

Taxonomic revision of the genus Xenogryllus Bolívar, 1890 (Orthoptera, Gryllidae, Eneopterinae, Xenogryllini)

Ranjana Jaiswara, Jiajia Dong, Libin Ma, Haisheng Yin, Tony Robillard

▶ To cite this version:

Ranjana Jaiswara, Jiajia Dong, Libin Ma, Haisheng Yin, Tony Robillard. Taxonomic revision of the genus Xenogryllus Bolívar, 1890 (Orthoptera, Gryllidae, Eneopterinae, Xenogryllini). Zootaxa, 2019, 4545 (3), pp.301. 10.11646/zootaxa.4545.3.1. hal-02866078

HAL Id: hal-02866078 https://hal.science/hal-02866078

Submitted on 12 Jun2020

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

- 1 Taxonomic revision of the genus *Xenogryllus* (Orthoptera, Gryllidae,
- 2 Eneopterinae)
- 3
- 4 **Page headlines:** Revision of *Xenogryllus*
- 5
- 6 Ranjana Jaiswara* (1,2)
- 7 Jiajia Dong* (1,3)
- 8 Libin Ma (3)
- 9 Haisheng Yin (4)
- 10 Tony Robillard (1)
- 11
- 12 1_Institut de Systématique, Evolution et Biodiversité (ISYEB), Muséum national
- 13 d'Histoire naturelle, CNRS, Sorbonne Université, EPHE, 57 rue Cuvier, CP 50, 75231
- 14 Paris Cedex 05, France
- 15 2_Department of Zoology, Panjab University, Chandigarh, 160014, India
- 3_College of Life Science, Shaanxi Normal University, 710119, Xi'an, Shaanxi, P.R.
 China
- 18 4_Shanghai Entomological Museum, Institute of Plant Physiology and Ecology,
- 19 Chinese Academy of Sciences, Shanghai 200032, China
- 20 * Equal contribution
- 21
- 22 Corresponding author: Tony Robillard, tony.robillard@mnhn.fr

23 Abstract

24 Subfamily Eneopterinae has been studied for its diversified acoustic modalities and 25 disjunct distribution. The genus Xenogryllus Bolívar, 1890 is one of the oldest genera of the subfamily, and the first genus of the tribe Xenogryllini. Xenogryllus is known for its 26 27 low-frequency calling songs and wide distribution across Africa and Asia. It is known 28 from five species and has never been subject to formal taxonomic revision and 29 description of acoustic features of their calling songs is lacking. Therefore, this study 30 consists of a detailed taxonomic revision of all the species. We re-describe or append 31 their taxonomic features using external morphological features and internal genitalic 32 structures. We update the list of Xenogryllus by describing three new species from 33 Africa. In addition, we provide keys to identify species and describe the calling songs of five species. 34

36 Introduction

37 The crickets of the subfamily Eneopterinae have been recently studied for the diversity of their communication signals and related structures (e.g., Robillard & 38 39 Desutter-Grandcolas 2004a, b; Robillard et al. 2013; ter Hofstede et al., 2015; 40 Schneider et al., 2017), and for their biogeographical patterns (Nattier et al. 2011; 41 Vicente et al. 2017; Dong et al. 2018). To investigate how this diversity occurred and 42 how the original traits evolved, it is necessary to understand the context in which it happened. The tribe Xenogryllini is the sister group of the most diversified Lebinthini 43 (Vicente et al. 2017), therefore, it serves as the best "control lineage" to understand the 44 45 evolution of unique features in the Lebinthini. A taxonomic, bioacoustics and phylogenetic focus on the tribe Xenogryllini is thus particularly needed. 46

The Xenogryllini are currently composed of two genera, *Xenogryllus* Bolívar, 1890 (six species) and *Pseudolebinthus* Robillard, 2006 (two species) according to Orthoptera Species File Online data base (Cigliano *et al.* 2018), but a taxonomic revision was presented as necessary by Robillard (2006). The members of the Xenogryllini produce calling songs with low dominant frequencies (Robillard & Desutter-Grandcolas 2011). However, their spectral and temporal features were never formally documented.

In the present study, we conduct a complete taxonomic revision of the genus *Xenogryllus* (Table 1) based on freshly collected specimens or procured from 17 natural history repositories. We evaluate the status, redescribe or amend description of the existing species, describe three new species, *Xenogryllus lamottei* Robillard n. sp., *X. maniema* Robillard & Jaiswara n. sp. and *X. mozambicus* Robillard n. sp., and provide and a key to species.

60

61 Material and methods

62 Material

63 The material examined belongs to the collections of 17 institutions (see Appendix 64 S1 for lists of material and abbreviations). Newly collected material comes from several 65 field work expeditions in Mozambique (2009), India (2015, 2017) and China (2012, 66 2014, 2017). Newly collected specimens are deposited in the collections of Zoological 67 Survey of India, Kolkata, Muséum national d'Histoire naturelle, Paris and Shaanxi Normal University, Xi'an. Political districts are mentioned for each specimen within lists 68 69 of materials, and square brackets are used for additional information not mentioned on 70 specimen labels or for translation of key information present on labels in languages 71 other than English.

72

73 Taxonomic description

74 Description of studied species follows terminologies as proposed by Robillard et 75 al. (2014). Observations of external morphological characters and dissection of male 76 and female genitalia were performed using Leica stereomicroscopes MZ16 (at MNHN, 77 Paris) and M205C (at IISER, Mohali). SEM observations were performed at the 78 Plateforme de Microscopie électronique of the MNHN, using a JEOL-JSM 840 79 electronic microscope (7kV), after a 60 s gold-coating. Terminologies for male FW 80 venation follow Ragge (1995) and Robillard & Desutter-Grandcolas (2004b). Male and 81 female genitalia were dissected either from dry preserved or freshly killed specimens. 82 Male genitalia were dissected by making a small slit between paraproct and subgenital 83 plate. Female copulatory papilla was dissected out by cutting the membrane between ovipositor and subgenital plate. Dissected genitalia were cleared in 10% cold KOH 84 solution and preserved in glass vials containing glycerine. Terminologies for genitalia 85 follow Desutter (1987), modified in Desutter-Grandcolas (2003) and Robillard & 86

Desutter-Grandcolas (2004a). Imaging of male and female genitalia were made using an AmScope MU1000 digital camera () or with a Canon EOS 40D Digital SLR camera (at MNHN), or with a MC120 HD, Leica, Germany (at IISER, Mohali). To highlight the structural components of male and female genitalia, water solution containing a drop of JBL Punktol was used. To fix orientations and stabilization of genitalia for photography, a clear and viscous Power Plast Hand Sanitizer was used following Su (2016).

93

94 Abbreviations used in taxonomic descriptions

General morphology: FI, FII, FIII, fore, median, hind femur; FW, forewing; TI, TII,
TIII, fore, median, hind tibia; Tarsomere I/II/III-1: basal segment of fore, median and
hind leg tarsomere.

Tegminal venation: 1A-4A, first to fourth anal veins; CuA, anterior cubitus; CuA1,
CuA2, ... first, second, ... branches of CuA; CuP, posterior cubitus; M, median vein; R,
radial vein; Sc, subcostal vein and its branches; c1-3, first to third cells of C alignment;
d1 cell (mirror), first cell(s) of D alignment; d2, second cell of D alignment; e1, first cell
of E alignment.

103 Species abbreviations used in figures: ene, *Xenogryllus eneopteroides;* Iam, 104 *Xenogryllus lamottei* n. sp.; mai, *Xenogryllus maichauensis*; man, *Xenogryllus* 105 *maniema* n. sp.; mar, *Xenogryllus marmoratus*; moz, *Xenogryllus mozambicus* n. sp.; 106 tra, *Xenogryllus transversus*; ulu, *Xenogryllus ululiu*.

107

108 Abbreviations of Institutions used in the lists of material

- 109
- 110 AMNH: American Museum of Natural History, New York, USA.
- 111 ECNU: East China Normal University, Shanghai, China.
- 112 DEI: Deutsches Entomologisches Institut, Müncheberg, Germany.

- 113 NHMUK: Natural History Museum (formerly British Museum of Natural History),
- 114 London, United Kingdom.
- 115 IISERM: Indian Institute of Science Education and Research Mohali, Punjab, India.
- 116 MNCN: Museo Nacional de Ciencias Naturales, Madrid, Spain.
- 117 MNHN: Muséum national d'Histoire naturelle, Paris, France.
- 118 MRAC: Musée Royal de l'Afrique Centrale, Tervuren, Belgium.
- 119 NHMW: Naturhistorisches Museum, Vienna, Austria.
- 120 NWAFU: Entomological Museum, Northwest A & F University, Yangling, China.
- 121 RMNH: National Natuurhistorisch Museum (formerly Rijksmuseum van Natuurlijke
- 122 Historie), Leiden, The Netherlands.
- 123 SNNU: College of Life Sciences, Shaanxi Normal University, Xi'an, China.
- 124 SIPPE: Institute of Plant Physiology and Ecology, Shanghai Institutes for Biological
- 125 Sciences, Chinese Academy of Sciences, China.
- 126 RBINS: Royal Belgian Institute of Natural Sciences, Brussels, Belgium.
- 127 ZFMK: Zoologisches Forschungsinstitut und Museum Alexander Koenig, Bonn,

128 Germany.

- 129 ZIN: Zoological Institute, Russian Academy of Sciences, S. Petersburg, Russia.
- 130 ZSI: Zoological Survey of India, Kolkata, India.
- 131

132 Measurements

BL, body length in dorsal view, from fastigium to apex of abdomen; FIIIL, length of FIII; FIIIW, width of FIII; TIIIL, length of TIII; FWL, forewing length; FWW, forewing width (at the level of maximal width at about 1/3 of FWL); HWT, hind wing tail length (part of the hind wings longer than the FWs); las, inner spines on TIII dorsal side above the spurs; lbs, inner spines on TIII dorsal side between the spurs; Oas, outer spines on TIII dorsal side above the spurs; Obs, outer spines on TIII dorsal side between the spurs; OL, ovipositor length; PronL, pronotum length; PronW, pronotum width; TallIs, spines of third hind tarsomere, not including the apical spines: Ids, inner dorsal spines;
Ods, outer dorsal spines; Ols, outer spines on lateral side of TallI.

142

143 Acoustic analyses

We document the temporal and spectral measurements of calling songs of Xenogryllus eneopteroides, X. maichauensis, X. marmoratus, X. mozambicus, X. transversus and X. ululiu. The calling songs of X. lamottei n. sp. and X. maniema n. sp. are unknown.

148 The basic cricket song terminology follows Ragge & Reynolds (1998). A basic 149 song unit is called syllable and corresponds to one opening-closure cycle of the male 150 forewings. The calling song of X. transversus was recorded using TASCAM DR07 MKII 151 at 44.1 k-samples/s sampling rate (RJ). Males of the species X. mozambicus n. sp. and 152 X. marmoratus were recorded with a modified Condenser Microphone Capsule CM16 (Avisoft Bioacoustics, Berlin), with a flat frequency response from 3 to 150 kHz (R. 153 154 Specht pers. comm.), connected to a Tascam HD-P2 digital recorder (96 kHz sampling 155 frequency, 16 bit) in the field, or using Avisoft Triggering Harddisk Recorder version 156 2.97 and an 8-Pre MOTU sound card at a sampling frequency of 96 k-samples/s (16 157 bit) (TR). The calling songs of X. maichauensis and X. ululiu were obtained from the 158 Orthoptera Species File Online (Cigliano et al. 2018) database and were recorded by 159 Sigfrid Ingrish with a Sony WM-D3 recorder with a Universum mono microphone.

Acoustic analyses were performed using the computer softwares Avisoft-SASLab Pro version 4.40 (Specht 2009) and Audacity version 2.0.0. Acoustic features were measured either manually on Audacity or using the automatic commands of Avisoft-SASLab Pro. Newly recording files are deposited in the Sound Library of the MNHN (inventory numbers mentioned as MNHN-SO*** in lists of material), and in the Orthoptera Sound Library of Indian Institute of Science Education and Research Mohali, Punjab, India.

167

- 168 Systematic part
- 169 Orthoptera
- 170 Gryllidae
- 171 Eneopterinae Saussure, 1874
- 172 Xenogryllini Robillard, 2004

173 Emended diagnosis. Crickets of contrasted sizes, generally light brown; head higher 174 than wide in facial view; fastigium rather wide; eyes of variable sizes, usually 175 ornamented with thin dorso-ventral dark stripes. TI with two tympana, inner one 176 covered by a sclerotized expansion, its membrane visible along a longitudinal slit only; 177 outer tympanum oval in shape, its surface smooth. TIII spurs characteristic of the 178 subfamily, with four pairs of subapical spurs and three apical spurs on each side, the 179 median the longest; subapical spurs rather straight, with curved apex as in Nisitrini. 180 Wings variable within the tribe in males and females. Male. Metanotum with 181 characteristic glandular structures, with a dense bunch of very long setae on scutum 182 anterior edge. Dorsal margin of subgenital plate with baso-lateral glandular structure or dorsal edge with a median invagination. *Male FW venation:* 2A and 3A veins narrowly 183 184 coupled at plectrum level; harp veins variable; mirror generally well developed and 185 rounded; bases of CuA2 and CuA3 fused. Male genitalia: Pseudepiphallic lophi welldeveloped, long and sclerotized, usually with an apical hook-like expansion; 186 187 pseudepiphallus with lateral membranous lobes more or less developed; rami fused 188 with pseudepiphallic sclerite, rather short with convergent apex; ectophallic arc faintly 189 or not sclerotized; endophallic sclerite with long latero-posterior arms connected to 190 ectophallic fold, and short anterior region; endophallic apodeme with wide lateral 191 lamellas. Female ovipositior apex not denticulate, variably pointed.

Distribution. Asia (Japan, China, India, Thailand, Myanmar, Vietnam, Pakistan,Bangladesh) and Sub-Saharan Africa.

194

195 Xenogryllus Bolívar, 1890

196 Type species: *Xenogryllus eneopteroides* Bolívar, 1890.

Xenogryllus Bolívar, 1890: 232; Kirby 1906: 106; Chopard 1968: 349; 1969: 307;
Vasanth 1993: 130; Robillard & Desutter-Grandcolas 2008: 67; Cigliano *et al.* 2018
(Orthoptera Species File Online).

200 Synonym names:

201 Dindymus Kirby, 1906: 88 - incorrect spelling of Dionymus.

202 *Dionymus* Brunner von Wattenwyl, 1893: 213 – Chopard 1968: 349.

203

204 Emended diagnosis. Genus of average to large size, differing from Pseudolebinthus by 205 FWs as long or longer than abdomen in both sexes (shorter in *Pseudolebinthus*), hind 206 wings longer than FWs, forming a short tail posterior to FWs. Eyes small and little 207 prominent, located on face. Face with a whitish or yellowish mask with dark spots. FIII 208 long and thin. Male. Dorsal disc of pronotum forming a wide trapezoid. FWs almost 209 completely overlapping, widened basally, usually forming a characteristic box around 210 abdomen, twice as wide as abdomen. FW cells with thin longitudinal wrinkles, including 211 harp and mirror, absent in *Pseudolebinthus*. Left and right FWs similar in sclerotization 212 and coloration (asymmetrical in Pseudolebinthus). Male genitalia: Pseudepiphallic 213 sclerite longer than rami; lophi long and sclerotized (shorter in *Pseudolebinthus*). Lophi 214 with apical hook-like inner expansions. Pseudepiphallic sclerite with lateral 215 membranous lobes. Rami strong and short, their apex convergent and hook-like. 216 Female. Dorsal disc of pronotum almost rectangular. FWs as long as in male, not 217 widened, reaching apex of abdomen. Ovipositor flattened longitudinally as in other eneopterine genera, its apex rounded and smooth. *Female copulatory papilla*: long,thin and conical.

220 Redescription. Genus of average to large size (Figs 1-2), generally light brown to 221 ochre, with a few dark patterns on FWs. Vertex with five wide longitudinal dark bands 222 (Fig. 4), including a wide median one prolonged on fastigium, two lateral bands more or 223 less marked, made of aggregates of brown dots, and two thin stripes posterior to eyes. 224 Eyes rather small, little prominent, ornamented with thin dorso-ventral dark stripes. 225 Fastigium forming a wide rectangular rostrum slightly prominent on face. Scapes yellow 226 with brown patterns, antennae light brown. Face with a whitish mask with dark spots 227 (Fig. 3). Lateral part of head and lateral lobes of pronotum almost homogeneously 228 brown. Legs usually homogeneously brown. TI with two tympana; inner tympanum 229 covered by a sclerotized expansion, its membrane visible along a small longitudinal slit 230 only; outer tympanum oval in shape, its surface smooth. TI with three apical spurs; 231 outer dorsal apical spur absent. TII with four apical spurs (two dorsal and two ventral). 232 FIII long and thin; TIII with four outer and four inner sub-apical spurs.

