



HAL
open science

Effects of marine Harmful Algal Blooms on bivalve cellular immunity and infectious diseases: a review

Malwenn Lassudrie, Helene Hegaret, Gary H. Wikfors, Patricia Mirella da Silva

► To cite this version:

Malwenn Lassudrie, Helene Hegaret, Gary H. Wikfors, Patricia Mirella da Silva. Effects of marine Harmful Algal Blooms on bivalve cellular immunity and infectious diseases: a review. *Developmental and Comparative Immunology*, 2020, 108, pp.103660. 10.1016/j.dci.2020.103660 . hal-02880026

HAL Id: hal-02880026

<https://hal.science/hal-02880026v1>

Submitted on 24 Jun 2020

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

Effects of marine Harmful Algal Blooms on bivalve cellular immunity and infectious diseases: a review

Malwenn Lassudrie¹, H el ene H egaret², Gary H. Wikfors³, Patricia Mirella da Silva⁴

¹ Ifremer, LER-BO, F- 29900 Concarneau, France.

² CNRS, Univ Brest, IRD, Ifremer, LEMAR, F-29280, Plouzan e, France.

³ NOAA Fisheries Service, Northeast Fisheries Science Center, Milford, CT 0640 USA.

⁴ Laboratory of Immunology and Pathology of Invertebrates, Department of Molecular Biology, Federal University of Para iba (UFPB), Para iba, Brazil.

Abstract

Bivalves were long thought to be “symptomless carriers” of marine microalgal toxins to human seafood consumers. In the past three decades, science has come to recognize that harmful algae and their toxins can be harmful to grazers, including bivalves. Indeed, studies have shown conclusively that some microalgal toxins function as active grazing deterrents. When responding to marine Harmful Algal Bloom (HAB) events, bivalves can reject toxic cells to minimize toxin and bioactive extracellular compound (BEC) exposure, or ingest and digest cells, incorporating nutritional components and toxins.

Several studies have reported modulation of bivalve hemocyte variables in response to HAB exposure. Hemocytes are specialized cells involved in many functions in bivalves, particularly in immunological defense mechanisms. Hemocytes protect tissues by engulfing or encapsulating living pathogens and repair tissue damage caused by injury, poisoning, and infections through inflammatory processes. The effects of HAB exposure observed on bivalve cellular immune variables have raised the question of possible effects on susceptibility to infectious disease. As science has described a previously unrecognized diversity in microalgal bioactive substances, and also found a growing list of infectious diseases in bivalves, episodic reports of interactions between harmful algae and disease in bivalves have been published. Only recently, studies directed to understand the physiological and metabolic bases of these interactions have been undertaken. This review compiles evidence from studies of harmful algal

effects upon bivalve shellfish that establishes a framework for recent efforts to understand how harmful algae can alter infectious disease, and particularly the fundamental role of cellular immunity, in modulating these interactions.

Experimental studies reviewed here indicate that HABs can modulate bivalve-pathogen interactions in various ways, either by increasing bivalve susceptibility to disease or conversely by lessening infection proliferation or transmission. Alteration of immune defense and global physiological distress caused by HAB exposure have been the most frequent reasons identified for these effects on disease. Only few studies, however, have addressed these effects so far and a general pattern cannot be established. Other mechanisms are likely involved but are understudied thus far and will need more attention in the future. In particular, the inhibition of bivalve filtration by HABs and direct interaction between HABs and infectious agents in the seawater likely interfere with pathogen transmission. The study of these interactions in the field and at the population level also are needed to establish the ecological and economical significance of the effects of HABs upon bivalve diseases. A more thorough understanding of these interactions will assist in development of more effective management of bivalve shellfisheries and aquaculture in oceans subjected to increasing HAB and disease pressures.

Keywords: Bivalve; Harmful Algal Blooms (HABs); Pathogen; Hemocyte; Disease.

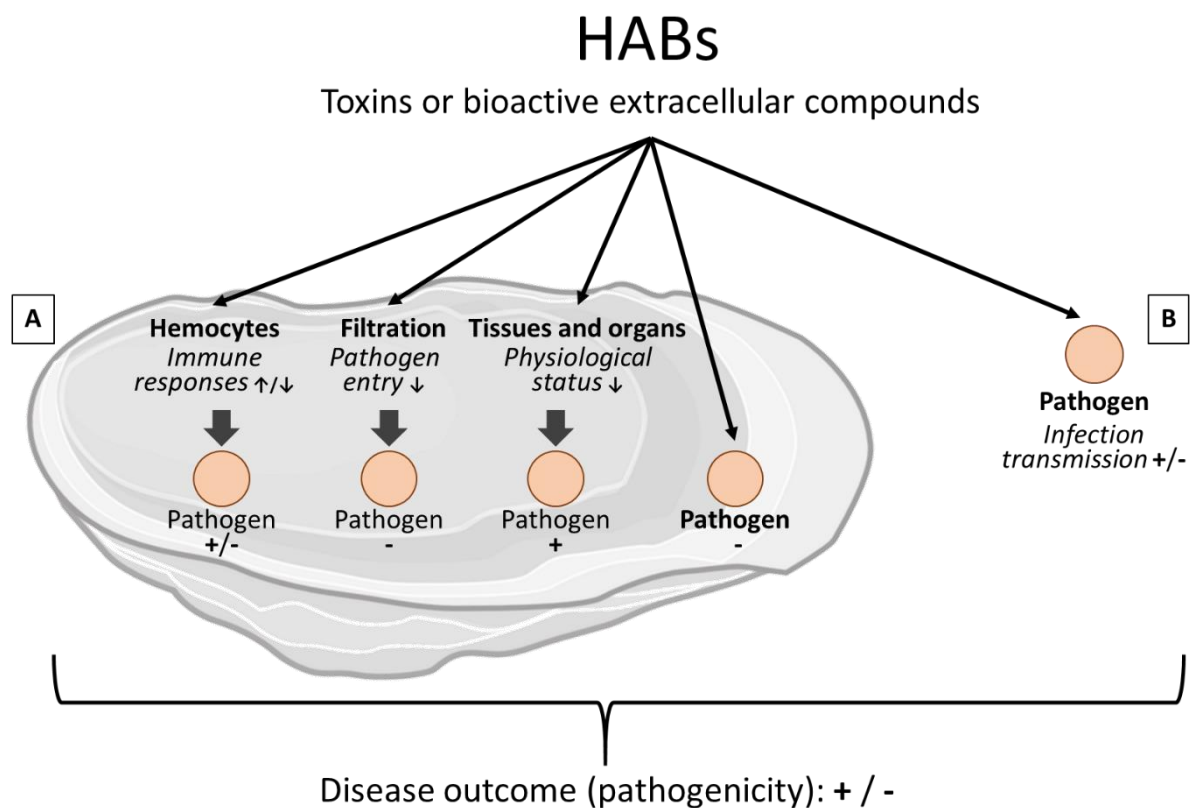
List of abbreviations

ASP: amnesic shellfish poisoning
AZA: azaspiracids
AZP: azaspiracid poisoning
BEC: bioactive extracellular compounds
BRD: brown ring disease
CFP: ciguatera fish poisoning
CTX: ciguatoxin
DA: domoic acid
DHC: differential hemocyte count
DMSP: dimethylsulfoniopropionate
DSP: diarrhetic shellfish poisoning
DTX: dinophysistoxin
HAB: harmful algal bloom
LC: lytic compounds
NSP: neurotoxic shellfish poisoning
OA: okadaic acid
OIE: World Organisation for Animal Health (historically: Office International des Epizooties)
OsHV-1: ostreid herpesvirus-1
PAMP: pathogen-associated molecular pattern
PbTx: brevetoxin
PCR: polymerase chain reaction
PLTX: palytoxin
PnTX: pinnatoxins
POMS: Pacific oyster mortality syndrome
PRP: receptor proteins
PSP: paralytic shellfish poisoning
PST: paralytic shellfish toxin
PTX: pectenotoxin
QPX disease: Quahog parasite unknown disease
QX disease: Queensland unknown disease
RLO: Rickettsia-like organisms
RNS: reactive nitrogen species
ROS: reactive oxygen species
SPX: spirolides
STX: saxitoxin
THC: total hemocyte count
YTX: yessotoxin

Highlights

- Review of the main studies on effects of marine harmful algae on bivalve immune cells (hemocytes) and on infectious disease
- Harmful algae can modulate host-pathogen interactions in bivalves on various way (increase or decrease the infection)
- Some experimental studies report the involvement of cellular immunity and global physiological weakness in the modulations
- Other mechanisms possibly involved: HAB effects on bivalve filtration; direct interaction of HABs with pathogens
- Future research recommendation: field studies, at population level, to establish the ecological and economical significance

Graphical Abstract



1 Introduction

Bivalve fisheries and aquaculture contribute to the economy of coastal countries worldwide. Mortality from infectious disease is the main threat to this industry, and mortality can be a consequence of interacting biotic and abiotic factors. Some evidence suggests the involvement of marine Harmful Algal Blooms (HABs) in bivalve disease outbreaks, particularly through effects upon bivalve immunity and defense against disease. This contribution reviews the published literature reporting effects of marine HABs upon cellular immunity and how they relate to bivalve infectious diseases, most of which have been studied under experimental conditions. Other mechanisms possibly involved in the effects of HABs on bivalve disease also are explored.

1.1 Bivalve diseases

Bivalve populations are affected by epizootics that decimate or weaken exploited stocks and therefore limit harvests (Barbosa Solomieu et al., 2015). Commercial trade between different regions of the world contributes to exotic species introduction and, as a consequence, to the spread of infectious diseases (Andrews, 1980; Renault, 1996). Bivalve diseases are caused by a variety of infectious agents (Zannella et al., 2017), mainly viruses (Arzul et al., 2017), bacteria (Travers et al., 2015), or protozoa (Robledo et al., 2014). A brief description of the most pathogenic agents for bivalves, mainly associated with OIE notifiable diseases, as well as occurrences and effects, follows.

In France, between 1967 and 1973, "gill disease" associated with an iridovirus, completely decimated populations of Portuguese oysters *Crassostrea angulata* (Comps, 1988), introduced from Portugal and Spain in 1868 (Marteil, 1976). This loss motivated the introduction in 1971 of the Pacific oyster *C. gigas* from Canada and Japan (Grizel and Héral, 1991); Pacific oyster resistance to this virus helped revive the French oyster industry. Nevertheless, in the early 1990s, Pacific oysters *C. gigas* were affected by "summer mortality" events associated with a complex interaction between temperature, genetics, and physiological status (reproductive period) of the oyster, as well as the presence of opportunistic infectious agents, such as bacteria *Vibrio aesturianus* and *V. splendidus*, and the ostreid herpesvirus-1 (OsHV-1) (Samain, 2011).

In 2008, this latter virus emerged as a more powerful variant (OsHV-1 μ Var, Segarra et al., 2010), causing Pacific Oyster Mortality Syndrome (POMS), which strongly affected young *C. gigas* in France. POMS spread afterwards into many European countries as well as other countries in Oceania (New Zealand and Australia) (Barbosa Solomieu et al., 2015; Gittenberger et al., 2016; Mortensen et al., 2016). The OsHV-1 recently has been detected by PCR in *C. gigas* cultured in southern Brazil, however without any associated mortality (Mello et al., 2018), suggesting that environmental conditions can play an important role in the development of outbreaks. In Europe and Australia, Pacific oyster mortalities associated with OsHV-1 occur when temperature exceeds 16-18°C (de Kantzow et al., 2016; Panel and Health, 2015; Paul-Pont et al., 2014). Disbalance of environmental conditions combined with temperature stress also may contribute to the development of POMS. For example, in Australia, a POMS outbreak was correlated with heavy rainfall, presence of a pathogenic *Vibrio* sp., and a toxic phytoplankton bloom (Jenkins et al., 2013). In recent years, concerted efforts were made to understand the development of POMS, leading to recognition that OsHV1- μ Var could weaken oyster defenses, thereby allowing opportunistic bacteria of the genus *Vibrio* to infect the host, leading to mortality (de Lorgeril et al., 2018; Petton et al., 2015).

Vibriosis is a major concern for both oyster hatcheries and field production, causing larval and spat damage, depending upon species. Vibriosis can result in impairment of velum structure and function (necrosis and detachment of velar cells) and deterioration of soft tissues, which result in mortalities (Dubert et al., 2017; Travers et al., 2015). The most pathogenic vibrios belong to the *Splendidus* and *Harveyi* clades or to the species *V. aestuarianus*, *V. tubiashii*, *V. coralliilyticus*, and *V. tapetis* (Travers et al., 2015). Although most *Vibrio* spp. can affect many bivalve species, *V. tapetis* appears to cause pathology only in Manila clams *Ruditapes philippinarum* and fishes (Levican et al., 2017; Reid et al., 2003). The resulting disease in clams is called Brown Ring Disease (BRD), because of the brown organic matrix deposit in the inner bivalve shell in responses to vibrio colonization. In advanced infections, *V. tapetis* can invade the circulatory system and lead to death (Paillard, 2004). The disease occurs in Europe, South Korea, and Japan (Travers et al., 2015).

Two species of protozoan parasite, *Marteilia refringens* and *Bonamia ostreae*, have strongly affected flat oyster *Ostrea edulis* production in France. *M. refringens* appeared in 1968 (Grizel et al., 1974) and *B. ostreae* in 1979 (Pichot et al., 1980), causing digestive gland and microcell diseases, respectively. Together these parasites caused a sharp decline in flat oyster production in France first, followed by in other countries in Europe (see special edition of the Diseases of

Aquatic Organisms journal, vol.110, 2014). Recently, *B. ostrea* was reported for the first time in New Zealand (Lane et al., 2016). Another member of this genus, *Bonamia exitiosa*, also caused mortalities in *Ostrea chilensis* in New Zealand (Hine et al., 2001) and in *O. puelchana* from wild and cultured populations in Argentina (Oehrens Kissner et al., 2014). The protozoan parasite *B. exitiosa* also has been reported in European flat oysters and can contribute to disease (Ramilo et al., 2014). In Australia, Winter Mortality disease causes mortalities in 2 and 3 year-old Sydney rock oysters *Saccostrea glomerata* in winter and early spring (Dove et al., 2013). The disease initially was suspected to be caused exclusively by the protistan parasite *Bonamia roughleyi* (Hill et al., 2014); however, the disease may actually result from a confluence of factors, including environmental conditions (Spiers et al., 2014). Another disease, called Queensland unknown (QX) disease caused by the haplosporidia parasite *Marteillia sydneyi*, affects the *S. glomerata* industry in Australia (Carrasco et al., 2015; Perkins and Wolf, 1976). The parasite *M. sydneyi* enters the host by gill and palp epithelia, with the progression of infection reaching the digestive tubules where parasite sporulation proceeds. This process causes the breakdown of the digestive gland, compromising feeding processes and leading to death (Kleeman et al., 2002).

Protozoan parasites from the *Perkinsus* genus are among the most pathogenic for oyster species worldwide (Villalba et al., 2011). The first outbreak occurred in 1946 in Louisiana (USA) with reported mortalities (up to 100%) of American oysters *C. virginica* that resulted in significant economic losses (Gouletquer et al., 1994). The disease is commonly referred to as "Dermo" in reference to the belief that it was a fungus, *Dermocystidium* (Mackin and Owen, 1950). Prevalence and body burden of *Perkinsus marinus* in *C. virginica* populations in the northeastern United States present a seasonal pattern strongly related to temperature, wherein maximum prevalence and intensity are reached in late summer - early autumn (Burreson and Ragone-Calvo, 1996). Occurrences of *P. marinus* in tropical regions, such as Mexico (Cáceres-Martínez et al., 2016) and Brazil (da Silva et al., 2014) were reported without mortalities of the native hosts, oysters *C. corteziensis* and *C. gasar*, respectively, but seasonal patterns of the disease seem to be similar to those in the USA. Intensities are predominantly light to moderate, without strongly deleterious, histopathological effects (Cáceres-Martínez et al., 2016; Queiroga et al., 2015; Scardua et al., 2017). Immunological functions, however, were impaired in intense infections (Queiroga et al., 2013), and genetic disturbances may occur because of widespread DNA methylation in *C. gasar* infected with *Perkinsus* spp. (Farias et al., 2017). Another representative species of this genus is *Perkinsus olseni* that affects clam *Ruditapes decussatus*

and *R. philippinarum* populations in Europe (Portugal, France, Spain and Italy) and Asia (South Korea and Japan). The intensity of the effect of this parasite is variable, being milder in France, despite the high prevalence (up to 100%) (Dang et al., 2010) and in Spain. Strong effect is observed in Portugal (Ruano et al., 2015), South Korea (Choi and Park, 2010) where mortalities were associated with stress of high seawater temperature (Nam et al., 2018), and Japan where high prevalence (100%) and intensity were associated with declining *R. philippinarum* populations (Waki et al., 2018).

1.2 Harmful Algal Blooms and co-occurrence with bivalve infectious diseases

HABs are a natural phenomenon characterized by localized and transient proliferation of some microalgal species favored by certain environmental conditions (Sellner et al., 2003). The frequency of HABs appears to have increased worldwide in recent decades and may be partly related to global change and human activities (Anderson et al., 2008; Glibert, 2017; Gobler et al., 2017; Hallegraeff, 1993; Wells et al., 2015). Toxic blooms in temperate areas tend to be punctual and seasonal phenomena, most often in spring and summer, and can last a few weeks to a few months. In tropical or sub-tropical areas, blooms can last much longer where no strong seasonal environmental changes occur (e.g. blooms of *Karenia brevis* persisted for over a year in Florida in 2018 (HABSOS, 2019)).

The harmful effect of HABs can either be direct (toxins or damaging algal morphology) or indirect (high concentration of microalgal cells leading to hypoxia). In this paper, we will discuss only HAB species producing described toxins (intra- or extracellular) and other uncharacterized bioactive extracellular compounds (BEC) with cytotoxic properties, which can affect bivalves.

The best-known microalgal biotoxins are those associated with symptoms in human consumers of contaminated seafood (see review from Berdalet et al., 2015; Gerssen et al., 2010; Lassus et al., 2016) as follows:

- 1) paralytic shellfish poisoning (PSP) is caused by saxitoxins (STXs, also referred to as Paralytic Shellfish Toxins, PSTs), produced by species of *Alexandrium*, *Gymnodinium* and *Pyrodinium*;

- 2) neurotoxic shellfish poisoning (NSP) caused by brevetoxins (PbTXs), mostly produced by *Karenia brevis*;
- 3) diarrhetic shellfish poisoning (DSP) associated with okadaic acid (OA) and dinophysistoxins (DTXs) produced by species of the genera *Dinophysis* and *Prorocentrum*;
- 4) amnesic shellfish poisoning (ASP) caused by domoic acid (DA) and analogs produced by species of *Pseudo-nitzschia* and *Nitzschia*;
- 5) ciguatera fish poisoning (CFP) caused by ciguatoxins (CTXs), produced by *Gambierdiscus* species.
- 6) Other microalgal toxins implicated in human toxic symptoms include azaspiracids (AZA), mostly produced by *Azadinium* and *Amphidoma* spp. and responsible for azaspiracid poisoning (AZP); palytoxins (PLTXs) and analogues, produced by *Ostreopsis* spp., are associated with food-borne poisoning as well as irritative symptoms through contact or aerosol inhalation. Other phycotoxins possibly toxic to humans, as suggested by potency in mouse bioassay or *in vitro* toxicological tests, include: spirolides (SPXs) produced by *Alexandrium ostenfeldii* and *A. peruvianum*; yessotoxins (YTXs) produced by species of *Protoceratium*, *Gonyaulax* and *Lingulodinium*; pectenotoxins (PTXs) produced by species of *Dinophysis*; and pinnatoxins (PnTXs) produced by *Vulcanodinium rugosum*.

Additionally, some HABs, mostly species not toxic to humans but known for being deleterious to marine fauna, are referred to as ichthyotoxic or fish-killing algae because they are mainly involved in mortality of farmed fishes (although most also affect shellfish and other marine organisms). The harmful mechanism of some of these algae is associated with hypoxia from high bloom density, mechanical irritation or clogging of gills, or production of compounds with cytotoxic and lytic activity. Some ichthyotoxins have been described already (e.g. prymnesins, karlotoxins, karmitoxin), and other compounds such as reactive oxygen species (ROS), polyunsaturated fatty acids, and mucopolysaccharides, appear to be involved in ichthyotoxic activity (Binzer et al., 2019; Dorantes-Aranda et al., 2015; Hallegraeff et al., 2017; Rasmussen et al., 2017, 2016). Most of the algal compounds responsible for ichthyotoxic activity, however, remain unknown or not well characterized (Rasmussen et al., 2016). Algae most involved in ichthyotoxic phenomena belong to the genera *Margalefidinium* (aka. *Cochlodinium*), *Chatonella*, *Pseudochatonella*, *Prymnesium*, *Chrysochromulina*, *Karenia*,

Karlodinium, *Pfiesteria*, *Heterocapsa*, *Heterosigma*, and *Akashiwo* (Hallegraeff et al., 2017; Lassus et al., 2016).

As sessile, filter-feeding organisms, bivalves interact directly with harmful microalgal blooms that occur along coasts worldwide. Bivalves can be in contact with both microalgal extracellular compounds, including BEC, as well as with intracellular toxins that can be released during digestion. Microalgal intracellular toxins often accumulate in bivalve tissues, mostly in the digestive gland (e.g. Lassudrie et al., 2014; Medhioub et al., 2012; see review of Landsberg, 2002). BEC have allelopathic, lytic, and oxidative activity reported upon microalgae, protists, or bivalve gametes (Arzul et al., 1999; Castrec et al., 2019; Flores et al., 2012; Le Goïc et al., 2014, 2013; Lelong et al., 2011; Long et al., 2018b; Tillmann and John, 2002). Toxins (intra- and extracellular) as well as BEC produced by HABs can be harmful to bivalves (e.g. Borcier et al., 2017; Castrec et al., 2018). HABs can affect physiological functions and ultimately survival of bivalves, at different life stages, and can result in major socio-economic and ecological problems (Landsberg, 2002; Shumway, 1990).

Considering spatial and temporal occurrence of HABs and infectious diseases, the probability for both potential stressors to affect bivalves simultaneously is high, as highlighted by Table 1, which lists (non-exhaustively) HABs reported to occur in regions and seasons where and when disease also occur. Only few studies, however, have reported co-occurrence of infectious disease outbreaks with HABs (Abi-Khalil et al., 2016; Jenkins et al., 2013). The low number of published observations probably is attributable more to a lack of reports than a lack of co-occurrence. The observations reported by these two field-studies suggested that *Alexandrium* spp. (Abi-Khalil et al., 2016; Jenkins et al., 2013) and/or *Pseudo-nitzschia* spp. (Jenkins et al., 2013) blooms in France and in Australia could increase oyster susceptibility to POMS. As causal relationship is difficult to infer from field observations, some experimental studies have focused on the interactive effects of both HABs and pathogen infection (summarized in Table 2).

The initial physiological status of the bivalve is one of the main conditions that modulates the outcome of a host-pathogen interaction (Samain and McCombie, 2007). Thus, the physio-pathological responses of bivalves exposed to both HABs and pathogens are the results of the interaction between: 1) the initial physiological status of the bivalve, 2) the effect of HABs, and 3) the effect of the pathogen upon the bivalve.

Since HABs interfere with bivalve immune cell variables and likely co-occur with disease infection, we review here the effects of HABs on hemocytes, including in bivalves co-exposed to pathogens, and their relationship with disease modulation. Other mechanisms by which infectious diseases can be modulated by HABs, through alteration of bivalve physiology, and through direct interactions between HABs and pathogens, are also explored.

