

Structural and Functional Partitioning of Bread Wheat Chromosome 3B

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Title: Structural and Functional Partitioning of Bread Wheat Chromosome 3B

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Abstract:

We produced a reference sequence of the 1 Gb chromosome 3B of hexaploid bread wheat. By sequencing 8,452 Bacterial Artificial Chromosomes in pools we assembled a sequence of 774 Mb carrying 5,326 protein-coding genes, 1,938 pseudogenes, and 85% of transposable elements. The distribution of structural and functional features along the chromosome revealed partitioning correlated with meiotic recombination. Comparative analyses indicated high wheat specific inter- and intra-chromosomal gene duplication activities that are potential sources of variability for adaption. In addition to providing a better understanding of the organization, function, and evolution of a large and polyploid genome, the availability of a high quality sequence anchored to genetic maps will accelerate the identification of genes underlying important agronomic traits.

One Sentence Summary*:* The reference sequence of the bread wheat chromosome 3B reveals partitioning of structural and functional features along the chromosome.

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Main text:

Introduction: Bread wheat (*Triticum aestivum* L.) is a staple food for 30% of the world population. It is a hexaploid species (6x=2n=42, AABBDD) that originates from two interspecific hybridizations estimated to have taken place ~0.5 million and 10,000 years ago (*1*). The predicted closest extant representatives of the ancestral parental diploid species (2n=14) are *Triticum urartu* (A genome), *Aegilops speltoides* (S genome related to the B genome) and *Aegilops tauschii* (D genome). Each of the three ancestral genomes is about 5.5 Gb in size and, therefore, results in a highly redundant 17 Gb hexaploid genome with 3 homoeologous sets of 7 chromosomes (1-7A, 1-7B, 1-7D) each carrying highly similar gene copies. Moreover, most of the genome was shaped by the amplification of transposable elements (TEs) that include highly repeated families and sequences (*2*). This high redundancy has complicated the assembly of a complete and properly ordered reference sequence of the bread wheat genome. A fully sequenced genome enables scientists and breeders to have access to a complete gene set, with the gene order along each chromosome, and identify candidate genes between markers associated with important traits. It also enables the identification of recent duplicates, which may be involved in species-specific evolution (*3*), and to trace their evolutionary history. Prior to obtaining a full genome sequence, the wheat gene space has been investigated through various genome and transcriptome survey sequencing approaches and through microarray hybridizations (*4-7*). Recently, whole genome shotgun sequencing of *cv.* Chinese Spring using Roche/454 technology and synteny driven assembly yielded around 95,000 gene models (N50=0.9 kb; (*8*)). Furthermore, the gene space of the diploid wild relatives *Ae. tauschii* (DD) and *T. urartu* (AA) has also been assembled and led to describe 43,150 and 34,879 genes, respectively (*9, 10*). While these sequences are useful templates for marker design and comparative analyses, as a result of assembly limitations of short-read based sequencing (*11, 12*), they are still very fragmented and a large

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fraction of the genes are unanchored to chromosomes. The maize (*Zea mays*) and potato (*Solanum tuberosum*) sequencing projects, both representing species with highly repetitive genomes, were able to avoid over-fragmentation by combining multiple sequencing technologies and through the use of DNA libraries with a diversity of insert sizes (*13, 14*).

The International Wheat Genome Sequencing Consortium (IWGSC) road map focuses on physically mapping and obtaining a high quality reference sequence of each of the 21 individual wheat chromosomes rather than approaching the hexaploid genome as a whole. This strategy relies on flow sorting individual chromosomes and/or chromosome arms from ditelosomic lines of the cultivar Chinese Spring to construct BAC libraries (*15*). The largest chromosome is 3B (~1 Gb). It was the first chromosome for which a BAC library was constructed (*16*) and a physical map achieved (*17*). A pilot sequencing study on 13 contigs (*2*) suggested that genes tend to be mainly clustered into small islands, the presence of a 2-fold gene density increase from the centromere towards the telomeres, and a high proportion of nonsyntenic genes interspersed within a conserved ancestral grass gene backbone. It provided a proof of principle for this strategy and opened the way for producing a reference sequence of the large and polyploidy wheat genome.

Sequencing and construction of a pseudomolecule: We used a hybrid sequencing and BAC pooling strategy to sequence 8,452 BAC clones from the minimal tiling path (MTP) that was established during the construction of the chromosome 3B physical map (*4, 18*). After the integration of BAC-end sequences, manual curation of the scaffolding, gap filling and correction of potential sequencing errors (*18*), we obtained a final assembly of 2,808 scaffolds representing 833 Mb with a N50 of 892 kb (i.e. half of chromosome sequence is assembled in scaffolds larger than 892 kb). We estimated that about 6% of the chromosome sequence were not present in the MTP BAC-based assembly through comparison with the 546,922 contigs assembled from whole chromosome shotgun sequencing of flow sorted 3B

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DNA (*19*). This suggests that the size of chromosome 3B is nearly 886 Mb *i.e.* about 11% smaller than originally predicted (*16, 20*). We built a pseudomolecule of chromosome 3B by ordering 1,358 scaffolds along the chromosome using an ordered set of 2594 anchor Single Nucleotide Polymorphism (SNP) markers. The pseudomolecule represents 774.4 Mb (93% of the complete sequence) with a scaffold N50 of 949 kb (Table S1). The order of markers was determined by linkage analysis of a recombination inbred lines (RILs) population derived from a cross between *T. aestivum* cultivars Chinese Spring (reference sequence) and Renan (a French elite cultivar) and refined by integrating linkage disequilibrium data from two panels and physical BAC contig information (*18*). This sequence corresponds to an annotation-directed improved high-quality draft (*21*) situated between the high quality finished rice genome sequence and the improved draft maize genome (*13*).

