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► To cite this version:

Chiara Lucchi, Anna Costa, Carmela Giordano, Giulia Curia, Marika Piat, et al.. Involvement of PPAR γ in the Anticonvulsant Activity of EP-80317, a Ghrelin Receptor Antagonist. *Frontiers in Pharmacology*, 2017, 8, 10.3389/fphar.2017.00676 . hal-02871324

HAL Id: hal-02871324

<https://hal.science/hal-02871324v1>

Submitted on 11 Jun 2021

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Involvement of PPAR γ in the Anticonvulsant Activity of EP-80317, a Ghrelin Receptor Antagonist

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Specialty section:

This article was submitted to
Neuropharmacology,
a section of the journal
Frontiers in Pharmacology

Received: 14 June 2017

Accepted: 08 September 2017

Published: 22 September 2017

Citation:

Lucchi C, Costa AM, Giordano C,
Curia G, Piat M, Leo G, Vinet J,
Brunel L, Fehrentz J-A, Martinez J,
Torsello A and Biagini G (2017)
Involvement of PPAR γ
in the Anticonvulsant Activity
of EP-80317, a Ghrelin Receptor
Antagonist. *Front. Pharmacol.* 8:676.
doi: 10.3389/fphar.2017.00676

Ghrelin, des-acyl ghrelin and other related peptides possess anticonvulsant activities. Although ghrelin and cognate peptides were shown to physiologically regulate only the ghrelin receptor, some of them were pharmacologically proved to activate the peroxisome proliferator-activated receptor gamma (PPAR γ) through stimulation of the scavenger receptor CD36 in macrophages. In our study, we challenged the hypothesis that PPAR γ could be involved in the anticonvulsant effects of EP-80317, a ghrelin receptor antagonist. For this purpose, we used the PPAR γ antagonist GW9662 to evaluate the modulation of EP-80317 anticonvulsant properties in two different models. Firstly, the anticonvulsant effects of EP-80317 were studied in rats treated with pilocarpine to induce *status epilepticus* (SE). Secondly, the anticonvulsant activity of EP-80317 was ascertained in the repeated 6-Hz corneal stimulation model in mice. Behavioral and video electrocorticographic (ECoG) analyses were performed in both models. We also characterized levels of immunoreactivity for PPAR γ in the hippocampus of 6-Hz corneally stimulated mice. EP-80317 predictably antagonized seizures in both models. Pretreatment with GW9662 counteracted almost all EP-80317 effects both in mice and rats. Only the effects of EP-80317 on power spectra of ECoGs recorded during repeated 6-Hz corneal stimulation were practically unaffected by GW9662 administration. Moreover, GW9662 alone produced a decrease in the latency of tonic-clonic seizures and accelerated the onset of SE in rats. Finally, in the hippocampus of mice treated with EP-80317 we found increased levels of PPAR γ immunoreactivity. Overall, these results support the hypothesis that PPAR γ is able to modulate seizures and mediates the anticonvulsant effects of EP-80317.

Keywords: 6-Hz corneal stimulation, EP-80317, ghrelin, peroxisome proliferator-activated receptor gamma, pilocarpine, status epilepticus, seizure

INTRODUCTION

Ghrelin and des-acyl ghrelin are neuroactive peptides prevalently produced in the stomach by X/A-like cells in rats, or P/D1 cells in humans (Rindi et al., 2002; Kojima, 2005; Chen et al., 2009). They regulate very important physiological functions, such as growth hormone secretion, food intake and metabolism by interacting with hypothalamic neurons (Nakazato et al., 2001). Ghrelin has also

been investigated for its possible role in regulating neuronal activity in other brain regions, since its receptor was found to be expressed out of the hypothalamus and especially in the hippocampus (Zigman et al., 2006). Interestingly, ghrelin was demonstrated to play an important role in learning and memory, cognitive functions in which the hippocampus is critically involved (Diano et al., 2006; Carlini et al., 2010; Li et al., 2013). Furthermore, ghrelin was also shown to modulate energy metabolism in areas other than the hypothalamus, as well as to be involved in the rewarding system of the brain (Egecioglu et al., 2010; Skibicka and Dickson, 2011; Skibicka et al., 2011; Perelló and Zigman, 2012). Overall, these evidences delineate a variety of physiological roles for ghrelin in the central nervous system (CNS).

Ghrelin has also been implicated in cerebral diseases. Specifically, it was shown to afford neuroprotection in models of cerebral ischemia (Spencer et al., 2013), Parkinson's disease (Bayliss and Andrews, 2013), and *status epilepticus* (SE) (Xu et al., 2009; Lucchi et al., 2013). Interestingly, this last neuroprotective effect occurred at a dosage unable to alter the epileptic activity. However, ghrelin was also found to modulate seizures induced by several, different approaches (Casillas-Espinosa et al., 2012; Portelli et al., 2012a; Kovac and Walker, 2013; Clynen et al., 2014; Curia et al., 2014; Dobolyi et al., 2014). Independent studies suggested an anticonvulsant activity of ghrelin in experimental paradigms by which seizures were induced using pentylenetetrazole (Obay et al., 2007), penicillin (Aslan et al., 2009), kainic acid (Lee et al., 2010), and pilocarpine (Portelli et al., 2012b). According to Lee et al. (2010), the anticonvulsant effects of ghrelin could be related to the interaction of this peptide with its established receptor. However, the ghrelin receptor was also demonstrated to possess a proconvulsant activity in basal condition, whereas the observed anticonvulsant properties of ghrelin ligands were explained by the induction of ghrelin receptor internalization (Portelli et al., 2012b).

Other than ghrelin, ghrelin receptor agonists such as hexarelin (Biagini et al., 2011), capromorelin (Portelli et al., 2012b), and JMV-1843 (Coppens et al., 2016), the inverse agonists A778193 and [D-Arg¹, D-Phe⁵, D-Trp^{7,9}, Leu¹¹] substance P (Portelli et al., 2012b), as well as the antagonist EP-80317 (Biagini et al., 2011), were all proven to be anticonvulsants. These data may suggest that all ghrelin receptor ligands could be able to desensitize neurons expressing the ghrelin receptor, so to block its proconvulsant activity (Portelli et al., 2012b). However, this hypothesis could not be supported in the case of the ghrelin receptor antagonist JMV-2959, which did not alter the induction of SE by pilocarpine (Biagini et al., 2011). Alternatively, it could be hypothesized that the variety of ghrelin-related peptides able to induce anticonvulsant effects may interact with different receptors that share a common anticonvulsant activity. Indeed, the existence of multiple ghrelin receptors has been hypothesized immediately following the discovery of ghrelin receptor (Davenport et al., 2005; Müller et al., 2015). In particular, it was evident that the protective effects exerted by ghrelin, des-acyl ghrelin and other related peptides in presence of an inflammatory reaction, especially in the cardiovascular system, were independent of

the ghrelin receptor (Bujold et al., 2009, 2013; Bulgarelli et al., 2009).

EP-80317 (Haic-D-Mrp-D-Lys-Trp-D-Phe-Lys-NH₂) is a hexapeptide with a primary structure similar to that of hexarelin, from which it differs for substitutions in the first and third amino acid residues causing the loss of the growth hormone (GH)-releasing properties possessed by the original peptide (Momany et al., 1981). EP-80317 is pharmacologically active on macrophages, and its activity on these cells was found to be dependent on activation of the peroxisome proliferator-activated receptor gamma (PPAR γ). However, the ability to activate PPAR γ is also common to ghrelin, des-acyl ghrelin and other ghrelin-related peptides, as all of them are able to interact with the CD36 scavenger receptor (Bujold et al., 2009, 2013). In the CNS, CD36 appears to be expressed prevalently although not exclusively in microglia (Glezer et al., 2009), whereas PPAR γ is found in neurons of various cerebral regions, including those known to be involved in the generation and propagation of seizure activity, such as the hippocampus and the piriform cortex (Moreno et al., 2004; Warden et al., 2016). For this reason, we tried to elucidate whether PPAR γ could be involved in the anticonvulsant effects of EP-80317. To this purpose, we designed experiments in which administration of EP-80317 was challenged by pretreatment with the PPAR γ inhibitor GW9662 (Wong et al., 2015), in rodents exposed to seizure induction. Additionally, we characterized the expression of PPAR γ in the hippocampus of mice treated with EP-80317.

MATERIALS AND METHODS

Animals and Treatments

A total of thirty-seven adult male Sprague-Dawley rats (Harlan, San Pietro al Natisone, Italy), ranging 260–270 g of body weight, and 66 4-week-old male CD-1 mice (Charles River, Calco, Italy) were used in this study. All animals were housed in a specific pathogen-free facility under controlled environment with *ad libitum* access to water and food.

EP-80317 was obtained through conventional solid phase synthesis, dissolved in a physiologic solution and administered through intraperitoneal (i.p.) injection (330 μ g/kg). GW9662 and dimethyl sulfoxide (DMSO) were purchased from Sigma-Aldrich Chemicals (St. Louis, MO, United States). GW9662 was dissolved in DMSO and diluted with saline and then administered (2 mg/kg, i.p.) (Rani et al., 2016).

All experiments were in compliance with the European Directive 2010/63/EU and carried out according to the national guidelines on animal experimental research of the Italian Ministry of Health (DM 126/2011 – B and DM 92/2013). The University of Modena and Reggio Emilia Ethics Committee approved the study protocol. All efforts were made to refine procedures to improve the welfare and to reduce the number of animals that were used.

Experimental Design

For the pilocarpine model, we subdivided rats into four groups for behavior and video electrocorticographic (ECoG) recordings:

(i) 10 control rats received saline (1 ml/kg, i.p.), (ii) 10 rats received EP-80317 alone, (iii) seven rats received GW9662 alone, and (iv) 10 rats received GW9662 10 min prior to EP-80317.

For the repeated 6-Hz corneal stimulation model, we considered a number of mice ($n = 39$) for behavior and video-ECoG recordings, and the others ($n = 27$) for immunohistochemical and immunofluorescence analysis. In particular, for behavior and video-ECoG recordings, we subdivided mice into the following groups: (i) nine control mice received saline (1 ml/kg, i.p.), (ii) 12 mice received EP-80317 alone (330 μ g/kg, i.p.), (iii) nine mice received GW9662 alone (2 mg/kg, i.p.), and (iv) nine mice received GW9662 10 min prior to EP-80317, both at the already indicated doses. For immunohistochemical analyses, four control mice were used to determine basal levels of the investigated marker and were neither treated nor stimulated. Still, they were handled and exposed to the same procedure as the others. Twelve were saline-treated mice, while the other 11 mice received an i.p. EP-80317 injection. Moreover, these animals were used for qualitative immunofluorescence experiments but, in this case, we used only three mice per treatment group.

