

# On the epigenetic origin of cancer stem cells

Audrey Vincent, Isabelle van Seuningen

### ▶ To cite this version:

Audrey Vincent, Isabelle van Seuningen. On the epigenetic origin of cancer stem cells. Biochimica et Biophysica Acta (BBA) - Reviews on Cancer, 2012, 10.1016/j.bbcan.2012.03.009. hal-02905665

### HAL Id: hal-02905665 https://hal.science/hal-02905665

Submitted on 29 Sep 2020

**HAL** is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers. L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

### **AUTHOR QUERY FORM**

s s'én	Journal: BBACAN	Please e-mail or fax your responses and any corrections to: E-mail: <u>corrections.essd@elsevier.spitech.com</u> Fax: +1 619 699 6721
ELSEVIER	Article Number: 87852	

Dear Author,

Please check your proof carefully and mark all corrections at the appropriate place in the proof (e.g., by using on-screen annotation in the PDF file) or compile them in a separate list. Note: if you opt to annotate the file with software other than Adobe Reader then please also highlight the appropriate place in the PDF file. To ensure fast publication of your paper please return your corrections within 48 hours.

For correction or revision of any artwork, please consult http://www.elsevier.com/artworkinstructions.

Any queries or remarks that have arisen during the processing of your manuscript are listed below and highlighted by flags in the proof. Click on the 'Q' link to go to the location in the proof.

Location in article	Query / Remark: <u>click on the Q link to go</u> Please insert your reply or correction at the corresponding line in the proof
<u>Q1</u>	Please confirm that given names and surnames have been identified correctly.
<u>Q2</u>	Please provide for the significance of Table 1 entries in italics emphasis.
<u>Q3</u>	Sentence was slightly modified; please check if appropriate.
<u>Q4, Q5</u>	Please supply the year of publication.
<u>Q6</u>	Please provide the volume number and page range for the bibliography in Ref. [57].
<u>Q7</u>	Please provide the volume number and page range for the bibliography in Ref. [78].

Thank you for your assistance.

# ARTICLE IN PR

Biochimica et Biophysica Acta xxx (2012) xxx-xxx

Contents lists available at SciVerse ScienceDirect

Review

1

9

7

8

21

22

23

38

37

Biochimica et Biophysica Acta



journal homepage: www.elsevier.com/locate/bbacan

#### Audrey Vincent, Isabelle Van Seuningen\* Q13 Inserm, UMR837, Jean-Pierre Aubert Research Center, Team 5 "Mucins, epithelial differentiation and carcinogenesis", Lille, France Université Lille Nord de France, 1 Place de Verdun, 59045 Lille cedex, France Centre Hospitalier Régional et Universitaire de Lille, Place de Verdun, 59037 Lille cedex, France ARTICLE INFO ABSTRACT Article history: Epigenetic mechanisms are the key component of the dynamic transcriptional programming that occurs 24 9 Received 2 December 2011 10 along the process of differentiation from normal stem cells to more specialized cells. In the development of 25 11 Received in revised form 9 March 2012 cancer and according to the cancer stem cell model, aberrant epigenetic changes may ensure the property of 26 12 Accepted 10 March 2012 cancer cells to switch cancer stem cell markers on and off in order to generate a heterogeneous population of 27 Available online xxxx 13 cells. The tumour will then be composed of tumourigenic (cancer stem cells) and non-tumourigenic (the side 28 18 population that constitutes the bulk of the tumour) cells. Characterizing epigenetic landscapes may thus help 29 17 Keywords: discriminate aberrant marks (good candidates for tumour detection) from cancer stem cell specific profiles. 30 18 Cancer In this review, we will give some insights about what epigenetics can teach us about the origin of cancer stem 31 19Cancer stem cell cells. We will also discuss how identification of epigenetic reprogramming may help designing new drugs 32 20Epigenetics

On the epigenetic origin of cancer stem cells

© 2012 Published by Elsevier B.V. 34

33

36

#### Contents <u>4</u>9

Differentiation

DNA methylation

Histone modifications

1	1.	Introduction—what is epigenetics?
2	2.	The role of epigenetic changes in normal embryonic and adult stem cell differentiation
3	3.	The cancer stem cell hypothesis and the establishment of an epigenetic tumour hierarchy
4	4.	The origin of CSC: what may we learn from epigenetic landscapes?
5	5.	Implications for cancer therapy
6	6.	Conclusion and perspectives
17	Acki	nowledgements
8	Refe	erences

that will specifically target cancer stem cells.

49

#### 1. Introduction-what is epigenetics? 50

Epigenetics define heritable changes in the pattern of gene expression 51that are not mediated by alterations in the primary nucleotide gene 5253sequence [1]. Among these stable but reversible changes, DNA methylation and specific post-translational modifications on NH2-terminal 54 histone tails are key mechanisms that control chromatin compacting. 55DNA methylation is almost entirely limited to CpG dinucleotides while 56

Abbreviations: ESC, embryonic stem cell; CSC, cancer stem cell; HP1, heterochromatin protein 1; PcG, polycomb group protein; HDAC, histone deacetylase; DNMT, DNA methyltransferase; BMI-1, B cell Moloney murine leukemia virus Insertion region

Corresponding author at: Inserm UMR837, Jean-Pierre Aubert Research Center, Team 5 "Mucins, epithelial differentiation and carcinogenesis", Rue Polonovski, 59045 Lille, France, Tel.: + 33 3 20 29 88 67; fax: + 33 3 20 53 85 62.

E-mail address: isabelle.vanseuningen@inserm.fr (I. Van Seuningen).

0304-419X/\$ - see front matter © 2012 Published by Elsevier B.V. doi:10.1016/j.bbcan.2012.03.009

histone modifications include a variety of modifications including 57 acetylation, methylation, phosphorylation, ubiquitination, biotinylation, 58 and SUMOylation. Crosstalks between the numerous writers of the 59 epigenetic code (i.e. DNA methyltransferases, histone acetyltransferases, 60 deacetylases, methyltransferases, demethylases, etc.) define whether 61 genes are activated or repressed [2]. Acetylation of histories H3 and H4 is 62 mostly associated with gene expression. On the contrary, DNA methyl- 63 ation, di- and trimethylation of H3 lysine 9 (H3K9) and trimethylation of 64 H3K27 elicit the compaction of chromatin leading to gene silencing 65 through the recruitment of heterochromatin protein 1 (HP1) and 66 polycomb group (PcG) proteins. 67

Adding a new layer of complexity to the epigenetic machinery, micro-68 RNAs (miRNA) known to target the 3'-untranslated regions (3'-UTR) of 69 mRNA transcripts for degradation or translation repression [3,4], have 70 also recently been shown in human cells to trigger transcriptional 71 silencing via chromatin remodelling [5]. 72

Please cite this article as: A. Vincent, I. Van Seuningen, On the epigenetic origin of cancer stem cells, Biochim. Biophys. Acta (2012), doi:10.1016/j.bbcan.2012.03.009

2

# **ARTICLE IN PRESS**

73 Epigenetic changes are increasingly regarded as key events in the 74 development of cancer. Aberrant DNA methylation profiles (largescale hypomethylation [6,7] associated with specific, non-random, 7576 hypermethylation of CpG islands [8]), histone modification landscapes and miRNA signatures are early features of carcinogenesis 77 occurring in preneoplastic lesions as well as in adjacent tissues. This 78 79has led to the hypothesis that epigenetic changes may predispose 80 cells to malignant transformation or even constitute one of the 81 primary transforming events.

