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Differential micropollutants bioaccumulation in European hake and their parasites *Anisakis* sp.

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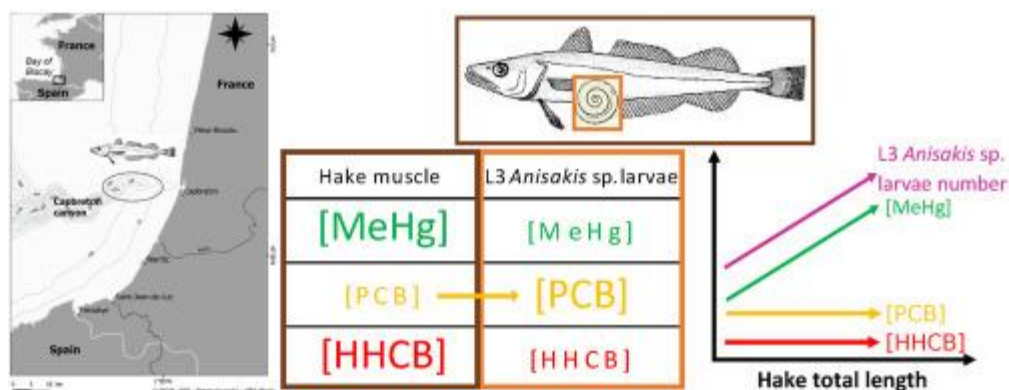
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Abstract :

Organisms are exposed to various stressors including parasites and micropollutants. Their combined effects are hard to predict. This study assessed the trophic relationship, micropollutants bioaccumulation and infection degree in a host-parasite couple. Carbon and nitrogen isotopic ratios were determined in hake *Merluccius merluccius* muscle and in its parasite *Anisakis* sp.. Concentrations of both priority (mercury species and polychlorinated biphenyls congeners) and emerging (musks and sunscreens) micropollutants were also measured for the parasite and its host, to detect potential transfer of contaminants between the two species. The results showed partial trophic interaction between the parasite and its host, in accordance with the *Anisakis* sp. life encysted in hake viscera cavity. PCB transfer between the two species may result from some lipids uptake by the parasite, while no relation occurred for the two other contaminants. Finally, a positive correlation was found between the number of *Anisakis* sp. larvae and the methylmercury contamination for hake, emphasizing the assumption that the contamination level in methylmercury can weaken immune system of the host enough to affect parasite infection degree.

Graphical abstract



Highlights

► Partial trophic interaction between L3 *Anisakis* sp. larvae and its host, hake. ► PCB transfer may result from the uptake of lipids from hake by the parasites. ► Highly MeHg level in hake can weaken immune system and increase *Anisakis* sp. number.

Keywords : Parasite-host system, Mercury, PCB, Emerging contaminants, Nematode

35 **Introduction**

36 The role of parasites in marine food webs is now recognized as important but largely overlooked
37 (Hudson et al., 2006). As they may have adverse effect on human health, too many parasites might
38 seem as organisms with no ecosystemic role that simply should be eradicated. However, they do not
39 necessarily have adverse effect on hosts nor cause economic or sanitary issues; they are important
40 and integral components of all ecosystems. Healthy ecosystems have healthy parasite communities
41 and assemblages (Marcogliese, 2005). They may be potentially used as indicators of species and
42 ecosystem diversity (Balbuena et al., 1995; Sures et al., 2017) because they reflect the presence of
43 organisms that contribute to their life cycles. They are also indicators of trophic relationships
44 between their successive hosts (Pascual et al., 1996). By strengthening predator/prey links, they
45 could be the glue that holds food webs together. In addition, since highly connected systems are
46 considered more resilient to environmental perturbations (Rooney and McCann, 2012; Sanders et al.,
47 2018; Thompson et al., 2012) and as parasitism results from biotic interactions, high level of
48 parasitism may track ecosystems with higher resistance to environmental perturbations. Due to the
49 contribution of parasites to population dynamics, Lafferty *et al.*, (2008) promote that they could be
50 integrated into food webs models. Accumulating different heavy metals in aquatic systems, they
51 could be used as bioindicators of pollution (Morsy et al., 2012; Sures, 2001). However, the
52 association between parasitism (infection degree), pollution and host health is a complex subject.
53 Cumulated effects of co-occurring parasites and pollutants can be either antagonists (contaminant
54 affects parasites and limits infestation) or synergistics (contaminant affects hosts immune system
55 and enhance infestation), precluding from easy understanding and prediction (Morrill et al., 2014).
56 By example, Sures et al. (2006) suggested an antagonistic relation between parasites and pollution in

57 eels. Infection by the swim-bladder nematode *Anguillicola crassus* increased cortisol concentrations
58 while simultaneously occurring metal and/or PCB contamination reduced plasma cortisol levels.

59 In the recent years, combining several trophic tracers was demonstrated efficient to understand
60 trophic transfers of organic matter. Carbon and nitrogen stable isotopes have been largely used in
61 trophic ecology, as predators' isotopic ratios are directly linked with isotopic ratios of their diet.
62 Carbon isotopic ratio do not vary much at each trophic level (theoretically +1‰), allowing the use of
63 this element as a tracer of organic matter source. On the contrary, nitrogen is gradually enriched
64 (theoretically +3.4‰), leading to high $\delta^{15}\text{N}$ values at high trophic levels (Layman et al., 2012).
65 Nevertheless, stable isotopes are not suited when trophic sources are not isotopically distinct. In
66 these cases, contaminants can also be used in combination (Pethybridge et al., 2018). By example, Hg
67 level was powerful to elucidate trophic pathways in the deep Mediterranean. As fish species
68 considered belonged to the same trophic pathway based on pelagic production, they exhibited
69 comparable isotopic ratios, but had different Hg levels, as they feed whether in the water column or
70 on the bottom, *ie* above or below the zone of Hg production (Cresson et al., 2014). Similarly, in the
71 Bay of Biscay, spatial variation of Hg contamination was used as a tracer of stock segregation for
72 hake, with higher Hg burdens in the southern individuals being interpreted as the occurrence of two
73 stocks in the Bay of Biscay (Chouvelon et al., 2014). Combined measurement of PCB and Hg in fish,
74 notably hake, is classical (e.g. Cresson et al., 2015; Dierking et al., 2009; Harmelin-Vivien et al., 2012),
75 as these compounds may affect different systems of the organisms (reproductive or neurologic
76 respectively) and results from different anthropic activities. As they have different chemical
77 properties, they are also powerful tracers of different exchanges of organic matter in food web.
78 Finally, investigating PCB content in different tissues allowed identifying intra-individual fluxes of
79 matter. Lipid transfers between muscle and eggs during oogenesis explained lower PCB burden in
80 females than in males (Bodiguel et al., 2009). Since isotopic relationship between host and parasites
81 is not as straight forward as between predator and prey (Thieltges et al., 2019), coupling several

82 trophic tracers can be powerful to gain better understanding of the organic matter exchange
83 between hake and *Anisakis* sp.

84 The nematode genus *Anisakis* is a cosmopolitan endoparasite. *Anisakis simplex* (s.s.) is the most
85 common species found in hake in NE Atlantic waters (Mattiucci et al., 2004). Its life cycle is
86 heteroxenous with crustaceans -mostly Euphausiids (Smith and Wootten, 1978), but also amphipods
87 or decapod larvae (Baird et al., 2014), being the first intermediate hosts. By the crustacean's
88 consumption, *Anisakis* sp. at their second larval stage (L2) are transformed into L3 in 3 to 8 days in
89 the hemocoel. The crustaceans are then ingested mainly by teleosts and cephalopods. *Anisakis* sp.
90 larvae L3 become encysted, enter hypobiosis, and do not undergo transformation. Fish and
91 cephalopods are thus considered as paratenic hosts. Cycle is completed in cetaceans' stomach, after
92 the consumption of teleosts and cephalopods, where L3 larvae are transformed in L4 larvae and then
93 mature adults. After sexual reproduction, eggs are expelled in the digestive tract of cetaceans and
94 evacuated with the faeces. Eggs develop freely in the water. Hatching occur in 20 to 27 days at 5 - 7
95 °C. The first larval stage (L1) develops inside the egg then after a second moult (L2), the larvae swims
96 freely in the water to join a crustacean (Baird et al., 2014; Buchmann and Mehrdana, 2016). The life
97 cycle of *Anisakis* is fairly well known, due to the importance of *Anisakis* sp. larvae L3 in food hygiene
98 and public health. As *Anisakis* sp. cannot complete its cycle in humans, ingestion of raw, brined,
99 marinated or undercooked fish fillet or cephalopods can lead to human health issues. Between 2010
100 and 2014, ~80 human anisakidosis (human allergy caused by *Anisakis* sp. infestation) have been
101 detected by French parasitology laboratories and hospitals, with 4 to 14 cases each year (ANSES,
102 2017; Dupouy-Camet et al., 2016). In addition, a recent study observed an increased importance of
103 *Anisakis* during the last decades with potential effects on fish and human health, and on fisheries
104 (Fiorenza et al., 2020).

105 Hake is an important demersal fishery resource. In the Bay of Biscay, this species is the first landed
106 with 42 536 tons caught in 2017 (Ifremer Fisheries Information system, 2018). Its diet varies with
107 ontogeny: juveniles mostly consume pelagic crustaceans, and adults are piscivores, with blue

108 whiting, horse mackerel, sardine and also young hake being important preys (Mahe et al., 2007; Rault
109 et al., 2017; Velasco and Olaso, 2000). Due to its diet, hake is one of fish species being an
110 intermediate host for the L3 *Anisakis* sp. larvae (Aibinu et al., 2019). In addition, its trophic position
111 drives biomagnification and high burdens in Hg (Chouvelon et al., 2014; Cossa et al., 2012) or PCB
112 (Bodiguel et al., 2009). Understanding the contamination and parasitism levels and their combined
113 effects on hake is thus of prime importance for ecological, economical and sanitary reasons.