Annotation of Genes, Transcribed Loci and Transposable Elements: Gene modeling led to predict 7,264 coding loci on the 3B pseudomolecule (Table 1), including 5,326 with a functional structure and 1,938 (27%) likely corresponding to pseudogenes. An additional 251 gene models and 188 pseudogenes were annotated in unanchored scaffolds. RNA-Seq data revealed that 71.4% of the predicted genes/pseudogenes are transcribed and led to identify 3,692 unannotated transcribed loci that may encode functional non-coding RNAs or unknown proteins, hereafter referred to as novel transcribed regions (NTRs, Table 1). In addition, 791 highly conserved non-coding RNA genes involved in RNA maturation and protein synthesis were also predicted (Table 1). Chromosome 3B appears to contain a high number of small nuclear RNA genes (U1 to U6) including 9 U1-snRNAs, 7 of which are tandemly duplicated. As a comparison, there are 14 U1-snRNAs in the entire *Arabidopsis thaliana* genome (www.plantgdb.org). The higher number of U1-snRNAs may reflect a higher level of duplication in the wheat genome. We found 53,288 complete and 181,058 truncated copies of transposable elements (TEs), belonging to 485 TE families and

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representing 85% (640 Mb) of the 3B pseudomolecule, through similarity-search approach. Further *de novo* repeat detection (*18*) identified 3.6% putatively new TEs.

We estimated the putative location of the centromeric region by plotting the density of the Long Terminal Repeats retrotransposons (LTR-RTs) CRW (Centromeric Retrotransposons of Wheat) and Quinta along the pseudomolecule. These LTR-RTs are recognized by the centromere-specific histone CenH3, and thus are centromere-functional sequences (*22*). Two major peaks covering a region of 122 Mb (265-387 Mb, Fig. S1) that includes 1 Mb previously shown as interacting with histone CenH3 (*22*) and encompassing the centromere of the orthologous rice chromosome 1 (*23*) were identified. This region was defined as the centromeric-pericentromeric region of chromosome 3B. A strong correlation has been observed between the size of the centromeres and the chromosomes in grasses (*24*) and it is likely that large chromosomes have centromeres larger than 10 Mb. This may be critical to ensure the structural rigidity of the pericentromeric regions needed for kinetochore co-orientation (*25*). Marker assignation to either short or long arm indicated the presence of a break point between 349.4 and 350.0 Mb that might be the position of the core centromere.

Variability in Recombination Rate and Gene Density Along the Chromosome: We found 787 crossover (CO) events on chromosome 3B in the Chinese Spring x Renan population with on average 2.6 COs per chromosome per individual, which is similar to maize (2.7-3.4; (*26*)). Distribution of meiotic recombination rate revealed extreme variations along the chromosome. While the average recombination rate is 0.16 cM/Mb, actual values range from 0 to 2.30 cM/Mb (per 10 Mb window; Fig. 1A). Segmentation analysis (*18*) revealed partitioning with the two distal regions of 68 Mb (region R1) and 59 Mb (region R3) on the short and long arms, respectively, showing recombination rates of 0.60 cM/Mb and 0.96 cM/Mb on average, and a large proximal region of 649 Mb (region R2) spanning the centromere with an average recombination rate of 0.05 cM/Mb (Table 2, Fig. 1A). This

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provides insight in the actual physical size of the highly recombinogenic regions previously detected at the end of the wheat chromosomes (*27, 28*). When a narrower window of 1 Mb was used, variations ranged from 0 to 12 cM/Mb (Fig. 1A), a range similar to that observed in maize (0.8-11.5 cM/Mb; (*26*)) and sorghum (0-10 cM/Mb; (*29*)). All crossover events occurred in only 13% of the chromosome in our population of 305 individuals. The largest region totally deprived of recombination corresponds to 150 Mb that includes the putative 122 Mb centromeric-pericentromeric region. This was confirmed by the linkage disequilibrium (LD) pattern (Fig. S2). Twenty-two regions showed a recombination ratio higher than 1.6 cM/Mb i.e. >10 times the average for this chromosome, and, thus, may contain recombination hot spots (Fig. 1A). However, no significant correlation was observed between the recombination rate and gene content, coding DNA, or TE content of these regions.

The 7,264 genes are not evenly distributed and gene density is increasing on both arms along the centromere-telomere axis correlating with the distance to the centromere (r_s=0.79, P<2.2e-16, R^2=0.61; Fig. 1B). Using a 10 Mb window, the average gene density estimate is 9±5 genes/Mb ranging from 1.3 in the centromeric-pericentromeric region up to 27.9 at the most telomeric end of the short arm, a pattern commonly observed in grass genomes. Variation of the gene density in wheat chromosome 3B is higher than for chromosomes in the more compact rice genome (*30*), lower than in sorghum where genes are mostly found in the telomeric regions (*31*), and in the same range as in maize which also contains a high percentage of TEs (*13*). Segmentation analysis revealed five major regions with contrasted gene densities (Fig. 1B) and a four-fold gradient of the gene density i.e. twice as much as suggested by the pilot study on chromosome 3B (*2*). The distal segments exhibiting the highest gene density (19 genes/Mb) correspond nearly to the highly recombinogenic R1 and R3 regions (Fig. 1A). The R2 region was subdivided into three segments with the lowest gene

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density (5 genes/Mb) in a 234 Mb segment encompassing the centromeric-pericentromeric region. As previously suggested (*2, 4*), there is no large region completely devoid of coding sequence (maximum of 3.7 Mb). We confirmed that the intergenic distances (IGDs) are extremely variable (average 104 kb+/-190 kb) and that a majority (73%) of the genes are organized in small islands, or "*insulae*" (*32*). This suggests that most of the intergenic regions are under selective constraint prevented from TE insertion. Indeed, only 29% of the IGDs are larger than 104 kb but they account for 81% of the chromosome size, demonstrating that TE-mediated genome expansion likely occurred within a limited number of intergenic regions.