### 2. The role of epigenetic changes in normal embryonic and adult stem cell differentiation

Both embryonic development and stem cell differentiation in adults involve crucial changes in gene expression profiles. Genes specific for each cell fate are activated while pluripotency factors are silenced, most likely by specific epigenetic rearrangements [9,10].

Embryonic stem cell (ESC) status involves multiple layers of molecular events designed to impose a flexible but precise control over the expression of genes important for development [11]. Several factors of pluripotency and self-renewal have been identified, including OCT4, NANOG, SOX2, FOXD3, REX1, KLF4, c-myc, Lin-28 and the Wnt signalling pathway [12,13].

94 Along with the expression of these specific factors, ESC harbours a particular epigenetic landscape with a highly dynamic and permissive 95 chromatin. This state is characterized by a distinct DNA methylation 96 profile, the genome of ESC being globally less methylated than 97 98 differentiated somatic cells, despite a high level of Dnmt3b expression [14,15]. Structural proteins such as HP1 and histone H1 also bind 99 more loosely to chromatin compared to differentiated somatic cells 100 [16,17]. These features are essential for chromatin rearrangements and 101 102therefore for global transcriptional programming upon differentiation. 103 More particularly, developmentally regulated genes are characterized by the presence of both active and repressive histone marks. In that case, 104 large genomic regions with trimethylated H3K27 colocalize with smaller 105regions containing permissive chromatin marks such as methylated 106 H3K4. This chromatin bivalency allows genes encoding specific transcrip-107 108 tion factors or early differentiation factors to be prepared for rapid transcriptional activation upon differentiation [18]. 109

PcG complexes, on the other hand, act to stabilize a repressive chromatin structure and have been shown to play important roles in the direct silencing of cell differentiation regulators such as Otx2, Satb2 and Tbx3 factors [19]. Among the actors of PcG complexes, the B cell Moloney murine leukemia virus insertion region protein BMI-1 serves as a key regulatory component and plays a critical role in the self-renewal of adult stem cells.

Many miRNAs are specifically expressed during differentiation and embryogenesis. Indeed, ESC expresses a unique set of miRNAs [20]. Moreover, some miRNA precursors (pre-miRNAs) expressed in stem cells are not processed and cleaved into mature miRNA until differentiation is initiated [21]. This strongly suggests that miRNA may have a role in the maintenance of the pluripotent cell state and in the regulation of early mammalian development.

Upon differentiation, ESC sees their epigenome rich in euchroma-124tin evolve toward a more compact heterochromatin structure. A 125unique DNA methylation profile is acquired, involving methylation in 126127domains mostly found in regions adjacent to CpG islands, named CpG island shores [22,23]. Acquisition of this specific profile reduces cell 128pluripotency and induces progressive activation of cell- and tissue-129 specific genes [9]. Heterochromatic markers, such as HP1 proteins 130modify their localization from a hyperdynamic to a more concentrat-131 ed state, stabilizing the new cell fate. Thus, the chromatin becomes 132progressively less permissive. This is characterized by a global 133 decrease in active histone marks such as acetylated histones H3 and 134 H4 [16] and by a specific increase in silencing-associated di- and 135136 trimethylated H3K9 [24]. Additionally, bivalent chromatin profiles are changed either to methylated H3K4 at genes specific for the particular 137 cell fate in which they are engaged or to trimethylated H3K27 at 138 genes specific for other cell fates [18]. 139

For instance, Oct4 and Nanog promoters shift from H3K4me3 to 140 H3K27me3 marks as cells differentiate [25] and undergo rapid DNA 141 methylation upon differentiation *in vitro* [13] whereas they are 142 unmethylated in undifferentiated human ESC [26]. Conversely, Oct4 143 and Nanog have been shown to interact with each other and associate 144 with Hdac1/2 and Metastasis Associated 1/2, in a complex named 145 NODE (Nanog and Oct4 associated DEacetylase). This complex 146 contains HDAC activity comparable to NuRD complex [27], hence 147 participating in the epigenetic landscaping specific for stemness. On 148 the contrary, because of changes in DNA methylation and histone 149 modifications in regions enriched for their binding sites, these factors 150 can no longer bind the promoters of their target genes in differentiated 151 cells [11,28].

Epigenetic mechanisms are thus a key component of the dynamic 153 regulation of differentiated *versus* stem cell identities. 154

# 3. The cancer stem cell hypothesis and the establishment of an 155 epigenetic tumour hierarchy 156

The clonal model of carcinogenesis, in which genetic and epigenetic 157 changes occur over time to confer a selective growth and survival 158 advantage to individual clones within a tumour, has long prevailed. 159 However, numerous studies conducted in hematologic malignancies as 160 well as in solid tumours have shown that only a proportion of the cancer 161 cells are able to initiate new tumour growth when xenografted in 162 immunodeficient NOD/SCID mice. The percentages of tumour-initiating 163 cells vary from 0.0001% [29] to 25% [30] depending on xenotransplan-164 tation assay conditions. In most human cancers (melanomas, sarcomas, 165 blood, brain, lung, gastrointestinal, hepatocellular, prostate, breast and 166 ovarian cancers), a panel of candidate markers has been described 167 that helps distinguish between the relatively rare subpopulations of 168 tumourigenic cells from non-tumourigenic (side population) cancer 169 cells (Table 1; [29]).