Pilocarpine Protocol and Behavioral Analysis

Pilocarpine (Sigma–Aldrich, Milan, Italy) was injected i.p. (380 mg/kg) to induce SE. The pilocarpine injection was preceded by methylscopolamine (1 mg/kg, i.p.; Sigma–Aldrich) to prevent the peripheral effects of cholinergic stimulation (Curia et al., 2008). EP-80317, GW9662 alone, or saline were injected i.p. 20 min after methylscopolamine and 10 min before pilocarpine. In the group of rats treated with GW9662 and then with EP-80317, the first drug was administered 10 min prior to the second one. In all rats experiencing SE, diazepam (20 mg/kg, i.p.; Hospira Italia, Naples, Italy) was injected 10 min after the SE onset to guarantee survival. This procedure stops convulsive seizures, leaving non-convulsive SE unaltered for hours (Gualtieri et al., 2012).

Drug-induced responses in rats were observed directly and graded by blind to treatment expert raters. Seizures were graded according to a modification of the Racine's scale (Racine, 1972). In particular, we considered: (i) non-convulsive seizures, ranked as stage 1 to stage 3; (ii) convulsive seizures, ranked as stage 4 to stage 5; (iii) SE (stage 6), considered as the stage in which rats either did not recover normal behavior (i.e., exploration, grooming, or motor reaction to stimuli) between one seizure and the other, or in which they displayed continuous shaking for more than 5 min (Lowenstein, 1999). Rats were sacrificed 72 h after SE to assess the absence of cerebral lesions caused by electrode implant, and the presence of injuries developed during the SE.

Corneal Stimulation Protocol and Behavioral Analysis

Mice were stimulated once and allowed to recover for 72 h before being stimulated up to four sessions. Corneal stimulation was performed as previously described (Giordano et al., 2015, 2016). Briefly, a topical eye anesthetic (0.4% oxybuprocaine

hydrochloride eye drops, Novesin, Novartis, Switzerland) was applied 10 min before stimulation. All mice received the injection of EP-80317, GW9662, or saline 10 min before each session of corneal stimulation. When GW9662 preceded EP-80317, it was administered 20 min before each session. Stimulation (fixed current intensity of 32 mA, pulse width of 0.2 ms, duration of 3 s, frequency of 6 Hz) was delivered via corneal electrodes connected to a stimulator (ECT Unit 5780; Ugo Basile, Comerio, Italy).

Seizure severity was ranked according to the following score: (i) stunned posture and eye blinking; (ii) head nodding, Straub tail and repetitive rhythmic movements (stereotypies) such as chewing; (iii) unilateral or alternating forelimb clonus; (iv) generalized tonic-clonic convulsions without loss of posture and rearings; (v) generalized tonic-clonic convulsions with loss of posture. Seizure scores were first recorded through direct observation, then reanalyzed on video recordings to quantify the duration of behavioral changes by an investigator unaware of the stimulation session. Recovery from seizures was defined as the reappearance of a normal exploratory behavior.

Electrodes Implantation for Electroencephalographic (ECoG) Recordings

For electrode implantation, anesthesia was induced with volatile isoflurane (4% induction and 1–2% maintenance) in rats, whereas mice were anesthetized with ketamine (150 μ g/g, i.p.) + xylazine (10 μ g/g, i.p.). After deep anesthesia was reached (assessed by deep breath, loss of tail and eye reflexes), the skin was shaved, disinfected with povidone-iodine 10% (Betadine[®] skin solution; Meda Pharma, Milano, Italy), cut and opened to expose the skull. Guiding holes were drilled and epidural electrodes (stainless steel $\varnothing = 1$ mm; PlasticsOne, Roanoke, VA, United States) were implanted in frontal (bregma 0 mm, 3.5 mm lateral from midline in rats and bregma 0 mm, 3 mm lateral from midline in mice) and occipital cortices (bregma -6.5 mm, 3.5 mm lateral from midline in rats and bregma -3.5 mm, 3 mm lateral from midline in mice) of both hemispheres. One electrode was implanted below the lambda on the midline in all animals and used as a reference. Electrodes were connected through steel wire to terminal gold pins (Bilaney Consultant GmbH, Düsseldorf, Germany) inserted in a plastic pedestal (PlasticsOne) cemented on heads. At the end of the surgery, gel containing 2.5 g lidocaine chloride, 0.5 g neomycin sulfate and 0.025 g fluocinolone acetonide (Neuflan[®] gel; Molteni Farmaceutici, Scandicci, FI, Italy) was applied to reduce acute pain and risk of infection. All animals were monitored until complete recovery from anesthesia, and were housed in single cages without grids or environmental enrichments to reduce risk of headset loss.

Video ECoG Recordings

For brain activity recording, animals were placed in cages with paper filter cover that allowed cable connection between headset and preamplifiers. Electrical brain activity was digitally filtered (0.3 Hz high-pass, 500 Hz low-pass), acquired at 1 kHz per channel, and stored on a personal computer after the mathematical subtraction of traces of recording electrodes

from trace of reference electrode, using a PowerLab8/30 amplifier connected to 4 BioAmp preamplifiers (ADInstruments; Dunedin, Otago, New Zealand). Videos were digitally captured through a camera connected to the computer and synchronized to the ECoG traces through LabChart 7 PRO internal trigger. Four days after electrode implantation, all animals were connected to the recording system and received the sequence of treatments described above. To facilitate handling and pharmacological manipulations, recordings were stopped, animals were temporarily disconnected while being injected and reconnected soon after. In particular, rats were recorded for 20 min after methylscopolamine administration, 10 min after EP-80317, GW9662 or saline injection, 10 min after SE onset and at least 2 h after diazepam administration. Rats that did not experience SE were recorded for at least 90 min after pilocarpine injection, until signs of normal behavior (exploring, grooming) reappeared. Mice were recorded after EP-80317, GW9662 or saline injection and 6-Hz corneal stimulation, until signs of normal behavior reappeared.

ECoG Analysis

Electrocorticographic traces were digitally filtered offline (band-pass: high 50 Hz, low 1 Hz) and manually analyzed using LabChart 7 PRO software (AD Instruments) by expert raters.

In rats, we measured the duration of each electrographic seizure, characterized as epileptiform ECoG patterns with trains of 150–250 ms long spikes with amplitudes at least twice as the previous 2 s baseline. Seizures occurring within 5 s of each other were defined as one epileptic event.

In mice, we quantified the ECoG signal of each seizure performing a power spectral analysis, that described the distribution of signal power over frequency (Dressler, 2004). In particular, the power spectrum was obtained by fast Fourier transformation of the ECoG waveforms (LabChart 7 PRO, ADInstruments) and used to obtain the mean power spectra for the experimental groups. The maximum value of power spectra (peak value) was considered for each mouse. Peak values of power, recorded from frontal and occipital electrodes, were evaluated to investigate whether modifications of ictal activity had occurred from the first to the fourth stimulation.

Immunohistochemistry

Control mice and mice receiving up to one or three sessions of 6-Hz corneal stimulation and treated with saline or EP-80317 were used for tissue analysis. Mice deeply anesthetized with isoflurane were transcardially perfused with phosphate buffered saline (PBS, pH 7.4) followed by Zamboni's fixative (pH 6.9), 14–17 h after the seizure. Brains were post-fixed at 4°C in the same fixative for 24 h, cryoprotected in 15 and 30% sucrose solutions (Vinet et al., 2016) and stored at –80°C until used. Horizontal sections of 50 μ m were cut using a freezing sliding microtome (Leica SM2000 R; Leica, Nussloch, Germany). For immunostaining, sections were washed in Tris-buffered saline (TBS) and incubated in 3% H₂O₂ in TBS (30 min) to quench endogenous peroxidase activity. Following another washing step, sections were blocked 1 h in TBS containing 2% bovine serum albumin, 0.3% Triton X-100 (Tx) and 5% normal goat serum.

Sections were then placed at 4°C with a polyclonal rabbit anti-PPAR γ (Ab209350; Abcam, Cambridge, United Kingdom, dilution 1:2000, 48 h). After washing, sections were incubated for 1 h with a biotinylated anti-rabbit secondary antibody (Vector Laboratories, Burlingame, CA, United States; 1:200), and later with the avidin-biotin-peroxidase complex (Elite ABC Kit; Vector Laboratories, Burlingame, CA, United States). The immunostaining was performed in 0.05% 3,3-diaminobenzidine tetrahydrochloride for 5 min (DAB, Sigma–Aldrich, Milan, Italy) and developed by adding 0.03% H₂O₂. Finally, sections were washed again in TBS, mounted on gelatin-coated slides and cover slipped with Eukitt (Eukitt®, O. Kindler GmbH & Co., Freiburg, Germany).

Image Analysis

Immunostained sections related to bregma from –8.04 mm to –5.04 mm for hippocampal CA1 and CA3 regions, and hilus of the dentate gyrus (DH) were analyzed using an Axioskop microscope (Carl Zeiss Vision GmbH, Munchen, Germany) equipped with a 10X objective. Images were digitally captured by a Sony CCD-IRIS B–W video camera, along the ventrodorsal direction of the brain (approximately 6–7 serial horizontal sections separated by 0.5 mm). A mouse brain atlas-C57BL/6J horizontal was used to assess brain sections. The image analysis was carried out using the KS300 software (Carl Zeiss Vision GmbH), as previously described (Biagini et al., 2005; Curia et al., 2013; Giordano et al., 2015, 2016). PPAR γ immunoreactivity was measured as field area values, corresponding to the addition of areas of the specific profiles obtained after discrimination from background staining. Background values in each section were obtained from areas devoid of specific immunostaining, such as the angular bundle. All measurements were taken bilaterally and the final values represent the left–right average.