2. Effects of HABs on bivalve cellular immunity and involvement in disease modulation

Compromised immunological status is a fundamental condition that contributes to disease acquisition and progression. Bivalve immune cells, called hemocytes, are responsive to disease infection, but can also be affected by environmental factors (e.g. temperature, salinity), including HABs. Therefore, the resulting effect on immunity and disease susceptibility has often been questioned.

In this section, after a brief description of the basics of bivalve immunity, we review the reported effects of HABs upon bivalve cellular effectors of immunity, the hemocytes, through the study of dual HAB-bivalve interactions (Table 3) and tripartite HAB-bivalve-pathogen interactions (Table 4).

2.1 Bivalve immunity and hemocytes

The immune system of bivalves is exclusively innate, lacking an adaptive memory as in vertebrates, although the existence of a form of immune memory in invertebrates is now being discussed, including in bivalves (see for example Lafont et al., 2019, 2017, for *C. gigas*). The immune cells, hemocytes, are implicated in multiple physiological functions in bivalves, including nutrition, shell mineralization and tissue repair and, the most important, immunity. These circulating cells perform defense functions against infectious agents, mainly by phagocytosis or encapsulation of infective agents (Cheng, 1996). The immune systems of bivalves also include soluble molecules in the hemolymph and mucus secretions that act in the

recognition and destruction or inactivation of invading organisms (Allam and Raftos, 2015; Song et al., 2010). It is now accepted that the immune system of bivalves is sophisticated and complex, presenting a high diversity of recognition molecules (Guo et al., 2015). Receptor proteins (PRPs) are inducible and secreted or are membrane proteins that recognize evolutionarily conserved pathogen-associated molecular patterns (PAMPs) (Allam and Raftos, 2015; Guo et al., 2015; Song et al., 2010). The ability to destroy microorganisms is associated with several immune molecules, including: hydrolytic enzymes, reactive oxygen and nitrogen species (ROS and RNS), and especially peptides and antimicrobial proteins which are widely studied in *C. gigas* (Destoumieux-Garzón et al., 2016; Schmitt et al., 2016, 2012b, 2012a). Hemocytes ontogeny, however, remains a mystery (Dyachuk, 2016; Li et al., 2017). Hemocytes circulate through the open circulatory system in the hemolymph and reach bivalve organs. Hemocytes are able to infiltrate tissues in response to infectious agents, such as bacteria and protozoa in a response termed inflammation (Carella et al., 2015; De Vico and Carella, 2012), and even in some cases transiting epithelial barriers and inadvertently bringing pathogens, such as *P. marinus*, inside the body (Lau et al., 2018) or expulsion of *M. sydneyi* (Kleeman et al., 2002). Total (THC) and differential hemocyte counts (DHC, granulocytes and hyalinocytes, aka. agranulocytes, discriminated by cell size and internal complexity), and functional measurements, phagocytosis activity particularly, are good indicators of bivalve immune status (Fournier et al., 2000; Vargas-Albores and Barracco, 2001).

2.2 Effects of HABs on bivalve hemocyte variables involved in immunity

In this review we focused on the most studied bivalve hemocyte characteristics modulated by HABs or associated toxins (section 2.2), as well as by combined exposure to HAB and pathogens (section 2.3). As we focused on immunological aspects of bivalves, articles reporting genotoxic effects on hemocytes were not included in this review. The most relevant results presented in 38 articles are summarized in Table 3 and 4.

THC, DHC, size, complexity, phagocytosis, and ROS production of circulating cells in the hemolymph are the most studied and common variables among the papers. Other hemocyte or plasma variables also were included, such as adhesion/aggregation, agglutination titer, phenoloxidase activity, viability, lysosomal membrane stability (cytotoxicity variables), and apoptosis. Apoptosis was added because programmed cell death may be particularly implicated in the immune defense against oyster intracellular pathogens (Sokolova, 2009), which has been

most-thoroughly investigated in the *P. marinus* – *C. virginica* model (Hughes et al., 2010; Lau et al., 2018c). The studies summarized in Table 3 include the most important commercial bivalve species worldwide, i.e., oysters primarily, followed by mussels, clams, and pectinids. A good representation of HAB species is included in published studies, such as producers of PSP toxins (*Alexandrium* spp.), DSP toxins (*Prorocentrum* and *Dynophysis* spp.), gymnodimin (*Karenia selliformis*), palytoxin (*Ostreopsis ovata*), karlotoxins (*Karlodinium veneficum*), and other toxic compounds still not well characterized from *Karenia mikimotoi*, *Heterosigma akashiwo*, and *Heterocapsa circularisquama*. The effects of purified phycotoxins were evaluated by *in vitro* incubation with hemocytes (STX, OA and PbTx) or by injection into the living bivalve (DA). No studies on hemocyte responses were carried out with algal producers of DA, YTX, CTX, or AZA toxins thus far (Table 3).

Bivalve hemocyte responses to a harmful alga depend upon the physiological state of the bivalve, but also on algal morphology and density, intraspecific variability of algal toxicity, and experimental conditions, which altogether may affect exposure to algal compounds, including through pre- and post-ingestive selection. Therefore, it is sometimes difficult to identify similar hemocyte response patterns, even considering the same alga and bivalve species. Two good examples of diversity of bivalve responses were obtained with the most-studied harmful algae *A. minutum* and *P. minimum*. In one experiment exposing *C. gigas* to *A. minutum*, only an increase in hemocyte phagocytosis was observed (Haber Korn et al., 2014). In contrast, in another study, THC and granulocytes increased and ROS production and phenoloxidase activity were not consistent (Haber Korn et al., 2010a). Regarding *P. minimum*, in one study, a simulated bloom increased *C. virginica* hemocyte phagocytosis, THC, granulocyte counts and viability, and aggregation decreased, while ROS varied (Hégaret and Wikfors, 2005a); whereas, the other experiment did not promote any hemocyte modulation (Hégaret and Wikfors, 2005b). Difference in hemocyte responses in these studies were attributed to differences between oyster populations and the means of algal exposure, i.e., cultured cells or a natural bloom. The latter study was later corroborated by an *in vitro* assay (Hégaret et al., 2011).

The species *A. catenella* (= *A. fundyense* as referred to in these studies; see last nomenclature in Fraga et al. 2015; Litaker et al. 2018) caused only mild effects upon hemocytes of adult *C. virginica* and *C. gigas* (Hégaret et al., 2007b) and *M. edulis* (Galimany et al., 2008b). In contrast, *A. catenella* and *A. tamarensis* caused cytotoxic effects in addition to impairment of immune responses in blue mussels *M. edulis* (Bianchi et al., 2019). Cytotoxic effects, however, were not directly related to PSTs. Indeed, both PST and non-PST strains that produced BEC

(also referred to as lytic compounds, LC), caused similar responses (Bianchi et al., 2019). Similarly, a highly-toxic, PST-producing strain of *A. catenella* (named *A. tamarensis* in the study; see new nomenclature in John et al., 2014; Fraga et al., 2015; Litaker et al., 2018) had no effects upon *R. philippinarum* or *M. arenaria* hemocytes; whereas, a non-PST strain of *A. tamarensis* did have effects, probably caused by bioactive compounds (Ford et al., 2008). Castrec et al. (2018) compared the effects of three *A. minutum* strains with contrasting PST and BEC characteristics. This study detected clear effects of PST on hemocyte mortality, hypothesized to involve apoptosis. PST effects upon hemocyte apoptosis were reported previously from *in vivo* (PST injection) and *in vitro* (PST exposure of hemocytes) assays with lions-paw scallops *Nodipecten subnodosus* (Estrada et al., 2014), in *C. gigas*, *in vivo* (*A. pacificum* exposure) (Medhioub et al., 2013), and *in vitro* (PST exposure of hemocytes) (Abi-Khalil et al., 2017). In other work with *A. minutum* and *C. gigas*, using *in vitro* exposures, similar responses, phagocytosis and ROS inhibition, were observed when hemocytes were exposed to PST-producing strains of *Alexandrium* spp. or directly to PSTs (Hégaret et al., 2011; Mello et al., 2013). The effects of PSTs themselves upon hemocytes seem to be relatively consistent, reducing phagocytosis (Astuya et al., 2015; Cao et al., 2018; Mello et al., 2013) and ROS (Astuya et al., 2015; Mello et al., 2013), regardless of bivalve species and administration mode (direct exposure or injection).

Separating effects of intracellular toxins and BEC has been recognized only recently because of a lack of awareness of the existence of BEC, and for practical reasons (availability of algal strains with different toxin and BEC characteristics). Correlations between hemocyte variables and toxin accumulation in tissues indicated that specific hemocyte responses to PST occur in *C. gigas* exposed to *Alexandrium* spp. (Haber Korn et al., 2010a; Lassudrie et al., 2016). This was later confirmed using different *A. minutum* strains with contrasting PST and BEC characteristics, showing some distinct hemocyte responses to PST vs. BEC (Bianchi et al., 2019; Castrec et al., 2018). Results of Lassudrie et al. (2016) with the oyster *C. gigas* exposed to *A. pacificum* (named *A. catenella*, in this study; see last nomenclature in John et al. 2014; Fraga et al. 2015; Litaker et al. 2018) indicated a relationship between PST levels and hyalinocyte variables, suggesting a particular role for this hemocyte type in response to PST; whereas, granulocyte modulations may respond to putative BEC. Other studies with *A. minutum*, however, did not support the hypothesis of a specific role of hyalinocytes in PST response (Castrec et al., 2018; Haber Korn et al., 2010a). Further studies are needed to decipher the functional responses of the different hemocyte types to PST vs. other bioactive compounds.

The toxic dinoflagellate *P. minimum* was also reported to have different effects upon *C. virginica* (Hégaret and Wikfors, 2005a, 2005b). Results of some experiments indicated increased phagocytosis and decreased aggregation and mortality (Hégaret and Wikfors, 2005a), other experiments did not detect any alteration in hemocyte variables (Hégaret and Wikfors, 2005b). A decrease of phagocytosis and of apoptotic hemocytes could be observed in clams *R. philippinarum* exposed to *P. minimum* (Hégaret et al., 2009). However, no effect on hemocyte was observed also in *M. edulis* exposed to *P. minimum*, (Galimany et al., 2008c). In contrast, a simulated bloom of *P. minimum* strongly affected all hemocyte variables of scallops *A. irradians irradians* (Hégaret and Wikfors, 2005b). This could indicate a higher sensitivity of scallops to some HABs like *P. minimum* than other bivalves, such as *C. virginica*. More studies are needed to support this hypothesis. The dinoflagellate *P. lima* and associated OA toxin were tested against mussels and clams and were shown to induce hemocyte apoptosis (Prado-Alvarez et al., 2013; Prego-Faraldo et al., 2016, 2015), except in one case of a natural bloom of *Prorocentrum* sp. and *Dinophysis* sp., and OA *in vitro* exposure (Prado-Alvarez et al., 2012). Effects of a simulated bloom of *P. lima* upon hemocyte functions were tested recently in the mussel *Perna perna*, and impaired phagocytosis, induced ROS production, and decreased THC were observed (Neves et al., 2019).

Although simulating a HAB under laboratory conditions may be a good approximation of reality, it is unquestionable that the best way to evaluate HAB effects upon bivalve cellular immune functions will be during a natural bloom; however, only three studies were conducted under those circumstances (Mello et al., 2010; Prado-Alvarez et al., 2012; Simões et al., 2015). During a natural bloom of *Dinophysis acuminata* in Santa Catarina state, South Brazil, some immune responses of three bivalve species belonging to different families were studied (Table 3). A reference group, animals collected at same site 30 days after the HAB exposure, were used for comparisons. Despite the differences in the concentration of algae during the blooms, it was clear that the bivalve species responded differently, *C. gigas* was the most resilient species; whereas, the mussel *P. perna* was the most affected, suggesting that this mussel may be a good bioindicator species. The sentinel candidature of mussels for environmental change and now for HABs was corroborated in another study with the mussel *M. galloprovincialis* exposed to the benthic dinoflagellate *Ostreopsis* cf. *ovata* with strong impairment (reduction) in granulocyte counts, phagocytosis activity, and lysosomal membrane stability as cytotoxic effects (Gorbi et al., 2013). In contrast, a study exposing oyster *C. gigas* hemocytes to brevetoxin did not reveal any cytotoxic effect (viability and apoptosis were evaluated) (Mello

et al., 2012). Even though no *Pseudo-nitzschia* or *Nitzschia* spp. blooms have been evaluated for effects upon immune cellular responses of bivalves, *M. edulis* was exposed to DA by intramuscular injection (Dizer et al., 2001). The acute effects of DA were seen in THC and phagocytosis, which increased initially, but after 7 days post-injection, variables normalized and hemocyte viability increased in a dose dependent manner during recovery.

2.3 Involvement of HAB effects on bivalve hemocytes in disease modulation?

The several effects of HABs upon hemocyte variables documented above led many authors to wonder about HAB effects upon immune functions and eventually upon disease susceptibility. These effects upon hemocytes have been hypothesized to modify the efficiency of bivalve immune functions, either by causing immunosuppression, particularly when functions such as phagocytosis are affected, or immunostimulation when considering increased THC or viability, for example. Few papers, however, report the combined effects of HABs and infectious agents upon immune functions of bivalves (Table 4). Most studied is the host-parasite model *R. philippinarum*-*P. olsenii* under the secondary effect of HABs *K. selliformis* and *K. mikimotoi* (da Silva et al., 2008; Hégaret et al., 2007a), *P. minimum* (Hégaret et al., 2009) and *A. ostentfeldii* (Lassudrie et al., 2014). Others evaluated the effects of HAB *A. catenella* (= *A. fundyense*), on *P. marinus* and *Bucephalus* sp. in oysters *C. virginica* (Lassudrie et al., 2015b), on trematodes Gymnophalidae in mussels *M. edulis* (Galimany et al., 2008b), on BRD caused by *V. tapetis* in clams *R. philippinarum* (Bricelj et al., 2011). The effect of *P. minimum* on Quahog Parasite Unknown (QPX) disease in clams *M. mercenaria* also was studied (Hégaret et al., 2010).

Many of these studies reported a repression of hemocyte responses during the interaction of bivalve – pathogen – HAB, which cannot be explained simply. Indeed, the combination of the two biotic factors causes hemocyte responses that might not correspond to the cumulative effect of each isolated factor, leading to antagonistic or synergistic responses. Additionally, the existence of combined effects of HAB and pathogen can modulate the bivalve response. For example *K. selliformis*, in a short exposure experiment, inhibited *R. philippinarum* hemocyte phagocytosis and increased hemocyte count in clams heavily infected with *P. olsenii*; whereas, the parasite itself did not affect the host (Hégaret et al., 2007a). Another toxic dinoflagellate, *A. ostentfeldii* had a very mild effect itself upon *R. philippinarum* hemocyte variables, but the combination with *P. olsenii* infection induced ROS production in lightly-infected clams comparable with the levels found in the heavily-infected animals unexposed to HAB (Lassudrie

et al., 2014). Likely, the interference between hemocyte responses to HAB and to pathogens are related to different functions.

It was hypothesized initially that effects of HABs upon hemocyte functionality would induce an immunocompromised state associated with increased susceptibility to disease. Such immunodepression, indicated by immunological changes and inflammatory response, was hypothesized by Galimany et al. (2008c) after observation of increased prevalence of trematodes in mussels exposed to *A. catenella* (= *A. fundyense*). Increase in parasite *P. marinus* prevalence was detected in the Eastern oysters exposed to *A. catenella* and concomitantly infected by trematodes *Bucephalus* sp. (Lassudrie et al., 2015b). This increase in parasite prevalence was associated with repression of hemocyte responses to the trematode infestation (Lassudrie et al., 2015b). The authors concluded that this higher susceptibility to *P. marinus* was likely the result of immunosuppression caused by HAB exposure in oysters already weakened by trematode infestation, as indicated by histological evidence.

Other studies reviewed in Table 4 could not link the hemocyte alteration caused by HABs with disease modulation.

3. Other bivalve physiological variables affected by HABs and possibly involved in disease modulation

HAB exposure can affect several physiological variables in bivalves. Here, we discuss the variables most likely involved in disease transmission or proliferation that have been reported to be modulated by HABs: feeding activity, and tissue alterations, often associated with inflammatory response.

3.1 Effects of HABs on feeding processes: possible involvement in pathogen transmission?

Filtration and ingestion rates could play an important role in the modulation of the tripartite interactions. Indeed, infection process is deeply related to the entry of pathogenic agents into

the bivalve body. Thus, it can be speculated that the lower the filtration rate, the lower the level of parasite inside the host.

Bivalves have the ability to adapt filtration and ingestion rates (feeding activity) to plankton composition. Indeed, decreases in clearance and filtration rates of several bivalve species in the presence of toxic microalgae have been widely reported in the literature (e.g. Contreras et al., 2011; Hégaret et al., 2007c; Jauffrais et al., 2012; Lassus et al., 2007, 1999; Shumway, 1990).

In oysters, reduction of *A. minutum* consumption seems to be mostly associated with algal BEC production, rather than to PST (Castrec et al., 2018). BECs irritate gills and, therefore, likely interfere with filtration or sorting functions and induce a protective behavioral response consisting of increased frequency of valve micro-closures (Castrec et al., 2018; Haberkorn et al., 2011; Tran et al., 2010). Conversely, contact with intracellular PSTs occurs mainly in the digestive organs after algal cell lysis. Pousse et al. (2018), however, demonstrated that a PST, non-BEC-producing strain of *A. minutum* also caused a decrease in feeding activity, thus suggesting the involvement of PST in filtration response. Furthermore, a recent study exposing mussels to *Alexandrium* spp. strains with different PST and BEC characteristics, also suggested that PSTs cause lower feeding activity (Bianchi et al., 2019). Such differences may, indeed, be caused by variability in both BEC and PST quality and quantity exposure in the two studies, or to different sensitivity of mussels and oysters.

Reduction in herpesvirus infection, associated with POMS, in juvenile *C. gigas* oysters was hypothesized to be associated with filtration decrease (Lassudrie et al., 2015a). No study to date, however, has investigated the specific links between effects of HABs upon feeding processes and infectious agent transmission and proliferation.

3.2 Tissue degeneration and inflammatory response caused by HABs: overall physiological weakness favorizing disease susceptibility?

Despite the ability of bivalves to avoid or reduce ingestion of some HAB species, they show histopathological effects resulting from the HAB exposures.

The most common tissues affected are epithelia from mantle, gills, digestive gland or foot, and the most common alterations found are: vacuolation, oedema, melanization of gills and mantle (putatively associated with oxidative stress), signs of irritation (mucus production), and

digestive tubules sloughing and atrophy (Haberkorn et al., 2010b; Hégaret et al., 2012, 2010, 2009; Lassudrie et al., 2014; Medhioub et al., 2012; Neves et al., 2019). Depending upon the organ, effects upon epithelia can be attributed to: (a) the extracellular compounds released by HABs, and (b) the intracellular toxins released after cell lysis at the end of the bloom or after direct contact with gills, or as a consequence of digestion. Indeed, the digestive gland is the organ accumulating most of the algal intracellular toxins (e.g. Lassudrie et al., 2014; Medhioub et al., 2012; see review of Landsberg, 2002).

Additionally, exposure to PST-producing HABs, or to purified toxins, can affect muscle integrity (Estrada et al., 2010; Haberkorn et al., 2010b; Hégaret et al., 2012, 2009), sometimes associated with altered escape response (Hégaret et al., 2012) or paralysis (Estrada et al., 2010).

Inflammatory response, consisting of hemocyte infiltration, often is observed in response to HAB or associated toxin exposure and likely is involved in repairing damaged tissues (e.g. Estrada et al., 2010; Hégaret et al., 2010; Hermabessiere et al., 2016; Lassudrie et al., 2015b, 2014; Medhioub et al., 2012). Hemocyte diapedesis through the digestive epithelia, and significative presence of hemocytes in the lumen of the intestine or around the gills, surrounding algal cells, and in the faeces, also suggest a mechanism comparable to encapsulation to protect the internal tissues from algal toxic compounds (Galimany et al., 2008c; Hégaret et al., 2009). These observations also lead some authors to assume an additional role for hemocytes in detoxification, i.e., transporting toxins or their products outside the tissues (Galimany et al., 2008c, 2008b; Hégaret et al., 2009). Detection of a type of lipophilic algal toxin, yessotoxins, in the hemocytes of mussels (Franchini et al., 2003), supports this hypothesis.

Degeneration of tissues and inflammatory response caused by HAB exposure are likely to increase susceptibility to pathogens by physiologically weakening bivalves and possibly interfering with hemocyte immune functions by recruiting hemocytes for tissue repair. This mechanism was suggested previously by Hégaret et al. (2012) and Lassudrie et al. (2015a) who observed concomitant tissue alterations and disease modulation under HAB exposure (Table 2).

4. Modulation of bivalve disease through direct effect of HABs on infectious agents

Direct interactions between HABs and infectious agents in the environment may affect bivalves indirectly by interfering with infectious agent transmission or proliferation. Two possible types of interactions are considered here: (a) effects of algal toxins and extracellular compounds on bivalve pathogens, which could either be beneficial or deleterious to pathogens, and (b) the possibility of microalgae, including HABs, to act as pathogen carriers.

4.1 Effects of algal toxins and extracellular compounds to bivalve pathogens

Bivalve pathogens may be in contact with toxic algal cells and associated toxic compounds directly before infecting the host (dissolved toxins or BEC) or during infection, within bivalve tissues (BEC or accumulated toxins).

Efforts to characterize the deleterious effects of extracellular compounds from HAB species upon various microorganisms have highlighted cytotoxic activity towards auto- and heterotrophic protists (Adolf et al., 2007; Arzul et al., 1999; Flores et al., 2012; Lelong et al., 2011; Long et al., 2018b, 2018a; Tillmann, 2003; Tillmann et al., 2008; Tillmann and John, 2002). As many species of HABs are mixotrophic, but also serve as prey, such extracellular bioactivity is believed to be related to predation and/or defense (anti-grazing) behavior (e.g. Adolf et al., 2007; Tillmann, 2003; Tillmann et al., 2008). Similarly, many other microalgae, including HAB species, can also feed on bacteria (Burkholder et al., 2008; Mitra et al., 2014).

Effects of HABs specifically upon protists, bacteria, and viruses pathogenic to bivalves have rarely been investigated (Table 2). Reports of decreased infections suggest that HABs could alter viability or virulence of bivalve pathogens (see further examples below, in da Silva et al., 2008; Hégaret et al., 2010; Lassudrie et al., 2015a). Another scenario considered is that HAB compounds could promote growth of pathogenic bacteria, as bacteria can consume organic nutrients excreted by microalgae (Seymour et al., 2017; Zhou et al., 2018). Among compounds excreted by microalgae, dimethylsulfoniopropionate (DMSP) is an organosulfur compound that is a potent chemoattractant to bacteria, which can metabolize it and use it as carbon and sulfur sources (Kiene et al., 2000; Simó, 2001). Not incidentally, harmful algal species are mostly

represented by dinoflagellates, which are important DMSP producers among phytoplankton groups (Caruana and Malin, 2014).

Many studies reported the allelopathic, lytic, and oxidative properties of *Alexandrium* spp. extracellular compounds toward different cells, such as microalgae, protists, or bivalve gametes (Arzul et al., 1999; Castrec et al., 2019; Flores et al., 2012; Le Goïc et al., 2014, 2013; Lelong et al., 2011; Long et al., 2018b; Tillmann and John, 2002). Although PST can be released extracellularly (Lefebvre et al., 2008; Persson et al., 2012), many studies reported the bioactivity of extracellular compounds produced by non-PST *Alexandrium* strains, demonstrating that these effects are independent from PSTs (Bianchi et al., 2019; Borcier et al., 2017; Castrec et al., 2019, 2020; Ford et al., 2008; Long et al., 2018b; Tillmann et al., 2007; Tillmann and John, 2002). Only a few studies focused on the characterization of these compounds, and their precise nature remains unknown (Flores et al., 2012; Ma et al., 2011, 2009).