This observation has led to the cancer stem cell (CSC) model in 171 which tumours would arise from rare to moderately frequent cells able 172 to initiate a hierarchy of proliferative and progressively differentiating 173 non-tumourigenic cells. This non-tumourigenic population of cells 174 would then form the bulk of the tumour. In this model, CSC would have 175 similar properties to normal adult stem cells or ESC. These properties 176 would include (i) the capacity of self-renewal potentially through 177 asymmetric division, (ii) the use of key regulatory pathways and 178 (iii) the establishment of dynamic epigenetic profiles. These profiles 179 would potentially help discriminate CSC from non-tumourigenic 180 cancer cells. Hence, the tumour hierarchical organization would be 181 comparable to that found in normal tissue, where epigenetic changes 182 trigger both cell differentiation and cell fate. CSC, however, are 183 thought to harbour higher proliferative rates and be capable of 184 causing constant expansion of existing tumours or forming new 185 tumours through metastatic processes. Moreover, the dependence 186 to the stem cell niche may provide a tremendous difference between 187 normal stem cells and CSC. In that case, the normal stem cell niche 188 would control the balance between cell proliferation, self-renewal 189 and differentiation [31], while the CSC niche would govern metastatic 190 colonization [32]. 191

Since clonogenicity implies that cancer cells accumulate selective 192 mutations over a relatively long period of time [33,34], it is rather 193 unlikely that genetic changes throughout a neoplastic clone would 194 result in rapid and extensive proliferation limited to only a subpopula- 195 tion of cancer cells. Hence, differences between tumourigenic and non-196 tumourigenic cancer cells may rather reside in their epigenetic profile. 197 Similar to stem cells giving rise to the panel of differentiated cells in each 198 organ, it is thought that CSC may give rise to the cellular heterogeneity of 199 the tumours by initiating epigenetic reprogramming (Fig. 1). Confirming 200

Please cite this article as: A. Vincent, I. Van Seuningen, On the epigenetic origin of cancer stem cells, Biochim. Biophys. Acta (2012), doi:10.1016/j.bbcan.2012.03.009

### ARTICLE IN PRESS

### A. Vincent, I. Van Seuningen / Biochimica et Biophysica Acta xxx (2012) xxx-xxx

#### t1.1 Table 1

Epigenetic regulation of cancer stemness markers.

t1.2 t1.3	Gene	Tumour type	CSC profile	Epigenetic regulation/function	Reference
t1.4	CD34	AMI.	CD34+/CD38-	415 bp CpG island <sup>a</sup>	
t1.5	CD38	Childhood B-ALL	CD133+/CD19-/CD38-	731 bp CpG island <sup>a</sup>	
t1.6	CD19	ALL and CML-BC	CD34 + /CD38 - /CD19 + t(12:21)	Chromatin remodeling	[74]
t1.7	CD24	Breast carcinoma	$CD44 + /CD24^{-/low}/Lin$	No differential methylation	[75]
				between CD44+CD24- and	
				CD44+CD24+	
				breast cancer cells	
t1.8	CD44	Head and neck cancer, prostate,	CD44+	DNA methylation	[76]
		gastric and colorectal carcinoma			
t1.9	Thy-1	Liver cancer	CD90+	DNA methylation	[77]
t1.10	ENG	Renal carcinoma	CD105+		
t1.11	CD117 (c-kit)	Ovarian adenocarcinoma	CD44+/CD117+	DNA methylation and miRNA	[78,79]
t1.12	Prominin-1	Brain tumour, glioblastoma, osteosarcoma, Ewing's	CD133+	Histone modifications and	[28,80-82]
		sarcoma, endometrial, hepatocellular, colon and lung		DNA methylation	
		carcinomas, ovarian and pancreatic adenocarcinoma			
t1.13	Bmi-1	Glioblastoma	CD133+/Bmi1+	Polycomb group protein	[83]
t1.14	ALCAM	Colorectal carcinoma	EpCAM <sup>m</sup> /CD44+/CD166+	DNA methylation	[84]
t1.15	NGFR	Melanoma	CD271+	DNA methylation	[85]
t1.16	ALDH1	Colorectal, breast, prostate and squamous cell	ALDH1+	DNA methylation	[86]
		carcinomas, pancreatic adenocarcinoma, osteosarcoma			
t1.17	EpCAM (ESA)	Colon carcinoma and Pancreatic adenocarcinoma	EpCAM <sup>III</sup> /CD44+ and CD44+/CD24 <sup>+</sup> /EpCAM+	DNA methylation	[87]
t1.18	ASCL1	Lung carcinoma	CD133+/ASCL1 <sup>III</sup> /ALDH <sup>III</sup>	Histone modifications	[88,89]
Q2t1.19	JARID1B	Melanoma		Histone demethylase	[90]
t1.20	PDPN	Squamous cell carcinoma	Podoplanin+	DNA methylation	[91]
t1.21	TACSID2	Prostate carcinoma	TROP2+/CD133+	DNA methylation	[92]

<sup>a</sup> CpG island information corresponds to the prediction of the UCSC Genome Browser on Human Mar. 2006 (NCBI36/hg18) Assembly with the following criteria: GC content of 50% or greater, length greater than 200 bp, ratio of observed number of CG dinucleotides to the expected number on the basis of the number of Gs and Cs in the segment greater t1.22 than 0.6.

201 this hypothesis, the lower level of DNA methylation and repressive histone marks such as H3K27me3 at tumour suppressor genes have 202been found in putative breast and liver CSC compared to their 203"differentiated" counterparts. Therefore, in these tumours, CSC harbours 204 a higher transcriptionally active chromatin state than "differentiated" 205cancer cells [35]. Similarly, histone modifications such as the marker of 206 207 activated gene H3K4me3 have been found predominantly in CSC of AML patients at genes characteristic for stemness, including Bmi1, Notch1 208 and Wnt1 [27]. In head and neck squamous cell carcinomas (HNSCC), 209CD44+ CSC also shows a unique epigenetic signature of 22 genes with a 210211 significant distinct methylation pattern compared with CD44- non-212 stem cells [36].

213 The dynamic property of cancer cells to switch CSC markers off (upon cell proliferation shown by the heterogeneity of the tumour 214 215 cell population [37]) and back on (as described in experimental models [38]) supports an epigenetic mechanism. Hence, virtually all 216217known CSC markers have been shown to be regulated either by DNA methylation or histone modifications (Table 1), or themselves are 218chromatin modifying enzymes (such as BMI-1 or JARID1B [39]). 219Additionally, miR-200c, miR-203 and miR-183, also play an important 220221 role in the regulation of stem cell markers, inducing differentiation in 222pancreatic and colorectal CSC [40]. Consistent with these findings, loss of these stemness-inhibiting miRNA has been shown to lead to 223the establishment of epigenetic patterns required for the formation 224 and maintenance of breast CSC [41]. Signatures of CSC have therefore 225been recently established in breast as well as in liver cancers [42,43]. 226Further analyses will be crucial (i) to understand how this 227

potential hierarchy is established and (ii) to discriminate CSC from 228 non-tumourigenic cancer cells based on their epigenetic landscapes. 229 Aberrant DNA methylation and histone marks that have been 230231 described in cancer for the past two decades may represent marks present in the bulk of the tumour and as such could be considered as 232good candidate markers for tumour detection. On the other hand, 233 234 epigenetic profiles that are specific for CSC may resemble the original 235 cancer-initiating cell, whether it is a dedifferentiated or a normal 236 stem cell, and could be considered as good therapeutic targets.