233 Male. Dorsal disc of pronotum forming a wide trapezoid FWs almost completely 234 overlapping, widened basally, forming a wide box around abdomen (Fig. 5), with 235 glandular structures on metanotum (Fig. 6). FW cells with thin longitudinal wrinkles. 236 Left and right FWs similar in sclerotization and coloration. FWs light brown, translucent, 237 with the following pattern of black spots on dorsal field: wide black transverse band 238 anterior to transverse part of 1A, including file angle in some species; four corners or 239 mirror and median area of chords with faint dark markings (Fig. 5). Harp with two 240 complete parallel oblique veins and one incomplete oblique vein between diagonal vein 241 and first complete oblique; harp distal angle concave. Mirror large and rounded, 242 separated in two sub-equal parts by a sinuous accessory vein. Cell d2 crescent-like, 243 thin and underlying mirror. Cell e1 very long and curved, underlying posterior margin of 244 mirror. Apical field well developed, triangular, including 4-5 cell alignments. Lateral field

245 crossed by numerous parallel projections of Sc. Male genitalia (Fig.7): Pseudepiphallic 246 sclerite longer than rami; pseudepiphallic lophi very long and sclerotized, with apical 247 hook-like inner expansions (Fig. 8). Pseudepiphallic sclerite with lateral membranous 248 lobes. Membrane at bases of pseudepiphallus forming a sclerotized plate; basal margin 249 with a ventral reinforcement. Rami usually strong with convergent hook-like apex; most 250 often with a ventral posterior expansion reaching base of pseudepiphallic membranous 251 lateral lobes. Pseudepiphallic parameres with one strong rectangular ventral lobe 252 covered with scale-like sculptures, with a basal lobe mostly membranous. Ectophallic 253 arc nearly membranous. Ectophallic apodemes usually strong. Ectophallic ventral 254 expansions well developed and sclerotized. Ectophallic fold entirely sclerotized 255 ventrally, its lateral expansions partly fused with lateral arms of endophallic sclerite. 256 Endophallic sclerite forming a flat plate with long latero-posterior arms and a short 257 medio-posterior triangular expansion. Endophallic apodeme with two wide lateral 258 lamellas and no dorsal crest.

259 Female. Pronotum dorsal disc almost rectangular. FWs as long as in male, reaching 260 apex of abdomen or slightly longer, not widened as in male; light brown with a dark 261 spot of variable size between veins CuP and CuA, near lateral angle, at ¼ of FW 262 length; with strong longitudinal veins and faint transverse ones. Lateral field crossed by 263 numerous parallel projections of Sc; area between R and Sc most often with a thin dark 264 brown band. Subgenital plate variably indented apically (Fig. 9). Ovipositor flattened 265 longitudinally, its apex rounded and smooth (Fig. 10). Female copulatory papilla: long, 266 thin and conical, its sclerotization variable (Fig. 11).

267 *Distribution.* Japan, Southern China, Taiwan, India, Sri Lanka, Thailand, Myanmar,
268 Vietnam, Bangladesh and Sub-Saharan Africa.

269 Natural history. Xenogryllus species live in open areas, in grassland and savannah.

- 270 Males sing at night from low bushy vegetation.
- 271

272 Key to species of Xenogryllus

273	1. Dorsal disc of pronotum carinated laterally. Eyes higher than long in lateral view,						
274	occupying		half	of		head	height
275						2	
276	- Dorsal dis	sc of pronotur	n not carinate	ed laterally.	Eyes as lo	ong as high	in lateral view,
277	restricted	to	dor	sal	third	of	head
278					4		
279							
280	2. Face alm	nost flat in late	eral view			. X. eneopt	eroides Bolívar,
281	1890						
282	-	Face	rounded		in	lateral	view
283						3	
284							
285	3. Face bul	bous in latera	l view. Size ra	ather larger	. Male geni	talia: Pseud	lepiphallic lophi
286	(Fig. 8F)	ended by	a small la	amella, the	eir inner	margin al	most straight,
287	membrano	JS				. X. mozan	<i>bicus</i> Robillard
288	n. sp.						
289	- Face less rounded in lateral view. Size rather shorter. Male genitalia: Pseudepiphallic						
290	lophi (Fig. 8E) ended by a long conical apex and with a widened inner membranous						
291	margin X. maniema Robillard &						
292	Jaiswara n.	sp.					
293							
294	4. Male	FWs well	widened	basally,	forming	a wide	box around
295	abdomen		5				

- Male FWs not widened, as wide as abdomen...... X. lamottei Robillard
n. sp.

298

307

308 6. Male genitalia: Pseudepiphallic lophi with pseudepiphallic lophi forming curved
309 elongate hooks widened apically (Fig. 8D). Female subgenital plate posterior margin
310 indentation rounded laterally, as wide as long (Fig. 9C)...... *X. maichauensis*311 Gorochov, 1992

315

316 7. Size larger. Male genitalia with larger pseudepiphallic sclerite; lophi proportionally
317 smaller (Fig. 8I), fused only basally, separated by a deep indentation. Female
318 subgenital plate posterior margin indentation very deep, longer than wide (Fig.
319 9G)..... X. transversus (Walker,
320 1869)

Size smaller. Male genitalia with smaller pseudepiphallic sclerite; lophi fused on half
of their length (Fig. 8H). Female subgenital plate posterior margin indentation weak,
wider than long (Fig. 9E)......X. *ululiu* Gorochov, 1990

325

326 Xenogryllus eneopteroides Bolívar, 1890

327 (Figs 1A-C; 3A-B; 4A; 5A; 6; 7A-C; 8A-B; 9A; 10A; 11A-B)

328

329 Xenogryllus eneopteroides Bolívar, 1890: 232 - Karsch 1893: 204; Kirby 1906: 106; 330 Giglio-Tos 1907: 26; Bolívar 1910: 544; Chopard 1954: 80; Chopard 1968: 349; 331 Chiffaud & Gillon 1984 (life history); Paris 1994[1993]: 69 (lectotype designated); 332 Robillard & Desutter-Grandcolas 2004a: 578; 2004b: 273 (morphological phylogeny); 333 2006: 644; 2008: 67; 2011: 637; Robillard 2006: 673; Robillard et al. 2007: 1267 (song 334 frequency); Nattier et al. 2011: 2201 (molecular phylogeny); Chintauan-Marguier et al. 335 2016: 62 (molecular phylogeny); Vicente et al. 2017: 2203 (historical biogeography); 336 Cigliano et al. 2018 (Orthoptera Species File Online).

337 *Xenogryllus eucopteroides -* Kevan & Knipper 1961: 368 (misspelling); Chopard 1968:
338 349.

339

Type material. Lectotype, *∂*, **Angola:** Duque de Bragança, Bayão (Paris, 1994)
[MNCN, examined on photograph].

Additional material examined. Benin: Bas-Dahomey, plateaux de Zagnamado et de
Ketou, forêt d'Aqua-queré (saison sèche), 1910 P. Ducorps, 1♀ (MNHN). Cameroun:
[L.] Conradt, 1♂, identified Xenogryllus eneopteroides Bol. (MNHN). Reg[ion] de Kribi,
1925, littoral, saison humide, Dr. Gromier, 1♂ (MNHN). Kamerun [Cameroun], Pipindi
[Bipindi], Eitel ded. 1♀, identified Xenogryllus eneopteroides Bol. (MNHN). 10 km N. of

Yaoundé, E. of Agronomy Station, Nkolbisson, 10.vii.1975, N. D. Jago, 13, identified 347 348 Xenogryllus eneopteroides Bolivar by B. C. Townsend, 1976 (NHMUK 010926525). 349 Central African Republic: République Centrafricaine, 75 km WNW Mbaiki, N'Gotto 350 [Ngoto], W Lobaye, savane arbustive, 7.iv.1995, nuit [night], L. Desutter-Grandcolas, 1 351 juvenile, n°14, molecular sample Xen-CA (MNHN-E0-ENSIF3159); 13, #1, sur plant 352 [on plant] (MNHN). Democratic Republic of the Congo: Congo, 20 km from L. Tumba 353 (towards Coquilhatville) [Mbandaka], 5.iii.1964, from grassy clearing in forest, 13, 19, 354 Tyson Roberts, Brit. Mus. 1982-71 (NHMUK 010926531, 010926572). Kalembe, 355 15.ix.1947, Miss. Tanganika, 1º (RBINS). Lukafa [Lukafu], Congo Belge, 6.xii.1938, 356 H.J. Brédo, 1♀, R.Mus.Nat. Belg.I.G.12.204 (RBINS). Belgian Congo, Aru, iii.1936, 5♂, 357 2^Q, H.J. Bredo, dry Acacia bush (NHMUK 010926536, 010926527, 010926578, 358 010926580, 010926539, 010926566, 010926569). Niangara, v-vi-1915, 28°0E 3°40"N, 13 (AMNH). Haut-Ituri: Faradje, 13, Blommaert, Musée du Congo, identified 359 Xenogryllus eneopteroides by L. Chopard, R. Det. N 2814 (MRAC). S.E. Katanga: 360 361 Ngaye: xi-xii.1931, 13, R.P. Claquin, Musée du Congo, identified Xenogryllus 362 eneopteroides by L. Chopard, R. Det. N 2814 (MRAC); 1932, 13, 12, R.P. Claquin, 363 Musée du Congo, identified Xenogryllus encopteroides by L. Chopard, R. Det. DD3509 364 (MRAC). Haut-Uele, vi.1925, 1^o, Musée du Congo, identified *Xenogryllus* 365 eneopteroides by L. Chopard, R. Det. N 2814 (MRAC). Kibali-Ituri: Geti, ii-iv.1937, 13, 366 Ch. Scops, Musée du Congo, identified Xenogryllus eneopteroides by L. Chopard, R. 367 Det. B 5318 (MRAC). Gabon: Plateaux Batéké, piste Ekalla, 30 km W Leconi [Lekoni], 368 milieu herbeux, 19.vi.1994L. Desutter-Grandcolas: 1 juvenile, n°61, jour [day], litière 369 [leaf litter], molecular sample Xen-GA (MNHN-EO-ENSIF3442); 1♂, n°3, nuit [night], 370 sur plante – base tronc [on plant, low on tree trunk] (MNHN). Bas Ogooué, 13, 371 identified Xenogryllus eneopteroides Bol. by L. Chopard (MNHN). Ntoum, 19.vii.[19]85, A. Pauly rec, 23, 22, 2 juveniles, lumière (RBINS). Ghana: Eastern region, 372 373 Hansowodze, 2 mls from Kade A.R.S. [Agricultural Research Station], 19.vii.1963, I.K. 374 Bacheampong, 4d (NHMUK 010926524, 010926567, 010926528, 010926571).

375 Ghana, Togoland plateau, grassy, summit above [Nkonya] Wurupong, 23.xii.1959, N.D. 376 Jago, 1∂, Brit. Mus. 1965-474 (NHMUK 010926535). Eastern region, Kade Agr. Res. 377 Stn [Agricultural Research Station], 11.vii.1963, I. K. Bacheampong, 23, Brit. Mus. 378 1965-474 (NHMUK 010926526, 010926538). Transvolta, Togoland, Amedzofe: 379 3.v.1959, (montane grass), 1[♀], N.D. Jago, Brit. Mus. 1965-474 (NHMUK 010926529); 25.v.[19]59, N.D. Jago, Brit. Mus. 1965-474 (NHMUK 010926579). Western Reg.[ion] 380 381 mls E of Elmina, 1°20W 5°05.5N, xii.1960, 1∂, 1♀, N.D. Jago, identified *Xenogryllus* eneopteroides by P.C. Tinning, 1966, Brit. Mus. 1965-474 (NHMUK 010926509, 382 383 010926511). T.v.T [Trans-Volta Togo], Ho-Hohoe rd. 10 miles from Volta bridge, iii.1960, N.D. Jago, Brit. Mus. 1965-474 (NHMUK 010926553). Western Reg. 384 385 Asankragua-Enchi rd., Tano R. Ferry, 24.ix.1962, 13, N.D. Jago, Brit. Mus. 1965-474 (NHMUK 010926510). Guinea: Kaoulenta [Kéoulenta], [Mount] Nimba (Guinée), ii-386 vi.[19]42, M. Lamotte, 13, 12, 2 juveniles (MNHN). Nimba (Guinée), [Mount] Pierre 387 Richaud, 900 m, M. Lamotte ii-iv.[19]42, 12, identified Xenogryllus eneopteroides Bol. 388 389 by L. Chopard; 1º (MNHN). Nimba (Guinée), Lamotte, Amiet, Vanderplaetsen, xii.56-390 v.57: Ziéla, 17.ii.[19]57, 1 juvenile; Ziéla, 20.iv.[19]57, 1 juvenile; sav. [savannah] entre 391 Ziéla et Gbakbré, 9.xii.1956, 1 juvenile; Zgpo. [Zouguépo], 900 m, androp, 18.iv.[1957], 392 2°; sav. [savannah] Ziéla, 3.ii.[19]57, 1 juvenile; sav. [savannah] Ziéla, 20-393 25.xii.[19]56, 4 juveniles; sav. [savannah] Kéoulenta, 5.xii.[19]56, 2 juveniles; forêt 394 Ziéla, 19.ii.[19]57, 1 juvenile; 1 juvenile, A.V. n°23, identified Xenogryllus eneopteroides 395 Bol. larva by L. Chopard; 1 juvenile, A.V. n°38; 1 juvenile, 87CD; 14.iii.[19]57, 1 396 juvenile, fge n°47 (MNHN). Mt. Nimba, Mt. Leclerc [Mount Jean Charles Leclerc], 1500 397 m, 23.iii.1991. M. Lamotte, n°319, 1º (MNHN); 1º, molecular sample X22-XenGU 398 (MNHN-EO-ENSIF1494). Nimba (Guinée), M. Lamotte, xii.56-v.57, 29.x.[1956], 1∂, 399 herbes A2 Ziela, identified Xenogryllus encopteroides Bol. by R. Roy, 1959 (MNHN). 400 Guinée Française, Diéké [Diecke], P. Chabamaud, 1920, 19, identified Xenogryllus eneopteroides Bol. by L. Chopard (MNHN). Ivory Coast: Lamto (Toumodi), Coll. 401 402 E.N.S. Paris, Lamotte et collab.: 1-10.ii.[19]62, 33, 22; 1963, 13, Xen-Robillard403 morpho; 25-31.xi.[19]62, 1[°]; 15-20.viii.[19]62, 1 juvenile; iv.[19]62, 1[°]; 7.xi.[19]62, 1[°]; 404 20.xi.[19]62, 1°; vi.[19]62, 1°; 4.vii.[19]62, 1°, mare sav. inondée [inundated 405 savannah] (MNHN). Forêt à 15 km de Lamto, 10.ix.1968, C. Girard, 12, 1 juvenile 406 (MNHN). Forêt de Yape, 1⁽¹⁾ (MNHN). Toumodi, xii.1930-iv.1931, Ch. Alluaud & P. A. 407 Chapuis, 1∂ (MNHN). Danané, i.1939, 1♀ (MNHN). Lamto (Toumodi), C. Girard: 12.ii.1968, savane [savannah], lisière forêt galerie [hedge of gallery forest] 6∂, 3♀, 5 408 409 juveniles; 13, identified Xenogryllus eneopteroides Bol. by R. Roy, 1974; 12.iii.1968, 410 lisière, forêt, savane, 1♂, molecular sample XenCl (MNHN-EO-ENSIF1481); 8♂, 2♀, 7 411 juveniles; 13, photos MEB [SEM photos] (MNHN-EO-ENSIF1518); 15-30.vii.1968, 412 savane [savannah], lisière forêt galerie [hedge of gallery forest] 13, 29, 3 juveniles; 27-413 28.ii.1968, 2♀, 1 juvenile, à la lumière; 10-20.iv.1968, 1♂, à la lumière (MNHN). Lamto 414 (Toumodi), Paul Planquette: 12, PNB 18; 1 juvenile, PNB 13B; 15.i.[19]64, 12, PNB 64 415 (MNHN). Lamto (Toumodi), 30.vi.[19]62, Coll. E.N.S. Paris, Lamotte et al. 13, identified 416 Xenogryllus eneopteroides by L. Chopard (MNHN). Nimba [Mount]: Lamotte, 1946, Z4, 417 1♀, identified Xenogryllus eneopteroides by R. Roy, 1967 (MNHN). Lamto, 1963, 1∂ 418 (MNHN). Assinie, 3295-85, Chaper, 13 (MNHN). Réserve du Banco [Banco national 419 park], R. Paulian & G. Delamare, 23, 29, 2 juveniles (MNHN). Bouaké (C.I.), 420 22.v.[19]62, coll. E.N.S. Paris, Lamotte et collab., 1♂ (MNHN). Taï, 18.viii.1978, G. 421 Couturier, 1♀ (MNHN). Lamto, 1962, P. Le Gall, 2♂, 2♀ (MNHN). Kenya: CRS 422 [Catholic Relief Service] Tebere, 0039.5S 3723E, 10.i.[19]72, I.A.D.R. 72/58, 1♀, Brit. Mus. 1982-71 (NHMUK 010926573). Nigeria: Lagos, Ikoyi, viii. 1951, L. Bala, 13, 423 424 identified Xeneogryllus eneopteroides by L. Chopard (MNHN). U.C. [University College] 425 Ibadan, 5.ix.1952, Tephrosia, leaf, Teph-66 (MNHN) coll. G.H. Caswell, 1^o (MNHN). 426 Ibadan, 6.vi.[19]23, 1♂, 1605 (NHMUK 010926581). Calabar, Nigeria, 10.iv.[18]82, 427 10.30 A.M., W. edge of [?t] Kwa swamp, E. edge of UNICAL staff guarters; about 50 428 cm up on base of tangle of turns e.g. Clitonia ? rubiginosa, notes 72, 1♀, C.I.E. coll 429 A.156 76 Pres by Comm Inst Ent B.M. 1985-1(NHMUK 010926537). Replublic of 430 **Congo:** Sibiti, xi-1963, mission A. Descarpentries et A. Villiers 1963-1964, 2³, 1²