Antibacterial compounds produced by microalgae have been widely reported in the literature (see for example Amaro et al., 2011; Kellam and Walker, 1989), including upon bacteria pathogenic to aquacultured species, in particular *Vibrio* spp. (Kokou et al., 2012). Antibacterial effects of HAB species, however, have rarely been studied. Lauritano et al. (2016) did not detect any activity of intracellular extracts of *Pseudo-nitzschia pseudodelicatissima*, *Alexandrium tamutum*, *A. andersonii* and *A. minutum* against different Gram-negative and Gram-positive bacteria infecting humans. PSTs target voltage-gated sodium channel receptors, resulting in modulations of sodium and potassium fluxes in many organisms, such as mammals (Catterall, 2000), bivalves (Boullot et al., 2017), and certain bacteria (Pomati et al., 2003). Such effects were also demonstrated in the bacterium *Vibrio fischeri* (Pomati et al., 2003). Therefore, it could be hypothesized that PST released by algal cells may interact with certain pathogenic bacteria *in situ* and result in modulation of infection in bivalves, although no study has demonstrated such effects so far.

Extracellular compounds from microalgae, including recognized HAB species, can have antiviral activity (see review by Amaro et al., 2011). For example, extracellular sulphated polysaccharides produced by *Gymnodinium impudicum* and *Margalefidinium polykrikoides* were reported to affect viruses pathogenic to human and other mammalian hosts (Hasui et al., 1995; Yim et al., 2004). The observation of a decrease in intensity of herpesvirus OsHV-1 μ Var infection in juvenile oysters *C. gigas* exposed experimentally to *A. pacificum* led the authors to

propose that the dinoflagellate extracellular compounds may also induce deleterious effects in viruses (Lassudrie et al., 2015a). The hypothesis was advanced that ROS produced in extracellular compounds of *Alexandrium* spp. (as evidenced by Flores et al., 2012) could have altered the OsHV-1 lipidic envelope and thereby infectivity. Despite these hypotheses, no study so far has confirmed the direct effects of HABs upon viruses pathogenic to bivalves.

Deleterious effects of other dinoflagellates upon bivalve parasites were reported. An *in vitro* experiment reported toxicity of a culture of *K. selliformis* to the Manila clam parasite *P. olseni*, as indicated by observation of a higher percentage of dead *P. olseni* cells and altered morphology (da Silva et al., 2008). These effects were believed to be responsible for decreasing intensity of *P. olseni* infection in clams *R. philippinarum* exposed experimentally to this alga (da Silva et al., 2008). Similarly, Hégaret et al. (2009) reported toxic effects of extracellular compounds from *P. minimum* to *P. olseni* cells during an *in vitro* experiment. These authors, however, did not observe any modulation of the *P. olseni* infection in Manila clams exposed to this alga, possibly because of the short duration of the exposure (6 days) (Hégaret et al., 2009).

Exposure to HABs were reported to modulate disease infections in bivalve, but only few studies have focused on the direct interaction between HABs and pathogenic agents so far (Table 2). Additional, *in vitro* exposures of pathogenic microorganisms to toxic microalgae and associated compounds are needed to better understand how microalgal extracellular compounds can alter bivalve infectious agent viability or virulence. Such methods, however, are limited by the inability to cultivate several pathogenic agents without the host (herpesvirus, certain bacteria and parasites). Field studies and developments in environmental DNA analyses may overcome this limitation.

4.2 Microalgae as vectors of infectious agents?

Phytoplankton dynamics may participate in disease transmission in bivalves as infectious agents may be attached to planktonic food particles. For example, viral particles of OsHV-1 were reported to attach to particles *in situ*, including algal food particles, leading the authors to hypothesize that plankton dynamics plays a key role in POMS transmission (Evans et al., 2015, 2014; Paul-Pont et al., 2013). Similarly, the acute viral necrobiotic disease virus, pathogen to the scallop *Chlamys farreri*, was shown to be carried by microalgae (Zhang et al., 2010).

Toxic dinoflagellates were reported to be vectors of human infectious agents, such as *Legionella pneumophila* or *Vibrio cholerae* (see review of Doucette, 1995; Rivera et al., 2013). No published study has researched so far the specific ability of toxic dinoflagellates to transport and disseminate bivalve pathogenic agents. The hypothesis of toxic microalgae acting as vectors for bivalve pathogenic agents could be further investigated to explore these phenomena.

5. Discussion and future research recommendations

This review, mainly based upon experimental studies, highlighted that HABs can modify disease processes in bivalves through several mechanisms (summarized in Figure 1). Modulation of hemocyte immune functions by HABs and associated compounds is one reported mechanism leading to the effects of HABs on disease in bivalves. HAB exposure can cause immunosuppression that promotes pathogen proliferation. Conversely, HAB exposure could stimulate immune responses. However, other mechanisms are likely involved, but are understudied thus far, and need more attention in the future. Indeed, HAB exposure can decrease bivalve filtration rate, which may in turn decrease pathogen acquisition. HABs can also cause tissue and organ lesions, increasing bivalve global physiological weakness and decreasing defense against pathogens. Pathogen viability or pathogenicity can be affected directly by HAB toxins or compounds in the seawater, which suggest they could also be affected in bivalve tissues exposed to HAB compounds. Finally, HABs could act as pathogen carriers and favor pathogen transmission.

Modulation of pathological condition in bivalves exposed to HABs and pathogens is the result of the interaction between the initial physiological status of the bivalve, the effects of the HABs, and the effects of the pathogen. It is challenging, therefore, to identify unique mechanisms applicable to all bivalves, and conclusions should be made for each bivalve population of interest and the specific HAB-pathogen combination. To best address a specific situation, we recommend experimental studies that simulate as close as possible the natural conditions. For example, experiments should be consistent with the seasonality of HABs and pathogen appearance in the field. To better understand the mechanisms involved, and for purposes of comparison with other studies, extensive information should be provided regarding the bivalve

initial condition (reproduction stage, age, origin, etc), and the HABs and pathogens (strain information, origin, growing conditions and stage, toxin production, etc) studied.

The ecological and economical significance of the effects of HABs upon bivalve disease are not well established, although experiments have shown clearly that such effects are likely. The study of these interactions in the field, and at the population level, is now needed to answer these questions.

The experimental studies and the field observations reported here only consider the immediate effects of a HAB exposure upon pathogen infections. Whether these acute effects can durably affect the dynamics of the host-pathogen interaction is a question that remains to be answered. From a selected number of HAB-bivalve-pathogen interaction studies reviewed herein, conceptual models of evolution of the host-pathogen dynamics can be proposed (Figure 2). We propose that the effect of a HAB on bivalve infectious disease would depend upon the temporal dynamics of both the disease and the HAB.

In the studies reviewed here, bacterial and viral infections led to rapid and intense consequences, i.e., acute effects (Figure 2A, B). POMS in France induces each year a mortality peak over a few weeks in oyster *C. gigas* spat. This disease is associated with herpesvirus and/or bacteria such as *V. tasmaniensis*. Lassudrie et al. (2015a) showed experimentally that a bloom of *A. pacificum* starting before herpesvirus exposure could alter transmission or proliferation of the virus. Conversely, Abi-Khalil et al. (2016) experimental results indicated that an *A. pacificum* bloom starting before injection of *V. tasmaniensis* could increase oyster mortality induced by the infection. These two experimental studies suggest that at the population level, mortality events could be modulated by toxic blooms occurring before the outbreak (Figure 2A, B).

Other papers reviewed here studied the effects of HABs on diseases associated with parasites *P. olseni* (da Silva et al., 2008; Lassudrie et al., 2014), *P. marinus* (Lassudrie et al., 2015b) and bacteria RLO (Rickettsia-like organisms; Hégaret et al., 2012). In the bivalve populations studied, these diseases do not induce severe mortality, resulting in long-lasting host-pathogen interactions. In the short-term, a toxic bloom can modulate the development of a bacterial (Hégaret et al., 2012) or parasitic (da Silva et al. 2008; Lassudrie et al., 2015b) infections (Figure 2C, D). Specifically, Lassudrie et al. (2015b) results indicated that an *A. catenella* (=

A. fundyense) bloom could promote *P. marinus* infection in oysters *C. virginica* that were already weakened by trematode infestation. The parasitic levels were not surveyed after the toxic algal exposure, it is therefore questionable whether this HAB would durably disrupt the *P. marinus* - *C. virginica* interaction, or whether the initial *P. marinus* levels would be quickly restored (Figure 2C). Nevertheless, it does not seem realistic that a short-term HAB exposure could have a long-term consequence. The equilibrium of the host-pathogen interaction would probably recover after exposure, as suggested by the results of da Silva et al. (2008) (Figure 2D). In this study, the intensity of infection by *P. olseni* in clams *R. philippinarum* decreased after 2 and 3 weeks of exposure of toxic dinoflagellate *K. selliformis*. Then, after 3 weeks of exposure at a lower *K. selliformis* concentration, the intensity of infection was restored to its original level (Figure 2D).

Another point to consider is that HABs are usually recurrent phenomena, whereas most studies assess the effect of a single HAB exposure. Long-term effects of repeated toxic blooms upon host-pathogen interaction dynamics at the population level should therefore be considered. Effects of HABs upon bivalve reproduction and on physiology through parental exposure were reported recently (e.g. Basti et al., 2013; Castrec et al., 2019, 2020; Rolton et al., 2015), and could ultimately alter bivalve population fitness in the case of repeated blooms. Adaptations of bivalve populations to recurrent toxic blooms, reflected by differential physiological responses between populations (Navarro et al., 2014), can lead to the emergence of resistant genotypes (Bricelj et al., 2010, 2005). The use of *in situ* monitoring and epidemiological models could contribute to understanding of effects of HABs upon the host-pathogen interactions at the population level.

Climate change conditions have been reported experimentally to alter physiology of bivalves and interaction with HABs (Farrell et al., 2015; Turner et al., 2016). Therefore, the HAB-bivalve-pathogen interaction likely will evolve under future climate conditions.

6. Conclusion

Increased disease susceptibility in bivalves caused by HAB exposure has been related to altered immune functions and global physiological distress. This pattern, however, is not always verified, as direct deleterious effects of HABs upon certain pathogenic agents can result in lower pathogen infection. Other processes likely interfere with disease infection under HAB exposure, such as filtration and direct interactions between pathogens and HABs. More studies are needed to understand the role of these processes in the complex tripartite bivalve-pathogen-HAB interaction. Future research should also focus on field observations to verify experimental results, and to identify the effect of HABs upon disease dynamics at the bivalve population level. Such considerations may provide clues for unexplained mortalities, and eventually improve shellfish management.

Declaration of interest

Nothing to declare.

Acknowledgments

The authors wish to thank Philippe Soudant and Caroline Fabioux (LEMAR, France) for their contribution to the first document in ML Ph.D. thesis that set the frame years ago for this review, and Raquel A. F. Neves (Universidade Federal do Estado do Rio de Janeiro, Brazil) for her help with listing HAB species from Brazil. Fundings from the PADDLE project (funding by the European Union's Horizon 2020 research and innovation program—grant agreement no. 73427) and from the French National program EC2CO (Ecosphère Continentale et Côtière, LET IT BI project) enabled to collaborate on this manuscript.

References

Abi-Khalil, C., Finkelstein, D.S., Conejero, G., Du Bois, J., Destoumieux-Garzon, D., Rolland, J.L., 2017. The paralytic shellfish toxin, saxitoxin, enters the cytoplasm and induces

- apoptosis of oyster immune cells through a caspase-dependent pathway. *Aquat. Toxicol.* 190, 133–141. <https://doi.org/10.1016/j.aquatox.2017.07.001>
- Abi-Khalil, C., Lopez-Joven, C., Abadie, E., Savar, V., Amzil, Z., Laabir, M., Rolland, J.-L., 2016. Exposure to the Paralytic Shellfish Toxin Producer *Alexandrium catenella* Increases the Susceptibility of the Oyster *Crassostrea gigas* to Pathogenic Vibrios. *Toxins (Basel)*. 8, 24. <https://doi.org/10.3390/toxins8010024>
- Adlard, R.D., Wesche, S.J., 2005. Aquatic Animal Health Subprogram: development of a disease zoning policy for *Marteilia sydneyi* to support sustainable production, health certification and trade in the Sydney rock oyster, in: Fisheries Research and Development Corporation Final Report 2001-214. South Brisbane, Australia: Queensland Museum, Brisbane, p. 46.
- Adolf, J.E., Krupatkina, D., Bachvaroff, T., Place, A.R., 2007. Karlotoxin mediates grazing by *Oxyrrhis marina* on strains of *Karlodinium veneficum*. *Harmful Algae* 6, 400–412. <https://doi.org/10.1016/j.hal.2006.12.003>
- Ajani, P., Brett, S., Krogh, M., Scanes, P., Webster, G., Armand, L., 2013. The risk of harmful algal blooms (HABs) in the oyster-growing estuaries of New South Wales, Australia. *Environ. Monit. Assess.* 185, 5295–5316. <https://doi.org/10.1007/s10661-012-2946-9>
- Ajani, P., Harwood, D., Murray, S., 2017. Recent Trends in Marine Phycotoxins from Australian Coastal Waters. *Mar. Drugs* 15, 33. <https://doi.org/10.3390/md15020033>
- Ajani, P.A., Hallegraeff, G.M., Allen, D., Coughlan, A., Richardson, A.J., Armand, L.K., Ingleton, T., Murray, S.A., 2016. Oceanography and Marine Biology, *Oceanography and Marine Biology: An Annual Review*. CRC Press. <https://doi.org/10.1201/9781315368597>
- Allam, B., Raftos, D., 2015. Immune responses to infectious diseases in bivalves. *J. Invertebr. Pathol.* 131, 121–136. <https://doi.org/10.1016/j.jip.2015.05.005>
- Amaro, H.M., Guedes, A.C., Malcata, F.X., 2011. Antimicrobial activities of microalgae: an invited review, in: Méndez-Vilas, A. (Ed.), *Science against Microbial Pathogens: Communicating Current Research and Technological Advances*. pp. 1272–1280.
- Anderson, D.M., Burkholder, J.M., Cochlan, W.P., Glibert, P.M., Gobler, C.J., Heil, C.A., Kudela, R.M., Parsons, M.L., Rensel, J.E.J., Townsend, D.W., Trainer, V.L., Vargo, G.A., 2008. Harmful algal blooms and eutrophication: Examining linkages from selected coastal regions of the United States. *Harmful Algae* 8, 39–53. <https://doi.org/10.1016/j.hal.2008.08.017>
- Andrews, J., 1980. A Review of Introductions of Exotic Oysters and Biological Planning for New Importations. *Mar. Fish. Rev.* 1–11.
- Aquino, E.P., Borges, G.C.P., Honorato-da-Silva, M., Passavante, J.Z.O., Silva-Cunha, M.G.G.S., 2015. Phytoplankton in a tropical estuary, Northeast Brazil: composition and life forms. *Check List* 11, 1633. <https://doi.org/10.15560/11.3.1633>
- Arzul, G., Seguel, M., Guzman, L., Erard-Le Denn, E., 1999. Comparison of allelopathic properties in three toxic *Alexandrium* species. *J. Exp. Mar. Bio. Ecol.* 232, 285–295.
- Arzul, I., Carnegie, R.B., 2015. New perspective on the haplosporidian parasites of molluscs. *J. Invertebr. Pathol.* 131, 32–42. <https://doi.org/10.1016/j.jip.2015.07.014>

- Arzul, I., Corbeil, S., Morga, B., Renault, T., 2017. Viruses infecting marine molluscs. *J. Invertebr. Pathol.* 147, 118–135. <https://doi.org/10.1016/j.jip.2017.01.009>
- Astuya, A., Carrera, C., Ulloa, V., Aballay, A., Núñez-Acuña, G., Hégaret, H., Gallardo-Escárate, C., 2015. Saxitoxin modulates immunological parameters and gene transcription in *Mytilus chilensis* hemocytes. *Int. J. Mol. Sci.* 16, 15235–15250. <https://doi.org/10.3390/ijms160715235>
- Barbosa Solomieu, V., Renault, T., Travers, M., 2015. Mass mortality in bivalves and the intricate case of the Pacific oyster, *Crassostrea gigas*. *J. Invertebr. Pathol.* 131, 2–10. <https://doi.org/10.1016/j.jip.2015.07.011>
- Basti, L., Endo, M., Segawa, S., 2011. Physiological, pathological, and defense alterations in Manila clams (short-neck clams), *Ruditapes philippinarum*, induced by *Heterocapsa circularisquama*. *J. Shellfish Res.* 30, 829–844.
- Basti, L., Nagai, K., Tanaka, Y., Segawa, S., 2013. Sensitivity of gametes, fertilization, and embryo development of the Japanese pearl oyster, *Pinctada fucata martensii*, to the harmful dinoflagellate, *Heterocapsa circularisquama*. *Mar. Biol.* 160, 211–219. <https://doi.org/10.1007/s00227-012-2079-2>
- Berdalet, E., Fleming, L.E., Gowen, R., Davidson, K., Hess, P., Backer, L.C., Moore, S.K., Hoagland, P., Enevoldsen, H., 2015. Marine harmful algal blooms, human health and wellbeing: Challenges and opportunities in the 21st century. *J. Mar. Biol. Assoc. United Kingdom.* 61–91. <https://doi.org/10.1017/S0025315415001733>
- Berthe, F.C.J., Le Roux, F., Adlard, R., Figueiras, A., Villalba, A., Reece, K.S., Ordás, M.C., Casas, S.M., Figueras, A., 2004. Marteiliosis in Molluscs: a review. *Aquat. Living Resour.* 17, 433–444. <https://doi.org/10.1051/alr>
- Bianchi, V.A., Langeloh, H., Tillmann, U., Krock, B., Müller, A., Bickmeyer, U., Abele, D., 2019. Separate and combined effects of neurotoxic and lytic compounds of *Alexandrium* strains on *Mytilus edulis* feeding activity and hemocyte function. *Fish Shellfish Immunol.* 84, 414–422. <https://doi.org/10.1016/j.fsi.2018.10.024>
- Binzer, S.B., Svenssen, D.K., Daugbjerg, N., Alves-de-Souza, C., Pinto, E., Hansen, P.J., Larsen, T.O., Varga, E., 2019. A-, B- and C-type prymnesins are clade specific compounds and chemotaxonomic markers in *Prymnesium parvum*. *Harmful Algae* 81, 10–17. <https://doi.org/10.1016/j.hal.2018.11.010>
- Borcier, E., Morvezen, R., Boudry, P., Miner, P., Charrier, G., Laroche, J., Hégaret, H., 2017. Effects of bioactive extracellular compounds and paralytic shellfish toxins produced by *Alexandrium minutum* on growth and behaviour of juvenile great scallops *Pecten maximus*. *Aquat. Toxicol.* 184, 142–154. <https://doi.org/10.1016/j.aquatox.2017.01.009>
- Boullot, F., Castrec, J., Bidault, A., Dantas, N., Payton, L., Perrigault, M., Tran, D., Amzil, Z., Boudry, P., Soudant, P., Hégaret, H., Fabioux, C., 2017. Molecular Characterization of Voltage-Gated Sodium Channels and Their Relations with Paralytic Shellfish Toxin Bioaccumulation in the Pacific Oyster *Crassostrea gigas*. *Mar. Drugs* 15, 21. <https://doi.org/10.3390/md15010021>
- Branco, S., Almeida, L.L., Alves-de-Souza, C., Oliveira, M.M.M., Proença, L.A.O., Menezes, M., 2019. Morphological and genetic characterization of bloom-forming Raphidophyceae

- from Brazilian coast. *Phycol. Res.* pre.12377. <https://doi.org/10.1111/pre.12377>
- Bricelj, V., Ford, S., Lambert, C., Barbou, A., Paillard, C., 2011. Effects of toxic *Alexandrium tamarense* on behavior, hemocyte responses and development of brown ring disease in Manila clams. *Mar. Ecol. Prog. Ser.* 430, 35–48. <https://doi.org/10.3354/meps09111>
- Bricelj, V.M., Connell, L., Konoki, K., MacQuarrie, S.P., Scheuer, T., Catterall, W.A., Trainer, V.L., 2005. Sodium channel mutation leading to saxitoxin resistance in clams increases risk of PSP. *Nature* 434, 763–767.
- Bricelj, V.M., MacQuarrie, S.P., Doane, J.A.E., Connell, L.B., 2010. Evidence of selection for resistance to paralytic shellfish toxins during the early life history of soft-shell clam (*Mya arenaria*) populations. *Limnol. Oceanogr.* 55, 2463–3590.
- Burkholder, J.M., Glibert, P.M., Skelton, H.M., 2008. Mixotrophy, a major mode of nutrition for harmful algal species in eutrophic waters. *Harmful Algae* 8, 77–93. <https://doi.org/10.1016/j.hal.2008.08.010>
- Burreson, E.M., Ragone-Calvo, L.M., 1996. Epizootiology of *Perkinsus marinus* disease of oysters in Chesapeake Bay, with emphasis on data since 1985. *J. Shellfish Res.* 15, 17–34.
- Cáceres-Martínez, J., Madero-López, L.H., Padilla-Lardizábal, G., Vásquez-Yeomans, R., 2016. Epizootiology of *Perkinsus marinus*, parasite of the pleasure oyster *Crassostrea corteziensis*, in the Pacific coast of Mexico. *J. Invertebr. Pathol.* 139, 12–18. <https://doi.org/10.1016/j.jip.2016.07.003>
- Cao, R., Wang, D., Wei, Q., Wang, Q., Yang, D., Liu, H., Dong, Z., Zhang, X., Zhang, Q., Zhao, J., 2018. Integrative Biomarker Assessment of the Influence of Saxitoxin on Marine Bivalves: A Comparative Study of the Two Bivalve Species Oysters, *Crassostrea gigas*, and Scallops, *Chlamys farreri*. *Front. Physiol.* 9, 1–14. <https://doi.org/10.3389/fphys.2018.01173>
- Carella, F., Feist, S.W., Bignell, J.P., De Vico, G., 2015. Comparative pathology in bivalves: Aetiological agents and disease processes. *J. Invertebr. Pathol.* 131, 107–120. <https://doi.org/10.1016/j.jip.2015.07.012>
- Carrasco, N., Green, T., Itoh, N., 2015. *Marteilia* spp. parasites in bivalves: A revision of recent studies. *J. Invertebr. Pathol.* 131, 43–57. <https://doi.org/10.1016/j.jip.2015.07.016>
- Caruana, A.M.N., Malin, G., 2014. The variability in DMSP content and DMSP lyase activity in marine dinoflagellates. *Prog. Oceanogr.* 120, 410–424. <https://doi.org/10.1016/j.pocean.2013.10.014>
- Castrec, J., Hégaret, H., Alunno-Bruscia, M., Picard, M., Soudant, P., Petton, B., Boulais, M., Suquet, M., Quéau, I., Ratiskol, D., Foulon, V., Le Goïc, N., Fabioux, C., 2019. The dinoflagellate *Alexandrium minutum* affects development of the oyster *Crassostrea gigas*, through parental or direct exposure. *Environ. Pollut.* 246, 827–836. <https://doi.org/10.1016/j.envpol.2018.11.084>
- Castrec, J., Hégaret, H., Huber, M., Le Grand, J., Huvet, A., Tallec, K., Boulais, M., Soudant, P., Fabioux, C., 2020. The toxic dinoflagellate *Alexandrium minutum* impaired oyster free living stages, embryos and larvae. *Harmful Algae* (in press).
- Castrec, J., Soudant, P., Payton, L., Tran, D., Miner, P., Lambert, C., Le Goïc, N., Huvet, A.,

