 could reflect an 'event time' neural correlate. 331

The 'event time' activity seems to encode the expected arrival of the reinforcement, or of the next event. We can wonder how it bridges the duration between the CS onset and the US, presumably through other patterns of activity such as sustained changes.

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#### 336 2.4 <u>Ramping activity</u>

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Ramping activity, when a neuron's firing rate increases or decreases gradually with passing time, either from baseline level or after an initial abrupt change in activity, has

been observed in a majority of studies (e.g., Donnelly et al., 2015; Fuster et al., 1982; 340 341 Knudsen et al., 2014; Kojima and Goldman-Rakic, 1982; Paz et al., 2006; Sakai, 1974; Soltysik et al., 1975) and may represent the gradual increase in expectation of the animal. 342 A neuron discharges more and more (or less and less) as time passes until it reaches a 343 threshold which induces a specific response that is time appropriate. Ramping activity 344 has been described in most brain structures, in diverse cortical areas, but also in 345 346 subcortical structures such as the hippocampus and the basal ganglia. It thus does not seem specific to a brain region. In the absence of probe trials, it can difficult to distinguish 347 between ramping and 'event time' cells, because we cannot see the post-expected 348 349 reinforcement activity. For example, Narayanan and Laubach (2009) showed ramping activity in the dorsomedial PFC, which seems to reach a plateau just before the arrival of 350 the reinforcement; adding probe trials without reinforcement would have differentiated 351 352 activity related to a specific time from simple accumulation of time, as it would have peaked and gone back to baseline level after the expected time of reinforcement in the 353 354 first case, but would have continued to increase as long as the stimulus was present in the second case. 355

Ramping activity has often been described in delayed matching-to-sample or non-356 matching-to-sample (i.e., working memory) tasks, and in expectation tasks, where the 357 animal waits for a stimulus. These tasks are not typical timing tasks, but have a temporal 358 component that can be modulated, *i.e.* the wait between the first and the second stimulus. 359 360 The increased activity during the delay could represent an encoding of the hazard rate, that is, the longer the duration, the more likely the stimulus is to appear (Heinen and Liu, 361 1997; Janssen and Shadlen, 2005; Leon and Shadlen, 2003; Lucchetti and Bon, 2001; 362 Renoult et al., 2006; Riehle et al., 1997). It is difficult to know whether expectation is 363 similar to timing, as it may involve different mechanisms, such as the accumulation of 364

activity over time or an increase in attention until a given stimulus has finished, ratherthan precise temporal control.

Trying to parse the role of ramping activity in increased expectation vs. precise 367 temporal control, Donnelly et al. (2015) used an attention task (5-CSRTT) in which a 5s 368 waiting period without making a nose poke is imposed after the onset of the trial, before 369 a brief (500ms) light indicates in which hole the rat must go for getting a reward, a task 370 371 which requires a high attention level to detect the light cue. The authors, comparing correct responses with premature responses (when the animal did not wait for the light 372 cue to respond), found that positive and negative ramping activity in both PFC and ventral 373 374 striatum started earlier in premature trials, but with similar ramping rate, so that the activity reached the threshold for action earlier on premature trials than on correct trials. 375 No ramping activity was observed in trials where the animal did not respond. When the 376 377 waiting period was varied, ramping activity on correct trials was found to reach its maximum at the earliest time of possible appearance of the visual stimulus, then 378 379 remaining at this level until the emission of a response. Thus, ramping activity up to a threshold may indeed be a correlate of precise temporal control, and premature response 380 may result from an aberrant start of a timing signal that may originate from somewhere 381 382 else.

Like any unbounded accumulator, however, it seems biologically impossible to encode long durations of more than a minute with ramping activity. There is a limit to the number of spikes a single cell can produce in a definite amount of time. This is where population coding might come into play by having different populations activating each other to represent longer durations than a single cell can.

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#### 389 **3.** <u>Can neurons form a network to encode time?</u>

391 As we have seen previously, single cells can encode durations but they are limited biologically in how high their firing rate can be, and other mechanisms have to be at play 392 to make them fire at a specific time (outside of the presence of external stimuli). 393 Therefore, to span complete and/or longer durations, other forms of temporal encoding 394 are necessary, at a network level. In addition to recordings of several individualized cells 395 at a time, other techniques in animals can give us a view of neural activity at the 396 population level, such as multi-unit recordings, local field potentials (LFPs), calcium 397 398 imaging, micro-dialysis and PET-scan (Positron Emission Tomography), but with a wide range of temporal resolution (from the millisecond to the minute). Patterns of activation 399 similar to the ones observed with single unit can be observed with other techniques that 400 401 have good temporal resolution, such as multi-units, LFP and, in a slightly less precise way, calcium imaging. In comparison to the studies of single cell encoding of durations, 402 fewer papers have focused on activity related to time at a population level in animals (see 403 Table 3 for explicit tasks and Table 4 for implicit tasks). Nonetheless, they cover a large 404 range of model species and of time intervals, as well as explicit and implicit tasks similar 405 406 to the ones studied with unit recordings.

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#### 408 3.1 <u>Sequential time cells</u>

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Sequential time cells (also known as 'time cells') fire one after the other, creating, as a population, a range of firing across time which forms a bridge of activity between events separated by a constant time interval, thus encoding as a whole the event's duration or the interval between events (Figure 2A). **Interestingly, the response field of cells that fire later in a sequence is larger than for cells firing early (e.g., Kraus et al., 2013), which** 

could be consistent with scalar timing. Sequential time cells have been described in the 415 416 hippocampus (Kraus et al., 2013; MacDonald et al., 2013, 2011; Pastalkova et al., 2008), in the PFC (Horst and Laubach, 2012; Jin et al., 2009; Kim et al., 2013; Kojima and 417 Goldman-Rakic, 1982; Oshio et al., 2008; Sakurai et al., 2004), in the premotor cortex 418 (Merchant et al., 2011), and in the basal ganglia (Jin et al., 2009; Mello et al., 2015). 419 MacDonald et al. (2011) have differentiated cells depending on whether or not they 420 421 modify their peak firing time when the duration of the interval is changed. They named 'absolute time cells' cells that show a peak response at a specific time point during the 422 interval with a pattern that does not rescale when the duration is modified. 'Relative time 423 424 cells' show a similar peak response at a specific time but their activity is rescaled depending on the duration of the timed interval. Other cells may either lose their activity 425 or change their activity to a non-similar and non-rescaled time point when the interval is 426 427 modified. Other studies have also highlighted the dichotomy between relative versus absolute time cells (Kojima and Goldman-Rakic, 1982; Merchant et al., 2011). 428

429 To describe these sequential time cells, MacDonald and collaborators (2011) used a go/no-go paradigm with a delay. The rats were trained to pair objects and odors, such that 430 they had to retain in memory for 10s the object that was presented at the beginning of a 431 432 trial to know whether or not they should dig into a scented pot to get a reward. During the 10s delay, neurons in the hippocampus fired sequentially to cover the whole duration with 433 a firing pattern that was rearranged when the duration was changed. Most neurons were 434 435 modulated by both space and time, and in a manner independent of locomotion, speed, or head placement. In another study, it was shown that very few neurons depend only on 436 time (MacDonald et al., 2013, 2011). Therefore, these sequential time cells seem similar, 437 or even identical, to the place cells described in the hippocampus (O'Keefe and 438 Dostrovsky, 1971) and may interact with those cells to form spatiotemporal maps of the 439

environment. More recently, Mello et al. (2015) have reported that 68% of cells recorded
in the dorsal striatum of rats performing a serial fixed-interval task (from 12s to 60s) were
relative time cells (i.e. active at specific time points during the fixed-interval). The pattern
of activity was modulated by the duration of the interval but not correlated to motor
responses, thus demonstrating a scalable population of neurons which conformed to the
scalar property. As can be seen from the studies described above, sequential time cells
have been evidenced in both explicit and implicit timing tasks.

These sequential time cells seem to constitute a "pure" time encoding which can support the encoding of long durations and, even, parallel encoding of multiple durations simultaneously. However, the questions remain of what makes these cells fire at a specific time (e.g., does it result from a local process or do they receive a temporal input from an upstream brain area), and of to separate the different sub-populations representing different durations in different contexts.

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#### 455 3.2 <u>Non-electrical activity measures</u>

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Using Positron Emission Tomography (PET) scan imaging in monkeys performing a 457 visual temporal discrimination task, Onoe et a. (2001) showed blood flow modulation 458 (increase in blood flow is correlated with an increase in neural activity) in specific 459 structures that was correlated with the length of the estimated interval (within a 0.4 to 460 1.5s range). These structures included the dorsolateral PFC, the posterior inferior parietal 461 cortex, the posterior cingulate cortex, and the basal ganglia. Meanwhile, micro-dialysis 462 can give access to dynamic release of neurotransmitters, albeit with a temporal resolution 463 of around a minute at the maximum resolution. In an olfactory Pavlovian aversive 464

conditioning in which CS-US trials were given at a regular pace (4 min), Hegoburu et al. 465 466 (2009) saw transient amino acid increases (GABA and glutamate) in the piriform cortex of rats (but not in the amygdala) near the expected time of the CS-US trial when the trial 467 was omitted after training. These increases may thus be correlates of the encoding of the 468 inter-trial interval (ITI). Unfortunately, these two techniques lack good temporal (from 469 tens of seconds to a few minutes) and/or spatial resolution (from a few millimeters to a 470 471 few centimeters) compared to other types of recordings, meaning that it is not possible to separate the different steps of accumulating, encoding and decoding duration. 472

Calcium imaging in behaving animals is a recent technique which looks at the activity 473 474 of single cells using fluorescent calcium indicators. This technique adds the very interesting aspect of being able to give spatiotemporal information on neuronal activation, 475 albeit at the expense of a lower ( $\sim 100$ ms) temporal precision compared to electrical unit 476 477 recordings (~40 µs). Furthermore, it also gives the opportunity to record from a large number of cells at the same time, while individualizing them spatially. Using this 478 479 technique in zebrafish larvae (which are transparent), entrainment to a visual stimulus was observed in the lateral habenula (similar to the mammalian habenula) through an 480 increased calcium metabolism at the expected time of arrival of a neutral stimulus (Cheng 481 et al., 2014). Also, in mice, a ramping pattern of activated neurons was detected in the 482 hippocampus during the delay between CS offset and US in a trace conditioning task 483 (Modi et al., 2014). While the authors did not observe a specific spatial organization of 484 the time-tuned cells, they observed that cells with highly correlated spontaneous activity 485 formed clusters that were modified with learning. 486