Relationships Between Gene Expression, Function, and Chromosome Location: Of all annotated genes on chromosome 3B, 71.4% are expressed in at least one of the 15 conditions analyzed (5 organs at 3 developmental stages each; Table S2), 33% in all conditions, and 5% in one only (Fig. S3). On average, genes are expressed in 10.8 of 15 conditions (considering all predictions) and expressed genes are transcribed into 5.8 alternative transcripts, or isoforms. Both the expression breadth and the average number of isoforms are distributed unevenly along the chromosome, with a clear decrease of the two parameters towards the telomeres (Fig. 1C-D). Segmentation revealed distal segments with boundaries similar to that of regions R1 and R3 and with genes expressed in fewer conditions than in the proximal region: 8.7 *versus* 11.7, respectively (p-value<2.2e-16, Welch t-test; Table 2). Similarly, the average number of alternative transcripts is higher in the proximal (6.5) than in the distal (4.3) regions (p-value<2.2e-16, Welch t-test; Table 2).

Gene ontology (GO) term enrichment was estimated for the R1, R2 and R3 regions (*18*) (Tables S3-5). The distal regions are enriched in many GO categories, some being related to adaptation ("response to abiotic stimulus", "response to stress"). Well-known examples of genes related to adaptation are those involved in resistance to pathogens. Chromosome 3B carries 171 genes putatively associated with disease resistance (*18*) and their distribution is

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highly biased with 135 (79%) of them located in the distal regions (while these regions contain 33% of the gene set). Such uneven distribution and the correlation with the distribution of crossovers suggest that meiotic recombination acts as a main driver for creating variability in distal regions of chromosome 3B.

To investigate whether such partitioning is a common pattern of large plant genomes, we analyzed the distribution of the gene expression breadth in maize and barley that both exhibit large genome size (>1 Gb) and increased recombination rates at chromosomal extremities (*33, 34*). In barley, segmentation analysis of the 7 chromosomes based on recombination data identified the same pattern as on chromosome 3B with two highly recombinogenic distal regions and a large non-recombinogenic region. Using expression data of 8 conditions (*34*), we also observed that the two high-recombination distal regions carry genes expressed in fewer conditions than those carried by the low-recombination proximal regions (5.9 versus 6.7; p-value=2.2e-16, Welch t-test; Fig. S4A). Using GO-terms, we found a significant enrichment of these regions in categories "cell death" and "defense response" which support previous findings that barley disease resistance genes are clustered in the distal regions (*34*), In contrast, in maize, although we observed partitioning of the recombination rate, no gene expression partitioning was detected using RNA-Seq data of 18 conditions (*35*). Overall the expression breadth is 13.2 and 12.7 in high and low recombination regions, respectively, with chromosome specific patterns (Fig. S4B). Nevertheless, high recombination regions are also enriched in GO categories "cell death" and "defense response to fungus and bacteria" suggesting that such genes are consistently found in distal recombinogenic regions in large plant genomes. These results suggest that the partitioning observed on wheat chromosome 3B is conserved in the *Triticeae* and may not reflect a general pattern of large genomes. Alternatively, it is possible that the active rearrangements observed in the maize

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genome have modified this pattern. Additional evidence will come once other large plant genomes (1 Gb) are sequenced and analyzed.

Uneven Distribution of Transposable Elements: LTR-RTs represent 66% of the chromosome 3B sequence (gypsy: 47%, copia: 16%, unclassified: 3%; Table 1) which is slightly lower than the ~75% of LTR-RT identified in the whole maize genome (*36*). Only 4% (3/85) of the LTR-RT families were found in single copies compared to 41% in maize (*36*) and 48% in the rice genome (*37*). Sixteen percent of the sequence is comprised of Class II DNA transposons that mostly correspond to CACTA elements (Table 1), compared to 3.2% in the maize genome (*13*). Only 6 families account for 50% of the wheat chromosome 3B TE fraction as previously suggested from partial sequence analyses (*2, 38*) and from observations in other large genomes (*36*). However, in contrast to the maize genome in which most of the intact elements are found in 1-10 copies (*36*), the majority of the TE families annotated on chromosome 3B has a higher number of copies (10 to 1000 copies). Estimated insertion dates for the most abundant LTR-RT families showed a major peak at 1.5 million years (MY) but also quite specific patterns of TE activity for each family (Fig. S5). Our data support the hypothesis (*2, 38*) that most of the transposable elements that shaped the B genome inserted before polyploidization (0.5 million years ago (MYA)) and have been less active since then. Distribution of recently inserted elements revealed that TE insertion occurred at a similar rate in the distal and proximal regions. In contrast, older insertions (>1.5 MYA) were 1.7 fold more abundant in the R2 region compared to the R1 and R3 regions, suggesting a higher rate of TE elimination in the distal ends of chromosome 3B.