- Quillien, V., Boullot, F., Amzil, Z., Hégaret, H., Fabioux, C., 2018. Bioactive extracellular compounds produced by the dinoflagellate *Alexandrium minutum* are highly detrimental for oysters. *Aquat. Toxicol.* 199, 188–198. <https://doi.org/10.1016/j.aquatox.2018.03.034>
- Catterall, 2000. From Ionic Currents to Molecular Review Mechanisms: The Structure and Function of Voltage-Gated Sodium Channels. *Neuron* 26, 13–25.
- Cheng, T.C., 1996. Haemocytes: Forms and functions, in: Kennedy, V., Newell, R., Eble, A. (Eds.), *The Eastern Oyster Crassostrea virginica*. College Park: Maryland Sea Grant, pp. 299–333.
- Choi, K.S., Park, K.I., 2010. Review on the Protozoan parasite *Perkinsus olseni* (Lester and Davis 1981) infection in asian waters, in: Ishimatsu, A., Lier, H.-J. (Eds.), *Coastal Environmental and Ecosystem Issues of the East China Sea*. TERRABUP; Nagasaki university, pp. 269–281.
- Comps, M., 1988. Epizootic diseases of oysters associated with viral infections. *Am. Fish. Soc. Spec. Publ.* 18, 23–37.
- Contreras, A.M., Marsden, I.D., Munro, M.H.G., 2011. Effects of short-term exposure to paralytic shellfish toxins on clearance rates and toxin uptake in five species of New Zealand bivalve. *Mar. Freshw. Res.* 63, 166–174. <https://doi.org/http://dx.doi.org/10.1071/MF11173>
- Cuellar-Martinez, T., Ruiz-Fernández, A.C., Alonso-Hernández, C., Amaya-Monterrosa, O., Quintanilla, R., Carrillo-Ovalle, H.L., Arbeláez M, N., Díaz-Asencio, L., Méndez, S.M., Vargas, M., Chow-Wong, N.F., Valerio-Gonzalez, L.R., Enevoldsen, H., Dechraoui Bottein, M.-Y., 2018. Addressing the Problem of Harmful Algal Blooms in Latin America and the Caribbean- A Regional Network for Early Warning and Response. *Front. Mar. Sci.* 5, 1–7. <https://doi.org/10.3389/fmars.2018.00409>
- da Silva, P.M., Scardua, M.P., Vianna, R.T., Mendonça, R.C., Vieira, C.B., Dungan, C.F., Scott, G.P., Reece, K.S., 2014. Two *Perkinsus* spp. infect *Crassostrea gasar* oysters from cultured and wild populations of the Rio São Francisco estuary, Sergipe, northeastern Brazil. *J. Invertebr. Pathol.* 119, 62–71. <https://doi.org/10.1016/j.jip.2014.04.005>
- da Silva, P.P.M., Hégaret, H., Lambert, C., Wikfors, G.H., Le Goïc, N., Shumway, S.E., Soudant, P., 2008. Immunological responses of the Manila clam (*Ruditapes philippinarum*) with varying parasite (*Perkinsus olseni*) burden, during a long-term exposure to the harmful alga, *Karenia selliformis*, and possible interactions. *Toxicon* 51, 563–573. <https://doi.org/10.1016/j.toxicon.2007.11.006>
- Dang, C., de Montaudouin, X., Caill-Milly, N., Trumbic, Z., 2010. Spatio-temporal patterns of perkinsosis in the Manila clam *Ruditapes philippinarum* from Arcachon Bay (SW France). *Dis. Aquat. Organ.* 91, 151–159.
- De Carli, G., 2014. Distribuição e abundância de dinoflagelados epi-bentônicos na costa Nordeste do Brasil. Dissertação de Mestrado, Universidade Federal do Estado do Rio de Janeiro.
- de Kantzow, M., Hick, P., Becker, J., Whittington, R., 2016. Effect of water temperature on mortality of Pacific oysters *Crassostrea gigas* associated with microvariant ostreid herpesvirus 1 (OsHV-1 μ Var). *Aquac. Environ. Interact.* 8, 419–428.

<https://doi.org/10.3354/aei00186>

- de Lorgeril, J., Lucasson, A., Petton, B., Toulza, E., Montagnani, C., Clerissi, C., Vidal-Dupiol, J., Chaparro, C., Galinier, R., Escoubas, J.-M., Haffner, P., Dégremont, L., Charrière, G.M., Lafont, M., Delort, A., Vergnes, A., Chiarello, M., Faury, N., Rubio, T., Leroy, M.A., Pérignon, A., Régler, D., Morga, B., Alunno-Bruscia, M., Boudry, P., Le Roux, F., Destoumieux-Garzón, D., Gueguen, Y., Mitta, G., 2018. Immune-suppression by OsHV-1 viral infection causes fatal bacteraemia in Pacific oysters. *Nat. Commun.* 9, 4215. <https://doi.org/10.1038/s41467-018-06659-3>
- De Rijcke, M., Van Acker, E., Nevejan, N., De Schampelaere, K.A.C., Janssen, C.R., 2016. Toxic dinoflagellates and *Vibrio* spp. act independently in bivalve larvae. *Fish Shellfish Immunol.* 57, 236–242. <https://doi.org/10.1016/j.fsi.2016.08.027>
- De Vico, G., Carella, F., 2012. Morphological features of the inflammatory response in molluscs. *Res. Vet. Sci.* 93, 1109–1115. <https://doi.org/10.1016/j.rvsc.2012.03.014>
- Destoumieux-Garzón, D., Rosa, R.D., Schmitt, P., Barreto, C., Vidal-Dupiol, J., Mitta, G., Gueguen, Y., Bachère, E., 2016. Antimicrobial peptides in marine invertebrate health and disease: Table 1. *Philos. Trans. R. Soc. B Biol. Sci.* 371, 20150300. <https://doi.org/10.1098/rstb.2015.0300>
- Dizer, H., Fischer, B., Harabawy, A.S., Hennion, M.-C., Hansen, P.-D., 2001. Toxicity of domoic acid in the marine mussel *Mytilus edulis*. *Aquat. Toxicol.* 55, 149–156. [https://doi.org/10.1016/S0166-445X\(01\)00178-3](https://doi.org/10.1016/S0166-445X(01)00178-3)
- Dorantes-Aranda, J.J., Seger, A., Mardones, J.I., Nichols, P.D., Hallegraef, G.M., 2015. Progress in Understanding Algal Bloom-Mediated Fish Kills: The Role of Superoxide Radicals, Phycotoxins and Fatty Acids. *PLoS One* 10, e0133549. <https://doi.org/10.1371/journal.pone.0133549>
- Doucette, G.J., 1995. Interactions between bacteria and harmful algae: a review. *Nat. Toxins* 3, 65–74.
- Dove, M., Nell, J.A., Mcorrie, S., O'connor, W.A., 2013. Assessment of Qx and Winter Mortality Disease Resistance of Mass Selected Sydney Rock Oysters, *Saccostrea glomerata* (Gould, 1850), in the Hawkesbury River and Merimbula Lake, NSW Australia. *J. Shellfish Res.* 32, 681–687. <https://doi.org/10.2983/035.032.0309>
- Dubert, J., Barja, J.L., Romalde, J.L., 2017. New Insights into Pathogenic Vibrios Affecting Bivalves in Hatcheries: Present and Future Prospects. *Front. Microbiol.* 8, 1–16. <https://doi.org/10.3389/fmicb.2017.00762>
- Dyachuk, V.A., 2016. Hematopoiesis in Bivalvia larvae: Cellular origin, differentiation of hemocytes, and neoplasia. *Dev. Comp. Immunol.* 65, 253–257. <https://doi.org/10.1016/j.dci.2016.07.019>
- Enríquez-Espinoza, T., Grijalva-Chon, J., Castro-Longoria, R., Ramos-Paredes, J., 2010. *Perkinsus marinus* in *Crassostrea gigas* in the Gulf of California. *Dis. Aquat. Organ.* 89, 269–273. <https://doi.org/10.3354/dao02199>
- Estrada, N., Ascencio, F., Shoshani, L., Contreras, R.G., 2014. Apoptosis of hemocytes from lions-paw scallop *Nodipecten subnodosus* induced with paralyzing shellfish poison from *Gymnodinium catenatum*. *Immunobiology* 219, 964–974.

<https://doi.org/10.1016/j.imbio.2014.07.006>

- Estrada, N., Rodríguez-Jaramillo, C., Contreras, G., Ascencio, F., 2010. Effects of induced paralysis on hemocytes and tissues of the giant lions-paw scallop by paralyzing shellfish poison. *Mar. Biol.* 157, 1401–1415. <https://doi.org/10.1007/s00227-010-1418-4>
- Evans, O., Hick, P., Dhand, N., Whittington, R., 2015. Transmission of Ostreid herpesvirus-1 in *Crassostrea gigas* by cohabitation: effects of food and number of infected donor oysters. *Aquac. Environ. Interact.* 7, 281–295. <https://doi.org/10.3354/aei00160>
- Evans, O., Paul-Pont, I., Hick, P., Whittington, R.J., 2014. A simple centrifugation method for improving the detection of Ostreid herpesvirus-1 (OsHV-1) in natural seawater samples with an assessment of the potential for particulate attachment. *J. Virol. Methods* 210, 59–66. <https://doi.org/10.1016/j.jviromet.2014.09.023>
- Farias, N.D., de Oliveira, N.F.P., da Silva, P.M., 2017. *Perkinsus* infection is associated with alterations in the level of global DNA methylation of gills and gastrointestinal tract of the oyster *Crassostrea gasar*. *J. Invertebr. Pathol.* 149, 76–81. <https://doi.org/10.1016/j.jip.2017.08.007>
- Farley, C., Wolf, P., Elston, R., 1988. A long-term study of ‘microcell’ disease in oysters with a description of a new genus, *Mikrocytos* (g. n.), and two new species, *Mikrocytos mackini* (sp. n.) and *Mikrocytos roughleyi* (sp. n.). *Fish. Bull.* 86, 581–593.
- Farrell, H., Seebacher, F., O’Connor, W., Zammit, A., Harwood, D.T., Murray, S., 2015. Warm temperature acclimation impacts metabolism of paralytic shellfish toxins from *Alexandrium minutum* in commercial oysters. *Glob. Chang. Biol.* 21, 3402–3413. <https://doi.org/10.1111/gcb.12952>
- Flores, H.S., Wikfors, G.H., Dam, H.G., 2012. Reactive oxygen species are linked to the toxicity of the dinoflagellate *Alexandrium* spp. to protists. *Aquat. Microb. Ecol.* 66, 199–209.
- Ford, S.E., Bricelj, V.M., Lambert, C., Paillard, C., 2008. Deleterious effects of a nonPST bioactive compound(s) from *Alexandrium tamarense* on bivalve hemocytes. *Mar. Biol.* 154, 241–253. <https://doi.org/10.1007/s00227-008-0917-z>
- Ford, S.E., Smolowitz, R., 2007. Infection dynamics of an oyster parasite in its newly expanded range. *Mar. Biol.* 151, 119–133. <https://doi.org/10.1007/s00227-006-0454-6>
- Fournier, M., Cyr, D., Blakley, B., Boermans, H., Brousseau, P., 2000. Phagocytosis as a biomarker of immunotoxicity in wildlife species exposed to environmental xenobiotics. *Am. Zool.* 40, 412–420.
- Fraga, S., Sampedro, N., Larsen, J., Moestrup, Ø, Calado, A., 2015. Arguments against the proposal 2302 by John & al. to reject the name *Gonyaulax catenella* (*Alexandrium catenella*). *Taxon* 64(3), 634–635. <http://dx.doi.org/10.12705/643.15>
- Franchini, A., Milandri, A., Poletti, R., Ottaviani, E., 2003. Immunolocalization of yessotoxins in the mussel *Mytilus galloprovincialis*. *Toxicon* 41, 967–970. [https://doi.org/10.1016/S0041-0101\(03\)00039-4](https://doi.org/10.1016/S0041-0101(03)00039-4)
- Galimany, E., Place, A.R., Jutson, M., Pipe, R.K., 2008a. The effects of feeding *Karlodinium veneficum* (PLY # 103 ; *Gymnodinium veneficum* Ballantine) to the blue mussel *Mytilus*

- edulis* 7, 91–98. <https://doi.org/10.1016/j.hal.2007.05.004>
- Galimany, E., Sunila, I., Hégaret, H., Ramón, M., Wikfors, G.H., 2008b. Experimental exposure of the blue mussel (*Mytilus edulis*, L.) to the toxic dinoflagellate *Alexandrium fundyense*: Histopathology, immune responses, and recovery. *Harmful Algae* 7, 702–711. <https://doi.org/10.1016/j.hal.2008.02.006>
- Galimany, E., Sunila, I., Hégaret, H., Ramón, M., Wikfors, G.H., 2008c. Pathology and immune response of the blue mussel (*Mytilus edulis* L.) after an exposure to the harmful dinoflagellate *Prorocentrum minimum*. *Harmful Algae* 7, 630–638. <https://doi.org/10.1016/j.hal.2008.01.001>
- Gerssen, A., Pol-Hofstad, I.E., Poelman, M., Mulder, P.P.J., Van den Top, H.J., De Boer, J., 2010. Marine Toxins: Chemistry, Toxicity, Occurrence and Detection, with Special Reference to the Dutch Situation. *Toxins (Basel)* 2, 878–904. <https://doi.org/10.3390/toxins2040878>
- Gittenberger, A., Voorbergen-Laarman, M.A., Engelsma, M.Y., 2016. Ostreid herpesvirus OsHV-1 μ Var in Pacific oysters *Crassostrea gigas* (Thunberg 1793) of the Wadden Sea, a UNESCO world heritage site. *J. Fish Dis.* 39, 105–109. <https://doi.org/10.1111/jfd.12332>
- Glibert, P.M., 2017. Eutrophication, harmful algae and biodiversity — Challenging paradigms in a world of complex nutrient changes. *Mar. Pollut. Bull.* 124, 591–606. <https://doi.org/10.1016/j.marpolbul.2017.04.027>
- Gobler, C.J., Doherty, O.M., Hattenrath-Lehmann, T.K., Griffith, A.W., Kang, Y., Litaker, R.W., 2017. Ocean warming since 1982 has expanded the niche of toxic algal blooms in the North Atlantic and North Pacific oceans. *Proc. Natl. Acad. Sci.* 114, 4975–4980. <https://doi.org/10.1073/pnas.1619575114>
- Gorbi, S., Avio, G.C., Benedetti, M., Totti, C., Accoroni, S., Pichierri, S., Bacchiocchi, S., Orletti, R., Graziosi, T., Regoli, F., 2013. Effects of harmful dinoflagellate *Ostreopsis cf. ovata* exposure on immunological, histological and oxidative responses of mussels *Mytilus galloprovincialis*. *Fish Shellfish Immunol.* 35, 941–950. <https://doi.org/10.1016/j.fsi.2013.07.003>
- Gouletquer, P., Héral, M., Rothschild, B.J., 1994. Causes of decline of oyster production (*Crassostrea virginica*) in the Maryland portion of the Chesapeake Bay: A literature study. *Haliotis* 23, 87–112.
- Green, T.J., Raftos, D., O'Connor, W., Adlard, R.D., Barnes, A.C., 2011. Disease Prevention Strategies for QX Disease (*Marteilia sydneyi*) of Sydney Rock Oysters (*Saccostrea glomerata*). *J. Shellfish Res.* 30, 47–53. <https://doi.org/10.2983/035.030.0108>
- Grizel, H., Comps, M., Bonami, J.R., Cousserans, F., Duthoit, J.L., Pennec, M., 1974. Epizooty of the common oyster *Ostrea edulis*, Part 1. Study of the agent of digestive gland disease in *Ostrea edulis* Linne. *Sci. Pêche* 240–241, 1–30.
- Grizel, H., Héral, M., 1991. Introduction into France of the Japanese oyster (*Crassostrea gigas*). *J. du Cons. Int. pour l' Exploit. la Mer* 399–403.
- Guo, X., He, Y., Zhang, L., Lelong, C., Jouaux, A., 2015. Immune and stress responses in oysters with insights on adaptation. *Fish Shellfish Immunol.* 46, 107–119.

<https://doi.org/10.1016/j.fsi.2015.05.018>

- Haberkorn, H., Lambert, C., Le Goïc, N., Guéguen, M., Moal, J., Palacios, E., Lassus, P., Soudant, P., 2010a. Effects of *Alexandrium minutum* exposure upon physiological and hematological variables of diploid and triploid oysters, *Crassostrea gigas*. *Aquat. Toxicol.* 97, 96–108. <https://doi.org/10.1016/j.aquatox.2009.12.006>
- Haberkorn, H., Lambert, C., Le Goïc, N., Moal, J., Suquet, M., Guéguen, M., Sunila, I., Soudant, P., 2010b. Effects of *Alexandrium minutum* exposure on nutrition-related processes and reproductive output in oysters *Crassostrea gigas*. *Harmful Algae* 9, 427–439. <https://doi.org/10.1016/j.hal.2010.01.003>
- Haberkorn, H., Lambert, C., Le Goïc, N., Quéré, C., Bruneau, A., Riso, R., Auffret, M., Soudant, P., 2014. Cellular and biochemical responses of the oyster *Crassostrea gigas* to controlled exposures to metals and *Alexandrium minutum*. *Aquat. Toxicol.* 147, 158–167. <https://doi.org/10.1016/j.aquatox.2013.12.012>
- Haberkorn, H., Tran, D., Massabuau, J.-C., Ciret, P., Soudant, P., 2011. Relationship between valve activity, microalgae concentration in the water and toxin accumulation in the digestive gland of the Pacific oyster *Crassostrea gigas* exposed to *Alexandrium minutum*. *Mar. Pollut. Bull.* 62, 1191–1197.
- HABSOS, 2019. *Karenia brevis* [WWW Document]. Harmful Algal Bloom Obs. Syst. URL <https://service.ncddc.noaa.gov/website/AGSViewers/HABSOS/maps.htm> (accessed 7.18.19).
- Hallegraeff, G., Dorantes-Aranda, J.J., Mardones, J., Seger, A., 2017. Review of progress in our understanding of fish-killing microalgae: implications for management and mitigation. In: Proença, L. A. O. and Hallegraeff, G.M. (eds). *Marine and Fresh-Water Harmful Algae. Proceedings of the 17th International Conference on Harmful Algae. International Society for the Study of Harmful Algae and Intergovernmental Oceanographic Commission of UNESCO 2017.*
- Hallegraeff, G.M., 1993. A review of harmful algal blooms and their apparent global increase. *Phycologia* 32, 79–99. <https://doi.org/10.2216/i0031-8884-32-2-79.1>
- Hasui, M., Matsuda, M., Okutani, K., Shigeta, S., 1995. *In vitro* antiviral activities of sulfated polysaccharides from a marine microalga (*Cochlodinium polykrikoides*) against human immunodeficiency virus and other enveloped viruses. *Int. J. Biol. Macromol.* 17, 293–297. [https://doi.org/10.1016/0141-8130\(95\)98157-T](https://doi.org/10.1016/0141-8130(95)98157-T)
- Hégaret, H., Brokordt, K.B., Gaymer, C.F., Lohrmann, K.B., García, C., Varela, D., 2012. Effects of the toxic dinoflagellate *Alexandrium catenella* on histopathological and escape responses of the Northern scallop *Argopecten purpuratus*. *Harmful Algae* 18, 74–83. <https://doi.org/10.1016/j.hal.2012.04.006>
- Hégaret, H., da Silva, P., Wikfors, G., Lambert, C., Debettignies, T., Shumway, S., Soudant, P., 2007a. Hemocyte responses of Manila clams, *Ruditapes philippinarum*, with varying parasite, *Perkinsus olseni*, severity to toxic-algal exposures. *Aquat. Toxicol.* 84, 469–479. <https://doi.org/10.1016/j.aquatox.2007.07.007>
- Hégaret, H., da Silva, P.M., Sunila, I., Shumway, S.E., Dixon, M.S., Alix, J., Wikfors, G.H., Soudant, P., 2009. Perkinsosis in the Manila clam *Ruditapes philippinarum* affects

- responses to the harmful-alga, *Prorocentrum minimum*. J. Exp. Mar. Bio. Ecol. 371, 112–120. <https://doi.org/10.1016/j.jembe.2009.01.016>
- Hégaret, H., da Silva, P.M., Wikfors, G.H., Haberkorn, H., Shumway, S.E., Soudant, P., 2011. *In vitro* interactions between several species of harmful algae and haemocytes of bivalve molluscs. Cell Biol. Toxicol. 27, 249–266.
- Hégaret, H., Smolowitz, R.M., Sunila, I., Shumway, S.E., Alix, J., Dixon, M., Wikfors, G.H., 2010. Combined effects of a parasite, QPX, and the harmful-alga, *Prorocentrum minimum* on northern quahogs, *Mercenaria mercenaria*. Mar. Environ. Res. 69, 337–344. <https://doi.org/10.1016/j.marenvres.2009.12.008>
- Hégaret, H., Wikfors, G., Soudant, P., Lambert, C., Shumway, S.E., Bérard, J.B., Lassus, P., 2007b. Toxic dinoflagellates (*Alexandrium fundyense* and *A. catenella*) have minimal apparent effects on oyster hemocytes. Mar. Biol. 152, 441–447. <https://doi.org/10.1007/s00227-007-0703-3>
- Hégaret, H., Wikfors, G.H., 2005a. Effects of natural and field-simulated blooms of the dinoflagellate *Prorocentrum minimum* upon hemocytes of eastern oysters, *Crassostrea virginica*, from two different populations. Harmful Algae 4, 201–209. <https://doi.org/10.1016/j.hal.2003.12.005>
- Hégaret, H., Wikfors, G.H., 2005b. Time-dependent changes in hemocytes of eastern oysters, *Crassostrea virginica*, and northern bay scallops, *Argopecten irradians* irradians, exposed to a cultured strain of *Prorocentrum minimum*. Harmful Algae 4, 187–199. <https://doi.org/10.1016/j.hal.2003.12.004>
- Hégaret, H., Wikfors, G.H., Shumway, S.E., 2007c. Diverse Feeding Responses of Five Species of Bivalve Mollusc When Exposed To Three Species of Harmful Algae. J. Shellfish Res. 26, 549–559. [https://doi.org/10.2983/0730-8000\(2007\)26\[549:dfrofs\]2.0.co;2](https://doi.org/10.2983/0730-8000(2007)26[549:dfrofs]2.0.co;2)
- Hermabessiere, L., Fabioux, C., Lassudrie, M., Boullot, F., Long, M., Lambert, C., Le Goïc, N., Gouriou, J., Le Gac, M., Chapelle, A., Soudant, P., Hégaret, H., 2016. Influence of gametogenesis pattern and sex on paralytic shellfish toxin levels in triploid Pacific oyster *Crassostrea gigas* exposed to a natural bloom of *Alexandrium minutum*. Aquaculture. <https://doi.org/10.1016/j.aquaculture.2016.01.001>
- Hill, K., Stokes, N., Webb, S., Hine, P., Kroeck, M., Moore, J., Morley, M., Reece, K., Burrenson, E., Carnegie, R., 2014. Phylogenetics of *Bonamia* parasites based on small subunit and internal transcribed spacer region ribosomal DNA sequence data. Dis. Aquat. Organ. 110, 33–54. <https://doi.org/10.3354/dao02738>
- Hine, P., Cochenec-Laureau, N., Berthe, F., 2001. *Bonamia exitiosus* n.sp. (Haplosporidia) infecting flat oysters *Ostrea chilensis* in New Zealand. Dis. Aquat. Organ. 47, 63–72. <https://doi.org/10.3354/dao047063>
- Hughes, F.M., Foster, B., Grewal, S., Sokolova, I.M., 2010. Apoptosis as a host defense mechanism in *Crassostrea virginica* and its modulation by *Perkinsus marinus*. Fish Shellfish Immunol. 29, 247–257. <https://doi.org/10.1016/j.fsi.2010.03.003>
- Jales, M.C., Antônio, F., Koenig, M.L., 2013. The reef ecosystem of Serrambi (Pernambuco State, Brazil): phytoplankton composition. Arq. Ciências do Mar 46, 27–39.
- Jauffrais, T., Contreras, A., Herrenknecht, C., Truquet, P., Séchet, V., Tillmann, U., Hess, P.,

