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Genetic analysis of partial resistance to bacterial leaf streak (*Xanthomonas campestris* pv. *cerealis*) in wheat

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Genetic variability of partial resistance to bacterial leaf streak was investigated in hexaploid winter wheat (*Triticum aestivum*), using 16 parental genotypes and 48 pure lines (F10) derived from a composite cross programme. Two experiments were undertaken in a controlled growth chamber. Seeds of all genotypes were grown under controlled conditions using a randomized block design with three replications. Each replication consisted of a row of 20 seedlings of each parent and pure line. An Iranian strain of bacterial leaf streak was used for the inoculation of 12-day-old seedlings. In a third experiment, eight genotypes from parents and F10 pure lines representing a large variability for partial resistance were inoculated with four other Iranian strains of bacterial leaf streak. A large genetic variability was observed amongst the 64 genotypes for partial resistance to the disease. Partial resistance heritability estimates were rather high (70%), indicating that the resistance factors may be transmitted by crossing. Amongst all genotypes investigated, 'DC²-30-N2' and 'IBPT-66' displayed the highest partial resistance to the disease. Significant correlations between strains in the third experiment show that a genotype resistant or susceptible to one strain will have similar reactions with other strains. No significant genetic gain was observed for partial resistance in the best pure line of the 48 lines studied, when compared with the best parental line. Increasing the number of pure lines is likely to result in the identification of genotypes that might prove to be more resistant.

INTRODUCTION

Bacterial leaf streak caused by various pathovars of *Xanthomonas campestris* is a serious worldwide bacterial disease of wheat (Boosalis, 1952; Duveiller, 1989). Losses attributed to bacterial leaf streak can be quite high under conditions favourable to the pathogen; up to 40% yield reductions were reported in susceptible wheat cultivars (Schaad & Forster, 1985). The disease also affects grain quality (Mehta, 1990). Bacterial leaf streak has been increasing on a world scale, and has now become one of the most important diseases of cereals (Mehta, 1990).

Foliar expression of the disease is characterized by longitudinal stripes that extend between the leaf veins and by the production of milky exudates

under humid conditions (Duveiller *et al.*, 1993). Contaminated seed is the main source of inoculum (Sands *et al.*, 1986; Forster & Schaad, 1988; Duveiller, 1989). Eradication of the bacterial inoculum from the wheat seed would be helpful, but known chemical and physical methods of seed treatment are not sufficiently effective (Forster & Schaad, 1988; Sun *et al.*, 1988; Fourest *et al.*, 1990). The susceptibility of wheat to bacterial leaf streak is similar under field and greenhouse conditions (Akhtar & Aslam, 1986). The disease reactions on the primary leaves are the same as those observed on the flag leaf (Milus & Mirlohi, 1994). Resistance amongst 19 wheat cultivars to 81 bacterial strains isolated from different cereals was highly variable, but the cultivar \times strain interaction was not significant and there was thus no evidence for races of the pathogen (Milus & Chalkly, 1994)

There is an increasing interest in wheat improvement programmes and in the development of cultivars with improved resistance to this disease.

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The development of easier, cheaper and widely applicable methods of integrated control requires the identification and incorporation of resistance genes into wheat breeding material. However, the genetic component of resistance to the disease is still not clearly understood. According to Woo & Smith (1962), resistance to bacterial leaf streak in wheat is controlled by a single dominant gene. The inheritance of resistance was also investigated by Duveiller *et al.* (1993) who showed that resistance to bacterial leaf streak or black chaff in five wheat lines was conditioned by five genes which differed in magnitude of expression of resistance. The aim of the investigations presented below was to assess the genetic variability of partial resistance to bacterial leaf streak in 64 hexaploid winter wheat (*Triticum aestivum*) genotypes: 16 parental genotypes and 48 pure lines (F10) derived from a composite cross programme.