114 As the actual magnitude of impact on the host, and a potential direct energy transfer between
115 *Anisakis* sp. and hake is unknown, this paper is aimed at investigating the relationship between
116 *Anisakis* sp. L3-larvae and hake. Combined use of stable isotopes and contaminants was employed to
117 investigate the potential transfer of organic matter and of contaminant (mercury, PCB and emergent
118 micropollutants) between host and parasite, and also the relationship between parasite infestation
119 and contamination levels.

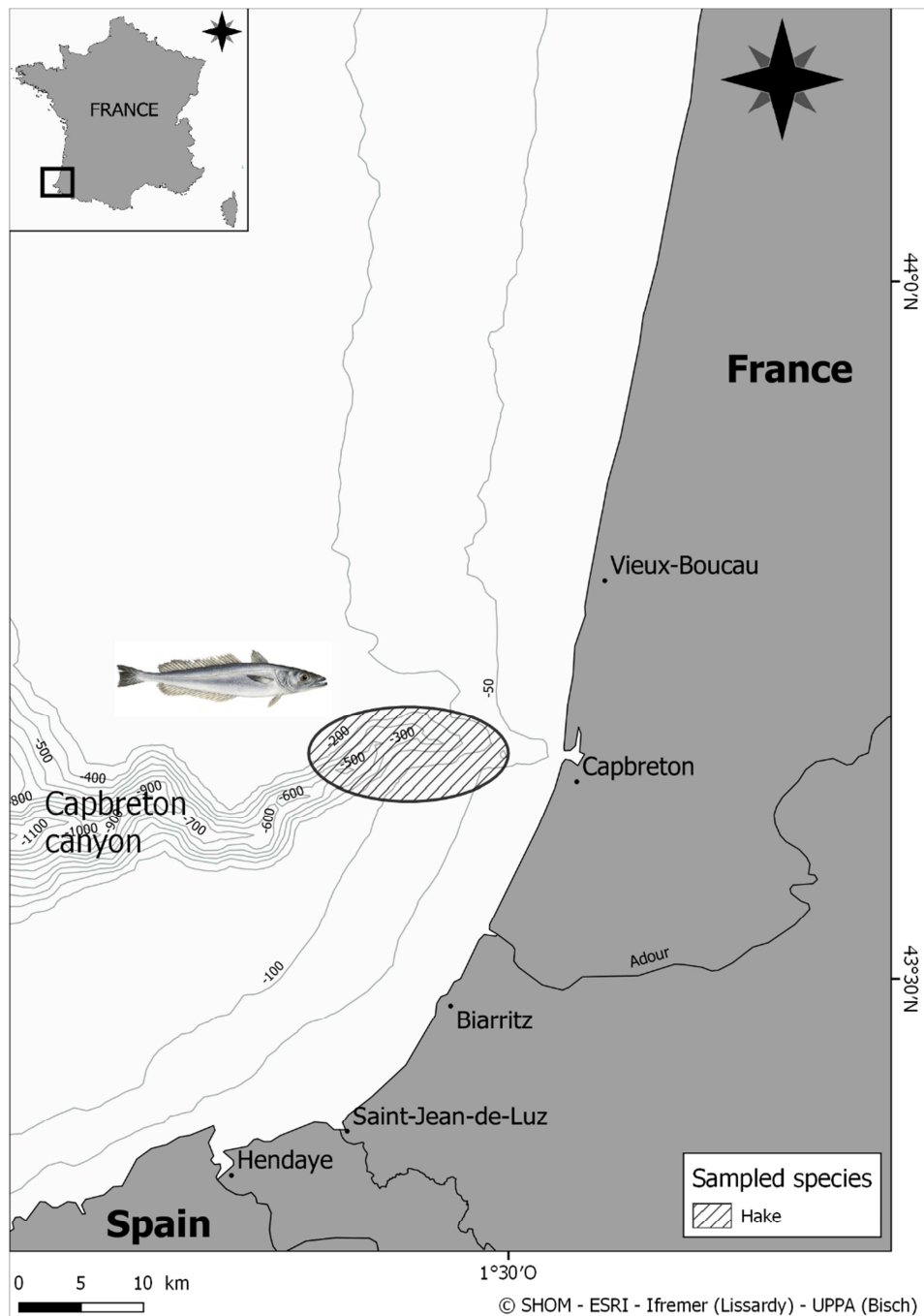
120 **Material and methods**

121 Sampling and samples preparation

122 A total of 114 hake individuals (*Merluccius merluccius*) were collected in 2017 to 2018 by fishermen
123 operating in the south of the Bay of Biscay, on the continental shelf from the Capbreton canyon, at
124 depth ranging between 150 and 200 m (Figure 1). Sampling occurred in February to June, ie before
125 and during the main reproduction period of hake. Individuals were stored on ice until dissection. In
126 the laboratory, each individual was identified, measured to the nearest mm (total length), weighted
127 to the nearest g. Sex was identified from a macroscopic examination of the gonads. On each hake,
128 muscle was collected for both stable isotope and contaminant analyses. A sample of white dorsal
129 muscle without skin and bones was taken on all individuals. A small part was used for isotopic
130 analysis and all remaining sample was used for contaminant analyses.

131 *Anisakis* sp. L3-larvae (referred as to *Anisakis* sp. hereafter) were systematically collected at the
132 opening of the visceral cavity of all hake individual, before their leaking to the organs. Parasites were

133 rinsed with distilled water, frozen at $-20\text{ }^{\circ}\text{C}$ for 48 h then decapsulated, counted and pooled to one
134 sample. The amount of matter needed to perform contaminant analyses was high ($> 2\text{ g}$ dry weight),
135 *i.e.* a weight corresponding to ~ 200 *Anisakis* sp. individuals. Analyses were thus performed in pools of
136 parasites collected in 20 hakes randomly selected among hakes with parasite's abundance higher
137 than 200 individuals. All samples (*Anisakis* sp. pools and hake muscles) were stored frozen, freeze-
138 dried and grinded.



139

140 **Figure 1: Map of hake sampling area.**

141 Carbon and nitrogen isotopic analysis

142 Isotopic analyses were performed on a small subsample of grinded hake muscle or of *Anisakis* pools
143 with a Thermo Scientific Delta V Advantage mass spectrometer coupled to a Thermo Scientific Flash
144 EA1112 elemental analyzer. Results are presented in the classical δ notation, $\delta X = \left(\frac{R_{sample}}{R_{standard}} - 1 \right) \times$
145 10^3 where X is ^{13}C or ^{15}N and R the ratio between heavy and light isotopes. Standard is Vienna-Pee
146 Dee Belemnite for $\delta^{13}\text{C}$ and atmospheric nitrogen for $\delta^{15}\text{N}$. Based on replicate measurements of
147 internal laboratory standards, the experimental precisions were 0.13‰ and 0.12‰ for $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$.
148 Carbon and nitrogen concentration were measured with the elemental analyzer and used to
149 calculate C/N ratios, classical proxy of lipid content in isotopic analyses. High C/N ratios are usually
150 interpreted as high lipid content that bias isotopic measurement. All hake C/N ratios were lower than
151 3.5, classically considered as the threshold values requiring lipid correction (Sweeting et al., 2006).
152 Previously published isotopic values were also retrieved in the literature for other fish and
153 invertebrate species living in the Bay of Biscay (Chouvelon et al., 2012).

154 Mercury species analysis

155 Methylmercury (MeHg) and inorganic mercury (IHg) were measured by capillary gas chromatography
156 (Focus GC, Thermo Electron) connected to an inductively coupled plasma mass spectrometer (ICPMS
157 X2 series, Thermo Electron). Due to its neurotoxicity, its ability to biomagnify along trophic network
158 (Bryan et al., 1979), and as it the predominant Hg form in fish (Bloom, 1992; Chouvelon et al., 2018).
159 MeHg has been used for the following analyses to represent mercury species. Methodology and
160 analytical set-up for the GC-ICP-MS for Hg speciation analysis are detailed in Monperrus et al.(2005).
161 Briefly, 100 mg of homogenized dry tissue (hake or *Anisakis* sp pool) were digested with 5 mL TMAH
162 (Tetramethylammonium hydroxide) under a microwave field and then centrifuged to remove solid
163 particles. The supernatant, stored at 4 °C, was then submitted to derivatization using sodium
164 tetraethylborate (3%). Quantification of Hg species was performed by species specific isotope
165 dilution, by adding the appropriate amount of isotopically enriched Hg standards (^{199}IHg and

166 $^{201}\text{MeHg}$, and by applying isotope pattern deconvolution for data processing (Rodríguez Martín-
167 Doimeadios et al., 2004). Total mercury (Hg) concentrations is equal to the sum of IHg and MeHg
168 concentrations. Results quality was checked by repeated analyses of blanks and of reference material
169 IAEA 407 (fish homogenate). Limits of quantification were set to 1.4 and 1.2 ng g^{-1} dw for THg and
170 MeHg, respectively.

171 Musk fragrances, UV-filters, and PCBs analyses

172 Personal care products such as musks and UV-filters are used in household products or cosmetic as
173 fragrance ingredients and in sunscreens to protect the skin against the harmful effects of ultraviolet
174 radiation, respectively. Because they are only partially eliminated by wastewater treatment plants,
175 they enter aquatic systems leading to organisms exposure (Ternes et al., 2004). Moreover, due to
176 their lipophilic nature and their great stability, these compounds can easily bioaccumulate and
177 biomagnify in the marine trophic food web (Cunha et al., 2018; Reiner and Kannan, 2011; Zhang et
178 al., 2013). On the contrary, PCB are historical contaminants in marine systems. Their toxicity drove
179 their ban in the 1990's, but due to their persistence, they are still present with high concentrations in
180 marine systems.