Double Immunofluorescence

We performed a PPAR γ /67-kDa glutamate decarboxylase (GAD67) qualitative double immunofluorescence to assess PPAR γ expression in interneurons. PPAR γ expression was further evaluated in different interneuron subsets by PPAR γ /parvalbumin (PV), PPAR γ /somatostatin-28 (SOM) or PPAR γ /vasoactive intestinal peptide (VIP) qualitative double immunofluorescences. For PPAR γ /GAD67 immunofluorescence, sections were washed in TBS at room temperature and unmasking in sodium citrate at 98°C and permeabilized for 1 h in TBS/0.02% Triton X-100 containing 5% normal goat serum. Concerning PPAR γ /PV, PPAR γ /SOM and PPAR γ /VIP immunofluorescences, sections were washed in TBS at room temperature and permeabilized for 1 h in TBS/0.1% Triton X-100 containing 5% normal goat serum. Then, sections were incubated for 48 h with primary antibodies: rabbit anti-PPAR γ (Ab209350; Abcam, Cambridge, United Kingdom, dilution 1:50), mouse anti-GAD67 (MAB5406; Chemicon International, Billerica, MA, United States, dilution 1:200), mouse anti-PPAR γ (E-8, sc-7273; Santa Cruz Biotechnology, Santa Cruz, CA, United States, dilution 1/50), rabbit anti-PV (Ab11427; Abcam, Cambridge, MA, United States, dilution 1:2000), rabbit anti-SOM (20089; Immunostar, Hudson, WI,

United States, dilution 1/1000) and rabbit anti-VIP (20077; Immunostar, Hudson, WI, United States, dilution 1/500). Following a washing step in TBS, sections were incubated for 3 h at room temperature with secondary antibodies Alexa Fluor 488-labeled goat anti-mouse antibody (A-11001; Invitrogen, Carlsbad, CA, United States, dilution 1:500) and Alexa Fluor 594-labeled goat anti-rabbit (A-11012; Invitrogen, Carlsbad, CA, United States, dilution 1:500). After rinsing sections in TBS, brain sections were placed on gelatinized glass slides, dried, and mounted with Mowiol after incubation with DAPI. Images were acquired using a Leica TCS SP2 (Leica Laser Technik, Heidelberg, Germany) confocal microscope. All images were taken using a 40X magnification.

Statistics

Chi-Square test (χ^2) was used to compare the percentage (%) of rats experiencing SE in the various treatment groups. Pairwise comparisons with Fisher's exact test (using α correction) were used to establish differences among groups. Additionally, the time interval (min) required to develop seizures and SE in the various groups was analyzed by Kruskal-Wallis test. Dunn's test was used for multiple comparisons. One-way analysis of variance (ANOVA) was used to compare electrographic seizure duration (sec) in each treatment groups.

The χ^2 test was also used to compare mice experiencing seizures with loss of posture in the various treatment groups. Again, pairwise comparisons with Fisher's exact test (using α correction) were used to establish differences among groups. Electrocorticographic recordings, analyzed as peak values of power spectra, were compared using two-way ANOVA, considering treatments as the between-factor and sessions as the within-factor. The Holm-Šidák test was used for multiple comparisons. All statistical analyses were performed using Sigmaplot 11 (Systat Software, San Jose, CA, United States). Unless otherwise indicated, results are shown as mean \pm standard error of the mean (SEM); p -values lower than 0.05 were considered as statistically significant.

RESULTS

EP-80317 Reduced the Percentage of Animals Developing SE after Pilocarpine Injection

Response to pilocarpine was assessed by expert raters who directly annotated the motor responses observed during seizure development according to the modified Racine's scale (Racine, 1972; Lucchi et al., 2013), and by offline analysis of video-ECoG recordings. Four ($n = 1$ saline group, $n = 2$ EP-80317 group, and $n = 1$ GW9662 group) animals died during continuous tonic-clonic seizures and were excluded from the analysis. Consistently with previous findings (Biagini et al., 2011), pilocarpine induced SE in 100% of saline-treated rats. Instead, only 50% of rats treated with EP-80317 developed SE ($p < 0.05$ vs. saline-treated rats, Fisher's exact test). Additionally, we also found that the percentages of rats developing SE after pilocarpine

administration in groups treated with GW9662 or GW9662+EP-80317 (respectively, 83% and 80%) were not significantly different from that observed in saline-treated rats (Table 1).

GW9662 Accelerated the Development of Convulsive Seizure and SE in the Pilocarpine Model

We calculated the time interval required to develop the first stage 1-3 and stage 4-5 seizure after pilocarpine administration, as median and interquartile range (IRQ) values. Moreover, we calculated the time interval required to develop SE (Table 2). No significant differences were found when comparing the latency periods of the first stage 1-3 seizure in the various treatment groups. Instead, the latency periods for developing the first stage 4-5 seizure were significantly different in GW9662-treated or GW9662+EP-80317-treated rats compared to, respectively, saline-treated rats ($p < 0.05$ for both groups, Dunn's test) and the EP-80317 group of treatment ($p < 0.05$). In addition, the latency periods for developing SE after the first stage 4-5 seizure were significantly shorter in GW9662-treated and GW9662+EP-80317-treated rats compared to saline-treated rats ($p < 0.05$).

Drug Treatments Did Not Affect Seizure Duration in Pilocarpine-Treated Rats

We measured the average seizure duration in traces obtained from ECoG recordings (Figure 1) performed in the various groups of treatment. No differences were found for animals treated, respectively, with saline (65.32 ± 8.75 s) or GW9662 (67.15 ± 10.95 s). Although the EP-80317 group presented a lower seizure duration (45.75 ± 9.24 s), this difference did not reach a statistically significant level. Mean seizure duration was also lower (49.91 ± 12.64 s) in GW9662+EP-80317 but, again, not enough to be statistically relevant.

EP-80317 Transiently Reduced the Seizure Severity and Duration in the Repeated 6-Hz Corneal Stimulation Model

Experimenters first visually monitored seizures induced through 6-Hz corneal stimulation and their pharmacological modulations. The first seizure duration was significantly shorter in mice treated with EP-80317 compared to control mice ($p < 0.05$, Holm-Šidák test), thus confirming the anticonvulsant properties of this molecule (Biagini et al., 2011;

TABLE 1 | Percentage (%) of rats developing *status epilepticus* (SE) in the various treatment groups.

Treatments	Percentage (%) of rats developing SE
Saline ($n = 9$)	100%
EP-80317 (EP, $n = 8$)	50%*
GW9662 (GW, $n = 6$)	83%
GW+EP ($n = 10$)	80%

* $p < 0.05$, EP-80317 vs. saline; Fisher's exact test.

TABLE 2 | Median and interquartile range values of latency periods for developing seizure and *status epilepticus* (SE) in the various treatment groups.

Latency (min)	Saline	EP-80317 (EP)	GW9662 (GW)	GW+EP
From pilocarpine injection to first stage 1-3 seizure	2.00 (1.75–3.50)	4.00 (3.00–11.75)	3.00 (2.00–4.00)	3.00 (3.00–4.00)
From pilocarpine injection to first stage 4-5 seizure	9.00 (6.75–10.25)	18.00 (8.00–21.25)	3.00 (2.00–4.00)*§	3.00 (3.00–4.00)*§
From pilocarpine injection to SE	17.00 (16.00–18.50)	13.00 (10.75–25.00)	14.00 (11.00–14.50)	16.50 (14.50–21.75)
From first stage 4-5 seizure to SE	8.00 (8.00–10.50)	5.50 (2.75–6.75)	3.00 (2.50–4.50)#	3.50 (3.00–6.50)#

* $p < 0.05$, GW9662 vs. saline and GW9662+EP-80317 vs. saline; Dunn's test. § $p < 0.05$, GW9662 vs. EP-80317 and GW9662+EP-80317 vs. EP-80317. # $p < 0.05$, GW9662 vs. saline and GW9662+EP-80317.