### 4. The origin of CSC: what may we learn from epigenetic landscapes? 237

Normal adult stem cells are characterized by the capacity to self- 238 renew. Their longevity in the stem cell niche allows normal stem 239 cells, or their immediate progeny, to be the first targets of malignant 240 transformation *via* accumulation of aberrant epigenetic profiles or 241 genetic mutations [44–46]. Hence, improper activation of the normal 242 self-renewal machinery could be a critical event in cancer initiation. 243 Supporting the hypothesis of normal stem cells being the pre-44 decessors of CSC, lineage-tracking studies in the colon have shown 245 that long-lived intestinal stem cells are susceptible to cancer-causing 246 mutations [45,47]. Additionally, genes that are targets of PCG proteins 247 and therefore known to be suppressed in normal stem cells show 248 hypermethylation correlated with aging. This locks the cells in an 249 undifferentiated state, therefore making it the first potential step 250 toward malignant transformation [48].

Finally, gene expression signatures reminiscent of ESC have been 252 found in various types of cancer especially poorly differentiated cancers, 253 and markers of CSC are usually similar to normal stem cell markers in 254 different organs [49]. For instance, Bmi1 has been described as a marker 255 of both adult normal stem cells and CSC in colon, breast and prostate, as 256 well as in hematopoietic and neuronal stem cells. In breast cancer, this 257 key regulatory component of the Polycomb Repressive Complex-1, 258 which regulates stem cell proliferation and blocks terminal differenti-259 ation to more committed cells through regulation of the Hedgehog 260 pathway, is also increased in tumour-initiating cells [50]. 261

Nevertheless, it is also interesting to note that CSC differs from 262 normal adult stem cells in that they do not retain the bivalent chromatin 263 pattern [35]. Additionally, studies analyzing the DNA methylation 264 profile of CSC indicate that epigenetic markers of stemness would 265 resemble the ESC (expression of Dnmt1 and Hdac1) rather than the 266 adult normal stem cell pattern (expression of MLL family of histone 267 methyltransferases) [51]. 268

Hence, the hypothesis of cell reprogramming should definitely 269 not be excluded. Indeed, while cellular differentiation has been 270 thought to be unidirectional, several recent studies have shown 271

Please cite this article as: A. Vincent, I. Van Seuningen, On the epigenetic origin of cancer stem cells, Biochim. Biophys. Acta (2012), doi:10.1016/j.bbcan.2012.03.009

### **ARTICLE IN PRESS**

A. Vincent, I. Van Seuningen / Biochimica et Biophysica Acta xxx (2012) xxx-xxx



Fig. 1. Normal stem cells *versus* CSC, an epigenetic change. In this simplified model, aberrant epigenetic programming would trigger either the reactivation of self-renewal in differentiated cells or the initiation of transformation in normal stem cells to give rise to CSC. CSC would in turn give rise to relatively differentiated cancer cells that may acquire additional epigenetic and genetic alterations and lead to highly proliferative cancer cells. CSC, differentiated cancer cells and highly proliferative cancer cells would constitute the heterogeneity of the tumour. Targeted therapy may force CSC to reenter the normal process of differentiation and could represent a relevant addition to conventional therapies that only target highly proliferative cancer cells.

that the reexpression of only four genes encoding the transcrip-272tion factors c-Myc, Oct4, Sox2 and Klf4 is sufficient to generate 273pluripotency. Expression of these factors can convert a terminally 274differentiated cell into a pluripotent cell by erasing the epigenetic 275pattern associated with cell differentiation. Therefore, epigenetic 276reprogramming that induces the expression of pluripotency factors in 277 differentiated cells would lead to aberrant stemness and cancer. 278Supporting this hypothesis, Oct4, c-myc, Sox2 and Klf4 have all been 279shown to be regulated by epigenetic mechanisms in differentiating 280 [52–54] or neoplastic cells [55]. More recently, Chaffer and collaborators 281 have discovered a striking property of differentiated epithelial breast 282 cells that are able to dedifferentiate and acquire stem-like properties, 283 284 unveiling a noticeable degree of plasticity of non-stem cell compart-285 ments [38].

While mouse models and *in vivo* experiments such as xenotrans- 286 plantation assays have brought great insight into the deciphering of 287 the mechanisms of cancer cell growth and metastasis, it is still 288 impossible, in patients, to retrospectively identify the cell-of-origin of 289 naturally occurring cancers. Therefore, it is yet unknown whether CSC 290 originates from normal adult stem cells, more mature progenitors or 291 even from differentiated cells. As discussed above, there is evidence 292 for each model. However, epigenetic events are more likely to 293 represent ideal dynamic changes for cells that enter a carcinogenetic 294 path. Indeed, according to the "epigenetic progenitor model of 295 cancer" proposed by Feinberg and collaborators early epigenetic 296 disruption of progenitor cells would disturb the normal balance 297 between undifferentiated and committed cells [56]. The epigenetic 298 progenitor landscape would therefore be reminiscent of the cell-of- 299

Please cite this article as: A. Vincent, I. Van Seuningen, On the epigenetic origin of cancer stem cells, Biochim. Biophys. Acta (2012), doi:10.1016/j.bbcan.2012.03.009

374

383

387

388

396

397

398

399

408

409

434

origin while the bulk of the tumour would acquire new epigenetic 300 301 and genetic alterations. The nature of epigenetic alterations involved in 302 CSC generation may strongly depend on the normal cell phenotype from 303 which they arise and cancers would acquire different characteristics according to their origin [57]. Supporting this hypothesis is the fact that 304 transformation of genetically identical but phenotypically different 305 human breast epithelial cells leads to distinct tumour types [58]. This 306 indicates that epigenetic presetting greatly influences the type of 307 308 generated tumour.