181 Therefore, concentrations of 9 musk fragrances (ADBI, AHMI, AHTN, ATII, HHCB, MA, MK, MM, MX),
182 5 UV-filters (3-BC, EHMC, 4-MBC, OC, OD-PABA) and 12 PCB congeners (CB 18, 28, 31, 44, 52,
183 101,118, 138, 149, 153, 180, 194) were determined in hake muscle and in *Anisakis* sp. pools by Gas
184 Chromatography-Mass Spectrometry (GC-MS). A Quick, Easy, Cheap, Effective, Rugged and Safe
185 (QuEChERS) method, combining an extraction and a clean-up step before the GC-MS analysis, has
186 been used. Methodology and analytical set-up are detailed in Miossec *et al.* (2018). Briefly,
187 micropollutants were extracted from an aliquot of 2 g of homogenized dry tissue (hake or *Anisakis* sp
188 pool) by a QuEChERS method. Then, 20 μL of the supernatant was injected and submitted to
189 derivatization using helium as carrier gas in a HP-5MS UI capillary column (30 m length \times 0.25 mm
190 diameter and 0.25 μm film thickness). Results were in good agreement with a sample spiked at 50
191 ng g^{-1} realized with all for target compounds and for each studied species (Supplementary material).

192 All concentrations were expressed relatively to dry weight. Concentrations measured for the 12 PCB
193 congeners were summed and expressed with the usual Σ PCB notation hereafter.

194

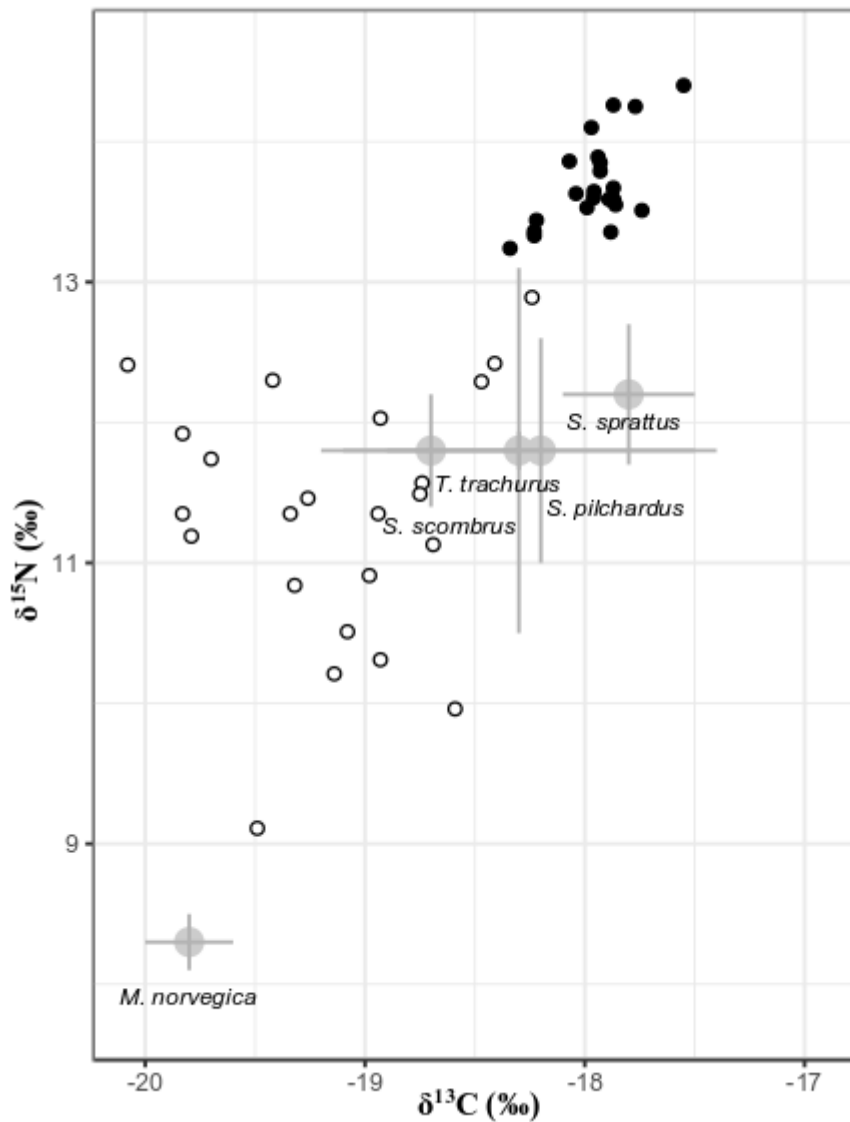
195 Data analyses

196 In order to investigate the potential contaminant transfer between the parasite and its host, the
197 relationship between contaminant concentration in hake and *Anisakis* sp. have been performed by
198 linear regressions.

199 Hake contamination level was related to its infection degree by *Anisakis* sp. in two steps. First, the
200 relationship between the number of *Anisakis* sp. found in hake and its total length and sex have been
201 investigate by an ANCOVA with total length as covariate and sex as factor, using data from all hake
202 individuals. Second, since the fish contamination can also increase with fish size, the infection degree
203 illustrated by the number of *Anisakis* sp. was fitted using a linear effect model as depending on total
204 length and micropollutant concentration of hake as continuous variable. This analysis was performed
205 on the subset of hakes the parasites of which were used for contaminant analyses. The model was
206 reduced by a bidirectional elimination procedure based on the Akaike Information Criterion (Borcard
207 et al., 2011). In order to determine active factors, significance of effects in the reduced model was
208 tested by *F* tests between nested models respecting marginality of the effects (type 2 tests; Fox and
209 Weisberg, 2011). The assumption of normality and homogeneity of variance were checked on
210 residual data. All statistical analyses were performed to the error threshold of 5%, using the “car”
211 package (Fox and Weisberg, 2011) in the statistical environment R Core Team (2018).

212 **Results**

213 Trophic ecology



214 **Figure 2: $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$ signatures for hake (●) and *Anisakis* sp. (○) from the Capbreton canyon . In order**
 215 **to complete the food web, pelagic fishes and Euphausiids are displayed in grey and are based on literature**
 216 **values (Chouvelon et al., 2012).**

217
 218
 219 Average isotopic ratios for hake were of $-17.96 \pm 0.18\text{‰}$ and $13.70 \pm 0.31\text{‰}$ for carbon and nitrogen,

220 respectively, and of $-19.13 \pm 0.51\text{‰}$ and $11.34 \pm 0.90\text{‰}$ for *Anisakis* sp., but with a large range of

221 values for both $\delta^{13}\text{C}$ (-20.8 – -18.2) and $\delta^{15}\text{N}$ (9.1 – 12.9) (Figure 2). In addition, C/N ratios were

222 higher for *Anisakis* sp. (5.4 ± 0.5) than hake (3.1 ± 0.1) (Table 1).

223

224 **Table 1: Stable isotopic signatures in carbon and nitrogen measured in hake muscles and *Anisakis* sp.**
 225 **pools (mean, standard deviation and range).**

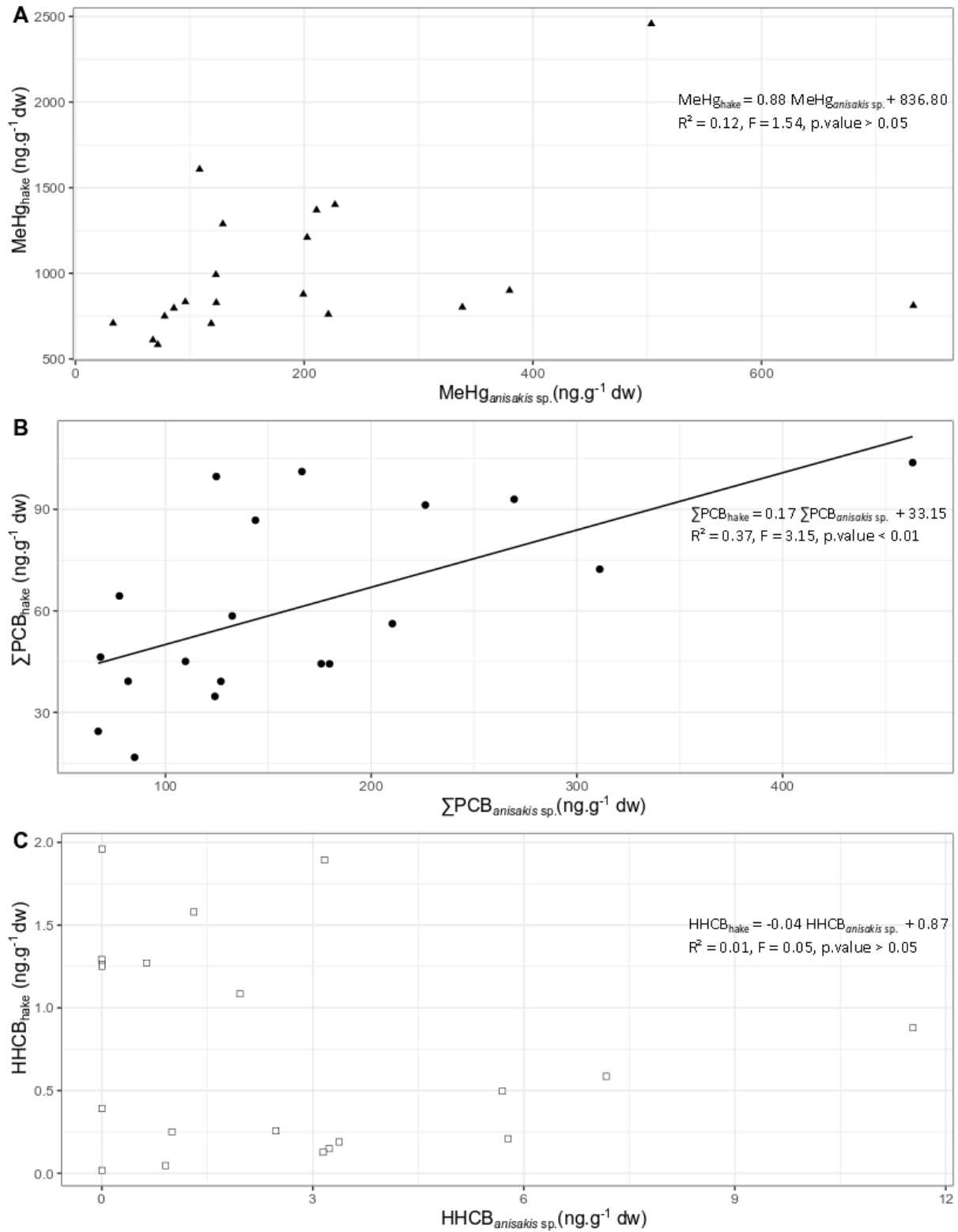
	<i>Merluccius merluccius</i>	<i>Anisakis</i> sp.
Total length (mm)	553 ± 45 (425 – 640)	–
δ¹³C (‰)	-17.9 ± 0.2 (-18.3 – -17.6)	-19.1 ± 0.5 (-20.8 – -18.2)
δ¹⁵N (‰)	13.7 ± 0.3 (13.2 – 14.4)	11.4 ± 0.9 (9.1 – 12.9)
C/N	3.1 ± 0.1 (3.1–3.2)	5.4 ± 0.5 (4.6–6.1)