(MNHN). Baie de Lagoa, envoi H. Deyrolle, 1^o (MNHN). Mayumbe, Luki, L. Tiebers [?], 431 432 1♀, Musée du Congo (MNHN). Odzala, x.1963, 3♂, 3♀, 1 juvenile (MNHN). Congo, 433 1963, 1º (MNHN). Ile [island] M'Bamou, 26.vii.1968, Fidèle, 1º (MNHN). Boma, Sundi, 434 P. Rolin, 1⁽¹⁾ (RBINS). Banana, F. Busschodts, 4⁽²⁾ (RBINS). Sierra Leone: Piste 435 Bandankoro, 29.v.1963, savane de plaine ayant brûlé [burnt lowland savannah], 436 mission ENS-IFAN aux Monts Loma, 13, identified Xenogryllus eneopteroides by R. 437 Roy, 1992; 2∂, 3♀ (MNHN). Piste Keimadugu, savane [savannah], 520 m, 29.v.1963, 438 mission ENS-IFAN aux Monts Loma, 23, 12 (MNHN). Firawa, savane [savannah], 439 1.vi.1963, mission ENS-IFAN aux Monts Loma, 13, 12 (MNHN). Njala, E. Hargreaves: 440 21.vii.[19]32, B.M. 1938-149, identified X. eneopteroides Bol. by L. Chopard, pres Imp. 441 Inst. Ent. Brit. Mus. 1965-474 (NHMUK 010926552); x.1936, 13, pres Imp. Inst. Ent. 442 Brit. Mus. 1965-474, 1⁽²⁾, identified Xenogryllus eneopteroides by B. Uvarov (NHMUK 010926534); 23.v.[19]25, 13, identified X. eneopteroides Bol. by L. Chopard, B. M. 443 444 1938-149 (NHMUK 010926530); xi.1935, 1♂, pres Imp. Inst. Ent. Brit. Mus. 1936-252, 445 13, identified Xenogryllus eneopteroides by B. Uvarov (NHMUK 010926576). Uganda: 446 Entebbe, 20.xi.1970, W.J. Bailey, B.M.1977-246, identified Xenogryllus eneopteroides 447 by B. C. Townsend, 1978 (NHMUK 010926570). Kisaru, 19.vii.1983, 13, H.B. Johnson, 448 at light (NHMUK 010926551). Zambia: Lusaka, about fifty metres from the Great East 449 Road in Chalimbana, Chonqwe Distr., 19.xii.2013, William van Niekerk, 13, identified 450 X. eneopteroides by William van Niekerk [online photograph]. Kapiri, 13.xi.1913, 22, L. 451 Charliers, Musée du Congo, identified Xenogryllus eneopteroides by L. Chopard, R. 452 Det. N 2814 (MRAC).

453 *Type locality.* Angola, Duque de Bragança, Bayão (=Kalandula).

Distribution. Sub-Saharan Africa: Angola, Benin, Cameroun, Central African Republic,
Democratic Republic of the Congo, Gabon, Ivory Coast, Kenya (one large female
specimen with unclear identification), Nigeria, Republic of Congo, Sierra Leone,
Uganda, Zambia.

Emended diagnosis. Species of average size, close to *X. mozambicus* n. sp. and *X. maniema* n. sp., from which it differs by face almost flat in lateral view (more rounded in other species), shorter male FWs (short apical field), and shape of lophi in male genitalia. Differing from *X. lamottei* n. sp. and Asian species by following characters: well-carinated lateral angles of dorsal disc of pronotum (also carinated in *X. mozambicus* and *X. maniema*), very short ovipositor and male genitalia.

Redescription. Species of average size (Fig. 1A-C), coloration gray brown little contrasted. Eyes large, lateral, occupying almost half of head height in lateral view (Fig. 3A-B). Face almost flat in lateral view, with typical whitish mask underlined by a black line below eyes and on mandibles. Pronotum dorsal disc strongly carinated laterally (Fig. 3B), coloration dark brown or brown, most often with a median dark brown band extended laterally near anterior margin, forming a T shape; lateral lobes almost homogeneously dark brown. First article of antennae dark brown.

471 *Male.* FWs very wide, dark coloration anterior to 1A including angle of 1A (Fig. 5A). File 472 with 658 stridulatory teeth (n=2) on transverse part of 1A. Harp longer than wide. Cell 473 c1 narrowed posteriorly, its fusion with cell b1 slightly shorter than its individualised 474 part. Cell c2 large, d2 long and narrow. Mirror large, wider than long and little rounded, 475 its inner angle forming an angle. Apical field short, wider than long, including four or 476 five cell alignments.

477 Male genitalia (Figs 7A-C, 8A-B). Pseudepiphallic lophi as long as rest of 478 pseudepiphallic sclerite, forming a long rectangle, sometimes slightly widened 479 preapically; lophi with a narrow membranous inner margin setose basally (Fig. 8A-B); 480 their bases fused until mid-length; apex pointed and divergent, ended by a thin lamella 481 without hook-like inner dorsal expansion (in dorsal view, anterior base of lamella 482 suggests that lamella may partly correspond to modified inner dorsal expansion of 483 other species); ventral blade of lophi without longitudinal wrinkles, but with a strong 484 ventral transverse carina, as in X. mozambicus n. sp. Pseudepiphallic parameres with

a strong rectangular ventral lobe and a basal membranous lobe. Rami strong, their
apex forming wide convergent hooks. Ectophallic apodemes strong, not lamellate.
Ectophallic lateral expansions, lateral sclerites of ectophallic fold and endophallic
sclerite partly fused, forming a wide ventral sclerotized plate, trifid apically; endophallic
apodemes made of wide lateral lamellas and a narrow dorsal crest.

Female (Fig. 1C). Dorsal disc of pronotum almost rectangular, its posterior margin
slightly bisinuate. FWs dorsal field with 7-10 longitudinal veins (m=8, n=10). Subgenital
plate with a shallow apical indentation (Fig. 9A). Ovipositor (Fig. 10A) very short, about
one third of FIII length.

494 *Female genitalia*. Copulatory papilla (Fig. 11A-B) conical and narrow, its apex rounded;
495 well-sclerotized except base and apex.

Life history traits. According to Chiffaud & Gillon (1984), *X. eneopteroides* is found in
lvory Coast at the end of the dry season, in parts of savannah protected from fires.
Their study revealed that the species feeds only on leaves and flowers of dicotyledones
which are specific to savannah.

500 *Calling song.* According to the sonogram showed in Desutter (1983), analysed by 501 Robillard & Desutter-Grandcolas (2004a, 2011), the calling song of *X. eneopteroides* is 502 a short echeme made of five long syllables (syllable duration = 65 ms, period = 106.5 503 ms. Echeme duration = 611 ms, echeme period = 1062 ms. The dominant frequency is 504 low (3.6 kHz) and corresponds to the first peak of the spectrum.

505 *Measurements.* See Table 2.

506 *Taxonomic discussion.* The material examined reveals numerous slight differences in 507 male genitalia (Fig. 8A-B) and size along the wide geographic distribution of the 508 species. In particular, specimens from Congo differ from the material from the western 509 coast of Africa. However, these differences are not sufficiently clear or are based on

- 510 too few individuals to delimit different species yet. More information and sampling will
- 511 be necessary to improve the study *Xenogryllus* in Africa.

512

513 Xenogryllus carmichaeli (Chopard, 1928)

514

515 Dionymus carmichaeli Chopard, 1928: 30.

516 Xenogryllus carmichaeli – Chopard 1968: 350; Yin & Liu 1995; Cigliano et al. 2018

517 (Orthoptera Species File Online).

518 Madasumma carmichaeli – Chopard 1969: 311.

519 *Nomen dubium* – this study.

520

521 Taxonomic discussion. According to the description of Chopard (1928), the female of 522 X. carmichaeli is similar to X. marmoratus in terms of shape, size and coloration. 523 However, the male specimen described by Chopard possesses five oblique veins, 524 which is not consistent with any other existing species of Xenogryllus, as noted by 525 Chopard himself. It is probable that the type series was heterogeneous, the male 526 belonging to another subfamily than Eneopterinae. The male type may belong to the 527 genus Madasumma, according the subsequent identification of the species by Chopard 528 (1969), while the female could belong to X. marmoratus. The type locality, "India, 529 Darjeeling District, alt. 100-300 ft, West Bengal", could fit with X. marmoratus, but also 530 X. transversus or X. maichauensis. Given these problems and the fact that the type 531 series is missing in ZSI, Kolkata (RJ, pers. obs.), we consider this species as nomen 532 dubium.

533

534 Xenogryllus lamottei Robillard n. sp.

535 (Figs 1H-I; 3C-D; 4B; 5B; 7D-E; 8C)

536

Type material. Holotype, ³, **Guinea:** Simandou [Mount], Guinée [Guinea], Cpt.[camp]
Fon Yenfédou, Ifan [Institut Français d'Afrique Noire], ix.1951, [M.] Lamotte (MNHNEO-ENSIF10685).

540 *Type locality:* Mount Simandou, Guinea.

541 *Distribution.* Species only known from the type locality in Guinea.

542 *Etymology.* The species is dedicated to the great French entomologist Marcel Lamotte

543 who collected the type specimen.

544 Diagnosis. Species of average size, characterized by male FWs not widened as in 545 other species of the genus; general morphology differing from the other African species 546 (X. eneopteroides, X. mozambicus n. sp. and X. maniema n. sp.) and more similar to 547 the Asian species (X. marmoratus, X. transversus, X. ululiu and X. maichauensis) by 548 the following characters: pronotum not carinated laterally (Fig. 3D); wrinkles on surface 549 of male FWs weak; eyes small, restricted to dorsal guarter of head (reaching 1/2 of head 550 in other African species); face almost flat in lateral view; male genitalia with short 551 pseudepiphallic lophi (Fig. 8C), close to that of X. ululiu (Fig. 8H), their apex short and 552 bifid, with a pre-apical dorsal hook-like expansion (absent in other African species); 553 ectophallic fold and latero-ventral expansions shorter than in other species.

Description. In addition to the characters of the genus, species of average size (Fig. 1H-I), coloration gray brown little contrasted. Fastigium longer than wide, thinner than in other species (Fig.4B), slightly widened apically. Face almost flat in lateral view (Fig. 3D), with typical pale mask with black spots underlined by a thick black line below eyes (Fig. 3C); mandibles dark brown; clypeus and labrum mottled with yellow and dark brown; maxillary palpi dark brown. Scapes and first article of antennae dark brown,

flagellum light brown. Eyes rounded and restricted to posterior quarter of head in lateral view. Pronotum dorsal disc not carinated laterally (Fig. 3D), brown, with lateral margins underlined by a yellow line, with a median dark brown band, ticker posteriorly; posterior margin almost straight. FIII narrow, ended by a long linear region. Abdomen slightly shorter than FWs. Cerci dark brown.

565 Male. FWs as wide as abdomen (Fig. 5B), not widened as in other species of the 566 genus; light brown, translucent, with weak wrinkles on surface. Dark coloration anterior 567 to 1A not including angle of file. CuP visible posteriorly until angle of 1A. Angle of 1A 568 straight. Harp wide, with two straight parallel oblique veins and a partial one, straight 569 and reaching harp mid-length. Cell c1 not narrowed posteriorly. Mirror almost rounded, 570 its inner limit forming a curve; underlined posteriorly by cells e1 and d2 fused together. 571 Apical field forming a narrow triangle made of four cell alignments (E-H). CuA thin and 572 brown; M thick, whitish; R and Sc almost fused, brown; M-R area dark mostly brown, its 573 dorsal margin whitish; lateral field translucent brown crossed by 22 projections of Sc.

574 Male genitalia (Fig. 7D-E). Pseudepiphallic lophi forming a short rectangle posterior to 575 wide base of pseudepiphallic sclerite; lophi (Fig. 8C) with a narrow membranous inner 576 margin, setose basally; apex with a black hook-like inner dorsal expansion; sclerotized 577 ventral blades of lophi with longitudinal wrinkles as in X. transversus and X. ululiu, but 578 without strong ventral carina as in X. eneopteroides and X. mozambicus. 579 Pseudepiphallic lateral membranous lobes small. Pseudephiphallic basal margin 580 reinforcement weak. Rami thin and straight, their apex narrowed but not hooked 581 innerly. Pseudepiphallic parameres ventral plate r-shaped,. Ectophallic arc not 582 sclerotized. Ectophallic apodemes thin and divergent. Ectophallic fold and latero-dorsal 583 expansions short. Endophallic sclerite and apodeme little differentiated.

584 *Female.* Unknown.

585 *Life history traits.* Unknown.

586 *Calling song.* Unknown.

587 *Measurements.* See Table 3.

588

589 Xenogryllus maichauensis Gorochov, 1992

590 (Figs 1L-O; 3E-F; 4C; 5C; 7L-N; 8D; 9C; 10B; 11E-G; 12A; 13)

591

592 Xenogryllus maichauensis Gorochov, 1992: 9 – Kim & Pham 2014: 60; Cigliano *et al.*593 2018 (Orthoptera Species File Online).

Xenogryllus transversus – Yin & Liu 1995: 96; Robillard *et al.* 2007: 1267 (song
frequency, wrong identification); He 2018: 516; Cigliano *et al.* 2018 (Orthoptera
Species File Online) (wrong identification).

597

598 *Type material. Holotype, ∂*, **Vietnam: Mai Châu** (ZIN) [examined through photographs
599 sent by A. Gorochov].