487

488 3.3 <u>Monotonic change of neuronal population activity across time</u>

Studying episodic memory, (Manns et al., 2007) have shown in rats that events 490 closer in time are associated with spatial ensemble activities in CA1 that are more 491 similar than for events farther in time. They taught the rats to choose an odor 492 according to when in a sequence of odors it had been presented before, and showed 493 differences in cell firing encoding between earlier and later odors on correct trials, 494 but not on incorrect trials. These ensembles of cells in CA1 change their patterns of 495 activity monotonically across time and could provide a temporal context, a way to 496 encode and differentiate similar memories that are separated in time. This has also 497 been shown in the hippocampus of primates (Naya and Suzuki, 2011). Within the 498 499 hippocampus, CA2, and to a lesser extent CA1, networks show changes of activity over time in stable spatial environments, providing a temporal coding at the scale of 500 hours, in contrast to a highly consistent time-independent CA3 network activity 501 502 (Mankin et al., 2015, 2012). These changes over time do not impede the decoding of the spatial information that is also encoded (Rubin et al., 2015). 503

Interestingly, sequential time cells that code time in the range of seconds to minutes may also exhibit this type of ensemble timing (in the range of hours to days). Indeed, using calcium imaging *in vivo* in mice running timed laps on a treadmill, Mau et al. (2018) showed that a population of sequential time cells in CA1 that encoded a 10s duration would change gradually over days, therefore encoding time on a larger scale as well.

510

#### 511 3.4 Local field potentials (LFPs)

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513 LFPs represent the sum of depolarization/hyperpolarization of a population of 514 neurons, reflecting action potentials as well as subthreshold electrical modulation such as

EPSPs (excitatory post-synaptic potentials) and IPSPs (inhibitory post-synaptic 515 516 potentials) (Buzsáki et al., 2012). It is important to note that LFPs represent both input activity (i.e. coming from upstream brain areas) as well as local computations and output, 517 in contrast to single unit or multi-unit recordings which only reflect spikes, and thus 518 output data of the recorded area. Some LFP modulations are time-locked to the onset of 519 a stimulus and are called event related potentials or ERP (Figure 2B). They can be 520 521 observed when averaging a large number of trials under constant conditions. The raw LFP signal (Figure 2C) recorded from a brain area can also be decomposed in different 522 frequency components (i.e., oscillations) that are considered to have different roles in 523 524 neural processing. Data on oscillations are often presented in the form of power spectrum 525 density (PSD), which represents the strength of different frequency bands in a signal. When looked at in a time-frequency manner, one can ask how the power of different 526 527 frequency bands varies across a trial. Most frequency bands - from slow oscillations (like delta and theta) to mid-range (like alpha and beta) and even high frequency oscillations 528 529 (like gamma and epsilon) - have been described in several mammalian species, and neural oscillations seem to be a conserved phenomenon across mammalian evolution (Buzsáki 530 et al., 2013). However, depending on the task and on the brain area under study, the exact 531 532 parameters of these oscillatory bands may differ. We will thus refer to the specific frequency ranges rather than the sole classification range, in order to keep accuracy and 533 avoid misinterpretation. 534

Neuronal oscillations have long been hypothesized as the major constituent of the internal clock (Buhusi and Meck, 2005; Treisman, 1963). Oscillations also seem to be involved in structuring events in time (for example events can be associated with the specific phase of an oscillation) (Kösem et al., 2014; Mizuseki et al., 2009). They present a further interest, as they provide potential comparison with human studies where most of the data are in the form of oscillatory activity measured in a non-invasive manner (Electroencephalogram (EEG) and Magnetoencephalogram (MEG)). In contrast to the large number of studies done using EEG and MEG in humans for the past two decades, studies on LFP and interval timing in animals are rather recent but reflect a growing interest in the neuroscience community.

545

#### 546 *3.4.1 Explicit tasks*

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Comparing an auditory temporal discrimination task with a simple auditory 548 549 reaction time task in rats, Onoda and collaborators showed that ERPs can be modulated by timing. They showed an involvement of the frontal cortex, the hippocampus and the 550 cerebellum in discriminating between 2s and 8s (Onoda et al., 2003) and of the frontal 551 552 cortex, striatum and thalamus for durations shorter than 2s (Onoda and Sakata, 2006). Matell and Meck (2004) have proposed, within the frame of SBF model, that the ERP 553 554 signal could be representative of a reset of cortical neurons at the beginning of a stimulus, which seems coherent with the observation of a time-dependent ERP in the frontal cortex 555 for all tested durations. These results confirm the lesions and inactivation studies showing 556 557 the importance of the striatum and PFC in interval timing while implying an involvement of other structures that have also been suggested to play a role in timing (i.e., cerebellum, 558 thalamus and hippocampus). However, ERPs do not give information on whether nor how 559 560 these structures are involved in further temporal processing beyond a reset mechanism.

Low frequency oscillations have been associated with explicit temporal processing in several structures, mostly in the hippocampus, striatum and PFC. Concerning the hippocampus, the results are somewhat contradictory depending on the durations studied. Nakazono et al. (2015) showed a transient increase in theta power (4 -

9 Hz) during the comparison between a 1s and a 3s stimulus in a temporal discrimination 565 566 task. However, looking at longer intervals (30s) in a peak interval task, Hattori and Sakata (2014) found that the power increase in the 6-12 Hz band was not modulated by time in 567 the hippocampus, while it was so in the striatum. Looking at both the striatum and the 568 PFC, Emmons et al. (2016) showed a correlation between timing behavior in rats engaged 569 in a fixed interval task (with intervals ranging from 3 to 16s) and the theta band (4 - 8)570 Hz) within the PFC as well as with the delta band (1 - 4 Hz) of the striatum. Interestingly, 571 timing behavior was impaired when disrupting dopamine signaling in the PFC, in 572 correlation with a decreased 4 Hz power in the PFC (Kim et al., 2016; Parker et al., 2014). 573 574 Furthermore, these impairments were rescued by stimulation of the lateral cerebellar nuclei projection to the thalamus (Parker et al., 2017). No study has reported an 575 involvement of higher frequencies in explicit timing, but it may reflect a simple neglect 576 577 of analysis. While these experiments show an important role of a PFC-striatum-thalamocerebellum network in timing behavior, further experiments are needed to give more 578 579 insight on potential larger networks (e.g., other cortical areas, hippocampus) and their specific involvement in the processing of time when the subject is engaged in explicit 580 timing. 581

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## 583 *3.4.2 Implicit tasks*

584

585 So far, eleven studies using various implicit timing tasks have reported changes in 586 neural oscillations in several brain areas and from low to high frequency ranges. In 587 entrainment tasks, in which a stimulus is presented at regular intervals, the observed 588 change is usually an increased activity at the entrained frequency even in absence of the 589 stimulus. For example, Abe et al. (2014) showed that hippocampal oscillations could be

entrained at a theta (4-12 Hz) frequency but not at higher frequencies (above 20 Hz).
Bartolo et al. (2014) showed entrainment in beta (10-30 Hz) and gamma (30-80 Hz)
ranges in the striatum of monkeys during an internally-driven rhythmic tapping task with
intervals ranging from 450ms to 1s.

Changes in power from low to mid frequency ranges have also been reported in 594 several cortical areas in tasks involving holding attention for a given amount of time. In 595 596 a sustained attention task, in which the rat had to pay attention for 8s to detect a visual stimulus that indicated which hole to choose between three to get a reward, Totah and 597 collaborators (2013b) reported changes during the delay on correct trials (incorrect trials 598 599 being any trials during which the animal chose the wrong hole) in two regions of the PFC, the anterior cingulate cortex (ACC) and the prelimbic cortex (PL). They observed an 600 increase in delta oscillations power (1-4 Hz) in a ramping manner in the ACC, while the 601 602 increase was sustained in the PL. Either of these patterns could be interpreted as a representation of expectation and time. Kilavik et al (2012) showed an increase in power 603 604 in a 12 to 40 Hz range in the motor cortex that was related to the temporal motor preparation of monkeys performing a reaching task with two different delays. The short 605 durations used (0.7 - 2 s range) may not have made the characterization of a ramping 606 pattern possible. Zold and Hussain Shuler (2015) described a modulation of a 6-9 Hz 607 frequency band in the primary visual cortex during a reward expectation task in rats, the 608 duration of which evolved with training from being initially related to physical parameters 609 610 of the stimulus used (i.e. the visual stimulus intensity) to being correlated to the reward time. However, as it was also correlated with the rate of reward (i.e. performance), it is 611 difficult to ascertain that it represented temporal coding rather than saliency/motivation 612 encoding. 613

Changes have also been reported in ranges from low to high frequency, most often in 614 615 the amygdala, but also in other subcortical and cortical areas, in associative conditioning tasks. In these tasks, in which the CS predicts the arrival of a US, at a given time, usually 616 at the end of the CS (delay conditioning) or after a stimulus-free period (trace 617 conditioning), changes in neural oscillations have been observed in a manner compatible 618 with increasing temporal expectancy for the US arrival. An increase in synchronicity 619 620 within the theta range (4-7 Hz) has been observed in the lateral nucleus of the amygdala (LA) in aversive conditioning in cats (Paré and Collins, 2000), while a sudden increase 621 in power in the gamma range (50-80 Hz) followed by a progressive return to baseline, 622 623 and a decrease in power of lower frequency oscillations (10-30 Hz), have been reported in the auditory cortex in a similar task in rats (Headley and Weinberger, 2013). In 624 appetitive Pavlovian trace conditioning in cats, Bauer et al. (2007) observed a ramping 625 626 low gamma activity (35-45 Hz) peaking just before the reward for the basolateral nucleus of the amygdala (BA) and earlier for the rhinal cortices. Interestingly it emerged with 627 628 training. However, in all these studies, the lack of recordings during post-reinforcement time and the use of a single CS-US interval render difficult to determine whether these 629 changes in neural activity really represent timing of the CS-US interval, rather than a 630 simple increased US expectancy as time passes. In a specifically designed Pavlovian 631 aversive task in which the US was embedded within a long 60s CS and probe non-632 reinforced CS alone trials were interleaved between CS-US pairing trials, allowing to 633 follow both the ascending and descending parts of the US expectancy, Dallérac et al. 634 (2017) showed a bell-shape pattern of increased power that followed closely the temporal 635 behavior pattern, in the theta (3-6 Hz) and gamma (60-70 Hz) ranges for both the BA and 636 the dorsomedial striatum. Importantly, these patterns shifted in time with a shift in the 637 CS-US interval and followed the scalar property of time. 638