226
 227 Micropollutants level according to species
 228 THg and MeHg concentrations were on average 6.4 and 4.8 times higher in hake than in *Anisakis* sp.
 229 pools, respectively (Table 2). No significant relationship between mercury concentration in hake and
 230 *Anisakis* sp. could be observed (Figure 3A). Among the 12 investigated PCB, congeners 101,118, 138,
 231 149, 153, and 180 were measured in all hake and parasite samples tested whereas PCB 194 was
 232 quantified in *Anisakis* sp. only. Unlike Hg, PCB concentrations were ~3 times higher in *Anisakis* sp.
 233 than in hake (Figure 3B). Three emergent compounds, *i.e.* one musk (ATNH) and two UV-filters
 234 (EHMC and OC) have been detected but in few individuals. Contrarily HHCB have been detected in
 235 almost all hake and *Anisakis* sp. samples tested and consequently been used to look at size and
 236 parasite effects. No relationship between HHCB concentration in host and parasite could be detected
 237 (Figure 3C).

238 **Table 2: Micropollutants concentrations measured in hake muscles and *Anisakis* sp. pools (mean,**
 239 **standard deviation and occurrence).**

	<i>Merluccius merluccius</i>	<i>Anisakis</i> sp.
Mercury species (ng g⁻¹ dw)		
THg	1543 ± 795 (20/20)	239 ± 196 (20/20)
MeHg	1000 ± 450 (20/20)	206 ± 176 (20/20)
PCB congeners (ng g⁻¹ dw)		
PCB 101	2 ± 1 (20/20)	11 ± 5 (19/20)
PCB 149	5 ± 2 (20/20)	17 ± 10 (20/20)

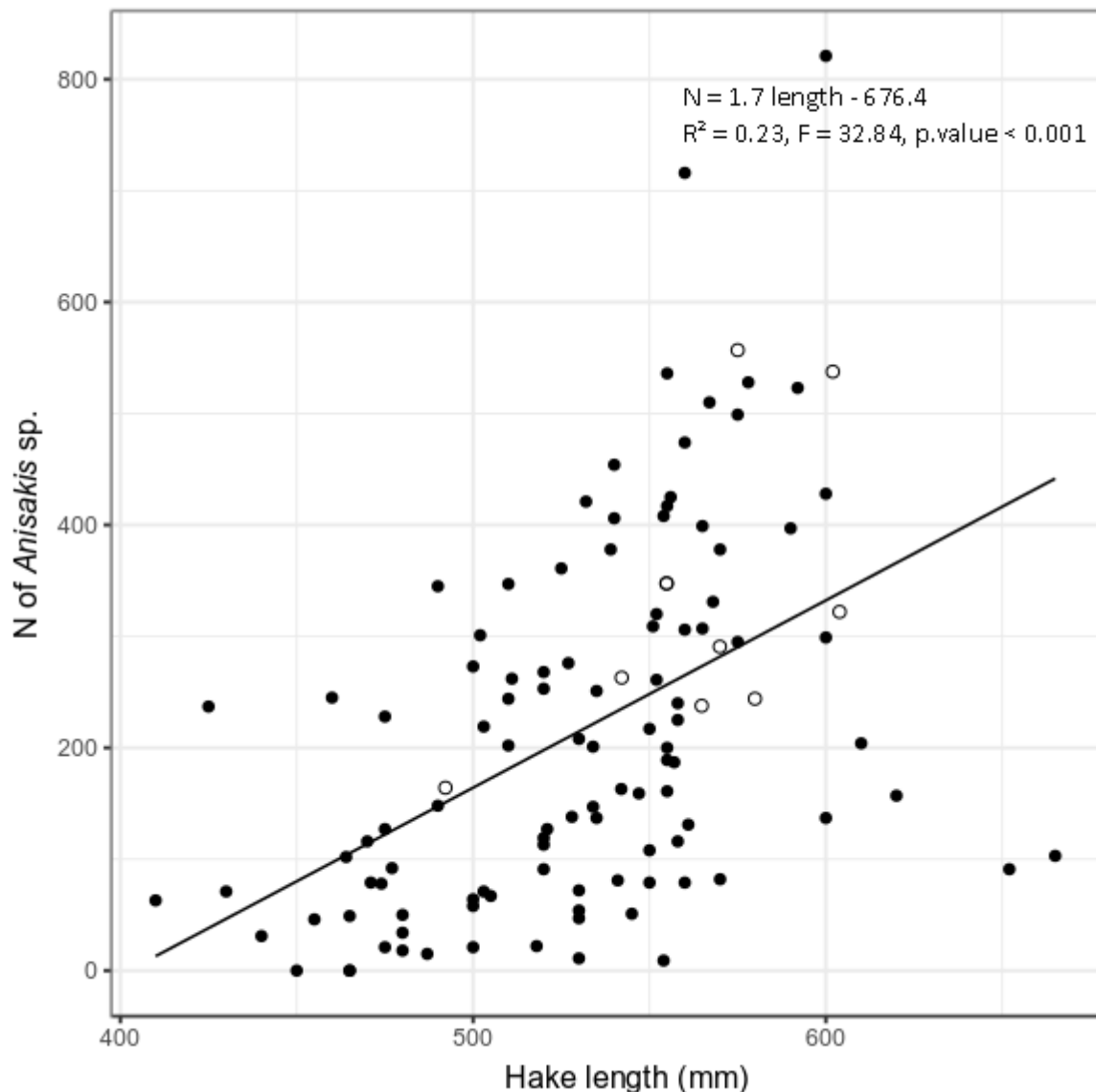
PCB 118	1 ± 1 (17/20)	8 ± 4 (20/20)
PCB 153	21 ± 9 (20/20)	51 ± 36 (20/20)
PCB 138	17 ± 8 (20/20)	46 ± 27 (20/20)
PCB 180	13 ± 7 (20/20)	32 ± 18 (20/20)
PCB 194	1 ± 1 (10/20)	<LOQ (0/20)
∑PCB	61 ± 28 (20/20)	165 ± 99 (20/20)
Musk fragrances (ng g⁻¹ dw)		
ATNH	1 ± 1 (4/20)	1 ± 2 (9/20)
HHCB	1 ± 1 (20/20)	3 ± 3 (14/20)
UV-filters (ng g⁻¹ dw)		
OC	<LOQ (0/20)	15 ± 22 (12/20)
EHMC	1 ± 1 (5/20)	<LOQ (0/20)



240

241 **Figure 3: Relationship between hake contamination and its *Anisakis sp.* pool contamination in MeHg (A),**
 242 **ΣPCB (B) and HHCB (C). Regression line is plotted only when significant (*i.e.* between hake and *Anisakis***
 243 ***sp.* ΣPCB only)**

244 Hake's contamination and degree of *Anisakis* sp. infection

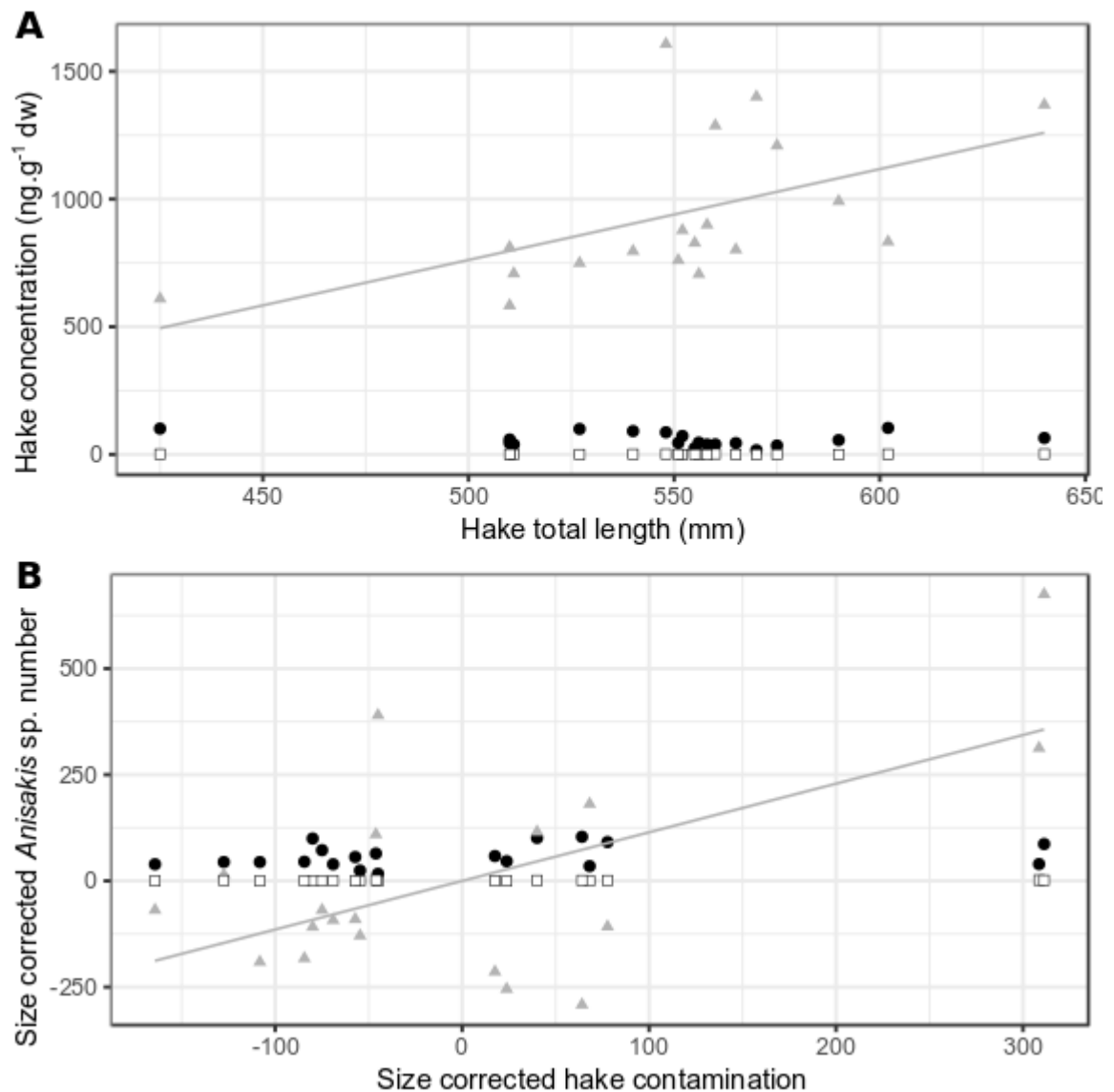


245
246
247
248

Figure 4: Relationship between the number (N) of *Anisakis* sp. (proxy of hake infection degree) and hake total length and sex (● female, ○ male).