600 Additional material examined. China: Yunnan, Mengla, E101°33' N21°28', 3.ix.1991, 601 Zuyao Liu, Tianqi Wang & Heisheng Yin: 1♂ (14062446), 2♀ (14062547, 14062622) 602 (SIPPE). Mengla Distr., 12-23.x.2014, coll. Zhang Tao, 14∂, 2♀, (SNNU). San Chahe, 603 Mongyang, E98°22' N25°23', 31.vii.1995, Xianwei Liu, Weinian Zhang, Xingbao Jin, 1 604 (14062620) (SIPPE). Xi Shuang Ban Na, 580 m, 10.ix.1993, Xinyue Chen, 3∂, 605 (14064963, 14064964, 14064966); 2♂, 7.ix.1993 (14064968, no number); 1♂, 606 5.ix.1993 (14064967) (SIPPE). Monglun Xi Shuang Ban Na, E101°15' N21°56', 607 5.ix.1993, Longlong Yang, 2♀, (14064969, 14064971); 7.ix.1993, Xinyae Chen, 1♀ 608 (14064970) (SIPPE). Jinhong, E101°15' N21°56°, Zuyao Liu, Tianqi Wang, Heisheng 609 Yin: 6.ix.1991, 3♂ (14062619, 14062546, 14062448); 8.ix.1991, 1♂ (14062447) 610 (SIPPE). Jinghong, [N22°0'6.97" 100°46'25.07"E], 9.ix.1991, Zuyao Liu, Tianqi Wang,

611 Heisheng Yin, 1^o (14062449) (SIPPE). Mohan Vill., 950 m, 3-ix.2005, coll. Xue Guoxi, 23 (NWAFU). Guangdong, Shen zhen, [22°33'59.69"N, 114° 2'25.53"E], 9.x.2012, 612 613 Zhang Tao [online photograph]. Thailand: Tak, Doi Musoe, 700 m, Agric. Res. Stn at 614 night, 9.x.1990, S. Ingrish: 1♀, 1♂, molecular sample XtrTh, identified Xenogryllus 615 transversus by S. Ingrish (ZFMK). Mae Salid, Monkrathing, 17° 30' N, 98° 5' E, 700 m, 19.ix.1989-21.ix.1989, S. Ingrisch, 13, mountain forest and agricultural land [day 616 617 collecting and night collecting following stridulation, Stridulation recorded by S.Ingrisch 0295DXEN.WAV [old ID CIGxentraSW03], identified Xenogryllus transversus by S. 618 Ingrish (ZFMK). Umphang, 1-6 km S., 15° 59' N, 98° 50' E, 16.x.1991, S. Ingrisch, 13, 619 bamboo forest, along road and trails, stridulation recorded by S. Ingrisch 620 621 0510XENO.WAV, identified Xenogryllus transversus by S. Ingrish (ZFMK). Chiang Mai, Roadside by Samoeng and Mae Rim, 18° 50' N, 99° 0', 13, 11.x.1991, S. 622 Ingrisch, 13, stridulation recorded by S. Ingrisch 0502XENO.WAV, [old ID] 623 624 CIGxentraSW05], roadside at night, identified Xenogryllus transversus by S. Ingrish 625 (ZFMK). Petchabun, Nam Nao, 16° 47' N, 101° 27' O, 1000 m, S. Ingrisch; 13.ix.1989-626 14.ix.1989, S. Ingrisch, 13, stridulation recorded by S. Ingrisch 0286XENO.WAV [old ID CIGxentraSW01], mixed Oak-Pine forest, grassy undergrowth, Bamboo thicket [day 627 628 collecting and night collecting following stridulation], identified Xenogryllus transversus 629 by S. Ingrish (ZFMK).

630 *Type locality.* Mai Châu, Vietnam.

631 *Distribution.* Southern China, Northern Vietnam and Northern Thailand.

Emended diagnosis. Species of large size, similar in size, venation and coloration to *X. transversus*, from which it differs mostly by male genitalia, with pseudepiphallic lophi
forming curved elongate hooks widened apically (Fig.8D), while lophi are straight in *X. transversus* (Fig. 8I).

636 *Redescription.* In addition to the characters of the genus, *X. maichauensis* has a large 637 size and a light brown or golden coloration (Fig. 1L-O). Fastigium slightly widened

apically as in *X. ululiu* and *X. transversus* (Fig. 4C). Eyes rather small, located on face,
restricted to the dorsal third of head in lateral view (Fig. 3E-F). Lateral angle of dorsal
disc of pronotum not carinated, with a thin yellow band underlined by a thin black line
anteriorly. Hind wings tail gray brown, twice as long a pronotum.

Male. Pronotum dorsal disc more rectangular than in *X. transversus*, with a wide median black longitudinal band, its posterior margin slightly bisinuate. FW venation (Fig. 5C): 1A forming a straight angle, with 299 stridulatory teeth (n=1) located on transverse part of 1A. Dark coloration anterior to 1A not including angle of file. Harp wide. Mirror large, well-rounded, its inner limit curved. Apical field forming a triangle longer than wide, with six cell alignments.

Male genitalia (Fig. 7L-N). Characteristic Y-shaped pseudepiphallic lophi forming curved elongate hooks, widened apically, with a sharp dorsal preapical expansion (Fig. 8D). Rami strong, convergent apically, with a ventral posterior expansion reaching base of pseudepiphallic lateral membranous lobes. Ectophallic apodemes long and thin. Ectophallic fold and endophallic sclerite almost fused, forming a wide sclerotized plate, trifurcate posteriorly; endophallic apodemes made of wide lateral lamellas.

Female. Head slightly wider than pronotum, with strong jaws visible from dorsal view (Fig. 1N-O). Dorsal disc of pronotum almost rectangular, its posterior margin slightly bisinuate. FWs light brown, anterior dark spot larger than in *X. ululiu, X. transversus* and *X. marmoratus*. Dorsal field with nine strong longitudinal veins. Subgenital plate with a V-shaped apical indentation with rounded edges (Fig. 9C). Ovipositor (Fig. 10B) as long as FIII.

660 *Female genitalia*. Copulatory papilla (Fig. 11E-F) conical, its apex rounded and 661 sclerotized.

662 *Life history traits.* In Vietnam, the type specimens were found in forest, on leaves of 663 bushes near a stream (A. Gorochov, pers. comm.). According to the information

associated with the recordings made by S. Ingrish in Thailand (available on the
Orthoptera Species Files online under the name *X. transversus* (Cigliano *et al.* 2018),
the species was found in mixed oak-pine forest on grassy undergrowth with bamboo
thicket. Males call at night on vegetation.

Calling song. (Figs 12A, 13) *X. maichauensis* was recorded in Thailand by S. Ingrish (Cigliano *et al.* 2018). At 24.5°C, the calling song is made of short echemes quickly repeated and composed of 2-4 long syllables (m = 3.02 ± 0.63), lasting for 179.5 ± 41.6 ms, with a period of 502 ± 42 ms. Within echemes, the first syllable has a lower amplitude than the next ones. Syllables are rather long (duration = 46.5 ± 1.7 ms) with a syllable period of 65.1 ± 1.7 ms (syllable duty cycle = 71%). The frequency spectrum shows a dominant frequency at 4.91 ± 0.07 kHz followed by two powerful harmonics.

675 *Measurements.* See Table 4.

Taxonomic discussion. The male genitalia of the specimen from Thailand shows slightly different pseudepiphallic lophi compared to the holotype from Vietnam and examined males from China, with more straight apical branches and less globular apex (Fig. 7L-M). Such differences are however based on too few observations to characterise a new species.

681

682 Xenogryllus maniema Robillard & Jaiswara n. sp.

683 (Figs 1J-K; 3G-H; 4D; 5D; 7F-G; 8E)

684

Type material. Holotype, ♂, Democratic Republic of the Congo: Lokandu, île [island]
Biawa, vii.1939, Lt. Vissers, coll. Mus. Congo (MNHN-EO-ENSIF10686). Paratypes
(5♂), Democratic Republic of the Congo: same information as holotype; 3♂,
identified Xenogryllus eneopteroides by L. chopard, R. Det. B.5318 (MRAC). Katanga:
Kafakumba, ix.1924, 1♂, G. F. Overlaet, coll. Musée du Congo, identified Xenogryllus

690 eneopteroides by L. Chopard, R. Det. N 2814 (MRAC). Congo Belge, Musosa
691 [Mususa], ix.1939, H. J. Brédo, 1³, I.G.13.212 (RBINS).

692 *Type locality.* Democratic Republic of the Congo, Lokandu, island Biawa.

Distribution. Species only known from forested areas in the eastern part of theDemocratic Republic of the Congo.

695 *Etymology.* The species is named after the type locality. Maniema, which means 696 "jungle" or "rain forest" in Kibangubangu dialect, is one of 26 provinces of the 697 Democratic Republic of the Congo.

698 Diagnosis. Species of average size, closer to X. mozambicus n. sp., from which it 699 differs by less rounded face in lateral view (Fig. 3H), pseudepiphallic lophi (Fig. 8E) 700 ended by a long conical apex and with a widened inner membranous margin 701 resembling that of in X. maichauensis. From X. eneopteroides and X. mozambicus, X. 702 maniema differs by absence of T-shaped median band on pronotum, absence of 703 transverse carina on ventral face of lophi, and by rami weak, without convergent hook-704 like apex. Differing from X. lamottei n. sp. and Asian species by strongly carinated 705 lateral angles of pronotum dorsal disc (also carinated in X. mozambicus and X. 706 eneopteroides).

Description. Species of average size, coloration yellow brown little contrasted (Fig. 1J-K). Eyes large, lateral, higher than long, occupying almost half of head height in lateral view (Fig. 3G-H). Face well-rounded in lateral view (less than in *X. mozambicus*), with typical whitish mask underlined by a black line below eyes and on mandibles. Pronotum dorsal disc strongly carinated laterally (Fig. 3H), coloration light brown, with a median dark brown band, not extended laterally near anterior margin; lateral lobes almost homogeneously brown. First article of antennae dark brown.

Male. FWs very wide (Fig. 5D), longer than abdomen; dark coloration anterior to 1A including angle of 1A. FW venation as in *X. eneopteroides;* apical field longer, forming

a long triangle made of five (n=2) cell alignments.

717 Male genitalia (Fig. 7F-G). Pseudepiphallic lophi twice longer than rest of 718 pseudepiphallic sclerite; with a wide membranous inner margin (Fig. 8E); their bases 719 fused until mid-length. Apex of lophi long and pointed, slightly convergent and ended 720 by a thin lamella, without inner dorsal expansions. Ventral blade of lophi with faint 721 longitudinal wrinkles, but without strong transverse carina as in X. eneopteroides. 722 Pseudepiphallic parameres with a strong rectangular ventral lobe. Rami rather weak, 723 their apex almost straight, not forming convergent hooks. Ectophallic apodemes strong, 724 not lamellate. Ectophallic lateral expansions, lateral sclerites of ectophallic fold and 725 endophallic sclerite partly fused, forming a wide ventral sclerotized plate, trifid apically; 726 endophallic apodemes made of wide lateral lamellas and a narrow dorsal crest.

727 *Female*. Unknown.

728 Life history traits. Unknown.

729 Calling song. Unknown.

730 Measurements. See Table 5.

731

732 Xenogryllus marmoratus (Haan, 1844)

733 (Figs 1D-G; 3I-J; 4E; 5E; 7H-I; 8G; 9B; 11CD; 12B; 14)

734

735 *Gryllus (Phalangopsis) marmoratus* Haan, 1844: 235.

Calyptotrypus marmoratus – Saussure 1878: 714 (the figures do not correspond to the
species *X. marmoratus*, which is consistent with the surprising distribution "Java, Iles
de la Sonde" proposed by Saussure, where the species is not distributed); Bolívar
1900[1899]: 805.

740 *Madasumma marmorata* – Chopard 1924: 56: Kirby 1906: 93.

Xenogryllus marmoratus – Chopard 1968: 350; Yin & Liu 1995: 96; Oshiro 1995: 43;
Walker 2010: 27; Robillard & Desutter-Grandcolas 2004a: 578 (calling song); 2004b:
273 (morphological phylogeny); 2006: 644 (molecular phylogeny); 2008: 67; 2011: 637;
Anso *et al.* 2016: 9 (molecular phylogeny); Chintauan-Marquier *et al.* 2016: 62
(molecular phylogeny); Vicente *et al.* 2017: 2203 (historical biogeography); Schneider *et al.* 2017 (tympanum morphology).

747 Xenogryllus marmoratus marmoratus Gorochov, 1992, in Gorochov & Belokobylskij

748 1992: 11 (nominal subspecies); Ichikawa et al. 2000: 276; 2006: 180; Storozhenko &

749 Paik 2007; Storozhenko et al. 2015: 145; Cigliano et al. 2018 (Orthoptera Species File

750 Online); Olivero & Robillard 2017: 1 (mating behavior).

751 Xenogryllus marmoratus unipartitus (Karny, 1915) – Gorochov & Belokobylskij 1992:
752 10 (subspecies).

753 Synonym names:

754 *Heterotrypus unipartitus* Karny, 1915 – Chopard, 1968: 350.

Common names: Pine cricket (English), matsumushi (Japanese = pine insect, waiting
insect); bao ta ling (Chinese = pagoda bell).

757

758 *Type material. Neotype,* [new designation], Japan: Honshu, collection Finot,
759 identified *Calyptotrypus* species nova by A. Finot (MNHN-EO-ENSIF1592).

Additional material examined. Japan: Honshu, Mie, ix.1957, F. Ohmachi, 13, 12, identified Xenogryllus marmoratus de Haan, Xma Robillard morpho (MNHN). Japon, 12, Exposition Universelle 1869, #1276-69, identified Dionymus marmoratus by L. Chopard (MNHN-EO-ENSIF1593). Japon [Japan, no precise locality and date], 13, 12, identified Dionymus marmoratus by L. Chopard, Xma Robillard morpho; 13, 22(MNHN); 12 (NHMW); 13, 20.679, identified X. marmoratus unipartitus by Karny (NHMW); 42, identified Calyptotrypus species nova, collection Finot (MNHN). Kobe, H.

Fruhstorfer, 1[♀], 24.125 (NHMW). Kyoto, I. Yamashiro, 1♂, Xma Robillard morpho 767 MEB, MEB Ziegler; 1º; 1º, molecular sample T. Robillard 2004 (MNHN). Kioto 768 769 [Kyoto], Y. Hirase, 13, identified Madasumma marmorata (Haan) and Calyptotrypus 770 marmoratus (Haan) by Hebard, 1924 (MNHN). Kanagawa Prefecture, Kanate, Ooi-Cho Ashigara-kami-gun [35° 23' 00"N, 139° 08' 00"E], H. Sakai, 14.viii.2010, 1∂, 2♀, 1 771 772 identified Xenogryllus marmoratus by A. Ichikawa (MNHN); 13, iuvenile. 773 enregistrement appel TR-male2 [call recording - MNHN-SO-2016-14364], identified 774 Xenogryllus marmoratus by A. Ichikawa (MNHN-EO-ENSIF1704); 13, enregistrement 775 appel TR-male1 [call recording, MNHN-SO-2016-14365], identified Xenogryllus 776 marmoratus by A. Ichikawa (MNHN-EO-ENSIF1598). Japon, environs de Tokyo, J. 777 Harmand, 1906 (MNHN). Kinki Distr.[ict], Wakayama Pref.[ecture], Hashimoto city, 778 19.x.1986, A. Ichikawa, 1∂, identified X. marmoratus by A. Ichikawa (RBINS). 779 Tsushima, H. Fruhstorfer, sept-oct [ix-x], 24.123 (NHMW). China: Guanxi, Jin Xiu, 10.x.1981, E110°11' N24°07', 1♀ (14062635) (SIPPE). Longzhou, 1995-viii18/23, 780 781 Xianwei Liu, Weinian Zhang, Xinbao Jin, 1^o (14062635) (SIPPE). **Chongqing**, Beibei, E106°23' N29°48', 1.x.2000, Zhou, 1 (14080765) (SIPPE). Anhui, Huangshan, 782 E118°19' 783 N29°43' [Shanghai market], viii.2012, T. Robillard, 13 (TR40), 784 enregistrement appel [call recording MNHN-SO-2018-36] (MNHN-EO-ENSIF1702); 13 785 (TR2), enregistrement appel [call recording MNHN-SO-2018-52], molecular sample X12 (MNHN-EO-ENSIF1594). Jiangsu, Zhenjiang, 32°12'0.00"N 119°27'0.00"E 786 [Shanghai market], viii.2012, T. Robillard, 13 (TR41), enregistrement appel [call 787 recording, MNHN-SO-2018-38] (MNHN-EO-ENSIF1707); 13 (TR3), enregistrement 788 789 appel [call recording MNHN-SO-2018-], molecular sample X13 (MNHN-EO-ENSIF3562). Shanghai, Padang Shanghai, E121° 32' N31° 13' [Shanghai market], 790 791 viii.2012, T. Robillard, 13, molecular sample X11, enregistrement appel [call recording, 792 MNHN-SO-2018-37] (MNHN-EO-ENSIF1599); 13 (MNHN). Shanghai, Prov. Klange[?], 793 Musée Meude, O. Piel, 30.viii.[19]30, 1♀, #1623, 1♂, 6.ix.[19]30 (MNHN). 794 **Guangdong**, Shenzhen Distr., Xichong, 27.ix.2014, coll. Zhang Tao, 4, 2 (SNNU).