While changes in oscillatory power points to the involvement of a given brain area 639 640 in timing - although the function of specific frequencies remains to be resolved - it does not give access to the recruitment of potential networks encompassing several structures. 641 Synchronicity of oscillations between different structures, also called coherence (Figure 642 2D), gives information about the communication between structures. When two structures 643 are highly coherent, it is thought to make information transfer easier because the other 644 645 structure is already primed to receive the spiking activity from the first (cf. the communication through coherence hypothesis, Fries, 2005). Only four studies have 646 reported increased coherent activity between brain areas, all of them involving the 647 648 amygdala in associative tasks. Pape and collaborators (2005) were the first to note a progressive increase in correlated theta (5-6 Hz) between CA1 of the hippocampus and 649 650 the lateral amygdala (LA) across the CS in an aversive Pavlovian delay conditioning 651 paradigm in mice. Popescu et al. (2009) detected an increase in gamma band (33-45 Hz) coherence between the BLA and the posterior striatum during the CS, with a peak 652 653 immediately before the arrival of the reward that was bigger for the CS<sup>+</sup> (associated with the US) than for the CS<sup>-</sup> (not associated with the US). This increased coherence emerged 654 with repeated training, and may thus represent a correlate of some expectancy-related 655 656 behavior rather than the initial temporal learning (which happens from the start of associative learning, Balsam and Gallistel, 2009). Totah and collaborators (2013b) 657 showed that coherence in a 8-12 Hz band between two sub-areas of the PFC (ACC and 658 PL) was increased in a ramping manner during the 8s preparatory period predicting a very 659 brief visual stimulus the rat had to detect. More recently, Dallérac et al. (2017) showed a 660 temporally-linked increase in coherence in the 3-6 Hz theta (but not the 60-70 Hz gamma) 661 range between BA and dorsomedial striatum that peaked at the expected time of an 662 aversive US arrival and returned to baseline level, in trials in which the US was omitted. 663

This increased coherence was shifted following a scalar-related variance when changing the CS-US duration. These results suggest that the dynamics of functional connectivity, as reflected through coherence, may indicate time-based recruitment of large neural networks. Although these types of investigation are yet too few to give a good idea of the network(s) involved in different timing tasks, it seems that the amygdala, prefrontal cortex and striatum may enter in active interaction during temporal expectancy.

- 670
- 671 3.5 Modulating neuronal activity
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673 Thanks to recent methodologies permitting a highly precise temporal control of cells' activity, there is an increasing interest in investigating how modulating activity 674 of specific neuronal populations could influence interval timing. In a task where rats 675 676 learned to optimize their wait time (~1.1s) after a visual cue in order to get the biggest reward, Namboodiri et al, in (2015), have shown that stimulating, for 200ms, 677 V1 neurons 300ms after the visual cue, shifted the entire distribution of the animal's 678 waiting times, and thus delayed its timed actions. Three papers from the 679 Narayanan's lab have looked at how to rescue behavioral timing impairment due to 680 681 a disruption of the medial frontal cortex (MFC) functioning. Stimulating D1 dopamine receptors in the MFC of DA-depleted transgenic mice or stimulation the 682 lateral cerebellar projection to the thalamus at 2Hz frequency in rats with D1-683 inhibited MFC, which both increased the ramping activity in the MFC (a well 684 described characteristic of timing, (Emmons et al., 2017; Kim et al., 2016; Parker et 685 al., 2015), or 20Hz activation of MFC axons in the dorsomedial striatum (DMS) 686 which rescued the time-related ramping activity in the DMS in MFC muscimol-687 inhibited rats, all these rescued the animal's temporal behavior in a 12s fixed 688

689 interval task that was otherwise disrupted by MFC dysfunction (Emmons et al., 2019;
690 Kim and Narayanan, 2019; Parker et al., 2017).

Soares and collaborators, in (2016), have shown that activation of the 691 dopaminergic neurons in the substantia nigra pars compacta in mice during the 692 entire interval produced a shift to the right of the temporal bisection curve (range 693 0.6-2.4s), suggesting a slowing down of internal time, whereas inhibition of those 694 neurons produced a shift in the opposite direction, compatible with an acceleration 695 of internal time. Targeting the GABAergic projection from the substantia nigra pars 696 reticulata (controlled by outputs from the striatum) to the superior colliculus in mice 697 698 performing in a 10s fixed-time schedule with delivery of sucrose reward, Toda et al. (2017) showed that, depending on the frequency of stimulation as well as when the 699 stimulation was done during a trial, stimulating this nigrotectal pathway not only 700 701 stopped the on-going licking behavior but could also produce a shift in anticipatory licking on the next trial, and thus disrupted interval timing. 702

All these studies not only confirm the suspected critical role of some brain areas (prefrontal cortex, striatum) or neuromodulators (dopamine), but also bring novel information on neural networks and the frequencies at which they should function for optimal interval timing. Nevertheless, more such studies aiming to modulate the activities previously correlated with passing time are necessary to help us distinguish the essential neural patterns for time from the non-essential.

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### 710 4. Open questions

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712 4.1 <u>Neural syntax of timing from single cells to networks</u>

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How the animal brain encodes time remains a challenging question. As we just reviewed above, a large range of activity patterns (ramping, sustained and phasic) could be considered to be associated with passing time and this suggests that it may be their combined activities that underlie timing.

At first, it is tempting to think that sequential time cells could be the main tool the 718 brain uses for time encoding; however, are they sufficient? Could these cells be at the 719 720 origin of the other patterns of activity that have been described or are they the end results of other neural computations? Sequential time cells may share the role of single unit's 721 722 sustained activity but for longer durations that cannot be coded by a single cell. Sequential 723 time cells need to be part of sets (i.e. functional populations) with each set involved in encoding a specific duration, this allows for multiple durations to be encoded and decoded 724 725 at the same time.

726 Oscillations may provide a good way to produce those sets either through the phase locking of spikes to different phases of a frequency band (i.e. whether spikes are 727 728 repeatedly more present at specific phases of the oscillations), or through phase amplitude coupling (PAC) of high frequency oscillations' power with low frequency oscillations' 729 phase. To our knowledge very few studies studies have looked at this aspect of neuronal 730 encoding in a timing task. Phase-locking of spikes to delta oscillations (centered at 1.6Hz) 731 in the ACC have been reported to correlate with the growing expectation of a reward in a 732 sustained attention task (Totah et al., 2013a). In that study, the phase-locking was 733 increased significantly more in correct than incorrect trials within the 2s period before the 734 presentation of the stimulus indicating which hole to choose to get a reward (2s before). 735 A similar pattern was observed in the PL, but with phase-locking of spikes to beta 736 oscillations (centered at 17 Hz). However, Nakazono et al. (2015), looking at single units 737 and theta oscillations in the hippocampus in a temporal discrimination task, showed that 738

only few cells were time-locked to the oscillations compared to the study by MacDonald
et al. (2013). The authors suggested that the two types of activity may have different roles,
albeit both related to time: theta oscillations might help the hippocampus to interact with
the PFC at the correct time, whereas the spikes may encode both temporal and
spatial/sensory information.

Clearly, more studies specifically devoted to timing are needed to decipher the respective roles of the different types of neural activities, how these patterns emerge, and how their intricate intertwined activities may possibly create time representations in the brain. In this venture, it will be critical to also identify which ones are required for timing per se versus other aspects of the task (e.g., motivation, motor preparation, ...).

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#### 750

#### 4.2 Neural correlates of time associated with models

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A number of models of interval timing have been proposed, but neural proofs can be contradictory and do not, for now, allow for choosing one specific timing model. As the subject is quite large and beyond the purpose of this review, we will only quickly discuss to what extent the available data support some of these timing models.

The main group of internal clock models are pacemaker accumulator models (PA). Taking into account the intrinsic functioning of neurons and known network connectivity, Matell and Meck (2000) proposed the Striatal Beat Frequency model (SBF). It involves numerous cortical oscillators (representing the pacemaker and accumulator), and the detection of their coincident activation by the medium spiny neurons of the striatum on which they project. At the start of a stimulus, the oscillators are synchronized (maybe by a dopamine burst, Kononowicz, 2015) while

keeping their own distinct frequencies, meaning that they will not remain 764 765 synchronized over time. When a significant event happens (e.g., at the end of the stimulus or at the appearance of a reinforcement), the state of these different 766 oscillators is encoded and stored by the striatum. By comparing the oscillators' 767 ongoing activity to the memorized patterns, it is possible for the striatum to 768 determine how much time has passed, provided that the oscillatory activity remains 769 770 very stable across time. Recently, replacing the memory and decision stage of the SBF by the ones from a well characterized cognitive architecture (the adaptive 771 control of thought-rational, ACT-R), which has defined default parameters, has 772 773 improved modeling of more complex temporal behaviors (such as timing of multiple overlapping intervals) (van Rijn et al., 2014). These systems from ACT-R may 774 involve other brain areas (e.g., the hippocampus) for memory and decision-making 775 776 processes.