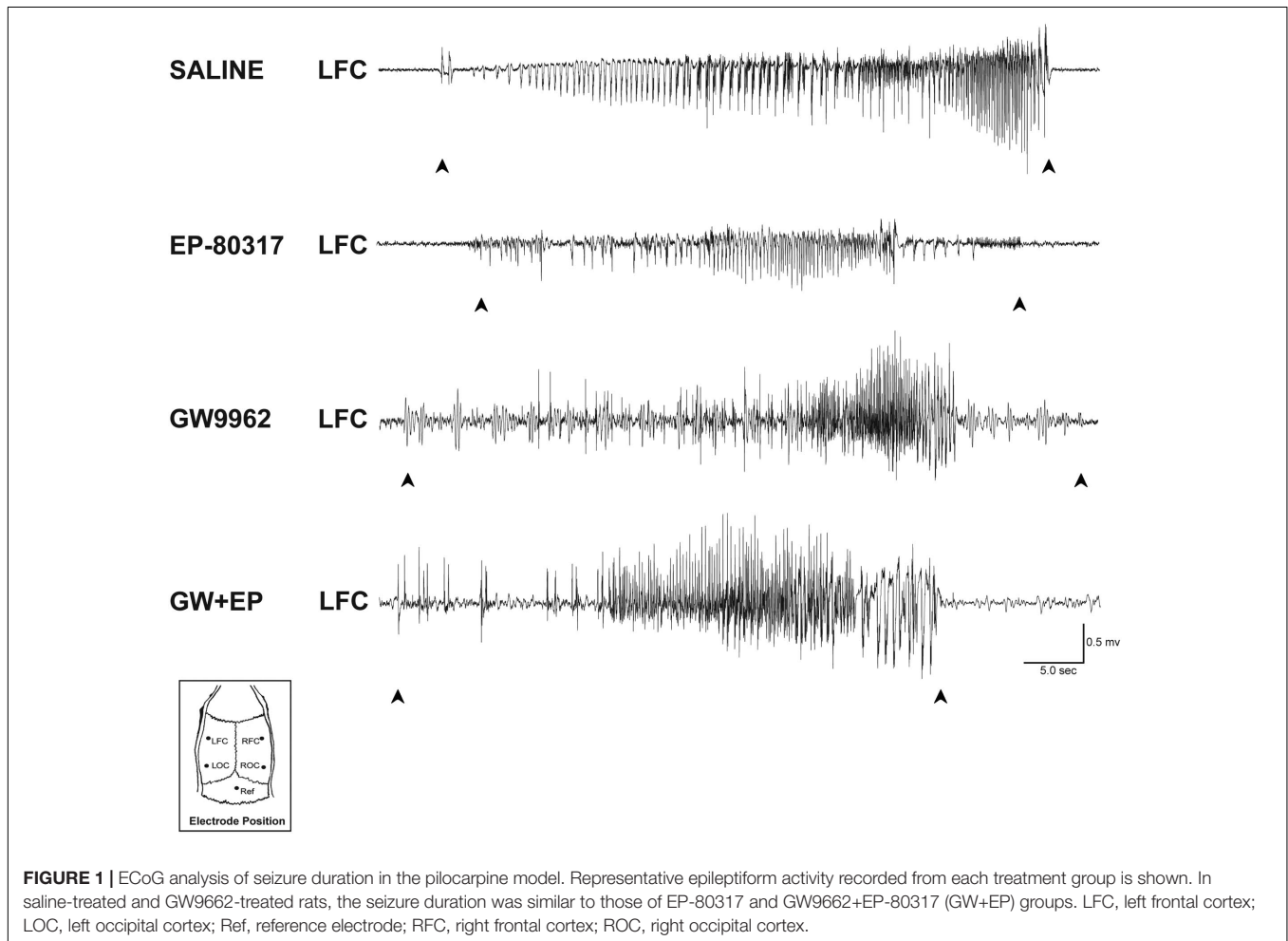


FIGURE 1 | ECoG analysis of seizure duration in the pilocarpine model. Representative epileptiform activity recorded from each treatment group is shown. In saline-treated and GW9662-treated rats, the seizure duration was similar to those of EP-80317 and GW9662+EP-80317 (GW+EP) groups. LFC, left frontal cortex; LOC, left occipital cortex; Ref, reference electrode; RFC, right frontal cortex; ROC, right occipital cortex.

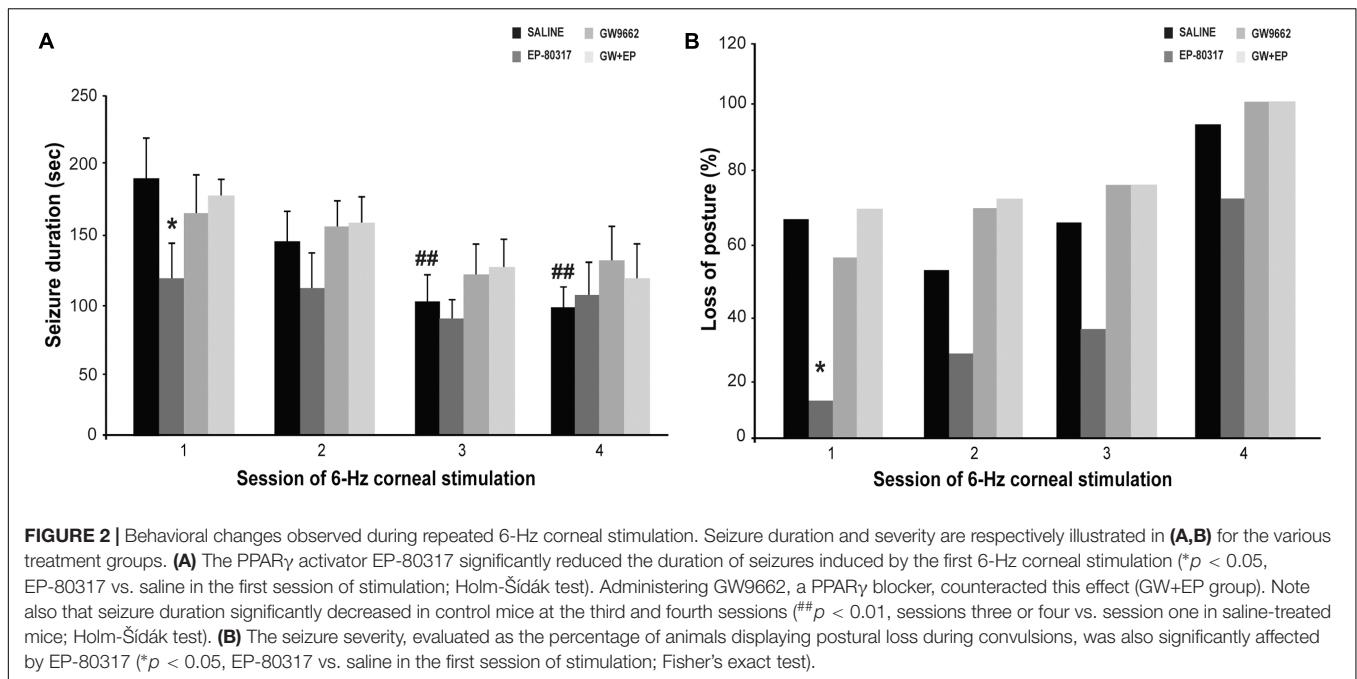
Giordano et al., 2016). No differences were noticed by comparing controls to mice treated with GW9662 alone or prior to EP-80317 (Figure 2A). Similarly seizure severity, defined as the percentage of mice displaying loss of posture during seizures, was also less pronounced in the presence of EP-80317 ($p < 0.05$, EP-80317 vs. saline in the first session of stimulation; Fisher's exact test). No differences were observed by comparing the other groups with saline-treated mice (Figure 2B).

As expected (Giordano et al., 2015), seizures significantly shortened by repeating 6-Hz corneal stimulation ($p < 0.01$, sessions three and four vs. session one in saline-treated mice; Holm-Šídák test). Consequently, no differences between mice treated with saline and EP-80317 were observed from the second

session onward (Figure 2A). Moreover, the severity of seizures progressed in all groups, including mice treated with EP-80317 (Figure 2B), which confirmed the appearance of resistance to the effects of EP-80317 (Giordano et al., 2016). Consistently, in the fourth session of 6-Hz corneal stimulation no differences were present within the various treatment groups.

EP-80317 Prevented the Increase in Power Spectra of Seizures Induced by Repeated 6-Hz Corneal Stimulation

Seizures induced by 6-Hz corneal stimulation and their pharmacological modulations were further monitored through



video ECoG recordings. In particular, we compared power spectra of seizures recorded from frontal (not shown) and occipital electrodes (Figure 3A). As previously reported (Giordano et al., 2015), power peaks did not change for ictal events recorded through frontal cortex electrodes, in all groups, whereas in the occipital recording we noticed the presence of an epileptogenic process. In particular, we found a significant increase in power peaks recorded from occipital electrodes in the fourth session, both in saline-treated and GW9662-treated mice ($p < 0.01$ for saline-treated mice; $p < 0.05$ for GW9662-treated mice, session four vs. session one; Holm-Šidák test; Figure 3B). Conversely, power peaks in mice treated with EP-80317 did not change from the first to the fourth session of 6-Hz corneal stimulation, suggesting an antiepileptogenic effect. This effect was not completely prevented by administering GW9662 prior to EP-80317, although a trend was present ($p = 0.09$). Interestingly, significantly lower values were found in the fourth session of mice treated with EP-80317 compared to saline-treated mice ($p < 0.05$; Figure 3B).

EP-80317 Significantly Increased PPAR γ Immunoreactivity in the Hippocampus of Mice Exposed to the First 6-Hz Corneal Stimulation

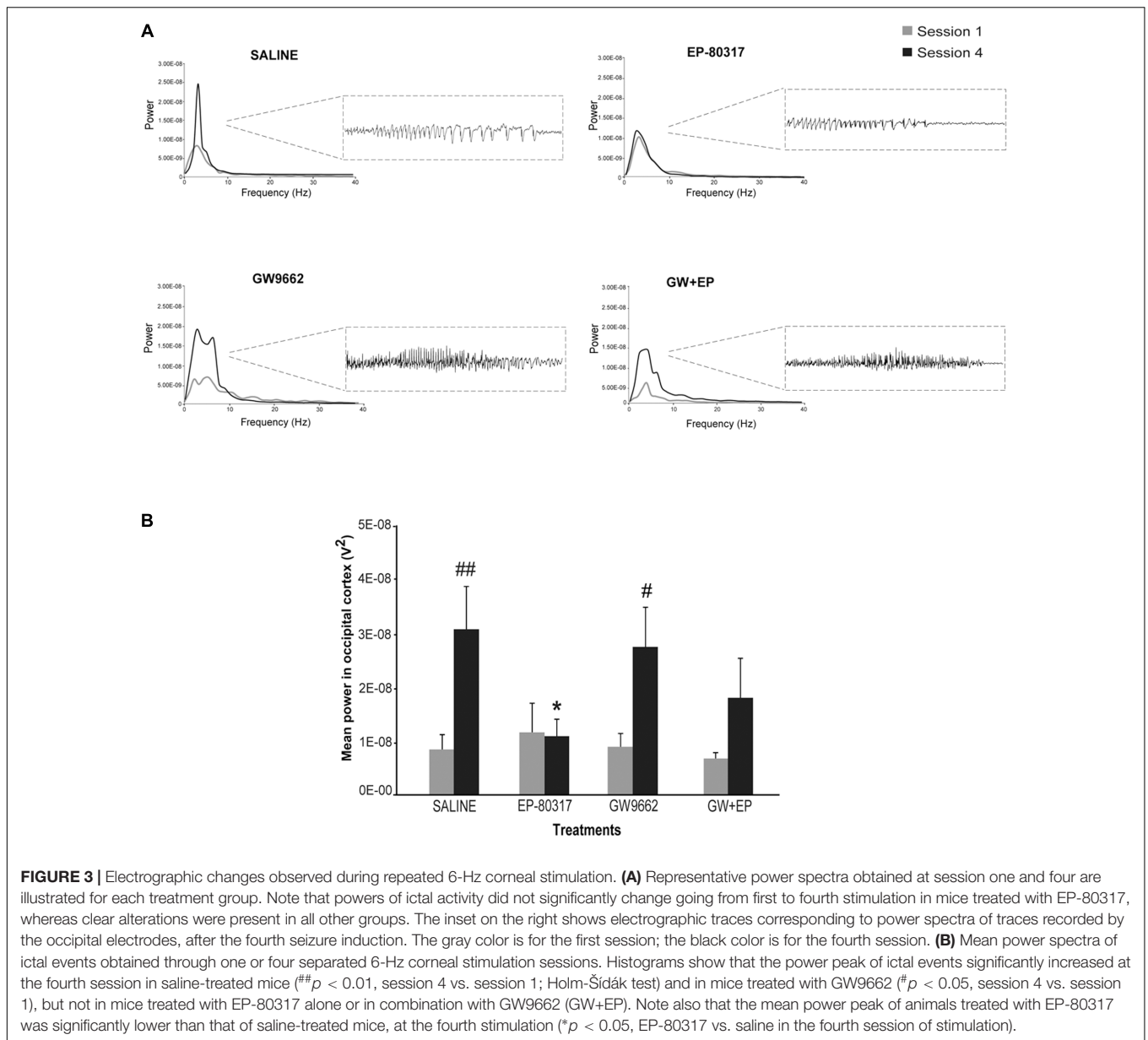
We evaluated the effects of repeated exposure to 6-Hz corneal stimulation on PPAR γ immunoreactivity in the *stratum pyramidale* of CA1 (Figure 4), in both *stratum pyramidale* and *stratum radiatum/lacunosum-moleculare* of CA3 (Figure 5), and finally in the DH (Figure 6). First, we quantified the changes occurring after the first and third session of seizure induction in pyramidal cells of the hippocampus and in mossy cells of