2012. Effect of *Azadinium spinosum* on the feeding behaviour and azaspiracid accumulation of *Mytilus edulis*. *Aquat. Toxicol.* 124–125, 179–187. <https://doi.org/10.1016/j.aquatox.2012.08.016>
- Jenkins, C., Hick, P., Gabor, M., Spiers, Z., Fell, S., Gu, X., Read, A., Go, J., Dove, M., O'Connor, W., Kirkland, P., Frances, J., 2013. Identification and characterisation of an ostreid herpesvirus-1 microvariant (OsHV-1 μ -var) in *Crassostrea gigas* (Pacific oysters) in Australia. *Dis. Aquat. Organ.* 105, 109–126. <https://doi.org/10.3354/dao02623>
- John U., Litaker R.W., Montresor M., Murray S., Brosnahan M.L., Anderson, D.M. 2014. Formal revision of the *Alexandrium tamarense* species complex (Dinophyceae) taxonomy: the introduction of five species with emphasis on molecular-based (rDNA) classification. *Protist* 165, 779–804. <https://doi.org/10.1016/j.protis.2014.10.001>
- Kellam, S.J., Walker, J.M., 1989. Antibacterial activity from marine microalgae in laboratory culture. *Br. Phycol. J.* 24, 191–194. <https://doi.org/10.1080/00071618900650181>
- Kiene, R.P., Linn, L.J., Bruton, J.A., 2000. New and important roles for DMSP in marine microbial communities. *J. Sea Res.* 43, 209–224. [https://doi.org/10.1016/S1385-1101\(00\)00023-X](https://doi.org/10.1016/S1385-1101(00)00023-X)
- Kleeman, S., Adlard, R., Lester, R.J., 2002. Detection of the initial infective stages of the protozoan parasite *Marteilia sydneyi* in *Saccostrea glomerata* and their development through to sporogenesis. *Int. J. Parasitol.* 32, 767–784. [https://doi.org/10.1016/S0020-7519\(02\)00025-5](https://doi.org/10.1016/S0020-7519(02)00025-5)
- Kokou, F., Makridis, P., Kentouri, M., Divanach, P., 2012. Antibacterial activity in microalgae cultures. *Aquac. Res.* 43, 1520–1527. <https://doi.org/10.1111/j.1365-2109.2011.02955.x>
- Lafont, M., Goncalves, P., Guo, X., Montagnani, C., Raftos, D., Green, T., 2019. Transgenerational plasticity and antiviral immunity in the Pacific oyster (*Crassostrea gigas*) against Ostreid herpesvirus 1 (OsHV-1). *Dev. Comp. Immunol.* 91, 17–25. <https://doi.org/10.1016/j.dci.2018.09.022>
- Lafont, M., Petton, B., Vergnes, A., Pauletto, M., Segarra, A., Gourbal, B., Montagnani, C., 2017. Long-lasting antiviral innate immune priming in the Lophotrochozoan Pacific oyster, *Crassostrea gigas*. *Sci. Rep.* 7, 13143. <https://doi.org/10.1038/s41598-017-13564-0>
- Landsberg, J.H.J.H., 2002. The Effects of Harmful Algal Blooms on Aquatic Organisms. *Rev. Fish. Sci.* 10, 113–390. <https://doi.org/10.1080/20026491051695>
- Lane, H., Webb, S., Duncan, J., 2016. *Bonamia ostreae* in the New Zealand oyster *Ostrea chilensis*: a new host and geographic record for this haplosporidian parasite. *Dis. Aquat. Organ.* 118, 55–63. <https://doi.org/10.3354/dao02960>
- Lassudrie, M., Soudant, P., Nicolas, J.-L., Fabioux, C., Lambert, C., Miner, P., Le Grand, J., Petton, B., Hégaret, H., 2015a. Interaction between toxic dinoflagellate *Alexandrium catenella* exposure and disease associated with herpesvirus OsHV-1 μ Var in Pacific oyster spat *Crassostrea gigas*. *Harmful Algae* 45, 53–61. <https://doi.org/10.1016/j.hal.2015.04.007>
- Lassudrie, M., Soudant, P., Nicolas, J.-L., Miner, P., Le Grand, J., Lambert, C., Le Goïc, N., Hégaret, H., Fabioux, C., 2016. Exposure to the toxic dinoflagellate *Alexandrium catenella*

- modulates juvenile oyster *Crassostrea gigas* hemocyte variables subjected to different biotic conditions. *Fish Shellfish Immunol.* 51, 104–115. <https://doi.org/10.1016/j.fsi.2016.02.017>
- Lassudrie, M., Soudant, P., Richard, G., Henry, N., Medhioub, W., da Silva, P.M., Donval, A., Bunel, M., Le Goïc, N., Lambert, C., de Montaudouin, X., Fabioux, C., Hégaret, H., 2014. Physiological responses of Manila clams *Venerupis (=Ruditapes) philippinarum* with varying parasite *Perkinsus olseni* burden to toxic algal *Alexandrium ostenfeldii* exposure. *Aquat. Toxicol.* 154, 27–38. <https://doi.org/10.1016/j.aquatox.2014.05.002>
- Lassudrie, M., Wikfors, G.H., Sunila, I., Alix, J.H., Dixon, M.S., Combot, D., Soudant, P., Fabioux, C., Hégaret, H., 2015b. Physiological and pathological changes in the eastern oyster *Crassostrea virginica* infested with the trematode *Bucephalus* sp. and exposed to the toxic dinoflagellate *Alexandrium fundyense*. *J. Invertebr. Pathol.* 126, 51–63. <https://doi.org/10.1016/j.jip.2015.01.011>
- Lassus, P., Amzil, Z., Baron, R., Séchet, V., Barillé, L., Abadie, E., Bardouil, M., Sibat, M., Truquet, P., Bérard, J., Gueguen, M., 2007. Modelling the accumulation of PSP toxins in Thau Lagoon oysters (*Crassostrea gigas*) from trials using mixed cultures of *Alexandrium catenella* and *Thalassiosira weissflogii*. *Aquat. Living Resour.* 67, 59–67.
- Lassus, P., Bardouil, M., Beliaeff, B., Masselin, P., Naviner, M., Truquet, P., 1999. Effect of continuous supply of the toxic dinoflagellate *Alexandrium minutum* Halim on the feeding behaviour of the Pacific oyster (*Crassostrea gigas* Thunberg). *J. Shellfish Res.* 18, 211–216.
- Lassus, P., Chomérat, N., Hess, P., Nézan, E., 2016. Toxic and harmful microalgae of the World Ocean. Denmark, International Society for the Study of Harmful Algae / Intergovernmental Oceanographic Commission of UNESCO. IOC Manuals and Guides, 68.
- Lau, Y.-T., Gambino, L., Santos, B., Pales Espinosa, E., Allam, B., 2018a. Transepithelial migration of mucosal hemocytes in *Crassostrea virginica* and potential role in *Perkinsus marinus* pathogenesis. *J. Invertebr. Pathol.* 153, 122–129. <https://doi.org/10.1016/j.jip.2018.03.004>
- Lau, Y.-T., Gambino, L., Santos, B., Pales Espinosa, E., Allam, B., 2018b. Regulation of oyster (*Crassostrea virginica*) hemocyte motility by the intracellular parasite *Perkinsus marinus*: A possible mechanism for host infection. *Fish Shellfish Immunol.* 78, 18–25. <https://doi.org/10.1016/j.fsi.2018.04.019>
- Lau, Y.-T., Santos, B., Barbosa, M., Pales Espinosa, E., Allam, B., 2018c. Regulation of apoptosis-related genes during interactions between oyster hemocytes and the alveolate parasite *Perkinsus marinus*. *Fish Shellfish Immunol.* 83, 180–189. <https://doi.org/10.1016/j.fsi.2018.09.006>
- Lauritano, C., Andersen, J.H., Hansen, E., Albrigtsen, M., Escalera, L., Esposito, F., Helland, K., Hanssen, K.Ø., Romano, G., Ianora, A., 2016. Bioactivity Screening of Microalgae for Antioxidant, Anti-Inflammatory, Anticancer, Anti-Diabetes, and Antibacterial Activities. *Front. Mar. Sci.* 3, 1–12. <https://doi.org/10.3389/fmars.2016.00068>
- Le Goïc, N., Hégaret, H., Boulais, M., Béguel, J.-P.P., Lambert, C., Fabioux, C., Soudant, P., 2014. Flow cytometric assessment of morphology, viability, and production of reactive oxygen species of *Crassostrea gigas* oocytes. Application to Toxic dinoflagellate

- (*Alexandrium minutum*) exposure. Cytom. Part A 85, 1049–1056. <https://doi.org/10.1002/cyto.a.22577>
- Le Goïc, N., Hégaret, H., Fabioux, C., Miner, P., Suquet, M., Lambert, C., Soudant, 2013. Impact of the toxic dinoflagellate *Alexandrium catenella* on Pacific oyster reproductive output: application of flow cytometry assays on spermatozoa. Aquat. Living Resources 26, 221–228.
- Lee, C.-K., Park, T.-G., Park, Y.-T., Lim, W.-A., 2013. Monitoring and trends in harmful algal blooms and red tides in Korean coastal waters, with emphasis on *Cochlodinium polykrikoides*. Harmful Algae 30, S3–S14. <https://doi.org/10.1016/j.hal.2013.10.002>
- Lefebvre, K. a, Bill, B.D., Erickson, A., Baugh, K. a, O'Rourke, L., Costa, P.R., Nance, S., Trainer, V.L., 2008. Characterization of intracellular and extracellular saxitoxin levels in both field and cultured *Alexandrium* spp. samples from Sequim Bay, Washington. Mar. Drugs 6, 103–116. <https://doi.org/10.3390/md20080006>
- Lelong, A., Haberkorn, H., Le Goïc, N., Hégaret, H., Soudant, P., 2011. A new insight into allelopathic effects of *Alexandrium minutum* on photosynthesis and respiration of the diatom *Chaetoceros neogracile* revealed by photosynthetic-performance analysis and flow cytometry. Microb. Ecol. 62, 919–930. <https://doi.org/10.1007/s00248-011-9889-5>
- Levicán, A., Lasa, A., Irgang, R., Romalde, J.L., Poblete-Morales, M., Avendaño-Herrera, R., 2017. Isolation of *Vibrio tapetis* from two native fish species (*Genypterus chilensis* and *Paralichthys adspersus*) reared in Chile and description of *Vibrio tapetis* subsp. *quintayensis* subsp. nov. Int. J. Syst. Evol. Microbiol. 67, 716–723. <https://doi.org/10.1099/ijsem.0.001705>
- Li, Y., Song, X., Wang, W., Wang, L., Yi, Q., Jiang, S., Jia, Z., Du, X., Qiu, L., Song, L., 2017. The hematopoiesis in gill and its role in the immune response of Pacific oyster *Crassostrea gigas* against secondary challenge with *Vibrio splendidus*. Dev. Comp. Immunol. 71, 59–69. <https://doi.org/10.1016/j.dci.2017.01.024>
- Litaker, W.; Sampedro, N., Calado, A., 2018. A practical guide to new nomenclature for species within the “*Alexandrium tamarense* species complex”, Harmful algae news 61: 13-15
- Long, M., Tallec, K., Soudant, P., Lambert, C., Le Grand, F., Sarthou, G., Jolley, D., Hégaret, H., 2018a. A rapid quantitative fluorescence-based bioassay to study allelochemical interactions from *Alexandrium minutum*. Environ. Pollut. 242, 1598–1605. <https://doi.org/10.1016/j.envpol.2018.07.119>
- Long, M., Tallec, K., Soudant, P., Le Grand, F., Donval, A., Lambert, C., Sarthou, G., Jolley, D.F., Hégaret, H., 2018b. Allelochemicals from *Alexandrium minutum* induce rapid inhibition of metabolism and modify the membranes from *Chaetoceros muelleri*. Algal Res. 35, 508–518. <https://doi.org/10.1016/j.algal.2018.09.023>
- Ma, H., Krock, B., Tillmann, U., Cembella, A., 2009. Preliminary Characterization of Extracellular Allelochemicals of the Toxic Marine Dinoflagellate *Alexandrium tamarense* Using a *Rhodomonas salina* Bioassay. Mar. Drugs 7, 497–522. <https://doi.org/10.3390/md7040497>
- Ma, H., Krock, B., Tillmann, U., Muck, A., Wielsch, N., Svatoš, A., Cembella, A., 2011. Isolation of activity and partial characterization of large non-proteinaceous lytic

- allelochemicals produced by the marine dinoflagellate *Alexandrium tamarense*. Harmful Algae 11, 65–72. <https://doi.org/10.1016/j.hal.2011.07.004>
- Machado, R.C.A., Feitosa, F.A.N., Koenig, M.L., Flores Montes, M.J., 2018. Spatial and seasonal variation of the phytoplankton community structure in a reef ecosystem in North-eastern Brazil. J. Mar. Biol. Assoc. United Kingdom 98, 557–566. <https://doi.org/10.1017/S0025315416001600>
- Mackin, J.G., Owen, H.M., 1950. Preliminary note on the occurrence of a new protistan parasite, *Dermocystidium marinum*. J. Perinat. Med. 111, 328–329. <https://doi.org/10.1515/jpme.1982.10.5.249>
- Mafra, L.L., Fernandes, L.F., Proença, L.A.O., 2006. Harmful algae and toxins in Paranaguá bay, Brazil: bases for monitoring. Brazilian J. Oceanogr. 54, 107–121. <https://doi.org/10.1590/S1679-87592006000200002>
- Mafra, L.L., Ribas, T., Alves, T.P., Proença, L.A.O., Schramm, M.A., Uchida, H., Suzuki, T., 2015. Differential okadaic acid accumulation and detoxification by oysters and mussels during natural and simulated *Dinophysis* blooms. Fish. Sci. 81, 749–762. <https://doi.org/10.1007/s12562-015-0882-7>
- Marteil, L., 1976. La conchyliculture française. 2eme partie. Biologie de l’huître et de la moule. Rev. des Trav. l’Institut des Pêches Marit. 40, 149–346.
- Medhioub, W., Lassus, P., Truquet, P., Bardouil, M., Amzil, Z., Sechet, V., Sibat, M., Soudant, P., 2012. Spirolide uptake and detoxification by *Crassostrea gigas* exposed to the toxic dinoflagellate *Alexandrium ostenfeldii*. Aquaculture 358–359, 108–115. <https://doi.org/10.1016/j.aquaculture.2012.06.023>
- Medhioub, W., Ramondenc, S., Vanhove, A.S., Vergnes, A., Masseret, E., Savar, V., Amzil, Z., Laabir, M., Rolland, J.L., 2013. Exposure to the neurotoxic dinoflagellate, *Alexandrium catenella*, induces apoptosis of the hemocytes of the oyster, *Crassostrea gigas*. Mar. Drugs 11, 4799–4814. <https://doi.org/10.3390/md11124799>
- Mello, D.F., da Silva, P.M., Barracco, M.A., Soudant, P., Hégaret, H., 2013. Effects of the dinoflagellate *Alexandrium minutum* and its toxin (saxitoxin) on the functional activity and gene expression of *Crassostrea gigas* hemocytes. Harmful Algae 26, 45–51. <https://doi.org/10.1016/j.hal.2013.03.003>
- Mello, D.F., Danielli, N.M., Curbani, F., Pontinha, V.A., Suhnel, S., Castro, M.A.M., Medeiros, S.C., Wendt, N.C., Trevisan, R., Magalhães, A.R.M., Dafre, A.L., 2018. First evidence of viral and bacterial oyster pathogens in the Brazilian coast. J. Fish Dis. 41, 559–563. <https://doi.org/10.1111/jfd.12755>
- Mello, D.F., de Oliveira, E.S., Vieira, R.C., Simoes, E., Trevisan, R., Dafre, A.L., Barracco, M.A., 2012. Cellular and Transcriptional Responses of *Crassostrea gigas* Hemocytes Exposed *in Vitro* to Brevetoxin (PbTx-2). Mar. Drugs 10, 583–597. <https://doi.org/10.3390/md10030583>
- Mello, D.F., Proença, L.A. de O., Barracco, M.A., 2010. Comparative Study of Various Immune Parameters in Three Bivalve Species during a Natural Bloom of *Dinophysis acuminata* in Santa Catarina Island, Brazil. Toxins (Basel). 2, 1166–1178. <https://doi.org/10.3390/toxins2051166>

- Menezes, M., Branco, S., Miotto, M.C., Alves-de-Souza, C., 2018. The Genus *Alexandrium* (Dinophyceae, Dinophyta) in Brazilian Coastal Waters. *Front. Mar. Sci.* 5, 1–13. <https://doi.org/10.3389/fmars.2018.00421>
- Mitra, A., Flynn, K.J., Burkholder, J.M., Berge, T., Calbet, A., Raven, J.A., Granéli, E., Glibert, P.M., Hansen, P.J., Stoecker, D.K., Thingstad, F., Tillmann, U., Våge, S., Wilken, S., Zubkov, M. V., 2014. The role of mixotrophic protists in the biological carbon pump. *Biogeosciences* 11, 995–1005. <https://doi.org/10.5194/bg-11-995-2014>
- Moreira-González, A.R., Fernandes, L.F., Uchida, H., Uesugi, A., Suzuki, T., Chomérat, N., Bilien, G., Mafra, L.L., 2019. Variations in morphology, growth, and toxicity among strains of the *Prorocentrum lima* species complex isolated from Cuba and Brazil. *J. Appl. Phycol.* 31, 519–532. <https://doi.org/10.1007/s10811-018-1587-y>
- Mortensen, S., Strand, Å., Bodvin, T., Alfjorden, A., Skår, C., Jelmert, A., Aspán, A., Sælemyr, L., Naustvoll, L., Albretsen, J., 2016. Summer mortalities and detection of ostreid herpesvirus microvariant in Pacific oyster *Crassostrea gigas* in Sweden and Norway. *Dis. Aquat. Organ.* 117, 171–176. <https://doi.org/10.3354/dao02944>
- Nam, K.-W., Jeung, H.-D., Song, J.-H., Park, K.-H., Choi, K.-S., Park, K.-I., 2018. High parasite burden increases the surfacing and mortality of the Manila clam (*Ruditapes philippinarum*) in intertidal sandy mudflats on the west coast of Korea during hot summer. *Parasit. Vectors* 11, 42. <https://doi.org/10.1186/s13071-018-2620-3>
- Nascimento, S.M., Corrêa, E. V., Menezes, M., Varela, D., Paredes, J., Morris, S., 2012. Growth and toxin profile of *Ostreopsis* cf. *ovata* (Dinophyta) from Rio de Janeiro, Brazil. *Harmful Algae* 13, 1–9. <https://doi.org/10.1016/j.hal.2011.09.008>
- Navarro, J.M., González, K., Cisternas, B., López, J. a, Chaparro, O.R., Segura, C.J., Córdova, M., Suárez-Isla, B., Fernandez-Reiriz, M.J., Labarta, U., 2014. Contrasting physiological responses of two populations of the razor clam *Tagelus dombeii* with different histories of exposure to Paralytic Shellfish Poisoning (PSP). *PLoS One* 9, e105794. <https://doi.org/10.1371/journal.pone.0105794>
- Nell, J., 2001. The history of oyster farming in Australia. *Mar. Fish. Rev.* 63, 14–25.
- Neves, R.A.F., Santiago, T.C., Carvalho, W.F., Silva, E. dos S., da Silva, P.M., Nascimento, S.M., 2019. Impacts of the toxic benthic dinoflagellate *Prorocentrum lima* on the brown mussel *Perna perna*: Shell-valve closure response, immunology, and histopathology. *Mar. Environ. Res.* 146, 35–45. <https://doi.org/10.1016/j.marenvres.2019.03.006>
- Núñez-Vázquez, E.J., Gárate-Lizarraga, I., Band-Schmidt, C.J., Cordero-Tapia, A., Lopez-Cortes, D.J., Sandoval, F.E.H., Heredia-Tapia, A., Bustillos-Guzman, J.J., 2011. Impact of Harmful Algal Blooms on wild and cultured animals in the Gulf of California. *J. Environ. Biol.* 32, 413–423.
- Oehrens Kissner, E., Doldan, M., Zaidman, P., Morsan, E., Kroeck, M., 2014. Bonamiosis status in natural *Ostrea puelchana* beds in San Matías Gulf (Patagonia, Argentina), 14 years after an epizootic. *Dis. Aquat. Organ.* 110, 135–142. <https://doi.org/10.3354/dao02707>
- Paillard, C., 2004. A short-review of brown ring disease, a vibriosis affecting clams, *Ruditapes philippinarum* and *Ruditapes decussatus*. *Aquat. Living Resour.* 17, 467–475.