Oscillations in the frontal cortex seem to be important for timing (e.g. Parker et 777 778 al, 2014, and duration-related increased neuronal oscillations have been reported in the dorsomedial striatum (Dallérac et al, 2017) that could reflect a read-out of 779 convergent oscillations from cortical neurons. However, no study has yet shown 780 781 increased coherence between striatum and prefrontal cortex during interval timing. Potentially adding on to the SBF model, data from our lab seems also to implicate 782 the amygdala as a modulator of cortico-striatal synapses to maintain the memory of 783 a previous duration when learning a new duration. So both memories are 784 maintained even though behavioral expression shifts quickly to the new duration 785 (Dallérac et al. 2017, see Figure 10). However, some of the expected signs of the SBF 786 model, such as phase reset at the beginning of the interval, have not been described 787 in neurophysiological studies (Kononowicz and van Wassenhove, 2016). 788

The TopDDM (Time Adaptative Opponent Poisson drift-diffusion model) is also a PA model, but based on the drift-diffusion model of decision-making (Balci and Simen, 2016; Simen et al., 2011). It considers, in very general terms, that the accumulation of time should be represented by a ramping activity which rate is inversely proportional to the duration encoded. As reported above, this type of activity has been described in multiple papers (e.g. Komura et al., 2001; Lebedev et al., 2008; Murakami et al., 2014).

There are also non-clock-based models; they require a population of units (can 796 be neurons or group of neurons) that respond differently across time, which is 797 798 consistent with the existence of sequential time cells. Included in those models is the TILT (Timing from Inverse Laplace transform) model from Shankar & Howard 799 (2012), as well as the multi-timescale model (MTS) from Staddon et al. (Staddon 800 801 and Higa, 1999), and the Spectral Timing Model (Grossberg and Merrill, 1992). TILT model can also explain the scalar property of time as well as the recency effect 802 of episodic memory (Howard et al., 2015). Its principle is that the presentation of a 803 stimulus will activate a certain number of t nodes which will maintain a stable firing 804 rate across the period of to-be-encoded time, and after the end of the interval decay 805 806 exponentially to result in the activation of T nodes through an inverse Laplace transform. This will produce units that are active at a specific time point after the 807 beginning of the stimulus. The different T nodes are activated sequentially and do 808 809 not influence one another, they are instead influenced by the activity in the t nodes. Activities resembling to t nodes (stable increased firing) and to T nodes (sequential 810 time cells) that when active during the early part of the interval have smaller spread 811 of activity than cells activated later have both been reported in the literature. 812

813 However, it remains to be determined to what extent they are found in all brain 814 areas.

815 Clearly, more studies are necessary to improve our models of timing, to try and
816 disprove some of them, and choose between clock and non-clock-based models.

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818 4.3 <u>Clock(s)</u>

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The question of how time is "written" in the brain opens another question of whether there is one clock, multiple clocks, or any clock at all. As Staddon & Higga (1999) have proposed, time may be encoded in the decaying function of memory strength, and would therefore be present in all brain areas that are involved in a specific task. If so, timerelated patterns of neural activities may simply reflect the resulting read-out of these memory strengths at each storage sites.

Neural correlates of time have been observed in most parts of the brain, depending on 826 the duration and the context of the task, making it difficult to determine the potential 827 origin of time. In particular, sequential time cells have been recorded not solely in the 828 hippocampus, but also in other brain areas that are involved in timing (PFC, premotor 829 830 cortex, basal ganglia). Also, as multiple durations are processed and encoded at every moment, context might be critical to distinguish which intervals are essential in a specific 831 situation. Indeed, duration and spatial location are learned and processed together as 832 shown through space-time-context binding (Malet-Karas et al., 2019). It would seem 833 important to store durations and contextual cues in the same anatomical regions, therefore 834 going in the direction of multiple clocks. 835

The possibility of multiple clocks is suggested by the fact that some individual neurons have an intrinsic oscillating activity, *in vitro* (e.g. Grace and Onn, 1989;

Hutcheon and Yarom, 2000; Steriade et al., 1993) and *in vivo* (Hyland et al., 2002).
Furthermore, it has been shown that Purkinje cells in the cerebellum can produce a
timed pause in activity by themselves, i.e. intracellularly (Johansson et al., 2014) and
can even encode two durations (Jirenhed et al., 2017). Therefore, a single cell might
represent the smallest unit for an internal clock.

This could lend some credence to the hypothesis that a small part of the brain is 843 844 sufficient for timing, without a need for inputs from external activity (Chubykin et al., 2013; Goel and Buonomano, 2016; Johnson et al., 2010). In effect, Johnson and 845 collaborators (2010) showed that chronic rhythmic stimulation in organotypic cortical 846 847 slices (auditory and somatosensory) can entrain the cell activity to reflect the interval between stimulations, in the hundreds of milliseconds range. Chubykin and colleagues 848 (2013) have also shown that it is possible to « teach » an interval of time to a slice of 849 850 primary visual cortex by using a carbachol infusion as a US and electrical stimulation of the underlying white matter as a CS. Changing the interval between the CS and the US 851 852 induced a shift of cortical layer 5 neurons responses to the new time. In vivo, Namboodiri et al. (2015) have demonstrated that it is possible for the primary visual cortex alone to 853 encode and influence highly stereotyped and quick motor actions. Although this remains 854 855 to be tested for longer durations, all these studies support the hypothesis that every cortical circuit has the capacity to time and that there are local clocks that are activated depending 856 on the type of task or stimuli (Karmarkar and Buonomano, 2007). However, these may 857 be restricted to rather simple timing demands and may not be sufficient for more complex 858 timing tasks. It is conceivable that, with increasing complexity and demand, rather than 859 processing time within a brain area, larger networks may be involved, and improvement 860 of communication between different brain areas during salient time periods may be 861 necessary to encode duration, especially when it is long. Interestingly, Emmons et al. 862

(2016) showed that the inactivation of the PFC inhibits the ramping activity in the 863 864 striatum and disrupts temporal behavior, suggesting a role of between brain connectivity in interval timing. Functional connectivity between brain areas can be measured in 865 various ways, through coherence (synchronization of the two LFP signals), PAC between 866 structures or phase-locking between oscillations and spikes (see a few examples at the 867 end of 3.3.2). Although the data are yet too few to draw a picture of time encoding at the 868 869 scale of large networks, it is a promising research avenue toward our understanding of the neural syntax of timing. 870

It is thus possible that simple timing processing can be done in the brain area involved in the task but that, for more complex paradigms, a larger network is at play (possibly including PFC and striatum). This seems to be the case during development, as preweaning age rats (around PN18-20), when prefrontal cortex, striatum and hippocampus are not yet adult-like (Tallot et al., 2015), can still learn and memorize simple intervals (Tallot et al., 2017). This simple/complex dichotomy may also apply in humans when comparing explicit and implicit timing in children *vs.* adults

878 (Droit-Volet and Coull, 2016).

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880

### 4.4 Temporal learning vs. temporal behavior

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One parameter that has been overlooked so far in the literature is the amount of exposure to the duration that is encoded. In effect, when looking at the experiments described in the present review, we can notice that most of them are done in well trained animals, and we can wonder whether different neurophysiological substrates may underlie timing depending on the level of training of the animal; especially because precise temporal behavior usually appears after at least a few hundred trials, although duration is discriminated and learned much earlier (Balsam et al., 2010). In addition, going even further in training, the animal may enter into habitual behavior, defined as insensitive to devaluation of the reward, a process which may not happen at the same rate depending on the duration and the task (Araiba et al., 2018). Habit may require a different neural encoding than initial learning and, even, than optimized behavioral output, although the network may remain the same (Doyère and El Massioui, 2016).

A single time interval can be learned in as little as one conditioning trial, as shown in 894 Diaz-Mataix et al. (2013), in which a change in CS-US interval is detected, that triggers 895 plasticity in the amygdala. Noticeably, this experiment is an example of a true 896 897 retrospective timing task, as the animal is "asked" only once whether the CS-US interval during reactivation differs from the one presented during conditioning. When adding 898 more training/testing sessions, the animal enters into prospective timing which is the 899 900 domain of most of the studies described in our review. Whether prospective timing is at play from the second trial or after several repetitions is not known, nor is it known whether 901 902 prospective timing would also engage time-stamping mechanisms, which retrospective timing likely relies on. 903

Importantly, learning a duration and optimizing temporal behavior may involve different mechanisms. This has been well exemplified by the study of Ohyama and Mauk (2001) using eyeblink conditioning. Rabbits were trained on a first duration only for a few sessions (i.e. before expression of temporal behavior) and then trained on a new duration for hundreds of trials, resulting, once the behavior had been optimized, in a dual response, one at each duration, and thus unravelling the initial, yet hidden, learning of the first duration.

911 Only very few studies have tried to look at how neural correlates emerges with 912 training. Using calcium imaging, Modi et al. (2014) evidenced the modification of spatial

patterns of neuronal activation depending on the level of training, which could represent 913 914 progressive learning of the duration and improvement in precision. In mice navigating in a virtual environment, Heys and Dombeck (2018) recorded sequential time cells' calcium 915 916 activity in the medial entorhinal cortex when animals spontaneously stopped to move and for the duration of inactivity, as a way to see how animals record passing time in the 917 absence of changes in space or in motor output. By changing the environment sufficiently 918 919 to get a global remapping of space and sequential time cells, they considered that they were looking at the initial encoding of time in that new environment. They showed that 920 sequential time cells' activity was very stable from the beginning of the session, from the 921 922 first trial even. Another proxy of early learning of duration can be to change the learned duration to a new one, forcing the system to re-encode. However, the temporal behavior 923 is shifted in time very quickly and it can be difficult to separate learning the new duration 924 925 and forgetting or eliminating responses to the previous one, each process possibly relying on different brain areas (Dallérac et al., 2017). 926

927 Beside the effects of training on the neural encoding of time, the behavioral output of the animal per se may modify how passing time is encoded, as has been shown in the 928 lateral entorhinal cortex (Tsao et al., 2018). On one hand, when behavior is stable across 929 930 trials, then encoding of time becomes relative to these trials, and it is difficult to differentiate a trial at the beginning of the session from a trial at the end of the session, 931 and time within a trial is very well encoded. On the other hand, in free exploration, the 932 933 flowing of time across the whole session is encoded, even if there are temporal cues intrinsic to the session that can separate it into trials. Therefore, behavioral output may 934 change how the animal perceives time. This is similar to what was implied in the 935 Behavioral Theory of Timing, where behavioral states are used as temporal counters 936 (Killeen and Fetterman, 1988). 937