the DH. PPAR γ levels were scantily detectable in all sampled regions of unstimulated control mice. PPAR γ immunoreactivity was not significantly changed in regions of interest of saline-treated mice. Notably, in animals treated with EP-80317 we observed a prominent increase in PPAR γ levels after the first 6-Hz corneal stimulation, reaching a statistically significant level in all the considered regions ($p < 0.05$ vs. controls, in CA1; $p < 0.001$ vs. controls and saline-treated mice, in CA3 and DH; Holm-Šidák test; Figures 4–6). After the third seizure induction, PPAR γ immunoreactivity returned to basal levels in all hippocampal regions.

To evaluate the response in interneurons, we analyzed the changes occurring in the *stratum radiatum/lacunosum-moleculare* of the CA3 (Table 3). Again, PPAR γ levels were barely detectable in unstimulated control mice and saline-treated mice, but a significant increase in PPAR γ levels was found in mice treated with EP-80317 after the first 6-Hz corneal stimulation ($p < 0.01$ vs. controls; $p < 0.05$ vs. saline-treated mice, Holm-Šidák test; Table 3). Consistently, PPAR γ levels returned to basal levels after the third seizure induction.

PPAR γ Is Expressed in Different Subsets of Interneurons

To confirm that PPAR γ was expressed in interneurons, we performed a double immunofluorescence experiment with PPAR γ and GAD67 antibodies. We found that not all interneurons coexpressed GAD67 and PPAR γ immunoreactivity (Figure 7, top panels). Thus, to assess the subset of interneurons expressing PPAR γ immunoreactivity, we evaluated PPAR γ /PV, PPAR γ /SOM and PPAR γ /VIP colabeling. PPAR γ -positive cells were frequently observed within PV interneurons (Figure 7, in which arrows indicate colabeling), and less frequently identified



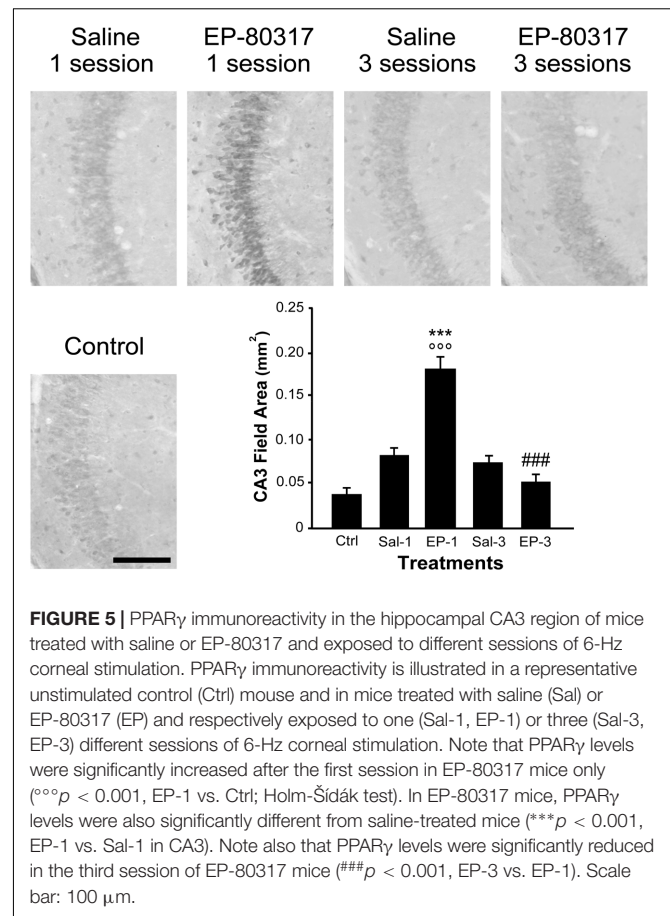
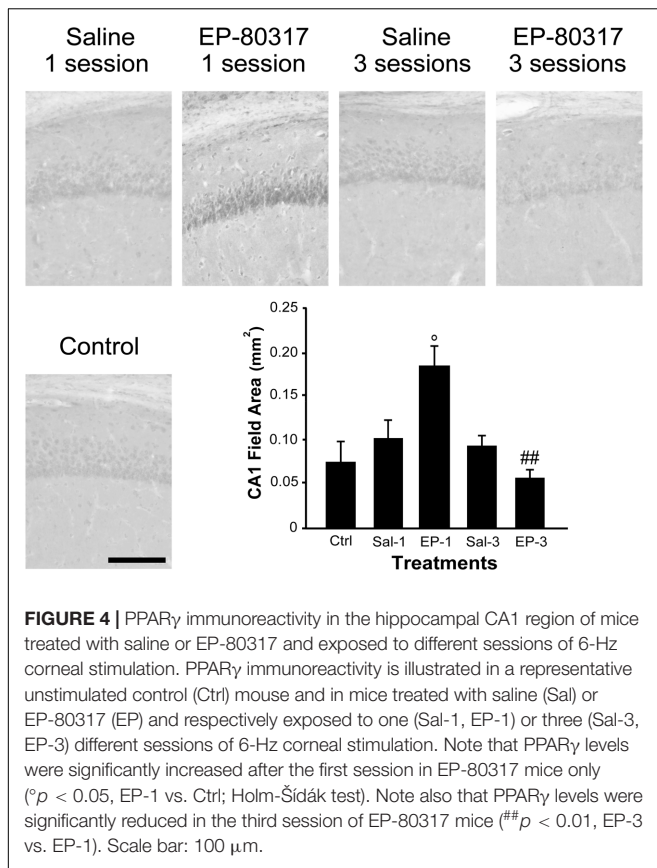
within SOM interneurons. In contrast, PPAR γ never colocalized with VIP in interneurons (**Figure 7**, in which arrowheads indicate lack of colabeling).

DISCUSSION

In the present investigation we confirmed that EP-80317 displays anticonvulsant effects in the pilocarpine model and in the repeated 6-Hz corneal stimulation model. As major findings of our investigation, we also demonstrated that GW9662, an inhibitor of PPAR γ activity, is able to counteract the anticonvulsant effects of EP-80317 in both the animal models, with the notable exception of the antiepileptogenic properties disclosed by analyzing the ECoG power spectrum in mice.

We also observed a proconvulsant activity of GW9662 when analyzing the latency period of stage 4-5 seizures, and the latency period of SE onset after the first stage 4-5 seizure in the pilocarpine model. Interestingly, PPAR γ immunoreactivity was transiently increased in pyramidal cells and interneurons belonging to PV and SOM subclasses, in coincidence with the anticonvulsant effects of EP-80317 in 6-Hz corneally stimulated mice.

We previously established that the ghrelin receptor antagonist EP-80317 is an anticonvulsant in the pilocarpine model (Biagini et al., 2011), as well as in the repeated 6-Hz corneal stimulation model (Giordano et al., 2016). We also hypothesized that its anticonvulsant activity could not be explained by an involvement of the ghrelin receptor, as we did not observe any anticonvulsant activity of either the ghrelin receptor

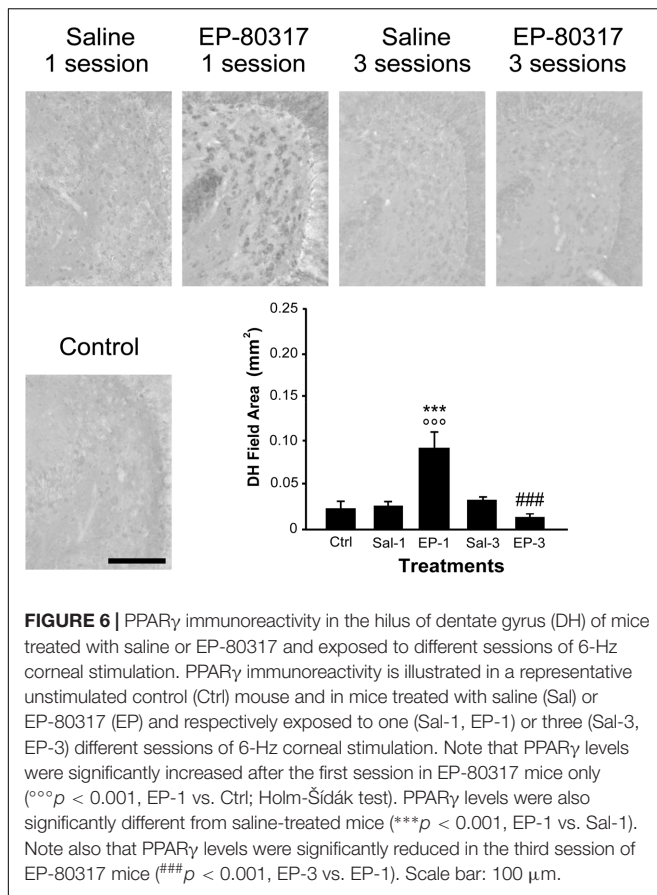


antagonist JMV-2959 (Biagini et al., 2011), nor of ghrelin, and of the ghrelin receptor agonist JMV-1843 (Biagini et al., 2011; Lucchi et al., 2013), all administered i.p. 10 min before injecting pilocarpine. At odds with our data, however, other investigators consistently showed that ghrelin produces some anticonvulsant effects in different seizure models (Portelli et al., 2012a,b). Furthermore, JMV-1843 also was found to be an anticonvulsant when used at high doses (Coppens et al., 2016). So, technical reasons, such as the more direct intracerebroventricular route of ghrelin administration in some cases (Portelli et al., 2012b), the earlier timing of ghrelin injection in others (Obay et al., 2007; Lee et al., 2010; Portelli et al., 2012b), and the different doses of the tested ghrelin receptor agonist (Coppens et al., 2016) may help in understanding why ghrelin receptor agonists in our hands did not display anticonvulsant properties (Biagini et al., 2011; Lucchi et al., 2013). Alternatively, it could be proposed that different receptors are involved in the anticonvulsant effects of ghrelin and its related peptides.