<https://doi.org/10.1051/alr:2004053>

- Panel, E., Health, A., 2015. Oyster mortality. *EFSA J.* 13, 4122. <https://doi.org/10.2903/j.efsa.2015.4122>
- Park, K.-I., Paillard, C., Le Chevalier, P., Choi, K.-S., 2006. Report on the occurrence of brown ring disease (BRD) in Manila clam, *Ruditapes philippinarum*, on the west coast of Korea. *Aquaculture* 255, 610–613. <https://doi.org/10.1016/j.aquaculture.2005.12.011>
- Park, K.-I., Tsutsumi, H., Hong, J.-S., Choi, K.-S., 2008. Pathology survey of the short-neck clam *Ruditapes philippinarum* occurring on sandy tidal flats along the coast of Ariake Bay, Kyushu, Japan. *J. Invertebr. Pathol.* 99, 212–219. <https://doi.org/10.1016/j.jip.2008.06.004>
- Paul-Pont, I., Dhand, N.K., Whittington, R.J., 2013. Spatial distribution of mortality in Pacific oysters *Crassostrea gigas*: reflection on mechanisms of OsHV-1 transmission. *Dis. Aquat. Organ.* 105, 127–138. <https://doi.org/10.3354/dao02615>
- Paul-Pont, I., Evans, O., Dhand, N.K., Rubio, A., Coad, P., Whittington, R.J., 2014. Descriptive epidemiology of mass mortality due to Ostreid herpesvirus-1 (OsHV-1) in commercially farmed Pacific oysters (*Crassostrea gigas*) in the Hawkesbury River estuary, Australia. *Aquaculture* 422–423, 146–159. <https://doi.org/10.1016/j.aquaculture.2013.12.009>
- Perkins, F.O., Wolf, P.H., 1976. Fine structure of *Marteilia sydneyi* sp. n.- haplosporidan pathogen of Australian oysters. *J. Parasitol.* 62, 528–538.
- Persson, A., Smith, B.C., Alix, J.H., Senft-Batoh, C., Wikfors, G.H., 2012. Toxin content differs between life stages of *Alexandrium fundyense* (Dinophyceae). *Harmful Algae* 19, 101–107. <https://doi.org/10.1016/j.hal.2012.06.006>
- Petton, B., Bruto, M., James, A., Labreuche, Y., Alunno-Bruscia, M., Le Roux, F., 2015. *Crassostrea gigas* mortality in France: the usual suspect, a herpes virus, may not be the killer in this polymicrobial opportunistic disease. *Front. Microbiol.* 6, 1–10. <https://doi.org/10.3389/fmicb.2015.00686>
- Pichot, Y., Comps, M., Tige, G., Grizel, H., Rabouin, M., 1980. Recherches sur *Bonamia ostreae*, gen. n. sp. n., parasite nouveau de l’huître plate *Ostrea edulis* L. *Rev. des Trav. l’Institut des Pêches Marit.* 43, 131–140.
- Pomati, F., Rossetti, C., Calamari, D., Neilan, B.A., 2003. Effects of Saxitoxin (STX) and Veratridine on bacterial Na⁺- K⁺ fluxes: a Prokaryote-based STX bioassay. *Appl. Environ. Microbiol.* 69, 7371–7376. <https://doi.org/10.1128/AEM.69.12.7371>
- Pousse, É., Flye-Sainte-Marie, J., Alunno-Bruscia, M., Hégaret, H., Jean, F., 2018. Sources of paralytic shellfish toxin accumulation variability in the Pacific oyster *Crassostrea gigas*. *Toxicon* 144, 14–22. <https://doi.org/10.1016/j.toxicon.2017.12.050>
- Prado-Alvarez, M., Flórez-Barrós, F., Méndez, J., Fernandez-Tajes, J., 2013. Effect of okadaic acid on carpet shell clam (*Ruditapes decussatus*) haemocytes by *in vitro* exposure and harmful algal bloom simulation assays. *Cell Biol. Toxicol.* 29, 189–197. <https://doi.org/10.1007/s10565-013-9246-1>
- Prado-Alvarez, M., Flórez-Barrós, F., Sexto-Iglesias, A., Méndez, J., Fernandez-Tajes, J., 2012. Effects of okadaic acid on haemocytes from *Mytilus galloprovincialis*: A comparison

- between field and laboratory studies. *Mar. Environ. Res.* 81, 90–93. <https://doi.org/10.1016/j.marenvres.2012.08.011>
- Prego-Faraldo, M., Valdíglesias, V., Laffon, B., Mendez, J., Eirin-Lopez, J., 2016. Early Genotoxic and Cytotoxic Effects of the Toxic Dinoflagellate *Prorocentrum lima* in the Mussel *Mytilus galloprovincialis*. *Toxins (Basel)*. 8, 159. <https://doi.org/10.3390/toxins8060159>
- Prego-Faraldo, M.V., Valdíglesias, V., Laffon, B., Eirín-López, J.M., Méndez, J., 2015. In Vitro Analysis of Early Genotoxic and Cytotoxic Effects of Okadaic Acid in Different Cell Types of the Mussel *Mytilus galloprovincialis*. *J. Toxicol. Environ. Heal. Part A* 78, 814–824. <https://doi.org/10.1080/15287394.2015.1051173>
- Proença, L., Tamanaha, M., Souza, N., 2001. The toxic dinoflagellate *Gymnodinium catenatum* Graham in southern Brazilian waters: Occurrence, pigments and toxins. *Atlântica* 23, 59–65.
- Proença, L.A.O., Schramm, M.A., M.S.Tamanaha, Alves, T.P., 2007. Diarrhoetic shellfish poisoning (DSP) outbreak in Subtropical Southwest Atlantic. *Harmful Algae News* 19–20.
- Queiroga, F.R., Marques-Santos, L.F., Hégaret, H., Soudant, P., Farias, N.D., Schlindwein, A.D., da Silva, P.M., 2013. Immunological responses of the mangrove oysters *Crassostrea gasar* naturally infected by *Perkinsus* sp. in the Mamanguape Estuary, Paraíba state (Northeastern, Brazil). *Fish Shellfish Immunol.* 35, 319–327. <https://doi.org/10.1016/j.fsi.2013.04.034>
- Queiroga, F.R., Vianna, R.T., Vieira, C.B., Farias, N.D., Da Silva, P.M., 2015. Parasites infecting the cultured oyster *Crassostrea gasar* (Adanson, 1757) in Northeast Brazil. *Parasitology* 142, 756–766. <https://doi.org/10.1017/S0031182014001863>
- Ramilo, A., González, M., Carballal, M., Darriba, S., Abollo, E., Villalba, A., 2014. Oyster parasites *Bonamia ostreae* and *B. exitiosa* co-occur in Galicia (NW Spain): spatial distribution and infection dynamics. *Dis. Aquat. Organ.* 110, 123–133. <https://doi.org/10.3354/dao02673>
- Rasmussen, S.A., Andersen, A.J.C., Andersen, N.G., Nielsen, K.F., Hansen, P.J., Larsen, T.O., 2016. Chemical Diversity, Origin, and Analysis of Phycotoxins. *J. Nat. Prod.* 79, 662–673. <https://doi.org/10.1021/acs.jnatprod.5b01066>
- Rasmussen, S.A., Binzer, S.B., Hoeck, C., Meier, S., de Medeiros, L.S., Andersen, N.G., Place, A., Nielsen, K.F., Hansen, P.J., Larsen, T.O., 2017. Karmitoxin: An Amine-Containing Polyhydroxy-Polyene Toxin from the Marine Dinoflagellate *Karlodinium armiger*. *J. Nat. Prod.* 80, 1287–1293. <https://doi.org/10.1021/acs.jnatprod.6b00860>
- Reid, H.I., Duncan, H.L., Laidler, L.A., Hunter, D., Birkbeck, T.H., 2003. Isolation of *Vibrio tapetis* from cultivated Atlantic halibut (*Hippoglossus hippoglossus* L.). *Aquaculture* 221, 65–74. [https://doi.org/10.1016/S0044-8486\(03\)00060-7](https://doi.org/10.1016/S0044-8486(03)00060-7)
- Renault, T., 1996. Appearance and spread of diseases among bivalve molluscs in the northern hemisphere in relation to international trade. *Rev. Sci. Tech. l'Office Int. des Epizoot.* 15, 551–561.
- Rhodes, L., Smith, K.F., 2019. A checklist of the benthic and epiphytic marine dinoflagellates

- of New Zealand, including Rangitāhua/Kermadec Islands. *New Zeal. J. Mar. Freshw. Res.* 53, 258–277. <https://doi.org/10.1080/00288330.2018.1492425>
- Rhodes, L.L., Smith, K.F., MacKenzie, L., Moisan, C., 2019. Checklist of the planktonic marine dinoflagellates of New Zealand. *New Zeal. J. Mar. Freshw. Res.* 0, 1–16. <https://doi.org/10.1080/00288330.2019.1626746>
- Rivera, I.N.G., Souza, K.M.C., Souza, C.P., Lopes, R.M., 2013. Free-Living and Plankton-Associated Vibrios: Assessment in Ballast Water, Harbor Areas, and Coastal Ecosystems in Brazil. *Front. Microbiol.* 3, 1–8. <https://doi.org/10.3389/fmicb.2012.00443>
- Robledo, J.A.F., Vasta, G.R., Record, N.R., 2014. Protozoan Parasites of Bivalve Molluscs: Literature Follows Culture. *PLoS One* 9, e100872. <https://doi.org/10.1371/journal.pone.0100872>
- Rolton, A., Soudant, P., Vignier, J., Pierce, R., Henry, M., Shumway, S.E., Bricelj, V.M., Volety, A.K., 2015. Susceptibility of gametes and embryos of the eastern oyster, *Crassostrea virginica*, to *Karenia brevis* and its toxins. *Toxicon* 99, 6–15. <https://doi.org/10.1016/j.toxicon.2015.03.002>
- Ruano, F., Batista, F.M., Arcangeli, G., 2015. Perkinsosis in the clams *Ruditapes decussatus* and *R. philippinarum* in the Northeastern Atlantic and Mediterranean Sea: A review. *J. Invertebr. Pathol.* 131, 58–67. <https://doi.org/10.1016/j.jip.2015.07.015>
- Samain, J., 2011. Review and perspectives of physiological mechanisms underlying genetically-based resistance of the Pacific oyster *Crassostrea gigas* to summer mortality. *Aquat. Living Resour.* 24, 227–236. <https://doi.org/10.1051/alr/2011144>
- Samain, J.F., McCombie, H., 2007. Mortalités estivales de l’huitre creuse *Crassostrea gigas*: Défi Morest, Editions Q. ed.
- Santiago, M.F., Silva-Cunha, M. da G.G. da, Neumann-Leitão, S., Costa, K.M.P. da, Palmeira, G.C.B., Porto Neto, F. de F., Nunes, F.S., 2010. Phytoplankton dynamics in a highly eutrophic estuary in tropical Brazil. *Brazilian J. Oceanogr.* 58, 189–205. <https://doi.org/10.1590/S1679-87592010000300002>
- Scardua, M., Vianna, R., Duarte, S., Farias, N., Correia, M., dos Santos, H., da Silva, P., 2017. Growth, mortality and susceptibility of oyster *Crassostrea* spp. to *Perkinsus* spp. infection during on-growing in northeast Brazil. *Rev. Bras. Parasitol. Veterinária* 26, 401–410. <https://doi.org/10.1590/s1984-29612017061>
- Schmitt, P., Lorgeril, J. de, Gueguen, Y., Destoumieux-Garzón, D., Bachère, E., 2012a. Expression, tissue localization and synergy of antimicrobial peptides and proteins in the immune response of the oyster *Crassostrea gigas*. *Dev. Comp. Immunol.* 37, 363–370. <https://doi.org/10.1016/j.dci.2012.01.004>
- Schmitt, P., Rosa, R.D., Destoumieux-Garzón, D., 2016. An intimate link between antimicrobial peptide sequence diversity and binding to essential components of bacterial membranes. *Biochim. Biophys. Acta - Biomembr.* 1858, 958–970. <https://doi.org/10.1016/j.bbamem.2015.10.011>
- Schmitt, P., Rosa, R.D., Duperthuy, M., de Lorgeril, J., Bachère, E., Destoumieux-Garzón, D., 2012b. The antimicrobial defense of the Pacific oyster, *Crassostrea gigas*. How diversity may compensate for scarcity in the regulation of resident/pathogenic microflora. *Front.*

Microbiol. 3, 1–17. <https://doi.org/10.3389/fmicb.2012.00160>

- Segarra, A., Pépin, J.F., Arzul, I., Morga, B., Faury, N., Renault, T., 2010. Detection and description of a particular Ostreid herpesvirus 1 genotype associated with massive mortality outbreaks of Pacific oysters, *Crassostrea gigas*, in France in 2008. *Virus Res.* 153, 92–99.
- Sellner, K.G., Doucette, G.J., Kirkpatrick, G.J., 2003. Harmful algal blooms: causes, impacts and detection. *J. Ind. Microbiol. Biotechnol.* 30, 383–406. <https://doi.org/10.1007/s10295-003-0074-9>
- Seymour, J.R., Amin, S.A., Raina, J.-B., Stocker, R., 2017. Zooming in on the phycosphere: the ecological interface for phytoplankton–bacteria relationships. *Nat. Microbiol.* 2, 17065. <https://doi.org/10.1038/nmicrobiol.2017.65>
- Shumway, S.E., 1990. A review of the effects of algal blooms on shellfish and aquaculture. *J. World Aquac. Soc.* 21, 65–104.
- Simó, R., 2001. Production of atmospheric sulfur by oceanic plankton: biogeochemical, ecological and evolutionary links. *Trends Ecol. Evol.* 16, 287–294. [https://doi.org/10.1016/S0169-5347\(01\)02152-8](https://doi.org/10.1016/S0169-5347(01)02152-8)
- Simões, E., Vieira, R.C., Schramm, M.A., Mello, D.F., De Almeida Pontinha, V., da Silva, P.M., Barracco, M.A., 2015. Impact of harmful algal blooms (*Dinophysis acuminata*) on the immune system of oysters and mussels from Santa Catarina, Brazil. *J. Mar. Biol. Assoc. United Kingdom* 95, 773–781. <https://doi.org/10.1017/S0025315414001702>
- Sokolova, I.M., 2009. Apoptosis in molluscan immune defense. *Invertebr. Surviv. J.* 6, 49–58.
- Song, L., Wang, L., Qiu, L., Zhang, H., 2010. Bivalve Immunity, in: Söderhäll, K. (Ed.), *Invertebrate Immunity*. Springer Science+Business Media, LLC. Landes Bioscience, USA, NY, USA, pp. 44–65.
- Spiers, Z., Gabor, M., Fell, S., Carnegie, R., Dove, M., O'Connor, W., Frances, J., Go, J., Marsh, I., Jenkins, C., 2014. Longitudinal study of winter mortality disease in Sydney rock oysters *Saccostrea glomerata*. *Dis. Aquat. Organ.* 110, 151–164. <https://doi.org/10.3354/dao02629>
- Tavares, J., Proença, L.A.O., Odebrecht, C., 2009. Assessing the harmful microalgae occurrence and temporal variation in a coastal aquaculture area, southern Brazil. *Harmful Microalgae A Mar. Aquac. Area* 31, 129–144.
- Tibiriçá, C.E.J.A., Fernandes, L.F., Mafra Jr, L.L., 2015. Seasonal and spatial patterns of toxigenic species of *Dinophysis* and *Pseudo-nitzschia* in a subtropical Brazilian estuary. *Brazilian J. Oceanogr.* 63, 17–32. <https://doi.org/10.1590/S1679-87592015071906301>
- Tillmann, U., 2003. Kill and eat your predator: a winning strategy of the planktonic flagellate *Prymnesium parvum*. *Aquat. Microb. Ecol.* 32, 73–84. <https://doi.org/10.3354/ame032073>
- Tillmann, U., Alpermann, T., John, U., Cembella, A., 2008. Allelochemical interactions and short-term effects of the dinoflagellate *Alexandrium* on selected photoautotrophic and heterotrophic protists. *Harmful Algae* 7, 52–64. <https://doi.org/10.1016/j.hal.2007.05.009>
- Tillmann, U., John, U., 2002. Toxic effects of *Alexandrium* spp. on heterotrophic

- dinoflagellates: an allelochemical defence mechanism independent of PSP-toxin content. *Mar. Ecol. Prog. Ser.* 230, 47–58. <https://doi.org/10.3354/meps230047>
- Tillmann, U., John, U., Cembella, A., 2007. On the allelochemical potency of the marine dinoflagellate *Alexandrium ostenfeldii* against heterotrophic and autotrophic protists. *J. Plankton Res.* 29, 527–543. <https://doi.org/10.1093/plankt/fbm034>
- Travers, M.-A., Boettcher Miller, K., Roque, A., Friedman, C.S., 2015. Bacterial diseases in marine bivalves. *J. Invertebr. Pathol.* 131, 11–31. <https://doi.org/10.1016/j.jip.2015.07.010>
- Tran, D., Haberkorn, H., Soudant, P., Ciret, P., Massabuau, J.-C., 2010. Behavioral responses of *Crassostrea gigas* exposed to the harmful algae *Alexandrium minutum*. *Aquaculture* 298, 338–345. <https://doi.org/10.1016/j.aquaculture.2009.10.030>
- Turner, L.M., Alsterberg, C., Turner, A.D., Girisha, S.K., Rai, A., Havenhand, J.N., Venugopal, M.N., Karunasagar, I., Godhe, A., 2016. Pathogenic marine microbes influence the effects of climate change on a commercially important tropical bivalve. *Sci. Rep.* 6, 32413. <https://doi.org/10.1038/srep32413>
- Vargas-Albores, F., Barracco, M.A., 2001. Defense mechanisms in bivalve mollusks, with emphasis in pectinids, in: Maeda-Martínez, A. (Ed.), *Los Moluscos Pectínidos de Iberoamérica: Ciencia y Acuicultura*. pp. 127–146.
- Villalba, A., Gestal, C., Casas, S.S.M., Figueras, A., 2011. Perkinsosis en moluscos, in: Figueras, A., Novoa, B. (Eds.), *Enfermedades de Moluscos Bivalves de Interés En Acuicultura*. Publicaciones Científicas y Tecnológicas de la Fundación Observatorio Español de Acuicultura, Madrid, pp. 183–242.
- Waki, T., Takahashi, M., Eki, T., Hiasa, M., Umeda, K., Karakawa, N., Yoshinaga, T., 2018. Impact of *Perkinsus olseni* infection on a wild population of Manila clam *Ruditapes philippinarum* in Ariake Bay, Japan. *J. Invertebr. Pathol.* 153, 134–144. <https://doi.org/10.1016/j.jip.2018.03.001>
- Wells, M.L., Trainer, V.L., Smayda, T.J., Karlson, B.S.O., Trick, C.G., Kudela, R.M., Ishikawa, A., Bernard, S., Wulff, A., Anderson, D.M., Cochlan, W.P., 2015. Harmful algal blooms and climate change: Learning from the past and present to forecast the future. *Harmful Algae* 49, 68–93. <https://doi.org/10.1016/j.hal.2015.07.009>
- Yim, J.H., Kim, S.J., Ahn, S.H., Lee, C.K., Rhie, K.T., Lee, H.K., 2004. Antiviral Effects of Sulfated Exopolysaccharide from the Marine Microalga *Gyrodinium impudicum* Strain KG03. *Mar. Biotechnol.* 6, 17–25. <https://doi.org/10.1007/s10126-003-0002-z>
- Zannella, C., Mosca, F., Mariani, F., Franci, G., Folliero, V., Galdiero, Marilena, Tiscar, P.G., Galdiero, Massimiliano, 2017. Microbial Diseases of Bivalve Mollusks: Infections, Immunology and Antimicrobial Defense. *Mar. Drugs* 15, 182. <https://doi.org/10.3390/md15060182>
- Zhang, J., Li, Y., Ren, W., Cai, Y., 2010. Studies on phytoplanktons carrying and spreading AVNV. *J. Fish. Chine* 34, 1254–1259.
- Zhou, J., Richlen, M.L., Sehein, T.R., Kulis, D.M., Anderson, D.M., Cai, Z., 2018. Microbial Community Structure and Associations During a Marine Dinoflagellate Bloom. *Front. Microbiol.* 9, 1–21. <https://doi.org/10.3389/fmicb.2018.01201>

Table 1: Main bivalve diseases (mainly OIE notifiable diseases), associated pathogen, distribution and temporal dynamics, main marine HAB genera hypothetically co-occurring with disease (HABs reported to occur in regions and seasons where and when disease also occur, suggesting that both HABs and disease co-occur), and marine HAB species reported to co-occur with disease outbreaks in the field.

Bivalve species	Disease	Pathogen	Effect	Distribution	Seasonal dynamic	Reference for disease reports	Hypothetical co-occurrence of main HAB genera *	Reported co-occurrence of HABs with disease outbreak
<i>Crassostrea gigas</i> (juvenile-spat)	Pacific Oyster Mortality Syndrome (POMS)	OsHV-1 microvariants (virus) and <i>Vibrio</i> spp. (bacteria)	Lethal	Southern and Western Europe	Summer to fall	(Barbosa Solomieu et al., 2015)	<i>Alexandrium</i> , <i>Pseudo-nitzschia</i> , <i>Dinophysis</i> , <i>Ostreopsis</i> , <i>Azadinium</i> , <i>Vulcanodinium</i> , <i>Protoceratium</i> , <i>Lingulodinium</i> , <i>Gonyaulax</i> , <i>Karlodinium</i>	<i>Alexandrium</i> spp. (Abi-Khalil et al., 2016)
<i>Crassostrea gigas</i> (juvenile-spat)	Pacific Oyster Mortality Syndrome (POMS)	OsHV-1 microvariants (virus) and <i>Vibrio</i> spp. (bacteria)	Lethal	Australia (Southern and Eastern coast), New-Zealand (Northern coast)	Summer to fall	(Barbosa Solomieu et al., 2015)	<i>Alexandrium</i> , <i>Pseudo-nitzschia</i> , <i>Ostreopsis</i> , <i>Gambierdiscus</i> , <i>Karenia</i> , <i>Karlodinium</i> , <i>Protoceratium</i> , <i>Gymnodinium</i> , <i>Dinophysis</i> , <i>Prorocentrum</i> , <i>Amphidinium</i> , <i>Vulcanodinium</i> (Ajani et al., 2017, 2013, 2016; Rhodes and Smith, 2019; Rhodes et al., 2019)	<i>Alexandrium</i> spp. and <i>Pseudo-nitzschia</i> spp. (Jenkins et al., 2013)
<i>Crassostrea gigas</i> (juvenile-spat)	Pacific Oyster Mortality Syndrome (POMS)	OsHV-1 microvariants (virus) and <i>Vibrio</i> spp. (bacteria)	Non-lethal	Brazil (South coast)	Unknown	(Mello et al., 2018)	<i>Dinophysis</i> spp., <i>Gymnodinium</i> , <i>Pseudo-nitzschia</i> , <i>Alexandrium</i> , <i>Prorocentrum</i> , <i>Chatonella</i> , <i>Heterosigma</i> , <i>Fibrocapsa</i> (Branco et al., 2019; Cuellar-Martinez et al., 2018; Mafra et al., 2015, 2006; Menezes et al., 2018; Moreira-González et al., 2019; Proença et al., 2001, 2007; Simões et al., 2015; Tavares et al., 2009; Tibiriçá et al., 2015)	NA
<i>Crassostrea gigas</i>	Perkinsosis	<i>Perkinsus marinus</i> (protozoan parasite)	Lethal	Mexico (Gulf of California)	Unknown	(Enríquez-Espinoza et al., 2010)	<i>Margalefidinium</i> , <i>Pseudo-nitzschia</i> , <i>Prorocentrum</i> , <i>Gymnodinium</i> , <i>Chatonella</i> , <i>Akashiwo</i> , <i>Heterosigma</i> (Núñez-Vázquez et al., 2011)	NA
<i>Crassostrea virginica</i>	Dermo / Perkinsosis	<i>Perkinsus marinus</i> (protozoan parasite)	Lethal	USA (East coast)	Maximum in summer and autumn	(Burreson and Ragone-Calvo, 1996; Ford and Smolowitz, 2007)	<i>Alexandrium</i> , <i>Karenia</i> , <i>Margalefidinium</i> , <i>Karlodinium</i> , <i>Pseudo-nitzschia</i> , <i>Prorocentrum</i> , <i>Prymnesium</i> , <i>Dinophysis</i>	NA
<i>Crassostrea corteziensis</i>	Perkinsosis	<i>Perkinsus marinus</i> (protozoan parasite)	Non-lethal	Mexico (Pacific coast)	Maximum in summer	(Cáceres-Martínez et al., 2016)	<i>Margalefidinium</i> , <i>Pseudo-nitzschia</i> , <i>Prorocentrum</i> , <i>Pyrodinium</i> , <i>Dinophysis</i> , <i>Gambierdiscus</i>	NA