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#### 4.5 <u>Animal vs. Human</u>

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Some aspects of timing may differ between humans and animals, potentially due to 941 the fact that animals need a lot of training to understand the task or express temporal 942 control in their behavior, while reading instructions is sufficient in most cases in humans. 943 944 The fact that there has not been much attempt to record brain activity in paradigms enabling comparison between animal and human data makes it difficult to extract a global 945 picture across species. Indeed, the types of tasks and of durations used are very different 946 947 between the human and animal research, as well as the types of recording. The neural correlates of time have been studied a lot in humans using different techniques ranging 948 949 from functional magnetic resonance imaging (fMRI) to magnetoencephalography 950 (MEG). We will not go into details since the subject has already been covered by several reviews (Allman et al., 2014; Coull et al., 2011; Merchant et al., 2013a). As a brief 951 952 summary, several structures have been detected as active during timing tasks across many different studies, such as the supplementary motor area (SMA), the pre-SMA, the 953 954 prefrontal cortex, the striatum, the inferior parietal cortex and the cerebellum (Brannon et 955 al., 2008; Coull and Nobre, 2008; Harrington et al., 2004; Lewis and Miall, 2003; Rubia and Smith, 2004; Wiener et al., 2010; Wittmann et al., 2011). The pre-SMA (or rostral 956 SMA) seems more involved in perceptually-based timing in the supra-second range, 957 958 whereas the caudal SMA may be more important for sensorimotor-based timing in the sub-second range (Schwartze et al., 2012). However, timing of a stimulus offset seems to 959 960 be encoded in the related sensory cortex (van Wassenhove and Lecoutre, 2015). In a metaanalysis, Wiener et al. (2010) highlighted the fact that the structures involved often 961 depend on the type of task and on the durations used. They concluded that two main 962

structures are involved in temporal encoding only: the prefrontal cortex and the SMA. In contrast, Harrington and collaborators (2010) showed that the striatum is the only structure that is more active during the encoding of durations than during their maintenance in memory. In their study, the SMA and pre-SMA had a high activity for both encoding and maintenance phases, while the cerebellum and frontal cortex were more active in the time paradigm only at the decision stage.

Unfortunately, the SMA and pre-SMA have not been studied much in animals' timing studies, in contrast to the prefrontal cortex, the striatum and the *substantia nigra*. Striatum and *substantia nigra* are deep brain structures that cannot be recorded easily in humans at least on a rapid timescale. Therefore, the question of to what extent time is encoded similarly between animals and humans may not find a complete answer - without technical improvement of recordings in humans - but there is space for advancement in species comparability.

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#### 977 4.6 <u>Concluding remarks</u>

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As we have seen in the previous pages, the neural processing and encoding of time 979 has been studied in animals for ~50 years using a large range of techniques, giving us 980 insight from individual neurons to the interaction of separate brain areas. However, 981 compared to other types of neural computation (such as spatial encoding), we are still 982 very much unsure of the exact neural correlates of interval timing. To process long 983 durations, interactions between populations of neurons, and even maybe between brain 984 areas, might be necessary (as ramping and sustained activity cannot be maintained over 985 long periods of time), but it has almost never been studied in that aspect. 986

In this review, we have pointed to studies where temporal patterns of brain activity 987 988 were observed, but for the most part, the studies described here did not directly have the purpose of searching for timing related activity, in particular in the case of implicit timing 989 990 tasks. This puts into focus the necessity to use the decades of psychological research on timing to create specific protocols and paradigms to determine how time can be processed 991 and encoded in the brain, and how these processes may be modified depending on task's 992 993 demand. For example, although testing for the scalar property of time is an essential parameter of a temporal task, very few studies have looked at it while recording brain 994 activity (see Dallérac et al., 2017; Mello et al., 2015). It is also very important to look at 995 996 both the pre-reinforcement as well as the post-reinforcement periods, for example by using non-reinforced probe trials, as some patterns of encoding cannot be detected when 997 looking only at the expectancy period. Also, specific controls may be necessary to isolate 998 999 temporal information from saliency of stimuli or of reinforcement. In addition, looking in a wide variety of brain areas, not necessarily involved in the task at hand, could help 1000 determine if timing is ubiquitous in the brain. Finally, cross-species comparison, using 1001 virtually identical tasks, is critical in order to dissociate general principles of timing 1002 mechanisms from species-specific time-associated behaviors, thus optimizing the 1003 1004 potential generalization of findings to human brain function.

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1011 <u>References</u>

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1625	Figure captions
1626	
1627	Figure 1: Schematic representation of the different patterns of changes in the number of
1628	spikes of single neurons in relation to the encoding of temporal durations (here two
1629	durations are represented, one in black and one in red). All of these patterns can also be
1630	observed in a negative variation.
1631	
1632	Figure 2: Schematic representation of possible encoding of time though modulation of
1633	population neural activity either through organization of single cells activity (A), or
1634	through local field potentials modulations (B-D).
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1636	Tables captions
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1638	Table 1: Single unit studies in explicit timing tasks
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1640	Table 2: Single unit studies in implicit timing tasks
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1642	Table 3: Population-level studies in explicit timing tasks
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1644	Table 4: Population-level studies in implicit timing tasks
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Tasks	Brain structures	Species	Durations (in s)	Results				References
				Sustained activity	Phasic response at onset or offset of cue	Event time	Ramping activity	
Time production	Motor Cx	rat	1 - 2	а		a	а	Knudsen et al. (2014) (a)
	Premotor Cx	monkey	2.5 - 4.5				а	Lebedev et al. (2008) (a)
	Parietal Cx	monkey	0.5 - 1				а	Jazayeri and Shadlen (2015) (a)
	Presupplementary Cx	monkey	2 - 8				a, b	Mita et al. (2009) (a); Roux et al.
	Supplementary motor							(2003) (b)
Time reproduction	Prefrontal Cx	monkey	2 - 7	а	a			Yumoto et al. (2011) (a)
	Parietal Cx	monkey	0.5 - 1				а	Jazayeri and Shadlen (2015) (a)
	Premotor Cx	monkey	0.45 - 1			а		Merchant et al. (2013b) (a)
Peak interval	Prefrontal Cx	rat	1.5 - 2.5			а	а	Xu et al. (2014) (a)
			10 - 40			b	а	Parker et al. (2014) (a); Matell et al. (2003) (b)
	Lateral agranular Cx	rat	10 - 40			а		Pang et al. (2001) (a)
	Medial agranular Cx	rat	10 - 20			а	а	Matell et al. (2011) (a)
	Primary visual Cx	rat and mice	1 - 2	a, b		a, b, c		Chubykin et al. (2013) (a); Liu et al. (2015) (b); Hussain Shuler and Bear (2006) (c.)
	Dorsal striatum	rat	10 - 40			a, b		Matell et al. (2003) (a); Portugal et al. (2011) (b)
Differential reinforcement of low rates (DRL)	Secondary motor Cx	rat	1 - 4		b		a, b	Murakami et al. (2014) (a); (2017) (b)
	Prefrontal Cx	monkey	2	а			а	Niki and Watanabe (1979) (a)
	Hippocampus	rat	15				а	Young and McNaughton (2000) (a)
Limited hold (LH)	Prefrontal Cx	pigeon	1.5				а	Kalenscher et al. (2006) (a)
	Ventral striatum	monkey	1			а		Shidara et al. (1998) (a)
	Primary visual Cx	rat	1.5	а		а		Namboodiri et al. (2015) (a)
Temporal discrimination	Parietal Cx	monkey	0.3 - 0.8				а	Leon and Shadlen (2003) (a)
	Frontal Cx	monkey	0.2 - 2	d	a, b, c, d		a, b, c, d	Genovesio et al. (2006) (a), (2009) (b); Oshio et al. (2006) (c), (2008) (d)
		rat	3 - 4				а	Kim et al. (2013) (a)
	Motor Cx	monkey	0.6 - 1.2		а		а	Roux et al. (2003) (a)
	Dorsal triatum	monkey and rat	0.2 - 2.4		a, b	с		Chiba et al. (2008)(a), (2015)(b); Gouvêa et al. (2015)(c)
	Hippocampus	rat	1 - 3		a			Nakazono et al. (2015) (a)
Fixed interval	Medial prefrontal Cx	rat and mice	12				a, b, c, d	Emmons et al. (2017)(a); Kim et al. (2016) (b); Parker et al. (2015) (c); (2017) (d)
	Cerebellum	rat	12				a, b	Emmons et al. (2017)(a); Parker et al. (2017) (b)
Serial fixed interval	Dorsal striatum	rat	10 - 60			a		Mello et al. (2015) (a)