EP-80317 was found to activate PPAR γ , as well as its cascade in apolipoprotein E-deficient mice (Bujold et al., 2009, 2013; Bulgarelli et al., 2009). Although PPAR γ was mainly localized in the liver and in adipocytes, in which it was characterized for its role in metabolism regulation (Demers et al., 2008; Li et al., 2014; Gorga et al., 2017), it was also found that PPAR γ agonists such as rosiglitazone or pioglitazone display antiseizure

effects (Adabi Mohazab et al., 2012; Hong et al., 2013). It has also been proposed that PPAR γ plays a key role in the antiseizure effects of cannabinoid agonists (Payandemehr et al., 2015), as well as in the anticonvulsant effects of the ketogenic diet (Jeong et al., 2011; Simeone et al., 2017). In agreement with these suggestions, our data support the involvement of PPAR γ in the anticonvulsant effects of EP-80317. Consistently, by inhibiting PPAR γ with the antagonist GW9662, the effects of rosiglitazone on seizure-like activity induced by Mg $^{2+}$ -free medium in hippocampal slices were partially prevented (Wong et al., 2015). In line with these *in vitro* studies, the effects of EP-80317 were counteracted by GW9662 in our *in vivo* experiments and, additionally, we have identified a proconvulsant effect of GW9662 in the pilocarpine model. Indeed, as GW9662 produced a decrease in the latency period of tonic-clonic seizures and accelerated the onset of SE, a question arises on whether the GW9662 and EP-80317 combination resulted in the simple summation of their independent pro/anticonvulsant effects or not, at least in the pilocarpine model.

The suggested ability to modulate seizures of PPAR γ could be explained by the fact that this nuclear receptor is also expressed in peripheral immune cells, microglia, and neurons (Moreno et al., 2004; Bujold et al., 2009; Warden et al., 2016). Especially by acting on immune cells, PPAR γ exerts a negative modulation of macrophages and microglia reactivity (Moreno et al., 2004;



Bujold et al., 2009; Warden et al., 2016), resulting in an anti-inflammatory action in the nervous tissue (Jeong et al., 2011). This effect may be important as several lines of evidence suggest that leukocytes and microglia are critical in establishing the onset of SE in the pilocarpine model (Fabene et al., 2008; Vezzani et al., 2015; Vinet et al., 2016). In particular, treatment with antibodies against leukocyte α_4 integrin, which is involved in inflammation, successfully prevented the onset of SE in pilocarpine-treated mice (Fabene et al., 2008). Additional evidences indicate that inflammation is required to establish the SE in pilocarpine-treated rodents (Marchi et al., 2009; Vezzani et al., 2015). Overall, these findings suggest that EP-80317 activation of the PPAR γ pathway in immune cells (Bujold et al., 2009, 2013; Bulgarelli et al., 2009) may have contributed to prevent the SE in pilocarpine-treated rats. However, we cannot rule out the possibility that other ghrelin analogs could share the same mechanism found for EP-80317. Indeed, further experiments are required to establish if PPAR γ may mediate the anticonvulsant effects of other ghrelin receptor agonists or antagonists.

The effects of EP-80317 on PPAR γ could be dependent on a direct modulation of the neuronal activity. As a matter of fact, EP-80317 markedly increased PPAR γ immunoreactivity in hippocampal pyramidal neurons and also in interneurons of 6-Hz corneally stimulated mice. It is interesting to notice that we have identified PPAR γ in interneurons and described its expression in different interneuron subpopulations for the first

time. Recently, the role of interneurons in seizure initiation and propagation has become more controversial (de Curtis and Avoli, 2016). In particular, the different interneuron subclasses were suggested to play specific roles in modulating seizures (Khoshkhoo et al., 2017). Here, we found that PV and SOM but not VIP interneurons expressed PPAR γ and, thus, were probably involved in the anticonvulsant effects of EP-80317. Interestingly, optogenetic inhibition of VIP interneurons affected both the seizure threshold and duration, whereas PV and SOM interneuron inhibition apparently reduced only the seizure duration (Khoshkhoo et al., 2017). According to these data, it is possible that the effects on seizure duration observed with EP-80317 in 6-Hz stimulated mice could be partly mediated by modulation of PV and SOM interneurons. It could also be of interest to evaluate other PPAR γ modulators, such as rosiglitazone or pioglitazone, in the pilocarpine model and, especially, in the repeated 6-Hz corneal stimulation model to examine their effects in exactly the same experimental conditions in which we characterized EP-80317.

Although all the EP-80317 anticonvulsant effects were counteracted by GW9662, a notable exception was found by analyzing the ECoG power spectra of 6-Hz corneally stimulated mice. In particular, we observed that the power of ECoG recordings increased significantly in saline-treated and GW9662-treated mice, but not in mice receiving EP-80317 or GW9662+EP-80317. These findings suggest that the already proposed antiepileptogenic activity of EP-80317 (Giordano et al., 2016) may be independent of PPAR γ activation. This hypothesis is in line with the time course of PPAR γ induction in the hippocampus, which has been characterized by a transient increase at the first session of seizure induction returning to basal levels at the third session. So, PPAR γ was presumably restored at basal levels when the effect of EP-80317 on the power of ECoG recordings was still present. Although we could not establish if this last finding was related to antagonism at the ghrelin receptor, it was certainly worth to further investigate this pharmacological activity of EP-80317 in view of the well-recognized need of antiepileptogenic drugs to prevent the appearance of epilepsy in subjects exposed to this risk, as in the case of patients which develop SE and, for this reason (Santamarina et al., 2015), may become affected by epilepsy.

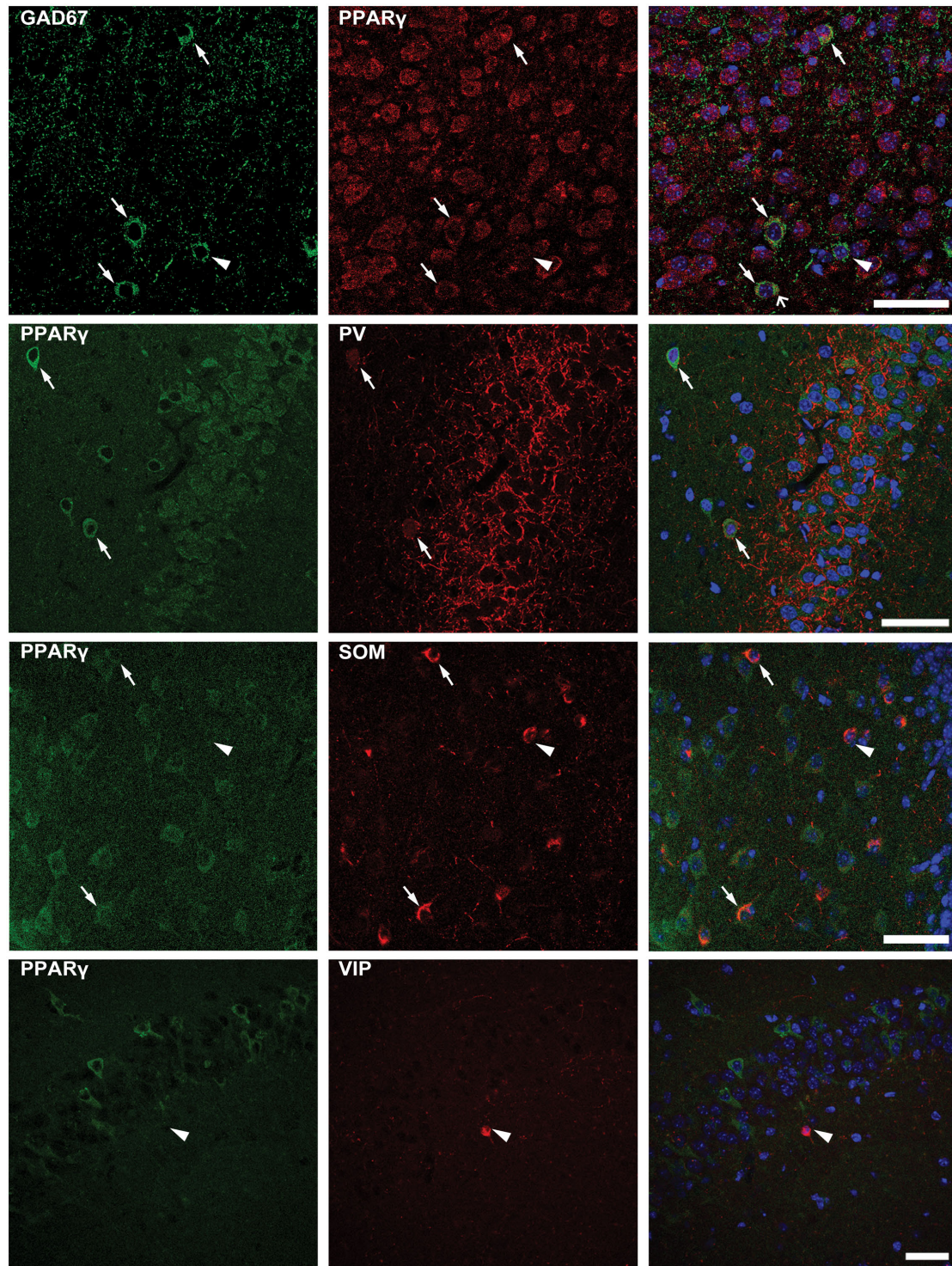


FIGURE 7 | Double immunofluorescence of interneurons expressing PPAR γ . Photomicrographs illustrating co-labeling with glutamate decarboxylase 67 (GAD67, green) and PPAR γ (red) in CA1 of a representative mouse are shown in top panels. Double immunofluorescence revealed the co-expression of GAD67 and PPAR γ in some (arrows), but not all interneurons (arrowhead point to a PPAR γ negative interneuron). Scale bar: 50 μ m. Photomicrographs illustrating co-labeling of PPAR γ (green) and parvalbumin (PV, red), somatostatin (SOM, red), or vasoactive intestinal peptide (VIP, red) in the hippocampus of mice exposed to repeated 6-Hz corneal stimulation are sequentially shown from the second row to bottom. Note that double immunofluorescence revealed the co-labeling of PPAR γ /PV in CA3, PPAR γ /SOM in the hilus of dentate gyrus (DH) (arrows), but not PPAR γ /VIP in CA1 (arrowhead). Scale bar: 50 μ m.