249 A significant increase of the infection by *Anisakis* sp. with the total length of its host was found
 250 ($F = 28.8, p.\text{value} < 0.05$) (Figure 4). However, no difference according to fish sex on the infection
 251 degree has been observed ($F = 1.4, p.\text{value} > 0.05$), despite males had a higher number of parasite
 252 than females, (327 ± 134 and 209 ± 164 for males and females respectively). In addition, larger hakes
 253 were significantly more contaminated in MeHg ($F = 6.8, p.\text{value} < 0.05$) but not in PCB ($F = 0.9,$
 254 $p.\text{value} > 0.05$) and in HHCB ($F = 6.0 \cdot 10^{-4}, p.\text{value} > 0.05$) (Figure 5A). The higher the MeHg

255 concentration in hake muscle, the higher the number of *Anisakis* sp. ($F= 10.5$, p .value < 0.01) (Figure
 256 5B). Similar relationships were not observed for other micropollutants tested (Σ PCB and HHCB).



257
 258 **Figure 5: Relationship between hake contamination level (▲MeHg, ● Σ PCB, □HHCB) and its total**
 259 **length (A). Relationship between hake infection degree and its contamination level (▲MeHg, ● Σ PCB,**
 260 **□HHCB) corrected by hake total length (B).**

261 Discussion

262 Trophic relationship between hake and *Anisakis* sp.

263 Stable isotopes are widely used to identify trophic interactions and to determine trophic positions of
 264 organisms in the food webs but for classical “prey-predator” or “primary producer - grazers”
 265 interactions. Direct applications of isotopic relationships calibrated for these interactions (*i.e.*
 266 fractionation factors of $\sim 1\text{‰}$ for carbon and of $\sim 3\text{‰}$ for nitrogen) to host-parasite relationship are

267 called into question (Thieltges et al., 2019). The present study revealed that the *Anisakis* sp. were
268 consistently depleted in $\delta^{15}\text{N}$ in relation to their host, as observed in Pinnegar et al. (2001) between
269 *Merlangius merlangus* (Gadiforme) and its parasite *Hysterothylacium aduncum* (Nematode
270 Ascaridoidea). Nitrogen isotopic ratios measured for *Anisakis* sp. was very close to literature values
271 for small pelagic fish - themselves known for zooplankton predation behavior. This similarity could be
272 interpreted like a classical of predator-prey relationship between *Anisakis* sp. and Euphausiids.
273 Isotopic differences between *Anisakis* sp. and Euphausiids are of $2.36 \pm 0.87\text{‰}$ and $1.17 \pm 0.53\text{‰}$ for
274 N and C, i.e. close to theoretical trophic enrichment factors between predator and preys, while lower
275 values for *Anisakis* sp. than for hake can track the absence of trophic relationships. *Anisakis* sp.
276 isotopic ratios would reflect thus a diet mostly based on the consumption of a previous host such as
277 Euphausiids, first known intermediate host (Smith and Wootten, 1978). This hypothesis would be
278 consistent with the fact that *Anisakis* sp. is encysted when in hakes' viscera cavity, potentially limiting
279 matter exchanges. Nevertheless, large variability in *Anisakis* sp. isotopic ratios may demonstrate
280 some variability in trophic behavior. In addition, isotopic relationship between host and parasites
281 was questioned and may appear not straightforward as the relationship established between preys
282 and predators (Pinnegar et al., 2001; Thieltges et al., 2019). The ability of parasites to use selectively
283 some macromolecules from its host may explain odd factors. Parasites may be example selectively
284 use ^{14}N -enriched ammonia excreted from host tissues for amino acid synthesis (Barrett, 1981).
285 Similarly, as lipids have lower $\delta^{13}\text{C}$ values than proteins, their selective use may blur isotopic
286 relationship, notably as isotopic ratios are measured for host muscle, a lipid poor tissue. Comparing
287 tissue-specific and parasites isotopic ratios may be a pertinent research pathway to better address
288 relationship established (Pinnegar et al., 2001). The isotopic variability measured in *Anisakis* sp. may
289 also reflect that the nature of compounds uptaken may vary along with nutritional needs of the
290 parasite. Future investigation of the actual pathway and macromolecules used by parasites, as well
291 as the calibration of accurate isotopic fractionation factors are needed before being able to use
292 stable isotope as tracers of host-parasites relationships.

293

294 Difference in micropollutants concentration between the parasite and its host

295 As diet constitutes the main contamination pathway for Hg and PCB, contaminant concentration in

296 preys is an essential parameter in understanding bioaccumulation pattern in predators (Cresson et

297 al., 2014; Harmelin-Vivien et al., 2012; Kenji et al., 2020). Since stable isotopes failed to ascertain the

298 relationship between hake and *Anisakis* sp., contaminant could be powerful tools to look differently

299 at this relationship. The absence of relationship between hake and *Anisakis* sp. for MeHg and HHCB

300 concentrations can be viewed as consistent with the absence of link between these species. This

301 hypothesis could nevertheless be formally confirmed in the absence of measurement in Euphausiids.

302 Previous studies documented MeHg behavior in fish, notably driven by the link between Hg and

303 some aminoacids (Harmelin-Vivien et al., 2012; Webb et al., 2006). The absence of relationship

304 between Hg concentration in hake and parasites may track the absence of major protein exchanges

305 between the two species. This assumption was confirmed by literature: nematodes such as *Anisakis*

306 that used fish as paratenic host with an encapsulated visceral life had lowest ability to concentrate

307 metals (Dural et al., 2011; Nachev and Sures, 2016)., notably when compared with other major

308 parasitic groups, such as tapeworms and acanthocephalans that live within the host's intestine (Sures

309 et al., 1999, 1997). Regarding other micropollutant tested, the pattern was somehow blurred. As

310 these compounds are emergent, they have received less scientific interest so far. Since their behavior

311 is less documented, mechanisms explaining the absence of a clear relationship between

312 concentrations in hake and *Anisakis* sp. are hard to identify. In hake, muscle may not be the most

313 suited organ to measure these compounds, and future studies could include measurement in other

314 organs with involved in detoxication metabolism, such as liver, spleen or gallbladder.

315 Fat content is usually low in most endoparasites. As they cannot produce their own long chain fatty

316 acids, they rely on their host to take up lipids (Maule and Marks, 2006). As PCBs are lipophilic, they

317 are usually transferred along with lipids. By example, oogenesis in hake is associated with PCB

318 transfer, explaining sexual difference in PCB concentration in hake (Bodiguel et al., 2009). Therefore,

319 the PCB concentration are usually lower in parasite than its host illustrating a low transfer between
320 the two species. In this study, the PCB content was nearly 3 times higher in *Anisakis* sp than in hake.
321 Even if the exchanges of organic contaminants between host and parasites have been less
322 investigated than the exchanges of metals most studies usually consider that parasites do not
323 accumulate organic contaminants (Yen Le et al. 2014). However, the significant relationship between
324 PCB concentrations in *Anisakis* sp. and hake could track some exchanges. Once ingested by its host,
325 the parasite crosses the stomach wall and encysts in the abdominal cavity. Cuticle synthesis requires
326 high fatty acids content, notably sterols, that *Anisakis* sp. presumably take up from its host (Mika et
327 al., 2010). This lipid uptake can explain increase of PCB concentration between hake and *Anisakis* sp.
328 Such a transfer would also be consistent with the high C/N ratio observed here, consistently with
329 previous studies that demonstrated different lipid levels all along parasite life cycle (Abollo and
330 Pascual, 2001). Nevertheless, this result is contradictory with the accepted hypothesis of an arrested
331 development in paratenic host, and by the fact that *Anisakis* L3 larvae do not use hake resources,
332 usually exemplified by lower PCB concentrations in parasites than in host (Sures, 2004; Yen Le et al.
333 2014). The results obtained here combining several trophic tracers may demonstrate that limited
334 exchanges occur: blurred isotopic and Hg patterns may be consistent with no proteins exchange
335 between hake and *Anisakis*; results based on PCB on the contrary may contrarily testify some
336 exchanges of lipids and lipid-associated contaminants, even if of minor intensity. Assuming a PCB
337 biomagnification factor of ~ 5 (i.e. a 5-time increase of PCB concentrations at each trophic level, (e.g.
338 Fisk et al., 2001) and keeping in mind that *Anisakis* tissues are lipid-rich (highlighted by high C/N
339 ratios), the high PCB concentration in *Anisakis* cannot be explained only by the consumption of
340 Euphausiids. Even if exchanges of lipid occur between Euphausiids and *Anisakis*, considering other
341 exchanges is needed to explain higher PCB concentration in *Anisakis* than in hake. If exchanges only
342 occurred between *Anisakis* and Euphausiids, the parasite could be considered to be at a trophic level
343 similar to that of zooplankton-feeding fish, i.e. one trophic level below hake. Thus, PCB
344 concentrations should be somehow lower. Nevertheless, the low sample size of the present study

345 and the impossibility to include Euphausiids and zooplankton-feeding fish in this study precluded
346 from reaching a formal conclusion. Further measurements of isotopic ratios, lipids and contaminants
347 concentrations is needed to accurately depict the complex exchanges of matter between host and
348 parasites.