MATERIALS AND METHODS

The experiments were carried out with 64 genotypes of hexaploid winter wheat (*T. aestivum*), including 48 randomized F10 pure lines derived from a composite cross population (IBPT) and their 16 parental genotypes. The composite cross population (IBPT) is a genetic pool developed by INRA (Institut National de la Recherche Agronomique, France). This pool was obtained from a pyramidal cross of 16 pure lines (Thomas *et al.*, 1991). A bulk was made with the hybrid plants obtained after the last cross and was multiplied for 3 years giving B0 population. Random samples of B0 were used in 1984 to found the B population in a multilocal network in France in the framework of a programme of dynamic management of wheat genetic resources. This population has been growing since 1984 in the same location at normal cultural density. At least 4000 individuals are grown each year. In the F10 generation of this composite population grown at our Department of Biotechnology and Plant Breeding (IBPT), 100 plants were randomly selected in order to study the genetic control of different agronomic traits. From the 100 above-mentioned F10 pure lines, 48 were selected at random for use in this study. The resistance or susceptibility of the F10 pure lines and their parents had not been studied previously.

Two successive experiments, with three replicates in each case, were performed in a growth chamber. Seeds were sterilized with sodium hypochlorite

solution (1.56% of available chlorine) for 5 min, washed in sterile distilled water and planted in plastic containers (60 × 40 × 8 cm) containing moistened vermiculite.

The bacterial strain (IBLS 20), which had been isolated from a wheat variety grown in Iran, and identified as *Xanthomonas campestris* pv. *cerealis* (Alizadeh & Rahimian, 1989) was used in our programme. The bacterial strain was grown at 26 ± 1°C and maintained on slants of nutrient agar medium (Difco) at 3 ± 1°C or was lyophilized for long-term storage. Each replication included one row of 20 seedlings per parent or pure lines and each tray contained eight rows. Seedlings were watered with Knop's nutrient solution (Arabi *et al.*, 1992) every day throughout the experiment. The containers were placed in a growth chamber at temperatures of 25 ± 1°C (day), 23 ± 1°C (night), and a relative humidity of 75–85%. Light intensity was 90 mol m⁻² s⁻¹, with a day length of 12 h.

One day before inoculation, the seedlings were sprayed with sterile distilled water and each container covered with a Perspex translucent lid to maintain near-saturated humidity, which is favourable to bacterial inoculation. Twelve-day-old seedlings were sprayed with a bacterial suspension standardized at about 10⁹ colony-forming units per mL in sterile distilled water (CFU mL⁻¹ SDW), prepared from a 24-h-old culture.

Twelve days after inoculation, the first leaves of the seedlings were scrutinized to determine the proportion of diseased leaf area (water soaking and translucent spots or stripes). Host reaction was rated from 1 (resistant) to 9 (susceptible) in relation to the proportion of leaf area showing disease symptoms as proposed by Alizadeh *et al.* (1994): 1: 0–5%; 2: 5–10%; 3: 10–20%; 4: 20–30%; 5: 30–40%; 6: 40–50%; 7: 50–60%; 8: 60–75% and 9: 75–100% of leaf area infected.

Statistical analyses were carried out in order to determine the genotype effect on the studied trait. The Newman–Keuls test was used for comparing means for partial resistance. Heritability was determined according to the equations:

$$MSG = r\delta^2G + \delta^2E,$$

$$\delta^2p = \delta^2G + \delta^2E,$$

$$h^2 = (\delta^2G/\delta^2p)'100,$$

where MSG is the mean square of genotypes, δ^2G is the genetic variance, δ^2E is the environmental

variance, δ^2p is the phenotypic variance, r is the number of blocks and h^2 is the heritability.

The mean (\bar{x}) of the 48 F10 pure lines and that of the 16 parents were compared. The best pure line was also compared with the best parent for partial resistance.

In order to determine the resistance and susceptibility of this material to other strains of bacterial leaf streak, a third experiment was made under the same conditions as before. In this experiment, eight genotypes (parents and F10 pure lines), representing a large variability for

resistance and susceptibility identified in the two previous experiments, were inoculated with four Iranian strains of bacterial leaf streak (IBLS 22, IBLS25, IBLS 29 et IBLS 41). Means (\bar{x}) of partial resistance were compared and correlations between the strains used in all experiments for partial resistance were estimated.