AUTHOR CONTRIBUTIONS

GB is responsible for the experimental design, contributed to the acquisition, analysis and interpretation of all the experiments, drafted and revised the work. CL, AC, and CG performed the experiments and contributed to acquisition and analysis of most of the results and revised the work. GC, MP, GL, and JV performed part of the experiments and revised the work. AT, LB, J-AF, and JM provided the anticonvulsant, contributed to data interpretation and revised the work.

REFERENCES

- Adabi Mohazab, R., Javadi-Paydar, M., Delfan, B., and Dehpour, A. R. (2012). Possible involvement of PPAR-gamma receptor and nitric oxide pathway in the anticonvulsant effect of acute pioglitazone on pentylenetetrazole-induced seizures in mice. *Epilepsy Res.* 101, 28–35. doi: 10.1016/j.eplepsres.2012.02.015
- Aslan, A., Yildirim, M., Ayyildiz, M., Güven, A., and Agar, E. (2009). The role of nitric oxide in the inhibitory effect of ghrelin against penicillin-induced epileptiform activity in rat. *Neuropeptides* 43, 295–302. doi: 10.1016/j.npep.2009.05.005
- Bayliss, J. A., and Andrews, Z. B. (2013). Ghrelin is neuroprotective in Parkinson's disease: molecular mechanisms of metabolic neuroprotection. *Ther. Adv. Endocrinol. Metab.* 4, 25–36. doi: 10.1177/2042018813479645
- Biagini, G., D'Arcangelo, G., Baldelli, E., D'Antuono, M., Tancredi, V., and Avoli, M. (2005). Impaired activation of CA3 pyramidal neurons in the epileptic hippocampus. *Neuromol. Med.* 7, 325–342. doi: 10.1385/NMM:7:4:325
- Biagini, G., Torsello, A., Marinelli, C., Gualtieri, F., Vezzali, R., Coco, S., et al. (2011). Beneficial effects of desacyl-ghrelin, hexarelin and EP-80317 in models of status epilepticus. *Eur. J. Pharmacol.* 670, 130–136. doi: 10.1016/j.ejphar.2011.08.020
- Bujold, K., Mellal, K., Zoccal, K. F., Rhainds, D., Brissette, L., Febbraio, M., et al. (2013). EP 80317, a CD36 selective ligand, promotes reverse cholesterol transport in apolipoprotein E-deficient mice. *Atherosclerosis* 229, 408–414. doi: 10.1016/j.atherosclerosis.2013.05.031
- Bujold, K., Rhainds, D., Jossart, C., Febbraio, M., Marleau, S., and Ong, H. (2009). CD36-mediated cholesterol efflux is associated with PPARgamma activation via a MAPK-dependent COX-2 pathway in macrophages. *Cardiovasc. Res.* 83, 457–464. doi: 10.1093/cvr/cvp118
- Bulgarelli, I., Tamiasso, L., Bresciani, E., Rapetti, D., Caporali, S., Lattuada, D., et al. (2009). Desacyl-ghrelin and synthetic GH-secretagogues modulate the production of inflammatory cytokines in mouse microglia cells stimulated by β -amyloid fibrils. *J. Neurosci. Res.* 87, 2718–2727. doi: 10.1002/jnr.22088
- Carlini, V. P., Gherzi, M., Schiöth, H. B., and de Barioglio, S. R. (2010). Ghrelin and memory: differential effects on acquisition and retrieval. *Peptides* 31, 1190–1193. doi: 10.1016/j.peptides.2010.02.021
- Casillas-Espinosa, P. M., Powell, K. L., and O'Brien, T. J. (2012). Regulators of synaptic transmission: roles in the pathogenesis and treatment of epilepsy: regulators of synaptic transmission. *Epilepsia* 53, 41–58. doi: 10.1111/epi.12034
- Chen, C.-Y., Asakawa, A., Fujimiya, M., Lee, S.-D., and Inui, A. (2009). Ghrelin gene products and the regulation of food intake and gut motility. *Pharmacol. Rev.* 61, 430–481. doi: 10.1124/pr.109.001958
- Clynen, E., Swijsen, A., Rajmakers, M., Hoogland, G., and Rigo, J.-M. (2014). Neuropeptides as targets for the development of anticonvulsant drugs. *Mol. Neurobiol.* 50, 626–646. doi: 10.1007/s12035-014-8669-x
- Coppens, J., Aourz, N., Walrave, L., Fehrentz, J.-A., Martinez, J., De Bundel, D., et al. (2016). Anticonvulsant effect of a ghrelin receptor agonist in 6Hz corneally kindled mice. *Epilepsia* 57, e195–e199. doi: 10.1111/epi.13463
- Curia, G., Gualtieri, F., Bartolomeo, R., Vezzali, R., and Biagini, G. (2013). Resilience to audiogenic seizures is associated with p-ERK1/2 dephosphorylation in the subiculum of Fmr1 knockout mice. *Front. Cell. Neurosci.* 7:46. doi: 10.3389/fncel.2013.00046
- Curia, G., Longo, D., Biagini, G., Jones, R. S. G., and Avoli, M. (2008). The pilocarpine model of temporal lobe epilepsy. *J. Neurosci. Methods* 172, 143–157. doi: 10.1016/j.jneumeth.2008.04.019

FUNDING

This study was supported by the Italian Ministry of Health (grant RF-2010-2309921 to GB).

ACKNOWLEDGMENT

JV is fellow of the Umberto Veronesi Foundation (<https://www.fondazioneveronesi.it/>).

- Curia, G., Lucchi, C., Vinet, J., Gualtieri, F., Marinelli, C., Torsello, A., et al. (2014). Pathophysiological of mesial temporal lobe epilepsy: is prevention of damage antiepileptogenic? *Curr. Med. Chem.* 21, 663–688. doi: 10.2174/0929867320666131119152201
- Davenport, A. P., Bonner, T. I., Foord, S. M., Harmor, A. J., Neubig, R. R., Pin, J.-P., et al. (2005). International Union of Pharmacology. LVI. Ghrelin receptor nomenclature, distribution, and function. *Pharmacol. Rev.* 57, 541–546. doi: 10.1124/pr.57.4.1
- de Curtis, M., and Avoli, M. (2016). GABAergic networks jump-start focal seizures. *Epilepsia* 57, 679–687. doi: 10.1111/epi.13370
- Demers, A., Rodrigue-Way, A., and Tremblay, A. (2008). Hexarelin signaling to PPAR γ in metabolic diseases. *PPAR Res.* 2008, 1–9. doi: 10.1155/2008/364784
- Diano, S., Farr, S. A., Benoit, S. C., McNay, E. C., da Silva, I., Horvath, B., et al. (2006). Ghrelin controls hippocampal spine synapse density and memory performance. *Nat. Neurosci.* 9, 381–388. doi: 10.1038/nn1656
- Dobolyi, A., Kékesi, K. A., Juhász, G., Székely, A. D., Lovas, G., and Kovács, Z. (2014). Receptors of peptides as therapeutic targets in epilepsy research. *Curr. Med. Chem.* 21, 764–787. doi: 10.2174/0929867320666131119154018
- Dressler, O. (2004). Awareness and the EEG power spectrum: analysis of frequencies. *Br. J. Anaesth.* 93, 806–809. doi: 10.1093/bja/aeh270
- Egecioglu, E., Jerlhag, E., Salomé, N., Skibicka, K. P., Haage, D., Bohlooly-Y, M., et al. (2010). Ghrelin increases intake of rewarding food in rodents: ghrelin and food reward. *Addict. Biol.* 15, 304–311. doi: 10.1111/j.1369-1600.2010.00216.x
- Fabene, P. F., Mora, G. N., Martinello, M., Rossi, B., Merigo, F., Ottoboni, L., et al. (2008). A role for leukocyte-endothelial adhesion mechanisms in epilepsy. *Nat. Med.* 14, 1377–1383. doi: 10.1038/nm.1878
- Giordano, C., Costa, A. M., Lucchi, C., Leo, G., Brunel, L., Fehrentz, J.-A., et al. (2016). Progressive seizure aggravation in the repeated 6-Hz corneal stimulation model is accompanied by marked increase in hippocampal p-ERK1/2 immunoreactivity in neurons. *Front. Cell. Neurosci.* 10:281. doi: 10.3389/fncel.2016.00281
- Giordano, C., Vinet, J., Curia, G., and Biagini, G. (2015). Repeated 6-Hz corneal stimulation progressively increases FosB/ Δ FosB levels in the lateral amygdala and induces seizure generalization to the hippocampus. *PLOS ONE* 10:e0141221. doi: 10.1371/journal.pone.0141221
- Glezer, I., Bittencourt, J. C., and Rivest, S. (2009). Neuronal expression of Cd36, Cd44, and Cd83 antigen transcripts maps to distinct and specific murine brain circuits. *J. Comp. Neurol.* 517, 906–924. doi: 10.1002/cne.22185
- Gorga, A., Rindone, G. M., Regueira, M., Pellizzari, E. H., Camberos, M. C., Cigorraga, S. B., et al. (2017). PPAR γ activation regulates lipid droplet formation and lactate production in rat Sertoli cells. *Cell Tissue Res.* doi: 10.1007/s00441-017-2615-y [Epub ahead of print].
- Gualtieri, F., Curia, G., Marinelli, C., and Biagini, G. (2012). Increased perivascular laminin predicts damage to astrocytes in CA3 and piriform cortex following chemoconvulsive treatments. *Neuroscience* 218, 278–294. doi: 10.1016/j.neuroscience.2012.05.018
- Hong, S., Xin, Y., HaiQin, W., GuiLian, Z., Ru, Z., ShuQin, Z., et al. (2013). The PPAR γ agonist rosiglitazone prevents neuronal loss and attenuates development of spontaneous recurrent seizures through BDNF/TrkB signaling following pilocarpine-induced status epilepticus. *Neurochem. Int.* 63, 405–412. doi: 10.1016/j.neuint.2013.07.010
- Jeong, E. A., Jeon, B. T., Shin, H. J., Kim, N., Lee, D. H., Kim, H. J., et al. (2011). Ketogenic diet-induced peroxisome proliferator-activated receptor- γ activation