795 Henan, Xinxian Distr., 4-8.ix.2014, coll. Ma Libin, 2^o (SNNU). Hainan, Wuzhishan Mt., 12;viii.2010, coll. Jiang Chaozhong, 13 (NWAFU). Zhejiang, Tianlongshan Mt., 796 797 8.vii.2009, coll. He Zhuqing, 13 (ECNU). Shaanxi, Ankang, 10.viii.2017, coll. Ma Libin, 798 13 (SNNU). Taiwan, Takao, Formosa, Sauter XI.[19]07, type of *Heterotrypus* 799 unipartitus, det. Karny, identified Dionymus formosanus n. sp. Type by T. Shiraki, (DEI, 800 Eberswalde), identified Xenogryllus marmoratus unipartitus (Karny) by A. V. Gorochov 801 (DEI) [examined on photo]. Sri Lanka: Ceylon, Kandy, H. Rolle Berlin SW.11, 1 802 (MNHN). South Korea: data from website "South Korea: Orthopteroids of Korea" 803 (http://www.jasa.pe.kr/pulmuchi/korthoptera/Xenogryllus-marmoratus.htm).

Distribution. Japan, China (including Taiwan and Hainan), South Korea
(http://www.jasa.pe.kr/pulmuchi/korthoptera/Xenogryllus-marmoratus.htm), India
(specimens observed by Chopard (1969)), Sri Lanka (1 male specimen).

Emmended diagnosis. Species characterised by its small size (Fig. 1E-G), head dorsal coloration with a faint median dark band on vertex, and male genitalia with very long and thin pseudepiphallic lophi (Fig. 8G); in male FWs (Fig. 5E), mirror wider than long, less rounded than in *X. maichauensis, X. ululiu* and *X. transversus*.

811 Redescription. Species of small to average size, the smallest for the genus (Fig. 1E-G). 812 Coloration almost homogeneously light ochre. Median dark band on head dorsum faint 813 on vertex (Fig. 4E), prolonged anteriorly by black fastigium, with three narrow lateral 814 bands on each side. Fastigium straight, not widened anteriorly. Antennae light brown, 815 first segment darker. Face relatively flat in lateral view (Fig. 3J). Pronotum dorsal disc 816 light brown, with a median brown band variably marked prolonging head coloration; 817 lateral margins not carinated, with a narrow yellow line; lateral lobes ochre, progressively lighter ventrally. 818

Male. FWs light brown, translucent (Fig. 5E); dark coloration anterior to 1A sometimes extended on angle of 1A. Transverse part of file almost straight, with 435 stridulatory teeth (n=1) on transverse part of 1A. Harp as wide as long. Fused part of cells c1 and

b1 as long as individualised part of c1. Cell d2 wider than in other species. Mirror wider
than long, little rounded, its inner limit forming a wide angle. Apical field short, including
five cell alignments.

825 Male genitalia (Fig. 7H-I). Pseudepiphallic sclerite longer than rami; pseudepiphallic 826 lophi very long (Fig. 8G), narrow and sclerotized, slightly convergent, ended by an 827 apical lamella curved dorsally and forming a small notch. Pseudepiphallic sclerite with 828 wide lateral membranous lobes. Basal reinforcement of pseudepiphallic sclerite strong. 829 Rami short with convergent hook-like apex. Pseudepiphallic parametes with a ventral 830 transverse crest absent in other species; inner apex of of pseudepiphallic parameres 831 sharp, slightly sclerotized apically. Ectophallic apodemes strong, not lamellate. 832 Ectophallic lateral arms strong, fused to lateral arms of ectophallic fold, and with lateral 833 arms of endophallic sclerite, forming together a wide gutter. Ectophallic fold entirely 834 sclerotized ventrally. Endophallic sclerite small, apodeme made of a thin anterior dorsal 835 crest and narrow lateral lamellas.

Female. FWs dorsal field with 8-9 strong longitudinal veins (m = 8, n = 10) and faint
transverse ones. Subgenital plate not indented apically (Fig. 9B). Ovipositor longer
than FIII, its apex rounded and smooth.

Female genitalia. Copulatory papilla (Fig. 11C-D) thin and conical, widened basally;
sclerotization strong basally and apically, its membrane plicate.

Life history traits. X. marmoratus lives on low vegetation in bushes and grass in secondary habitats and along roads. The species has a nocturnal activity; males call at night from the vegetation. See Olivero & Robillard (2017) for description of peculiar mating behaviours.

Calling song (Figs 12B, 14). At 26.5°C, the calling song of *X. marmoratus* is made of relatively long syllables, each corresponding to one FW closure. Syllables form a twopart echeme: 1-3 singleton syllables (m = 2, n = 10), followed by a chirp made of three

848 syllables, sometimes duplicated. The amplitude profile of the call shows that the 849 singletons are usually less loud than the chirp. Total echeme duration is 711 ± 33 ms, 850 for an echeme period of 6776 ± 368 ms (echeme duty cycle = 10.5%). The syllables 851 have the following characteristics: 1^{st} syllable duration = 51.8 ± 1.5 ms (period = 318.3) 852 \pm 35.6 ms); 2nd syllable duration = 50.3 \pm 1 ms; chirp duration = 183.8 \pm 1 ms; chirp syllable duration = 49.5 ± 1.9 ms (period = 68 ± 0.8 ms). The frequency spectrum 853 854 shows a pure tone fundamental peak corresponding to the dominant frequency at ca. 855 5.9 \pm 0.2 kHz, with a clear harmonic series, the third harmonic (*ca.* 18 kHz) being 856 almost as powerful as the fundamental peak.

857 *Measurements.* See Table 6.

Taxonomic discussion. The types of De Haan, supposed to be in Leiden, were not found in RMNH in 2006 (TR, pers. obs.), with no record mentioning them as loaned (Rob De Vries, curator of the Orthopteran collection in RMNH, pers. com. 2006; confirmation by Luc Willemse, current curator of the Orthopteran collection in RMNH, pers. com. 2018). Specimens possibly belonging to the original type series were also searched for in other European museums, but could not be found. We thus designate a neotype from Japan that will serve as a reference for future systematic works.

865 Gorochov (1992) distinguished two subspecies, X. m. marmoratus (Haan, 1844) and X. 866 m. unipartitus (Karny, 1915). In his study, he considered Gryllus (Phalangopsis) 867 marmoratus Haan, 1844 (type from Japan), as the nominal subspecies Xenogryllus 868 marmoratus marmoratus, while he established the subspecies X. m. unipartitus based 869 on the species Heterotrypus unipartitus Karny, 1915, which was described from one 870 female from Taiwan (Takao, examined on photo). The type of H. unipartitus clearly 871 corresponds to a female of Xenogryllus, however, given the continuous variation 872 observed across the wide distribution of X. marmoratus, and the limited information 873 available based on female morphology, there is no reason to maintain these two 874 subspecies. We thus synonymise the subspecies under X. marmoratus, and H.

unipartitus becomes a junior synonym of *X. marmoratus*, as previously proposed by
Chopard (1968).

877

878 Xenogryllus mozambicus Robillard n. sp.

879 (Figs 2A-D; 3K-L; 4H; 5F; 7J-K; 8F; 9D; 11H-J; 12C; 15; 16)

880

881 Xenogryllus eniopteroides (wrong spelling of X. eneopteroides) – Toms 1984: 309.

882

883 *Type material. Holotype, ∂*, **Mozambique: Cabo Delgado,** Pantanos de Nhica, zone 884 herbacée à l'Est du camp [herbaceous area E of camp], 10°45'19,1"S 40°13'00"E, 122 885 m, 29.xi.2009, T. Robillard, nuit, enregistrement appel [call recording MNHN-SO-2018-886 140] (TR475) (MNHN-EO-ENSIF1517). Allotype, Q, Mozambique: Cabo Delgado, 887 mare SE de Nhica, bras mort de la Rovuma [pond SE Nhica, dead arm of Rovuma 888 river], 10°45'41,9"S 40°13'31,7"E, 124 m, 27.xi.2009, T. Robillard, mort en élevage 889 [dead in captivity], (MNHN-EO-ENSIF1539). *Paratypes* (8♂, 4♀): **Mozambique: Cabo** 890 **Delgado**, same information as holotype: 1♂, 1♀, morts en élevage [dead in captivity] 891 (MNHN-EO-ENSIF1576-ENSIF1556). Same information as allotype: 23, mort en 892 élevage [dead in captivity] (MNHN-EO- ENSIF1557, ENSIF1508). Mare SE de Nhica, 893 bras mort de la Rovuma [pond SE Nhica, dead arm of Rovuma river], 10°45'41,9"S 894 40°13'31,7"E, 124 m, nuit, zone herbacée en bord de piste [herbaceous area near 895 track] T. Robillard: 22.xi.2009, 1∂ (MNHN-EO-ENSIF1554), 3♀ (MNHN-EO-896 ENSIF1502, ENSIF1503, ENSIF1505); 23.xi.2009, 13 (TR208), enregistrement appel 897 Take 224 [call recording MNHN-SO-2018-133] (MNHN-EO-ENSIF3079); 1∂ (TR206), 898 enregistrement appel Take 222 [call recording MNHN-SO-2018-134], molecular sample 899 X7-XenMoz (MNHN-EO-ENSIF1515). Mare SW de Nhica, bras mort de la Rovuma
[pond SW Nhica, dead arm of Rovuma river], 10°52'09,5"S 40°06'47,1"E, 122 m,
24.xi.2009, nuit, zone herbacée en bord de piste [herbaceous area near track] T.
Robillard: 1 (TR224), hautes herbes (h=1.3 m) [on high grass], enregistrement appel
Take 228 [call recording MNHN-SO-2018-135], molecular sample X23 (MNHN-EOENSIF1559); 1 , mort en élevage (MNHN-EO-ENSIF1581).

Additional material examined. Mozambique: Delagoa [Maputo Bay], xii.1898, 1 ♂,
identified Phormincter species nova by A. Finot (MNHN). South Africa: Zululand,
Mtunzini, 8.i.1952, R. Toms, 1♂, R146, identified Xenogryllus eneopteroides by B. C.
Townsend, 1984, B.M.1983-166 (NHMUK 010926568). Malawi: Fish Eagle Bay Lodge,
herbaceous area near lake Malawi (Mal7), S13°02'21.1" E34°19'34.8", 503 m,
6.x.2018, night, 2♂, call recording, 1♀, T. Robillard, K. Salazar & R. Murphy (MNHN).

911 *Type locality.* Mozambique, Cabo Delgado, Pantanos of Nhica, 10°45'19,1"S
912 40°13'00"E.

913 Distribution. Mozambique, South Africa, Malawi.

914 *Etymology.* Species named after the type locality.

915 Diagnosis. Species of average to large size, close to X. encopteroides and X. maniema 916 n. sp. From X. encopteroides, the new species differs by rather larger size, face more 917 rounded, almost globular in lateral view, absence of T-shaped median band on 918 pronotum (also absent in X. maniema) larger mirror, and slight differences in male 919 genitalia. From X. maniema n. sp., X. mozambicus mostly differs by male genitalia with 920 larger lophi (Fig. 8F) and presence of transverse carina on ventral face of lophi (also 921 present in X. eneopteroides). As X. maniema and X. eneopteroides, X. mozambicus 922 differs from X. lamottei n. sp. and Asian species by strongly carinated lateral angles of 923 pronotum.

924 Description. Species of average size (Fig. 2A-D), coloration light brown little contrasted.
925 Eyes little prominent laterally (Fig. 3K), higher than long, occupying almost half of head

height in lateral view. Face globular in lateral view (Fig. 3L), with typical whitish mask
underlined by a black line below eyes and on mandibles. Pronotum dorsal disc strongly
carinated laterally, coloration light brown, with a median dark brown band, not extended
laterally near anterior margin; lateral lobes almost homogeneously dark brown. First
article of antennae dark brown.

Male. FWs very wide, longer than abdomen; dark coloration anterior to 1A including
angle of 1A (Fig. 5F). Stridulatory file with 505 teeth (n=1) on transverse part of 1A. FW
venation as in *X. eneopteroides,* mirror wider and apical field longer, forming a long
triangle including 5-6 (n=8) cell alignments.

Male genitalia (Fig. 7J-K). Similar to *X. eneopteroides,* except longer pseudepiphallic
lophi (Fig 8F); transverse carina on ventral face of lophi present but weak.

937 *Female* (Fig. 2C-D). FWs dorsal field elongate, with 8-9 longitudinal veins (m = 9, n =
938 6). Subgenital plate with a shallow apical indentation (Fig. 9D). Ovipositor short, about
939 0.6 times FIII length.

940 *Female genitalia.* Copulatory papilla (Fig. 11H-J) as in *X. eneopteroides,* conical and
941 narrow, well-sclerotized except base and apex.

942 Life history traits. X. mozambicus lives in areas of wet savannah (Fig. 15). Individuals 943 of both sexes are found only at night in herbaceous vegetation, and males sing at night 944 from the vegetation (*ca.* 30-80 cm high). Toms (1984) demonstrated that this species 945 have directional calls and turn while calling, giving rise to changes in sound intensity.

946 *Calling song* (Figs 12C, 16). At 21.5 °C, males of *X. mozambicus* emit almost 947 continuously long bouts of highly musical calling songs. After a warming phase, call 948 bouts are made of successions of echemes of 23 ± 2 syllables, barely separated from 949 each other (echeme duration = 3.13 ± 0.29 s; echeme period 3.21 ± 0.30 s, echeme 950 duty cycle = 97.6 %), with a regular amplitude profile, except for the last syllable, which 951 is more intense. Within echemes, syllables are initially organized in 4 ± 1 doublets,

952 which are followed by a series of similar syllables. Syllables are very long (syllable 953 duration = 126 ± 5 ms), with a relatively short syllable period of 137 ± 8 ms (syllable 954 duty cycle = 92%). All syllables are characterized by a large amplitude modulation: in 955 initial doublets within echemes, the first syllables are less intense than the second ones 956 and they start with a lower amplitude than their end (initial amplitude < ending 957 amplitude), contrary to the second syllables of the doublets (initial amplitude > ending 958 amplitude); the rest of the syllables have a higher starting amplitude, except for the last 959 syllable of each echeme, which has a higher ending amplitude. The frequency 960 spectrum shows a pure-tone dominant frequency at 3.3 ± 0.01 kHz, followed by a rich 961 harmonic content including three powerful harmonics at ca. 6.6, 9.9, 13.2 kHz, 962 especially visible in ending syllables.

963 Measurements. See Table 7.

964 *Taxonomic discussion.* The specimens observed by Toms (1984) and identified 965 *Xenogryllus eniopteroides* (wrong spelling for *X. eneopteroides*) from Zululand 966 (Mtunzini and Eastern Transvaal (Clanor)), South Africa, belong to *X. mozambicus* n. 967 sp. according to the male specimen observed from the series of Toms (NHMUK 968 010926568).

969

970 Xenogryllus transversus (Walker, 1869)

- 971 (Figs 2E-H; 3O-P; 4E; 5G; 7O-P; 8I; 9G; 11M-O; 12D; 17)
- 972

973 *Platydactylus transversus* Walker, 1869: 88.

- 974 *Cardiodactylus? transversus* Saussure 1878: 665.
- 975 *Madasumma transversa* Kirby 1906: 94; Chopard 1968: 350.

- *Xenogryllus transversus* Chopard 1969: 310; Bhowmik 1976: 46; 1985: 67; Saeed *et al.* 1990: 309 (redescription); Yin & Liu 1995: 96; Nattier *et al.* 2011: 2199 (molecular phylogeny); Vicente *et al.* 2017: 2203 (historical biogeography); He 2018: 516; Cigliano *et al.* 2018 (Orthoptera Species File Online).
- 980 Synonym names:
- 981 *Dionymus calcaratus* Brunner von Wattenwyl, 1893: 213 Chopard 1968: 350.
- 982 Calyptotrypus roonwali Bhowmik, 1977: 35 Vasanth 1993: 131.
- 983

984 *Type material. Holotype, ∂*, **Bangladesh:** Silhat [= Silhet / Sylhet] (NHMUK010362937)
985 [examined].