The TE density distribution was not random (Fig. 1E) with a lower density in the R1 (73%) and R3 (68%) regions compared to the R2 region (88%, Table 2). The 122 Mb centromeric-pericentromeric region displayed the highest density (93%) of TEs. Beneath the global TE distribution pattern, each superfamily presents its own specificities (Fig. S6). For

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example, CACTA transposons are more abundant in the distal gene-rich regions (Table 2), supporting *in situ* hybridization findings at the whole genome level (*39*). In addition, the distribution of TE families varied on the basis of their relative distance to genes (*18*) (Fig. S7). DNA transposons Mutator, Harbinger and MITEs are found in close vicinity to genes whereas LTR-RTs and CACTAs tend to be located at much larger distances from the genes. For instance, the 17,479 annotated MITEs were significantly found associated with genes (r=0.89; p-value<1e-10) as previously observed in plant genomes (*40*).

Synteny Between Chromosome 3B and Related Grass Genomes: Comparative genomics in grasses has been used to define syntenic relationships between different species (*41, 42*) and to provide insight into their evolution since the divergence from a common ancestor 50-70 MYA (*43*). We compared the wheat chromosome 3B genes (Ta3B) with the closest sequenced relative, *Brachypodium distachyon* (common ancestor: 32-39 MY; (*44*)), and with one representative of each of the *Ehrhartoideae* and *Panicoideae grass* subfamilies: *Oryza sativa indica* (rice, (*30*)) and *Sorghum bicolor (31)*, respectively. Wheat chromosomes of group 3 are syntenic with chromosome 1 of rice (Os1), chromosome 3 of sorghum (Sb3), and the distal parts of *B. distachyon* chromosome 2 (Bd2). We first investigated potential gene loss following polyploidization by using the conserved and syntenic genes found on chromosomes Os1, Bd2 and Sb3. These represent the grass core genes that are expected to be present on wheat homoeologous group 3, unless they have been lost by fractionation following polyploidization. The finding that 94% of the conserved genes are also present on the 3B sequence (Fig. 2A), which represents 94% of the chromosome (see above), suggests that no major gene loss occurred in the B subgenome yet. This is confirmed at the whole genome level by the results of the chromosome survey sequences (*19*). In contrast, 2,065 genes on chromosome 3B (34.6%; including pseudogenes) shared similarity with genes on non-orthologous chromosomes in the other grass genomes. This proportion of nonsyntenic

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genes is much higher than the 5% (between 149 and 207) of nonsyntenic genes found in the other grass species analyzed (Fig. 2A, Table S6). It confirms previous results showing substantial modifications and rearrangements of the wheat gene space (*2*). When looking at the conservation of the gene order, collinear genes represent 42 to 68% of the genes present on Os1, Bd2 and Sb3, whereas they represent less than 30% of the Ta3B genes (including pseudogenes; Table S7 and Fig. S8). The spatial distribution of syntenic and nonsyntenic genes along the 3B pseudomolecule (Fig. 2B) shows an increased proportion of nonsyntenic genes in the R1 (44%) and R3 (53%) regions compared to the R2 region (28%; Table 2). This supports the hypothesis that accelerated evolution occurred in the wheat lineage compared to other grasses (*2, 45, 46*) with insertions of nonsyntenic genes intercalated in the ancestral grass genome backbone via gene duplications or translocations that preferentially occurred in the distal recombining regions.

Origin and Evolution of Nonsyntenic Genes: With such a high proportion of nonsyntenic genes, one key question is whether these genes are under selection pressure or in the process of becoming pseudogenes. On the basis of the coding sequence structure, 32% of the nonsyntenic genes (*vs*. 17% of syntenic genes) were annotated as likely pseudogenes or gene fragments. This ratio is not surprising given that TE activity can duplicate gene fragments that are dead upon arrival. Expression patterns revealed that a majority of the nonsyntenic genes (69% *vs.* 82% of syntenic genes) are expressed in at least one condition tested (Table S8) thereby suggesting that a large fraction of these relocated genes are unlikely to be pseudogenes and may contribute to recent wheat genome evolution, and, therefore, to adaptation. Interestingly, a majority (51%) of the genes expressed in a single condition corresponds to nonsyntenic genes whereas 80% of the genes that are expressed in all 15 conditions are syntenic genes (Fig. S9). This suggests that nonsyntenic genes are involved in specific processes that may be related to adaptation while syntenic genes tend to be associated

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with essential biological processes. This hypothesis is supported by the fact that putative resistance genes identified on chromosome 3B are mainly nonsyntenic genes (*18*). In addition, GO term enrichment of nonsyntenic genes revealed an overrepresentation of genes involved in response to stress (Table S9).

The fact that chromosome 3B exhibits a higher number of genes than its orthologs in other grasses and that at least 94% of the ancestral grass gene backbone is conserved, indicate that most insertions of nonsyntenic genes result from inter-chromosomal duplication with retention of the parental copy. To test this hypothesis, we used the sequences of the 18 bread wheat chromosomes non-homoeologous to group 3 chromosomes (*19*) to search for potential parental copies of chromosome 3B genes elsewhere in the genome (*18*) (Table S10). A paralog was identified for 87% of the nonsyntenic genes (*18*), with no bias regarding the chromosomal origin of the inter-chromosomally duplicated genes (Fig. S10). Duplications of DNA fragments to different locations in a genome have been shown to result from double strand break (DSB) repair (in which a copy of the foreign DNA is used as filler to repair the break) or capture by active TEs (*46, 47*). We analyzed the composition of the regions flanking the syntenic *versus* nonsyntenic genes (20 kb on each side) and found a high association of nonsyntenic genes with a class II transposon superfamily: 41% more CACTAs were found around nonsyntenic genes than around syntenic genes (Fig. S11). CACTA transposons are known to capture genes (*31, 48*) and may have contributed significantly to inter-chromosomal gene duplications in wheat.