<i>Crassostrea gasar</i>	Perkinsosis	<i>Perkinsus marinus</i> and <i>P. beihaiensis</i> (protozoan parasites)	Sub-lethal	Brazil (Northeast coast)	Sergipe State: Maximum in summer and autumn Other States: no seasonal pattern	(da Silva et al., 2014; Queiroga et al., 2015, 2013)	<i>Nitzschia</i> , <i>Dinophysis</i> , <i>Alexandrium</i> , <i>Ostreopsis</i> , <i>Prorocentrum</i> , <i>Gambierdiscus</i> (Aquino et al., 2015; De Carli, 2014; Jales et al., 2013; Machado et al., 2018; Menezes et al., 2018; Nascimento et al., 2012; Santiago et al., 2010) **	NA
<i>Ostrea edulis</i>	Digestive gland disease	<i>Marteilia refringens</i> (protozoan parasite)	Lethal	Europe	Maximum in summer (>17°C). Winter and early spring: absence or few parasites.	(Berthe et al., 2004; Carrasco et al., 2015)	<i>Alexandrium</i> ., <i>Pseudo-nitzschia</i> ., <i>Dinophysis</i> , <i>Ostreopsis</i> , <i>Azadinium</i> , <i>Vulcanodinium</i> , <i>Protoceratium</i> , <i>Lingulodinium</i> , <i>Gonyaulax</i> , <i>Karlodinium</i>	NA
<i>Ostrea edulis</i>	Microcell disease	<i>Bonamia ostrea</i> (protozoan parasite)	Lethal	Europe	Unclear. Maximum prevalence in winter.	(Arzul and Carnegie, 2015)	<i>Alexandrium</i> ., <i>Pseudo-nitzschia</i> ., <i>Dinophysis</i> , <i>Ostreopsis</i> , <i>Azadinium</i> , <i>Vulcanodinium</i> , <i>Protoceratium</i> , <i>Lingulodinium</i> , <i>Gonyaulax</i> , <i>Karlodinium</i>	NA
<i>Saccostrea glomeraa</i>	Winter mortality disease	<i>Bonamia roughleyi</i> (protozoan parasite)	Lethal	Australia (East coast)	Winter and early spring	(Dove et al., 2013; Farley et al., 1988; Spiers et al., 2014)	<i>Alexandrium</i> , <i>Pseudo-nitzschia</i> , <i>Ostreopsis</i> , <i>Gambierdiscus</i> , <i>Karlodinium</i> , <i>Prorocentrum</i> , <i>Dinophysis</i> , <i>Amphidinium</i> (Ajani et al., 2017, 2013)	NA
<i>Saccostrea glomeraa</i>	Queensland unknown (QX) disease	<i>Marteillia sydneyi</i> (protozoan parasite)	Lethal	Australia (East coast)	Unclear	(Adlard and Wesche, 2005; Green et al., 2011; Nell, 2001; Perkins and Wolf, 1976)	<i>Alexandrium</i> , <i>Pseudo-nitzschia</i> , <i>Ostreopsis</i> , <i>Gambierdiscus</i> , <i>Karlodinium</i> , <i>Prorocentrum</i> , <i>Dinophysis</i> , <i>Amphidinium</i> (Ajani et al., 2017, 2013)	NA
<i>Ruditapes philippinarum</i>	Perkinsosis	<i>Perkinsus olseni</i> (protozoan parasite)	Lethal (Portugal) and sub-lethal	Europe (Western and South)	Spain, and Portugal: Maximum in spring and summer. France: no seasonality.	(Ruano et al., 2015; Villalba et al., 2011)	<i>Alexandrium</i> ., <i>Pseudo-nitzschia</i> ., <i>Dinophysis</i> , <i>Ostreopsis</i> , <i>Azadinium</i> , <i>Protoceratium</i> , <i>Lingulodinium</i> , <i>Gonyaulax</i> , <i>Karlodinium</i>	NA
<i>Ruditapes philippinarum</i>	Perkinsosis	<i>Perkinsus olseni</i> (protozoan parasite)	Lethal	East Asia (Korea, Japan, China)	South Korea: Maximum late summer; Japan: Maximum in summer and autumn	(Choi and Park, 2010; Waki et al., 2018)	<i>Margalefidinium</i> , <i>Chatonella</i> , <i>Karenia</i> , <i>Heterocapsa</i> , <i>Gymnodinium</i> , <i>Heterosigma</i> , <i>Alexandrium</i> , <i>Dinophysis</i> , <i>Prorocentrum</i> , <i>Gambierdiscus</i> , (Lee et al., 2013)	NA
<i>Ruditapes philippinarum</i>	Brown Ring Disease (BRD)	<i>Vibrio tapetis</i> (bacterium)	Lethal	Korea and Japan	Undertemined	(Park et al., 2008, 2006)	<i>Margalefidinium</i> , <i>Chatonella</i> , <i>Heterosigma</i> , <i>Prorocentrum</i> , <i>Alexandrium</i> , <i>Akashiwo</i> (Lee et al., 2013)	NA
<i>Ruditapes philippinarum</i>	Brown Ring Disease (BRD)	<i>Vibrio tapetis</i> (bacterium)	Lethal	France, Spain	Winter and spring: high prevalence and mortality	(Paillard, 2004)	<i>Alexandrium</i> ., <i>Pseudo-nitzschia</i> , <i>Dinophysis</i> , <i>Ostreopsis</i> , <i>Azadinium</i> , <i>Protoceratium</i> , <i>Lingulodinium</i> , <i>Gonyaulax</i> , <i>Karlodinium</i>	NA
Most bivalves	Vibriosis	<i>Vibrio</i> spp. (bacteria)	Lethal – sub-lethal	Worldwide	Variable and undetermined	(Travers et al., 2015)	Potentially all HABs	NA

* Main references used in this column are HAEDAT: Harmful Algae Events Database (<http://haedat.iode.org/index.php>), and (Lassus et al., 2016), in addition to other references mentioned.

** Important HAB taxa may be unlisted due to scarce information related to absence of HAB monitoring in this region

NA: Not available

Table 2. Literature survey of the main studies reporting the effects of marine harmful algal blooms (HABs) on bivalve infectious diseases or pathogens. This table summarizes HAB effects on pathogen infection in bivalves, effects of direct HAB exposure to pathogens, effects of HABs and their combination with pathogens on bivalve physiological variables, and mechanisms identified to induce disease modulation. Individual effects of pathogens on bivalves are not listed here. Detailed hemocyte responses are listed in Table 4.

Bivalve species; stage	Pathogen (infection type; conditions compared)	HAB species Exposure type	Effects of HAB on pathogen infection	Direct effects of HAB on pathogen	Bivalve variables studied	Main effects of HAB and combination with pathogen on physiology of bivalves	Identified reason for disease modulation	Reference
<i>Crassostrea virginica</i> ; adult	<i>Perkinsus marinus</i> (<i>P.m.</i>) (natural; non infected vs. infected); <i>Bucephalus</i> sp. (<i>B.</i>) (natural; non infected vs. infected)	<i>Alexandrium catenella</i> (= <i>A. fundyense</i> as referred to in this study)* Simulated bloom ; 2x10 ³ cells mL ⁻¹ 4 times a day ; 3 weeks	<u>Effect of HAB alone:</u> Prevalence & intensity (=, <i>P.m.</i> & <i>B.</i>) <u>Combined effect HAB-B:</u> Prevalence of <i>P.m.</i> (↑) in <i>B.</i> -infected oysters, Intensity (=).	NA	Viability; Condition index; Circulating hem.; Histopathology; Expression of targeted immune-related genes in gills; Toxin accumulation; Pathogen infection	<u>Effect of HAB in <i>B.</i>-uninfected bivalves</u> - Hem. infiltration (kidney) <u>Combined effect HAB - <i>B.</i>-infected bivalves:</u> - Antagonistic: Suppression of hem. inflammatory response to <i>B.</i> by HAB (circulating hem. and infiltration in tissues) - Synergistic: Tissue degeneration in adductor muscle	Immunosuppression caused by HAB exposure combined to physiological weakness caused by <i>B.</i> infection	(Lassudrie et al., 2015b)
<i>Crassostrea gigas</i> ; juvenile	Herpesvirus OsHV-1 μ Var (experimentally by cohabitation (<i>Exp.</i>) or naturally (<i>Nat.</i>); non infected vs. infected)	<i>Alexandrium pacificum</i> (named <i>A. catenella</i> in this study)* Simulated bloom ;- 3.5x10 ² cells mL ⁻¹ continuously ; 4 days	<u>Exp.:</u> Prevalence (↓), Intensity (=); <u>Nat.:</u> Prevalence (=), Intensity (↓)	NA	Viability; Toxin accumulation; Pathogen infection	<u>Combined effect HAB-pathogen:</u> Toxin accumulation (↓)	NA	(Lassudrie et al., 2015a)
<i>Crassostrea gigas</i> ; juvenile	<i>Vibrio tasmaniensis</i> LGP32. (experimental by injection; infected vs. non infected)	<i>Alexandrium pacificum</i> (named <i>A. catenella</i> in this study)*, (<i>A.p.</i> , PST strain) <i>Alexandrium tamarense</i> *, (<i>A.t.</i> , non-PST strain)	NA	NA	Viability; Toxin accumulation	<u>Effect of HAB for all infection status:</u> none <u>Combined effect HAB-pathogen:</u> Viability (↓ <i>A.p.</i> ; = <i>A.t.</i>)	NA	(Abi-Khalil et al., 2016)

		Simulated bloom; 1-3x10 ³ cells mL ⁻¹ continuously ; 10 days						
<i>Mytilus edulis</i> ; larva	<i>Vibrio splendidus</i> (V.s.); <i>Vibrio coralliilyticus</i> (V.c.) (experimentally by incubation; infected vs non infected)	<i>Prorocentrum lima</i> (P.l.) ; <i>Prorocentrum micans</i> (P.m.) ; <i>Alexandrium ostenfeldii</i> (A.o.) ; <i>Alexandrium minutum</i> (A.m.) ; <i>Karenia mikimotoi</i> (K.m.) ; <i>Protoceratium reticulatum</i> (P.r.) ; <i>Prorocentrum cordatum</i> (P.c.) Simulated bloom ; 10 ² , 5x10 ² , 2.5x10 ³ & 1.2x10 ⁴ cells mL ⁻¹ once ; 5 days	NA	NA	Viability; Larval development; Phenoloxydase activity (except for A.m, A.o, P.r, with V.s. and V.c., and for P.m., K.m. with V.s.)	<u>Effect of HAB (at highest concentrations) for all infection status:</u> - Larval viability and development rate ↓ (except P.c.) - Immune function: modulation of phenoloxydase activity depending on algal species (↑ P.c. and P.m., ↓ K.m. and P.l.) <u>Combined effect of HAB-pathogen:</u> none	NA	(De Rijcke et al., 2016)
<i>Mytilus edulis</i> ; adult	Gymnophallidae trematodes (natural; not considered as a factor)	<i>Alexandrium catenella</i> (= <i>A. fundyense</i> as referred to in this study)* Simulated bloom; 3x10 ³ cells mL ⁻¹ every 90min; 9 days.	Prevalence (↑)	NA	Viability; Histopathology; Circulating hem.; Toxin accumulation; Pathogen infection	<u>Effect of HAB for all infection status:</u> - Inflammatory response: hem. infiltration in gonad, and stomach and intestine lumen, circulating hem. modulations. - Mucus secretory cells in gills - Oxidative stress: ceroidosis in tissues	Immunosuppression caused by HAB exposure	(Galimany et al., 2008b)
<i>Perna perna</i> ; adults	RLO, copepods (Cop.), Nematopsis sp. (Nem.), Trematodes (Trem.), Tylocephalum sp. (Tyl.); (natural; not considered as a factor)	<i>Prorocentrum lima</i> Simulated bloom; 9x10 ² cells mL ⁻¹ once a day; 1, 2 & 4 days.	RLO, Cop.: Prevalence (↓) Others: Prevalence (=)	NA	Survival; Shell-valve closure response; Circulating hem.; Histopathology (incl. pathogen infection)	<u>Effect of HAB for all infection status:</u> - Alteration of shell valve-closure response - Circulating hem. modulations - In digestive gland, alteration (atrophy of digestive tubules) and inflammatory response (hem. infiltrations)	NA	(Neves et al., 2019)

<i>Ruditapes philippinarum</i> ; adult	<i>Perkinsus olseni</i> (natural; 2 classes of infection: very light-light vs. moderate-heavy)	<i>Alexandrium ostenfeldii</i> Simulated bloom ; 10 ³ cells mL ⁻¹ continuously; 7 days	Prevalence (=); Intensity (NS trend ↓)	NA	Viability; Condition index; Circulating hem. and plasma; Histopathology; Activity of antioxidant and digestive enzymes; Toxin accumulation; Pathogen infection	<u>Effect of HAB for all infection status:</u> - Tissue alterations in digestive gland, gills and mantle - Inflammatory response: hem. infiltration in gills and digestive gland, hem. diapedesis in digestive gland epithelia, circulating hem. modulations - Metabolic and oxidative processes: modulation of antioxidant enzyme activity in gills and hem. modulations - <u>Combined effect HAB - highest infection class:</u> Antagonistic: Suppression of anti/pro-oxidative responses to highest parasite burden (CAT activity in gills, ROS in hem.)	NA	(Lassudrie et al., 2014)
<i>Ruditapes philippinarum</i> ; adults	<i>Perkinsus olseni</i> (natural; 2 classes of infection: null-light vs. moderate-heavy)	<i>Karenia selliformis</i> <u>Bivalve exposure:</u> simulated bloom; 10 ² -10 ³ cells mL ⁻¹ continuously; 6 weeks. <u>Pathogen exposure:</u> <i>in vitro</i> , HAB whole culture or exudates, cell ratio 1:250 (HAB:pathogen); 30 min, 1h30, 3h30.	Prevalence and intensity (↓) after 2 and 3 weeks, and recovery after decrease of HAB concentration	Mortality (whole culture; exudates), altered morphology (whole culture)	Viability; Condition index; Circulating hem. and plasma; Pathogen infection	<u>Effect of HAB for all infection status</u> Circulating hem. modulations <u>Combined effect HAB-pathogen:</u> none	Direct toxicity of HAB to pathogen	(da Silva et al., 2008)
<i>Ruditapes philippinarum</i> ; adults	<i>Perkinsus olseni</i> (natural; 2 classes of infection: null-light vs. moderate-heavy)	<i>Prorocentrum minimum</i> <u>Bivalve exposure:</u> simulated bloom; 10 ⁵ cells mL ⁻¹ continuously; 6 days. <u>Pathogen exposure:</u> <i>in vitro</i> , HAB whole culture or exudates, cell ratio 1:50 (HAB:pathogen); 30 min	Prevalence and intensity (=)	Mortality and altered morphology (whole culture; exudates)	Viability; Condition index; Circulating hem. and plasma; Histopathology; Apoptosis; Pathogen infection	<u>Effect of HAB for all infection status</u> - Condition index (↓) - Circulating hem. modulations - Tissue alterations (adductor and foot muscles, digestive gland, ova); hem. infiltration in gonad; bacteria and hem. (viable and apoptotic) in intestine lumen. <u>Combined effect HAB-pathogen:</u> - Synergistic: Tissue degeneration in gonad	NA	(Hégaret et al., 2009)

<i>Ruditapes philippinarum</i> , adults	<i>Perkinsus olseni</i> , (natural, 2 classes of infection: null-light vs. moderate-heavy)	<i>Karenia mikimotoi</i> (<i>K.m.</i>); <i>Karenia selliformis</i> (<i>K.s.</i>); Simulated bloom; <i>K.m.</i> : 10^3 cells mL ⁻¹ ; <i>K.s.</i> : 5×10^2 cells mL ⁻¹ ; continuously: 3 & 6 days	Prevalence and intensity (=)	NA	Condition index; Circulating hem.; Plasma; Pathogen infection;	<u>Effect of HAB for all infection status:</u> Circulating hem. modulations, stronger for <i>K.s.</i> than <i>K.m.</i> <u>Combined effect HAB-pathogen:</u> Synergistic effects: circulating hem. modulations for <i>K.s.</i> after 3 days	NA	(Hégaret et al., 2007a)
<i>Ruditapes philippinarum</i> ; juveniles (<i>Juv.</i>); adults (<i>Ad.</i>)	<i>Vibrio tapetis</i> (<i>Juv.</i> : experimental by injection; <i>Ad.</i> : natural, 4 stages)	<i>Alexandrium catenella</i> (named <i>A. tamarense</i> in this study)* Simulated bloom. <i>Juv.</i> : 5×10^1 cells mL ⁻¹ continuously; 3 days. <i>Ad.</i> : 10^2 cells mL ⁻¹ continuously; 16h.	<i>Juv.</i> : Apparent prevalence and intensity (↓) (assessed by shell mark symptoms). <i>Ad.</i> : None	NA	<i>Juv. & Ad.</i> : Viability; Burrowing; Toxin accumulation; Disease stage (based on shell-mark symptoms); <i>Juv.</i> : Growth; <i>Ad.</i> : Condition index, Circulating hem.	<u>Effect of HAB</u> , all infection status; <i>Juv.</i> : Viability (↓); Burrowing incapacitation; Growth (↓); <i>Ad.</i> : Burrowing incapacitation; Circulating hem. modulations <u>Combined effect HAB-pathogen:</u> <i>Juv.</i> : Cumulative: Higher mortality	<i>Juv.</i> : Unclear if apparent decrease of disease severity traduces host-pathogen modulation, or a decrease of defense mechanisms (possibly related to shell mark symptoms)	(Bricelj et al., 2011)
<i>Mercenaria mercenaria</i> ; adults	QPX (natural; infected vs. non infected)	<i>Prorocentrum minimum</i> Simulated bloom; 2×10^4 cells mL ⁻¹ continuously; 5 days.	NA	NA	Viability; Circulating hem. and plasma; Histopathology; Pathogen infection	<u>Effect of HAB in non-infected bivalves:</u> - Circulating hem. modulations - Tissue alteration and inflammatory response (hem. infiltrations) in gills, hemocytic aggregates in all organs <u>Effect of HAB in infected bivalves:</u> - Circulating hem.: morphology - Hem. infiltrations in tissues	NA	(Hégaret et al., 2010)
<i>Argopecten purpuratus</i> , adults	RLO, (natural; not considered as a factor)	<i>Alexandrium catenella</i> * Simulated bloom; 5×10^3 cells mL ⁻¹ continuously; 6 days	Prevalence (↑)	NA	Survival; Escape response, Heat-shock protein (hsp70); Histopathology (incl. pathogen infection); Toxin accumulation	<u>Effect of HAB for all infection status:</u> - Altered escape response to predator - Tissue alteration in muscle - Melanization of epithelia	Physiological weakness and decreased defense associated to pathologies caused by HAB	(Hégaret et al., 2012)

(↑): increase; (↓): decrease; no change: (=); NA: not available (not investigated); NS: non significant; Hem.: hemocytes; QPX: Quahog Parasite Unknown; RLO: Rickettsia-like organisms; incl.: including. *: Nomenclature for species within the *A. tamarense* complex has changed since the publication of these studies, current accepted names indicated here were deduced from John et al. (2014); Fraga et al. (2015); Litaker et al. (2018), where *A. catenella* corresponds to group I, *A. tamarense* to group III, *A. pacificum* to group IV).

Table 3. Literature survey of most recently reported effects of marine harmful algal blooms (HABs) or their toxins on bivalve hemocyte variables involved in cellular immunity.

Bivalve species	HAB/phyco toxin	Experiment type	Hemocyte parameters	Reference
<i>Mytilus chilensis</i>	Saxitoxins	<i>In vitro</i> 1–100 nM, 4h and 16h.	Viability (=), ROS (↓ 10 nM, ↑ 100 nM), phagocytosis (↓)	(Astuya et al., 2015)
<i>Crassostrea gigas</i>	Saxitoxins	<i>In vitro</i> 0.8, 1 and 3.3 μM, 3.5h.	Apoptosis (↑).	(Abi-Khalil et al., 2017)
<i>Crassostrea gigas</i> (C.g.) <i>Chlamys farreri</i> (C.f.)	Saxitoxins	Injection 10–20 μg STX eq. 100 g ⁻¹ shellfish meat, 12, 48, and 96 h p.i.	Phagocytosis (↓ 12 and 96h, C.g. and C.f.), ROS (↑ 48h and 96h, C.f.).	(Cao et al., 2018)
<i>Crassostrea gigas</i>	<i>Alexandrium minutum</i> (A.m.); Saxitoxins (STX)	<i>In vitro</i> A.m.: 2 x 10 ⁴ cells mL ⁻¹ . STX: 0.05 μg L ⁻¹ for 4h.	Size (↑ GR, A.m.), and complexity (=), viability (=), phagocytosis (↓), ROS (↓) for A.m and STX	(Mello et al., 2013)
<i>Crassostrea gigas</i> (diploid and triploid, in April and in May)	<i>Alexandrium minutum</i>	Simulated bloom 5 x 10 ³ cells mL ⁻¹ , for 4 days.	THC (↑) GRC (↑), viability (=), phagocytosis (=), ROS (↑ April GR, ↓ May GR and HY), agglutination titer (=), phenoloxidase (↓ April, ↑ May).	(Haberkorn et al., 2010a)
<i>Crassostrea gigas</i>	<i>Alexandrium minutum</i>	Simulated bloom 5 x 10 ³ cells mL ⁻¹ , flow of 14 mL min ⁻¹ , for 4 days.	THC (=), phagocytosis (↑), ROS (=), phenoloxidase (=).	(Haberkorn et al., 2014)
<i>Crassostrea gigas</i>	<i>Alexandrium minutum</i> (PST, BEC, and PST+BEC strains)	Simulated bloom 3.1 x 10 ³ cells mL ⁻¹ , for 4 days (no control algae).	THC (=), HYC (PST < PST+BEC, BEC), Size (=), complexity (=), viability (PST < PST+BEC, BEC), phagocytosis (=), ROS (=).	(Castrec et al., 2018)
<i>Crassostrea virginica</i> (C.v.)	<i>Alexandrium catenella</i> (= <i>A. fundyense</i> as referred to in this study*; A.c.)	Simulated bloom A.c.: 1, 2.5, 5 x 10 ² cells mL ⁻¹ , for 1, 7 days.	THC (=), size (=), complexity (=), viability (=), phagocytosis (=), ROS (=), for C.v and C.g.	(Hégaret et al., 2007b)
<i>Crassostrea gigas</i> (C.g.)	<i>A. pacificum</i> (named <i>A. catenella</i> in this study*; A.p.)	A.p.: 75 cells mL ⁻¹ , for 14 h, then 1.5 x 10 ² cells mL ⁻¹ , for 10 h day for 4 days.		
<i>Crassostrea gigas</i> (juveniles)	<i>Alexandrium pacificum</i> (named <i>A. catenella</i> in this study)*	Simulated bloom 10 ² cells mL ⁻¹ for 4 to 9 days.	THC (↑), GRC (↑), HYC (↑), size (↑ HY, ↑ GR day 4), complexity (↑ HY, ↑ GR), viability (=), ROS (↑).	(Lassudrie et al., 2016)
<i>Ruditapes philippinarum</i> , <i>Mya arenaria</i>	<i>Alexandrium catenella</i> (named <i>A. tamarense</i> in this study)* (PST) <i>Alexandrium tamarense</i> (nonPST)	<i>In vitro</i> Filtered extracts of 5 x 10 ⁵ cells mL ⁻¹	Adherence (↓ nonPST), phagocytosis (↓ nonPST).	(Ford et al., 2008)
<i>Mytilus edulis</i>	<i>Alexandrium catenella</i> (PST and PST+LC strains) <i>A. tamarense</i> (LC strain)	Simulated bloom 500 cells mL ⁻¹ , for 3 and 7 days.	THC (↓ PST and LC, = PST+LC, 3 days ; = PST, PST+LC and LC, 7 days), viability and lysosomal membrane stability (↓ for all, 3 and 7 days), phagocytosis (↑ PST, 3 days ; = for all, 7 days), ROS (↓ for all, 3 days).	(Bianchi et al., 2019)