309 Therefore, epigenetic landscapes, if they are maintained through 310 CSC self-renewal, would more likely indicate the exact origin of CSC. 311Confirming this hypothesis, symmetric DNA methylation patterns have proven to be very informative to deduce the clonal expansion 312 313 dynamics of CSC and the mechanisms of tumour hierarchy in colorectal cancers [59]. 314

#### 5. Implications for cancer therapy 315

CSCs are more and more considered as the roots of cancer and 316 therefore identifying and targeting CSC for elimination appears as 317 critical for cancer therapy. Most currently used cancer therapies **O3**318 target fast growing cells. If CSC share, as it is thought, the more slowly 319 320 cycling characteristics of their normal counterparts, known treat-321 ments would rather lead to an enrichment in CSC. Recent studies conducted in epithelial cancers and in melanomas have indeed 322 shown that CSC candidates harbour lower proliferation rates than the 323 bulk of the tumour [39,60-62]. Thus, they are mostly resistant to 324 325 current treatments, including radiotherapy and conventional chemotherapy [63-65]. Differentiation therapy through epigenetic drugs, such 326 HDAC or DNMT inhibitors, represents however a particularly interesting 327 and promising approach, as it is known that CSC keeps plasticity for 328 329 differentiation [65]. For instance, the demethylating agent 5-aza-2'deoxycytidine has been shown to inhibit self-renewing population of 330 331 CD34+ cells [66].

These epigenetic drugs have proven valuable as new weapons 332 against cancer, especially leukemia. However, their broad spectrum of 333 inhibition, targeting whole families of enzymes may not be perfectly 334 335 suited to most solid tumours. This is especially true for the specific targeting of CSC which harbours a peculiar epigenetic landscape with 336 the expression of specific epigenetic regulators or enzymes when 337 compared to the rest of the tumour. More targeted epigenetic-based 338 therapies may thus greatly improve patient outcome, as each epigenetic 339 modifier enzyme could be a potential individual target for therapeutic 340 manipulation [44]. 341

For instance, Bmi-1 is currently considered as a potent novel 342 343 target for drug discovery [67]. It has been shown that Bmi-1 silencing, 344either by siRNA or epigenetic drugs such as HDAC inhibitors (sodium butyrate or valproic acid), enhances the sensitivity of cancer cells to 345chemoradiation and inhibits tumour growth [68]. Additionally, 346 therapies using properties of miRNA to specifically inhibit stemness 347 effectors and CSC markers have provided very promising results. For 348 349example, miR-34a inhibits prostate CSC and metastasis by directly 350 repressing CD44 [69], while miR-128 inhibits BMI1 in breast cancer and gliomas [70,71], miR-34a specifically targets Notch in pancreatic 351cancer [72], and miR-199b-5p negatively regulates HES1 in medul-352353 loblastomas [73].

354 Identifying the epigenetic reprogramming that occurs during the acquisition of CSC properties and understanding the differences between 355 normal adult stem cells and CSC will therefore help designing new drugs 356 that will specifically target CSC without altering tissue homeostasis. 357

#### 6. Conclusion and perspectives 358

A new challenge in medical science is to determine the origin of 359 cancer resistance to most current therapies. Evidence have been 360 361 brought that in the battle against cancer, CSC, if they exist, and nonCSC will both need to be targeted with specific strategies to eliminate 362 both the bulk of a tumour and its potential roots. However, as most 363 studies have focused on entire tumours with heterogeneous cell 364 populations, our knowledge on human CSC, a small population of 365 cells so far only identifiable by their capacity to generate new 366 tumours in immunodeficient mice, remains very limited. This issue 367 will more likely be overcome with the use of new tools such as next 368 generation sequencing that allows genome-wide epigenetic profiling 369 of a single stem cell. Hence, defining the early epigenetic reprogram- 370 ming or deprogramming that gives rise to the cancer cell population 371 instead of normal differentiated cells will be crucial for the discovery 372 of new therapeutic strategies of cancer. 373

### Acknowledgements

We thank Pr Michael Goggins (Johns Hopkins University, School of 375 Medicine, Baltimore, USA) for critically reading the manuscript. Dr 376 Audrey Vincent is the recipient of a postdoctoral fellowship from the 377 Association pour la Recherche sur le Cancer (ARC). This work is 378 supported by a grant from la Ligue Nationale contre le Cancer (Equipe 379 Labellisée Ligue 2010, IVS). Dr Isabelle Van Seuningen is the recipient 380 of a grant "Contrat Hospitalier de Recherche Translationnelle"/CHRT 381 2010, AVIESAN. 382