Additional examined material. India: Meghalaya, Jaintia Hills, 12, Narpuh R.F. 986 Umpyrsung - off R. Lubha Sunapor Town, N25°06.683' E92°21.559', 149 ft. 987 988 N25°06.983' E92°21,559', Elev.149ft, 19.x.2004, molecular sample Xtr-X2, Svenson 989 (MNHN-EO-ENSIF87). India, 1[°], #6752 (MNHN). **Sikkim**, 1[°], identified *Dionymus* 990 calcaratus Br. by L. Chopard (MNHN-EO-ENSIF1516). Lebong, 3000 ft, IX.1908, 12, 991 H.M.L., identified *Dionymus calcaratus* Br. (NHMUK010362936). Manipur, Imphal 992 Valley, alt 715 m., 11.x.1945, 1∂, HT of Calyptotrypus roonwali Bhowmik (ZSI). 993 Jammu, Tawi river, 32.715736° 74.860403°, secondary area, 30.VII, 11 pm, 13, 994 chorus, molecular sample X26 (MNHN-EOENSIF4395), 13 (MNHN). Punjab, IISER Mohali, 30.663169° 76.724212°, 6♂ (MJO 765, 766, 411, 715, 413), 2 ♀ (MJO 36, 995 996 MJO 726), molecular sample Xtr-715, 765, 766, R. Jaiswara (IISER Mohali). Myanmar: Bhamo, Birmania, Fea ix.1886, Museo Civ. Genova, 19.176, syntype of 997 Dionymus calcaratus (NHMW). Pakistan: Mansehra, 34.333882° 73.201062°, 27.ix, 998 999 1³, W. A. Panhwar, molecular sample X25 (MNHN-EO-ENSIF4388).

1000 *Type locality.* Bangladesh, Silhet.

1001 *Distribution*. Myanmar, India (North and East), Bangladesh, Pakistan.

Emended diagnosis. Species of large size, similar in size, venation and coloration to *X*. *maichauensis*, from which it differs by male genitalia, with long sclerotized pseudepiphallic lophi ended by a sharp apex with a dorsal preapical pointed expansion (Fig. 8I); from *X. ululiu, X. transversus* differs by its larger size and by larger pseudepiphallic sclerite in male genitalia, with lophi proportionally smaller, separated by a deep indentation (lophi basally fused in *X. ululiu*).

1008 *Redescription.* In addition to the characters of the genus, *X. transversus* has a large 1009 size and a golden coloration (Fig. 2E-H). Fastigium 1.5 times wider than scape, slightly 1010 widened apically as in *X. ululiu* and *X. maichauensis*. Eyes rather small, located on 1011 face, restricted to the dorsal third of head in lateral view (Fig. 3O-P). Lateral angle of 1012 dorsal disc of pronotum not carinated, as in *X. maichauensis* and *X. ululiu*, with a thin 1013 yellow band underlined by a black line anteriorly. Hind wings tail gray brown, nearly as 1014 long as long as pronotum.

1015 *Male.* Pronotum dorsal disc forming a wide trapezoid, with a wide median black 1016 longitudinal band pronlonging vertex coloration; posterior margin slightly bisinuate. FW 1017 venation (Fig. 5G): 1A forming a 90° angle, with 265 stridulatory teeth (n=1) on 1018 transverse part of 1A. Dark coloration anterior to 1A including angle of file. Harp wide. 1019 Mirror large, well-rounded, its inner limit forming a wide curve. Apical field forming a 1020 triangle longer than wide, with six cell alignments.

Male genitalia (Fig. 7O-P). Pseudepiphallic sclerite elongate, with a wide basis; lophi separated by a large indentation (Fig. 8I), similar to that of *X. ululiu*, with a sharp apex and a strong dorsal preapical expansion. Rami strong, convergent apically. Ectophallic apodemes strong, lamellate apically. Ectophallic fold and endophallic sclerite almost fused, forming a long ventral gutter, wider than in *X. ululiu*, trifurcate posteriorly. Endophallic apodemes made of lateral lamellas.

Female (Fig. 2G-H). Head slightly wider than pronotum. Dorsal disc of pronotum almost
 rectangular, its posterior margin slightly bisinuate. FWs slightly longer than abdomen,

light brown, with anterior dark spot very small; dorsal field with 10-11 (n = 3) strong
longitudinal veins. Subgenital plate with a deep V-shaped apical indentation with sharp
edges (Fig. 9G). Ovipositor as long as FIII.

1032 *Female genitalia*. Copulatory papilla conical (Fig. 11M-O), wider than in *X*.
1033 *maichauensis*, its apex rounded and sclerotized.

1034 Life history traits. X. transversus lives in open secondary habitats, where males usually 1035 call in chorus from 120-180 cm height dense shrubs. In IISER Mohali (Northern India), 1036 two males were seen calling from a bamboo plantation on one occasion, from a height 1037 of 60 cm, where one male was probably trying to mount the other (RJ personal obs.). In 1038 Jammu (Northern India), males were found calling from cannabis plant. Calling activity 1039 starts late in the evening, almost around 22:00 hrs and continues until 03:00 hrs in the 1040 morning. Spacing between calling males of this species is guite variable, the closest 1041 males being spaced by approximately one meter horizontally.

1042 Calling song. (Figs 12D, 17). The calling songs of three males were recorded from 1043 IISER Mohali campus. At 25-27 °C, the calling song of X. transversus is made of short 1044 echemes quickly repeated and lasting for 197 ± 4 ms (echeme period = 873 ± 13 ms), 1045 and composed of 4-5 long syllables (m = 4.5 ± 0.5). Within echemes, the syllables 1046 usually show increasing amplitudes. Syllables are rather long (duration = 31 ± 4 ms) 1047 with a syllable period of 46 ± 3 ms (syllable duty cycle = 71%). The frequency spectrum 1048 shows a pure-tone dominant frequency at 4.5 ± 0.5 kHz followed by two powerful 1049 harmonics.

1050 *Measurements.* See Table 8.

1051 *Taxonomic discussion.* We confirm the status of junior synonyms of *Calyptotrypus* 1052 *roonwali* and *Dionymus calcaratus* according to re-examination of type specimens (HT 1053 of *C. roonwali* and one male ST of *D. calcaratus*).

1054

1055 Xenogryllus ululiu Gorochov, 1990

1056 (Figs 2I-L; 3M-N; 4G; 5H; 7Q-R; 8H; 9E; 11K-L; 12E; 18)

1057

Xenogryllus ululiu Gorochov, 1990: 14 – Gorochov 1992: 10; Kim & Pham 2014: 60;
Robillard *et al.* 2007: 1267 (song frequency); Cigliano *et al.* 2018 (Orthoptera Species
File Online).

Type material. Holotype, ∂, **Vietnam: Gia Lai**, Kannack, A.V. Gorochov (ZIN) [not
examined]. *Paratype (1∂),* **Vietnam: Gia Lai**, Kannack, A.V. Gorochov: 8-16.ii.1988,
1*∂* (MNHN-EO-ENSIF2809) [examined].

1064 Additional material examined. Vietnam: Gia Lai, 2.ii.1993, 12, identified Xenogryllus 1065 ululiu by A.V. Gorochov, molecular sample XulV (MNHN-EO-ENSIF2810); 7.XI.1993, 1066 1⁽²⁾, molecular sample XulV2, A.V. Gorochov. Thailand: Petchabun, Nam Nao, 16° 47' 1067 N, 101° 27' O, 1000 m, S. Ingrisch; 13.ix.1989-14.ix.1989, S. Ingrisch, 13, Stridulation recorded by S. Ingrisch 0291XENO.WAV, mixed Oak-Pine forest, grassy undergrowth, 1068 1069 Bamboo thicket [day collecting and night collecting following stridulation], identified 1070 Xenogryllus ululiu by S. Ingrish, molecular sample XulTh (ZFMK). Loei, NamNao NP at 1071 night, 13(14.ix.1989, S. Ingrisch, 13, identified Xenogryllus ululiu by S. Ingrish, 1072 molecular sample XulTh (ZFMK). Cambodia: Kampong Spoe, northern part of Elefan 1073 mts [Dâmrei Mountains], Kiri-Rom National Park, 130 km NNE Sihanoukville, 300-1074 500m, 27.ix-1.x.2003, A. Gorochov, M. Berezin, 1⁽²⁾, molecular sample X21-XulCam2, 1075 1º (ZIN). Kaoh Kong, central part of Elefan mountains [Dâmrei Mountains], vill. 1076 Styeng-Chkhral (100 km NE of Sihanoukville), 300-500m, 27.viii-6.ix.2003, A. 1077 Gorochov, L. Anisyutkin, 1♂, molecular sample X20-XulCam1 (MNHN-EO-1078 ENSIF4385), 13 (ZIN). Campot, environs of Sihanoukville (=Kampong Som), 14-1079 21.ii.1998, A.V. Gorochov, 1^o (ZIN). Sihanoukville, citi near Siam bay, environs, 22-26.VIII.2003, A. Gorochov, L. Anisyutkin, 13, molecular sample X19-XulSi (ZIN). 1080

1081 China: Guangdong, Shenzhen District, Wutongshan Mt., 21.ix.2014, coll. Zhang Tao,
1082 5♂, 3♀, (SNNU).

1083 *Type locality.* Vietnam, Gia Lai, Kannack.

1084 *Distribution* (Fig. 16). Vietnam, Thailand, Cambodia, Southern China.

Emended diagnosis. Species average to large size, close to *X. transversus* and *X. maichauensis* in terms of shape and coloration, from which it differs by smaller size, mirror slightly wider than long (less rounded than in in *X. transversus* and *X. maichauensis*), and male genitalia looking like a stockier version of that of *X. transversus*, with pseudepiphallic lophi basally fused together (Fig. 8H), with a sharp posterior apex and a strong dorsal hook-like expansion. In females, head larger than in other species, wider in dorsal view than rest of body.

1092 *Redescription.* In addition to the characters of the genus, *X. ululiu* has an average size 1093 and a light brown or golden brown coloration (Fig. 2I-L). Head (Fig. 3M-N): Fastigium 1094 slightly widened apically, as in *X. transversus* and *X. maichauensis* (Fig. 4G); eyes 1095 rather small, restricted to the dorsal third of head in lateral view (Fig. 3N). Pronotum 1096 dorsal disc with a wide median black longitudinal band, its lateral angles not carinated, 1097 with a thin yellow band underlined by a black line ventrally. Hind wings tail gray brown, 1098 near twice as long a pronotum.

1099 *Male.* FW venation (Fig. 5H): 1A forming a straight angle. Dark coloration anterior to 1A 1100 not including angle of file. Harp wide, with one short incomplete oblique vein and two 1101 complete ones slightly bisinuate. Mirror large, slightly wider than long, less rounded 1102 than in *X. maichauensis* and *X. transversus,* its inner margin curved as in these 1103 species. Apical field forming a triangle longer than wide, with 6-7 (n = 5) cell 1104 alignments.

1105 *Male genitalia* (Fig. 7Q-R). Pseudepiphallic lophi stocky, basally fused together, with a 1106 sharp apex and a strong dorsal preapical expansion (Fig. 8H). Rami strong, convergent

apically. Ectophallic apodemes strong, lamellate apically. Ectophallic fold and
endophallic sclerite almost fused, forming a long ventral gutter, trifurcate posteriorly
and extended anteriorly between rami. Endophallic apodemes made of anterior lateral
lamellas.

Female. Head larger than in male (Fig. 2K-L), wider than rest of body in dorsal view.
Dorsal disc of pronotum almost rectangular, its posterior margin slightly bisinuate. FWs
light brown, anterior dark spot small. Dorsal field with nine strong longitudinal veins.
Subgenital plate with a wide, apical U-shaped indentation with sharp edges (Fig. 9E).
Ovipositor as long as FIII.

1116 *Female genitalia.* Copulatory papilla small, conical, its apex rounded and sclerotized1117 (Fig. 11K-L).

Life history traits. According to S. Ingrish who recorded the species' call in Thailand, males of *X. ululiu* sing at night in grassy undergrowth of mixed Oak-Pine forest or bamboo thicket (Cigliano *et al.* 2018).

1121 *Calling song* (Figs 12E, 18). One male from Thailand was recorded by S. Ingrish at 1122 21°C. The song consists of echemes during 566 \pm 57 ms, with a period of 2041 \pm 358 1123 ms. Echemes are made of 4 or 5 syllables (m=5), with syllable duration = 108 \pm 3.8 ms 1124 and syllable period = 120.3 \pm 3 ms. Within echemes, the three starting syllables have a 1125 lower amplitude than the final 1-2 syllables. Dominant frequency is 4.8 \pm 0.4 kHz and 1126 corresponds to the fundamental frequency of the spectrum.

1127 Measurements. See Table 9.

1128

1129 Discussion

1130 The taxonomic revision of *Xenogryllus* is based on the thorough study of 1131 specimens from natural history collections belonging to 17 institutions and on field work 1132 in Mozambique, China and India. Our study reveals that the genus is relatively little

diversified compared to genera of other tribes of the subfamily Encopterinae. The
revision clarifies the status of all previously described species and subspecies (Table
1), and added three new species from Africa.

1136 The large geographical coverage of the available data for Xenogryllus suggest 1137 that the most common species of the genus are now clearly defined, formally described 1138 or redescribed and included in identification keys. However two of the three new species described here are based on few specimens only (six males from three 1139 1140 localities in Congo for X. maniema n. sp. and a unique male from Guinea for X. lamottei 1141 n. sp.). As a consequence, while some species of Xenogryllus have wider distributions 1142 across Asia and Africa, where they occur mostly in savannahs and in open shrubby 1143 areas, other species may have more limited distributions, perhaps in association with 1144 more constrainted and limited forested habitats. This would involve an undefined 1145 proportion of endemic taxa yet to discover.

1146

1147 Acknowledgements

1148 We acknowledge the Indo-French Centre for Promotion of Advanced Research 1149 (CEFIPRA) for funding this project (PIs: Rohini Balakrishnan & Laure Desuter-1150 Grandcolas), and the National Biodiversity Authority of the Government of India for 1151 providing the necessary permits to allow us to carry out the fieldwork. The collaboration 1152 between France and China was facilitated by CAMPUS FRANCE and the program 1153 PHC XU GUANGQI (2012). This work was conducted in the context of the PhD thesis 1154 of JD, which was funded by China Scholarship Council (CSC), the Innovation Funds of 1155 Graduate Programs of Shaanxi Normal University [2012CXB019], the National Science 1156 Foundation of China [Grant No. 31402006] and a supporting grant from "La Société 1157 des Amis du Muséum" and the Institut de Systématique, Evolution, Biodiversité 1158 (ISYEB). We thank Jérôme Constant and Carole Paleco (RBINS) for their help during 1159 the study of the collections of the Royal Belgian Institute of Natural sciences, Brussels,

1160 funded by the SYNTHESYS European program (BE-TAF-6640); Andrej Gorochov 1161 (ZIN) for providing samples from the collections of Saint Petersburg; Rob de Vries, 1162 Caroline Pepermans and Luc Willemse (RMNH) for their help with the Leiden 1163 collections, and the following persons for additional loans of material or photos: 1164 Merdedes Paris (MNCN), Stéphane Hanot (MRAC), Stephan Blank (Müncheberg) and 1165 Sigfrid Ingrisih (ZFMK). We also thank Simon Poulain (MNHN) for his help 1166 photographing the specimens, and Karen Salazar (MNHN) for her help with line 1167 drawing figures. We also thank Rohini Balakrishnan (CES, IISc Bangalore) and Manjari 1168 Jain, (IISER Mohali) for supporting RJ with logistics and lab space to carry out part of 1169 this work in India. We also thank the Orthopterist Society for supporting RJ with one-1170 month project grant to visit MNHN in 2017. We thank Jean-Yves Rasplus (CBGP, 1171 Montpellier) and Oliver Pascal for organising the 2009 expedition in Mozambique; this 1172 expedition was made possible by generous contributions from the Prince Albert II of 1173 Monaco Foundation and the Stavros Niarchos Foundation; we would like to thank Pro-1174 Natura International and the Muséum national d'Histoire naturelle, Paris, for organising 1175 the expedition under their Our Planet Reviewed programme, and the Instituto de Investigação Agrária de Moçambique for helping with the administration and the 1176 1177 collecting permits issue.