349

350 Relationship between fish contamination and degree of infection

351 Previous studies demonstrated that combined effects of contaminants and parasites on host are hard
352 to identify and predict. In the present study, parasites number and Hg contamination were both
353 correlated with hake length. Size effect of Hg increase is classical in hake and explained by
354 bioaccumulation and chronic exposure to contaminant (Cossa et al., 2012; Cresson et al., 2015;
355 Harmelin-Vivien et al., 2012). The same mechanism were proposed to explain higher parasite
356 number in large individuals (Morsy et al., 2012). In addition, the parasite number in hake was
357 generally higher in males than in females despite being non-significant. This may be explained by the
358 sexual dimorphism in fish size. A slower growth rate of males (de Pontual et al., 2006) implies a
359 longer expose time to parasite. Moreover, larger fish increase the capacity to have larger number,
360 size and various species of prey generating more interaction in the ecosystem and thus in turn
361 increasing the probability of exposition to infected prey (Hudson et al., 2006; Morsy et al., 2012).
362 Such an effect of sexual dimorphism and differential growth rate was already proposed to explain
363 difference in contamination in hake (Cossa et al., 2012; Harmelin-Vivien et al., 2012).

364 In the present study, fish contamination (notably in MeHg) and degree of parasitism are correlated,
365 even after removing size effect on both parameters. Such a result might be consistent with a
366 synergistic effect of contaminant and parasites. Contaminant may presumably weaken hake's
367 immune system, leading to enhanced parasitic infestation. Previous studies demonstrated that MeHg
368 at concentrations lower than levels measured in the present study (300 to 400 ng g⁻¹ in fish muscle)
369 weaken immune system of marine fish species (Guardiola et al., 2016; Ren et al., 2019). In addition
370 an increase of parasite load in highly contaminated fish was also observed in other studies (Sagerup

371 et al., 2009; Sures and Knopf, 2012). Other studies also demonstrated some effect of PCBs on
372 immune system, but at concentrations way higher than concentrations measured here (Sures and
373 Knopf, 2012).

374 **Conclusions**

375 The present data indicated that average contamination levels in parasite were lower than in its host
376 for Hg and emergent micropollutants but not for PCB and that isotopic relationship was not
377 straightforward. These results testified some complex trophic transfers, consistent with functional
378 optimization of the parasite, and its encysted nature in hake. In addition, parasites number increases
379 in highly Hg contaminated hake individual, presumably as a result of a weakened immune system of
380 the host. Present results call for extended analyses of the complex relationship established between
381 hosts and parasites, notably regarding the nature and the intensity of the organic matter fluxes. They
382 are nevertheless consistent with the current idea of the importance of parasites as important drivers
383 of marine ecosystems.

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389 **References**

- 390 Abollo, E., Pascual, S., 2001. Element concentration variability in the whaleworm *Anisakis simplex s.l.*
391 *Parasitol. Int.* 50, 115–119. [https://doi.org/10.1016/S1383-5769\(01\)00069-1](https://doi.org/10.1016/S1383-5769(01)00069-1)
- 392 Aibinu, I.E., Smooker, P.M., Lopata, A.L., 2019. *Anisakis* Nematodes in Fish and Shellfish- from
393 infection to allergies. *Int. J. Parasitol. Parasites Wildl.* 9, 384–393.
394 <https://doi.org/10.1016/j.ijppaw.2019.04.007>
- 395 ANSES, 2017. Fiche de description de danger biologique transmissible par les aliments / *Anisakis spp.*,
396 *Pseudoterranova spp.* <https://www.anses.fr/fr/system/files/BIORISK2016SA0071Fi.pdf>
- 397 Baird, F.J., Gasser, R.B., Jabbar, A., Lopata, A.L., 2014. Foodborne anisakiasis and allergy. *Mol. Cell.*
398 *Probes* 28, 167–174. <https://doi.org/10.1016/j.mcp.2014.02.003>

- 399 Balbuena, J.A., Aznar, F.J., Fernández, M., Raga, J.A., 1995. Parasites as indicators of social structure
400 and stock identity of marine mammals, in: Blix, A.S., Walløe, L., Ulltang, Ø. (Eds.),
401 Developments in Marine Biology, Whales, Seals, Fish and Man. Elsevier Science, pp. 133–139.
402 [https://doi.org/10.1016/S0163-6995\(06\)80017-X](https://doi.org/10.1016/S0163-6995(06)80017-X)
- 403 Barrett, J., 1981. Biochemistry of Parasitic Helminths., London: Macmillan. ed.
- 404 Bloom, N.S., 1992. On the Chemical Form of Mercury in Edible Fish and Marine Invertebrate Tissue.
405 Can. J. Fish. Aquat. Sci. 49, 1010–1017. <https://doi.org/10.1139/f92-113>
- 406 Bodiguel, X., Loizeau, V., Le Guellec, A.-M., Rouspard, F., Philippon, X., Mellon-Duval, C., 2009.
407 Influence of sex, maturity and reproduction on PCB and p,p'DDE concentrations and
408 repartitions in the European hake (*Merluccius merluccius*, L.) from the Gulf of Lions (N.W.
409 Mediterranean). Sci. Total Environ. 408, 304–311.
410 <https://doi.org/10.1016/j.scitotenv.2009.10.004>
- 411 Borcard, D., Gillet, F., Legendre, P., 2011. Numerical Ecology with R. Springer.
- 412 Bryan, G.W., Waldichuk, M., Pentreath, R.J., Darracott, A., 1979. Bioaccumulation of Marine
413 Pollutants. Philos. Trans. R. Soc. Lond. B. Biol. Sci. 286, 483–505.
- 414 Buchmann, K., Mehrdana, F., 2016. Effects of anisakid nematodes *Anisakis simplex* (s.l.),
415 *Pseudoterranova decipiens* (s.l.) and *Contracaecum osculatatum* (s.l.) on fish and consumer
416 health. Food Waterborne Parasitol., Foodborne Parasites in Cold Climates 4, 13–22.
417 <https://doi.org/10.1016/j.fawpar.2016.07.003>
- 418 Chouvelon, T., Caurant, F., Cherel, Y., Simon-Bouhet, B., Spitz, J., Bustamante, P., 2014. Species- and
419 size-related patterns in stable isotopes and mercury concentrations in fish help refine marine
420 ecosystem indicators and provide evidence for distinct management units for hake in the
421 Northeast Atlantic. ICES J. Mar. Sci. 71, 1073–1087. <https://doi.org/10.1093/icesjms/fst199>
- 422 Chouvelon, T., Cresson, P., Bouchoucha, M., Brach-Papa, C., Bustamante, P., Crochet, S., Marco-
423 Miralles, F., Thomas, B., Knoery, J., 2018. Oligotrophy as a major driver of mercury
424 bioaccumulation in medium-to high-trophic level consumers: A marine ecosystem-
425 comparative study. Environ. Pollut. 233, 844–854.
426 <https://doi.org/10.1016/j.envpol.2017.11.015>
- 427 Chouvelon, T., Spitz, J., Caurant, F., Mèndez-Fernandez, P., Autier, J., Lassus-Débat, A., Chappuis, A.,
428 Bustamante, P., 2012. Enhanced bioaccumulation of mercury in deep-sea fauna from the Bay
429 of Biscay (north-east Atlantic) in relation to trophic positions identified by analysis of carbon
430 and nitrogen stable isotopes. Deep Sea Res. Part Oceanogr. Res. Pap. 65, 113–124.
431 <https://doi.org/10.1016/j.dsr.2012.02.010>
- 432 Cossa, D., Harmelin-Vivien, M., Mellon-Duval, C., Loizeau, V., Averty, B., Crochet, S., Chou, L., Cadiou,
433 J.-F., 2012. Influences of Bioavailability, Trophic Position, and Growth on Methylmercury in
434 Hakes (*Merluccius merluccius*) from Northwestern Mediterranean and Northeastern Atlantic.
435 Environ. Sci. Technol. 46, 4885–4893. <https://doi.org/10.1021/es204269w>
- 436 Cresson, P., Bouchoucha, M., Morat, F., Miralles, F., Chavanon, F., Loizeau, V., Cossa, D., 2015. A
437 multitracer approach to assess the spatial contamination pattern of hake (*Merluccius*
438 *merluccius*) in the French Mediterranean. Sci. Total Environ. 532, 184–194.
439 <https://doi.org/10.1016/j.scitotenv.2015.06.020>
- 440 Cresson, P., Fabri, M.C., Bouchoucha, M., Brach Papa, C., Chavanon, F., Jadaud, A., Knoery, J.,
441 Miralles, F., Cossa, D., 2014. Mercury in organisms from the Northwestern Mediterranean
442 slope: Importance of food sources. Sci. Total Environ. 497–498, 229–238.
443 <https://doi.org/10.1016/j.scitotenv.2014.07.069>
- 444 Cunha, S.C., Trabalón, L., Jacobs, S., Castro, M., Fernandez-Tejedor, M., Granby, K., Verbeke, W.,
445 Kwadijk, C., Ferrari, F., Robbens, J., Sioen, I., Pocurull, E., Marques, A., Fernandes, J.O.,
446 Domingo, J.L., 2018. UV-filters and musk fragrances in seafood commercialized in Europe
447 Union: Occurrence, risk and exposure assessment. Environ. Res. 161, 399–408.
448 <https://doi.org/10.1016/j.envres.2017.11.015>
- 449 de Pontual, H., Groison, A.L., Piñeiro, C., Bertignac, M., 2006. Evidence of underestimation of
450 European hake growth in the Bay of Biscay, and its relationship with bias in the agreed