RESULTS AND DISCUSSION

The genetic variability of parental genotypes

Table 1 Genetic variability for resistance of parental genotypes and F10 pure lines of a composite cross in hexaploid wheat

Varieties or F10 lines	Experiment I	Experiment II	Varieties or F10 lines	Experiment I	Experiment II
<i>Parents</i>					
C45	5.79a	5.58a	Kaukaz	4.02b	3.03a
Clement 2	6.05a	5.24ab	Tjb36	3.93b	4.33abcd
Rivoli	4.64ab	3.47cd	Vc75Rv53	3.97b	3.60cd
Weihemphe	4.55ab	3.90bcd	Oxley	3.95b	3.91bcd
Loros cr	4.60ab	3.21cd	Top	3.29bc	3.22cd
Mironovskaia	4.38ab	4.78abc	Clément 1	3.18bc	2.84d
C3275	4.55ab	4.19abcd	C6156	2.00c	1.60e
D48	4.60ab	4.54cd	De ² 30N2	1.93c	1.58e
<i>F10 lines</i>					
IBPT 1	5.83cde	4.50fgh	IBPT 23	6.95ab	5.73def
IBPT 2	5.19efg	4.62fgh	IBPT 25	3.72ijk	4.23ghi
IBPT 4	4.48ghi	6.86abc	IBPT 28	3.91ghi	3.49jk
IBPT 6	4.02ghi	3.47jk	IBPT 29	5.73cde	4.24ghi
IBPT 7	6.35bcd	3.95ijk	IBPT 33	5.72cdef	6.07cde
IBPT 10	4.50fgh	4.47fgh	IBPT 34	3.55klm	3.40jk
IBPT 12	4.86fgh	5.66def	IBPT 37	5.14efg	5.70def
IBPT 16	5.52def	5.69def	IBPT 38	5.05efg	6.32bcd
IBPT 17	6.24bcd	4.09ijk	IBPT 40	3.69klm	4.75gh
IBPT 19	4.60fgh	3.94ijk	IBPT 43	5.24def	5.67def
IBPT 22	5.62cde	5.73def	IBPT 44	4.12ghi	4.30ghi
IBPT 45	4.81fgh	5.65def	IBPT 72	4.36ghi	6.18bcd
IBPT 48	3.76ijk	4.67fgh	IBPT 74	5.19efg	5.05fgh
IBPT 50	5.17efg	5.04fgh	IBPT 75	5.57def	5.38efg
IBPT 51	4.67fgh	5.77def	IBPT 76	5.44def	4.40ghi
IBPT 54	5.26def	6.38bcd	IBPT 81	4.88fgh	4.54fgh
IBPT 55	5.67cde	5.2fgh	IBPT 84	3.28lm	3.51jk
IBPT 57	6.29bcd	4.92fgh	IBPT 85	5.17efg	4.50fgh
IBPT 59	4.83fgh	4.52fgh	IBPT 86	6.43bcd	6.83abc
IBPT 60	3.29lm	5.51efg	IBPT 87	6.83abc	5.98cde
IBPT 63	4.07ghi	5.58def	IBPT 88	4.71fgh	4.58fgh
IBPT 64	3.79hij	5.25efg	IBPT 89	7.31a	7.67a
IBPT 66	2.41m	2.77k	IBPT 90	6.22bcd	7.33ab
IBPT 69	4.89fgh	5.96cde	IBPT 92	5.20efg	5.90cde

Means with different letters are significantly different at $P = 0.05$ level (Newman-Keuls test). The values represent the mean of host reaction rate, scale range 1 to 9, from three replications

and F10 pure lines tested in experiments I and II is shown in Table 1. Analyses of variance for the 16 parents, the 48 randomized (F10) pure lines and the whole set of 64 genotypes showed the occurrence of a highly significant genotype effect. Parental genotype 'DC²-30-N2' showed the highest level of partial resistance, and the two pure lines, 'IBPT-66' and 'IBPT-84', had the highest values for resistance to bacterial leaf streak in both experiments, whereas 'IBPT-89' was characterized by the highest susceptibility (Table 1).

There was thus a large genetic variability amongst the 64 genotypes tested for resistance to *X. campestris* pv. *cerealis*. These results suggest that the genotype is a major determinant of resistance to bacterial leaf streak. Genotype susceptibility and resistance to bacterial leaf streak has been also reported in wheat by Hagborg (1974), Duveiller (1990), Demir & Ustun (1992) and in triticale by Cunfer & Scolari (1982).

A summary of the data from experiments I and II is shown in Table 2. The difference between the mean of the disease scores of pure lines (PL) and that of the parents (P) was not significant in the first experiment but was significant in the second trial, indicating that the 48 randomly selected pure lines were probably not completely representative of the whole population.