$ \begin{array}{ c c c c c c c c c c c c c c c c c c c$	Tasks	Brain structures	Species	Durations (in s)	Results				References
Parkovian aversive delay         Audiory Cx         rat         2         a, b         Annany et al. (1099) (b): Quik et al. (1097)           Parkovian aversive trace         Performal Cx         rat         30         a         Pendyam et al. (2013) (a)           Pavlovian aversive trace         Performal Cx         rat         30         a         Pendyam et al. (2013) (a)           Pavlovian aversive trace         Performal Cx         rat         30         a         Ginamitian add Relectiona could (10)           Pavlovian appetitive delay         Anaguita         monkey         2         a         Bermadore et al. (2012) (a)           Pavlovian appetitive delay         Anaguita         monkey         1         6         a         Performal Cx           Pavlovian appetitive trace         Orbiofontal Cx         mice         2         a         Pavlovian appetitive trace         Basolaterial argyatalia         rotio at al. (2008) (a)           Delayed matching to sample / working         Parietal CX         monkey         0.5 - 2         a         Basolaterial argyatalia         rotio at al. (2008) (a)           memory         Performal Cx         monkey         0.5 - 2.5         a         Basolaterial argyatalia         rotio at al. (2009) (a). (b). Favlar at al. (2009) (b). Favlar at al. (2009) (c). The roti at al. (2009) (c). The roti at a					Sustained activity	Phasic response at onset or offset of cue	Event time	Ramping activity	
$ \begin{array}{  c                                  $	Pavlovian aversive delay	Auditory Cx	rat	2			a, b		Armony et al. (1998) (a); Quirk et al. (1997)
Basal anyalah         rat         30         a         Pendoyan across of the second secon		Prefrontal Cx	rat	30	а				Pendyam et al. (2013) (a)
Parkovian aversive trace         Performal Cs         rat         20         s.b         Additional Additional (2005) (a)           Pavlovian appetitive delay         Angulal         monkey         2          a         Berlon (2005) (a)           Nakosa accumbers         rat         10         a          Day et al (2006) (a)           Nakosa accumbers         rat         10         a          Day et al (2006) (a)           Nakosa accumbers         rat         10         a          Day et al (2006) (a)           Ventral legmental area         monkey         1-16         a          Fortile et al. (2008) (a)           Previovian appetitive trace         Orbitiofrontal CX         mice         2         a          Anse and Shaden(2005) (a)           Parkot AL CX         monkey         0.5 - 2.5         a         Janssen and Shaden(2005) (a)            memory         Perfornal CX         monkey         0.5 - 2.5         a         a         Restinant and Louber (2006) (a)           Marce CX         monkey         0.5 - 0.5         a         a         Restinant and Louber (2006) (a)           Marce CX         monkey         0.5 - 0.5         a         a		Basal amygdala	rat	30	а				Pendyam et al. (2013) (a)
Hippocampus         mbibit         10-20         a, b         McRebra of al (2003) (a)           Pavlovian appetitive delay         Amrgydala         monkey         2         a         Bernukez et al (2012) (a)           Nackaus accumbens         raf         10         a         Day et al (2006) (a)           Subtanti nigna         monkey         1 - 16         a         Forillo et al (2008) (a)           Pavlovian appetitive trace         Orbitofortal Cs         mice         2         a         Territo et al (2006) (a)           Basolateral anyagotial         en c         1.5         a         Paz et al (2006) (a)           Delayed matching to sample / working         Parietal Cx         monkey         0.5 - 2         a         Amsteral Ansates and Shadden (2005) (a)           memory         Prefrontal Cx         monkey         0.5 - 25         a         Amsteral Ansates and Shadden (2006) (a); Tir et al. (2009) (b); Starer et al. (2009) (b); Rossi-Pool et al. (2009)	Pavlovian aversive trace	Prefrontal Cx	rat	20				а	Gilmartin and McEchron (2005) (a)
Pavlovian appetitive delay       Auggala       monkey       2       a       Bermadez et al. (2012) (a)         Nucleas accurses       nat       Day et al. (2006) (a)       Day et al. (2006) (a)         Ventral tegmental ara       monkey       1 - 16       a       Forillo et al. (2008) (a)         Pavlovian appetitive trace       Orbitoffontal Cx       monkey       1 - 16       a       Forillo et al. (2008) (a)         Delayed matching to sample / working       Parisel CX       monkey       0.5 - 2       a       Pave et al. (2005) (a)         Delayed matching to sample / working       Parisel CX       monkey       0.5 - 60       b, g       c       b, e       a, b, c, d, f, g       Brody et al. (2005) (a)         memory       Prefrontal Cx       monkey       0.5 - 2.5       a       Augeate et al. (2006) (b)       Statter et al. (2006) (b)         Motor Cx       monkey       0.5 - 2.5       a       a       Rearbinam et al. (2006) (b)       Statter et al. (2006) (b)         Motor Cx       monkey       0.5 - 2.5       a       a       Rearbinam et al. (2006) (b)       Statter a		Hippocampus	rabbit	10 - 20			a, b		McEchron et al. (2003) (a)
Nucleas accumbens         rat         10         a         Day et al. (2006) (a)           Subuniani agentitive trace         Orbinofontal Cx         monkey         1 - 16         a         Findle out al. (2008) (a)           Pavlovian appetitive trace         Orbinofontal Cx         mice         2         a         Zotu et al. (2005) (a)           Delayed matching to sample / working         Pareital CX         monkey         0.5 - 2         a         Pavlovian all Shadlen (2005) (b)           msmory         Prefinatal CX         monkey         0.5 - 60         b, g         c         b, c         a, b, c         R, Brody et al. (2005) (c) hours et al. (2003) (c) hours et al. (2009)	Pavlovian appetitive delay	Amygdala	monkey	2				а	Bermudez et al. (2012) (a)
		Nucleus accumbens	rat	10	а				Day et al. (2006) (a)
Vertral tegeneral area         monkey         1 · 1 · 6         a         Findle stal. (2009) (a)           Pavlovian appetitive trace         Orbitofontal Cx         mice         2         a         Zhou et al. (2015) (a)           Basolaterial amygdala         cat         1.5         a         Paz et al. (2006) (a)           Delayed matching to sample / working         Parical Cx         monkey         0.5 - 2         a         Jamsen and Shudden (2005) (a)           memory         Perfontal Cx         monkey         0.5 - 60         b, g         c         a, b, c d, f, g         Poyle atl. (2004) (c); Jin et al. (1982) (f); Horster at Al. (1982) (f); Horster at Al. (2010) (g), Sakari at Alexander (1971) (b); Ruster et al. (2014) (g)           memory         Orbitofontal Cx         monkey         0.5 - 2.5         a         monkey         Reschand Olson (2005) (a)           Inferotemporal Cx         monkey         0.5 - 2.5         a         a         Reutiman et al. (2004) (b); C); Jin et al. (1992) (f); Mole cal. (2014) (g); Sakari et al. (1992) (h); Sakari et al. (1992) (h); Sakari et al. (2093) (a)           Inferotemporal Cx         monkey         1.5 - 0         a         Reutiman et al. (2004) (a)           Titual mum         monkey         2 - 4         a, b, c         a, b         Histoskar et al. (1993) (h); Jin et al. (2004) (a)           Subiciuum<		Subtantia nigra	monkey	1 - 16		а			Fiorillo et al. (2008) (a)
Pavlovian appetitive trace         Obioinformal Cx         mice         2         a         Zhou et al. (2015) (a)           Delayed matching to sample / working memory         Parienal Cx         monkey         0.5 - 2         a         Parienal Cx         a         Paze et al. (2006) (a)           Delayed matching to sample / working memory         Prefrontal Cx         monkey         0.5 - 60         b, g         c         b, e         a, b, c, f, g         Brody et al. (2003) (a): Fuster and Alexander h, i, j           Orbitofrontal Cx         monkey         0.5 - 2.5         a         monkey         0.6 - 9.5         monkey         for an off colority (b): Fuster et al. (920) (c). In et al. (2009) (b): Skathman-Rakic (1982) (c): Machene et al. (2010) (g); Rainer et al. (2009) (b): Rost-rot et al. (2010) (g); Rainer et al. (2009) (b): Rost-rot al. (2009) (a)           Inferotemporal Cx         monkey         5 - 8         a         Resech and Olson (2005) (a)           Inferotemporal Cx         monkey         5 - 8         a         Resultmant et al. (2009) (a): Obmae et al. (2008) (b): Rost-rot et al. (2019) (a)           Thalama         monkey         1 - 5 - 0         a         a, b         c         Naryama and Labakeh (2009) (a): Obmae et al. (2008) (b): Rost-rot oct al. (2019) (a)           Thalama         monkey         2 - 5         a         a         Schout et al. (1992) (a)		Ventral tegmental area	monkey	1 - 16		а			Fiorillo et al. (2008) (a)
Basolateral amygdala         cat         1.5         a         Par et al. (2006) (a)           Delayed matching to sample / working         Parietal CX         monkey         0.5 - 2         a         Janssen ad Shadlen (2005) (a)           memory         Preffonal CX         monkey         0.5 - 60         b, g         c         b, e, d, f, g         Brody et al. (2003) (a): Fuster and Alexander (1971) (b): Fuster et al. (1982) (c): Jin et al. (1982) (c): Jin et al. (1982) (c): Jin et al. (1992) (b): Sakari et al. (2004) (d): Savari et al. (2004) (d): Machers et al. (2004) (d): Savari et al. (2004) (d): Machers et al. (2004) (d): Machers et al. (2004) (a)           Motor CX         monkey         0.5 - 2.5         a         Reseth and Olson (2005) (a)           Inferotemporal CX         monkey         5 - 8         a         Reseth and Olson (2005) (a)           Motor CX         monkey         15 - 60         a         a, b. C         Narayaran and Laubach (2009) (a): Ohmac et al. (2008) (b): Tersimbax et al. (2009) (c): Tersimbax et al. (2004) (f): Tersimax et al. (2004) (f): Ters	Pavlovian appetitive trace	Orbitofrontal Cx	mice	2	a				Zhou et al. (2015) (a)
Delayed matching to sample / working memoryParefontal Cxmonkey $0.5 - 2$ aJanssen and Shadlen (2005) (a)memoryPreffontal Cxmonkey $0.5 - 60$ b, gcb, cf, jBrody et al. (2003) (a): Fuster and Alexander (1982) (c): Jin et al. (2009) (d). Joseph and Barone (1987) (c): Korina and Goldman-Rakit (1982) (c): Jin et al. (2009) (d). Joseph and Barone (1987) (c): Korina and Goldman-Rakit (1982) (f): Machers et al. (2010) (g): Rainer et al. (2009) (d). Joseph and Barone (1987) (f): Statuiner et al. (2004) (fi): Tsujimoto and Sawagachi, (2005) (j)Orbitofrontal Cxmonkey $0.5 - 2.5$ aRoesch and Olson (2005) (a)Inferotemporal Cxmonkey $5 - 3$ aNarayanan and Laubach (2009) (a); Ohmae et al. (2006) (b); Rossi-Pool et al. (2019) (c): Rossi-Pool et a		Basolateral amygdala	cat	1.5				а	Paz et al. (2006) (a)
memoryPrefrontal Cxmonkey $0.5 \cdot 60$ b, gcb, ea, b, c, d, f, g, h, i, jBrody et al. (2003) (a); Fuster and Alexander (1971) (b); Fuster et al. (1982) (c); Jin et al. (2009) (d), Joseph and Barone (1987) (e); Kajima and Goldman-Rakie (1982) (f); Machens et al. (2010) (d), Joseph and Barone (1987) (e); Machens et al. (2010) (g); Rainer et al. (2009) (d); Rainer et al. (2009) (d), Joseph and Barone (1987) (e); Machens et al. (2010) (d), Joseph and Barone (1987) (e); Machens et al. (2010) (d), Joseph and Barone (1987) (e); Machens et al. (2010) (d), Joseph and Barone (1987) (e); Machens et al. (2010) (d), Statistical et al. (2009) (d); Raiser et al. (2009	Delayed matching to sample / working	Parietal Cx	monkey	0.5 - 2				а	Janssen and Shadlen (2005) (a)
Orbitofrontal Cx     monkey     0.5 - 2.5     a     Reesch and Olson (2005) (a)       Inferotemporal Cx     monkey     5 - 8     a     Reutimann et al. (2004) (a)       Motor Cx     monkey     0.5 - 3     a     b     a, b, c     Narayanan and Laubach (2009) (a); Ohma et al. (2019) (c)       Premotor CX     monkey     15 - 60     a     a, b, c     Narayanan and Laubach (2009) (a); Ohma et al. (2019) (c)       Thalamus     monkey     15 - 60     a     a, b     Fuster and Alexander (1971) (a), (1973) (b)       Striatum     monkey     2 - 4     a, b, c     c     a, b     Hikosaka et al. (1989) (a); Jin et al. (2009)       Ventral striatum     monkey     2 - 5     a     a     a     Schulz et al. (1992) (a)       Subiculum     rat     1 - 30     a     a     Hippocampus     ad (2014) (a)       Expectation     Primary auditory Cx     rat     0.3 - 1.5     a     Jaramillo and Zador (2011) (a)       Premotor CX     monkey and     3 - 6     b     a, b     Donnelly et al. (2015) (a); Sakai (1974) (b)       rat     rat     up to 80 min     a     Taso et al. (2018)     Motor (CX     monkey and     0 - 5       Lateral entorhinal CX     rat     up to 80 min     a     Taso et al. (2016) (a)     Miyazaki et al	memory	Prefrontal Cx	monkey	0.5 - 60	b, g	с	b, e	a, b, c ,d, f, g, h, i , j	Brody et al. (2003) (a); Fuster and Alexander (1971) (b); Fuster et al. (1982) (c); Jin et al. (2009) (d), Joseph and Barone (1987) (e); Kojima and Goldman-Rakic (1982) (f); Machens et al. (2010) (g); Rainer et al. (1999) (h); Sakurai et al. (2004) (i); Tsujimoto and Sawaguchi, (2005) (j)
$ \begin{array}{ c c c c c c c c c c c c c c c c c c c$		Orbitofrontal Cx	monkey	0.5 - 2.5		а			Roesch and Olson (2005) (a)
$ \begin{array}{c ccccccccccccccccccccccccccccccccccc$		Inferotemporal Cx	monkey	5 - 8				а	Reutimann et al. (2004) (a)
$ \begin{array}{c c c c c c c c c c c c c c c c c c c $		Motor Cx Premotor Cx	monkey	0.5 - 3	a	b		a, b, c	Narayanan and Laubach (2009) (a); Ohmae et al. (2008) (b); Rossi-Pool et al. (2019) (c)
$ \begin{array}{c c c c c c c c c c c c c c c c c c c $		Thalamus	monkey	15 - 60	а			a,b	Fuster and Alexander (1971) (a), (1973) (b)
Ventral striatummonkey2 - 5aaaSchultz et al. (1992) (a)Subiculumrat1 - 30aaaHampson and Deadwyler (2003) (a)Hippocampusmonkey0.7 - 1.2aSakon et al. (2014) (a)ExpectationPrimary auditory Cxrat0.3 - 1.5aJaramillo and Zador (2011) (a)Prefrontal Cxmonkey and3 - 6ba, bDonnelly et al. (2015) (a); Sakai (1974) (b)ratrat0 - 5a, bLucchetti and Bon (2001) (a); Lucchetti et al. (2005) (b)Motor Cxmonkey and0 - 5a, bLucchetti and Bon (2001) (a); Lucchetti et al. (2005) (b)Lateral entorhinal Cxratup to 80 minaTsao et al. (2018)Posterior thalamusrat0 - 2aMiyazaki et al. (2011) (a)Ventral tegmental arearat8aaTotah et al. (2013) (a); Hikosaka et al. (2005) (b)Ventral tegmental arearat8abDonnelly et al. (2015) (a); Kakamassi et al. (2008) (b)Ventral striatumrat1 - 5abDonnelly et al. (2015) (a); Khamassi et al. (2008) (b)		Striatum	monkey	2 - 4	a, b, c		c	a, b	Hikosaka et al. (1989) (a); Jin et al. (2009) (b); Tremblay et al. (1998) (c)
Subiculumrat1 - 30aaHampson and Deadwyler (2003) (a)Hippocampusmonkey0.7 - 1.2aSakon et al. (2014) (a)ExpectationPrimary auditory Cxrat0.3 - 1.5aJaramillo and Zador (2011) (a)Prefrontal Cxmonkey and3-6ba, bDonnelly et al. (2015) (a); Sakai (1974) (b)ratnatnatnatnatnatnatMotor Cxmonkey and0 - 5a, bLucchetti and Bon (2001) (a); Lucchetti et al. (2005) (b)Lateral entorhinal Cxratup to 80 minaTsao et al. (2018)Posterior thalamusrat0 - 2aMiyazaki et al. (2011) (a)Dorsal rapherat1.5 - 20aMiyazaki et al. (2011) (a)Ventral tegmental arearat8aaTotah et al. (2013) (a); Hikosaka et al. (2009) (b)Ventral tegmental arearat1.5 - 4.5ca, b, cApicella et al. (2001) (a); (b); Sardo et al. (2000) (c)Ventral striatumrat1 - 5aDonnelly et al. (2015) (a); Khamassi et al. (2008) (b)		Ventral striatum	monkey	2 - 5			а	а	Schultz et al. (1992) (a)
Hippocampusmonkey0.7 - 1.2aSakon et al. (2014) (a)ExpectationPrimary auditory Cxrat0.3 - 1.5aJaramillo and Zador (2011) (a)Prefrontal Cxmonkey and3-6ba, bDonnelly et al. (2015) (a); Sakai (1974) (b)ratMotor Cxmonkey and0 - 5a, bLucchetti and Bon (2001) (a); Lucchetti et al. (2005) (b)Lateral entorhinal Cxratup to 80 minaTsao et al. (2018)Posterior thalamusrat0 - 2aKomura et al. (2001) (a)Dorsal rapherat1.5 - 20aMiyazaki et al. (2011) (a)Ventral tegmental arearat8aTotah et al. (2013b) (a)Striatum (caudate and putamen)monkey1.5 - 4.5ca, b, cApicella et al. (1992) (a); Hikosaka et al. (2008) (b)Ventral striatumrat1 - 5abDonnelly et al. (2015) (a); Khamassi et al. (2008) (b)		Subiculum	rat	1 - 30	a			а	Hampson and Deadwyler (2003) (a)
ExpectationPrimary auditory Cx ratrat0.3 - 1.5aJaramillo and Zador (2011) (a)Prefrontal Cx ratmonkey and rat3-6ba, bDonnelly et al. (2015) (a); Sakai (1974) (b)Motor Cx Premotor Cx Lateral entorhinal Cxmonkey and rat0 - 5a, bLucchetti and Bon (2001) (a); Lucchetti et al. (2005) (b)Premotor Cx Lateral entorhinal Cx Posterior thalanusratup to 80 minaTsao et al. (2018)Posterior thalanus Ventral tegmental area putamen)rat1.5 - 20aMiyazaki et al. (2011) (a)Ventral tegmental area putamen)rat1.5 - 4.5caTotah et al. (2013b) (a)Ventral striatum ratrat1 - 5abDonnelly et al. (2005) (b)Ventral striatum ratrat1 - 5abDonnelly et al. (2013b) (a)Ventral striatum ratrat1 - 5abDonnelly et al. (2015) (a); Khamassi et al. (2008) (b)Ventral striatum ratrat1 - 5abDonnelly et al. (2015) (a); Khamassi et al. (2008) (b)		Hippocampus	monkey	0.7 - 1.2				а	Sakon et al. (2014) (a)
Prefrontal Cxmonkey and rat3-6ba, bDonnelly et al. (2015) (a); Sakai (1974) (b)Motor Cxmonkey and rat0 - 5a, bLucchetti and Bon (2001) (a); Lucchetti et al. (2005) (b)Premotor Cxratup to 80 minaTsao et al. (2018)Lateral entorhinal Cxrat0 - 2aKomura et al. (2001) (a)Posterior thalamusrat0 - 2aMiyazaki et al. (2011) (a)Dorsal rapherat1.5 - 20aMiyazaki et al. (2013b) (a)Ventral tegmental arearat8aTotah et al. (2013b) (a)Striatum (caudate and putamen)monkey1.5 - 4.5ca, b, cVentral striatumrat1 - 5aDonnelly et al. (2015) (a); Khamassi et al. (2008) (b)Out of the striaturerat1 - 5aDonnelly et al. (2015) (a); Khamassi et al. (2008) (b)	Expectation	Primary auditory Cx	rat	0.3 - 1.5		а			Jaramillo and Zador (2011) (a)
Motor Cxmonkey and0 - 5a, bLucchetti and Bon (2001) (a); Lucchetti et al. (2005) (b)Premotor Cxratup to 80 minaTsao et al. (2018)Lateral entorhinal Cxratup to 80 minaKomura et al. (2001) (a)Posterior thalamusrat0 - 2aMinyazki et al. (2011) (a)Dorsal rapherat1.5 - 20aMinyazki et al. (2011) (a)Ventral tegmental arearat8aTotah et al. (2013b) (a)Striatum (caudate andmonkey1.5 - 4.5ca, b, cApicella et al. (1992) (a); Hikosaka et al. (1989) (b); Sardo et al. (2000) (c )Putamer)rat1 - 5abDonnelly et al. (2015) (a); Khamassi et al. (2008) (b)		Prefrontal Cx	monkey and rat	3-6			b	a, b	Donnelly et al. (2015) (a); Sakai (1974) (b)
Premotor $Cx$ rat $(2005)$ (b)Lateral entorhinal $Cx$ ratup to 80 minaTsao et al. (2018)Posterior thalamusrat0 - 2aKomura et al. (2001) (a)Dorsal rapherat1.5 - 20aMiyazaki et al. (2011) (a)Ventral tegmental arearat8aTotah et al. (2013b) (a)Striatum (caudate and putamen)monkey1.5 - 4.5ca, b, cApicella et al. (1992) (a); Hikosaka et al. (1989) (b); Sardo et al. (2000) (c )Ventral striatumrat1 - 5abDonnelly et al. (2015) (a); Khamassi et al. (2008) (b)		Motor Cx	monkey and	0 - 5				a, b	Lucchetti and Bon (2001) (a); Lucchetti et al.
Lateral entorhnal Cxratup to 80 mmaTsao et al. (2018)Posterior thalamusrat0 - 2aKomura et al. (2001) (a)Dorsal rapherat1.5 - 20aMiyazaki et al. (2011) (a)Ventral tegmental arearat8aTotah et al. (2013b) (a)Striatum (caudate and putamen)monkey1.5 - 4.5ca, b, cApicella et al. (1992) (a); Hikosaka et al. (1989) (b); Sardo et al. (2000) (c )Ventral striatumrat1 - 5abDonnelly et al. (2015) (a); Khamassi et al. (2008) (b)		Premotor Cx	rat						(2005) (b)
Posterior thalamusrat0 - 2aKomura et al. (2001) (a)Dorsal rapherat1.5 - 20aMiyazaki et al. (2011) (a)Ventral tegmental arearat8aTotah et al. (2013b) (a)Striatum (caudate and putamen)monkey1.5 - 4.5ca, b, cApicella et al. (1992) (a); Hikosaka et al. (1989) (b); Sardo et al. (2000) (c )Ventral striatumrat1 - 5abDonnelly et al. (2015) (a); Khamassi et al. (2008) (b)		Lateral entorhinal Cx	rat	up to 80 min				а	Isao et al. (2018)
Dorsal rapherat1.5 - 20aMiyazaki et al. (2011) (a)Ventral tegmental arearat8aTotah et al. (2013b) (a)Striatum (caudate and putamen)monkey1.5 - 4.5ca, b, cApicella et al. (1992) (a); Hikosaka et al. (1989) (b); Sardo et al. (2000) (c)Ventral striatumrat1 - 5abDonnelly et al. (2015) (a); Khamassi et al. (2008) (b)		Posterior thalamus	rat	0-2				a	Komura et al. $(2001)$ (a)
Ventral tegmental area     rat     8     a     1 of table tal. (2013b) (a)       Striatum (caudate and monkey     1.5 - 4.5     c     a, b, c     Apicella et al. (1992) (a); Hikosaka et al. (1989) (b); Sardo et al. (2000) (c)       Putamen)     rat     1 - 5     a     b     Donnelly et al. (2015) (a); Khamassi et al. (2008) (b); (b); Sardo et al. (2009) (c)		Dorsal raphe	rat	1.5 - 20	a				Miyazaki et al. (2011) (a)
Striatum (caudate and monkey 1.5 - 4.5 c a, b, c Apicella et al. (1992) (a); Hikosaka et al. (1989) (b); Sardo et al. (2000) (c) Ventral striatum rat 1 - 5 a b Donnelly et al. (2015) (a); Khamassi et al. (2008) (b) Caula libration of the string of the		Ventral tegmental area	rat	8				a	Totah et al. (2013b) (a)
Ventral striatum rat 1 - 5 a b Donnelly et al. (2015) (a); Khamassi et al. (2008) (b)		Striatum (caudate and putamen)	monkey	1.5 - 4.5	с			a, b, c	Apicella et al. (1992) (a); Hikosaka et al. (1989) (b); Sardo et al. (2000) (c)
		Ventral striatum	rat	1 - 5			a	b	Donnelly et al. (2015) (a); Khamassi et al. (2008) (b)
Cerebellum mice I a wagner et al. (2017) (a)		Cerebellum	mice	1				а	Wagner et al. (2017) (a)
EntrainmentPremotor Cxmonkey0.45 - 1ba, b, cbCrowe et al. (2014)(a); Merchant et al. (2011)(b) (2013b)(c)	Entrainment	Premotor Cx	monkey	0.45 - 1	b		a, b, c	b	Crowe et al. (2014)(a); Merchant et al. (2011) (b) (2013b) (c)
Striatum monkey 0.45 - 1 a Bartolo et al. (2014) (a)		Striatum	monkey	0.45 - 1			a		Bartolo et al. (2014) (a)