- 451 method of age estimation. ICES J. Mar. Sci. 63, 1674–1681.
452 <https://doi.org/10.1016/j.icesjms.2006.07.007>
- 453 Dierking, J., Wafo, E., Schembri, T., Lagadec, V., Nicolas, C., Letourneur, Y., Harmelin-Vivien, M., 2009.
454 Spatial patterns in PCBs, pesticides, mercury and cadmium in the common sole in the NW
455 Mediterranean Sea, and a novel use of contaminants as biomarkers. Mar. Pollut. Bull. 58,
456 1605–1614. <https://doi.org/10.1016/j.marpolbul.2009.07.008>
- 457 Dupouy-Camet, J., Touabet-Azouzi, N., Fréalle, E., Van Cauteren, D., Yera, H., Moneret-Vautrin, A.,
458 2016. Incidence de l'anisakidose en France. Enquête rétrospective 2010-2014. Bull Epidémiol
459 Hebd (5-6), 64–70. http://www.invs.sante.fr/beh/2016/5-6/2016_5-6_1.html
- 460 Dural, M., Genc, E., Sangun, M.K., Güner, Ö., 2011. Accumulation of some heavy metals in
461 *Hysterothylacium aduncum* (Nematoda) and its host sea bream, *Sparus aurata* (Sparidae)
462 from North-Eastern Mediterranean Sea (Iskenderun Bay). Environ. Monit. Assess. 174, 147–
463 155. <https://doi.org/10.1007/s10661-010-1445-0>
- 464 Fiorenza, E.A., Wendt, C.A., Dobkowski, K.A., King, T.L., Pappaionou, M., Rabinowitz, P., Samhoury,
465 J.F., Wood, C.L., 2020. It's a wormy world: Meta-analysis reveals several decades of change in
466 the global abundance of the parasitic nematodes *Anisakis* spp. and *Pseudoterranova* spp. in
467 marine fishes and invertebrates. Glob. Change Biol. 26, 2854–2866.
468 <https://doi.org/10.1111/gcb.15048>
- 469 Fisk, A.T., Hobson, K.A., Norstrom, R.J., 2001. Influence of Chemical and Biological Factors on Trophic
470 Transfer of Persistent Organic Pollutants in the Northwater Polynya Marine Food Web.
471 Environ. Sci. Technol. 35, 732–738. <https://doi.org/10.1021/es001459w>
- 472 Fox, J., Weisberg, S., 2011. An {R} Companion to Applied Regression.
- 473 Guardiola, F.A., Chaves-Pozo, E., Espinosa, C., Romero, D., Meseguer, J., Cuesta, A., Esteban, M.A.,
474 2016. Mercury Accumulation, Structural Damages, and Antioxidant and Immune Status
475 Changes in the Gilthead Seabream (*Sparus aurata* L.) Exposed to Methylmercury. Arch.
476 Environ. Contam. Toxicol. 70, 734–746. <https://doi.org/10.1007/s00244-016-0268-6>
- 477 Harmelin-Vivien, M., Bodiguel, X., Charmasson, S., Loizeau, V., Mellon-Duval, C., Tronczyński, J.,
478 Cossa, D., 2012. Differential biomagnification of PCB, PBDE, Hg and Radiocesium in the food
479 web of the European hake from the NW Mediterranean. Mar. Pollut. Bull. 64, 974–983.
480 <https://doi.org/10.1016/j.marpolbul.2012.02.014>
- 481 Hudson, P.J., Dobson, A.P., Lafferty, K.D., 2006. Is a healthy ecosystem one that is rich in parasites?
482 Trends Ecol. Evol. 21, 381–385. <https://doi.org/10.1016/j.tree.2006.04.007>
- 483 Ifremer Fisheries Information system, 2018. Activité des navires de pêche 2017 : façade Atlantique.
484 <https://archimer.ifremer.fr/doc/00490/60174/>
- 485 Kenji, Y., Keisuke, M., Gen, K., Shigeaki, K., Yasuhisa, H., Akito, M., Megumi, Y., 2020. Food sources
486 are more important than biomagnification on mercury bioaccumulation in marine fishes.
487 Environ. Pollut. 113982. <https://doi.org/10.1016/j.envpol.2020.113982>
- 488 Lafferty, K.D., Allesina, S., Arim, M., Briggs, C.J., De Leo, G., Dobson, A.P., Dunne, J.A., Johnson, P.T.J.,
489 Kuris, A.M., Marcogliese, D.J., Martinez, N.D., Memmott, J., Marquet, P.A., McLaughlin, J.P.,
490 Mordecai, E.A., Pascual, M., Poulin, R., Thielges, D.W., 2008. Parasites in food webs: the
491 ultimate missing links. Ecol. Lett. 11, 533–546. <https://doi.org/10.1111/j.1461-0248.2008.01174.x>
- 493 Layman, C.A., Araujo, M.S., Boucek, R., Hammerschlag-Peyer, C.M., Harrison, E., Jud, Z.R., Matich, P.,
494 Rosenblatt, A.E., Vaudo, J.J., Yeager, L.A., Post, D.M., Bearhop, S., 2012. Applying stable
495 isotopes to examine food-web structure: an overview of analytical tools. Biol. Rev. Camb.
496 Philos. Soc. 87, 545–562. <https://doi.org/10.1111/j.1469-185X.2011.00208.x>
- 497 Mahe, K., Amara, R., Bryckaert, T., Kacher, M., Brylinski, J.M., 2007. Ontogenetic and spatial variation
498 in the diet of hake (*Merluccius merluccius*) in the Bay of Biscay and the Celtic Sea. ICES J. Mar.
499 Sci. 64, 1210–1219. <https://doi.org/10.1093/icesjms/fsm100>
- 500 Marcogliese, D.J., 2005. Parasites of the superorganism: Are they indicators of ecosystem health? Int.
501 J. Parasitol., Annual Scientific Meeting of the Australian Society for Parasitology, Freemantle
502 Esplanade Hotel, 2004 Highlights 35, 705–716. <https://doi.org/10.1016/j.ijpara.2005.01.015>

- 503 Mattiucci, S., Abaunza, P., Ramadori, L., Nascetti, G., 2004. Genetic identification of *Anisakis* larvae in
504 European hake from Atlantic and Mediterranean waters for stock recognition. *J. Fish Biol.* 65,
505 495–510. <https://doi.org/10.1111/j.0022-1112.2004.00465.x>
- 506 Maule, A.G., Marks, N.J., 2006. *Parasitic FlatWorms: Molecular Biology, Biochemistry, Immunology*
507 *and Physiology*. URL <https://www.cabi.org/bookshop/book/9780851990279/> (accessed
508 1.27.20).
- 509 Mika, A., Gołebowski, M., Szafranek, J., Rokicki, J., Stepnowski, P., 2010. Identification of lipids in the
510 cuticle of the parasitic nematode *Anisakis simplex* and the somatic tissues of the Atlantic cod
511 *Gadus morhua*. *Exp. Parasitol.* 124, 334–340. <https://doi.org/10.1016/j.exppara.2009.11.013>
- 512 Miossec, C., Lancelleur, L., Monperrus, M., 2018. Adaptation and validation of QuEChERS method for
513 the simultaneous analysis of priority and emerging pollutants in sediments by gas
514 chromatography—mass spectrometry. *Int. J. Environ. Anal. Chem.* 98, 695–708.
515 <https://doi.org/10.1080/03067319.2018.1496245>
- 516 Monperrus, M., Tessier, E., Veschambre, S., Amouroux, D., Donard, O., 2005. Simultaneous
517 speciation of mercury and butyltin compounds in natural waters and snow by propylation
518 and species-specific isotope dilution mass spectrometry analysis. *Anal. Bioanal. Chem.* 381,
519 854–862. <https://doi.org/10.1007/s00216-004-2973-7>
- 520 Morrill, A., Provencher, J.F., Forbes, M.R., 2014. Testing for dual impacts of contaminants and
521 parasites on hosts: the importance of skew. *Environ. Rev.* 22, 445–456.
522 <https://doi.org/10.1139/er-2014-0026>
- 523 Morsy, K., Bashtar, A.-R., Abdel-Ghaffar, F., Mehlhorn, H., Quraishy, S.A., El-Mahdi, M., Al-Ghamdi, A.,
524 Mostafa, N., 2012. First record of anisakid juveniles (Nematoda) in the European seabass
525 *Dicentrarchus labrax* (family: Moronidae), and their role as bio-indicators of heavy metal
526 pollution. *Parasitol. Res.* 110, 1131–1138. <https://doi.org/10.1007/s00436-011-2600-4>
- 527 Nachev, M., Sures, B., 2016. Environmental parasitology: Parasites as accumulation bioindicators in
528 the marine environment. *J. Sea Res., Ecology and Evolution of Marine Parasites and Diseases*
529 113, 45–50. <https://doi.org/10.1016/j.seares.2015.06.005>
- 530 Pascual, S., González, Á.F., Arias, C., Guerra, Á., 1996. Biotic relationships of *Illex coindetii* and
531 *Todaropsis eblanae* (Cephalopoda, Ommastrephidae) in the Northeast Atlantic: Evidence
532 from parasites. <https://doi.org/10.1080/00364827.1996.10413624>
- 533 Pethybridge, H.R., Choy, C.A., Polovina, J.J., Fulton, E.A., 2018. Improving Marine Ecosystem Models
534 with Biochemical Tracers. *Annu. Rev. Mar. Sci.* 10, 199–228.
535 <https://doi.org/10.1146/annurev-marine-121916-063256>
- 536 Pinnegar, J.K., Campbell, N., Polunin, N.V.C., 2001. REGULAR PAPERS Unusual stable isotope
537 fractionation patterns observed for fish host—parasite trophic relationships. *J. Fish Biol.* 59,
538 494–503. <https://doi.org/10.1111/j.1095-8649.2001.tb02355.x>
- 539 R Core Team, 2018. R: A language and environment for statistical computing. R Foundation for
540 Statistical Computing, Vienna, Austria. URL <https://www.R-project.org/>.
- 541 Rault, J., Le Bris, H., Robert, M., Pawlowski, L., Denamiel, M., Kopp, D., 2017. Diets and trophic niches
542 of the main commercial fish species from the Celtic Sea. *J. Fish Biol.* 91, 1449–1474.
543 <https://doi.org/10.1111/jfb.13470>
- 544 Reiner, J.L., Kannan, K., 2011. Polycyclic Musks in Water, Sediment, and Fishes from the Upper
545 Hudson River, New York, USA. *Water. Air. Soil Pollut.* 214, 335–342.
546 <https://doi.org/10.1007/s11270-010-0427-8>
- 547 Ren, Z., Liu, J., Huang, W., Cao, L., Cui, W., Dou, S., 2019. Antioxidant defenses and immune
548 responses of flounder *Paralichthys olivaceus* larvae under methylmercury exposure. *Comp.*
549 *Biochem. Physiol. Part C Toxicol. Pharmacol.* 225, 108589.
550 <https://doi.org/10.1016/j.cbpc.2019.108589>
- 551 Rodríguez Martín-Doimeadios, R.C., Tessier, E., Amouroux, D., Guyoneaud, R., Duran, R., Caumette,
552 P., Donard, O.F.X., 2004. Mercury methylation/demethylation and volatilization pathways in
553 estuarine sediment slurries using species-specific enriched stable isotopes. *Mar. Chem.,*