### References

- V.E.A. Russo, R.A. Martienssen, A.D. Riggs, Epigenetic Mechanisms of Gene 384
  - Regulation, Cold Spring Harbor Laboratory Press, New York, 1996. 385 386
- H. Santos-Rosa, C. Caldas, Eur. J. Cancer 41 (2005) 2381-2402.
- D.P. Bartel, Cell 116 (2004) 281-297. [3] L. He, G.J. Hannon, Nat. Rev. Genet. 5 (2004) 522-531.
- S. Gonzalez, D.G. Pisano, M. Serrano, Cell Cycle 7 (2008) 2601-2608.
- 389 A.J. Bannister, P. Zegerman, J.F. Partridge, E.A. Miska, J.O. Thomas, R.C. Allshire, T. 390 [6]
- Kouzarides, Nature 410 (2001) 120-124. 391 Y. Schlesinger, R. Straussman, I. Keshet, S. Farkash, M. Hecht, J. Zimmerman, E. 392
- Eden, Z. Yakhini, E. Ben-Shushan, B.E. Reubinoff, Y. Bergman, I. Simon, H. Cedar, 393 Nat. Genet. 39 (2007) 232-236. 394395
- J.G. Herman, S.B. Baylin, N. Engl. J. Med. 349 (2003) 2042-2054.
- N. Hattori, K. Shiota, FEBS J. 275 (2008) 1624-1630.
- A. Vincent, I. Van Seuningen, Differentiation 78 (2009) 99-107. [10] V.V. Lunyak, M.G. Rosenfeld, Hum. Mol. Genet. 17 (2008) R28-R36. [11]
- K.S. O'Shea, Biol. Reprod. 71 (2004) 1755-1765 [12]
- [13] S. Yeo, S. Jeong, J. Kim, J.S. Han, Y.M. Han, Y.K. Kang, Biochem. Biophys. Res. 400 Commun. 359 (2007) 536-542. 401 C. Allegrucci, L.E. Young, Hum. Reprod. Update 13 (2007) 103-120. 402
- M. Bibikova, E. Chudin, B. Wu, L. Zhou, E.W. Garcia, Y. Liu, S. Shin, T.W. Plaia, J.M. 403 [15] Auerbach, D.E. Arking, R. Gonzalez, J. Crook, B. Davidson, T.C. Schulz, A. Robins, A. 404 Khanna, P. Sartipy, J. Hyllner, P. Vanguri, S. Savant-Bhonsale, A.K. Smith, A. 405 Chakravarti, A. Maitra, M. Rao, D.L. Barker, J.F. Loring, J.B. Fan, Genome Res. 16 406 (2006) 1075-1083. 407
- [16] L. Chen, G.Q. Daley, Hum. Mol. Genet. 17 (2008) R23-R27
- P. Collas, A. Noer, S. Timoskainen, J. Cell. Mol. Med. 11 (2007) 602-620. [17]
- [18] B.E. Bernstein, T.S. Mikkelsen, X. Xie, M. Kamal, D.J. Huebert, J. Cuff, B. Fry, A. 410 Meissner, M. Wernig, K. Plath, R. Jaenisch, A. Wagschal, R. Feil, S.L. Schreiber, E.S. 411 Lander, Cell 125 (2006) 315-326. 412
- [19] D. Pasini, A.P. Bracken, J.B. Hansen, M. Capillo, K. Helin, Mol. Cell. Biol. 27 (2007) 413 3769-3779. 414 415
- [20] H.B. Houbaviy, M.F. Murray, P.A. Sharp, Dev. Cell 5 (2003) 351-358
- [21] A. Rybak, H. Fuchs, L. Smirnova, C. Brandt, E.E. Pohl, R. Nitsch, F.G. Wulczyn, Nat. 416 Cell Biol. 10 (2008) 987-993. 417
- [22] A. Doi, I.H. Park, B. Wen, P. Murakami, M.J. Aryee, R. Irizarry, B. Herb, C. Ladd-418 Acosta, J. Rho, S. Loewer, J. Miller, T. Schlaeger, G.Q. Daley, A.P. Feinberg, Nat. 419 Genet. 41 (2009) 1350-1353. 420
- [23] R.A. Irizarry, C. Ladd-Acosta, B. Wen, Z. Wu, C. Montano, P. Onyango, H. Cui, K. 421 Gabo, M. Rongione, M. Webster, H. Ji, J.B. Potash, S. Sabunciyan, A.P. Feinberg, Nat. 422 Genet. 41 (2009) 178-186 423
- [24] B. Wen, H. Wu, Y. Shinkai, R.A. Irizarry, A.P. Feinberg, Nat. Genet. 41 (2009) 424 246-250. 425
- [25] G. Pan, S. Tian, J. Nie, C. Yang, V. Ruotti, H. Wei, G.A. Jonsdottir, R. Stewart, J.A. 426 Thomson, Cell Stem Cell 1 (2007) 299-312. 427
- M.A. Lagarkova, P.Y. Volchkov, A.V. Lyakisheva, E.S. Philonenko, S.L. Kiselev, Cell 428 [26] Cycle 5 (2006) 416-420. 429
- J. Yamazaki, M.R. Estecio, J. Jelinek, D. Graber, Y. Lu, L. Ramagli, S. Liang, S.M. [27] 430 Kornblau, J.P. Issa, ASH Annual Meeting, vol. 116, American Society of 431 Hematology, 2010. 432 433
- [28] H. You, W. Ding, C.B. Rountree, Hepatology 51 (2010) 1635-1644.
- [29] S. Sell, Am. J. Pathol, 176 (2010) 2584-494.

Please cite this article as: A. Vincent, I. Van Seuningen, On the epigenetic origin of cancer stem cells, Biochim. Biophys. Acta (2012), doi:10.1016/j.bbcan.2012.03.009