Tasks	Brain structures	Species	Durations (s)	Results		References		
				PSD	Coherence	Other	_	
Time production	Parietal Cx	monkey	1			multiunits	Schneider and Ghose (2012)	
Temporal discrimination	Prefrontal Cx	monkey	0.2 - 2			Single unit recordings	Oshio et al. (2008)	
		rat	3 - 4			Single unit recordings	Kim et al. (2013); Tiganj et al (2017)	
	Frontal Cx Hippocampus Cerebellum	rat	2 - 8			ERP	Onoda et al. (2003)	
	Frontal Cx Striatum Thalamus	rat	0.5 - 2			ERP	Onoda and Sakata (2006)	
	Hippocampus	rat	1 - 3	theta (4-9 Hz)			Nakazono et al. (2015)	
				<u> </u>		Single unit recordings	Sakurai (2002)	
	Subtantia nigra	mice	0.6 - 2.4			Calcium imaging	Soares et al. (2016)	
	Dorsolateral prefrontal Cx Posterior inferior parietal Cx Posterior cingulate Cx Striatum	monkey	0.4 - 1.5			Increased blood flow	Onoe et al. (2001)	
Peak interval	Striatum	rat	30	theta (6-12 Hz)			Hattori and Sakata (2013)	
Limited Hold (LH)	Medial entorhinal Cx	mice	6			Calcium imaging	Heys and Dombeck (2018)	
Fixed interval	Medial prefrontal Cx	rat and mice	12	delta (4 Hz)			Parker et al. (2014); (2017)	
				delta (1 - 4 Hz)			Kim et al. (2016)	
	Medial prefrontal Cx Striatum	rat	3 - 12	delta (1 - 4 Hz) theta (4 - 8 Hz)			Emmons et al. (2016)	
	Striatum	rat	12	, ,		Single unit recordings	Emmons et al. (2019)	