1178

1179 **References**

Anso, J., Barrabe, L., Desutter-Grandcolas, L., Jourdan, H., Grandcolas, P., Dong, J. &
Robillard, T. (2016) *Pixibinthus*, a new cricket genus endemic in New Caledonia:
phylogenetic study of a potential relict lineage from maquis minier vegetation. *PLoS One*, 11, e0150920. https://doi.org/10.1371/journal.pone.0150920.
Audacity Team (2016) Audacity(R): Free Audio Editor and Recorder [Computer

1185 application]. Version 2.0.0 retrieved 12 January 2016 from1186 https://audacityteam.org/.

- 1187 Bhowmik, H.K. (1976) On the gryllid fauna (Gryllidae: Orthoptera) of the districts of the
- 1188 North Bengal, with description of three new species. *Indian Museum Bulletin*, 11,
 1189 42–48.
- 1190 Bhowmik, H.K. (1977) Studies on Indian Crickets (Orthoptera: Insecta) with
- 1191 Descriptions of Two New Species. *Records of Zoological Survey of India*, 73,
 1192 368–382.
- 1193 Bhowmik, H.K. (1985) Contribution to the gryllid fauna of the Western Himalayas 1194 (Orthoptera: Gryllidae). *Records of the Zoological Survey of India*, 73, 1–74.
- Bolivar, I. (1890) Ortópteros de Africa del Museo de Lisboa. *Jornal de Sciencias Mathematicas, Physicas e Naturaes*, Series 2, 73–232.
- Bolívar, I. (1900[1899]) Les Orthoptères de St-Joseph's College à Trichinopoly (Sud de
 l'Inde). Annales de la Société Entomologique de France, 68, 761–812.
- Bolivar, I. (1910) Fauna de la Guinea Espanola. *Memorias de la Real Sociedad Espanola de Historia Natural,* Tomo 1, 525–544.
- Brunner von Wattenwyl, C. (1893) Révision du système des Orthoptères et description
 des espèces rapportées par M. Leonardo Fea de Birmanie. *Annali del Museo civico di storia naturale di Genova*, 2, 13, 5–230.
- Chintauan-Marquier, I.C., Legendre, F., Hugel, S., Robillard, T., Grandcolas, P., Nel,
 A., Zuccon, D. & Desutter-Grandcolas, L. (2016) Laying the foundations of
 evolutionary and systematic studies in crickets (Insecta, Orthoptera): a multilocus
- 1207 phylogenetic analysis. *Cladistics*, 32, 54–81. <u>https://dx.doi.org/10.1111/cla.12114</u>
- 1208 Chopard, L. (1928) Revision of the Indian Gryllidae. *Records of Indian Museum*, 30, 1–
 1209 36.
- 1210 Chopard, L. (1954) La Réserve naturelle intégrale du Mt. Nimba, III. Orthoptères
 1211 Ensifères, *Mémoires de l'Institut Français d'Afrique Noire*, 40, 25–97.

1212 Chopard L. (1968) Gryllides. Fam. Gryllidae: Subf. Mogoplistinae, Myrmecophilinae,
1213 Scleropterinae, Cachoplistinae Pteroplistinae, Pentacenturinae, Phalangopsinae,
1214 Trigonidinae, Eneopierinae; Fam.Oecanthidae, Gryllotalpidae. *In:* Beier, M. (Ed.),
1215 Orthopterorum Catalogus. Vol. 12. Uitgeverij Dr. W. Junk N.V.'s, Gravenhage,
1216 pp. 213–500.

- 1217 Chopard, L. (1969) *The fauna of India and adjacent countries.* Orthoptera. Vol. 2.
 1218 *Grylloidea.* Zoological Survey of India, Calcutta: Baptist Mission Press, 421 pp.
- 1219 Cigliano, M.M., H. Braun, D.C. Eades & Otte, D. (2018) Orthoptera Species File.
 1220 Version 5.0/5.0. [Retrieved 10 May 2018]. http://Orthoptera.SpeciesFile.org
- 1221 Desutter, L. (1987) Structure et évolution du complexe phallique des Gryllidea
 1222 (Orthoptera) et classification des genres néotropicaux de Grylloidea. 1re partie.
 1223 Annales de la Société entomologique de France, Nouvelle Série, 23, 213–239.
- Desutter-Grandcolas, L. (2003) Phylogeny and the evolution of acoustic
 communication in extant Ensifera (Insecta, Orthoptera). *Zoologica Scripta*, 32,
 525–561. https://doi.org/10.1046/j.1463-6409.2003.00142.x
- Dong, J., Kergoat, G.J., Vicente, N., Rahmadi, C., Xu, S. & Robillard, T. (2018)
 Biogeographic patterns and diversification dynamics of the genus Cardiodactylus
- Saussure (Orthoptera, Grylloidea, Eneopterinae) in Southeast Asia. *Molecular Phylogenetics and Evolution*, 129 (2018) 1–14
- 1231 https://doi.org/10.1016/j.ympev.2018.06.001

1232 Giglio-Tos, E. (1907) Ortotteri africani. Parte II, Blattodea, Mantodea, Phasmodea,

- Locustodea, Gryllodea. Bulletino dei Musei di Zoologia ed Anatomia comparata,
 della R. Universita di Torino, 22, 563, 1–26.
- Gorochov, A.V. (1990) New and insufficiently studied crickets (Orthoptera, Gryllidae)
 from Vietnam and some other territories. *In:* Gorochov, A.V. [Ed.]. *News of*systematics and faunistics of Vietnam insects. Part 1. Trudy Zoologicheskogo

- Instituta. Akademiia nauk SSSR. Leningrad [= Proceedings of the Zoological
 Institute, USSR Academy of Sciences, Leningrad] 209, pp. 3–28.
- 1240 Gorochov, A.V. (1992) New and little known crickets (Orthoptera, Gryllidae) from 1241 Vietnam. *Trudy Zoologicheskogo Instituta Akademiya Nauk SSSR*, 245, 3–16.
- Haan, D. (1844) Verhandelingen over de Natuurlijke Geschiedenis der Nederlansche
 Overzeesche Bezittingen, 24, 235. https://doi.org/10.5962/bhl.title.114730
- 1244 Ichikawa, A. Murai, T. Honda, E. (2000) Monograph of Japanese crickets (Orthoptera;
 1245 Grylloidea). *Bulletin of the Hoshizaki Green Foundation*, 4, 257–332.
- 1246 Ichikawa, M. & Honda, E. (2000) Bulletin of the Hoshizaki Green Foundation, 4, 276.
- 1247 Karny, V.H. (1915) Orthoptera et Ootbecaria. Supplementa Entomologica, 4, 56–108.
- 1248 Karsch, F. (1893) Verzeichniss der von Herrn Dr. Paul Preuss in Kamerun 1249 gesammelten Grillen. *Entomologische Nachrichten*, 19, 199–208.
- 1250 Kevan, D.K. & Knipper, H. (1961) Geradflügler aus Ostafrika. *Beiträge zur* 1251 *Entomologie*, 11, 356–413.
- 1252 Kirby, W. F. (1906) A synonymic catalogue of Orthoptera (Vol. 2). Orthoptera
 1253 Saltatoria. Part I. (Achetidae et Phasgonuridae). The trustees of the British
 1254 Museum, London, pp. viii+562
- Nattier, R., Robillard, T., Desutter-Grandcolas, L., Couloux, A., & Grandcolas, P. (2011)
 Older than New Caledonia emergence? A molecular phylogenetic study of the
 eneopterine crickets (Orthoptera: Grylloidea). *Journal of Biogeography*, 38,
 2195–2209. https://doi.org/10.1111/j.1365-2699.2011.02563.x
- Oki, S., Hayashi, I., Iwatsuki, N. & Morikawa, K. (2002) Study on sound generating
 mechanism of *Xenogryllus marmoratus*. *Proceedings of Dynamics & Design Conference*, 95.

- Olivero, P. & Robillard, T. (2017) Same-sex sexual behavior in *Xenogryllus marmoratus* (Haan, 1844) (Grylloidea: Gryllidae: Eneopterinae): Observation in the wild from YouTube. *Journal of Orthoptera Research*, 26,1–5.
 https://doi.org/10.3897/jor.26.14569
- 1266 Oshiro, Y. (1995) *Studies on the Gryllidae from the Ryukyu Islands*. Nakimushikai,
 1267 Okinawa, 131 pp.
- Ragge, D.R. (1955) *The wing-venation of the Orthoptera Saltatoria, with notes on Dictyopteran wing-venation*. British Museum (Natural History), London, 159 pp.
- 1270 Ragge, D.R., Reynolds, W.J. & Natural History Museum (London, England) (1998).
- 1271 The Songs of the Grasshoppers and Crickets of Western Europe. Harley Books,
- 1272 Colchester, England, 612 pp. https://doi.org/10.2307/3503528
- 1273 Robillard, T. & Desutter-Grandcolas, L. (2004a) High-frequency calling in Eneopterinae 1274 crickets (Orthoptera, Grylloidea, Eneopteridae): adaptive radiation revealed by 1275 phylogenetic analysis. *Biological Journal of the Linnean Society*, 83, 577–584.
- 1276 https://doi.org/10.1111/j.1095-8312.2004.00417.x
- Robillard, T. & Desutter-Grandcolas, L. (2004b) Phylogeny and the modalities of
 acoustic diversification in extant Eneopterinae (Insecta, Orthoptera, Grylloidea,
 Eneopteridae). *Cladistics*, 20, 271–293. https://doi.org/10.1111/j.10960031.2004.00025.x
- Robillard, T. & Desutter-Grandcolas, L. (2006) Phylogeny of the cricket subfamily
 Eneopterinae (Orthoptera, Grylloidea, Eneopteridae) based on four molecular loci
 and morphology. *Molecular Phylogenetics and Evolution*, 40, 643–661.
- 1284 https://doi.org/10.1016/j.ympev.2005.10.019
- 1285 Robillard, T. (2006) Phylogenetic systematics of *Pseudolebinthus*, a new genus of 1286 Eneopterinae crickets (Orthoptera, Grylloidea, Eneopteridae) from south-east

1287 Africa. Systematic Entomology, 31, 671–683. https://doi.org/10.1111/j.13651288 3113.2006.00347.x

Robillard, T. & Desutter-Grandcolas, L. (2008) Clarification of the taxonomy of extant
crickets of the subfamily Eneopterinae (Orthoptera: Grylloidea; Gryllidae). *Zootaxa*, 1789, 66–68.

Robillard, T. & Desutter-Grandcolas, L. (2011) Evolution of calling songs as
multicomponent signals in crickets (Orthoptera: Grylloidea: Eneopterinae). *Behaviour*, 148, 627–672.

Robillard, T., Grandcolas, P. & Desuter-Grandcolas, L. (2007) A shift toward harmonics
for high-frequency spectra in Eneopterinae crickets (Orthoptera, Grylloidea,
Eneopteridae). *Canadian Journal of Zoology*, 85, 1264–1275.
https://doi.org/10.1139/z07-106

Robillard T, Montealegre-Z, F., Desutter-Grandcolas, L., Grandcolas, P. & Robert, D.
(2013) Mechanisms of high-frequency song generation in brachypterous crickets
and the role of ghost frequencies. *Journal of Experimental Biology*, 216, 2001–
2011.

Saeed, A. & Yousuf, M. (1990) New record of family Eneopteridae (Grylloidea:
Orthoptera) from Pakistan. *Pakistan Journal of Zoology*, 22, 309.

Saussure, H. D. (1878) Mélanges Orthoptérolgiques. Memoires de la Société de
physique et d'histoire naturelle de Genève, 25, 1–836.

Schneider, E.S., Römer, H., Robillard, T. & Schmidt, A.K.D. (2017) Hearing with
exceptionally thin tympana: Ear morphology and tympanal membrane vibrations
in encopterine crickets. *Scientific Reports*, 7, 15266.
https://doi.org/10.1038/s41598-017-15282-z

1311 Storozhenko, S.Y. & Paik, J.C. (2007) *Orthoptera of Korea*. Dalnauka, Vladivostok, 232
1312 pp.

- 1313 Storozhenko, S.Y., Kim, T.W. & Jeon, M.J. (2015) *Monograph of Korean Orthoptera*.
 1314 National Institute of Biological Resources, 145 pp.
- 1315 Su, Y.N. (2016) A simple and quick method of displaying liquid-preserved
 1316 morphological structures for microphotography. *Zootaxa*, 4208, 592–593.
 1317 https://doi.org/10.11646/zootaxa.4208.6.6
- ter Hofstede, H.M., Schoneich, S., Robillard, T. & Hedwig, B. (2015) Evolution of a
 communication system by sensory exploitation of startle behavior. *Current Biology*, 25, 1–8. https://doi.org/10.1016/j.cub.2015.10.064
- Vasanth, M. (1993) Studies on crickets (Insecta: Orthoptera: Gryllidae) of northeast
 India. *Records of the Zoological Survey of India*, Occasional Paper, Calcutta,
 1323 132, 1–178.
- Vicente, N., Kergoat, G.J., Dong, J., Yotoko, K., Legendre, F., Nattier, R. & Robillard,
 T. (2017) In and out of the Neotropics: historical biogeography of Eneopterinae
 crickets. *Journal of Biogeography*, 44, 2199–2210.
 https://doi.org/10.1111/jbi.13026
- Walker, F. (1869) Catalogue of the Specimens of Dermaptera Saltatoria and
 Supplement to the Blattariae in the Collection of the British Museum, 224 pp.
 https://doi.org/10.5962/bhl.title.8149
- Walker, B.L. (2010) *Toxic Archipelago: a history of industrial disease in Japan*. The
 University of Washington press, 352 pp.
- Yin, H. & Liu, X.W. (1995) Synopsis on the classification of Grylloidea and
 Gryllotalpoidea from China. Shanghai Science and Technology and Literature
 Press, Shanghai.

- 1337 **Tables**
- 1338
- 1339 Table 1.

List of species and revised status of *Xenogryllus* species based on Robillard (2006) and Orthoptera Species File Online (Cigliano *et al.* 2018). Junior synonym names for which the status is not updated are not considered in this list but are presented in the revision under the senior name.

Таха	Status
Xenogryllus Bolívar 1890	valid genus (emended diagnosis, redescription)
X. eneopteroides Bolívar, 1890	valid species (redescription, type species)
X. carmichaeli (Chopard, 1928)	nomen dubium (new status)
X. lamottei Robillard n. sp.	new species
X. maichauensis Gorochov, 1992	valid species (redescription)
X. maniema Robillard & Jaiswara n. sp.	new species
X. marmoratus (Haan, 1844)	valid species (redescription, neotype, synonymy of subspecies)
X. mozambicus Robillard n. sp.	new species
X. transversus (Walker, 1869)	valid species (redescription)
X. ululiu Gorochov, 1990	valid species (redescription)

1344

1345

Table 2. Measurements of *Xenogryllus eneopteroides*

	BL	PronL	PronW	FWL	FWW	HWT	FIIIL		FIIIW
Males (n=6)	20-27.1	3.4 -3.9	6.1-7	21.5-24.	2 8-8.3	7.4-8.3	20.0-22.4		3.1-3.6
(Male mean)	(22.7)	(3.6)	(6.7)	(23.0)	(8.2)	(7.8)	(21.1)		(3.4)
Females (n=5)	16.9-22.2	3.8-4.1	5.8-6.0	20.0-20.	9 4.9-5.3	7.3-8.4	22.1-22.6		2.2-3.6
(Female mean)	(19.4)	(4)	(5.9)	(20.5)	(5.1)	(7.9)	(22.4)		(3)
	TIIIL			TIIIs			Tallis		OL
		las	lbs	Oas	Obs	lds	Ods	Ols	
Males (n=6)	20.0-21.5	9-13	9-11	18-21	8-13	0	6-7	3-6	-
(Male mean)	(20.2)	(10.5)	(9.8)	(19.3)	(10.8)	0	(6)	(5)	-
Females (n=5)	21.6-25.5	9-10	10-13	18-20	13-14	0	6-7	3-6)	8-9
(Female mean)	(23.6)	(9.5)	(11.5)	(19.0)	(13.5)	0	(6)	(5)	(8.42)

Table 3. Measurements of Xenogryllus lamottei n. sp.