- 554 Special Issue in honor of Dr. William F. Fitzgerald 90, 107–123.
555 <https://doi.org/10.1016/j.marchem.2004.02.022>
- 556 Rooney, N., McCann, K.S., 2012. Integrating food web diversity, structure and stability. *Trends Ecol.*
557 *Evol.* 27, 40–46. <https://doi.org/10.1016/j.tree.2011.09.001>
- 558 Sagerup, K., Savinov, V., Savinova, T., Kuklin, V., Muir, D.C.G., Gabrielsen, G.W., 2009. Persistent
559 organic pollutants, heavy metals and parasites in the glaucous gull (*Larus hyperboreus*) on
560 Spitsbergen. *Environ. Pollut.* 157, 2282–2290. <https://doi.org/10.1016/j.envpol.2009.03.031>
- 561 Sanders, D., Thébault, E., Kehoe, R., Veen, F.J.F. van, 2018. Trophic redundancy reduces vulnerability
562 to extinction cascades. *Proc. Natl. Acad. Sci.* 115, 2419–2424.
563 <https://doi.org/10.1073/pnas.1716825115>
- 564 Smith, J.W., Wootten, R., 1978. Anisakis and Anisakiasis, in: Lumsden, W.H.R., Muller, R., Baker, J.R.
565 (Eds.), *Advances in Parasitology*. Academic Press, pp. 93–163.
566 [https://doi.org/10.1016/S0065-308X\(08\)60573-4](https://doi.org/10.1016/S0065-308X(08)60573-4)
- 567 Sures, B., 2006. How parasitism and pollution affect the physiological homeostasis of aquatic hosts. *J.*
568 *Helminthol.* 80, 151–157. <https://doi.org/10.1079/joh2006346>
- 569 Sures, B., 2004. Environmental parasitology: relevancy of parasites in monitoring environmental
570 pollution. *Trends Parasitol.* 20, 170–177. <https://doi.org/10.1016/j.pt.2004.01.014>
- 571 Sures, B., 2001. The use of fish parasites as bioindicators of heavy metals in aquatic ecosystems: a
572 review. *Aquat. Ecol.* 35, 245–255. <https://doi.org/10.1023/A:1011422310314>
- 573 Sures, B., Knopf, K., 2012. Individual and combined effects of cadmium and 3,3',4,4',5-
574 pentachlorobiphenyl (PCB 126) on the humoral immune response in European eel (*Anguilla*
575 *anguilla*) experimentally infected with larvae of *Anguillicola crassus* (Nematoda). [WWW
576 Document]. URL <https://www.ncbi.nlm.nih.gov/pubmed/15151150> (accessed 11.20.19).
- 577 Sures, B., Nachev, M., Pahl, M., Grabner, D., Selbach, C., 2017. Parasites as drivers of key processes in
578 aquatic ecosystems: Facts and future directions. *Exp. Parasitol., Proceedings of the 27th*
579 *Meeting of the German Society for Parasitology 2016* 180, 141–147.
580 <https://doi.org/10.1016/j.exppara.2017.03.011>
- 581 Sures, B., Siddall, R., Taraschewski, H., Sures, B., Siddall, R., Taraschewski, H., 1999. Parasites as
582 Accumulation Indicators of Heavy Metal Pollution. *Parasitol. Today* 15, 16–21.
583 [https://doi.org/10.1016/S0169-4758\(98\)01358-1](https://doi.org/10.1016/S0169-4758(98)01358-1)
- 584 Sures, B., Taraschewski, H., Siddall, R., 1997. Heavy metal concentrations in adult acanthocephalans
585 and cestodes compared to their fish hosts and to established free-living bioindicators.
586 *Parassitologia* 39, 213–218.
- 587 Sweeting, C.J., Polunin, N.V.C., Jennings, S., 2006. Effects of chemical lipid extraction and arithmetic
588 lipid correction on stable isotope ratios of fish tissues. *Rapid Commun. Mass Spectrom.* 20,
589 595–601. <https://doi.org/10.1002/rcm.2347>
- 590 Ternes, T.A., Joss, A., Siegrist, H., 2004. Peer Reviewed: Scrutinizing Pharmaceuticals and Personal
591 Care Products in Wastewater Treatment. *Environ. Sci. Technol.* 38, 392A–399A.
592 <https://doi.org/10.1021/es040639t>
- 593 Thieltges, D.W., Goedknegt, M.A., O'Dwyer, K., Senior, A.M., Kamiya, T., 2019. Parasites and stable
594 isotopes: a comparative analysis of isotopic discrimination in parasitic trophic interactions.
595 *Oikos* 128, 1329–1339. <https://doi.org/10.1111/oik.06086>
- 596 Thompson, R.M., Brose, U., Dunne, J.A., Hall, R.O., Hladysz, S., Kitching, R.L., Martinez, N.D., Rantala,
597 H., Romanuk, T.N., Stouffer, D.B., Tylianakis, J.M., 2012. Food webs: reconciling the structure
598 and function of biodiversity. *Trends Ecol. Evol.* 27, 689–697.
599 <https://doi.org/10.1016/j.tree.2012.08.005>
- 600 Velasco, F., Olaso, I., 2000. Hake food consumption in the southern Bay of Biscay estimated from a
601 gastric evacuation model. *ICES*. [https://doi.org/ICES CM2000/Q:11](https://doi.org/ICES%202000/Q:11).
- 602 Webb, M.A.H., Feist, G.W., Fitzpatrick, M.S., Foster, E.P., Schreck, C.B., Plumlee, M., Wong, C.,
603 Gundersen, D.T., 2006. Mercury Concentrations in Gonad, Liver, and Muscle of White
604 Sturgeon *Acipenser transmontanus* in the Lower Columbia River. *Arch. Environ. Contam.*
605 *Toxicol.* 50, 443–451. <https://doi.org/10.1007/s00244-004-0159-0>

606 Zhang, X., Xu, Q., Man, S., Zeng, X., Yu, Y., Pang, Y., Sheng, G., Fu, J., 2013. Tissue concentrations,
 607 bioaccumulation, and biomagnification of synthetic musks in freshwater fish from Taihu
 608 Lake, China. *Environ. Sci. Pollut. Res. Int.* 20, 311–322. [https://doi.org/10.1007/s11356-012-](https://doi.org/10.1007/s11356-012-1095-6)
 609 1095-6
 610

611 Supplementary material

612 Analytical performances of the method used including limits of quantification (LOQ), recoveries (%) and
 613 precisions.

	LOQ	<i>Merluccius merluccius</i>		<i>Anisakis</i> sp.	
		Recovery values (%)	Precision (%)	Recovery values (%)	Precision (%)
PCB congeners (ng g⁻¹ dw)					
PCB 18	0.1	46.2	0.2	70.7	13.0
PCB 28	0.2	68.3	13.1	82.7	10.6
PCB 31	0.2	68.3	13.1	82.7	10.6
PCB 44	0.1	57.5	15.2	81.5	7.8
PCB 52	0.1	65.9	20.0	84.1	8.1
PCB 101	0.1	62.8	21.8	84.1	4.4
PCB 118	0.1	69.3	17.1	84.7	2.9
PCB 138	0.2	51.7	6.2	68.8	21.6
PCB 149	0.1	59.1	16.5	79.8	6.1
PCB 153	0.2	59.6	5.3	77.1	24.4
PCB 180	0.3	53.9	7.6	79.6	8.9
PCB 194	0.5	61.4	11.3	67.4	17.6
Musk fragrances (ng g⁻¹ dw)					
ADBI	0.1	102.1	2.9	87.4	3.3
AHMI	0.1	98.2	2.6	83.2	20.4
AHTN	0.1	110.0	12.8	105.9	32.2
ATII	0.1	108.8	11.5	102.1	15.1
HHCB	0.1	111.6	14.7	100.8	7.5
MA	0.9	102.9	4.0	75.0	8.9

MK	0.2	104.0	5.5	92.1	9.1
MM	0.1	110.1	13.0	91.7	1.8
MX	0.1	108.0	10.5	94.1	12.1
UV-filters (ng g⁻¹ dw)					
3-BC	1.4	127.0	30.1	108.4	11.2
EHMC	0.2	111.7	14.8	123.8	11.3
4-MBC	0.7	108.3	10.8	102.1	20.6
OC	1.9	104.0	5.4	93.2	1.2
OD-PABA	0.1	112.8	16.0	67.0	29.3

Highlights:

- Partial trophic interaction between L3 *Anisakis* sp. larvae and its host, hake.
- PCB transfer may result from the uptake of lipids from hake by the parasites.
- Highly MeHg level in hake can weaken immune system and increase *Anisakis* sp. number.

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Declaration of interests

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

The authors declare the following financial interests/personal relationships which may be considered as potential competing interests:

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