We also investigated the time since duplication of nonsyntenic genes via the analysis of nucleotide substitution rates (Ks) (*18*). In total, 62% of these duplications were older than 10 MY and, thus, are likely shared within other *Triticeae* species whereas 37% are potentially wheat-specific. Comparison with the barley genome survey sequence data (*34*) showed that at least 29% of the 3B nonsyntenic genes (vs. 51% of the syntenic genes) are orthologous with

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barley chromosome 3H, confirming that part of the nonsyntenic genes were relocated before the divergence of wheat and barley 10 to 14 MYA.

We next asked if the high gene duplication activity is also observed at the intra-chromosomal level. We identified 809 gene families with two or more copies comprising 2,216 genes on chromosome 3B, which is about 3 times more than in rice, *Brachypodium,* and sorghum (Table S11). This indicates that, in proportion, more than twice as many genes were duplicated or retained after intra-chromosomal duplications in wheat (~37%) compared to the other three grasses (~15-18%). About 46% of the duplicated genes of chromosome 3B are found in tandem whereas 54% are dispersed duplicates ((*18*); Table S12). In other grass species, a majority of the duplicated genes are organized in tandem. Given the high inter-chromosomal duplication activity observed in our analyses (see above), it is possible that some dispersed duplicates on chromosome 3B originated via independent inter-chromosomal duplications rather than through intra-chromosomal duplications thereby leading to overestimate these latter. However, even when considering syntenic dispersed duplicates, *i.e.* those genes that have remained at their ancestral locus and have undergone intra-chromosomal duplication, 23% of the whole gene set appears to have originated from recent intra-chromosomal duplications which is still higher than in other grass species. Thus, we conclude that both inter- and intra-chromosomal rates of duplication are higher in wheat than in the other grass species analyzed so far. Interestingly, inter-chromosomal duplicates were distributed uniformly along the chromosome whereas the proportion of tandem duplicates slightly increased in the distal regions (Fig. 2D). This suggests that long distance and tandem duplications likely arose through different mechanisms. Finally, expression analysis of intra-chromosomal duplicated genes indicated that 49% of the families show expression of all copies in at least one condition. Similar to what was observed for the inter-chromosomal duplicated genes, the intra-chromosomal duplicated genes tend to be expressed in fewer

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conditions as compared to non-duplicated genes (Fig. S9; Table S8), suggesting they may be undergoing subfunctionalization.

QTL mining: As exemplified in rice and other crops, a reference genome sequence provides a resource for gene discovery, marker development, and allele mining in support of crop improvement (*49*). We identified 153,190 and 35,579 Insertion Site-Based Polymorphism (ISBP; (*50*)) and microsatellite markers, respectively, along the 3B chromosome. We also located 121 quantitative trait loci (QTLs) for 50 different traits on chromosome 3B (Table S13). Using these data, we conducted a meta-analysis that integrates QTLs defined in independent studies (*51*) and identified 18 metaQTLs with confidence intervals covering between 1.5 and 620 Mb of the chromosome 3B sequence. The largest one encompasses the centromeric region where recombination is suppressed. Five metaQTLs with small intervals (<10 Mb) that contain between 23 and 266 protein-coding genes and between 511 and 4,049 markers are suitable for fine mapping (Table S14).

Discussion: We present a reference sequence of chromosome 3B that can be used to precisely delineate structural and functional features along a chromosome and establish correlations between recombination intensity, gene density, gene expression, and evolution rate. Our results indicate that during evolution, regions with distinct features become delineated along chromosome 3B, including relatively small distal regions that are preferential targets for recombination, adaptation and genomic plasticity. Whether our observations reflect a general pattern for the wheat genome will need to be confirmed by the analysis of other chromosome reference sequences. Already, some of the features such as the CACTA distribution, the high rate of intra-chromosomal duplication, the absence of major gene loss since polyploidization and the gradient of gene density have been confirmed at the whole genome level (*19, 39*). Moreover, the ordered chromosome 3B sequence allowed us to distinguish duplicated genes and provided evidence for superimposed mechanisms of gene

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duplications. The high level of gene duplication (allopolyploidy, inter- and intra-chromosomal duplications) provides the wheat genome with a vast reservoir of functional genes that likely contribute to wheat adaptation.

On the basis of this work, the IWGSC has already defined an adapted BAC pooling strategy to reach the same sequence quality while reducing sequencing cost for the remaining chromosomes. While progress in sequencing technologies and cost reductions allow for more cost efficient sequence production, the challenge of bioinformatics and limitations of current sequencing technologies remain (*12*). Solving this issues and improving methods to efficiently anchor and orientate scaffolds within pseudomolecules should make the assembly of high quality reference sequences of complex genomes routine work in the future. There is no doubt that, as witnessed after the release of the rice genome sequence (*49*), the number of genes cloned from wheat will grow exponentially in the near future thereby enabling wheat researchers and breeders to cope with the urgent need of improving wheat yield in the face of climate change and food security challenges (*52*).

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Materials and Methods:

Sequencing, assembly and scaffolding: 8452 BACs representing the Minimal Tiling Path of wheat chromosome 3B were pooled into 922 BAC-pools. Each pool was used to create a barcoded Roche/454 8-kb long paired-end library. In total, 150 sequencing runs were performed, leading to an average of 36-fold sequence coverage. After assembly with Newbler (Roche), we integrated 42,551 BAC end sequences to validate and improve scaffolding. Illumina reads generated from sorted DNA of chromosome 3B were used to fill gaps within scaffolds and correct potential sequencing errors remaining in the consensus sequence (*18*).