Resistance (expressed as the lowest disease score) of the best parent (BP) was significantly superior to that of the best pure line (BPL) in the second experiment. This could be due to the limited number (48) of pure lines studied and the consequent failure to accumulate favourable resistance alleles. Increasing the number of pure lines is likely to result in the identification of

Table 2 Genetic gain for partial resistance to bacterial leaf streak in hexaploid wheat

Genotypes	Experiment I	Experiment II
Parents (P)	4.09	3.76
Pure lines (PL)	4.99	5.12
PL-P	0.90 NS	1.36*
Best Parent (BP)	1.93	1.58
Best Pure Line (BPL)	2.41	2.77
BPL-BP	0.48 NS	1.19*
LSD (0.05)	1.01	1.01

*Significant at $P = 0.05$ level; NS, not significant; LSD, least significant difference.

Table 3 Heritability of resistance to bacterial leaf streak in hexaploid winter wheat

Genotypes	Experiment I	Experiment II
δ^2G -Parents (P)	1.10	1.36
h^2	72.80	78.61
δ^2G -Pure Lines (L)	0.96	1.01
h^2	72.11	73.18
δ^2G -(P + L)	1.19	1.42
h^2	73.68	78.02

δ^2G , Genetic variance; h^2 , heritability (%).

genotypes that might prove to be as resistant as the best parents.

The heritability values of various cultivars show that the parents (P), the F10 pure lines (PL) as well as parents and pure lines (P + PL) were characterized by nearly the same heritability (Table 3). Heritability estimates were relatively high (70%); thus, it should be relatively easy to breed for resistance.

The genetics of resistance to bacterial diseases in wheat has been hardly investigated so far. Under artificial disease pressure in the field, resistance to *X. campestris* pv. *undulosa* in bread wheat appeared to be conditioned by five genes which differed in strength of resistance expression (Duveiller *et al.*, 1993). As far as partial resistance to bacterial leaf streak is concerned, diallel analysis of five genotypes and their 20 F1 hybrids also showed highly significant general and specific combining abilities, proving the importance of genetic effects in controlling the disease (Alizadeh *et al.*, 1994).

Analysis of variance for the eight genotypes of the third experiment (parents and F10 pure lines), representing a large variability for resistance and susceptibility, inoculated with the (IBLS20) strain of *X. campestris* pv. *cerealis* in the previous experiments, and four other strains showed a high significant genotype effect with all of strains studied (Table 4). Significant correlations between strains (r : 0.87–0.971) show that a genotype resistant or susceptible to one strain will have similar reactions with other strains. Milus & Chalkly (1994) also showed a high variability of the resistance of 19 wheat cultivars to 81 bacterial strains, and each cultivar had a similar susceptibility phenotype.

Finally, we may conclude that partial resistance to bacterial leaf streak in hexaploid wheat is genotype-dependent and highly heritable.

Table 4 Genetic variability for resistance of eight wheat genotypes to five strains of *Xanthomonas campestris* pv. *cerealis*

Varieties	Experiment II		Experiment III		
	IBLS 20	IBLS 22	IBLS 25	IBLS 29	IBLS 41
C61	1.60e	4.00f	2.00hi	5.00e	2.00hi
Dc ² 30N2	1.58e	3.00g	1.75i	2.25h	3.00g
Clement 2	5.24ab	9.00a	9.00a	8.00b	9.00a
C45	5.58a	8.00b	9.00a	6.50c	8.00b
IBPT 66	2.77k	3.90f	3.00g	3.95f	4.00f
IBPT 84	3.51jk	4.00f	4.00f	5.00e	6.00d
IBPT 89	7.67a	9.00a	8.00b	7.75b	9.00a
IBPT 90	7.33ab	9.00a	9.00a	9.00a	9.00a

Means with different letters are significantly different at $P = 0.05$ level (Newman–Keuls test).

The values represent the mean of host reaction rate, scale range 1 to 9 from three replications.

Correlations (ddl = 6): r (IBLS 20 and IBLS 22) = 0.93; r (IBLS 20 and IBLS 25) = 0.914; r (IBLS 20 and IBLS 41) = 0.943; r (IBLS 20 and IBLS 29) = 0.893; r (IBLS 22 and IBLS 25) = 0.971; r (IBLS 22 and IBLS 29) = 0.939; r (IBLS 22 and IBLS 41) = 0.943; r (IBLS 25 and IBLS 29) = 0.901; r (IBLS 25 and IBLS 41) = 0.962; r (IBLS 29 and IBLS 41) = 0.874; All 'r' values are significant at $P = 0.01$ level.