	BL	PronL	PronW	FWL	FWW	HWT	FIIIL		FIIIW	
Holotype male	24.2	3.1	4.6	20	6.6	5.7	19.5		3.3	
	TIIIL			ΓIIIs			TallIs		OL	
		las	lbs	Oas	Obs	lds	Ods	Ols	-	
Holotype male	?	?	?	?	?	?	?	?	-	
1350										

Table 4. Measurements of Xenogryllus maichauensis

	BL	PronL	PronW	FWL	FWW	HWT	FIIIL		FIIIW	
Thailand										
Male	33.3	3.8	5.7	24	8.2	8.4	22.9		4.1	
Female	23.7	3.7	5	24.6	5.5	9.4	22.5		4.2	
China										
Males (n=5)	30.6-32.8	3.3-3.9	4.8-5.1	21.3-24	7.7-8.1	4-5.8	21-22		3.5-4.6	
(Male mean)	(31.4)	(3.7)	(5)	(22.7)	(7.8)	(5.2)	(21.6)		(3.9)	
Females (n=2)	24-27.4	3.4-3.9	4.8-5.3	15.5-18	4.4-5.5	5-8.5	22.1-22		3.7-5.5	
(Females mean)	(25.7)	(3.6)	(5)	(18.5)	(5.1)	(6.2)	(22)		(4.9)	
	TIIIL			TIIIs			Tallis		OL	
		las	lbs	Oas	Obs	lds	Ods	Ols		
Thailand										
Male	24.4	12	11	19	13	0	3	6	-	
Female	23	10	12	20	17	0	4	4	20.7	
China										
Males (n=5)	20-21	11-15	10-12	15	12-13	0	3	3	-	
(Male mean)	(20.5)	(12.3)	(11)	(15)	(12)	(0)	(3)	(3)	-	
					· · · · -			<u> </u>	00 0 0 1 0	
Females (n=2)	21-22	8-11	10-12	14-20	14-17	0	4-5	3-4	23.6-24.2	
Females (n=2) (Females mean)	21-22 (21.3)	8-11 (9)	10-12 (11)	14-20 (17)	14-1 <i>7</i> (16)	0 (0)	4-5 (5)	3-4 (4)	23.6-24.2 (23.9)	

Table 5. Measurements of Xenogryllus maniema n. sp.

	BL	PronL	PronW	V	FWL	FWW	HWT	FIIIL		FIIIW	
Holotype male	21.8	2.9	4		18.3	6.6	6.4	?		?	
Males (n=5)	19.8-22.9	2.7-3	3.8-4.4	1	18.5-21.3	6.4-7.6	5.6-6.4	18.3-1	9.1	3.1-3.2	
(Male mean)	(21.3)	(2.9)	(4.1)		(19.3)	(6.9)	(5.8)	(18.8)		(3.1)	
	TIIIL			TIIIs				TallIs		OL	
		las	lbs	Oas	Ob	s	lds	Ods	Ols		
Holotype male	?	?	?	?	?		?	?	?	-	
Males (n=5)	18.3-19.1	7-13	8-15	16-20	11-	16	0	5-9	7-10	-	
(Male mean)	(18.7)	(10)	(12)	(18)	(14	·)	(0)	(7)	(8)	-	
1354											

Table 6. Measurements of	Xenogryllus	marmoratus
--------------------------	-------------	------------

	BL	PronL	PronW	FWL	FWW	HWT	FIIIL		FIIIW
Neotype male	19.4	2.9	4.1	15.5	6.7	3.5	15.3		2.8
Males (n=4)	18.6-22	2.5-3	3.9-4.8	15.5-17	6.4-6.7	3.1-4.3	14.2-15	5.8	2.6-3.4
(Male mean)	(20.4)	(2.9)	(4.3)	(16.5)	(6.5)	(3.4)	(15.3)		(3)
Females (n=5)	18.8-20	2.1-3.1	3.1-4.3	11.5-15	2.8-4	1.2-6.3	11.8-16	6.3	2.2-3.2
(Female mean)	(19.1)	(2.7)	(3.7)	(13.4)	(3.3)	(3.4)	(14.4)		(2.7)
· · · · ·	TIIL			TIIIs	· · ·	, <i>,</i> ,	Tallis	6	ÔL
		las	lbs	Oas	Obs	lds	Ods	Ols	
Neotype male	11.1	9	11	14	16	0	4	5	-
Males (n=4)	11.1-15.8	5-12	10-12	11-20	11-16	0-2	3-7	5-7	-
(Male mean)	(14.1)	(9)	(11)	(16)	(5)	(1)	(5)	(5)	-
Females (n=5)	12.1-14.9	4-10	7-12	13-18	9-13	0-2	2-5	5-10	14.1-19
(Female mean)	(13.3)	(8)	(9)	(16)	(11)	(1)	(3)	(7)	(16.6)

Table 7. Measurements of Xenogryllus mozambicus n. sp.

	BL	PronL	PronW	FWL	FWW	HWT	FIIIL		FIIIW
Holotype male	25.5	3.3	4.5	20.5	7.3	7.5	21		3.6
Males (n=5)	22.5-25.5	2.9-3.3	4.3-4.5	21-22	7-7.2	7-7.5	19-20		3.1-3.9
(Male mean)	(23.8)	(3.2)	(4.4)	(21.2)	(7.2)	(7.3)	(20)		(3.5)
Female allotype	21.9	2.9	4.4	17.5	3.8	9	20.5		4
Females (n=4)	21.5-22.1	3-3.3	4.2-4.5	18-21	.5 4-7.7	7.5-8.5	19-20		3.5-4
(Female mean)	(21.8)	(3)	(4.3)	(19.2)	(5.3)	(7.9)	(19.9)		(3.7)
	TIIIL			TIIIs			TallIs		OL
		las	lbs	Oas	Obs	lds	Ods	Ols	-
Holotype male	20	13	10	15	19	1	8	4	-
Males (n=5)	18-20	9-13	9-14	11-16	16-21	1-2	8-10	4-5	-
(Male mean)	(18.2)	(11)	(12)	(14)	(18)	(1)	(9)	(5)	-
Female allotype	20	9	12	14	17	0	7	6	?
Females (n=4)	19-20	9-15	9-12	14-18	13-18	0-2	6-8	4-6	11.5-13.4
(Female mean)	(19.7)	(16)	(10)	(16)	(17)	(1)	(7)	(5)	(12.5)

Table 8. Measurements of Xenogryllus transversus

-	BL	PronL	PronW	FWL	FWW	HWT	FIIIL		FIIIW
Males (n=6)	27.8-34.6	3.4 -3.9	6.1-7	21.5-24.2	8-8.3	7.4-8.3	20.0-22.4		3-4
(Male mean)	(31.4)	(3.6)	(6.7)	(23.0)	(8.2)	(7.8)	(21.1)		(3.7)
Females	28.5-30	3.8-4.1	5.8-6.0	20.0-20.9	4.9-5.3	7.3-8.4	22.1-22.6		4.2-4.8
(n=2)									
(Female	(29.3)	(4)	(5.9)	(20.5)	(5.1)	(7.9)	(22.4)		(4.5)
mean)									
	TIIIL			TIIIs			TallIs		OL
		las	lbs	Oas	Obs	lds	Ods	Ols	_
Males (n=6)	20.0-21.5	9-13	9-11	18-21	8-13	0	2-5	2-3	-
(Male mean)	(20.2)	(10.5)	(9.8)	(19.3)	(10.8)	(0)	(3)	(3)	-
Females	21.6-25.5	9-10	10-13	18-20	13-14	0	2-3	4-5	25.8-27
(n=2)									
(Female	(23.6)	(9.5)	(11.5)	(19.0)	(13.5)	(0)	(3)	(5)	(26.4)
mean)			. ,						

1358 1359

Table 9. Measurements of Xenogryllus ululiu

	BL	PronL	PronW	FWL	FWW	HWT	FIIIL		FIIIW
Male paratype	28.2	3.5	4.3	22.2	7.9	7.1	22		3.6
Males (n=5)	23.3-28.2	3.3-3.5	4.2-4.8	20-22.2	7.1-8.1	3.3-7.9	19-22.7		3.3-4
(Male mean)	(26)	(3.4)	(4.5)	(21.5)	(7.7)	(5.8)	(21.3)		(3.7)
Females (n=3)	23.7-25.9	3.4-4	4.5-5	17.2-23.1	3.3-4	5.1-8.2	20-21.5		3.4-4.3
(Female mean)	(24.5)	(3.7)	(4.8)	(20.2)	(3.8)	(6.9)	(20.5)		(3.8)
	TIIL			TIIIs			Tallis		OL
		las	lbs	Oas	Obs	lds	Ods	Ols	
Male paratype	21.8	12	10	20	14	1	4	6	-
Males (n=5)	19.9-21.8	11-12	10-11	16-20	14-16	0-1	4-7	5-8	-
(Male mean)	(20.8)	(11)	(10)	(18)	(15)	(0)	(5)	(6)	-
Females (n=3)	19.8-20.7	7-10	10-11	12-16	14-16	Ô	3-5	5-7	13.9-17.6
(Female mean)	(20.1)	(9)	(11)	(14)	(15)	(0)	(4)	(6)	(17)

1360 Legends of figures

1361

Figure 1. (A-C) *Xenogryllus eneopteroides*: male in dorsal (A) and lateral (B) views,
female in dorsal view (C); (D-G) *Xenogryllus marmoratus* male in dorsal (D) and
lateral (E) views, female in dorsal (F) and lateral (G) views; (H-I) *Xenogryllus lamottei* n. sp.: male holotype in dorsal (H) and lateral (I) views; (J-K) *Xenogryllus maniema* n. sp.: male paratype in dorsal (J) and lateral (K) views;
(L-O) *Xenogryllus maichauensis*: male in dorsal (L) and lateral (M) views,
female in dorsal (N) and lateral (O) views. Scale bar: 1 cm.

Figure 2. (A-D) *Xenogryllus mozambicus* n. sp.: male in dorsal (A) and lateral (B)
views, female in dorsal (C) and lateral (D) views; (E-H) *Xenogryllus transversus*:
male in dorsal (E) and lateral (F) views, female in dorsal (G) and lateral (H)
views; (I-L) *Xenogryllus ululiu*: male in dorsal (I) and lateral (J) views, female in
dorsal (K) and lateral (L) views. Scale bar: 1 cm.

- Figure 3. Heads of *Xenogryllus* in facial (left panel) and lateral views (right panel): (A B) *Xenogryllus eneopteroides;* (C-D) *Xenogryllus lamottei* n. sp.; (E-F)
 Xenogryllus maichauensis; (G-H) *Xenogryllus maniema* n. sp.; (I-J) *Xenogryllus marmoratus.* (K-L); *Xenogryllus mozambicus* n. sp.; (M-N) *Xenogryllus ululiu;* (O-P) *Xenogryllus transversus.* Scale bar: 1 mm.
- Figure 4. Heads of Xenogryllus in dorsal views: (A) Xenogryllus eneopteroides; (B)
 Xenogryllus lamottei n. sp.; (C) Xenogryllus maichauensis; (D) Xenogryllus
 maniema n. sp.; (E) Xenogryllus marmoratus; (F) Xenogryllus transversus; (G)
 Xenogryllus ululiu; (H) Xenogryllus mozambicus n. sp. Scale bar: 1 mm.
- Figure 5. Male forewing venation in dorsal view: (A) Xenogryllus eneopteroides; (B)
 Xenogryllus lamottei n. sp.; (C) Xenogryllus maichauensis; (D) Xenogryllus
 maniema n. sp.; (E) Xenogryllus marmoratus; (F) Xenogryllus mozambicus n.
 sp.; (G) Xenogryllus transversus; (H) Xenogryllus ululiu. Scale bar: 5 mm.
- Figure 6. Male metanotal glandular structures in *Xenogryllus eneopteroides*: (A)
 glandular pit (drawing by G. Hodebert (MNHN) modified from Robillard &
 Desutter-Grandcolas, 2004b), (B-C), SEM view of glandular pores on scutum.
 Scale bars: 1 mm (A), 10 μm (B), 1 μm (C).
- Figure 7. Male genitalia: (A-C) *Xenogryllus eneopteroides*, dorsal (A), ventral (B) and
 lateral (C) views; (D-E) *Xenogryllus lamottei* n. sp., ventral (D), lateral (E) views;
 (F-G) *Xenogryllus maniema* n. sp., ventral (F), lateral (G) views; (H-I)
 Xenogryllus marmoratus, ventral (H), lateral (I) views; (J-K) *Xenogryllus*

1395mozambicusn.sp.,ventral(J),lateral(K)views;(L-N)Xenogryllus1396maichauensis,pseudepiphallic lophi in ventral (L-M) and lateral (N)views,male1397fromSouthChina(L),malefromThailand(M,N);(O-P)Xenogryllus1398transversus,dorsal(O),ventral(P)views;(Q-R)Xenogryllusululiu,1399pseudepiphallic lophi in dorsal(Q) and ventral(R)views.Scale bar:1 mm.

Figure 8. Right pseudepiphallic lophi of male genitalia in dorsal view: (A-B) Xenogryllus 1400 1401 eneopteroides, male from Central African Republic (A), male from Ivory Coast 1402 (B); (C) Xenogryllus lamottei n. sp.; (D) Xenogryllus maichauensis; (E) 1403 Xenogryllus maniema n. sp.; (F) Xenogryllus mozambicus n. sp.; (G) 1404 Xenogryllus marmoratus; (H) Xenogryllus ululiu; (I) Xenogryllus transversus. 1405 Symbols: Preapical hook-like expansion figured in gray or with a black arrow; 1406 dotted parts represent membranous areas; thin lines represent folds and reliefs 1407 in sclerites. Scale bar: 1 mm.

Figure 9. Apex of female subgenital plate and base of ovipositor in ventral view: (A) *Xenogryllus eneopteroides*; (B) *Xenogryllus marmoratus*; (C) *Xenogryllus maichauensis*; (D) *Xenogryllus mozambicus*; (E) *Xenogryllus transversus*; (F) *Xenogryllus ululu.* Scale bar: 1 mm.

1412 Figure 10. Apex of female ovipositor: (A) *Xenogryllus eneopteroides*; (B) *Xenogryllus maichauensis*. Scale bar: 1 mm.

Figure 11. Female copulatory papilla: (A-B) *Xenogryllus eneopteroides*, in ventral (A)
and lateral (B) views; (C-D) *Xenogryllus marmoratus*, in ventral (C) and lateral
(D) views; (E-G) *Xenogryllus maichauensis*, in ventral (E), dorsal (F) and lateral
(G) views; (H-J) *Xenogryllus mozambicus* n. sp., in ventral (H), dorsal (I) and
lateral (J) views; (K-L) *Xenogryllus ululiu*, in ventral (K) and lateral (L) views; (MO) *Xenogryllus transversus*, in ventral (M), dorsal (N) and lateral (O) views.
Scale bar: 0.5 mm.

Figure 12. Comparison of calling songs, oscillogram of five echemes: (A) Xenogryllus
 maichauensis; (B) Xenogryllus marmoratus; (C) Xenogryllus mozambicus; (D)
 Xenogryllus transversus; (E) Xenogryllus ululiu. Scale bar: 1 s.

Figure 13. Calling song of *Xenogryllus maichauensis*: (A) oscillogram of 5 echemes;
(B) detailed oscillogram (upper panel) and sonogram (lower panel) of 1
echeme; (C) frequency spectrum.

Figure 14. Calling song of *Xenogryllus marmoratus*: (A) oscillogram of 5 echemes; (B)
detailed oscillogram (upper panel) and sonogram (lower panel) of 1 echeme;
(C) detailed oscillogram of one syllable; (D) frequency spectrum.

Figure 15. (A-D) *Xenogryllus mozambicus* n. sp.: Savannah habitat in Mozambique
(A), male (B) and female (C) in bushy vegetation at night; male (D) predated by
Reduviidae sp.

- Figure 16. Calling song of *Xenogryllus mozambicus*: (A) oscillogram of 5 echemes; (B)
 detailed oscillogram (upper panel), amplitude envelope (middle panel) and
 sonogram (lower panel) of 1 echeme; (C) detailed oscillogram (upper panels)
 and sonograms (lower panels) of one the two initial syllables (left) and of the
 final syllable (right); (D) frequency spectrum.
- Figure 17. Calling song of *Xenogryllus transversus*: (A) oscillogram of 5 echemes; (B)
 detailed oscillogram (upper panel) and sonogram (lower panel) of 1 echeme;
 (C) frequency spectrum.

Figure 18. Calling song of *Xenogryllus ululiu*: (A) oscillogram of 5 echemes; (B) detailed oscillogram (upper panel) and sonogram (lower panel) of 1 echeme; (C) frequency spectrum.

1444









1449	Fig.3
1450	-




























Fig. 15