- [30] E. Quintana, M. Shackleton, M.S. Sabel, D.R. Fullen, T.M. Johnson, S.I. Morrison, 435436 Nature 456 (2008) 593-598.
- 437 [31] T.M. Yeung, L.A. Chia, C.M. Kosinski, C.J. Kuo, Cell. Mol. Life Sci. 68 (2011) 438 2513-2523
- 439[32] I. Malanchi, A. Santamaria-Martinez, E. Susanto, H. Peng, H.A. Lehr, J.F. Delaloye, J. **O4**440 Huelsken, Nature 481 85-9.
  - 441 [33] S Jones W D Chen G Parmigiani F Diehl N Beerenwinkel T Antal A Traulsen M.A. Nowak, C. Siegel, V.E. Velculescu, K.W. Kinzler, B. Vogelstein, J. Willis, S.D. 442 Markowitz Proc Natl Acad Sci U S A 105 (2008) 4283-4288 443
  - [34] S. Yachida, S. Jones, I. Bozic, T. Antal, R. Leary, B. Fu, M. Kamiyama, R.H. Hruban, J.R. 444 Eshleman, M.A. Nowak, V.E. Velculescu, K.W. Kinzler, B. Vogelstein, C.A. Iacobuzio-Donahue, Nature 467 (2010) 1114–1117. 445446
  - [35] H. Yasuda, K. Soejima, H. Watanabe, I. Kawada, I. Nakachi, S. Yoda, S. Nakayama, R. 447 448 Satomi, S. Ikemura, H. Terai, T. Sato, S. Suzuki, Y. Matsuzaki, K. Naoki, A. Ishizaka, Int. J. Oncol. 37 (2010) 1537-1546. 449
  - [36] H.H. Zhang, A. Stone, S.A. Schichman, B.R. Smoller, J.Y. Suen, C.Y. Fan, Mod. Pathol. 45045123 (2010) 1266.
  - 452[37] Y.P. Choi, H.S. Shim, M.Q. Gao, S. Kang, N.H. Cho, Cancer Lett. 307 (2011) 62-71.
  - 453[38] C.L. Chaffer, I. Brueckmann, C. Scheel, A.J. Kaestli, P.A. Wiggins, L.O. Rodrigues, M. 454Brooks, F. Reinhardt, Y. Su, K. Polyak, L.M. Arendt, C. Kuperwasser, B. Bierie, R.A. 455Weinberg, Proc. Natl. Acad. Sci. U. S. A. 108 (2011) 7950-7955.
  - 456 [39] A. Roesch, M. Fukunaga-Kalabis, E.C. Schmidt, S.E. Zabierowski, P.A. Brafford, A. 457 Vultur, D. Basu, P. Gimotty, T. Vogt, M. Herlyn, Cell 141 (2010) 583-594.
  - 458[40] U. Wellner, J. Schubert, U.C. Burk, O. Schmalhofer, F. Zhu, A. Sonntag, B. 459Waldvogel, C. Vannier, D. Darling, A. zur Hausen, V.G. Brunton, J. Morton, O. 460 Sansom, J. Schuler, M.P. Stemmler, C. Herzberger, U. Hopt, T. Keck, S. Brabletz, T. 461 Brabletz, Nat. Cell Biol. 11 (2009) 1487-1495.
  - 462 [41] D. Iliopoulos, M. Lindahl-Allen, C. Polytarchou, H.A. Hirsch, P.N. Tsichlis, K. Struhl, 463 Mol. Cell 39 (2010) 761-772.
  - 464 [42] J.G. Sun, R.X. Liao, J. Qiu, J.Y. Jin, X.X. Wang, Y.Z. Duan, F.L. Chen, P. Hao, Q.C. Xie,
  - 465Z.X. Wang, D.Z. Li, Z.T. Chen, S.X. Zhang, J. Exp. Clin. Cancer Res. 29 (2010) 174.
  - 466 [43] R. Li, N. Qian, K. Tao, N. You, X. Wang, K. Dou, J. Exp. Clin. Cancer Res. 29 (2010) 467 169
  - 468M.R. Alison, S.M. Lim, L.J. Nicholson, J. Pathol. 223 (2011) 148-162. [44]
  - 469 N. Barker, R.A. Ridgway, J.H. van Es, M. van de Wetering, H. Begthel, M. van den [45] 470 Born, E. Danenberg, A.R. Clarke, O.J. Sansom, H. Clevers, Nature 457 (2009) 471 608 - 611
  - [46] S.J. Leedham, A.T. Thliveris, R.B. Halberg, M.A. Newton, N.A. Wright, Stem Cell Rev. 472 473(2005) 233-241.
  - 474 [47] L. Zhu, P. Gibson, D.S. Currle, Y. Tong, R.J. Richardson, I.T. Bayazitov, H. Poppleton, 475S. Zakharenko, D.W. Ellison, R.J. Gilbertson, Nature 457 (2009) 603-607.
  - 476 [48] A.E. Teschendorff, U. Menon, A. Gentry-Maharaj, S.J. Ramus, D.J. Weisenberger, H. Shen, M. Campan, H. Noushmehr, C.G. Bell, A.P. Maxwell, D.A. Savage, E. Mueller-477 Holzner, C. Marth, G. Kocjan, S.A. Gayther, A. Jones, S. Beck, W. Wagner, P.W. 478Laird, I.J. Jacobs, M. Widschwendter, Genome Res. 20 (2010) 440-446. 479
  - L.A. Mathews, F. Crea, W.L. Farrar, Differentiation 78 (2009) 1-17. 480 [49]
  - [50] S. Liu, G. Dontu, I.D. Mantle, S. Patel, N.S. Ahn, K.W. Jackson, P. Suri, M.S. Wicha, 481 482 Cancer Res. 66 (2006) 6063-6071.
  - 483 [51] D.J. Wong, H. Liu, T.W. Ridky, D. Cassarino, E. Segal, H.Y. Chang, Cell Stem Cell 2 484 (2008) 333-344.
  - [52] J.Y. Li, M.T. Pu, R. Hirasawa, B.Z. Li, Y.N. Huang, R. Zeng, N.H. Jing, T. Chen, E. Li, H. 485Sasaki, G.L. Xu, Mol. Cell. Biol. 27 (2007) 8748-8759. 486
- **O5**487 S. Barrand, I.S. Andersen, P. Collas, Biochem. Biophys. Res. Commun. 401 611-7.
  - 488 [54] L. Laurent, E. Wong, G. Li, T. Huynh, A. Tsirigos, C.T. Ong, H.M. Low, K.W. Kin Sung, I. Rigoutsos, J. Loring, C.L. Wei, Genome Res. 20 (2010) 320-331. 489
  - 490 [55] C. Zhang, Z. Li, Y. Cheng, F. Jia, R. Li, M. Wu, K. Li, L. Wei, Clin. Cancer Res. 13 491 (2007) 944 - 952
  - 492[56] A.P. Feinberg, R. Ohlsson, S. Henikoff, Nat. Rev. Genet. 7 (2006) 21-33.
- **O6**493 Ì57Ì P. Scaffidi, T. Misteli, Cold Spring Harb. Symp. Quant. Biol. (2010).
  - T.A. Ince, A.L. Richardson, G.W. Bell, M. Saitoh, S. Godar, A.E. Karnoub, J.D. Iglehart, 494 [58] R.A. Weinberg, Cancer Cell 12 (2007) 160-170. 495
  - 496 [59] K.D. Siegmund, P. Marjoram, Y.J. Woo, S. Tavare, D. Shibata, Proc. Natl. Acad. Sci. U. 497S. A. 106 (2009) 4828-4833.
  - 498 [60] J.L. Dembinski, S. Krauss, Clin. Exp. Metastasis 26 (2009) 611-623.
  - [61] M.Q. Gao, Y.P. Choi, S. Kang, J.H. Youn, N.H. Cho, Oncogene 29 (2010) 2672–2680. 499
  - 566