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Anchoring scaffolds: SNP discovery was performed through sequence capture for 52,265 loci flanking TE junctions representing an average density of one locus per 16.2 kb (*18*). Out of 39,077 SNPs distributed along the chromosome, a subset of 3,075 evenly distributed (38.2+/-9.4 SNPs/10 Mb) SNPs was selected to genotype 1025 lines from recombination inbred and association panels. An anchor genetic map was built first by linkage analysis and integration of linkage disequilibrium data. A consensus map comprising 5,318 markers was also built using 40 different genetic maps to anchor additional scaffolds (*18*). Finally, a position in the pseudomolecule was inferred for scaffolds without marker information but belonging to an anchored physical BAC contig.

Sequence annotation: Gene modeling was performed using an improved version of the TriAnnot pipeline (*53*). Non-coding RNA genes were predicted using three different programs (*18*) and predictions were manually curated. Predictions of transposable elements (TEs) and reconstruction of the pattern of nested insertions were performed through the development a of specific program (*18*) that automatically curate similarity-search results obtained with a dedicated databank, comprising 4,929 known wheat TEs classified into 521 families.

Gene expression analyses: Thirty RNA samples, corresponding to RNAs extracted in duplicates from five organs (root, leaf, stem, spike, and grain) at three developmental stages each from hexaploid wheat *cv.* Chinese Spring (*4*), were used for gene expression analyses. RNA-Seq libraries were constructed using the IlluminaTruSeq™ RNA sample preparation Kit and sequenced. An average of 50+/-11 million paired end reads per sample were mapped on the chromosome 3B scaffolds and used to reconstruct transcripts and estimate transcript abundance in units of fragments per kb of exon per million mapped reads (FPKM). Regions with FPKM values higher than zero were considered as expressed.

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Distribution and segmentation analyses: Distributions of recombination rate, gene and TE densities, and expression breadth, were calculated within a sliding window of 10 Mb (and 1 Mb for the recombination rate) with a step of 1 Mb along the chromosome sequence using a homemade Perl script. Segmentation analyses of these distributions were performed using the R package *changepoint* v1.0.6 (*54*) with Segment Neighbourhoods method and BIC penalty on the mean change.

Comparative genomics, gene duplications and molecular evolution: We performed an all-by-all BLASTP (cutoff Evalue: 1e-5) comparison between the amino-acid sequences of predicted genes of wheat chromosome 3B, rice (MSU version 7.0), *Brachypodium* (*Brachypodium* Sequencing Initiative, 2.0), and sorghum (phytozome, version 1.4). We filtered out genes with no homology with at least one other gene in a compared species (cutoff 35% amino acid identity and 35% sequence overlap). Syntenic genes were defined as genes with a reciprocal best BLAST hit on an orthologous chromosome in at least one other species. Nonsyntenic genes were defined as genes for which best BLAST hit was on a non-orthologous chromosome in the other species. Clustering of orthologous and paralogous genes was performed using OrthoMCL (e-value cutoff: 1e-5, percent match cutoff: 35%; (*55*). All 3B genes clustered into the same family were considered intra-chromosomal duplicates. 3B genes clustered in a family with wheat gene models annotated on another chromosome (*19*), not including genes from group 3, were considered as inter-chromosomal duplicates. Tandem duplicates were defined as genes in the same family with 5 or less spacer genes separating them on the pseudomolecule, and dispersed duplicates were defined as having more than 5 spacer genes. Synonymous (Ks) and nonsynonymous (Ka) substitution rates were calculated based on ClustalW 2.1 (*56*) coding sequence alignments by the Nei and Gojobori method using codeml (part of the PAML package; (*57*)). Age of gene divergence was estimated by the equation Ks/2r, where r=6.5e-9.

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The chromosome 3B BAC library and the pools of the MTP BAC clones are available upon request under a Material Transfer Agreement at the French Plant Genomic Center, INRA-CNRGV.

Annotation data and browser are available at https://urgi.versailles.inra.fr/gb2/gbrowse/wheat_annot_3B. Sequences and annotations of the reference pseudomolecule and unassigned scaffolds have been deposited in ENA (project PRJEB4376) under the accession numbers HG670306 and CBUC010000001-

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Figure legends

Figure 1. Structural and functional partitioning of wheat chromosome 3B. Distribution and segmentation analysis of (A) meiotic recombination rate (cM/Mb in sliding window of 10 Mb in black, and 1 Mb in red); (B) gene density (CDS/Mb); (C) expression breadth; (D) average number of alternative spliced transcripts per expressed gene; (E) TE content along the 3B pseudomolecule. Distal regions of the chromosome R1 and R3 are represented in red; C: centromeric/pericentromeric region (black). The borders of these regions are indicated in Mb. Sliding window size: 10 Mb, step: 1 Mb.

Figure 2: Inter- and intra-specific comparative analyses of the gene content of wheat chromosome 3B. (A) Venn diagram displaying the number of genes conserved between wheat chromosome 3B (Ta3b, blue) and orthologous chromosomes in rice (Os1, red), *Brachypodium* (Bd2, green), and sorghum (Sb3, purple). The number of nonsyntenic genes is indicated in bold for each species. Distribution along the 774 Mb of the chromosome 3B pseudomolecule of the relative proportion of (B) syntenic (blue) vs. nonsyntenic (red) genes, (C) inter-chromosomal duplications (duplicates in red, group 3 specific genes in blue), and (D) tandem (yellow) and dispersed (red) intra-chromosomal duplications and singletons (blue). Chromosome 3B is represented at the bottom with distal regions in red and the centromeric/pericentromeric region in black.