Bycrossing different genotypes, resistance to this disease should be transferred to susceptible ones.

REFERENCES

- Akhtar MA, Aslam M, 1986. *Xanthomonas campestris* pv. *undulosa* on wheat. *Rachis* **5**, 34–7.
- Alizadeh A, Rahimian H, 1989. Bacterial leaf streak of gramineae in Iran. *EPPO Bulletin* **19**, 113–7.
- Alizadeh A, Benetti V, Sarrafi A, Barrault G, Albertini L, 1994. Genetic analysis of partial resistance to an Iranian strain of bacterial leaf streak (*X. campestris* pv. *hordei*) in barley. *Plant Breeding* **113**, 323–6.
- Arabi MI, Barrault G, Sarrafi A, Albertini L, 1992. Variation in the resistance of barley cultivars and the pathogenicity of *Drechslera teres* f.sp. *maculata* and *D. teres* f.sp. *teres* isolated from France. *Plant Pathology* **41**, 180–86.
- Boosalis MG, 1952. The epidemiology of *Xanthomonas translucens* (J. J. & R.) Dowson on cereals and grasses. *Phytopathology* **42**, 387–95.
- Cunfer BM, Scolari BL, 1982. *Xanthomonas campestris* pv. *translucens* on triticale and other small grains. *Phytopathology* **72**, 683–6.
- Demir G, Ustun N, 1992. Studies on bacterial streak disease *Xanthomonas campestris* pv. *translucens* (Jones et al.) Dye of wheat and other gramineae. *Journal of Turkish Phytopathology* **21**, 33–40.
- Duveiller E, 1989. Research on '*Xanthomonas translucens*' of wheat and triticale at CIMMYT. *EPPO Bulletin* **19**, 97–103.
- Duveiller E, 1990. Seed detection of *Xanthomonas campestris* pv. *undulosa* using a modification of Wilbrink's agar. *Parasitica* **46**, 3–17.
- Duveiller E, Ginkel MV, Thijssen M, 1993. Genetic analysis of resistance to bacterial leaf streak caused by *Xanthomonas campestris* pv. *undulosa* in bread wheat. *Euphytica* **66**, 35–43.
- Forster RL, Schaad NW, 1988. Control of black chaff of wheat with seed treatment and a foundation seed health program. *Plant Disease* **72**, 935–8.
- Fourest E, Rehms LD, Sands DC, Bjarko M, Lund RE, 1990. Eradication of *Xanthomonas campestris* pv. *translucens* from barley seed with dry heat treatments. *Plant Disease* **74**, 816–8.
- Hagborg WAF, 1974. Notes of bacterial diseases of cereals and some other crop plants. *Canadian Plant Disease Survey* **54**, 129–51.
- Mehta YR, 1990. Management of *Xanthomonas campestris* pv. *undulosa* and *hordei* through cereal seed testing. *Seed Science and Technology* **8**, 467–76.
- Milus EA, Chalkly DB, 1994. Virulence of *Xanthomonas campestris* pv. *translucens* on selected wheat cultivars. *Plant Disease* **78**, 612–5.
- Milus EA, Mirolohi AF, 1994. Use of disease reactions to identify resistance in wheat to bacterial streak. *Plant Disease* **78**, 157–61.
- Sands DC, Mizrak G, Hall VN, Kim HK, Bockelman HE, Golden MJ, 1986. Seed transmitted bacterial diseases of cereals: epidemiology and control. *Arab Journal of Plant Protection* **4**, 125–7.
- Schaad NW, Forster RL, 1985. A semiselective agar medium for isolating *Xanthomonas campestris*

- pv. *translucens* from wheat seeds. *Phytopathology* **75**, 260–3.
- Sun FZ, Yang LYF, Wang YJ, 1988. Study on the control of black chaff of wheat TF-128. *Acta Phytophylactica Sinica* **15**, 79–83.
- Thomas G, Rousset M, Pichon M, Trottet M, Doussinaut G, Picard E, 1991. Méthodologie de population artificielle à 16 parents, base de cette étude méthodologique. *Agronomie* **11**, 359–68.
- Woo SH, Smith GS, 1962. A genetic study of leaf sheath, barbs, auricle hairs, and reaction to stem rust and 'black chaff' in crosses of (ND105 ND1) and ND113 with Conley wheat. *Botanic Bulletin Academia Sinica* **3**, 195–203.