- decreases neuroinflammation in the mouse hippocampus after kainic acid-induced seizures. *Exp. Neurol.* 232, 195–202. doi: 10.1016/j.expneurol.2011.09.001
- Khoshkhoo, S., Vogt, D., and Sohal, V. S. (2017). Dynamic, cell-type-specific roles for GABAergic interneurons in a mouse model of optogenetically inducible seizures. *Neuron* 93, 291–298. doi: 10.1016/j.neuron.2016.11.043
- Kojima, M. (2005). Ghrelin: structure and function. *Physiol. Rev.* 85, 495–522. doi: 10.1152/physrev.00012.2004
- Kovac, S., and Walker, M. C. (2013). Neuropeptides in epilepsy. *Neuropeptides* 47, 467–475. doi: 10.1016/j.npep.2013.10.015
- Lee, J., Lim, E., Kim, Y., Li, E., and Park, S. (2010). Ghrelin attenuates kainic acid-induced neuronal cell death in the mouse hippocampus. *J. Endocrinol.* 205, 263–270. doi: 10.1677/JOE-10-0040
- Li, E., Chung, H., Kim, Y., Kim, D. H., Ryu, J. H., Sato, T., et al. (2013). Ghrelin directly stimulates adult hippocampal neurogenesis: implications for learning and memory. *Endocr. J.* 60, 781–789. doi: 10.1507/endocrj.EJ13-0008
- Li, Z., Xu, G., Qin, Y., Zhang, C., Tang, H., Yin, Y., et al. (2014). Ghrelin promotes hepatic lipogenesis by activation of mTOR-PPAR signaling pathway. *Proc. Natl. Acad. Sci. U.S.A.* 111, 13163–13168. doi: 10.1073/pnas.1411571111
- Lowenstein, D. H. (1999). Status epilepticus: an overview of the clinical problem. *Epilepsia* 40, S3–S8. doi: 10.1111/j.1528-1157.1999.tb00872.x
- Lucchi, C., Curia, G., Vinet, J., Gualtieri, F., Bresciani, E., Locatelli, V., et al. (2013). Protective but not anticonvulsant effects of ghrelin and JMV-1843 in the pilocarpine model of status epilepticus. *PLOS ONE* 8:e72716. doi: 10.1371/journal.pone.0072716
- Marchi, N., Fan, Q., Ghosh, C., Fazio, V., Bertolini, F., Betto, G., et al. (2009). Antagonism of peripheral inflammation reduces the severity of status epilepticus. *Neurobiol. Dis.* 33, 171–181. doi: 10.1016/j.nbd.2008.10.002
- Momany, F. A., Bowers, C. Y., Reynolds, G. A., Chang, D., Hong, A., and Newlander, K. (1981). Design, synthesis, and biological activity of peptides which release growth hormone in vitro*. *Endocrinology* 108, 31–39. doi: 10.1210/endo-108-1-31
- Moreno, S., Farioli-Vecchioli, S., and Cerù, M. P. (2004). Immunolocalization of peroxisome proliferator-activated receptors and retinoid X receptors in the adult rat CNS. *Neuroscience* 123, 131–145. doi: 10.1016/j.neuroscience.2003.08.064
- Müller, T. D., Nogueiras, R., Andermann, M. L., Andrews, Z. B., Anker, S. D., Argente, J., et al. (2015). Ghrelin. *Mol. Metab.* 4, 437–460. doi: 10.1016/j.molmet.2015.03.005
- Nakazato, M., Murakami, N., Date, Y., Kojima, M., Matsuo, H., Kangawa, K., et al. (2001). A role for ghrelin in the central regulation of feeding. *Nature* 409, 194–198. doi: 10.1038/35051587
- Obay, B. D., Tasdemir, E., Tümer, C., Bilgin, H. M., and Sermet, A. (2007). Antiepileptic effects of ghrelin on pentylenetetrazole-induced seizures in rats. *Peptides* 28, 1214–1219. doi: 10.1016/j.peptides.2007.04.003
- Payandemehr, B., Ebrahimi, A., Gholizadeh, R., Rahimian, R., Varastehmoradi, B., Gooshe, M., et al. (2015). Involvement of PPAR receptors in the anticonvulsant effects of a cannabinoid agonist, WIN 55,212-2. *Prog. Neuropsychopharmacol. Biol. Psychiatry* 57, 140–145. doi: 10.1016/j.pnpbp.2014.11.005
- Perelló, M., and Zigman, J. M. (2012). The role of ghrelin in reward-based eating. *Biol. Psychiatry* 72, 347–353. doi: 10.1016/j.biopsych.2012.02.016
- Portelli, J., Michotte, Y., and Smolders, I. (2012a). Ghrelin: an emerging new anticonvulsant neuropeptide. *Epilepsia* 53, 585–595. doi: 10.1111/j.1528-1167.2012.03423.x
- Portelli, J., Thielemans, L., Ver Donck, L., Loyens, E., Coppens, J., Aourz, N., et al. (2012b). Inactivation of the constitutively active ghrelin receptor attenuates limbic seizure activity in rodents. *Neurotherapeutics* 9, 658–672. doi: 10.1007/s13311-012-0125-x
- Racine, R. J. (1972). Modification of seizure activity by electrical stimulation. II. Motor seizure. *Electroencephalogr. Clin. Neurophysiol.* 32, 281–294. doi: 10.1016/0013-4694(72)90177-0
- Rani, N., Bharti, S., Bhatia, J., Nag, T. C., Ray, R., and Arya, D. S. (2016). Chrysin, a PPAR- γ agonist improves myocardial injury in diabetic rats through inhibiting AGE-RAGE mediated oxidative stress and inflammation. *Chem. Biol. Interact.* 250, 59–67. doi: 10.1016/j.cbi.2016.03.015
- Rindi, G., Necchi, V., Savio, A., Torsello, A., Zoli, M., Locatelli, V., et al. (2002). Characterisation of gastric ghrelin cells in man and other mammals: studies in adult and fetal tissues. *Histochem. Cell Biol.* 117, 511–519. doi: 10.1007/s00418-002-0415-1
- Santamarina, E., Gonzalez, M., Toledo, M., Sueiras, M., Guzman, L., Rodriguez, N., et al. (2015). Prognosis of status epilepticus (SE): relationship between SE duration and subsequent development of epilepsy. *Epilepsy Behav.* 49, 138–140. doi: 10.1016/j.yebeh.2015.04.059
- Simeone, T. A., Matthews, S. A., Samson, K. K., and Simeone, K. A. (2017). Regulation of brain PPAR γ 2 contributes to ketogenic diet anti-seizure efficacy. *Exp. Neurol.* 287, 54–64. doi: 10.1016/j.expneurol.2016.08.006
- Skibicka, K. P., and Dickson, S. L. (2011). Ghrelin and food reward: the story of potential underlying substrates. *Peptides* 32, 2265–2273. doi: 10.1016/j.peptides.2011.05.016
- Skibicka, K. P., Hansson, C., Alvarez-Crespo, M., Friberg, P. A., and Dickson, S. L. (2011). Ghrelin directly targets the ventral tegmental area to increase food motivation. *Neuroscience* 180, 129–137. doi: 10.1016/j.neuroscience.2011.02.016
- Spencer, S., Miller, A., and Andrews, Z. (2013). The role of ghrelin in neuroprotection after ischemic brain injury. *Brain Sci.* 3, 344–359. doi: 10.3390/brainsci3010344
- Vezzani, A., Dingleline, R., and Rossetti, A. O. (2015). Immunity and inflammation in status epilepticus and its sequelae: possibilities for therapeutic application. *Expert Rev. Neurother.* 15, 1081–1092. doi: 10.1586/14737175.2015.1079130
- Vinet, J., Vainchtein, I. D., Spano, C., Giordano, C., Bordini, D., Curia, G., et al. (2016). Microglia are less pro-inflammatory than myeloid infiltrates in the hippocampus of mice exposed to status epilepticus. *Glia* 64, 1350–1362. doi: 10.1002/glia.23008
- Warden, A., Truitt, J., Merriman, M., Ponomareva, O., Jameson, K., Ferguson, L. B., et al. (2016). Localization of PPAR isotypes in the adult mouse and human brain. *Sci. Rep.* 6:27618. doi: 10.1038/srep27618
- Wong, S.-B., Cheng, S.-J., Hung, W.-C., Lee, W.-T., and Min, M.-Y. (2015). Rosiglitazone suppresses *in vitro* seizures in hippocampal slice by inhibiting presynaptic glutamate release in a model of temporal lobe epilepsy. *PLOS ONE* 10:e0144806. doi: 10.1371/journal.pone.0144806
- Xu, J., Wang, S., Lin, Y., Cao, L., Wang, R., and Chi, Z. (2009). Ghrelin protects against cell death of hippocampal neurons in pilocarpine-induced seizures in rats. *Neurosci. Lett.* 453, 58–61. doi: 10.1016/j.neulet.2009.01.067
- Zigman, J. M., Jones, J. E., Lee, C. E., Saper, C. B., and Elmquist, J. K. (2006). Expression of ghrelin receptor mRNA in the rat and the mouse brain. *J. Comp. Neurol.* 494, 528–548. doi: 10.1002/cne.20823

Conflict of Interest Statement: AT, GB, and other inventors share a patent on the possible therapeutic use of growth hormone secretagogues for epileptic disorders (patent 0001399610 – 2013; http://www.uibm.gov.it/uibm/dati/Avanzata.aspx?load=info_list_uno&id=1800628&table=Invention&#ancoraSearch).

The other authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

The reviewer FW and handling Editor declared their shared affiliation.

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