Tasks	Brain structures	Species	Durations	Results			References
			(in s)	PSD	Coherence	Other	-
Pavlovian appetitive delay	Substantia nigra	monkey	2 - 16			Multiunits	Kobayashi and Schultz (2008)
	Striatum Basolateral amygdala	cat	3		gamma (35-45 Hz)		Popescu et al. (2009)
Pavlovian aversive delay	Auditory Cx	rat	10	gamma (50-80 Hz) beta (10-50 Hz)			Headley and Weinberger (2011); (2013)
	Dorsomedial striatum Basolateral amygdala	rat	10-30	theta (3-6 Hz) gamma (60-70 Hz)	theta (3-6 Hz)		Dallérac et al. (2017)
	Lateral amygdala	cat	15	(00 /0112)		Multiunits and cross- correlograms	Paré and Collins (2000)
	Hippocampus Lateral amygdala	mice	10		theta (5-6 Hz)		Pape et al. (2005)
Pavlovian appetitive trace	Rhinal Cx Basolateral amygdala	cat	1.5	gamma (35-45 Hz)			Bauer et al. (2007)
	Medial entorhinal Cx	mice	5.5			Calcium imaging	Heys and Dombeck (2018)
Pavlovian aversive trace	Hippocampus	mice	0.25			Calcium imaging	Modi et al. (2014)
Delayed matching to sample / working memory	/ Prefrontal Cx	monkey	0.5 - 20			Single unit recordings	Fuster et al. (1982); Jin et al. (2009); Narayanan and Laubach (2009); Tiganj et al. (2018)
		rat	0 - 20			Single unit recordings	Horst and Laubach (2012)
	Striatum	monkey	2 - 4			Single unit recordings	Soltysik et al. (1975)
	Hippocampus	monkey	0.5 - 1			Single unit recordings	Naya & Suzuki (2011)
		rat	10 - 20			Single unit recordings	Gill et al. (2012); Kraus et al. (2013); MacDonald et al. (2011), (2013); Manns et al. (2007); Pastalkova et al. (2008); Salz et al. (2016): Robinson et al. (2017)
Expectation	Anterior cingulate Cx Prelimbic Cx	rat	8	delta (1 - 4 Hz) theta (8 - 12 Hz)			Totah et al. (2013a)
	Hippocampus (CA1)	mice	10	10 12 1121		Calcium imaging	Mau et al. (2018)
	Primary visual Cx	rat	1 - 2	theta (6-9 Hz)			Zold and Shuler (2015)
	Lateral entorhinal Cx	rat	up to 80 min			Single unit recordings	Tsao et al. (2018)
	Motor Cx	monkey	0.7 - 2	beta (12-40 Hz)			Kilavik et al. (2012)
Entrainment	Lateral habenula	zebrafish	20 - 30			Calcium imaging	Cheng et al. (2014)
	Hippocampus	mice	0.15	theta (4-12 Hz)			Abe et al. (2014)
	Striatum	monkey	0.45 - 1	beta (10 - 30 Hz) gamma (30 - 80 Hz)	1		Bartolo et al. (2014)
	Piriform Cx	rat	240			glutamate and GABA micro- dialwais	Hegoburu et al. (2009)
Free exploration	Hippocampus	rat	hours to days			Single unit recordings	Mankin et al. (2012), (2015)
						Calcium imaging	Rubin et al. (2015)