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Table 1. General features of the 3B pseudomolecule

Pseudomolecule sequence				
	Length (bps)		774,434,471	
	G+C content		46.16%	
Protein coding genes		all	full genes	pseudogenes
	#genes	7,264	5,326	1,938
	average size (bps) of coding sequences (+/-standard deviation)	1,095+/-807	1,187+/-821	840+/-710
	average number of exons (+/-standard deviation)	4.2+/-4.4	4.4+/-4.6	3.6+/-3.8
	gene density (kb^{-1})	107	145	400
	#expressed genes	5,185	4,125	1,060
	#gene with alternative splicing	3,185	2,596	589
	%genes with alternative splicing	61%	63%	56%
	average no. isoforms/expressed gene	5.8	5.8	5.8
	NTRs	3,692		
Non coding RNA genes				
	tRNA	589		
	5S rRNA	85		
	Others (snRNA, snoRNA)	117		
	Total	791		
Transposable elements (TEs)				
Class I	Copia	15.6%		
	Gypsy	46.9%		
	Unclassified LTR-retrotransposons	3.5%		
	LINEs	1.2%		
	SINEs	0.01%		
	Total class I	**67.1%**		
Class II	CACTA	16.4%		
	Harbinger	0.19%		
	Mariner	0.19%		
	Mutator	0.43%		
	hAT	0.02%		
	Unclassified class II with TIRs	0.22%		
	Unclassified class II	0.10%		
	Helitron	0.01%		
	Total class II	**17.6%**		
Unclassified repeats		**0.81%**		
Total TEs		**85.5%**		

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Table 2. Distribution of features in the three regions of chromosome 3B as defined from the recombination segmentation along the chromosome

	R1	R2	R3
size (Mb)	68	648	59
recombination rate (cM/Mb)	0.60	0.05	0.96
Genes			
predicted gene density (Mb^{-1})	19	7	19
number of predicted genes/pseudogenes	1,318	4,845	1,101
# full genes	910 (69%)	3682 (76%)	734 (67%)
# pseudogenes/gene fragments	408 (31%)	1163 (24%)	367 (33%)
mean intergenic distance (kb)	49	130	52
# Expressed predicted genes	823 (62%)	3629 (75%)	733 (67%)
# Expressed full genes	621	2963	541
# Expressed pseudogenes/fragments	202	666	192
average expression breadth (per expressed gene; /15)	8.8	11.7	8.6
average FPKM (per expressed gene)	141	255	156
average number of isoforms (per expressed gene)	4.2	6.5	4.4
Proportion of nonsyntenic genes*	44%	28%	53%
Proportion of intra-chromosomally duplicated genes*	49%	33%	42%
Proportion of tandemly duplicated genes*	24%	14%	22%
Proportion of dispersed duplicated genes*	26%	18%	20%
Proportion of inter-chromosomally duplicated genes*	36%	33%	37%
Transposable Elements	73.0%	88.3%	68.4%
Copia	14.7%	15.8%	14.1%
Gypsy	31.7%	50.3%	27.1%
CACTA	18.7%	15.9%	19.5%