<i>Crassostrea virginica</i> (native and non-native populations)	<i>Prorocentrum minimum</i>	Natural and simulated bloom 10 ⁴ cells mL ⁻¹ , 2 days. Native: exposed to recurrent blooms; Non-native: naïve from bloom exposure.	THC (↑), GRC (↑), viability (↑), phagocytosis (↑), ROS (↑ native, = non-native), aggregation (↓).	(Hégaret and Wikfors, 2005a)
<i>Crassostrea virginica</i> (C.v.), <i>Argopecten irradians irradians</i> (A.i.i.)	<i>Prorocentrum minimum</i>	Simulated bloom 10 ⁵ cells mL ⁻¹ , for 1h, and 1, 2, 4 and 7 days.	C.v.: THC (=), size (=), complexity (=), viability (=), phagocytosis (=), ROS (=), aggregation (=). A.i.i.: GRC (↓), HYC (↑), viability (↑ GR, ↓ HY), phagocytosis (↓), ROS (↑), aggregation (↑ HY, ↓ GR).	(Hégaret and Wikfors, 2005b)
<i>Mytilus edulis</i>	<i>Prorocentrum minimum</i>	Simulated bloom 10 ⁴ cells mL ⁻¹ , for 9 days.	Viability (=), phagocytosis (=), ROS (=), adhesion (=), apoptosis (=).	(Galimany et al., 2008c)
<i>Perna perna</i>	<i>Prorocentrum lima</i>	Simulated bloom 900 cells mL ⁻¹ , for 1, 2 and 4 days.	THC (↓ 2 & 4 days), size (↑2 & 4 days), phagocytosis (↑ 2 days), ROS (↑ 4 days).	(Neves et al., 2019)
<i>Ruditapes decussatus</i>	<i>Prorocentrum lima</i> (P.l.); Okadaic acid (OA)	P.l.: Simulated bloom 2 x 10 ² and 2 x 10 ⁴ cells mL ⁻¹ , for 24h and 48h. OA: <i>In vitro</i> 10, 50, 100 and 500 nM, for 2 and 4h.	P.l.: Viability (↓ 24 h), apoptosis (↑ 24 h). OA: Viability (↓ 100 and 500 nM, 2 and 4 h), phagocytosis (↓ all concentrations), apoptosis (↑ 100 and 500 nM, 4 h).	(Prado-Alvarez et al., 2013)
<i>Mytilus galloprovincialis</i>	<i>Prorocentrum lima</i>	Simulated bloom 10 ³ and 10 ⁵ cells mL ⁻¹ , 24h and 48h.	Apoptosis (↑ 48h, both concentrations).	(Prego-Faraldo et al., 2016)
<i>Mytilus galloprovincialis</i>	<i>Prorocentrum</i> and/or <i>Dinophysis</i> ; OA	Natural bloom; OA: <i>In vitro</i> 10, 50 and 100 nM, for 2 and 4 h.	Natural bloom: Apoptosis (↓) OA: Apoptosis (↓ 100 nM, 4h).	(Prado-Alvarez et al., 2012)
<i>Mytilus galloprovincialis</i>	OA	<i>In vitro</i> 10, 50, 100, 200 and 500 nM, for 1 and 2 h.	Apoptosis (↑ 500 nM, 2h).	(Prego-Faraldo et al., 2015)
<i>Perna perna</i> (P.p.), <i>Crassostrea gigas</i> (C.g.)	<i>Dinophysis acuminata</i>	Natural bloom 2 sites: PA: 2950 cells L ⁻¹ PZ: 4150 cells L ⁻¹	P.p.: THC (↓ PA, ↑ PZ), DHC (=), hemagglutinating titter (=), phenoloxidase (↑PA, = PZ), C.g.: THC (=), DHC (=), hemagglutinating titter (=), phenoloxidase (↓ PZ).	(Simões et al., 2015)
<i>Perna perna</i> (P.p.), <i>Anomalocardia brasiliana</i> (A.b.), <i>Crassostrea gigas</i> (C.g.)	<i>Dinophysis acuminata</i>	Natural bloom 17,600 cells L ⁻¹ .	P.p.: THC (↑), GRC (↓), phenoloxidase (↓). A.b.: THC (↑), GRC (=), phenoloxidase (=). C.g.: THC (=), GRC (=), phenoloxidase (=). Apoptosis and hemagglutinating titter (=, all spp.).	(Mello et al., 2010)

<i>Mercenaria mercenaria</i> (M.m.), <i>Mya arenaria</i> (M.a.), <i>Crassostrea virginica</i> (C.v.), <i>Crassostrea gigas</i> (C.g.), <i>Ruditapes philippinarum</i> (R.p.)	<i>Heterosigma akashiwo</i> (H.a.) <i>Prorocentrum minimum</i> (P.m.) <i>Alexandrium catenella</i> (= <i>A. fundyense</i> as referred in this study) (Ac.) <i>Alexandrium minutum</i> (A.m.) <i>Karenia selliformis</i> (K.s.) <i>Karenia mikimotoi</i> (K.m.)	<i>In vitro</i> H.a.: 10 ⁵ cells mL ⁻¹ P.m.: 10 ⁵ cells mL ⁻¹ A.c.: 10 ⁴ cells mL ⁻¹ A.m.: 5 x 10 ⁴ cells mL ⁻¹ K.s.: 4-7 x 10 ³ cells mL ⁻¹ K.m.: 4-7 x 10 ³ cells mL ⁻¹ For 4 h	H.a.: M.m.: viability (↓ WC), phagocytosis (=), ROS (=), adhesion (↓ S); M.a.: phagocytosis (=), ROS (↑ WC), adhesion (=); C.v.: viability (↓ WC), phagocytosis (=), adhesion (=). P. m.: M.m.: size (↓ WC), complexity (↓ WC), adhesion (↑ S), phagocytosis (↓ WC); M.a.: ROS (=), adhesion (=), phagocytosis (↓ WC); C.v.: All parameters (=). A.c.: M.m.: phagocytosis (↓ WC and S), ROS (=), adhesion (↓ WC); M.a.: phagocytosis (↓ WC), ROS (=), adhesion (=); C.v.: viability (↓ WC), remaining parameters (=). A.m.: C.g.: viability (↓ WC and S), ROS (↓ WC), remaining parameters (=). K.s.: R.p.: size (↓ WC), viability (↓ WC and S), phagocytosis (↓ WC), ROS (↓ WC and S), adhesion (↓ WC). K.m.: R.p.: ROS (↓ WC), adhesion (↓ WC), other parameters (=).	(Hégaret et al., 2011)
<i>Crassostrea gigas</i>	Brevetoxin	<i>In vitro</i> 3, 30, 100, 300 and 1000 µg L ⁻¹ , for 1, 4 and 12 h.	Viability (=), apoptosis (=).	(Mello et al., 2012)
<i>Mytilus galloprovincialis</i>	<i>Ostreopsis cf. ovata</i>	Simulated bloom 1 x 10 ⁵ cells L ⁻¹ , for 7 and 14 days.	GRC (↓), HYC (↑), phagocytosis (↓), lysosomal membrane stability (↓), for both durations.	(Gorbi et al., 2013)
<i>Mytilus edulis</i>	Domoic acid	Injection 1, 10, 100 and 500 ng g body weight ⁻¹ , 48h and 7 days p.i.	THC (↑ 48h, 500 ng), viability (all concentrations: = 48h; ↑ 7 days), phagocytosis (↑ 48h, 100 ng).	(Dizer et al., 2001)
<i>Ruditapes philippinarum</i>	<i>Heterocapsa circularisquama</i>	Simulated bloom 10 ³ cells mL ⁻¹ , for 24 and 48h.	THC (↑ 24h).	(Basti et al., 2011)
<i>Crassostrea gigas</i>	<i>Alexandrium pacificum</i> (named <i>A. catenella</i> in this study)* (PST) <i>Alexandrium tamarense</i> (nonPST)	Simulated bloom, 10 ³ cells mL ⁻¹ , for 48h.	Apoptosis (PST>nonPST, 29h)	(Medhioub et al., 2013)
<i>Nodipecten subnodosus</i>	Saxitoxins	Injection: 140 µg STX eq per 0.2 mL injection, 24h p.i. <i>In vitro</i> : 10–20 µg STX eq.mL ⁻¹ , 24h p.i.	Apoptosis (↑)	(Estrada et al., 2014)
<i>Nodipecten subnodosus</i>	Saxitoxins	Injection Low dose: 6.25 MU, 0.2mL, 12-72h p.i. High dose: 100 MU, 0.2mL, 40 days p.i.	THC (↓) (low dose, 12-24h p.i., high dose 10-40 days p.i.)	(Estrada et al., 2010)
<i>Mytilus edulis</i>	<i>Karlodinium veneficum</i>	Simulated bloom 6.25 x 10 ¹ cells mL ⁻¹ , for 3 and 6 days	GRC (↓ 3 days, = 6 days), HYC (↑ 3 days, = 6 days), ROS (↑), phagocytosis (↑ 6 days)	(Galimany et al., 2008a)

THC: total hemocyte count; DHC: differential hemocyte count; GR: granulocytes; HY: hyalinocytes (a.k.a. agranulocytes); GRC: granulocyte count; HYC: hyalinocyte count; ROS: reactive oxygen species production; PST: paralytic shellfish toxin; STX: Saxitoxin; LC: lytic compounds; BEC: bioactive extracellular compounds; OA: Okadaic acid; PA: Praia Alegre, Brazil; PZ: Praia de Zimbros, Brazil; MU: mouse units; p.i.: post injection; ↓: decrease, ↑: increase, or = : no difference compared to control non-toxic treatments; < decrease or

> increase compared to other treatments. *: Nomenclature for species within the *A. tamarense* complex has changed since the publication of these studies, current accepted names are indicated here and were deduced from John et al. (2014), Fraga et al. (2015) and Litaker et al. (2018), where *A. catenella* (= *A. fundyense*) corresponds to group I, *A. tamarense* to group III, *A. pacificum* to group IV.

Table 4. Literature survey of most recently reported effects of marine harmful algae blooms (HABs) combined with pathogen infection on bivalve hemocyte variables involved in cellular immunity.

Bivalve species	Pathogen	HAB species	Experiment type	Effects of HABs and pathogens on hemocytes	Effect of HAB on pathogen infection (^h : considered to be related to hemocytes)	Reference
<i>Ruditapes philippinarum</i>	<i>Perkinsus olseni</i>	<i>Karenia mikimotoi</i> (K.m.) <i>Karenia selliformis</i> (K.s.)	Simulated bloom K.m.: 10 ³ cells mL ⁻¹ , K.s.: 5 x 10 ² cells mL ⁻¹ , for 3 and 6 days.	<u>Effect of HAB</u> : viability (↑, K.s., K.m.), size and complexity (↓, K.s.), phagocytosis (=), ROS (=), agglutination titer (=). <u>Effect of parasite</u> : all parameters (=). <u>Combined effect</u> : THC (M-H: ↑ K.s.), phagocytosis (M-H: ↓ K.s) (after 3 days).	Prevalence (=), intensity (=).	(Hégaret et al., 2007a)
<i>Ruditapes philippinarum</i>	<i>Perkinsus olseni</i>	<i>Karenia selliformis</i>	Simulated bloom 10 ² - 10 ³ cell mL ⁻¹ , for 6 weeks.	<u>Effect of HAB</u> : viability (=), size (↓), phagocytosis (=), ROS (=), agglutination titer (=), apoptosis (↓). <u>Effect of parasite</u> : all parameters (=). <u>Combined effect</u> : none	Intensity of infection (↓, after 2 & 3 weeks), associated with direct toxicity of HAB to parasite <i>in vitro</i>	(da Silva et al., 2008)
<i>Ruditapes philippinarum</i>	<i>Perkinsus olseni</i>	<i>Prorocentrum minimum</i>	Simulated bloom 10 ³ cells mL ⁻¹ , for 6 days. <i>In vitro</i> <i>P. minimum</i> - <i>P. olseni</i> – (1:500)	<u>Effect of HAB</u> : THC (=), size (↓), complexity (=), viability (=), phagocytosis (↓), ROS (=), agglutination titer (=). <u>Effect of parasite</u> : all parameters (=). <u>Combined effect</u> : none	Prevalence (=), intensity (=).	(Hégaret et al., 2009)
<i>Ruditapes philippinarum</i>	<i>Perkinsus olseni</i>	<i>Alexandrium ostenfeldii</i>	Simulated bloom 10 ³ cells mL ⁻¹ , for 7 days.	<u>Effect of HAB</u> : THC (=), size (↑), complexity (=), viability (=), phagocytosis (=), ROS (↑), agglutination titer (=). <u>Effect of parasite</u> : M-H, size (↑) ROS (↑) <u>Combined effect</u> : ROS (antagonistic: repression of ↑).	Intensity of infection (↓ non-significant) ^h	(Lassudrie et al., 2014)
<i>Crassostrea virginica</i>	<i>Perkinsus marinus</i> (P.m.) <i>Bucephalus</i> sp. (B.)	<i>Alexandrium catenella</i> (= <i>A. fundyense</i> as referred to in this study)*	Simulated bloom 1-2 x 10 ³ cells mL ⁻¹ , for 3 weeks.	<u>Effect of HAB</u> : THC (=), size (=) and complexity (=), ROS (=). <u>Effect of B.</u> : THC (↑), viability (↑). <u>Effect of P.m.</u> : all parameters (=). <u>Combined effect HAB-B</u> : THC (repression of ↑ normally induced by B.)	Prevalence of <i>P. m.</i> ↑ in <i>B.</i> infected oysters ^h	(Lassudrie et al., 2015a)
<i>Mercenaria mercenaria</i>	QPX	<i>Prorocentrum minimum</i>	Simulated bloom 2 x 10 ⁴ cells mL ⁻¹ , for 5 days.	<u>Effect of HAB</u> : THC (=), size (↑), complexity (=), viability (=), phagocytosis (↓), ROS (↑), adhesion (=), apoptosis (=). <u>Combined effect</u> : phagocytosis (↓), size (↓).	NA	(Hégaret et al., 2010)

<i>Mytilus edulis</i>	Gymnophalli dae trematode	<i>Alexandrium catenella</i> (= <i>A. fundyense</i> as referred to in this study)*	Simulated bloom 4 x 10 ³ cells mL ⁻¹ , for 9 days.	<u>Effect of HAB</u> : THC (↓), GRC (↓), phagocytosis (=), ROS (=), complexity (↓, GR and HY), size (GR ↓, HY =). <u>Combined effect</u> : none	Prevalence (↑) ^h	(Galimany et al., 2008b)
<i>Ruditapes philippinarum</i> (adults)	<i>Vibrio tapetis</i>	<i>Alexandrium catenella</i> (named <i>A. tamarensis</i> in this study)*	Simulated bloom. 10 ² cells mL ⁻¹ , for 16h.	<u>Effect of HAB</u> : THC (↑), GRC (↑), phagocytosis (↓)	None	(Bricelj et al., 2011)

THC: total hemocyte count; GR: granulocytes; HY: hyalinocytes (a.k.a. agranulocytes); GRC: granulocyte count; ROS: reactive oxygen species production; VL-L: very-light and light *P. olsenii* infection intensity; M-H: moderate to heavy *P. olsenii* infection intensity; QPX: Quahog Parasite Unknown; ↓: decrease; ↑: increase or =: no difference compared to control non-toxic algal treatments. *: Nomenclature for species within the *A. tamarensis* complex has changed since the publication of these studies, current accepted names are indicated here and were deduced from John et al. (2014), Fraga et al. (2015) and Litaker et al. (2018), where *A. catenella* (= *A. fundyense*) corresponds to group I.

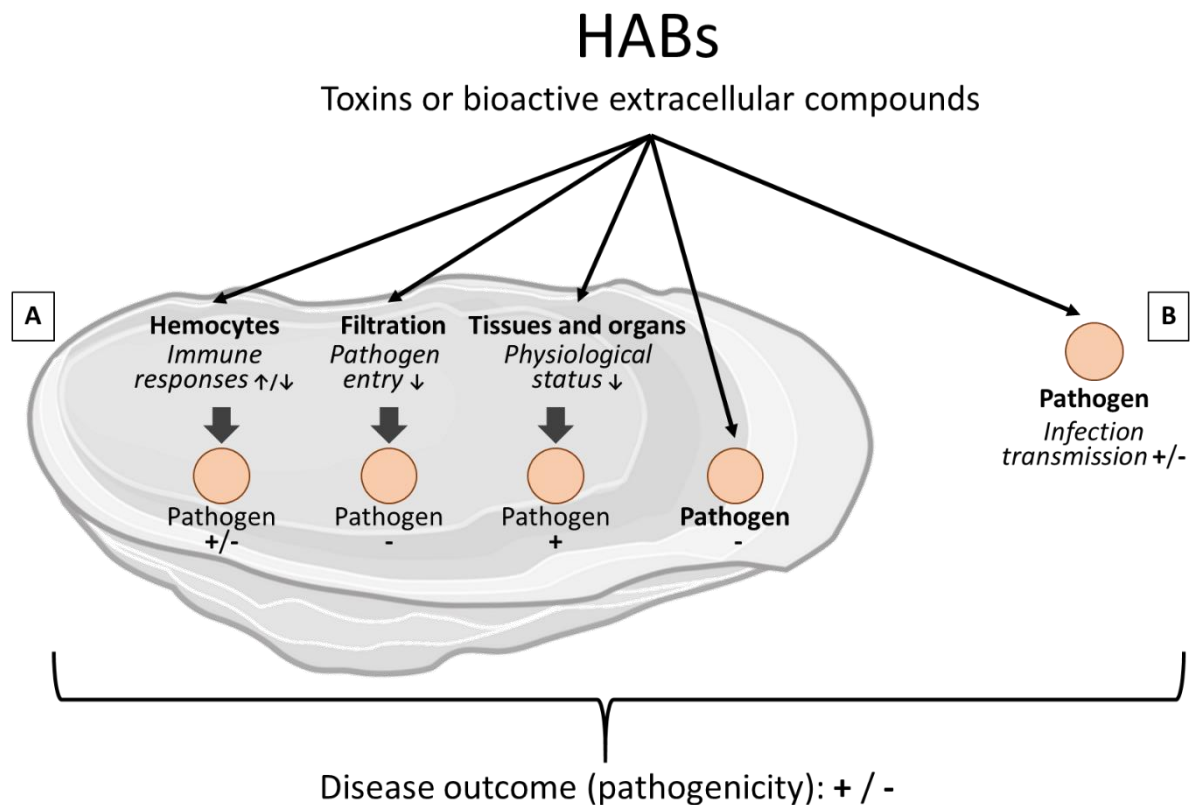


Figure 1. Summary of the different mechanisms reported or supposed to be involved in the effects of harmful algal blooms (HABs) upon bivalve diseases, according to the literature. Arrows indicate increase or decrease of the adjoining process, + and – indicate positive and negative effect, respectively, on the pathogen transmission or proliferation. **A.** When reaching bivalves, HABs and compounds produced can affect hemocyte functions, either by inhibiting them (\downarrow) causing immunosuppression and favor pathogen proliferation (+), or, conversely, stimulate immune responses (\uparrow), which could inhibit pathogen proliferation (-). HABs can decrease filtration rates that may in turn decrease pathogens entry (\downarrow) and reduce its transmission (-). HABs can cause tissue and organs lesions, altering physiological status (\downarrow) and therefore decreasing defense against pathogens, that can proliferate (+). Pathogen in bivalve tissues could also be affected by HAB compounds directly, altering its viability or pathogenicity (-). **B.** HABs and compounds produced can also interact directly with pathogens in the seawater, altering pathogen integrity or putatively acting as pathogen carrier, therefore preventing (-) or favouring (+) pathogen transmission.

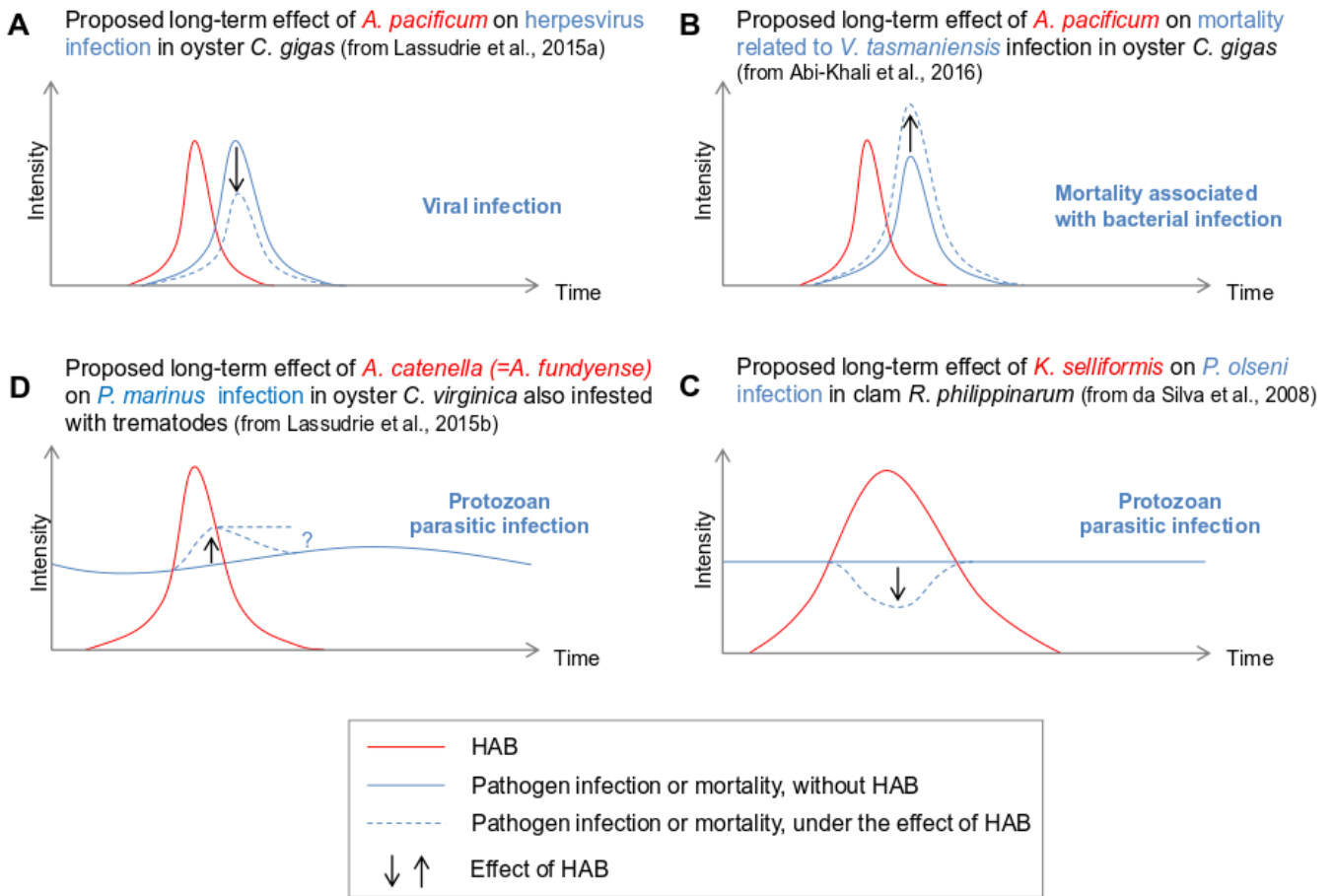


Figure 2. Hypothetical scenarios of the long-term effect of HABs on disease dynamics in bivalves, proposed based on short-term experimental studies. The evolution of the HAB (red), and of the pathogen infection or the mortality rate (without the effect of HAB: solid blue line; with the effect of HAB: interrupted blue line) are represented on the y-axis, according to a temporal scale, represented on the x-axis. **A.** Proposed effect of *Alexandrium catenella* bloom on herpesvirus OsHV-1 infection in oyster *Crassostrea gigas* spat (from Lassudrie et al., 2015a). **B.** Proposed effect of *A. catenella* on mortality associated with bacterium *Vibrio tasmaniensis* infection in oyster *C. gigas* spat (from Abi-Khalil et al., 2016). **C.** Proposed effect of *A. fundyense* on protozoan parasite *Perkinsus marinus* infection in oyster *C. virginica* also infested with trematodes (from Lassudrie et al., 2015b). **D.** Proposed effect of *Karenia selliformis* on protozoan parasite *P. olseni* infection in clam *Ruditapes philippinarum* (from da Silva et al., 2008).