- Viale, P.G. Pelicci, P.P. Di Fiore, Cell 140 (2010) 62-73. 501
- [63] S. Bao, Q. Wu, R.E. McLendon, Y. Hao, Q. Shi, A.B. Hjelmeland, M.W. Dewhirst, D.D. 502 Bigner IN Rich Nature 444 (2006) 756–760 503
- S.P. Hong, J. Wen, S. Bang, S. Park, S.Y. Song, Int. J. Cancer 125 (2009) 2323-2331. 504 [64] [65] R. Roy, P. Willan, R. Clarke, G. Farnie, Breast Cancer Res. 12 (Suppl. 1) (2010) 05. 505
- Z. Hu, S. Negrotto, X. Gu, R. Mahfouz, K.P. Ng, Q. Ebrahem, E. Copelan, H. Singh, J.P. 506 [66]
- Maciejewski, Y. Saunthararajah, Mol. Cancer Ther. 9 (2010) 1536-1543. 507
- [67] L. Cao, J. Bombard, K. Cintron, J. Sheedy, M.L. Weetall, T.W. Davis, J. Cell. Biochem. 508 509(2011).
- [68] Y.C. Chen, C.J. Chang, H.S. Hsu, Y.W. Chen, L.K. Tai, L.M. Tseng, G.Y. Chiou, S.C. 510 Chang, S.Y. Kao, S.H. Chiou, W.L. Lo, Oral Oncol. 46 (2009) 158-165. 511
- [69] C. Liu, K. Kelnar, B. Liu, X. Chen, T. Calhoun-Davis, H. Li, L. Patrawala, H. Yan, C. 512 Jeter, S. Honorio, J.F. Wiggins, A.G. Bader, R. Fagin, D. Brown, D.G. Tang, Nat. Med. 513 17 (2011) 211-215. 514
- J. Godlewski, M.O. Nowicki, A. Bronisz, S. Williams, A. Otsuki, G. Nuovo, A. [70] 515 Raychaudhury, H.B. Newton, E.A. Chiocca, S. Lawler, Cancer Res. 68 (2008) 516 9125-9130 517
- Y. Zhu, F. Yu, Y. Jiao, J. Feng, W. Tang, H. Yao, C. Gong, J. Chen, F. Su, Y. Zhang, E. [71] 518 Song, Clin. Cancer Res. 17 (2011) 7105-7115. 519
- [72] D. Nalls, S.N. Tang, M. Rodova, R.K. Srivastava, S. Shankar, PLoS One 6 (2011) 520 e24099 521
- [73] L. Garzia, I. Andolfo, E. Cusanelli, N. Marino, G. Petrosino, D. De Martino, V. 522 Esposito, A. Galeone, L. Navas, S. Esposito, S. Gargiulo, S. Fattet, V. Donofrio, G. 523 Cinalli, A. Brunetti, L.D. Vecchio, P.A. Northcott, O. Delattre, M.D. Taylor, A. 524Iolascon, M. Zollo, PLoS One 4 (2009) e4998. 525
- [74] K. Walter, P.N. Cockerill, R. Barlow, D. Clarke, M. Hoogenkamp, G.A. Follows, S.J. 526Richards, M.J. Cullen, C. Bonifer, H. Tagoh, Oncogene 29 (2010) 2927-2937. 527528
- [75] M.J. Meyer, J.M. Fleming, M.A. Ali, M.W. Pesesky, E. Ginsburg, B.K. Vonderhaar, Breast Cancer Res. 11 (2009) R82. 529
- [76] N.S. Verkaik, J. Trapman, J.C. Romijn, T.H. Van der Kwast, G.J. Van Steenbrugge, Int. 530J. Cancer 80 (1999) 439-443. 531
- [77] Y.Y. Sanders, A. Pardo, M. Selman, G.J. Nuovo, T.O. Tollefsbol, G.P. Siegal, J.S. 532 Hagood, Am. J. Respir. Cell Mol. Biol. 39 (2008) 610-618. 533
- X.N. Gao, J. Lin, Y.H. Li, L. Gao, X.R. Wang, W. Wang, H.Y. Kang, G.T. Yan, L.L. Wang, [78] 534535 O7 L. Yu, Oncogene (2011).
- [79] K. Koinuma, R. Kaneda, M. Toyota, Y. Yamashita, S. Takada, Y.L. Choi, T. Wada, M. 536Okada, F. Konishi, H. Nagai, H. Mano, Carcinogenesis 26 (2005) 2078-2085. 537
- [80] T. Baba, P.A. Convery, N. Matsumura, R.S. Whitaker, E. Kondoh, T. Perry, Z. Huang, 538R.C. Bentley, S. Mori, S. Fujii, J.R. Marks, A. Berchuck, S.K. Murphy, Oncogene 28 539(2009) 209-218. 540
- [81] A.M. Friel, L. Zhang, M.D. Curley, V.A. Therrien, P.A. Sergent, S.E. Belden, D.R. 541Borger, G. Mohapatra, L.R. Zukerberg, R. Foster, B.R. Rueda, Reprod. Biol. 542 Endocrinol. 8 (2010) 147. 543
- [82] P. Schiapparelli, M. Enguita-German, J. Balbuena, J.A. Rey, P. Lazcoz, J.S. 544Castresana, Oncol. Rep. 24 (2010) 1355-1362. 545
- [83] A. Tong, L. Wu, Q. Lin, Q.C. Lau, X. Zhao, J. Li, P. Chen, L. Chen, H. Tang, C. Huang, 546 Y.Q. Wei, Proteomics 8 (2008) 2012-2023. 547
- [84] J.A. King, F. Tan, F. Mbeunkui, Z. Chambers, S. Cantrell, H. Chen, D. Alvarez, L.A. 548Shevde, S.F. Ofori-Acquah, Mol. Cancer 9 (2010) 266. 549
- [85] M. Kanduri, N. Cahill, H. Goransson, C. Enstrom, F. Ryan, A. Isaksson, R. Rosenquist, 550Blood 115 (2010) 296-305. 551
- [86] A. Tong, L. Gou, Q.C. Lau, B. Chen, X. Zhao, J. Li, H. Tang, L. Chen, M. Tang, C. Huang, 552Y.Q. Wei, J. Proteome Res. 8 (2009) 1037-1046. 553
- [87] G. Yu, X. Zhang, H. Wang, D. Rui, A. Yin, G. Qiu, Y. He, Oncol. Rep. 20 (2008) 5541061-1067. 555
- [88] R.R. Williams, V. Azuara, P. Perry, S. Sauer, M. Dvorkina, H. Jorgensen, J. Roix, P. 556McQueen, T. Misteli, M. Merkenschlager, A.G. Fisher, J. Cell Sci. 119 (2006) 132-140. 557
- [89] B. Jin, B. Yao, J.L. Li, C.R. Fields, A.L. Delmas, C. Liu, K.D. Robertson, Cancer Res. 69 558(2009) 7412-7421. 559
- Y. Xiang, Z. Zhu, G. Han, X. Ye, B. Xu, Z. Peng, Y. Ma, Y. Yu, H. Lin, A.P. Chen, C.D. 560[90] Chen, Proc. Natl. Acad. Sci. U. S. A. 104 (2007) 19226-19231. 561
- [91] B. Hantusch, R. Kalt, S. Krieger, C. Puri, D. Kerjaschki, BMC Mol. Biol. 8 (2007) 20. 562
- I. Ibragimova, I. Ibanez de Caceres, A.M. Hoffman, A. Potapova, E. Dulaimi, T. Al-Saleem, 563G.R. Hudes, M.F. Ochs, P. Cairns, Cancer Prev. Res. (Phila.) 3 (2010) 1084-1092. 564565

A. Vincent, J. Van Seuningen / Biochimica et Biophysica Acta xxx (2012) xxx-xxx

6

[62] S. Pece, D. Tosoni, S. Confalonieri, G. Mazzarol, M. Vecchi, S. Ronzoni, L. Bernard, G. 500