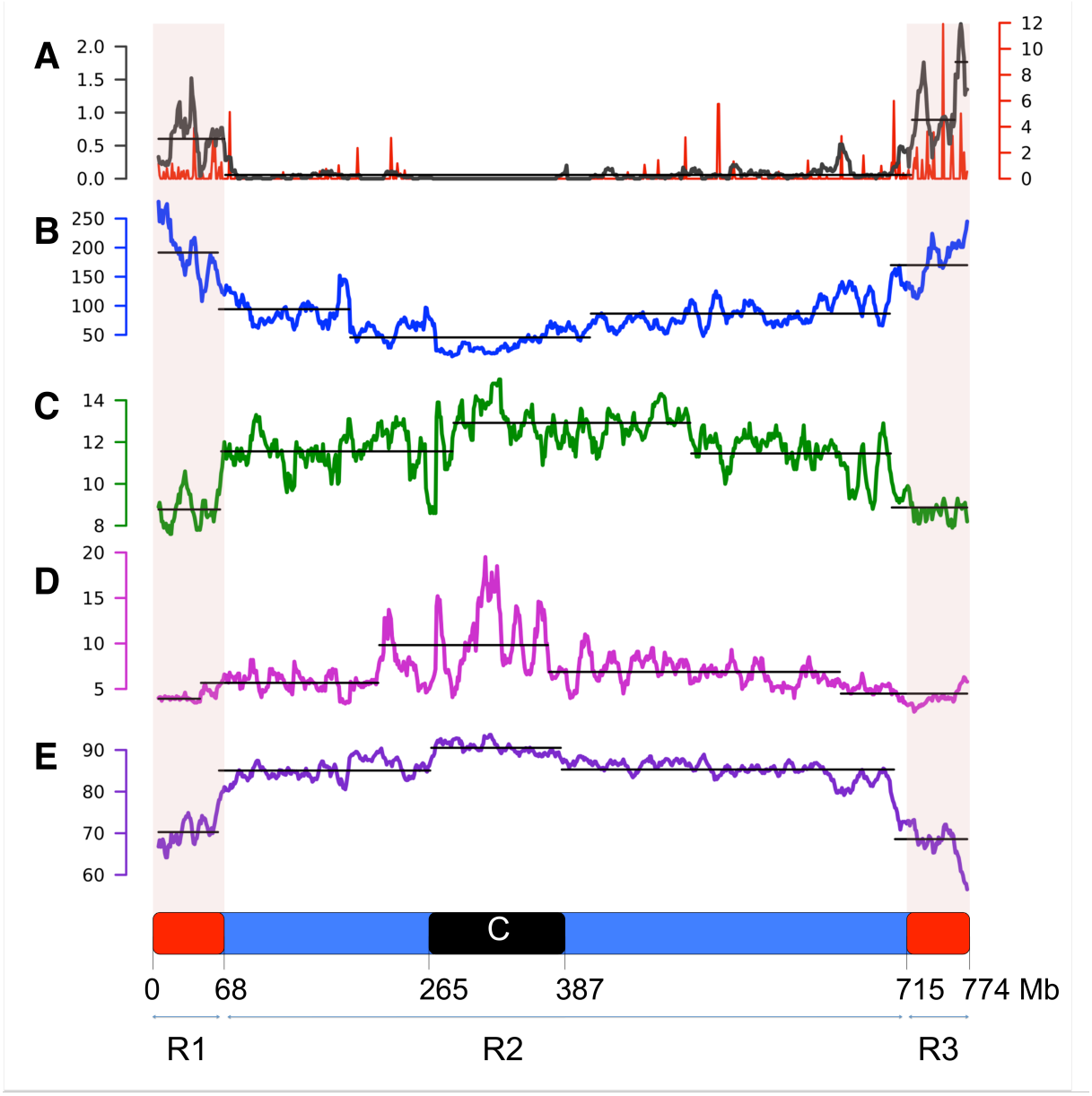
IGD: intergenic distance

*number of duplicated genes (filtered set; including pseudogenes) divided by the total number of genes in each region.

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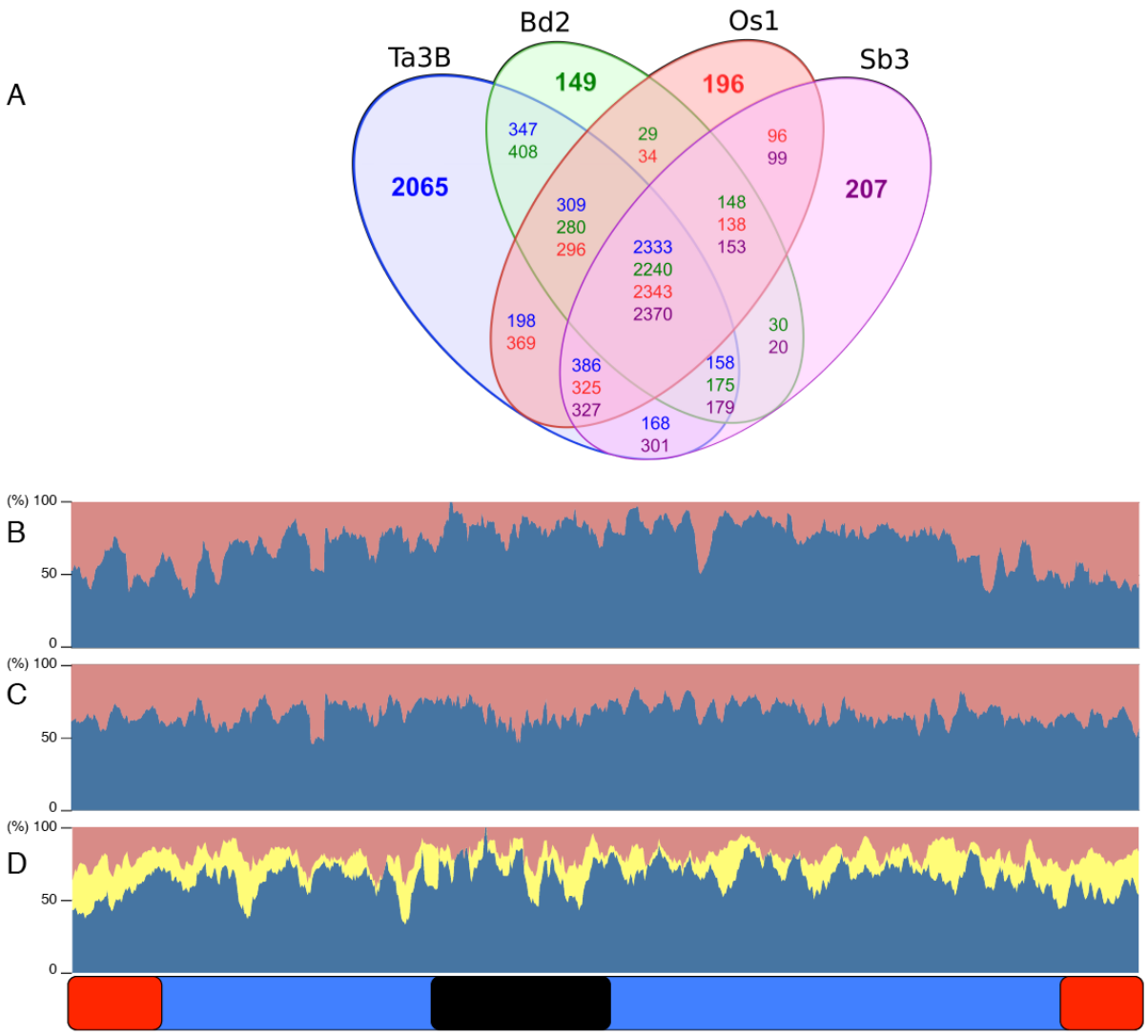
Figure 1.



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Figure 2.



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